

Recovery of deep-sea meiofauna community in Kaikōura Canyon following an earthquake-triggered turbidity flow (#93989)

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Recovery of deep-sea meiofauna community in Kaikōura Canyon following an earthquake-triggered turbidity flow

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Turbidity flows can transport massive amounts of sediment across large distances with dramatic, long-lasting impacts on deep-sea benthic communities. The 2016 M_w 7.8 Kaikōura Earthquake triggered a canyon-flushing event in Kaikōura Canyon, New Zealand, which included significant submarine mass wasting, debris, and turbidity flows. This event provided an excellent opportunity to investigate the effects of large-scale natural disturbance on benthic ecosystems. Benthic meiofauna community structure before and after the event was analysed from a time series of sediment cores collected 10 years and 6 years before, and 10 weeks, 10 months, and 4 years after the disturbance. Immediately after the 2016 event abundances of all meiofauna dramatically decreased. Four years later the meiofauna community had recovered and was no longer distinguishable from the pre-event community. However, the nematode component of the community was similar, but not fully comparable to the pre-event community by 4 years after the disturbance. Community recovery was systematically correlated to changes in the physical characteristics of the habitat caused by the disturbance, using physical and biochemical variables derived from sediment cores, namely: sediment texture, organic matter, and pigment content. While these environmental variables explained relatively little of the overall variability in meiofauna community structure, particle size, food availability and quality were significant components. The minimum threshold time for the meiofauna community to fully recover was estimated to be between 3.9 and 4.7 years, although the predicted recovery time for the nematode community was longer, between 4.6 and 5 years. We consider the management implications of this study in comparison to the few studies of large-scale disturbances in the deep sea, in terms of their relevance to the efficacy of the marine reserve that encompasses Kaikōura Canyon, along with potential implications for our understanding of the impacts of anthropogenic seafloor disturbances, such as seabed mining.

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Abstract

Turbidity flows can transport massive amounts of sediment across large distances with dramatic, long-lasting impacts on deep-sea benthic communities. The 2016 Mw 7.8 Kaikōura Earthquake triggered a canyon-flushing event in Kaikōura Canyon, New Zealand, which included significant submarine mass wasting, debris, and turbidity flows. This event provided an excellent opportunity to investigate the effects of large-scale natural disturbance on benthic ecosystems. Benthic meiofauna community structure before and after the event was analysed from a time series of sediment cores collected 10 years and 6 years before, and 10 weeks, 10 months, and 4 years after the disturbance. Immediately after the 2016 event abundances of all meiofauna dramatically decreased. Four years later the meiofauna community had recovered and was no longer distinguishable from the pre-event community. However, the nematode component of the community was similar, but not fully comparable to the pre-event community by 4 years after the disturbance. Community recovery was systematically correlated to changes in the physical characteristics of the habitat caused by the disturbance, using physical and biochemical variables derived from sediment cores, namely: sediment texture, organic matter, and pigment content. While these environmental variables explained relatively little of the overall variability in meiofauna community structure, particle size, food availability and quality were significant components. The minimum threshold time for the meiofauna community to fully recover was estimated to be between 3.9 and 4.7 years, although the predicted recovery time for the nematode community was longer, between 4.6 and 5 years. We consider the management implications of this study in comparison to the few studies of large-scale disturbances in the deep sea, in terms of their relevance to the efficacy of the marine reserve that encompasses Kaikōura Canyon, along with potential implications for our understanding of the impacts of anthropogenic seafloor disturbances, ~~such as seabed mining~~.

Introduction

Disturbance is a key process that underpins the structure of all marine communities (Sousa, 2001). By creating heterogeneity and redistributing limiting resources (space, refuge, nutrients, etc.) disturbances structure ecological succession, increase habitat variability, and enhance biodiversity (Sousa, 1984; Willig and Walker, 1999). Many physical and biological factors determine the rate and pattern of resilience, resistance, and/or recovery of a community after a disturbance (Sousa, 1984). Here, as in Bigham et al. (2023a; 2023b), resilience refers to the amount of disturbance that an ecosystem or its components can experience before changing to an alternative state, which is sometimes referred to as ecological resilience (Holling, 1996). Resistance is defined as the ability of an ecosystem, or its components, to remain unchanged from its initial state despite a disturbance (Walker et al., 2004). In contrast, recovery is defined as the return time after a disturbance for an ecosystem, or its components, to attain a stable state (Folke et al., 2004). Some of the largest benthic disturbances in the marine environment are caused by subaqueous sediment-density flows, which occur worldwide (Bigham et al., 2021). Sediment density flows occur when the material in submarine landslides mixes with water and creates high-density parcels of turbid water that travel downslope beneath less dense water (Kuenen and Migliorini, 1950; Talling, 2014). These turbulent, sediment-laden gravity flows are hydrodynamically complex, and a single event can contain multiple flow types with spatial and temporal variability (Haughton et al., 2003; Talling et al., 2007; Paull et al., 2018). As such, many terms and classification schemes have been proposed to differentiate and recognise flow types, although confusion around the interpretation and application of these terms persists (Kuenen and Migliorini, 1950; Lowe, 1979; Talling et al., 2013). Herein, as in Bigham et al. (2023a; 2023b), the term “turbidity flow” *sensu stricto* Kuenen and Migliorini (1950) will mainly be used because it is the commonly used overarching term for sediment density flows in the ecological literature (cf. Bigham et al., 2021). Turbidity flows impact the benthic faunal communities in their path through both erosional and depositional processes (Bigham et al., 2021), but it is not clear to what extent these communities are resilient to the impacts of these different disturbances. Studies from the 2011 Tōhoku Earthquake showed rapid recovery (within 1.5 years) of the meiofaunal communities following a triggered turbidity flow (Kitahashi et al., 2014, 2016; Nomaki et al., 2016). Turbidity flows pose a particular recolonisation challenge to meiofauna because they typically can only migrate laterally into relatively small, disturbed patches (Chandler and Fleeger, 1983; Gallucci et al., 2008; Gollner et al., 2013) and otherwise must be dispersed passively (Ptatscheck and Traunspurger, 2020). Despite these functional limitations and the potential for large-scale disturbances from turbidity flows, researchers of the Tōhoku studies hypothesised that meiofauna were more resilient to turbidity flows than macrofauna due to their faster turnover times and lower sensitivity to changes in environmental factors (Kitahashi et al., 2014, 2016; Nomaki et al., 2016). Studies of turbidity flows hundreds to thousands of years old have suggested that the impact of the disturbance is still detectable thousands of years after the event for all three benthic metazoan size classes (meio-, macro- and megafauna), although results were somewhat ambiguous for meiofauna specifically (e.g., Briggs et al., 1996; Griggs et al., 1969; Woods and Tietjen, 1985; Lamshead et al. 2001). Furthermore, such studies of older turbidity flows are often confounded by local patterns of surface productivity and terrigenous

inputs that have occurred in the intervening time (Richardson et al., 1985; Richardson and Young, 1987; Thurston et al., 1994; Briggs et al., 1996; Thurston et al., 1998). Many studies, even ones of more recent turbidity flows, lack sufficient pre-disturbance data to fully interpret the impacts of turbidity flows on benthic communities (Bigham et al., 2021). Kaikōura Canyon off eastern New Zealand is the site of a recent and large earthquake-triggered turbidity flow (Mountjoy et al., 2018), with the additional context of pre-event benthic data (De Leo et al., 2010; Leduc et al., 2014).

Kaikōura Canyon on the northeastern side of the South Island, New Zealand, has been dubbed a benthic productivity hotspot due to an abundant macro- and megafaunal biological community with biomasses 100 times higher than those seen in (non-chemosynthetic) deep-sea habitats below 500 m (De Leo et al., 2010). The canyon also supports a distinct nematode community in response to high food availability and high frequency of disturbance, and which contributes significantly to regional meiofaunal diversity (Leduc et al., 2014). High organic carbon content and elevated meiofaunal biomass in Kaikōura Canyon, relative to another New Zealand canyon on the opposite side of the South Island (Hokitika Canyon), was inferred to be related to land-derived organic matter as a dominant food source (Leduc et al., 2020). On 14th November 2016, the Mw 7.8 Kaikōura Earthquake triggered a highly complex “full canyon-flushing event” that reshaped the canyon floor and transported an estimated 850 metric megatons of sediment and 7.5 metric megatons of organic carbon through the canyon, and into the slope basin via the Hikurangi Channel (Mountjoy et al., 2018). The flushing event was geomorphologically complex, including submarine landslides and other local mass wasting episodes that generated debris and cascading turbidity flows down the canyon walls, forming a large flow down the canyon axis (for simplicity, as above, this event is hereafter referred to as the “turbidity flow”). Analysis of time-series imagery from the canyon found that the seafloor and near-seafloor megafauna community structure was recovering, with full recovery predicted 4.6-5.2 years after the turbidity flow (Bigham et al., 2023a). Analysis of the macrofauna community structure in the canyon substrate also suggested ongoing recovery with full recovery predicted 5.6-6.7 years after the turbidity flow (Bigham et al., 2023b). With comprehensive repeat datasets from before and after the turbidity flow, Kaikōura Canyon provides a unique opportunity to also explore meiofauna community resilience to the impact of turbidity flows on deep-sea fauna.

The present study compares meiofauna community structure in Kaikōura Canyon before and after the turbidity flow event in 2016 to determine the community response to the event in relation to changes in the environmental characteristics of the habitat caused by the disturbance. The management implications for the Hikurangi Marine Reserve, which envelopes the Kaikōura Canyon head and much of its middle reach, and the potential for turbidity flows to be considered as proxies for predicting the impacts of widespread physical disturbances caused by large-scale deep-sea mining in the future are also considered, as they were for the megafauna and macrofauna studies (Munoz-Royo et al. 2022; Bigham et al. 2023a, 2023b).

Materials & Methods

Site descriptions

Kaikōura Canyon is located off the northeastern coast of New Zealand's South Island (Figure 1).

The canyon is 60 km long, broadly sinuous, ranges in depth from 20 m to > 2000 m, is generally U-shaped in profile, and is the primary headwater source for the 1500-km long Hikurangi Channel, which transports sediments from the axial mountain chain of the South Island to a distal abyssal fan-drift northeast of New Zealand (Lewis, 1994). The Kaikōura Canyon incises into the narrow continental shelf, the head shoaling to within 500 m of the shore (Lewis and Barnes, 1999). The November 2016 “full canyon flushing” event caused significant erosion and deposition on the canyon floor (Mountjoy et al., 2018). These areas of impact, indicated by the measured bathymetric changes (Figure 1), informed the post-event benthic sampling campaigns, as did the location of pre-disturbance sampling in the canyon. Impacts to the megafauna and macrofauna communities from this same site and disturbance can be found in Bigham et al. (2023a; 2023b).

