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Ultrastructure of the Jurassic serpulid tubes - phylogenetic and paleoecological implications

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The ultrastructural diversity of the Middle and Late Jurassic serpulid tubes from the Polish Basin has been investigated. The inspection of 12 taxa representing the two major serpulid clades allowed for the identification of three ultrastructure kinds – irregularly oriented prismatic structure (IOP), spherulitic prismatic structure (SPHP), and simple prismatic structure (SP). Six of the studied species are single-layered and six species possess two distinct layers. Ultrastructural diversity corresponds to certain serpulid clades. The members of Filograninae have single-layered tube walls composed of possibly plesiomorphic, irregularly oriented prismatic structure (IOP). Two-layered tubes occur solely within the clade Serpulinae, where the external, denser layer is built of either the ordered spherulitic (SPHP) or simple prismatic microstructure (SP), and the internal layer is composed of irregularly oriented prismatic structure (IOP). Apart from phylogenetic signals provided by the tube ultrastructure, it can be used in analyzing paleoecological aspects of tube-dwelling polychaetes. Compared to the more primitive, irregularly oriented microstructures of Filograninae, the regularly oriented microstructures of Serpulinae need a higher level of biological control over biomineralization. The advent of the dense outer protective layer (DOL) in serpulids, as well as the general increase in ultrastructure diversity, was likely a result of the evolutionary importance of the tubes for serpulids, enhanced also by the intensified predation during the Mesozoic Marine Revolution. Such diversity of the tube ultrastructural fabrics allowed for maximizing functionality by utilizing a variety of morphogenetic programs. The biomineralization system of serpulids remains more complex compared to other tube-dwelling polychaetes. Physiologically more expensive tube formation allows for mechanical strengthening of the tube by building robust, strongly ornamented tubes and firm attachment to the substrate. Contrary to sabellids, which perform a fugitive strategy, an increased tube durability allows serpulids a competitive advantage over other encrusters.

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11

12 **Abstract**

13 The ultrastructural diversity of the Middle and Late Jurassic serpulid tubes from the
14 Polish Basin has been investigated. The inspection of 12 taxa representing the two major serpulid
15 clades allowed for the identification of three ultrastructure **kinds** – irregularly oriented prismatic
16 structure (IOP), spherulitic prismatic structure (SPHP), and simple prismatic structure (SP). Six
17 of the studied species are single-layered and six species possess two distinct layers.

18 Ultrastructural diversity corresponds to certain serpulid clades. The members of Filograninae
19 have single-layered tube walls composed of possibly plesiomorphic, irregularly oriented
20 prismatic structure (IOP). Two-layered tubes occur solely within the clade Serpulinae, where the
21 external, denser layer is built of either the ordered spherulitic (SPHP) or simple prismatic
22 microstructure (SP), and the internal layer is composed of irregularly oriented prismatic structure
23 (IOP). Apart from phylogenetic signals provided by the tube ultrastructure, it can be used in
24 analyzing paleoecological aspects of tube-dwelling polychaetes. Compared to the more
25 primitive, irregularly oriented microstructures of Filograninae, the regularly oriented
26 microstructures of Serpulinae need a higher level of biological control over biomineralization.

27 The advent of the dense outer protective layer (DOL) in serpulids, as well as the general increase
28 in ultrastructure diversity, was likely a result of the evolutionary importance of the tubes for
29 serpulids, enhanced also by the intensified predation during the Mesozoic Marine Revolution.
30 Such diversity of the tube ultrastructural fabrics allowed for maximizing functionality by
31 utilizing a variety of morphogenetic programs. The biomineralization system of serpulids
32 remains more complex compared to other tube-dwelling polychaetes. Physiologically more
33 expensive tube formation allows for mechanical strengthening of the tube by building robust,
34 strongly ornamented tubes and firm attachment to the substrate. Contrary to sabellids, which
35 perform a fugitive strategy, an increased tube durability allows serpulids a competitive advantage
36 over other encrusters.

37

38 **Introduction**

39 Numerous polychaete families produce habitation tubes (Rouse and Pleijel, 2001).
40 Tubicolous polychaetes may either agglutinate exogenous material, such as sand particles and
41 shell fragments, using a proteinaceous cement to form a tube (e.g., Stewart et al., 2004; Zhao et
42 al., 2005; Fournier, Etienne and Le Cam, 2010; Vinn and Luque, 2013) or produce secretions by
43 themselves utilizing a variety of glands (e.g., Hausen, 2005; Tanur et al., 2010). Secreted tubes
44 may be composed of organic substances, such as proteins and polysaccharides (e.g., Barnes,
45 1965; Chamoy et al., 2001; Nishi and Rouse, 2013), and mineral substances such as calcium
46 carbonate (Weedon, 1994; Fischer, Pernet and Reitner, 2000; Vinn et al., 2008). Amongst tube-
47 dwelling polychaetes producing hard, mineralized exoskeletons are the families Sabellidae,
48 Cirratulidae, and Serpulidae. Whereas calcareous sabellids (Perkins, 1991; Vinn, ten Hove and
49 Mutvei, 2008; Słowiński, Banasik and Vinn, 2023) and cirratulids (Reish, 1952; Fischer, Oliver
50 and Reitner, 1989; Fischer, Pernet and Reitner, 2000; Taylor et al., 2010; Kočí et al., 2021) are
51 restricted to one (*Glomerula*) and two genera (*Dodecaceria*, *Diplochaetetes*) in each family
52 respectively, only serpulids dwell exclusively in tubes composed of calcium carbonate (Vinn et
53 al., 2008).

54 Thus, having a hard, mineralized exoskeleton, serpulids are the most abundant
55 polychaetes in the fossil record, appearing as far back as the Middle Permian (Sanfilippo et al.,
56 2017; 2018). However, their hard parts are not strictly related to the organism's soft body and
57 form only a habitation tube serving, among others, for protection against predators (e.g., Morton

58 and Harper, 2009; Klompmaker, 2012). Although tubes' taxonomic usefulness is not without
59 validity, as many genera possess easily recognizable tubes, which are diagnostic displaying
60 different longitudinal and transverse elements (see e.g., Ippolitov et al., 2014), homeomorphy is
61 still a common feature in serpulid tube morphology (e.g., Kupriyanova and Ippolitov, 2015) due
62 to convergence and high ecophenotypic plasticity. This leads to certain discrepancies between
63 paleontological and biological classification, the latter of which is based solely on the molecular
64 data, soft parts, and their reciprocal relationships (Kupriyanova, Macdonald and Rouse, 2006; ten
65 Hove and Kupriyanova, 2009). Moreover, it makes the linkage of contemporary species with
66 their ancestors difficult due to the lack of fully reliable taxonomic tools.

67 Serpulid tubes can be composed of up to four layers; however, most of them are single-
68 layered (Vinn et al., 2008). They exhibit different ultrastructural fabrics, depending on how
69 calcium carbonate crystals are arranged and oriented (see Vinn et al., 2008). During the
70 progressing expansion of the tube, an animal secretes consecutive growth lamellae, which
71 correspond to a single growth episode. Such lamellae may be either straight, perpendicular to the
72 direction of growth (e.g., Bałuk and Radwański, 1997) or much more frequently chevron-shaped
73 (see Weedon, 1994). Serpulid ultrastructures may provide phylogenetic signals in the case of
74 Jurassic taxa but also serve as a record of the physiological changes of the worm during its entire
75 lifespan, providing important ecological signatures that may be successfully implemented in
76 paleoecological analyses. Moreover, serpulid tubes may exhibit additional skeletal structures
77 such as tubulae, alveolar structures, or internal tube structures, helping in unraveling their true
78 systematics (e.g., Pillai, 1993; Pillai and ten Hove, 1994; Jäger, 2005).

79 Recent studies have proven a tube microstructure to be a useful tool in deciphering the
80 true affinity of fossil serpulid (e.g., Bornhold and Milliman, 1973; Vinn, ten Hove and Mutvei,
81 2008; Vinn and Kupriyanova, 2011; Kupriyanova and Ippolitov, 2012; Vinn, 2013; Buckman,
82 2020; Buckman and Harries, 2020), and other calcareous tube-dwelling polychaete species
83 (Vinn, ten Hove and Mutvei, 2008; Taylor et al., 2010; Słowiński, Banasik and Vinn, 2023).
84 Anyhow, the majority of them were conducted based on single species, and/or single specimens.

85 In the present study, we performed a microstructural investigation of the Jurassic serpulid
86 tubes based on the representatives of the two main serpulid clades according to the newest
87 systematics (Kupriyanova, ten Hove and Rouse, 2023). We provide paleoecological implications

88 of different microstructures and consequently biomineralization system and highlight some
89 phylogenetic signals and evolutionary patterns of distinct microstructures.

90

91 **Material and its provenance**

92 The examined fossils consist of 47 specimens of serpulids representing 12 taxa (table 1).
93 The studied material has been selected from an ample collection (see Słowiński et al., 2020;
94 2022) concerning the well-preserved specimens, which allowed for a reliable ultrastructural
95 investigation. All taphonomically altered tubes have been discarded from further studies. Almost
96 all specimens encrust a variety of invertebrate fossils, oncoids, and hiatus concretions, and have
97 been derived from various localities with Middle and Upper Jurassic (upper Bajocian – lower
98 Kimmeridgian) deposits representing an array of paleoenvironments. The investigated material is
99 stored at the Institute of Earth Sciences in Sosnowiec, abbreviated GIUS 8-3589 (Callovian of
100 Zalas), GIUS 8-3730 (Gnaszyn Dolny), GIUS 8-3745 (Bołęcín), GIUS 8-3746 (Oxfordian of
101 Zalas), GIUS 8-3747 (Małogoszcz), GIUS 8-3750 (Ogrodzieniec-Świertowiec), GIUS 8-3751
102 (Krzyworzeka and Żarki). In ascending stratigraphical order, the localities are as follows:

103 Ogrodzieniec-Świertowiec: serpulids from this outcrop heavily encrust oncoids of
104 cyanobacterial genesis (Zatoń et al., 2012). The oncoids have been derived from condensed,
105 sandy, and carbonate layers, being overlain by clay sediments. This and the subsequent three
106 localities are situated within the area of the Polish Jura (Fig. 1). The deposits occur in the form of
107 dark siliciclastic sediments belonging to the Częstochowa Ore-bearing Clay Formation (e.g.,
108 Majewski, 2000; Matyja and Wierzbowski, 2000; Zatoń et al., 2011; Leonowicz, 2015). The late
109 Bajocian–early Bathonian age of the oncoid-bearing deposits was based on ammonites (Zatoń
110 and Taylor, 2009a; Zatoń et al., 2012), dinoflagellate cysts, and foraminifers (Słowiński, 2019).
111 Due to the origin of oncoids, the paleoenvironment is interpreted to be shallow marine, located
112 within the photic zone, and slightly below a fair-weather wave base (Zatoń et al., 2012).

113 Gnaszyn Dolny: serpulid tubes derived from this locality in majority encrust belemnite
114 rostra and oyster shells. The fossils come from middle Bathonian mudstones originating in
115 marine environment, below the storm wave base (Gedl and Kaim, 2012; Gedl et al., 2012).

116 Krzyworzeka: polychaetes from this site encrust carbonate hiatus concretions. The
117 concretion-bearing sediments from this locality are confined to the upper Bathonian (Poulsen,
118 1998). The deposits from Krzyworzeka belong to the northern sedimentary region of the Polish

119 Jura and originated in a calm outer shelf environment (Zatoń et al., 2011; Sadlok and Zatoń,
120 2020), located below the storm wave base (Różycki, 1953; Leonowicz, 2013).

121 Żarki: the deposits from which the serpulids encrusting hiatus concretions have been
122 derived are confined to the northern sedimentary region of the Polish Jura, and are represented by
123 the upper Bathonian mudstones (Zatoń and Taylor, 2009b). Sediments within this location
124 originated in a relatively calm but interrupted by episodic storm events (Leonowicz, 2015),
125 marine paleoenvironment (Marynowski et al., 2007; Leonowicz, 2012; 2013).