Sampling and sample processing

Sediment core samples were collected from the R/V *Tangaroa* using an Ocean Instruments MC-800A multicorer (internal core diameter = 9.52 cm), which can collect up to eight cores in a single deployment. These cores were processed for different analyses, such as meiofauna community or sediment characteristics as well as macrofauna community (see Bigham et al. 2023b). Samples from eight sites along the axis of Kaikōura Canyon (depth range 400-1300 m) were collected 6 years before the turbidity flow and 10 months and 4 years after the event (voyages TAN1006, TAN1708, and TAN2011, respectively). Samples from two of the eight main sites were also collected 10 years before and 10 weeks after the disturbance (TAN0616 and TAN1701, respectively) (Figure 1C). See Table 1 for site and sampling details and Figure 1B for site locations. Bathymetric difference mapping by Mountjoy et al. (2018) identified zones along the canyon length after the flushing event that were net erosional (downcut) or depositional (elevation gain). As can be seen on the map in Figure 1, the samples come from sites broadly occupying two different disturbance regimes – most of the samples come from sites where the net change was erosional, but two of the deepest sample sites, K06 and K07 are from depositional zones.

For this study, one core per site was analysed for meiofaunal community analyses, except for samples from 10 weeks after (TAN1701) where two cores were analysed and 6 years before (TAN1006) where two to three cores were analysed (see Table 1 for details). Each meiofauna sample consists of a syringe subcore (internal diameter 26 or 29 mm) to 5 cm depth. The subcore was sectioned at 0-1 cm and 1-5 cm depth and preserved in 100% buffered formalin, though only depth integrated 0-5 cm results are reported here. In the laboratory, samples were sieved through a 1 mm mesh with fresh water to remove macrofauna, and through a 45 µm mesh size to retain meiofauna. Ludox flotation was used to extract meiofauna from the remaining sediment (Somerfield and Warwick, 1996). Samples were transferred to a Bogorov tray and all meiofauna present in the sample were identified to major taxa (e.g., nematodes, annelids, harpacticoid copepods, kinorhynchs) and counted using a compound microscope (x100 magnification).

Nematodes were transferred to a mixture of dilute ethanol and glycerol in a cavity block and left under a fume hood for at least 48 h to allow water and ethanol to evaporate, leaving the sample material in pure glycerol (Somerfield and Warwick, 1996). Nematodes were mounted on slides in pure glycerol and sealed with paraffin wax. Nematode body volumes were estimated using

ImageJ to measure length and maximum body width for all eight sites. Body volumes were converted to dry weight (DW) based on a relative density of 1.13 and a DW: wet weight (WW) ratio of 0.25 (Feller and Warwick, 1988). Nematodes present in the samples from sites K2 and K3 (green circles in Figure 1B) 6 years before and 10 weeks, 10 months, and 4 years after the turbidity flow were identified to species/morphospecies using a compound microscope (x1000 magnification) and percentage of juveniles to adults was recorded. The percentage of juveniles was averaged for each time point. One additional core per site and per time point was analysed for sediment parameters. These are the same environmental parameters used in Bigham et al. (2023b) for comparison with the macrofaunal community in the canyon and full details can be found there.

Statistical analysis

Many of the same statistical analyses used in this study were also used in Bigham et al. (2023a; 2023b) and further details on the methodology can be found there. As was the case in the previous two studies, unless otherwise noted statistical analyses were carried out using routines in PRIMER 7 (Clarke and Gorley, 2018) with PERMANOVA + (Anderson et al., 2008). Meiofaunal communities typically comprise both permanent meiofaunal taxa (e.g., nematodes and harpacticoid copepods, which spend their entire life cycle in the sediment) and temporary meiofaunal taxa (i.e., juveniles of macrofaunal-sized taxa, such as most polychaetes and molluscs, which typically have a pelagic larval stage) (Warwick 1988). Temporary meiofauna are typically excluded from analyses of meiofauna communities due to being larger, having a highly patchy distribution, and occurring at low densities relative to permanent meiofauna (Higgins and Thiel 1988). However, juvenile macrofauna are an important indicator of overall community response to disturbance from the turbidity flow as they provide information on macrofauna taxa that would not be captured in the macrofauna analyses alone (Bigham et al., 2023b). Since this study is concerned with the full community response, both permanent and temporary meiofauna were used in the community analysis. Nematodes are typically the most abundant taxon of meiofauna communities (Giere, 2008). They are an important group for indicating impact and recovery from disturbance and specific genera are used as indicator taxa for disturbance (Boyd et al., 2000; Lamshead et al., 2001; Van Gaever et al., 2009; Leduc and Pilditch, 2013; Semprucci et al., 2015; Zeppilli et al., 2015; Ingels et al., 2020). However, due to the time constraints of high-level taxonomic identification (Miljutin et al., 2010) only the two sites (K2 and K3) with the most time points had nematodes identified to species level. Therefore, where appropriate, statistical analyses were undertaken on both the complete meiofaunal community and nematode species community separately. It was preferred to use a single point to represent 'Before' the turbidity flow event for the statistical analysis. Therefore, an analysis of similarity test was used to confirm there was no significant difference in community structure between data from 10 years and 6 years before (ANOSIM, $R = 0.466$, $p = 0.089$, Number of permutations: 45). The exception to using the combined Before data was for the Distance-based linear models (DISTLM) only data from 6 years before was used because environmental data was not available from 10 years before the disturbance event.

212 Community structure

213 Analyses were run on data from the 0-5 cm sediment depth layer for both the non-species level
 214 meiofauna data, including nematodes (referred to hereafter as “meiofauna” community data), as
 215 well as the nematodes identified to species level (referred to hereafter as “nematode”
 216 community data). For the meiofauna community analyses replicate cores from the same site
 217 were averaged. For the nematode community analyses replicate cores from the same site were
 218 kept separate.

219 Similarity matrices for the multivariate community structure data were made using the zero-
 220 adjusted Bray-Curtis similarity measure of square root transformed abundances (Clarke and
 221 Gorley 2018). Variability in the community structure through times was tested using
 222 PERMANOVAs with Type III (partial) sums of squares, unrestricted permutation of raw data and
 223 9999 permutations. A pair-wise PERMANOVA was only run on the meiofauna data, ~~not the~~
 224 ~~nematode~~ data since there were only two sites per time point ~~for the nematode data~~. The results
 225 of these multivariate community structure analysis were visualized using two-dimensional non-
 226 metric multi-dimensional (nMDS) plots. The centroids, the point at the centre of the data cloud,
 227 provide a simplified view of the overall patterns. The SIMPER routine was run on meiofauna and
 228 nematode data to determine the contribution of taxa to within and between community similarity
 229 for each time point. A cut-off of 70% was used in the SIMPER routine to identify key taxa
 230 contributing to the similarity/dissimilarity. Further, the RELATE test of cyclicity (Correlation
 231 method: Spearman rank (ρ), Number of permutations: 9999), MVDISP, and PERMDISP
 232 (Number of permutations: 9999) routines were used to evaluate the patterns observed in the
 233 meiofauna nMDS plot. Permutational Analysis of Multivariate Dispersion (PERMDISP) (Number
 234 of permutations: 9999) was used to determine the significance of differences in the multivariate
 235 dispersion (Anderson et al., 2008). It was not possible to run the RELATE test, MVDISP, or
 236 PERMDISP on the nematode data due to the small sample size.

237 Environmental drivers

238 Distance-based linear models (DISTLM) were used to assess the effect of environmental
 239 parameters on the meiofauna and nematode community structure. The predictor variables were
 240 those sediment parameters detailed in Bigham et al. (2023b), which were used to characterise
 241 food availability and physical sediment habitat, as well as the water depth at which the sample
 242 was taken. Correlation between environmental variables was checked before running the
 243 DISTLM using draftsman plots and correlation matrices. When Pearson's r was >0.8 between
 244 variables, one of the variables was excluded from the analysis; if more than one variable
 245 correlated with others, the variable with the most correlations was kept. Both DISTLMs were run
 246 with stepwise variable selection, the Akaike Information Criterion (AIC) and 9999 permutations.
 247 Both marginal and sequential tests were conducted. Marginal tests examine a single variable
 248 separately, while the sequential test takes in to account the previously tested variables when
 249 examining each variable (Anderson 2008). The best models proposed by the DISTLM were
 250 visualized with distance-based redundancy analysis (dbRDA) ordination plots.

252 Predicting recovery

While a result of no significance difference between the meiofauna community structure before and 4 years after the turbidity flow indicates that recovery has occurred (see Results) it does not provide an estimate of the trajectory of the recovery, nor an indication when recovery may have occurred before the time point that the non-significant result was apparent. To estimate the recovery trajectory, rates of recovery were predicted by fitting three common models of population growth (linear, exponential, logistic; Lundquist et al. 2010; Solé et al. 2010) using the generalized linear model and nonlinear least squares functions in R (R Core Team, 2022). The models were fitted to the observed similarity at the three post-event time points for the 0-5 cm sediment layer for the meiofauna and the nematode data. The meiofauna and nematode community were predicted to be recovered when they at least reached the level of within-group similarity exhibited by the pre-turbidity flow community (i.e., 79% and 46.1%, respectively).

Results

Community structure

The Kaikōura Canyon meiofauna community has recovered overall following the disturbance by the turbidity flow. The community structure differed significantly between prior to the disturbance and 10 months after the event (PERMANOVA, $p < 0.0001$, Table 2), but by 4 years after the turbidity flow the community structure was no longer significantly different ($p = 0.1703$, Table 2). In contrast, the main PERMANOVA test for the nematode community indicates a significant difference in the community structure among all time points, for the two-site subset of data ($df = 3$, $SS = 13,174$, $MS = 4,391.2$, $Pseudo-F = 2.4428$, $P(perm) = 0.0014$). The recovery pattern of the meiofauna and nematode communities are illustrated in the multivariate ordinations of community similarity. The nMDS plots show clustering of samples by time point, with samples from Before the turbidity flow and 4 years after clustered most distinctly on the left-hand side of the plot, and samples taken 10 weeks and 10 months after the event spread out on the right-hand side (Figure 2A and C). The centroids of the meiofauna and nematode community sample data, with trajectories overlaid, are also displayed to provide a simplified illustration of the pattern (Figure 2B and D). The test for cyclicity for the meiofauna community was not significant ($\rho = 0.061$, $p = 7.42\%$), meaning that the meiofauna community's pattern of recovery was not comparable to a simple, equal distance cyclical recovery. Community variability (dispersion) for the meiofauna community was greatest in the weeks and months after the turbidity flow (10 weeks: 1.165, 10 months after: 1.56), then decreased as recovery progressed towards the original community structure (4 years after: 0.72) and was significantly different between all time points ($Pseudo-F = 7.1157$, $P(perm) < 0.001$). The SIMPER analysis of the meiofauna community found that nematodes consistently contributed between 69 and 76% of the within community similarity, with copepods also being key contributors 10 months after the turbidity flow and nauplii key contributors 10 weeks after the disturbance event (Table 3). The nematode community SIMPER analysis found that between four and nine species of nematodes explained within time point community similarity. *Hopperia beaglese* had the highest contribution to within time point community similarity Before the turbidity flow, while *Daptonema* sp. 18 was the highest contributor 10 weeks, 10 months, and 4 years after the event (Table 6).