126 Bołecin: the outcrop is situated approximately 6 km east of Chrzanów. The serpulid tubes
127 have been found within sandy limestones, presumably referring to the so-called “Balin Oolite.”
128 Serpulid tubes have been found fixed to a variety of fossils including bivalves, gastropods, and
129 ammonites, as well as belemnite rostra. These condensed sediments (see Tarkowski et al., 1994;
130 Mangold et al., 1996) deposited presumably in an open marine paleoenvironment and are late
131 Bathonian-early Callovian in age (Mangold et al., 1996).

132 Zalas: the fossil material collected from this locality comes from two sites: 1) an active
133 quarry with sandy limestones forming a hardground of middle-earliest late Callovian age
134 (Giżejewska and Wieczorek, 1977; Dembicz and Praszkie, 2007); here, the serpulids encrust a
135 wide variety of invertebrate fossils, notably large *Ctenostreon proboscideum* (J. Sowerby and
136 G.B. Sowerby, 1820) bivalves. Hardground deposits originated in a deeper open marine setting
137 corresponding to a shelf environment (Dembicz and Praszkie, 2007; Zatoń, Wilson and Zavar,
138 2011); 2) overlying, lower Oxfordian light limestones hosting sponge bioherms (Matyja, 2006;
139 Matyszkiewicz, Kochman and Duś, 2012), to which serpulid tubes were attached.

140 Małogoszcz: serpulid fauna has been derived from an active quarry located in close
141 proximity to the town of Małogoszcz. This site is located in the southwestern part of the
142 Mesozoic margin of the Holy Cross Mountains. The tubes are attached to bivalve shells,
143 particularly the oyster *Actinostreon gregareum* (J. Sowerby, 1815). Lower Kimmeridgian
144 fossiliferous deposits are referred to as the Skorków Lumachelle (Kutek, 1994; Machalski, 1998;
145 Zatoń and Machalski, 2013). These shell beds were deposited during episodic storms in a
146 relatively shallow marine paleoenvironment (Machalski, 1998).

147

148 **Methods**

149 Before the preparation of specimens for the SEM examination, they were coated with
150 ammonium chloride and photographed using the Canon EOS camera. All previously selected
151 samples were cut longitudinally, polished, and etched with a 5% solution of acetic acid for one
152 minute prior to the SEM study. Part of these tube portions were oriented and mounted in epoxy
153 resin before polishing. Subsequently, the serpulid tube microstructure investigations were
154 performed on a SEM Zeiss EVO MA15 under a back-scattered electron detector (BSE) in a low
155 vacuum regime (Department of Geology, University of Tartu, Estonia). The sections were also
156 inspected under an environmental scanning electron microscope (ESEM) Philips XL30, and a
157 Quanta 250 scanning electron microscope using BSE imaging (both at the Institute of Earth
158 Sciences in Sosnowiec). The beam voltage was operated at 20 kV.

159 **Results**

160 During the examination of 12 taxa within eight genera representing two major clades,
161 three kinds of ultrastructure were distinguished: irregularly oriented prismatic structure (IOP),
162 spherulitic prismatic structure (SPHP), and simple prismatic structure (SP) (Table 1).

163 ***Filograninae* (BI)**

164 Members of the clade *Filograninae*, formerly referred to as BI, are represented in the
165 herein material by two genera: *Metavermia* and *Filogranula*, the latter one consisting of two
166 species: *F. runcinata* (J. de C. Sowerby, 1829) and *F. spongiophila* (J. Słowiński, O. Vinn, M.
167 Jäger et al., 2022).

168 The tube wall of *Metavermia* cf. *striatissima* (F.T. Fürsich, T.J. Palmer, and K.L.
169 Goodyear, 1994) is single-layered, and composed of an irregularly oriented prismatic structure
170 (IOP) (Fig. 2). Minute (maximally a few μm), needle-like crystals are deployed more or less
171 evenly within the entire tube wall. Their crystallization axes are longitudinal and lack a uniform
172 orientation.

173 The tube of *Filogranula runcinata* is presumably single-layered and is composed of a
174 spherulitic, regularly oriented prismatic microstructure (SPHP) (Fig. 3). It is formed by crystals
175 of prismatic shape exhibiting a slightly spherulitic arrangement. Some internal parts of the tube
176 are built of irregularly oriented, tiny, elongated crystals indicating an irregularly oriented
177 prismatic structure (IOP). However, the boundary between the two putative layers is transitional
178 and the latter microstructure occurs rather like inclusions in certain areas of the tube wall.
179 Growth lines are apparent across almost the entire tube. Some minor, external parts of the section

180 may appear like fine homogeneous granular microstructure; however, more likely it corresponds
181 to the differences in preservation between the external and internal parts of the tube due to the
182 diagenetic alteration of irregularly oriented prismatic microstructure.

183 The tube of *Filogranula spongiophila* consists of a single layer that is composed of a
184 simple, regularly oriented prismatic structure (SP) (Fig. 4). Growth increments are visible across
185 almost the whole tube (Fig. 4C-E).

186

187

188 ***Serpulinae, tribe Serpulini (AI)***

189 This tribe is represented in the investigated material by one genus comprising two
190 species: *Cementula spiroloinites* (Münster in Goldfuss, 1831) and *Cementula radwanskae* (J.
191 Słowiński, O. Vinn, M. Jäger et al., 2022).

192 The tube wall of *C. spiroloinites* is single-layered and consists of a simple prismatic
193 structure (SP) (Fig. 5). This microstructure is formed by parallel prismatic crystals oriented
194 perpendicularly to each growth line showing incremental zonation. The crystals are arranged
195 perpendicularly or obliquely to the tube wall.

196 The tube microstructure of the investigated specimens of *C. radwanskae* is altered in
197 most places. However, some places exhibiting faint growth increments indicate possibly a simple
198 (SP) or spherulitic prismatic structure (SPHP) constituting a single layer (Fig. 6). Certain areas
199 with tiny, elongated irregularly oriented crystals presumably are an effect of a diagenetic
200 distortion of the tube wall.

201 ***Serpulinae, tribe Ficopomatini (AII)***

202 The tribe Ficopomatini, formerly referred to as AII, includes here 5 genera with 7 species.
203 *Propomatoceros lumbricalis* (E.F. Schlotheim, 1820) possesses a tube wall consisting of two or
204 three layers (Fig. 7). The majority of the examined specimens have two-layered tubes. The
205 external part is formed by a spherulitic prismatic structure (SPHP), which consists of parallel
206 crystals of a slightly spherulitic arrangement. The inner part has an irregularly oriented prismatic
207 microstructure (IOP) comprising elongated, but relatively short, tiny crystals with inconsistent
208 orientation axes. In some areas of the tubes' wall, the IOP structure is diagenetically altered
209 superficially resembling a fine homogeneous granular structure. The putative three-layered tube
210 wall (Fig. 7D) is built of the external and internal (uncertain) parts composed of a spherulitic

211 prismatic structure (SPHP). The middle part shows a fine homogeneous granular microstructure,
212 which may be either a primary microstructure or similarly to other specimens, an obliterated IOP
213 structure. In contrast to two-layered *P. lumbricalis* tubes, no areas exhibiting irregularly oriented
214 prismatic structures have been found. As such, it is not evident, whether these microstructures
215 are primary or have undergone diagenetic alteration. The external parts exhibit incremental
216 zonation with prominent chevron-shaped growth lines. The crystals are continuously and
217 regularly positioned through successive growth increments.

218 In the herein material *Nogrobs* is represented by three species: *N. aff. quadrilatera* (A.
219 Goldfuss, 1831), *N. aff. tetragona* (J. de C. Sowerby, 1829), and *N? aff. tricristata* (A. Goldfuss,
220 1831), which, however, share all the major microstructural characters. The tube walls of all the
221 species have two layers, which are separated from each other with a sharp boundary. The
222 external layer is composed of a simple prismatic structure (SP), whereas the internal layer is
223 composed of very thin, short, needle-like crystals, which make up an irregularly oriented
224 prismatic structure (IOP) (Fig. 8).

225 *Mucroserpula* sp. (H. Regenhardt, 1961) possesses a two-layered tube wall (Fig. 9). The
226 internal layer is composed of an irregularly oriented prismatic structure (IOP) consisting of
227 bunches of densely packed, short but elongated crystals lacking uniform orientation axes. The
228 external layer is composed of a regular spherulitic prismatic structure (SPHP), which constitutes
229 the major part of the tube wall. Within this layer, crystals are oriented perpendicularly with
230 respect to each incremental zone and have a somewhat prismatic arrangement. The chevron-
231 shaped growth lines are very well-visible alongside almost the whole tube length. The border
232 between the two layers is somewhat transitional.

233 The tube wall of *Placostegus planorbiformis* (Münster in Goldfuss, 1831) is single-
234 layered, and composed of a simple prismatic structure (SP) (Fig. 10). All crystals are more or
235 less parallel within each incremental zone. Within the outermost part of the tube, the crystals are
236 oriented perpendicularly to the tube wall.

237 The specimens of undetermined serpulid, Serpulidae sp. possess a two-layered tube wall
238 (Fig. 11). The external part features a regularly oriented simple prismatic structure (SP). The
239 internal, thinner part, separated from the external with a sharp boundary is formed by an
240 irregularly oriented prismatic structure (IOP) composed of densely packed, minute, elongated
241 crystals.

242

243 **Discussion**244 ***Tube ultrastructure evolution and its phylogenetic constraints***

245 Three distinct kinds of ultrastructure have been identified within 12 taxa corresponding to
246 the two of three main clades of serpulids – Filograninae and Serpulinae (Kupriyanova, ten Hove
247 and Rouse, 2023). These microstructures comprise irregularly oriented prismatic structure (IOP),
248 spherulitic prismatic structure (SPHP), and simple prismatic structure (SP) (see Table 1), which
249 are among the most prevalent microstructure kinds in fossil serpulids (Vinn et al., 2008; Vinn,
250 2020). Six of these taxa have single-layered tubes and six are two-layered, of which one taxon
251 perhaps may possess either two or three ultrastructural layers (see discussion). The majority of
252 both Cenozoic and contemporary serpulids are single-layered, only about one-third of serpulid
253 species have at least two or up to four distinct ultrastructural layers (Vinn et al., 2008). During
254 the Jurassic, the percentage of multi-layered serpulid species was lower, constituting
255 approximately 25% (Vinn and Furrer, 2008). It may have resulted from the evolutionary pattern
256 where more complex tube walls with at least two layers were more common from the beginning
257 of the Cenozoic onward – the growing complexity of microstructures combined with relatively
258 fast biomineralization likely enhanced the strengthening abilities of the tubes. It can also be
259 explained by the vagaries of the fossil record resulting in incomplete preservation of the
260 outermost layers, which might be similar to the case of calcareous sabellids (Vinn, ten Hove and
261 Mutvei, 2008; Słowiński, Banasik and Vinn, 2023).

262 Sanfilippo (1998) presented a concept to utilize the ultrastructural diversity of serpulids to
263 assess their generic determination. Further studies revealed that microstructure characters may be
264 species-dependent (Vinn, 2007; Vinn et al., 2008; Kupriyanova and Ippolitov, 2015), which
265 limits the application of tube microstructures in deciphering the generic affiliation of serpulids.
266 Regardless, ultrastructural fabrics of tubes may still be used to approach the relationships
267 between distinct kinds of structures throughout the evolution of the main serpulid clades (e.g.,
268 Vinn et al., 2008; Vinn and Kupriyanova, 2011; Vinn, 2013; Ippolitov and Rzhavsky, 2014;
269 2015a; b).