At the meiofauna community level, community dissimilarity among time points was explained by up to five taxa. Dissimilarity was highest between samples from Before and 10 weeks after the turbidity flow at 57.75%, with only a small decrease in dissimilarity to 51.67% between Before and 10 months after the event. Ten weeks and 10 months after the disturbance the average abundances of all key contributing taxa were lower than the average abundances from before the event. Nematodes and kinorhynchs were key contributors to the differences between the community at 10 weeks after and Before the turbidity flow. Along with nematodes and kinorhynchs, nauplii contributed to differences between the Before community and the community 10 months after the event. Dissimilarity was lowest, at 20.75%, between samples from before the turbidity flow and 4 years after the event. Nauplii, copepods, and gastrotrichs were key contributors to dissimilarity between the community 4 years after the turbidity flow and the Before community. Four years after the event, nematode average abundances were similar to their pre-disturbance levels, while copepod and nauplii average abundances exceeded pre-event levels. Kinorhynch and gastrotrich abundances at 4 years after the event remained depressed compared to before the turbidity flow (Table 6).

Nematode community dissimilarity among time points can be explained by many species. *Sabatieria* sp. A, *Hopperia beaglense*, *Microlaimus* sp. 34, *Cervonema kaikouraensis*, and *Daptonema* sp. 18 were consistently among the highest contributors to the observed dissimilarity. Dissimilarity was highest, at 81.79%, between the community Before the disturbance and the community sampled 10 weeks after the turbidity flow. This difference in the two communities was represented by a large decrease in the abundance of all key contributory taxa. Dissimilarity decreased to 77.50% between the Before community and the community 10 months after the turbidity flow. At this stage in community recovery the dissimilarity was characterized with continued low average abundances for most key contributory taxa except for *Daptonema* sp. 18, which had nearly tripled in abundance from its pre-event average abundance. *Campylaimus* sp. 6, *Leptolaimus* sp. 14, and *Sphaerolaimus* sp. 1 were not observed 10 months after the disturbance despite being present 10 weeks after the event. Community dissimilarity was lowest, at 59.71%, between samples from Before the turbidity flow and 4 years after the event. By 4 years after the turbidity flow, the average densities of contributory species had begun to increase, though had not attained pre-event levels. In contrast, *Leptolaimus* sp. 14 continued to be absent, while *Daptonema* sp. 18 was now 12 times pre-event levels, and *Endeolophos* sp. 3 had increased to an average abundance three times pre-event levels (Table 6).

Environmental drivers

Of the 8 environmental variables included in the DISTLM analysis the marginal test identified two variables (TOM% and PN%) as significant ($p\text{-value} \leq 0.05$) explanatory variables for the meiofauna community structure, and one variable (C:N (molar)) as significant for the nematode community structure. The best DISTLM model ($AIC = 152.82$, $R^2 = 0.28971$, $RSS = 12,481$) for meiofauna community structure included 3 variables, only one (%TOM) of which was significantly correlated to the community structure and explained 14% of the sample variation across all time points (see sequential test under meiofauna in Table 7). While the best DISTLM model ($AIC = 56.28$, $R^2 = 0.9117$, $RSS = 1,578.9$) for the nematode community structure

included 6 variables, of which only one (C:N (molar); explaining 33% of variation) was significantly correlated to the community structure and explained 91% of the sample variation across all time points (see sequential test under nematodes in Table 7). The first two axes of the dbRDA plots explained 24.9% and 3.8% of total community variation for meiofauna, and 37.8% and 17.9% for nematodes (Figure 3). For the meiofauna, dbRDA1 accounted for most of the variation among the samples; it was primarily correlated with %TOM, sediment Chl *a* concentrations, and C:N (molar). dbRDA2 accounts for a much smaller portion of the variation, primarily that for community variation between 6 years before the turbidity flow and the other time points and is correlated with negative sediment skewness and higher percentages of sediment less than 16 μm (Figure 3A). For the nematodes, dbRDA1 accounts for the variation in samples between 10 weeks after the turbidity flow and the other three time points; it is primarily correlated with C:N (molar) and the ratio of Chl *a* to phaeopigments. dbRDA2 also accounts for a large amount of variation, primarily between the samples taken 6 years before and 10 months and 4 years after the disturbance event; this axis correlates to Chl *a* concentrations and percent particulate nitrogen (Figure 3B). The sediment samples taken 10 weeks, 10 months after, and 4 years after the turbidity flow all had higher percentages of TOM, nitrogen, and sediment particles greater than 16 μm compared to 6 years before the event. In contrast, all had lower concentrations of Chl *a*, ratio of Chl *a* to phaeopigments, ratio of C:N (molar), and a slightly negative skewed distribution of sediment grain size (Figure 4A, B, C, D, E, F, G). Additionally, highly correlated but removed variables would likely also explain the same variation in community structure described above.

Predicting recovery

Since the PERMANOVA test indicated that there was no significant difference in the meiofauna community structure between the Before community and four years after turbidity flow, this community can be considered recovered. To determine the recovery trajectory and when recovery may have occurred prior to the final sampling point, recovery rates for the meiofauna community were estimated using three different population growth models (linear, exponential, and logistic). These models confirmed the results from the PERMANOVA test and predicted that the impacted meiofauna community would exhibit the same within-group level of similarity as the pre-disturbance community (79%; the threshold used for predicted recovery) between 3.9 and 4.0 years after the turbidity flow (Figure 5). The same three population growth models were used to estimate recovery rates for the nematode community, which the PERMANOVA test indicated was still significantly different 4 years after the turbidity flow. The models predicted that the impacted nematode community could exhibit the same within group level of similarity as the pre-disturbance community (46%) between 4.6 and 5.0 years after the turbidity flow.

Nematode juvenile percentage

The highest percentage of juveniles was observed 10 weeks after the turbidity flow, at 53.1%. The percentage of juveniles at 10 months and 4 years after the disturbance was 39.6% and

377 28.1%, respectively. The percentage of juvenile nematodes 6 years before the turbidity flow was
378 41.6% (Figure 6).

379 Discussion

380 Impact of turbidity flow on meiofauna and nematode community structure

381 The meiofauna community sampled in Kaikōura Canyon was not resistant to disturbance
382 caused by the 2016 Kaikōura Earthquake-triggered turbidity flow, but it appears that the
383 community is resilient because by 4 years after the event the community had largely recovered.
384 However, when considering the nematode component separately – the largest component of
385 the meiofaunal community – using species level identification data (for a sub-set of the study
386 sites), it appears that nematodes were still on a trajectory to recovery, as it had not yet
387 recovered 4 years after turbidity flow disturbance.

388 It is evident that the meiofauna community was significantly altered by the disturbance with 
389 dissimilarity highest between Before and 10 weeks after the turbidity flow: meiofauna, 57.8%;
390 nematodes, 81.8%. The community was in a similar state 10 months after the event, though
391 dissimilarity between the community and the Before event community had decreased:
392 meiofauna, 51.7%; nematodes, 77.5%. The level of dissimilarity between Before the turbidity
393 flow and 4 years after the event had decreased considerably for both meiofauna and nematode
394 communities (20.8%, 59.7%, respectively), and the meiofauna community was no longer
395 significantly different from community sampled before the disturbance. However, while the
396 nematode community was beginning to resemble the pre-disturbance community, there was still
397 a significant difference in community structure. These findings were supported by the sample
398 dispersion values for the meiofauna community which were highest after the disturbance but
399 had returned to a level similar to pre-disturbance by 4 years after. While recovery is occurring
400 the trajectory of the recovery is not comparable to a simple cyclical pattern, which assumes that
401 roughly the same amount of recovery will occur between each time step, indicating another
402 pattern may better describe the meiofauna community's recovery (see below).

403 The meiofauna community before the disturbance was dominated by nematodes (Leduc et al.
404 2014, 2020; Figure 7A). The key nematode species included *Hopperia beaglense*, *Cervonema*
405 *kaikouraensis*, *Camplyaimus* sp. 6, *Leptolaimus* sp. 14, and *Sabatieria* sp. A. The difference in
406 the community Before and 10 weeks after the disturbance event is characterised by a large
407 decrease in abundance of all key taxa (Figure 7B). For example, the abundance of *Sabatieria*
408 sp. A decreased from 355 ind./10 cm² to 11 ind./10 cm² 10 months after the disturbance. This
409 drastic abundance reduction in most taxa is to be expected given the evacuation of substrate
410 from the canyon head, which would have removed most if not all of the living meiofauna
411 community that resided within those sediments prior to the event. Similar removal of all or most
412 fauna has been documented in other studies where substantial amounts of near-surface
413 material are removed, such as harbour and aggregate dredging (Kenny and Rees, 1994;
414 Szymlfenig et al. 2006). From 10 weeks to 10 months after the turbidity flow, the abundance
415 levels of most key taxa remained depressed compared to pre-disturbance levels (Figure 7C).
416 Some taxa saw minor increases from 10 weeks to 10 months after the turbidity flow, likely due
417 to their recovery (i.e., *Sabatieria* sp. A, 10 weeks: 11 ind./10 cm²; 10 months: 18 ind./10 cm²).
418 While other taxa decreased in abundance or were not seen at all 10 months after the

disturbance. For example, kinorhynchs decreased from 2 ind./10 cm² to <1 ind./10 cm² and the nematodes *Camplyaimus* sp. 6 and *Leptolaimus* sp. 14 were not observed despite being present at 10 weeks after the turbidity flow (potentially due to fecundity levels, see below). Decreases in kinorhynch abundances have been reported following organic enrichment and associated increases in sulphide concentrations (Mirto et al., 2012; Dal Zotto et al., 2016). Alternatively, with such low abundances post-turbidity flow the missing taxa may have been present in the overall habitat but not sampled by the two cores analysed for this study. The exception to these small changes in abundance at 10 months after was *Daptonema* sp. 18 which increased to almost three times pre-disturbance abundances. Consistent with these Kaikōura observations, nematodes in the *Daptonema* genus are opportunistic, non-selective deposit feeders that are commonly found in disturbed, organic-rich sediment (Vanreusel, 1990; Schratzberger and Jennings, 2002; Moreno et al., 2008; Liao et al., 2020). Four years after the disturbance, the meiofauna community was no longer significantly different then the community before the disturbance, although not identical. Key taxa in the meiofauna community at 4 years after were nematodes, which had recovered to near pre-disturbance abundance levels, nauplii and copepods, which were slightly more abundant than they had been before the disturbance, and kinorhynchs and gastrotrichs, which had much lower abundances than before the turbidity flow event (Figure 7D). Copepods, gastrotrichs, and nauplii are considered to be more sensitive to stress than nematodes (Murrell and Fleeger, 1989; De Troch et al., 2013; Pusceddu et al., 2013; Zeppilli et al., 2015) providing some explanation for the longer time taken compared to nematodes to re-establish after the turbidity flow in Kaikōura Canyon. A similar relative abundance response by nematodes and copepods to a turbidity flow disturbance was observed after the Tōhoku Earthquake-triggered turbidity flow, where nematode densities remained unchanged after the disturbance, but harpacticoid copepod densities were negatively impacted by the disturbance and it wasn't until months to years after the event that they increased (Kitahashi et al. 2014, 2018). The percentage of juvenile nematodes peaked 10 weeks after the turbidity flow before decreasing 10 months and 4 years after. Conversely, meiofaunal annelids, which mostly comprise juvenile polychaetes (Warwick, 1988), were least abundant 10 weeks after the turbidity flow but steadily increased in abundance at the 10 months and 4 years after time points. The increase in annelids over time after the disturbance indicates recruitment into the macroinfaunal community. The differences in juvenile abundance between these two groups is likely due to the differences in life histories. The peak of juvenile nematodes shortly after the disturbance suggests that their initial recruitment occurred primarily via juveniles rather than adults, probably due to the transport of juveniles from nearby unimpacted locations via sediment resuspension by currents (Ptatscheck & Transpurger 2020). Polychaete recruitment depends on the availability of larvae in the water column, which can be highly variable in time and space depending on reproductive cycles, abundance of adult populations, larval mortality and hydrodynamics (Qian 1999). The separate species level analysis of the nematode community provided, in particular, some additional understanding of the status of this important taxon 4 years after the turbidity flow, when this component of the meiofauna community had yet to fully recover. *Daptonema* sp. 18 dominated the community at the final sampling timepoint but with abundances 12 times higher than pre-disturbance levels. ~~As noted above, *Daptonema* species are opportunistic, non-~~