270 The most recent phylogenetic analyses indicate that serpulids are split into three major
271 clades – Filograninae, Spirorbinae, and Serpulinae, the last of which is further subdivided into
272 two tribes: Serpulini and Ficopomatini (Kupriyanova, ten Hove and Rouse, 2023). The family

273 Serpulidae until recently was maintained to comprise two major clades referred to as B and A,
274 separated into BI, BII, AI, and AII (Kupriyanova, Macdonald and Rouse, 2006, Kupriyanova et
275 al., 2009; Kupriyanova and Nishi, 2010). Formerly recognized clades BI and BII generally may
276 be related to Filograninae and Spirorbinae, whereas AI and AII, both settled within Serpulinae,
277 correspond to the tribes Serpulini and Ficopomatini, respectively. Apart from Spirorbinae
278 (former BII), members of all clades are present in the material studied.

279 Clade Filograninae is possibly rooted even in the Permian with primitive tubes such as
280 *Filograna* (Sanfilippo et al., 2017; Ramsdale, 2021), and abundantly represented in the fossil
281 record during the Mesozoic by many strongly ornamented tubes with several keels, e.g.,
282 *Vermiliopsis*, *Metavermlia*, and *Filogranula*, the latter two of which are present in the material
283 investigated. *Metavermlia* cf. *striatissima* is single-layered and possesses irregularly oriented
284 prismatic microstructure (IOP), which has been also found in the recent *Metavermlia*
285 *multicristata* (Vinn et al., 2008). It is characteristic of this clade and is also the most common
286 microstructure in the recent serpulids encompassing ca 60% of species (Vinn, 2007). IOP
287 microstructure most often builds the wall of single-layered tubes, or alternatively the inner part
288 of multi-layered tubes (see Vinn et al., 2008).

289 The representatives of Filograninae are generally characterized in the fossil record by
290 more or less uniform fabrics of tube microstructures forming irregularly oriented structures,
291 notably irregularly oriented prismatic structure (IOP). Secretion of such microstructures seems to
292 be governed by the lower biological control of an animal, compared to more advanced
293 microstructures. The first confirmed appearance of such ultrastructure dates back to the Early
294 Jurassic, possibly being present already in the Triassic (Vinn, Jäger and Kirsimäe, 2008). Such a
295 simpler way of tube formation may indicate its plesiomorphic character and earlier origin.

296 Another representative of this clade in the investigated material is the genus *Filogranula*
297 comprising the recently discovered new species *F. spongiophila* (Słowiński, Vinn, Jäger et al.,
298 2022), which has been studied with respect to ultrastructure for the first time, and by *F.*
299 *runcinata*. The tube wall of *F. spongiophila* is single-layered and has a simple prismatic structure
300 (SP), whereas *F. runcinata* possesses one well-preserved layer composed of spherulitic prismatic
301 structure (SPHP) and a dubious IOP layer consisting of somewhat distorted, occasionally
302 occurring areas of tiny, irregularly oriented crystals. As a result, the tube wall is interpreted as
303 single-layered, composed of SPHP structure. Such microstructure occurs commonly in the clade

304 Serpulinae comprising both formerly established “A” clades. The systematic position of the
305 fossil *Filogranula* seems to be complicated, as the Recent *Filogranula* is considered to be settled
306 within Filograninae, relatively closely related to *Vermiliopsis* and *Metavermilia* (Kupriyanova,
307 ten Hove and Rouse, 2023). While the Cretaceous *Filogranula cincta* (A. Goldfuss, 1831) is
308 classified as belonging to Filograninae too, the Jurassic *Filogranula runcinata* was considered to
309 be placed in Serpulinae, within the tribe Ficopomatini (see Ippolitov et al., 2014; Kočí and Jäger,
310 2015), due to its prismatic microstructure, characteristic of the serpulid tubes appearing
311 transparent in the Recent species, such as e.g., *Placostegus*. On the other hand, prismatic
312 structures are not a fully satisfactory character in differentiating between former A and B clades.
313 Ippolitov and Rzhavsky (2014; 2015a; b) found out that some spirorbins may also have their
314 tubes composed of regularly oriented prismatic microstructures, in spite of the fact, that
315 Spirorbinae is phylogenetically closely related to Filograninae, which commonly form
316 irregularly oriented structures.

317 Serpulinae constitutes two well-supported tribes – Serpulini and Ficopomatini
318 (Kupriyanova, ten Hove and Rouse, 2023). The first unequivocal members of Serpulinae very
319 likely existed since the Triassic (Assmann, 1937) if not even since the Permian (Sanfilippo et al.,
320 2017; 2018). In contrast to Filograninae, which have mostly non-oriented structures, serpulids
321 within this lineage have a more advanced biomineralization system and are capable of forming
322 complex microstructures, such as e.g., oriented, lamello-fibrillar, or oriented-fibrillar (see Vinn
323 et al., 2008; Vinn and Kupriyanova, 2011). During the Jurassic, serpulid microstructure diversity
324 was relatively modest (ten kinds; see Vinn, 2020) compared to an array of contemporary and
325 Cenozoic ones (Vinn, 2007). The most sophisticated microstructures in serpulids evolved in the
326 Eocene (e.g., Vinn, 2008; Buckman, 2020). Although such complex microstructures as lamello-
327 fibrillar or regularly ridged prismatic appeared quite late in the evolution of serpulids, the
328 oriented prismatic structures evolved at the latest during the Middle Jurassic. Simple and
329 spherulitic prismatic microstructures are very common and characteristic of the Jurassic
330 Serpulinae and are considered to be apomorphic. Oriented prismatic structures are unknown
331 from any genus of Filograninae; however, they were found to persist in the Cretaceous clade
332 Spirorbinae (Ippolitov and Rzhavsky, 2015a; b), which may mean, that either it is plesiomorphic,
333 or the prismatic structures have evolved at least twice.

334 The tribe Serpulini is represented in the Jurassic by characteristically coiled *Spiraserpula*
335 and related *Cementula*, which is present in the herein material. *C. spiroloinites* and *C. radwanskae*
336 are both single-layered and possess their tube walls built of simple prismatic microstructure (SP),
337 though in the latter it is somewhat obliterated. Simple and spherulitic prismatic microstructures
338 exhibit the predominantly uniform orientation of calcium carbonate crystals which determines
339 the optical transparency of the tubes (Zibrowius and ten Hove, 1987), supported also by the
340 dense arrangement of crystals and their large size (Vinn and Kupriyanova, 2011). In contrast, the
341 recent tubes with IOP microstructure most often are optically opaque (Vinn et al., 2008).

342 All the main members of the Jurassic representatives of Ficopomatini are present in the
343 investigated material. They contain genera with various morphotypes comprising a robust,
344 single-keeled *Propomatoceros*, a three-keeled *Mucroserpula*, quadrangular tubes attributed most
345 often to *Nogrobs*, and a well-defined, planispirally coiled *Placostegus*. Serpulids in this tribe
346 commonly possess two layers of tube walls having the outer dense layer built of ordered
347 prismatic structures (simple or spherulitic), and the internal layer composed of irregularly
348 oriented prismatic structure. Two-layered tubes appeared in the Jurassic (Vinn and Furrer, 2008)
349 and have their external layer denser than the internal, which is composed of thinner mineral
350 microstructure. Dense outer protective layers (DOL) have been found in tube walls of serpulids
351 inhabiting diverse environments (see Vinn and Kupriyanova, 2011). Consequently, rather than
352 being dependent on the environment, the advent of DOLs during the Jurassic appears to be a
353 significant evolutionary adaptation of serpulids. Except for one species in the clade Filograninae,
354 dense, outer layers exist exclusively in the clade Serpulinae (Vinn and Kupriyanova, 2011).

355 *Propomatoceros lumbricalis* and *Mucroserpula* possess here two well-distinguishable
356 layers: internal IOP and external SPHP. A single *Propomatoceros* specimen may have three
357 layers, which, however, is ambiguous, as its internal layer may be diagenetically obliterated, and
358 therefore all the specimens are considered two-layered. As discussed previously (Ippolitov et al.,
359 2014; Słowiński et al., 2022), it cannot be ruled out, that at least in some populations,
360 *Mucroserpula* and *Propomatoceros* may represent the same genus. The diagnostic characters of
361 both genera are highly transitional, subjective, and dependent to a large extent on different
362 variables, such as ontogenetic stage or diagenesis (see Słowiński et al., 2022). Nonetheless, the
363 microstructural fabrics of the two genera in our investigation are essentially the same and typical
364 of Ficopomatini.

365 *Nogrobs* consists here of three species: *N. aff. quadrilatera*, *N? aff. tricristata*, and *N. aff.*
366 *tetragona*. All of them are two-layered composed of internal IOP and external SP microstructure.
367 Apart from *Nogrobs*, Jurassic quadrangular fossils of serpulids that share morphological
368 similarities with this genus were attributed by various authors to different, partially uncertain
369 genera - *Tetraserpula*, *Tetraditrupa*, *Glandifera*, *Tubulostium*, *Tectorotularia*, and *Ditrupula*.
370 Kupriyanova and Ippolitov (2015) examined and reviewed a number of extant and fossil taxa
371 having tusk-shaped, tetragonal in cross-section tubes, concluding, that these morphologically
372 similar forms belong to several different genera, most likely being an effect of convergence. Not
373 surprisingly, these recent taxa cannot be synonymized with the fossil *Nogrobs*. Nevertheless, the
374 authors claim that at least the majority of fossil *Nogrobs* species may be members of a single
375 clade, as opposed to morphologically-related recent species. According to its external tube layer
376 composed of simple prismatic microstructure (responding to a transparent tube), the three
377 investigated species fit well Ficopomatini, confirming previous ultrastructural studies
378 (Kupriyanova and Ippolitov, 2012). It has to be noted, however, that the recent *Nogrobs*
379 *grimaldii* (P. Fauvel, 1909) has been found to possess an opaque tube (Kupriyanova and Nishi,
380 2011).

381 The only exception not having a two-layered tube wall within this tribe is *Placostegus*,
382 represented here by *P. planorbiformis*, which has a single layer made entirely of a simple
383 prismatic structure (SP). Similarly, contemporary *P. tridentatus* (J.C. Fabricius, 1779) possesses
384 a tube wall composed of a simple prismatic microstructure resulting in a completely transparent
385 tube (ten Hove and Kupriyanova, 2009: 8, fig. 1F; Vinn and Kupriyanova, 2011).

386 The undetermined serpulid taxon Serpulidae sp., described recently by Słowiński et al.
387 (2022) has been studied here with respect to its microstructure. Although the investigated
388 specimens externally resemble a few genera, such as *Propomatoceros*, *Placostegus*, and
389 *Metavermlia*, they do not exactly fit any of those taxa (see Słowiński et al., 2022). The
390 specimens possess a tube wall composed of two layers – external simple prismatic (SP), and
391 internal irregularly oriented prismatic microstructure (IOP), which is of a very close resemblance
392 to other members of Ficopomatini, notably *Nogrobs*.

393

394 ***Ecological implications and comparisons with sabellids***

395 Serpulids perform biologically controlled biomineralization where their cellular activity
396 regulates the nucleation and extracellular growth of the calcium carbonate crystals by the ion
397 uptake from the surrounding water (see Neff, 1971a; b) using a secretory epithelium, mediated
398 and controlled by the organic matrix (Vinn, Kirsimäe and ten Hove, 2009; Vinn, 2021a). As a
399 result, serpulids accomplish a specified crystal orientation within the tube wall, which is
400 reflected by their variety of ultrastructural fabrics - the growth direction of crystals may be
401 anisotropic, semi-oriented, or oriented (Weedon, 1994; Vinn et al., 2008). Apart from a matrix-
402 mediated crystallization (Vinn, 2021a), an alternative explanation has been proposed recently to
403 define the crystal orientation mechanism, which is based on a variable application of the
404 serpulid's collar rotational force between the formation of ordered and unordered microstructures
405 (Buckman and Harries, 2020). Considering the fact that the same growth increments may occur
406 across zones with diverse ultrastructures makes (Vinn, 2021a; b) this model strongly disputable.

407 The serpulid biomineralization system differs from that of other tube-dwelling
408 polychaetes. By the formation of the cylindrical and **parable** layers (see Jäger, 1983) serpulids
409 are capable of forming multi-layered tubes arranged in distinct microstructures (see Vinn et al.,
410 2008). Importantly, the parable layer is formed by adding secretory increments to the rim of the
411 worm's aperture allowing it to actively modify the external morphology according to the tube's
412 sculpture, but also depending on the temporary ecological requirements. Such a solution results
413 in a wide array of serpulid tube characters including attachment structures and base widenings of
414 the tube, which combined with the ability to form several layers greatly improves the durability
415 of the tube. Fossil cirratulids (Vinn, 2009; Taylor et al., 2010) and sabellids (Vinn, ten Hove and
416 Mutvei, 2008; Słowiński, Banasik and Vinn, 2023) possess single-layered tube wall composed of
417 a spherulitic prismatic structure. The presence of only cylindrical layer in sabellids strongly
418 impairs their biomineralization abilities. Unlike serpulids having mostly chevron-shaped growth
419 lines (see Weedon, 1994), they form their tubes by secreting calcareous material along distinct
420 growth lamellae oriented parallel to the tube wall (Vinn, ten Hove and Mutvei, 2008; Słowiński
421 et al., 2022). Subsequent increments are added to the internal surface of the tube, and therefore
422 *Glomerula* is unable to modify its simple tube architecture. A much longer secretion zone in
423 sabellids compared to serpulids enables them for a fast calcification incurring comparably lower
424 physiological costs, which allows them to considerably prolong their tubes. On the other hand,
425 such fast dispersal may be required by the diminishing inside of the tube, insufficient for the

426 growing worm. Interestingly, cirratulids perform a double-phased, combined controlled, and
427 influenced biomineralization. The products of an influenced biomineralization such as
428 agglutinated xenolithic granules within a calcareous matrix, as well as the anisotropic orientation
429 of different-sized crystals, indicate, that cirratulids govern even a weaker than sabellids control
430 over biomineralization (Guido et al., 2024).

431 A much more advanced biomineralization system of serpulids and the resulting variety of
432 microstructures are an important evolutionary adaptation for this taxon, which is an obligatory
433 tube-dweller. The emergence of multi-layered tubes with dense, outer protective layers during
434 the Jurassic could have been triggered by the intensified predation during the Mesozoic Marine
435 Revolution (Vermeij, 1977). With the skeleton development, serpulids could perform
436 competitive strategies against other organisms instead of avoiding competition or temporary
437 paleoenvironmental vagaries. Gradual increase in abundance of serpulids during the Mesozoic
438 and Cenozoic (see Ippolitov et al., 2014 for a review), and their advantage over other organisms
439 allowed for higher plasticity and divergence of different morphotypes (Vinn et al., 2023 (in
440 press)). Different morphogenetic programs in turn enhanced their functional utilization not only
441 by mechanical strengthening of the tubes but also by optimizing their space by planispiral
442 coiling, providing a higher feeding tier, avoiding being overgrown or coated by sediment thanks
443 to upward growth. Free-living serpulids with tusk-shaped tubes were presumably adapted to live
444 in or on the surface of soft sediment during episodes of increased sedimentation rates (see Vinn
445 et al., 2023 for a review (in press)). Sabellid biomineralization system allowed them to quickly
446 elongate their tubes but these polychaetes did not exhibit such diverse morphotypes. Instead,
447 they could grow irregularly away from the initial point of encrustation and perform an
448 opportunistic, fugitive strategy (see Taylor, 2016; Słowiński, Banasik and Vinn, 2023).
449 Additionally, it could be the outcome of biomineralization's lack of significance in this group as
450 calcareous sabellids were restricted to a single genus in the family.

451

452

453

454 **Conclusions**

455 The first thorough assessment of the ultrastructural diversity of Middle and Late Jurassic
456 serpulid tubes has been conducted. The obtained data reveal a characteristic of Jurassic serpulids,

457 a relatively low diversity of ultrastructural fabrics, which generally correspond to certain clades.
458 Amongst 12 taxa representing two (Filograninae and Serpulinae) of the three main serpulid
459 clades, six of them possess tube walls composed of a single layer, and six are two-layered. There
460 are certain evolutionary trends in tube ultrastructures. The representatives of the clade
461 Filograninae are single-layered and have their tube walls built of a primitive, irregularly oriented
462 prismatic microstructure (IOP). The majority of members of possibly apomorphic clade
463 Serpulinae possess two-layered tube walls, where the denser, external layer is composed of
464 oriented, prismatic microstructures (either spherulitic (SPHP) or simple(SP)), and the internal is
465 irregularly oriented prismatic (IOP). The exceptions are *Placostegus planorbiformis* and the
466 genus *Cementula*, which are single-layered, and built of simple prismatic structure (SP).

467 Serpulid tube ultrastructures reflect their biomineralization abilities providing important
468 paleoecological signatures. Formation of the regularly oriented microstructures of Serpulinae
469 requires a higher biological control over biomineralization compared to the more primitive,
470 anisotropic microstructures of Filograninae. The development of serpulid ultrastructure diversity
471 was likely triggered by the evolutionary importance of the tubes for this group. The formation of
472 dense outer protective layers in serpulids was presumably induced by increased predation rates
473 during the Mesozoic Marine Revolution. A variety of microstructure kinds and the ability to
474 form multi-layered tubes allowed serpulids to employ different morphogenetic programs, which
475 had an impact on their functional utilization.

476 The differences in the biomineralization system between serpulids and other tube-
477 dwelling polychaetes resulted from the importance of tubes for the former taxon. The complex
478 biomineralization system of serpulids resulted in a multiplicity of forms and an ability to form
479 robust, strongly ornamented tubes, which mechanically strengthened their durability. Although
480 higher energy expenditure of skeletal secretion decreased the rate of tube formation, the solid
481 attachment and skeleton robustness allowed for a competitive advantage over other encrusters.
482 Conversely, sabellid primitive biomineralization abilities presumably were elicited by the
483 unimportance of skeleton for this taxon resulting in a simplicity of forms. On the other hand, it
484 enabled a fast spreading over substrate shunning competition and disadvantageous conditions by
485 utilizing an opportunistic strategy of quick tube elongation.

486

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490

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Figure 1

Figure 1: Geology of the investigated area.

(A) geological sketch-map of Poland without the Cenozoic cover showing three sampled sites; HCM-Holy Cross Mountains; PJ-Polish Jura; 1-Bolećin; 2-Zalas; 3-Małogoszcz. **(B)** geological map of the Polish Jura area without Quaternary cover, with sampled localities indicated (modified after Zatoń, Marynowski and Bzowska, 2006).

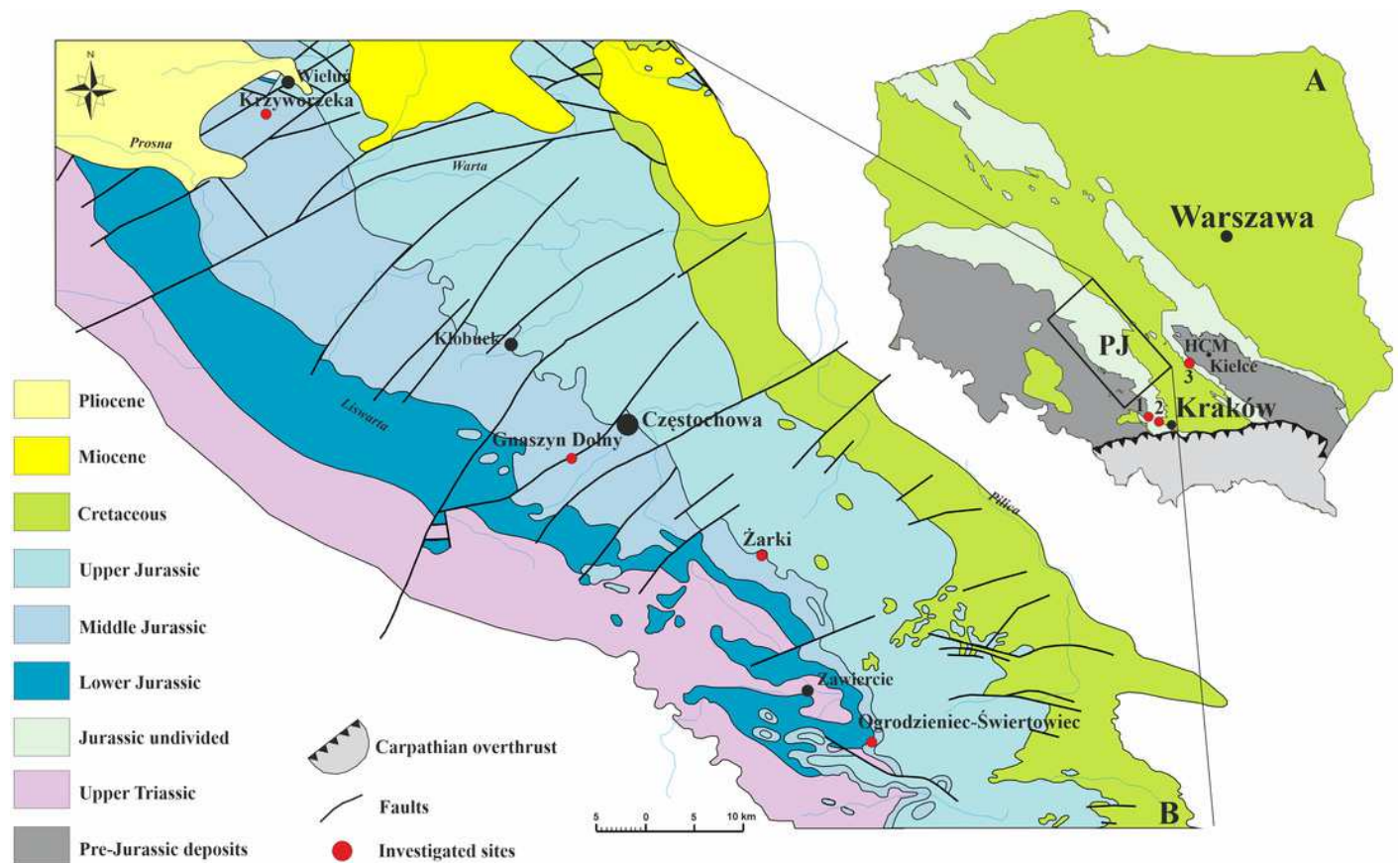


Figure 2

Figure 2: Selected specimens of *Metavermilia* cf. *striatissima* (F.T. Fürsich, T.J. Palmer, and K.L. Goodyear, 1994) from the Jurassic of Poland.

(A, B) *Metavermilia* cf. *striatissima* encrusting the interior of the boring *Gastrochaenolites* from the upper Bathonian of Krzyworzeka, Polish Jura (**A.** GIUS 8-3751/3; **B.** GIUS 8-3751/8). **(C, D)** Longitudinal sections of the tubes from the upper Bathonian of Krzyworzeka with visible irregularly oriented prismatic structure (IOP). **(C)** General look of the ground, polished and etched tube with visible thin, single-layered walls (GIUS 8-3751/3). **(D)** Close-up of the tube wall section showing densely packed, tiny, elongated prismatic crystals of irregular arrangement (GIUS 8-3751/3).