~~selective deposit feeders that are commonly found in disturbed, organic rich sediment~~
~~(Vanreusel, 1990; Schratzberger and Jennings, 2002; Moreno et al., 2008; Liao et al., 2020).~~
 Other species such as *Sabatieria* sp. A were near pre-disturbance abundance levels, while
Campylaimus sp. 6 was observed but in very low abundances, and *Leptolaimus* sp. 14 was still
 not observed 4 years after the turbidity flow. Similarly, nematodes from the genus *Leptolaimus*
 were rare or absent in fresh iceberg disturbance scours in the Weddell Sea (Lee et al., 2001),
 despite otherwise often being a dominate taxa (Vanaverbeke et al., 1997; Vanreusel et al.,
 2000). This impact on these nematodes was attributed to relatively low fecundity of this genus
 (Lee et al., 2001). Two species that were not identified as key taxa before the turbidity flow,
Endeolophos sp. 3 and *Microlaimus* sp. 34, were identified as key taxa 4 years after the
 disturbance. The genus *Microlaimus* makes up an important fraction of the nematode
 community in the Congo Channel, which is regularly disturbed by turbidity flows (Van Gaever et
 al., 2009). The genus is considered to be an opportunistic coloniser and is often among the first
 taxon to recolonise physically disturbed patches (e.g., Lee et al., 2001; Raes et al., 2010).
 Overall, 4 years after the turbidity flow disturbance the meiofauna and nematode community of
 Kaikōura Canyon has reattained pre-disturbance character of high abundance, low diversity,
 and dominance by a few species/taxa that are typically associated with high food and high
 levels of disturbance experienced in the canyon (Leduc et al. 2014). A similar meiofauna
 community pattern has been observed at other locations disturbed by turbidity flows (Hess et
 al., 2005; Hess and Jorissen, 2009; Lamshead et al., 2001; Tsujimoto et al., 2020; Van Gaever
 et al., 2009) (see below). While the meiofauna community was not significantly different from the
 pre-disturbance community and could be considered recovered, the analysis of species level
 nematode data for two of the eight sites indicated that at this level the community was still
 significantly different from the pre-disturbance community, and therefore recovery was
 incomplete. Using the species level nematode data, recovery was predicted to occur between
 4.6 and 5.0 years after the turbidity flow. These results indicate that while disturbance and
 community recovery can be detected using coarse taxonomic groups (Warwick, 1988; Olsford
 et al., 2003; Musco et al., 2011), the use of species data give a more nuanced understanding of
 change and will likely indicate a longer recovery period than if a coarse taxonomic level is used.
 Predictions of recovery time suggest that a linear model may best describe the pattern of
 recovery the meiofauna exhibited, but this may be due to the limited number of repeated
 samples and additional points are necessary to help establish the recovery pattern.

Comparison with other studies of turbidity flow disturbances

Other meiofauna communities impacted by turbidity flows have generally recovered rapidly from
 the disturbance. Overall, the meiofauna community impacted by the Tōhoku Earthquake-
 triggered turbidity flow recovered by 1.5 years after the disturbance (Kitahashi et al., 2014,
 2016). However, the foraminiferal component of the community was not yet considered
 recovered by this time (Tsujimoto et al. 2020), which contrasts with the foraminifera community
 of the Cap Breton Canyon which was considered recovered ~1.5 years after a turbidity flow in
 this canyon (Hess 2005; Hess and Jorissen 2009). The difference between the recovery time of
 the Kaikōura Canyon meiofauna community and the Japan Trench slope community is notable
 since the Kaikōura Canyon sites are mostly in relatively shallower water depths (400-1,300 m)

compared to the majority of the sites considered in the Tōhoku study (100-6,000 m), because it is generally held that organisms at deeper depths will take longer to recover from disturbances (Nomaki et al. 2016). The difference in these recovery times is likely due to scale of the disturbance at the locations. The Tōhoku Earthquake turbidity flow was less confined by seabed morphology and had a wider, but a reduced sedimentation impact on the Japan Trench slope (1-5 cm of deposition, 0.2 km³ of transported sediment; Kitahashi et al. 2014; Kioka et al., 2019) than the Kaikōura Earthquake turbidity flow had on Kaikōura Canyon (average erosion of 5.6 m, 0.9 km³ of transported sediment; Mountjoy et al., 2018). Similarly, while the Cap Breton turbidity flow occurred in a canyon, it was considerably smaller (8-18 cm of deposition; Anschutz et al., 2002) than the Kaikōura Canyon turbidity flow.

Additionally, following the Tōhoku turbidity flow and the Cap Breton Canyon turbidity flow there was an apparent commensurate decrease in the distribution in the meiofauna community to the sediment subsurface (Hess et al. 2005; Kitahashi et al. 2014; Nomaki et al. 2016; Tsujimoto et al. 2020), potentially in response to burial of organic carbon or other structuring factors (see below). These vertical changes in distribution have also been observed in meiofauna communities from the Congo Canyon that have been impacted by turbidity flows (Galéron et al., 2009; Van Gaever et al., 2009). Data from the present study do not have the same vertical resolution, because sediment slices were taken from 0-1 cm and 1-5 cm rather than 1 cm slices to 5 cm achieved for the Japan Trench slope samples. Hence, it is not possible to assess similar fine-scale changes in vertical distribution in the Kaikōura Canyon meiofauna following the turbidity flow. However, evidence from the megafauna and macrofauna components of the canyon community indicate that the overall community distribution did not change to be deeper in the substrate, and the distribution of some organisms may have instead changed towards the seafloor surface (Bigham et al., 2023a, 2023b).

All three benthic size classes in Kaikōura Canyon were characterised by opportunistic species generally thought to be rapid colonisers and or those with traits that allow them to thrive in habitats with high food availability and high levels of disturbance. The estimated time to recovery for the meiofauna community in Kaikōura Canyon based on the coarse taxonomic level (3.9-4.0 years) is less than that predicted for both the megafauna (4.6-5.2 years; Bigham et al., 2023a) and macrofauna (5.6-6.7 years; Bigham et al., 2023b) communities. It has previously been hypothesised that meiofauna are more resilient to turbidity flow disturbances due to their rapid turnover times and lower sensitivity to changes in environmental factors (Kitahashi et al. 2014; 2016; Nomaki et al. 2016). However, these new recovery estimates for the Kaikōura meiofauna may be an underestimate due to lower taxonomic resolution of these data (cf. Smith and Simpson, 1993; Lasiak, 2003; Bates et al., 2007) with a more complete recovery from the disturbance, as indicated by the nematode species level analysis, predicted to take longer (4.6-5 years), which is on par with the recovery estimate for the megafauna community (Bigham et al., 2023a) but faster than the macrofauna community (Bigham et al., 2023b).

Changes in environmental factors and potential influences on the meiofauna community

The influence of environmental variables on community structure was modelled to provide further explanation for the pattern of meiofauna community structure observed in Kaikōura Canyon following the turbidity flow. The best model for describing the patterns of similarity

observed in the meiofauna and nematode communities among the time points accounted for approximately 29% and 91% of the total variation, respectively. The amount of explanation for the meiofauna community is low, though not unusual for studies of deep-sea meiofauna communities (e.g., Zeppilli et al., 2013; Román et al., 2016), but the amount of explanation for the nematode community is quite high, likely due to the relatively small dataset of only two sites and the species level taxonomic resolution of the nematode dataset.

The Kaikōura Canyon meiofauna community structure has previously been linked to high food availability in the canyon (Leduc et al., 2014, 2020) and the findings from the environmental modelling in the present study suggest the same inference. The community structure over time post-event was best explained by the quantity and quality of the available organic matter and the skewness of the sediment, similar to the results for the macrofauna from Kaikōura Canyon (Bigham et al., 2023b). Post-turbidity flow, the organic matter content of the sediments increased but the overall quality of that organic matter decreased (as reflected in the decrease of Chl *a* to phaeopigment ratios in the sediment). The decrease observed in the concentrations of Chl *a* in the sediments (typically associated with the productivity of phytoplankton) after the turbidity flow, and the related change in the ratio of Chl *a* and phaeopigment concentrations, indicates that there was a decrease in the relative lability of the organic matter in the sediments. This change may have been due to the significant erosion caused by the canyon flushing event (Mountjoy et al., 2018) uncovering older, less labile organic matter or due to an increase in terrestrial material entering the canyon (e.g., Gibbs et al. 2020) following landsliding in the surrounding catchments and hinterland also triggered by the earthquake (Dellow et al., 2017; Croissant et al., 2019; Massey et al., 2020; Thomsen et al., 2020). The overall post-turbidity flow sediment particle size was negatively skewed reflecting an increase in finer particles. An increase in organic matter tends to be closely associated with an increase in fine sediments (Keil et al., 1994; Mayer, 1994; Milliman, 1994) so this change may simply reflect that increase in available organic matter, but it also reflects changes to the arrangement and structure of the physical environment. The physical environment has also been shown to drive changes in the fauna, particularly meiofauna which as the smallest size class that live in the interstitial spaces between sediment particles experiencing changes in the sediment matrix more strongly than larger fauna (Tietjen, 1976; Heip et al., 1985; Etter and Grassle, 1992; Leduc et al., 2012a).

The key variables identified by the environmental models for the nematode community are all connected to food quantity and quality. The most important variable was the ratio of C:N (molar). The relatively low C:N ratios inside the canyon 6 years before the disturbance was attributed to higher overall contributions of “fresh” marine organic matter (Gibbs et al. 2020). Ten weeks after the turbidity flow the C:N ratios were higher for both sites and were still high 10 months after the disturbance, which may be due, as noted above, to the canyon-flushing removing the fresher more labile organic matter and/or exposed older organic matter (Okey 1997). By four years after the disturbance, the C:N ratio attained pre-disturbance levels for all sites, indicating that availability of labile organic matter had returned to pre-disturbance levels (Gibbs et al. 2020). Other important variables identified by the model were an increase in %TOM after the turbidity flow, a decrease in Chl *a* concentrations, and the Chl *a* to phaeopigment ratio in the sediments, and an increase in percent nitrogen in the sediment, reflecting an increase in food availability but a decrease in the quality of that food, as suggested by the relatively elevated C:N ratios (see also above for meiofauna community overall). Other,

non-turbidity flow, studies have found correlations between deep-sea nematode density and distribution and food quality (Levin et al. 1991; Neira et al. 2001; Gallucci et al. 2008). The relatively small amount of variation in community structure explained by the model for the meiofauna may be because of the coarse taxonomic resolution of the data, as evidenced by the higher variation explained by the higher resolution nematode data, or because of other unmeasured biological or environmental factors are instead mainly responsible for the recovery process. For example, the oxygenation and chemical conditions of the sediments have been hypothesized and found to structure meiofauna communities after other turbidity flows. Though not measured sediment oxygen levels were postulated as a driving factor for meiofauna communities impacted by turbidity flows in Cap Breton Canyon (Anschutz et al., 2002; Hess et al., 2005; Hess and Jorissen, 2009). A study of meiofauna following the Tōhoku Earthquake-triggered turbidity flow found that sediment oxygen levels were a key structuring factor for meiofauna-sized copepods (Nomaki et al., 2016). Additionally, oxygen limitation has been proposed more broadly as a direct control on deep-sea meiofauna composition at higher taxonomic levels (e.g., copepods and nauplii density; Levin, 1991; Neira et al., 2001). In Kaikōura Canyon, a study of sediment mixing depth from 4 years after the disturbance found that the maximum mixing depth was 2.19 cm, which may be mediating sediment oxygenation in the canyon (Hale et al., in review).