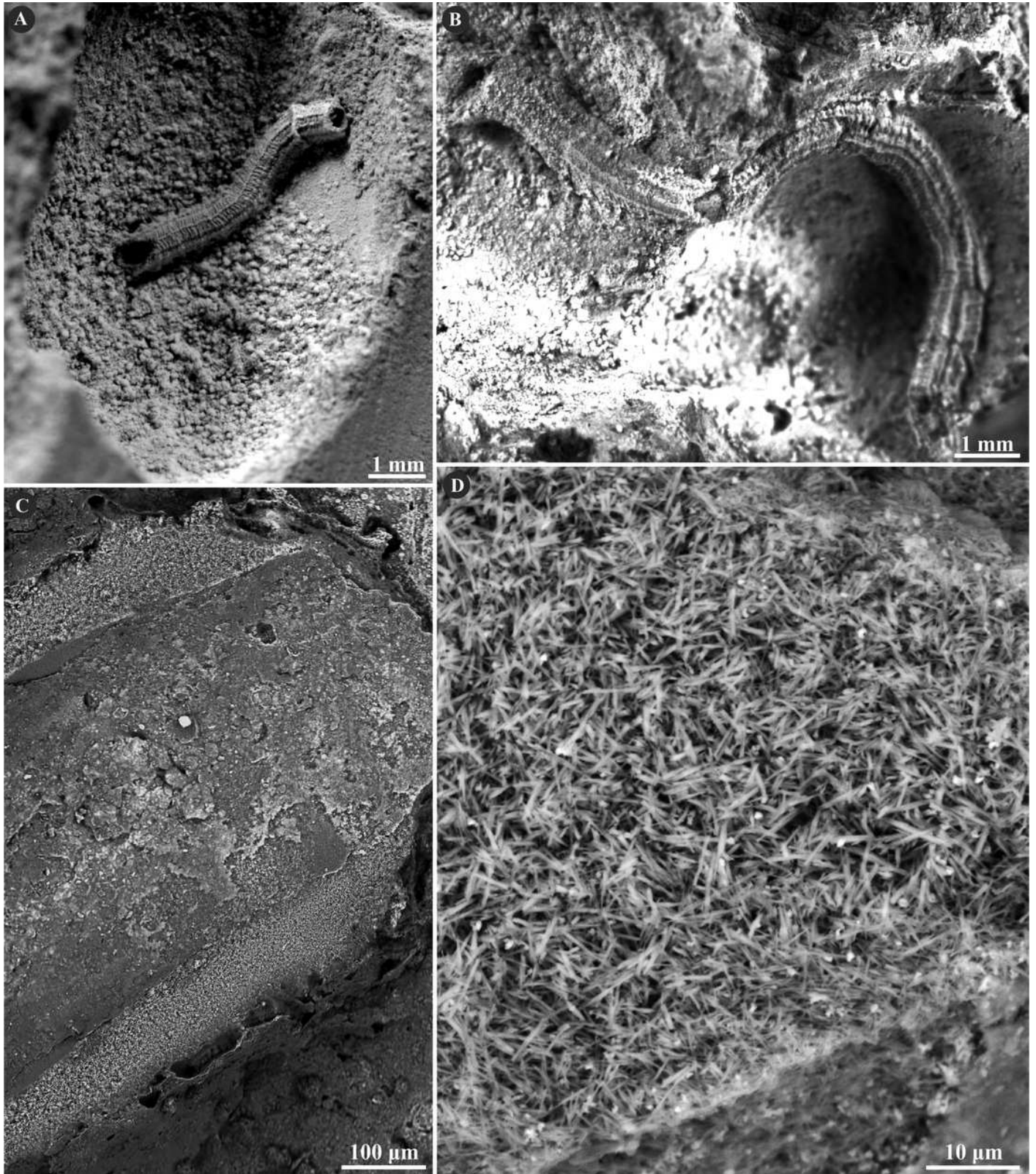


Figure 3

Figure 3: Selected specimens of *Filogranula runcinata* (J. de C. Sowerby, 1829) from the Jurassic of Poland.

(A, B) *Filogranula runcinata* from the middle Bathonian of Gnaszyn Dolny (**A.** GIUS 8-3730/10) and the Callovian of Zalas (**B.** GIUS 8-3589/6) encrusting shell fragments. **(C-F)** Longitudinal sections of the tubes showing a single-layered tube wall built of spherulitic prismatic structure (SPHP) with well-visible growth increments (GIUS 8-3730/10).

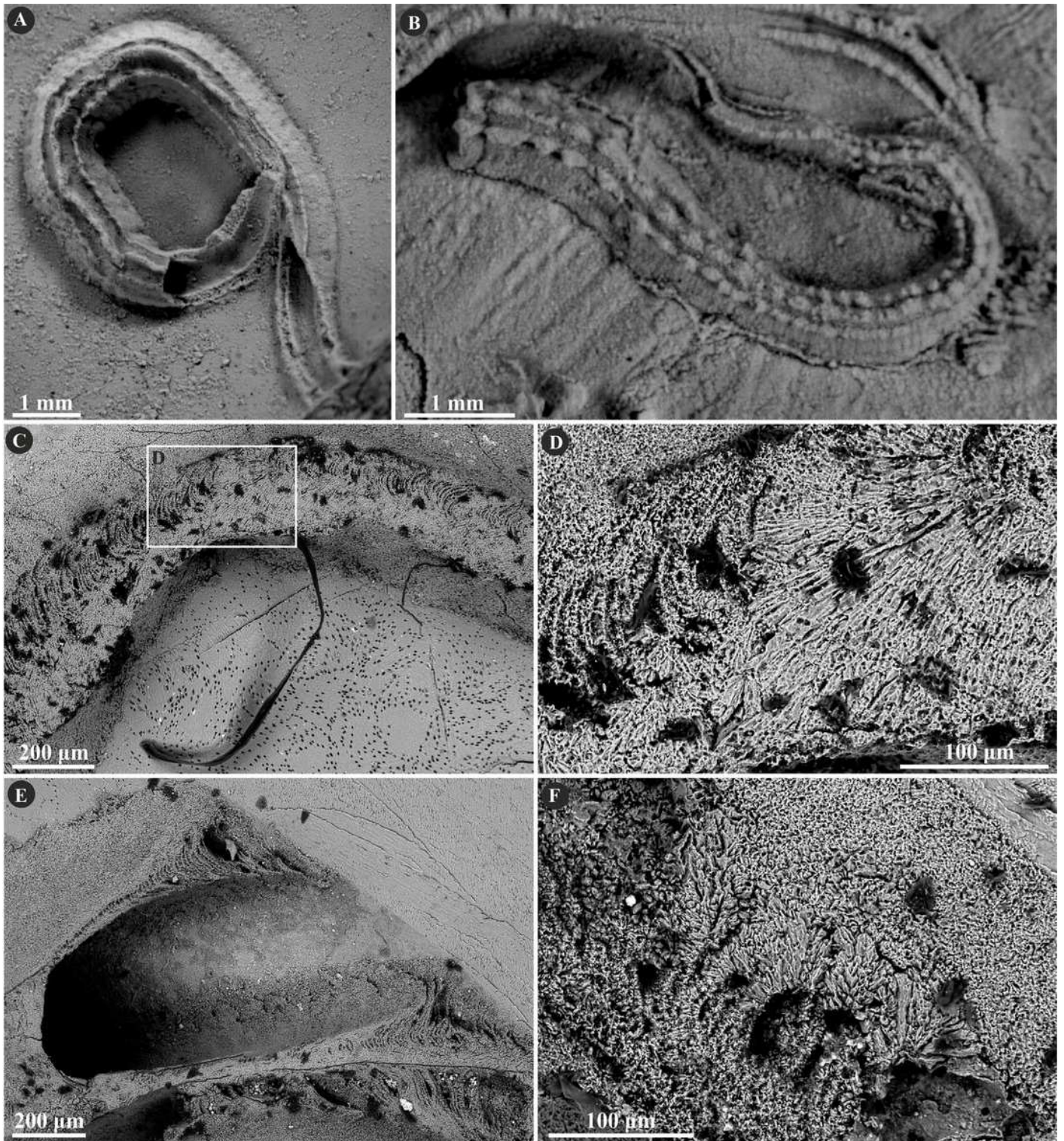


Figure 4

Figure 4: Selected specimens of *Filogranula spongiophila* (J. Słowiński, O. Vinn, M. Jäger, et al., 2022) from the Jurassic of Poland.

(A, B) *Filogranula spongiophila* from the Oxfordian of Zalas (**A.** GIUS 8-3746/2; **B.** GIUS 8-3746/3) encrusting sponge fragments. **(C-E)** Longitudinal sections of the tubes showing the simple prismatic structure (SP) with well-visible growth increments (GIUS 8-3746/9).

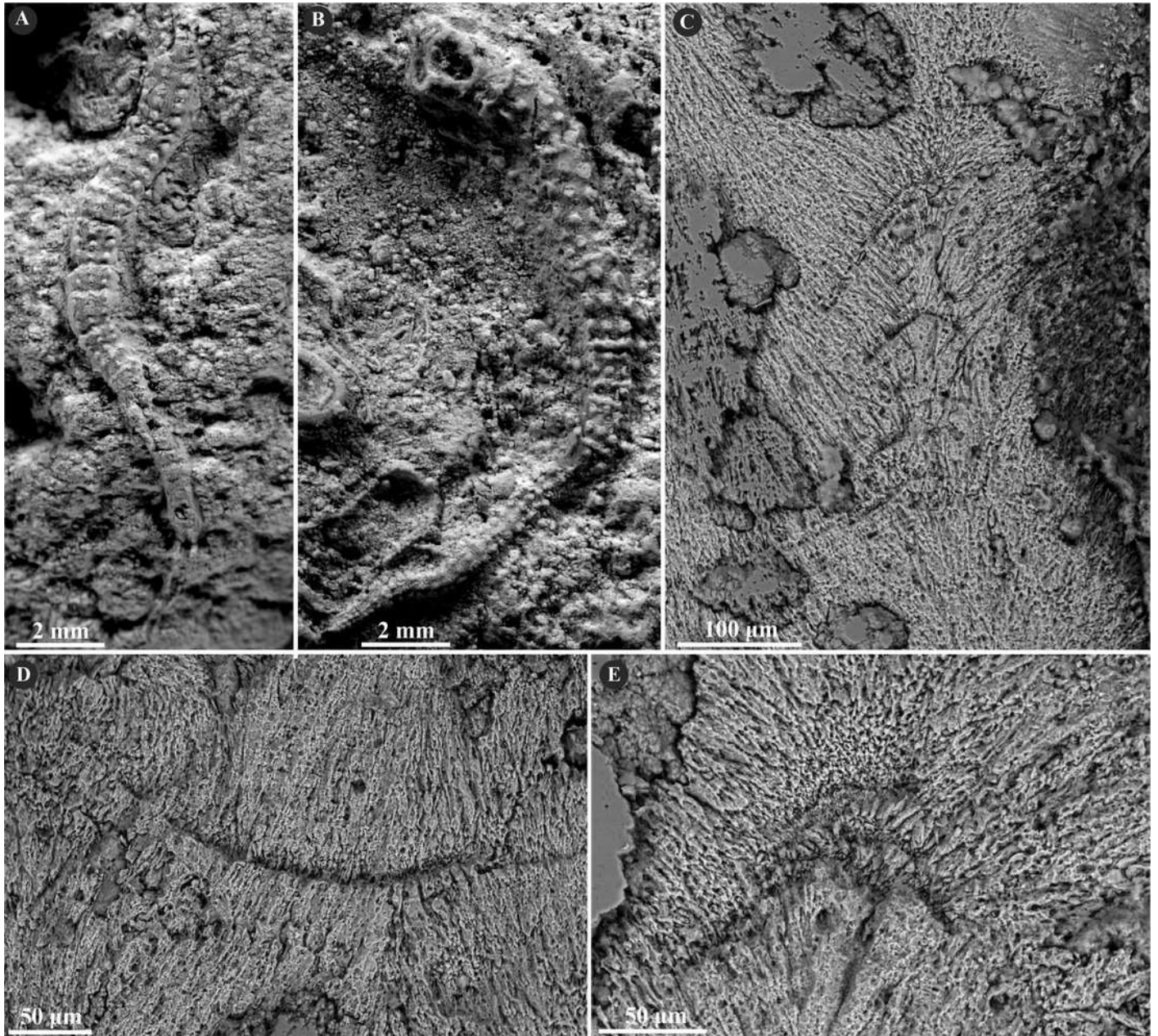


Figure 5

Figure 5: Selected specimens of *Cementula spirolinites* (Münster in Goldfuss, 1831) from the Jurassic of Poland.