Management implications

Kaikōura Canyon was designated part of the Hikurangi Marine Reserve in 2014 because it is a benthic productivity hotspot (De Leo et al. 2010) and provides wider ecosystem services (Fernandez-Arcaya et al. 2017), including hosting an abundant marine mammal and avifauna (e.g., Guerra et al., 2020). Concerns were raised following the 2016 Kaikōura Earthquake-triggered canyon flushing event that the efficacy of the reserve had been impacted. Results from this study show that overall, the meiofauna community had largely recovered 4 years after the turbidity flow. However, a more complete recovery from the disturbance, as indicated by the nematode species level analysis, was predicted to take longer (a minimum of 4.6-5 years, i.e., somewhere between 2023 and 2024) and additional samples are necessary to test this prediction. These additional samples would better establish the shape of recovery trajectory patterns and to see if recovery time falls higher on the curve and closer to maximum predicted time to recover.

Natural disturbances in the deep sea have been considered as potential proxies for anthropogenic disturbance with varying levels of validity (Angel and Rice 1996; Tyler, 2003). Debris and turbidity flows create large-scale erosional and depositional disturbances, and thus, could be considered as proxies for some anthropogenic disturbances, such as deep-sea seabed mining where extraction and dredging/turnover of the seafloor can occur. However, results from Bigham et al. (2023a, 2023b) on the recovery of the megafauna and macrofauna component of the Kaikōura Canyon benthic community suggest that the impacts of a turbidity flow on a benthic community was not readily transferable to understanding the impact of future deep-sea mining. This conclusion appears to also be the case for the meiofauna component of the Kaikōura Canyon benthic community. Studies of the impact of small-scale experimental deep-sea mining-related disturbances on meiofauna have shown that fauna at abyssal sites have not

recovered to baseline levels after decades (Miljutin et al. 2011; de Jonge et al. 2020). In contrast, this study estimates that the Kaikōura Canyon nematode community structure could be recovered as soon as 4 years (meiofauna) and 4.6 years (nematodes) after the disturbance, although recovery could take up to 8 years or longer if different levels of community similarity were used as the threshold for recovery. The discrepancy in recovery timing and general lack of transferability between this natural disturbance and seabed mining is likely due to meiofauna communities in Kaikōura Canyon being subjected to much higher levels of natural disturbance from submarine landslides and turbidity flows than abyssal plains where mining for polymetallic nodules may occur in the future. Furthermore, as discussed above, the meiofauna in the canyon are likely to be more adapted to be resilient to these large-scale disturbances. For example, the genera and species of nematodes within Kaikōura Canyon are atypical of deep-sea nematode communities and instead are typically associated with high food availability and high disturbance levels (Leduc et al., 2012b, 2014, 2020). Further, there are discrepancies in the habitat type as well as the scale of the disturbances. In the case of polymetallic nodules, the nodules themselves constitute a unique habitat with meiofauna communities living on and in them that are distinct from the surrounding soft sediments, and which would be predominantly removed by the mining (Thiel et al., 1993; Bussau et al., 1995; Veillette et al., 2007a, 2007b). In contrast the habitat on the floor of Kaikōura Canyon is a mostly uniform soft sediment. Erosion and deposition of sediment by the canyon-flushing event in Kaikōura Canyon was on the scale of metres to tens of metres (Mountjoy et al., 2018), much greater than the tens of centimetres to metres of erosion (Levin et al., 2001) and millimetres of deposition (Thiel et al., 2001) that are expected to occur from seabed mining. The minimum areal extent of the impact from the turbidity flow in Kaikōura Canyon was approximately 220 km² (Mountjoy et al., 2018), which although it is comparable to the hundreds km² per year impacted area envisaged for manganese nodule mining in the abyss (Ardron et al., 2019) seabed mining is expected to occur over successive and multiple years, and therefore may ultimately extend hundreds to thousands of square kilometres (Smith et al., 2008). As such, the recovery estimates from the Kaikōura Canyon study of the impact of turbidity flows on benthic communities are not likely to be good proxies for the recovery of such communities from deep-sea mining on abyssal plains.

Conclusions

The meiofauna community, identified at a coarse taxonomic level, sampled from sediment cores from Kaikōura Canyon appears to be resilient to the earthquake-triggered turbidity flow and has apparently recovered 4 years after the event. However, analysis of species level nematode data (~~for a subset of study sites~~) indicates that this component of the community had not yet recovered by this timepoint and is predicted to take a minimum of 4.6 years to recover. Future sampling at the same sites ~~remains key~~ to ascertain if or when the meiofaunal communities will fully recover. The pattern of resilience for the meiofauna community is somewhat in contrast to those for the megafauna and macrofauna communities examined in previous studies (Bigham et al., 2023a, 2023b).  With data from all three size classes available from Kaikōura Canyon it is now possible to synthesize the overall community resilience and examine inter-size class interaction dynamics during recovery.

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References

- Anderson, M. J., Gorley, R. N., & Clarke, K. R. (2008). PERMANOVA+ for PRIMER: Guide to Software and Statistical Methods. In *Plymouth, UK*.
- Angel, M. v., & Rice, T. L. (1996). The Ecology of the Deep Ocean and Its Relevance to Global Waste Management. *The Journal of Applied Ecology*, 33(5). <https://doi.org/10.2307/2404673>
- Anschutz, P., Jorissen, F. J., Chaillou, G., Abu-Zied, R., & Fontanier, C. (2002). Recent turbidite deposition in the eastern Atlantic: Early diagenesis and biotic recovery. *Journal of Marine Research*, 60(6). <https://doi.org/10.1357/002224002321505156>
- Ardron, J. A., Simon-Lledó, E., Jones, D. O. B., & Ruhl, H. A. (2019). Detecting the Effects of Deep-Seabed Nodule Mining: Simulations Using Megafaunal Data From the Clarion-Clipperton Zone. *Frontiers in Marine Science*, 6. <https://doi.org/10.3389/fmars.2019.00604>
- Barnthouse, L. W. (2004). Quantifying population recovery rates for ecological risk assessment. *Environmental Toxicology and Chemistry*, 23(2), 500–508. <https://doi.org/10.1897/02-521>
- Bates, C. R., Scott, G., Tobin, M., & Thompson, R. (2007). Weighing the costs and benefits of reduced sampling resolution in biomonitoring studies: Perspectives from the temperate rocky intertidal. *Biological Conservation*, 137(4). <https://doi.org/10.1016/j.biocon.2007.03.019>
- Berkenbusch, K., Probert, P. K., & Nodder, S. D. (2011). Comparative biomass of sediment benthos across a depth transect, Chatham Rise, Southwest Pacific Ocean. *Marine Ecology Progress Series*, 425. <https://doi.org/10.3354/meps09014>
- Bigham, K. T., Rowden, A. A., Leduc, D., & Bowden, D. A. (2021). Review and syntheses: Impacts of turbidity flows on deep-sea benthic communities. *Biogeosciences*, 18(5), 1893–1908. <https://doi.org/10.5194/bg-18-1893-2021>
- Bigham, K. T., Rowden, A. A., Bowden, D. A., Leduc, D., Pallentin, A., Chin, C., Mountjoy, J. J., Nodder, S. D., & Orpin, A. R. (2023a). Deep-sea benthic megafauna hotspot shows indication of resilience to impact from massive turbidity flow. *Frontiers in Marine Science*, 10. <https://doi.org/10.3389/fmars.2023.1180334>
- Bigham, K. T., Rowden, A. A., Leduc, D., Bowden, D. A., Nodder, S. D., Orpin, A. R., & Halliday, J. (2023b). Deep-sea macrofauna community recovery in Kaikōura canyon following an earthquake-triggered turbidity flow. *Deep Sea Research Part I: Oceanographic Research Papers*, 202, 104192. <https://doi.org/10.1016/j.dsr.2023.104192>
- Boyd, S. E., Rees, H. L., & Richardson, C. A. (2000). Nematodes as sensitive indicators of change at dredged material disposal sites. *Estuarine, Coastal and Shelf Science*, 51(6). <https://doi.org/10.1006/ecss.2000.0722>
- Briggs, K. B., Richardson, M. D., & Young, D. K. (1996). The classification and structure of megafaunal assemblages in the Venezuela Basin, Caribbean Sea. *Journal of Marine Research*, 54(4). <https://doi.org/10.1357/0022240963213736>
- Bussau, C., Schriever, G., & Thiel, H. (1995). Evaluation of abyssal metazoan meiofauna from a manganese nodule area of the eastern South Pacific. *Vie et Milieu*, 45(1).