(A) *Cementula spirolinites* from the Oxfordian of Zalas encrusting a sponge fragment (GIUS 8-3746/4). **(B-D)** Longitudinal sections of the tubes showing the simple prismatic structure (SP). **(B)** GIUS 8-3746/10; **(C, D)** GIUS 8-3746/11.

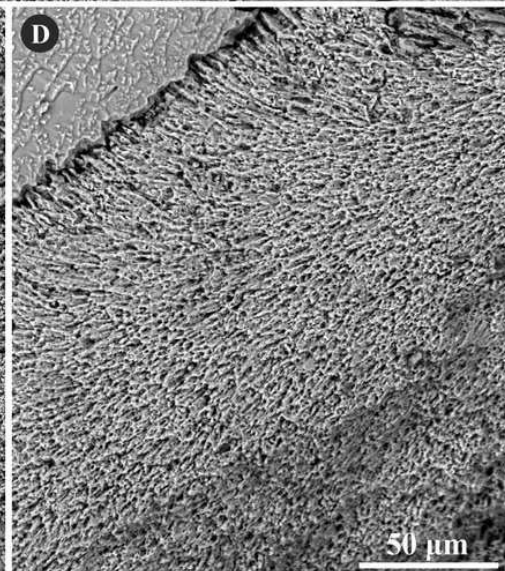
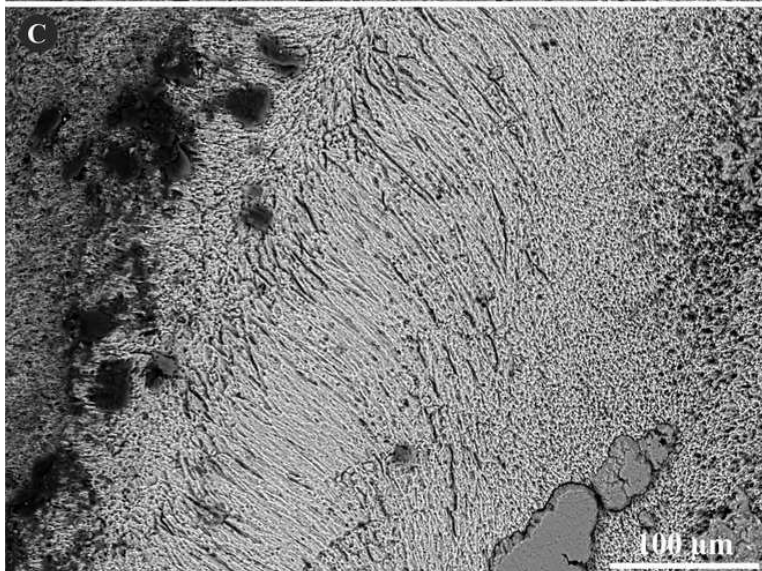
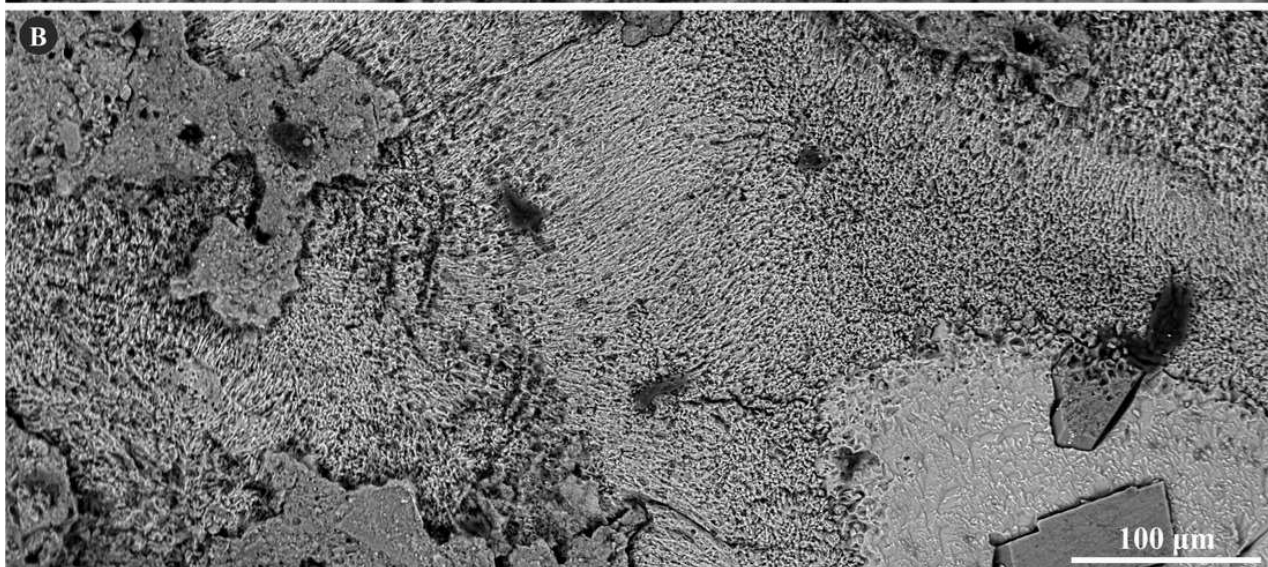


Figure 6

Figure 6: Selected specimens of *Cementula radwanskae* (J. Słowiński, O. Vinn, M. Jäger, et al., 2022) from the Jurassic of Poland.

(A, B) *Cementula radwanskae* from the Callovian of Zalas encrusting shell fragments (**A.** 8-3589/7; **B.** GIUS 8-3589/10). **(C-E)** Longitudinal sections of the tubes showing mostly altered, presumably simple prismatic microstructure (SP); GIUS 8-3589/18. Note the weak growth increments in **C**.

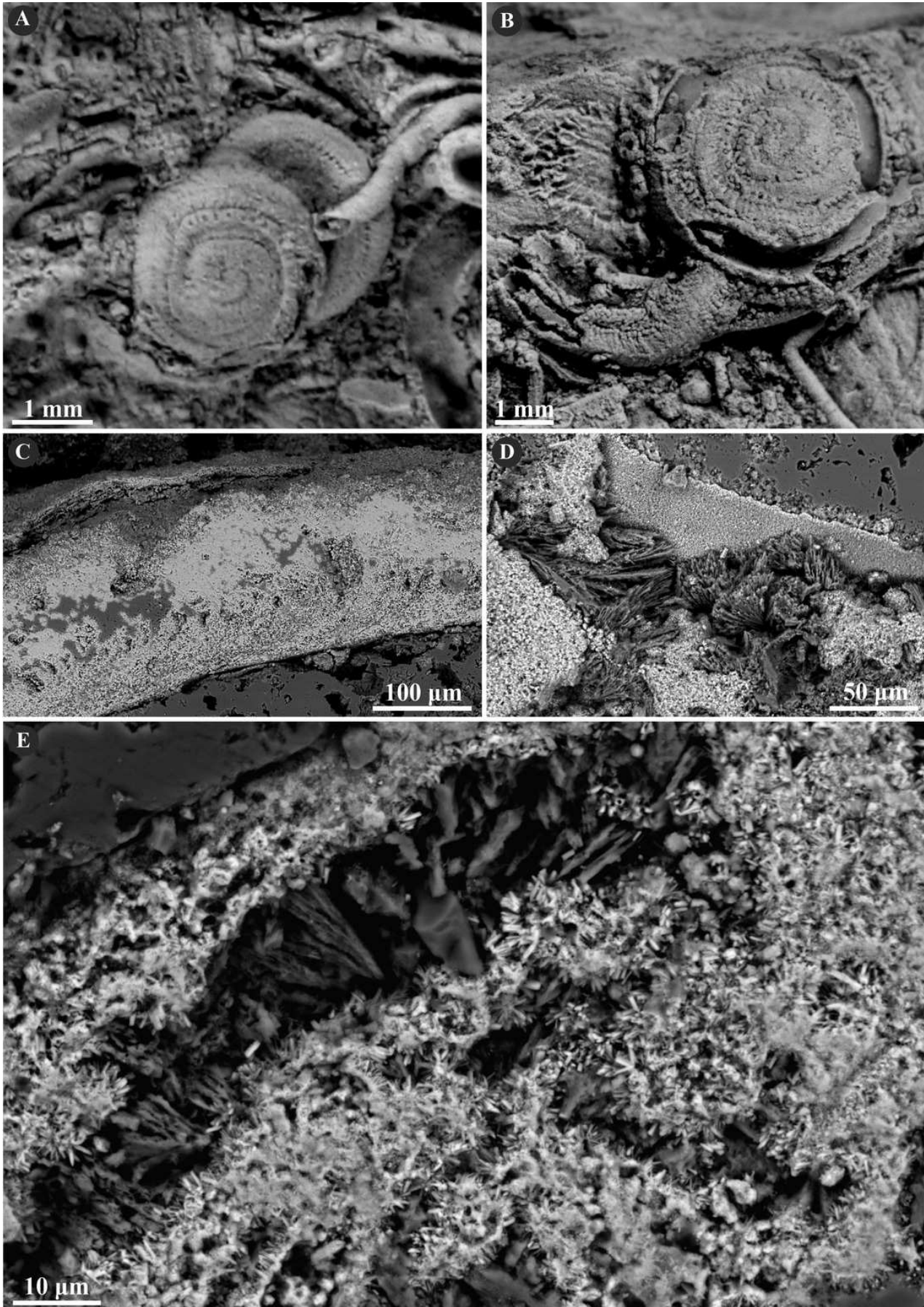


Figure 7

Figure 7: Selected specimens of *Propomatoceros lumbricalis* (E.F. Schlotheim, 1820) from the Jurassic of Poland.

(A, B) *Propomatoceros lumbricalis* encrusting a shell fragment from the Callovian of Zalas (**A.** GIUS 8-3589/11; **B.** GIUS 8-3589/12). **(C)** *Propomatoceros lumbricalis* encrusting an oyster shell from the middle Bathonian of Gnaszyn Dolny (GIUS 8-3730/12). **(D-J)** Sections of the tubes showing the ultrastructural details. **(D)** The longitudinal section of the *P. lumbricalis* tube from the upper Bathonian–lower Callovian of Bołęcín showing the putative three-layered tube (GIUS 8-3745/5). The external layer shows spherulitic prismatic structure (SPHP) with growth increments; the middle part exhibits presumably altered irregularly oriented prismatic structure (IOP) superficially resembling homogeneous structure; the internal layer shows uncertain spherulitic prismatic structure. **(E)** The longitudinal section of the two-layered *P. lumbricalis* tube from the middle Bathonian of Gnaszyn Dolny showing the external spherulitic prismatic structure (SPHP) with growth increments and the internal irregularly oriented prismatic structure (IOP) (GIUS 8-3730/30). **(F)** A close-up showing the internal layer built of irregularly oriented prismatic structure (IOP) with short, needle-like crystals of different length and orientation (GIUS 8-3730/30). **(G)** The longitudinal section of the *P. lumbricalis* tube from the lower Kimmeridgian of Małogoszcz (GIUS 8-3747/5) showing the spherulitic arrangement of crystals within the external tube layer. **(H, I)** Longitudinal sections of the two *P. lumbricalis* tubes from the middle Bathonian of Gnaszyn Dolny showing the external spherulitic prismatic structure (SPHP) with well-visible growth increments and the internal, partially recrystallized irregularly oriented prismatic structure (IOP) (**H.** GIUS 8-3730/31; **I.** GIUS 8-3730/32). **(J)** The lateral section of the *P. lumbricalis* tube from the middle Bathonian of Gnaszyn Dolny showing tubulae subdivided densely by septae (GIUS 8-3730/33).