- Chandler, G. T., & Fleeger, J. W. (1983). Meiofaunal colonization of azoic estuarine sediment in Louisiana: Mechanisms of dispersal. *Journal of Experimental Marine Biology and Ecology*, 69(2). [https://doi.org/10.1016/0022-0981\(83\)90066-7](https://doi.org/10.1016/0022-0981(83)90066-7)
- Clarke, K. R., & Gorley, R. N. (2018). Getting started with PRIMER v7. *Primer-E*, 1.
- Clarke, K., & Green, R. (1988). Statistical design and analysis for a “biological effects” study. *Marine Ecology Progress Series*, 46, 213–226. <https://doi.org/10.3354/meps046213>
- Clarke, K. R., Chapman, M. G., Somerfield, P. J., & Needham, H. R. (2006). Dispersion-based weighting of species counts in assemblage analyses. *Marine Ecology Progress Series*, 320, 11–27. <https://doi.org/10.3354/meps320011>
- Croissant, T., Steer, P., Lague, D., Davy, P., Jeandet, L., & Hilton, R. G. (2019). Seismic cycles, earthquakes, landslides and sediment fluxes: Linking tectonics to surface processes using a reduced-complexity model. *Geomorphology*, 339, 87–103. <https://doi.org/10.1016/j.geomorph.2019.04.017>
- Dal Zotto, M., Santulli, A., Simonini, R., & Todaro, M. A. (2016). Organic enrichment effects on a marine meiofauna community, with focus on Kinorhyncha. *Zoologischer Anzeiger*, 265. <https://doi.org/10.1016/j.jcz.2016.03.013>
- de Jonge, D. S. W., Stratmann, T., Lins, L., Vanreusel, A., Purser, A., Marcon, Y., Rodrigues, C. F., Ravara, A., Esquete, P., Cunha, M. R., Simon-Lledó, E., van Breugel, P., Sweetman, A. K., Soetaert, K., & van Oevelen, D. (2020). Abyssal food-web model indicates faunal carbon flow recovery and impaired microbial loop 26 years after a sediment disturbance experiment. *Progress in Oceanography*, 189. <https://doi.org/10.1016/j.pocean.2020.102446>
- De Leo, F. C., Smith, C. R., Rowden, A. A., Bowden, D. A., & Clark, M. R. (2010). Submarine canyons: Hotspots of benthic biomass and productivity in the deep sea. *Proceedings of the Royal Society B: Biological Sciences*, 277(1695). <https://doi.org/10.1098/rspb.2010.0462>
- De Troch, M., Roelofs, M., Riedel, B., & Grego, M. (2013). Structural and functional responses of harpacticoid copepods to anoxia in the Northern Adriatic: An experimental approach. *Biogeosciences*, 10(6). <https://doi.org/10.5194/bg-10-4259-2013>
- Dellow, S., Massey, C., Cox, S., Archibald, G., Begg, J., Bruce, Z., Carey, J., Davidson, J., della Pasqua, F., Glassey, P., Hill, M., Jones, K., Lyndsell, B., Lukovic, B., McColl, S., Rattenbury, M., Read, S., Rosser, B., Singeisen, C., ... Little, M. (2017). Landslides caused by the Mw7.8 Kaikōura earthquake and the immediate response. *Bulletin of the New Zealand Society for Earthquake Engineering*, 50(2). <https://doi.org/10.5459/bnzsee.50.2.106-116>
- Etter, R. J., & Grassle, J. F. (1992). Patterns of species diversity in the deep sea as a function of sediment particle size diversity. *Nature*, 360(6404). <https://doi.org/10.1038/360576a0>
- Fernandez-Arcaya, U., Rotllant, G., Ramirez-Llodra, E., Recasens, L., Aguzzi, J., Flexas, M. M., Sanchez-Vidal, A., López-Fernández, P., García, J. A., & Company, J. B. (2013). Reproductive biology and recruitment of the deep-sea fish community from the NW Mediterranean continental margin. *Progress in Oceanography*, 118, 222–234. <https://doi.org/10.1016/j.pocean.2013.07.019>
- Folk, R. L., & Ward, W. C. (1957). Brazos River bar; a study in the significance of grain size parameters. *Journal of Sedimentary Research*, 27(1). <https://doi.org/10.1306/74d70646-2b21-11d7-8648000102c1865d>
- Folk, R. (1966). A review of grain-size parameters. *Sedimentology*, 6(1966).
- Folke, C., Carpenter, S., Walker, B., Scheffer, M., Elmqvist, T., Gunderson, L., & Holling, C. S. (2004). Regime shifts, resilience, and biodiversity in ecosystem management. In *Annual Review of Ecology, Evolution, and Systematics* (Vol. 35). <https://doi.org/10.1146/annurev.ecolsys.35.021103.105711>
- Frutos, I., & Sorbe, J. C. (2017). Suprabenthic assemblages from the Capbreton area (SE Bay of Biscay). Faunal recovery after a canyon turbidity disturbance. *Deep-Sea Research Part I: Oceanographic Research Papers*, 130. <https://doi.org/10.1016/j.dsr.2017.10.007>
- Galéron, J., Menot, L., Renaud, N., Crassous, P., Khripounoff, A., Treignier, C., & Sibuet, M. (2009). Spatial and temporal patterns of benthic macrofaunal communities on the deep continental margin in the Gulf of Guinea. *Deep-Sea Research Part II: Topical Studies in Oceanography*, 56(23), 2299–2312. <https://doi.org/10.1016/j.dsr2.2009.04.011>
- Gallucci, F., Moens, T., Vanreusel, A., & Fonseca, G. (2008). Active colonisation of disturbed sediments by deep-sea nematodes: Evidence for the patch mosaic model. *Marine Ecology Progress Series*, 367. <https://doi.org/10.3354/meps07537>

- Gibbs, M., Leduc, D., Nodder, S. D., Kingston, A., Swales, A., Rowden, A. A., Mountjoy, J., Olsen, G., Ovenden, R., Brown, J., Bury, S., & Graham, B. (2020). Novel Application of a Compound-Specific Stable Isotope (CSSI) Tracking Technique Demonstrates Connectivity Between Terrestrial and Deep-Sea Ecosystems via Submarine Canyons. *Frontiers in Marine Science*, 7. <https://doi.org/10.3389/fmars.2020.00608>
- Giere, O. (2008). Meiobenthology: The microscopic motile fauna of aquatic sediments. In *Meiobenthology: The Microscopic Motile Fauna of Aquatic Sediments*. <https://doi.org/10.1007/978-3-540-68661-3>
- Gollner, S., Miljutina, M., & Bright, M. (2013). Nematode Succession at deep-sea hydrothermal vents after a recent volcanic eruption with the description of two dominant species. *Organisms Diversity and Evolution*, 13(3). <https://doi.org/10.1007/s13127-012-0122-2>
- Griggs, G. B., Carey, A. G., & Kulm, L. D. (1969). Deep-sea sedimentation and sediment-fauna interaction in Cascadia Channel and on Cascadia Abyssal Plain. *Deep-Sea Research and Oceanographic Abstracts*, 16(2). [https://doi.org/10.1016/0011-7471\(69\)90071-0](https://doi.org/10.1016/0011-7471(69)90071-0)
- Guerra, M., Dawson, S., Sabadel, A., Slooten, E., Somerford, T., Williams, R., Wing, L., & Rayment, W. (2020). Changes in habitat use by a deep-diving predator in response to a coastal earthquake. *Deep-Sea Research Part I: Oceanographic Research Papers*, 158. <https://doi.org/10.1016/j.dsr.2020.103226>
- Haughton, P. D. W., Barker, S. P., & McCaffrey, W. D. (2003). "Linked" debrites in sand-rich turbidite systems - Origin and significance. *Sedimentology*, 50(3), 459–482. <https://doi.org/10.1046/j.1365-3091.2003.00560.x>
- Heip, C., Vincx, M., & Vranken, G. (1985). The Ecology of Marine Nematodes. *Oceanography and Marine Biology: An Annual Review*, 23, 399–489.
- Hess, S., & Jorissen, F. J. (2009). Distribution patterns of living benthic foraminifera from Cap Breton canyon, Bay of Biscay: Faunal response to sediment instability. *Deep Sea Research Part I: Oceanographic Research Papers*, 56(9), 1555–1578. <https://doi.org/10.1016/j.dsr.2009.04.003>
- Hess, S., Jorissen, F. J., Venet, V., & Abu-Zied, R. (2005). Benthic foraminiferal recovery after recent turbidite deposition in cap breton canyon, bay of biscay. *Journal of Foraminiferal Research*, 35(2). <https://doi.org/10.2113/35.2.114>
- Higgins, R., & Thiel, H. (1988). *Introduction to the Study of Meiofauna*. Smithsonian Institution Press.
- Holling, C. S. (1996). Engineering resilience versus ecological resilience. In *Engineering within ecological constraints* (Issue 1996).
- Ingels, J., Vanreusel, A., Pape, E., Pasotti, F., Macheriotou, L., Arbizu, P. M., Sørensen, M. V., Edgcomb, V. P., Sharma, J., Sánchez, N., Homoky, W. B., Woulds, C., Leduc, D., Gooday, A. J., Pawlowski, J., Dolan, J. R., Schratzberger, M., Gollner, S., Schoenle, A., ... Zeppilli, D. (2020). Ecological variables for deep-ocean monitoring must include microbiota and meiofauna for effective conservation. *Nature Ecology & Evolution*, 5(1). <https://doi.org/10.1038/s41559-020-01335-6>
- Kawagucci, S., Yoshida, Y. T., Noguchi, T., Honda, M. C., Uchida, H., Ishibashi, H., Nakagawa, F., Tsunogai, U., Okamura, K., Takaki, Y., Nunoura, T., Miyazaki, J., Hirai, M., Lin, W., Kitazato, H., & Takai, K. (2012). Disturbance of deep-sea environments induced by the M9.0 Tohoku Earthquake. *Scientific Reports*, 2, 1–7. <https://doi.org/10.1038/srep00270>
- Keil, R. G., Montluçon, D. B., Prah, F. G., & Hedges, J. I. (1994). Sorptive preservation of labile organic matter in marine sediments. *Nature*, 370(6490). <https://doi.org/10.1038/370549a0>
- Kenny, A. J., & Rees, H. L. (1994). The effects of marine gravel extraction on the macrobenthos: Early post-dredging recolonization. *Marine Pollution Bulletin*, 28(7). [https://doi.org/10.1016/0025-326X\(94\)90130-9](https://doi.org/10.1016/0025-326X(94)90130-9)
- Kioka, A., Schwestermann, T., Moernaut, J., Ikehara, K., Kanamatsu, T., McHugh, C. M., dos Santos Ferreira, C., Wiemer, G., Haghipour, N., Kopf, A. J., Eglinton, T. I., & Strasser, M. (2019). Megathrust earthquake drives drastic organic carbon supply to the hadal trench. *Scientific Reports*, 9(1), 1–10. <https://doi.org/10.1038/s41598-019-38834-x>
- Kitahashi, T., Jenkins, R. G., Nomaki, H., Shimanaga, M., Fujikura, K., & Kojima, S. (2014). Effect of the 2011 Tohoku Earthquake on deep-sea meiofaunal assemblages inhabiting the landward slope of the Japan Trench. *Marine Geology*, 358. <https://doi.org/10.1016/j.margeo.2014.05.004>
- Kitahashi, T., Watanabe, H., Ikehara, K., Jenkins, R. G., Kojima, S., & Shimanaga, M. (2016). Deep-sea meiofauna off the Pacific coast of Tohoku and other trench slopes around Japan: a comparative