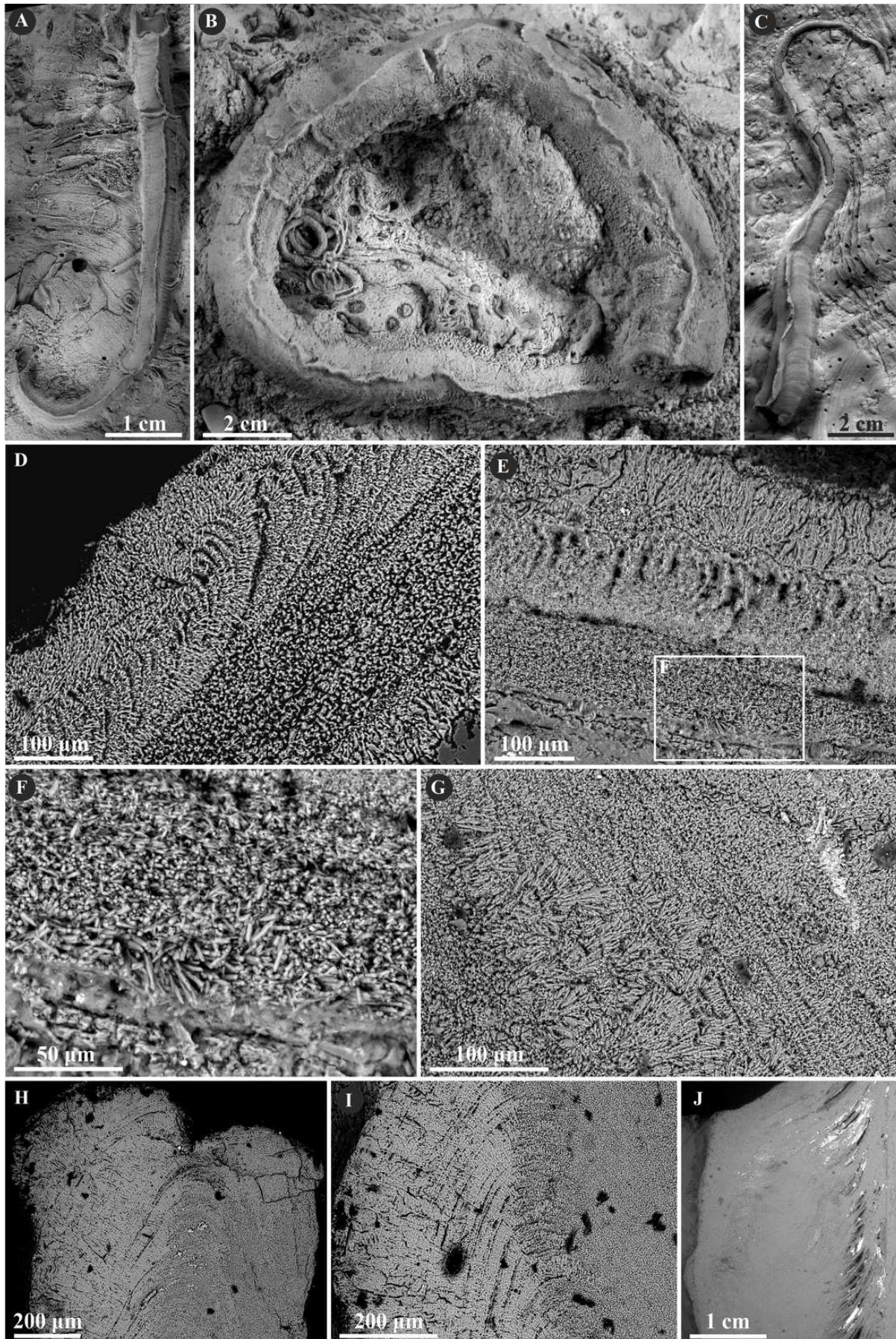


Figure 8

Figure 8: Selected specimens of *Nogrobs* from the Jurassic of Poland.

(A) Two specimens of *Nogrobs* aff. *quadrilatera* (A. Goldfuss, 1831) encrusting a belemnite rostrum from the middle Bathonian of Gnaszyn Dolny (GIUS 8-3730/17). **(B)** *Nogrobs?* aff. *tricristata* (A. Goldfuss, 1831) encrusting a belemnite rostrum from the middle Bathonian of Gnaszyn Dolny (GIUS 8-3730/19). **(C, D)** Free-lying tubes of *Nogrobs* aff. *tetragona* (J. de C. Sowerby, 1829) (**C.** GIUS 8-3730/24; **D.** GIUS 8-3730/25); lateral (**C₁**) and cross-section (**C₂**, **D**) view. **(E, F, G, I, J)** The longitudinal sections of the *Nogrobs* aff. *tetragona* (**E, F.** GIUS 8-3730/34), *Nogrobs?* aff. *tricristata* (**G.** GIUS 8-3730/20), and *Nogrobs* aff. *quadrilatera* (**I.** GIUS 8-3730/35; **J.** GIUS 8-3730/36) tubes from the middle Bathonian of Gnaszyn Dolny showing two distinct layers: the external simple prismatic structure (SP) with distinct growth lines and internal irregularly oriented prismatic structure (IOP). **(H, K)** A close-up of the longitudinal sections of the internal tube layer of *Nogrobs?* aff. *tricristata* (**H.** GIUS 8-3730/20) and *Nogrobs* aff. *quadrilatera* (**K.** GIUS 8-3730/36) showing irregularly oriented prismatic structure (IOP) with short, needle-like crystals of different length and orientation.

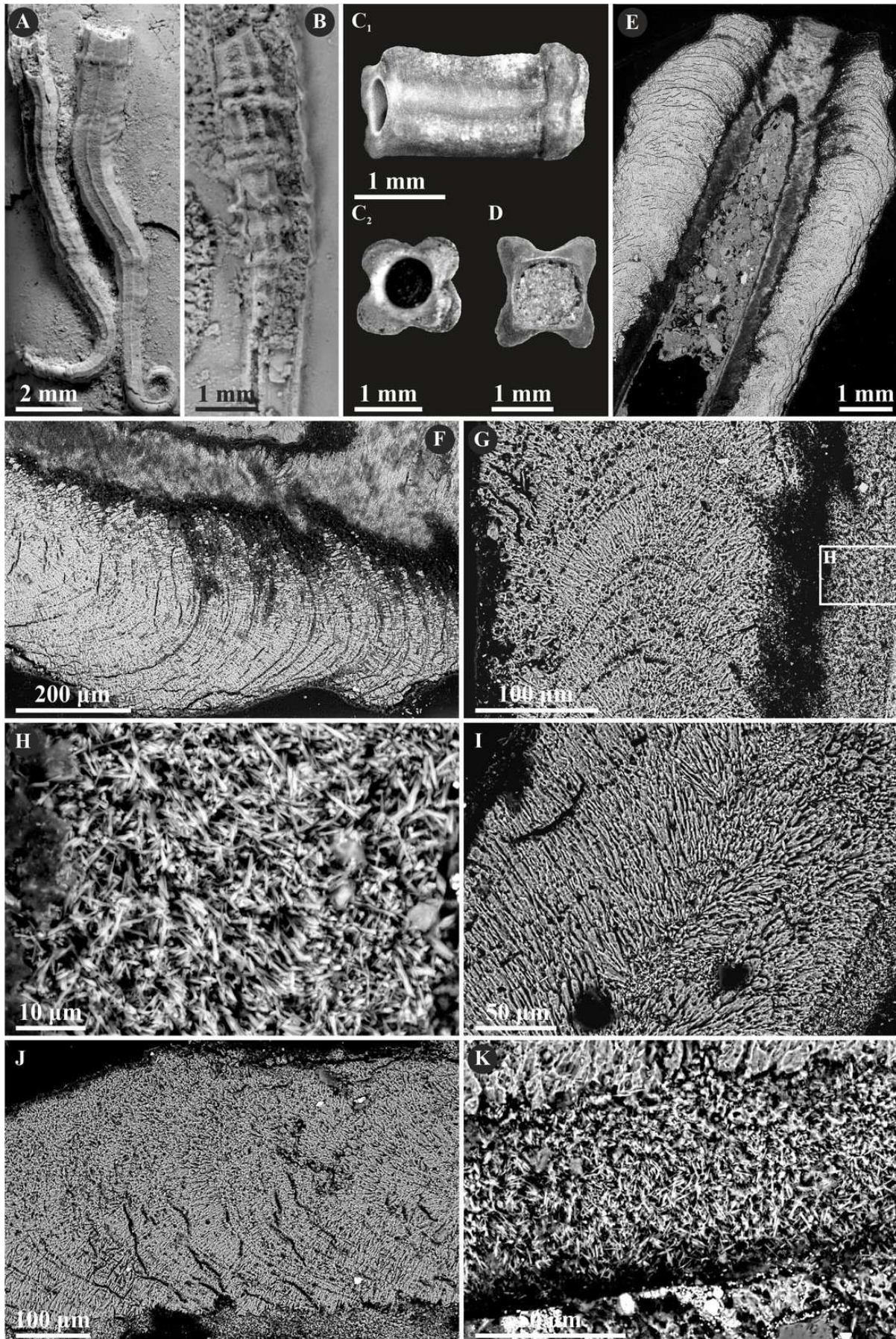


Figure 9

Figure 9: Selected specimens of *Mucroserpula* (H. Regenhardt, 1961) from the Jurassic of Poland.

(A, B) *Mucroserpula* sp. encrusting a fragment of a shell from the middle Bathonian of Gnaszyn Dolny (GIUS 8-3730/26); top **(A)** and cross-section view **(B)**. **(C-E)** longitudinal sections of the *Mucroserpula* sp. tube from the middle Bathonian of Gnaszyn Dolny showing two distinct layers: the external spherulitic prismatic structure (SPHP) with distinct, chevron-shaped growth lines **(C)** and internal irregularly oriented prismatic structure (IOP) (GIUS 8-3730/26). **(F)** A close-up of the longitudinal section of the internal tube layer showing irregularly oriented prismatic structure (IOP) with short, needle-like crystals of different length and orientation (GIUS 8-3730/26).

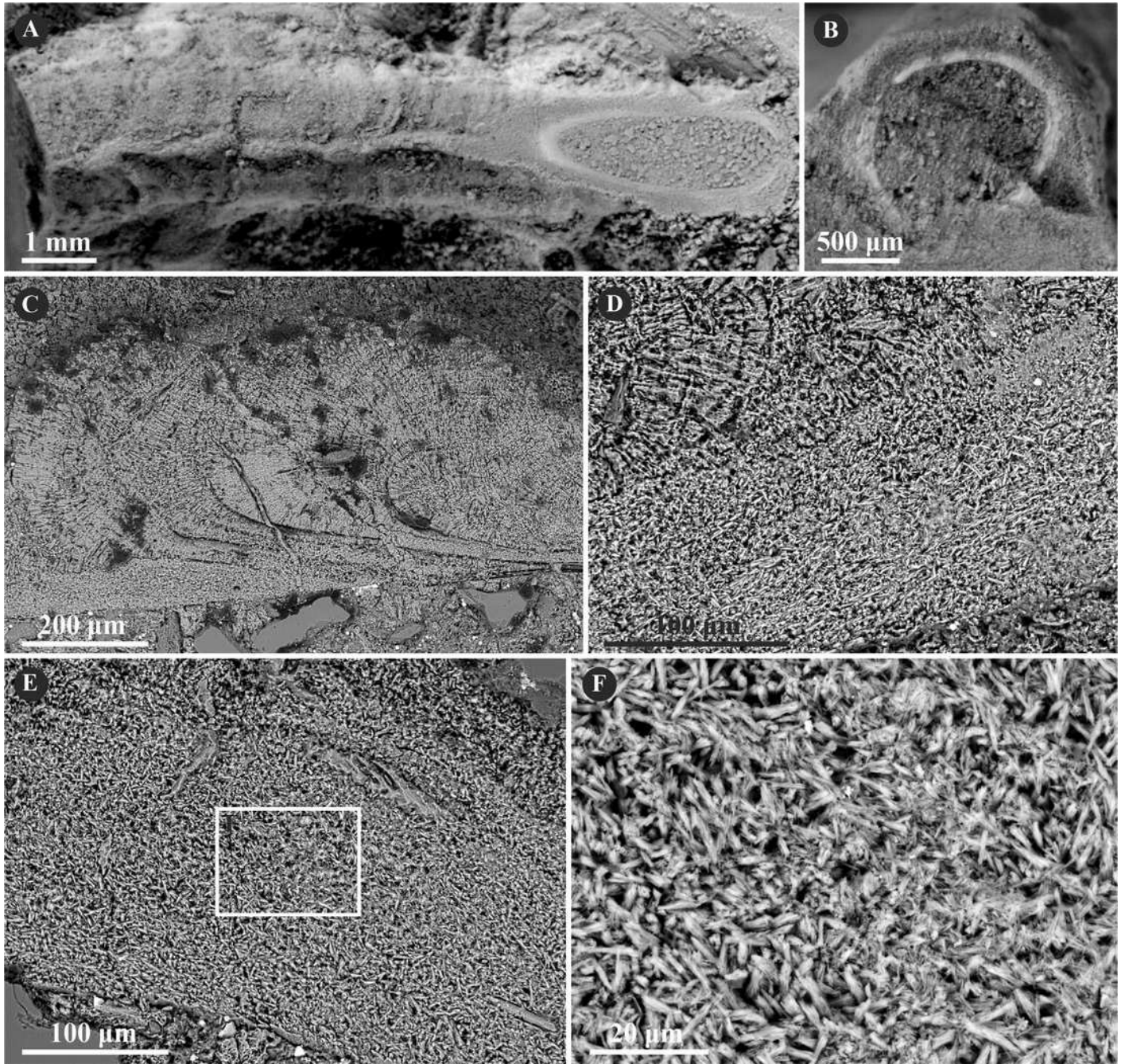


Figure 10

Figure 10: Selected specimens of *Placostegus planorbiformis* (Münster in Goldfuss, 1831) from the Jurassic of Poland.

(A) *P. planorbiformis* encrusting a sponge fragment from the Oxfordian of Zalas (GIUS 8-3746/8). **(B-D)** Longitudinal sections of the tubes showing the simple prismatic structure (SP) with well-visible growth increments (GIUS 8-3746/8).

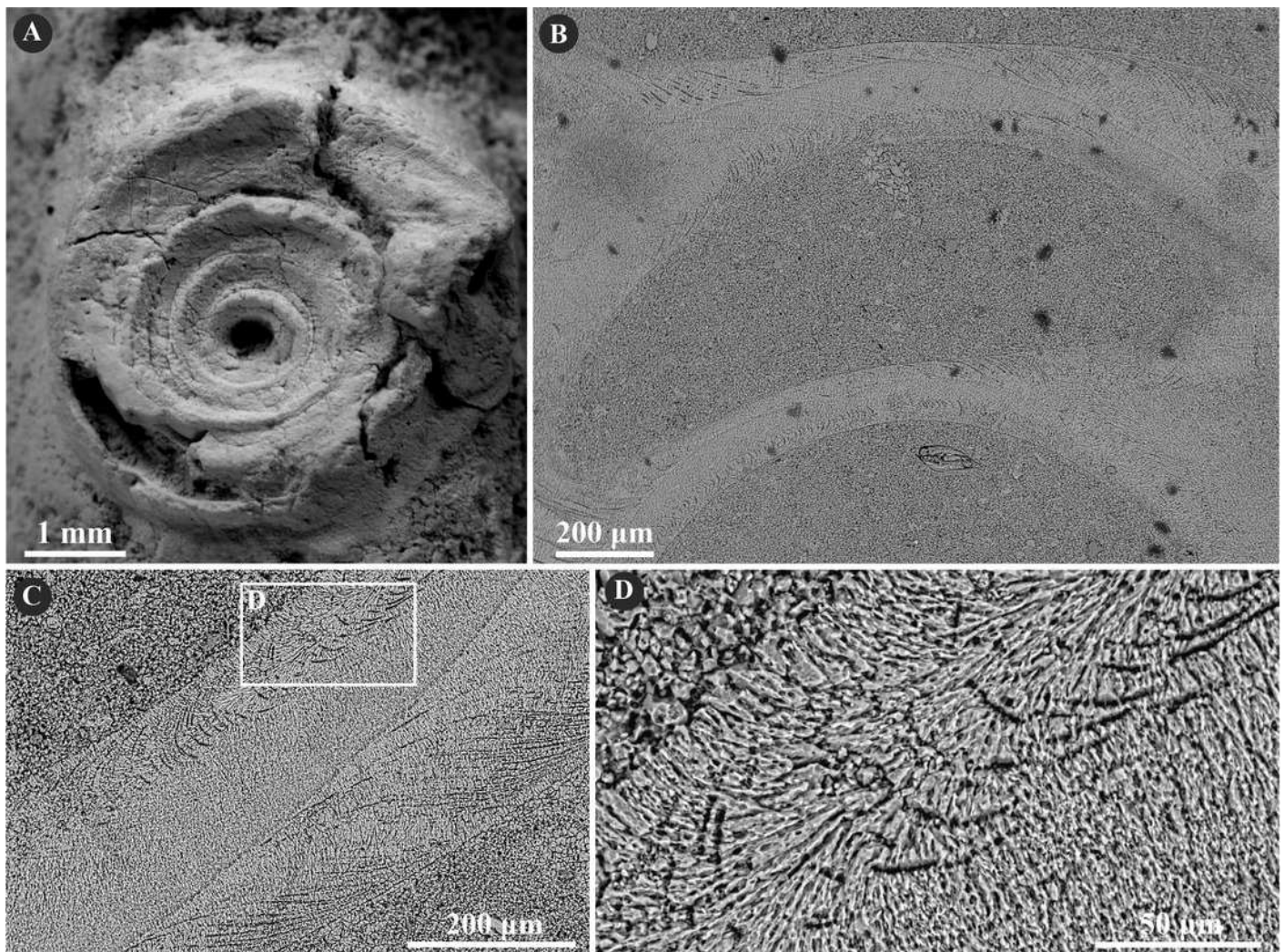


Figure 11

Figure 11: Selected specimens of Serpulidae sp. from the Jurassic of Poland.

(A) Dense aggregation of closely spaced Serpulidae sp. 3 (see Słowiński et al., 2022) encrusting a fragment of a belemnite rostrum from the middle Bathonian of Gnaszyn Dolny (GIUS 8-3730/28). **(B-E)** Longitudinal sections of the Serpulidae sp. tube from the middle Bathonian of Gnaszyn Dolny showing two distinct layers: the external simple prismatic structure (SP) and internal irregularly oriented prismatic structure (IOP) (GIUS 8-3730/28).

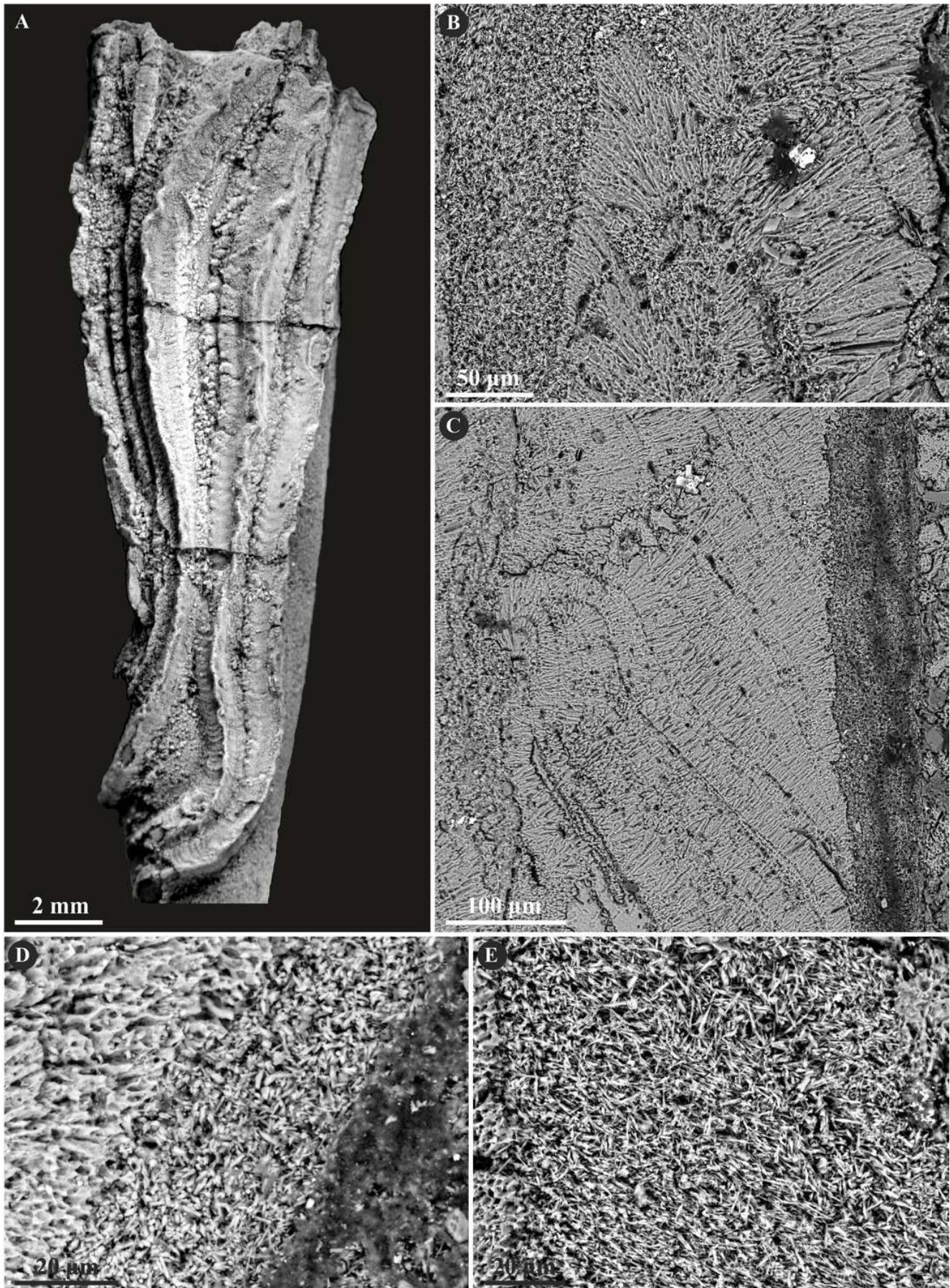


Table 1 (on next page) 

Table 1: Ultrastructural diversity of the Jurassic serpulid tubes from Poland.

IOP - irregularly oriented prismatic structure, SPHP - spherulitic prismatic structure, SP - simple prismatic structure. Numbers (1, 2, 3) indicate the position of the layer within the tube counted from the exterior. Square brackets denote partially preserved or equivocal microstructure precluding a fully reliable recognition. Green, yellow, and blue correspond to Filograninae, Serpulini, and Ficopomatini respectively.

Taxon	Ultrastructure			Number of individuals investigated
	IOP	SPHP	SP	
<i>Metavermilia cf. striatissima</i>	1			3
<i>Filogranula runcinata</i>	[2]	1		5
<i>Filogranula spongiophila</i>			1	3
<i>Cementula spirolinites</i>			1	9
<i>Cementula radwanskae</i>			[1]	4
<i>Propomatoceros lumbricalis</i>	2	1, [3]		8
<i>Nogrobs aff. quadrilatera</i>	2		1	3
<i>Nogrobs? aff. tricristata</i>	2		1	1
<i>Nogrobs aff. tetragona</i>	2		1	2
<i>Mucroserpula sp.</i>	2	1		3
<i>Placostegus planorbiformis</i>			1	3
Serpulidae sp.	2		1	3
Total				47

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