- study before and after the 2011 off the Pacific coast of Tohoku Earthquake. *Journal of Oceanography*, 72(1), 129–139. <https://doi.org/10.1007/s10872-015-0323-3>
- Kitahashi, T., Jenkins, R. G., Kojima, S., & Shimanaga, M. (2018). High resilience of harpacticoid copepods in the landward slope of the Japan Trench against disturbance of the 2011 Tohoku Earthquake. *Limnology and Oceanography*, 63(6). <https://doi.org/10.1002/lno.11006>
- Kuenen, PH. H., & Migliorini, C. I. (1950). Turbidity currents as a cause of graded bedding. *The Journal of Geology*, 58(2), 91–127.
- Lambshead, P. J. D., Tietjen, J., Glover, A., Ferrero, T., Thistle, D., & Gooday, A. J. (2001). Impact of large-scale natural physical disturbance on the diversity of deep-sea North Atlantic nematodes. *Marine Ecology Progress Series*, 214. <https://doi.org/10.3354/meps214121>
- Lasiak, T. (2003). Influence of taxonomic resolution, biological attributes and data transformations on multivariate comparisons of rocky macrofaunal assemblages. *Marine Ecology Progress Series*, 250. <https://doi.org/10.3354/meps250029>
- Leduc, D., Rowden, A. A., Nodder, S. D., Berkenbusch, K., Probert, P. K., & Hadfield, M. G. (2014). Unusually high food availability in Kaikoura Canyon linked to distinct deep-sea nematode community. *Deep-Sea Research Part II: Topical Studies in Oceanography*, 104. <https://doi.org/10.1016/j.dsr2.2013.06.003>
- Leduc, D., & Pilditch, C. A. (2013). Effect of a physical disturbance event on deep-sea nematode community structure and ecosystem function. *Journal of Experimental Marine Biology and Ecology*, 440. <https://doi.org/10.1016/j.jembe.2012.11.015>
- Leduc, D., Nodder, S. D., Rowden, A. A., Gibbs, M., Berkenbusch, K., Wood, A., de Leo, F., Smith, C., Brown, J., Bury, S. J., & Pallentin, A. (2020). Structure of infaunal communities in New Zealand submarine canyons is linked to origins of sediment organic matter. *Limnology and Oceanography*, 65(10). <https://doi.org/10.1002/lno.11454>
- Leduc, D., Rowden, A. A., Probert, P. K., Pilditch, C. A., Nodder, S. D., Vanreusel, A., Duineveld, G. C. A., & Witbaard, R. (2012). Further evidence for the effect of particle-size diversity on deep-sea benthic biodiversity. *Deep-Sea Research Part I: Oceanographic Research Papers*, 63. <https://doi.org/10.1016/j.dsr.2011.10.009>
- Leduc, D., Rowden, A. A., Bowden, D. A., Probert, P. K., Pilditch, C. A., & Nodder, S. D. (2012). Unimodal relationship between biomass and species richness of deep-sea nematodes: Implications for the link between productivity and diversity. *Marine Ecology Progress Series*, 454. <https://doi.org/10.3354/meps09609>
- Lee, H., Gerdes, D., Vanhove, S., & Vincx, M. (2001). Meiofauna response to iceberg disturbance on the Antarctic continental shelf at Kapp Norvegia (Weddell Sea). *Polar Biology*, 24(12). <https://doi.org/10.1007/s003000100301>
- Levin, L. A. (1991). Interactions between metazoans and large, agglutinating protozoans: Implications for the community structure of deep-sea benthos. *Integrative and Comparative Biology*, 31(6). <https://doi.org/10.1093/icb/31.6.886>
- Lewis, K. B. (1994). The 1500-km-long Hikurangi Channel: Trench-axis channel that escapes its trench, crosses a plateau, and feeds a fan drift. *Geo-Marine Letters*, 14(1). <https://doi.org/10.1007/BF01204467>
- Lewis, K. B., & Barnes, P. M. (1999). Kaikoura Canyon, New Zealand: Active conduit from near-shore sediment zones to trench-axis channel. *Marine Geology*, 162(1). [https://doi.org/10.1016/S0025-3227\(99\)00075-4](https://doi.org/10.1016/S0025-3227(99)00075-4)
- Liao, J. X., Wei, C. L., & Yasuhara, M. (2020). Species and Functional Diversity of Deep-Sea Nematodes in a High Energy Submarine Canyon. *Frontiers in Marine Science*, 7. <https://doi.org/10.3389/fmars.2020.00591>
- Lowe, D. R. (1979). Sediment gravity flows: their classification and some problems of application to natural flows and deposits. *Geology of Continental Slopes*. <https://doi.org/10.2110/pec.79.27.0075>
- Lundquist, C. J., Thrush, S. F., Coco, G., & Hewitt, J. E. (2010). Interactions between disturbance and dispersal reduce persistence thresholds in a benthic community. *Marine Ecology Progress Series*, 413. <https://doi.org/10.3354/meps08578>
- Massey, C. I., Townsend, D., Jones, K., Lukovic, B., Rhoades, D., Morgenstern, R., Rosser, B., Ries, W., Howarth, J., Hamling, I., Petley, D., Clark, M., Wartman, J., Litchfield, N., & Olsen, M. (2020). Volume Characteristics of Landslides Triggered by the MW 7.8 2016 Kaikōura Earthquake, New

- Zealand, Derived From Digital Surface Difference Modeling. *Journal of Geophysical Research: Earth Surface*, 125(7). <https://doi.org/10.1029/2019JF005163>
- Mayer, L. M. (1994). Surface area control of organic carbon accumulation in continental shelf sediments. *Geochimica et Cosmochimica Acta*, 58(4). [https://doi.org/10.1016/0016-7037\(94\)90381-6](https://doi.org/10.1016/0016-7037(94)90381-6)
- McClanahan, T. R., Graham, N. A. J., Calnan, J. M., & MacNeil, M. A. (2007). Toward pristine biomass: Reef fish recovery in coral reef marine protected areas in Kenya. *Ecological Applications*, 17(4). <https://doi.org/10.1890/06-1450>
- Miljutin, D. M., Gad, G., Miljutina, M. M., Mokievsky, V. O., Fonseca-Genevois, V., & Esteves, A. M. (2010). The state of knowledge on deep-sea nematode taxonomy: How many valid species are known down there? In *Marine Biodiversity* (Vol. 40, Issue 3). <https://doi.org/10.1007/s12526-010-0041-4>
- Miljutin, D. M., Miljutina, M. A., Arbizu, P. M., & Galéron, J. (2011). Deep-sea nematode assemblage has not recovered 26 years after experimental mining of polymetallic nodules (Clarion-Clipperton Fracture Zone, Tropical Eastern Pacific). *Deep-Sea Research Part I: Oceanographic Research Papers*, 58(8), 885–897. <https://doi.org/10.1016/j.dsr.2011.06.003>
- Milliman, J. D. (1994). Organic matter content in U.S. Atlantic continental slope sediments: Decoupling the grain-size factor. *Deep-Sea Research Part II*, 41(4–6). [https://doi.org/10.1016/0967-0645\(94\)90049-3](https://doi.org/10.1016/0967-0645(94)90049-3)
- Mirto, S., Gristina, M., Sinopoli, M., Maricchiolo, G., Genovese, L., Vizzini, S., & Mazzola, A. (2012). Meiofauna as an indicator for assessing the impact of fish farming at an exposed marine site. *Ecological Indicators*, 18. <https://doi.org/10.1016/j.ecolind.2011.12.015>
- Moreno, M., Vezzulli, L., Marin, V., Laconi, P., Albertelli, G., & Fabiano, M. (2008). The use of meiofauna diversity as an indicator of pollution in harbours. *ICES Journal of Marine Science*, 65(8). <https://doi.org/10.1093/icesjms/fsn116>
- Mountjoy, J. J., Howarth, J. D., Orpin, A. R., Barnes, P. M., Bowden, D. A., Rowden, A. A., Schimel, A. C. G., Holden, C., Horgan, H. J., Nodder, S. D., Patton, J. R., Lamarche, G., Gerstenberger, M., Micallef, A., Pallentin, A., & Kane, T. (2018). Earthquakes drive large-scale submarine canyon development and sediment supply to deep-ocean basins. *Science Advances*, 4(3). <https://doi.org/10.1126/sciadv.aar3748>
- Muñoz-Royo, C., Ouillon, R., el Mousadik, S., Alford, M. H., & Peacock, T. (2022). An in situ study of abyssal turbidity-current sediment plumes generated by a deep seabed polymetallic nodule mining preprototype collector vehicle. *Science Advances*, 8(38). <https://doi.org/10.1126/sciadv.abn1219>
- Murrell, M. C., & Fleeger, J. W. (1989). Meiofauna abundance on the Gulf of Mexico continental shelf affected by hypoxia. *Continental Shelf Research*, 9(12). [https://doi.org/10.1016/0278-4343\(89\)90057-5](https://doi.org/10.1016/0278-4343(89)90057-5)
- Musco, L., Mikac, B., Tataranni, M., Giangrande, A., & Terlizzi, A. (2011). The use of coarser taxonomy in the detection of long-term changes in polychaete assemblages. *Marine Environmental Research*, 71(2). <https://doi.org/10.1016/j.marenvres.2010.12.004>
- Neira, C., Sellanes, J., Levin, L. A., & Arntz, W. E. (2001). Meiofaunal distributions on the Peru margin: Relationship to oxygen and organic matter availability. *Deep-Sea Research Part I: Oceanographic Research Papers*, 48(11). [https://doi.org/10.1016/S0967-0637\(01\)00018-8](https://doi.org/10.1016/S0967-0637(01)00018-8)
- Nomaki, H., Mochizuki, T., Kitahashi, T., Nunoura, T., Arai, K., Toyofuku, T., Tanaka, G., Shigeno, S., Tasumi, E., Fujikura, K., & Watanabe, S. (2016). Effects of mass sedimentation events after the 2011 off the Pacific coast of Tohoku Earthquake on benthic prokaryotes and meiofauna inhabiting the upper bathyal sediments. *Journal of Oceanography*, 72(1). <https://doi.org/10.1007/s10872-015-0293-5>
- Oguri, K., Kawamura, K., Sakaguchi, A., Toyofuku, T., Kasaya, T., Murayama, M., Fujikura, K., Glud, R. N., & Kitazato, H. (2013). Hadal disturbance in the Japan Trench induced by the 2011 Tohoku-Oki earthquake. *Scientific Reports*, 3, 1–6. <https://doi.org/10.1038/srep01915>
- Okey, T. A. (1997). Sediment flushing observations, earthquake slumping, and benthic community changes in Monterey Canyon head. *Continental Shelf Research*, 17(8). [https://doi.org/10.1016/S0278-4343\(96\)00067-2](https://doi.org/10.1016/S0278-4343(96)00067-2)
- Olsgard, F., Brattegard, T., & Holthe, T. (2003). Polychaetes as surrogates for marine biodiversity: Lower taxonomic resolution and indicator groups. *Biodiversity and Conservation*, 12(5). <https://doi.org/10.1023/A:1022800405253>

- 947 Paull, C. K., Talling, P. J., Maier, K. L., Parsons, D., Xu, J., Caress, D. W., Gwiazda, R., Lundsten, E. M.,
948 Anderson, K., Barry, J. P., Chaffey, M., O'Reilly, T., Rosenberger, K. J., Gales, J. A., Kieft, B.,
949 McGann, M., Simmons, S. M., McCann, M., Sumner, E. J., ... Cartigny, M. J. (2018). Powerful
950 turbidity currents driven by dense basal layers. *Nature Communications*, 9(1).
951 <https://doi.org/10.1038/s41467-018-06254-6>
- 952 Ptatscheck, C., & Traunspurger, W. (2020). The ability to get everywhere: dispersal modes of free-living,
953 aquatic nematodes. *Hydrobiologia*, 847(17), 3519–3547. [https://doi.org/10.1007/s10750-020-](https://doi.org/10.1007/s10750-020-04373-0)
954 [04373-0](https://doi.org/10.1007/s10750-020-04373-0)
- 955 Pusceddu, A., Mea, M., Canals, M., Heussner, S., Durrieu De Madron, X., Sanchez-Vidal, A., Bianchelli,
956 S., Corinaldesi, C., Dell'Anno, A., Thomsen, L., & Danovaro, R. (2013). Major consequences of
957 an intense dense shelf water cascading event on deep-sea benthic trophic conditions and
958 meiofaunal biodiversity. *Biogeosciences*, 10(4), 2659–2670. [https://doi.org/10.5194/bg-10-2659-](https://doi.org/10.5194/bg-10-2659-2013)
959 [2013](https://doi.org/10.5194/bg-10-2659-2013)
- 960 Qian, P. Y. (1999). Larval settlement of polychaetes. *Hydrobiologia*, 402.
961 <https://doi.org/10.1023/A:1003704928668>
- 962 R Core Team. (2022). *R: A language and environment for statistical computing*. R Foundation for
963 *Statistical Computing*.
- 964 Raes, M., Rose, A., & Vanreusel, A. (2010). Response of nematode communities after large-scale ice-
965 shelf collapse events in the Antarctic Larsen area. *Global Change Biology*, 16(5).
966 <https://doi.org/10.1111/j.1365-2486.2009.02137.x>
- 967 Richardson, M. D., Briggs, K. B., & Young, D. K. (1985). Effects of biological activity by abyssal benthic
968 macroinvertebrates on a sedimentary structure in the Venezuela Basin. *Marine Geology*, 68(1–4).
969 [https://doi.org/10.1016/0025-3227\(85\)90015-5](https://doi.org/10.1016/0025-3227(85)90015-5)
- 970 Richardson, M. D., & Young, D. K. (1987). Abyssal benthos of the Venezuela Basin, Caribbean Sea:
971 standing stock considerations. *Deep Sea Research Part A, Oceanographic Research Papers*,
972 34(2). [https://doi.org/10.1016/0198-0149\(87\)90079-3](https://doi.org/10.1016/0198-0149(87)90079-3)
- 973 Román, S., Vanreusel, A., Romano, C., Ingels, J., Puig, P., Company, J. B., & Martin, D. (2016). High
974 spatiotemporal variability in meiofaunal assemblages in Blanes Canyon (NW Mediterranean)
975 subject to anthropogenic and natural disturbances. *Deep-Sea Research Part I: Oceanographic*
976 *Research Papers*, 117. <https://doi.org/10.1016/j.dsr.2016.10.004>
- 977 Schratzberger, M., & Jennings, S. (2002). Impacts of chronic trawling disturbance on meiofaunal
978 communities. *Marine Biology*, 141(5). <https://doi.org/10.1007/s00227-002-0895-5>
- 979 Sciberras, M., Hiddink, J. G., Jennings, S., Szostek, C. L., Hughes, K. M., Kneafsey, B., Clarke, L. J.,
980 Ellis, N., Rijnsdorp, A. D., McConnaughey, R. A., Hilborn, R., Collie, J. S., Pitcher, C. R.,
981 Amoroso, R. O., Parma, A. M., Suuronen, P., & Kaiser, M. J. (2018). Response of benthic fauna
982 to experimental bottom fishing: A global meta-analysis. *Fish and Fisheries*, 19(4).
983 <https://doi.org/10.1111/faf.12283>
- 984 Semprucci, F., Losi, V., & Moreno, M. (2015). A review of Italian research on free-living marine
985 nematodes and the future perspectives on their use as Ecological Indicators (Ecolnds). In
986 *Mediterranean Marine Science* (Vol. 16, Issue 2). <https://doi.org/10.12681/mms.1072>
- 987 Smith, S. D. A., & Simpson, R. D. (1993). Effects of pollution on holdfast macrofauna of the kelp *Ecklonia*
988 *radiata*: discrimination at different taxonomic levels. *Marine Ecology Progress Series*, 96(2).
989 <https://doi.org/10.3354/meps096199>
- 990 Smith, C. R., Levin, L. A., Koslow, A., Tyler, P. A., & Glover, A. G. (2008). The near future of the deep-sea
991 floor ecosystems. In *Aquatic Ecosystems: Trends and Global Prospects*.
992 <https://doi.org/10.1017/CBO9780511751790.030>
- 993 Solé, R. v., Saldaña, J., Montoya, J. M., & Erwin, D. H. (2010). Simple model of recovery dynamics after
994 mass extinction. *Journal of Theoretical Biology*, 267(2). <https://doi.org/10.1016/j.jtbi.2010.08.015>
- 995 Somerfield, P. J., & Warwick, R. M. (1996). Meiofauna in marine pollution monitoring programmes. A
996 laboratory manual. . *Directorate of Fisheries Research(MAFF), Lowestoft(UK)*.
- 997 Sousa, W. P. (1984). The Role of Disturbance in Natural Communities. *Annual Review of Ecology and*
998 *Systematics*, 15(1), 353–391. <https://doi.org/10.1146/annurev.es.15.110184.002033>
- 999 Sousa, W. P. (2001). Natural Disturbance and the Dynamics of Marine Benthic Communities. In M. D.
1000 Bertness, S. D. Gaines, & M. E. Hay (Eds.), *Marine Community Ecology* (pp. 85–130). Sinauer
1001 Associates, Inc.

- Szymelfenig, M., Kotwicki, L., & Graca, B. (2006). Benthic re-colonization in post-dredging pits in the Puck Bay (Southern Baltic Sea). *Estuarine, Coastal and Shelf Science*, 68(3–4). <https://doi.org/10.1016/j.ecss.2006.02.018>
- Talling, P. J., Amy, L. A., Wynn, R. B., Blackburn, G., & Gibson, O. (2007). Evolution of turbidity currents deduced from extensive thin turbidites: Marnoso Arenacea Formation (Miocene), Italian Apennines. *Journal of Sedimentary Research*, 77(3–4). <https://doi.org/10.2110/jsr.2007.018>
- Talling, P. J. (2014). On the triggers, resulting flow types and frequencies of subaqueous sediment density flows in different settings. *Marine Geology*, 352, 155–182. <https://doi.org/10.1016/j.margeo.2014.02.006>
- Thiel, H., Schriever, G., Bussau, C., & Borowski, C. (1993). Manganese nodule crevice fauna. *Deep-Sea Research Part I*, 40(2). [https://doi.org/10.1016/0967-0637\(93\)90012-R](https://doi.org/10.1016/0967-0637(93)90012-R)
- Thiel, H., Schriever, G., Ahnert, A., Bluhm, H., Borowski, C., & Vopel, K. (2001). The large-scale environmental impact experiment DISCOL - Reflection and foresight. *Deep-Sea Research Part II: Topical Studies in Oceanography*, 48(17–18), 3869–3882. [https://doi.org/10.1016/S0967-0645\(01\)00071-6](https://doi.org/10.1016/S0967-0645(01)00071-6)
- Thomsen, M. S., Metcalfe, I., Siciliano, A., South, P. M., Gerrity, S., Alestra, T., & Schiel, D. R. (2020). Earthquake-driven destruction of an intertidal habitat cascade. *Aquatic Botany*, 164. <https://doi.org/10.1016/j.aquabot.2020.103217>
- Thurston, M. H., Bett, B. J., Rice, A. L., & Jackson, P. A. B. (1994). Variations in the invertebrate abyssal megafauna in the North Atlantic Ocean. *Deep-Sea Research Part I*, 41(9). [https://doi.org/10.1016/0967-0637\(94\)90100-7](https://doi.org/10.1016/0967-0637(94)90100-7)
- Thurston, M. H., Rice, A. L., & Bett, B. J. (1998). Latitudinal variation in invertebrate megafaunal abundance and biomass in the North Atlantic Ocean Abyss. *Deep Sea Research Part II: Topical Studies in Oceanography*, 45(1–3), 203–224. [https://doi.org/10.1016/S0967-0645\(97\)00077-5](https://doi.org/10.1016/S0967-0645(97)00077-5)
- Tietjen, J. H. (1976). Distribution and species diversity of deep-sea nematodes off North Carolina. *Deep Sea Research and Oceanographic Abstracts*, 23(8). [https://doi.org/10.1016/s0011-7471\(76\)80018-6](https://doi.org/10.1016/s0011-7471(76)80018-6)
- Tsujimoto, A., Nomura, R., Arai, K., Nomaki, H., Inoue, M., & Fujikura, K. (2020). Changes in deep-sea benthic foraminiferal fauna caused by turbidites deposited after the 2011 Tohoku-oki earthquake. *Marine Geology*, 419. <https://doi.org/10.1016/j.margeo.2019.106045>
- Tyler, P. A. (2003). Disposal in the deep sea: Analogue of nature or faux ami? *Environmental Conservation*, 30(1). <https://doi.org/10.1017/S037689290300002X>
- van Gaever, S., Galéron, J., Sibuet, M., & Vanreusel, A. (2009). Deep-sea habitat heterogeneity influence on meiofaunal communities in the Gulf of Guinea. *Deep-Sea Research Part II: Topical Studies in Oceanography*, 56(23). <https://doi.org/10.1016/j.dsr2.2009.04.008>
- Vanaverbeke, J., Soetaert, K., Heip, C., & Vanreusel, A. (1997). The metazoan meiobenthos along the continental slope of the Goban Spur (NE Atlantic). *Journal of Sea Research*, 38(1–2). [https://doi.org/10.1016/S1385-1101\(97\)00038-5](https://doi.org/10.1016/S1385-1101(97)00038-5)
- Vanreusel, A. (1990). Ecology of the Free-Living Marine Nematodes from the Voordelta (Southern Bight of the North-Sea) .1. Species Composition and Structure of the Nematode Communities. *Cahiers De Biologie Marine*, 31(4).
- Vanreusel, A., Clough, L., Jacobsen, K., Ambrose, W., Jutamas Jivaluk, Ryheul, V., Herman, R., & Vincx, M. (2000). Meiobenthos of the central Arctic Ocean with special emphasis on the nematode community structure. *Deep-Sea Research Part I: Oceanographic Research Papers*, 47(10). [https://doi.org/10.1016/S0967-0637\(00\)00007-8](https://doi.org/10.1016/S0967-0637(00)00007-8)
- Veillette, J., Sarrazin, J., Gooday, A. J., Galéron, J., Caprais, J. C., Vangriesheim, A., Étoubleau, J., Christian, J. R., & Kim Juniper, S. (2007). Ferromanganese nodule fauna in the Tropical North Pacific Ocean: Species richness, faunal cover and spatial distribution. *Deep-Sea Research Part I: Oceanographic Research Papers*, 54(11). <https://doi.org/10.1016/j.dsr.2007.06.011>
- Veillette, J., Juniper, S. K., Gooday, A. J., & Sarrazin, J. (2007). Influence of surface texture and microhabitat heterogeneity in structuring nodule faunal communities. *Deep-Sea Research Part I: Oceanographic Research Papers*, 54(11). <https://doi.org/10.1016/j.dsr.2007.06.012>
- Walker, B., Holling, C. S., Carpenter, S. R., & Kinzig, A. (2004). Resilience, adaptability and transformability in social-ecological systems. *Ecology and Society*, 9(2). <https://doi.org/10.5751/ES-00650-090205>

- Warwick, R. M. (1988). The level of taxonomic discrimination required to detect pollution effects on marine benthic communities. *Marine Pollution Bulletin*, 19(6). [https://doi.org/10.1016/0025-326X\(88\)90596-6](https://doi.org/10.1016/0025-326X(88)90596-6)
- Warwick, R. M., & Clarke, K. R. (1993). Increased variability as a symptom of stress in marine communities. *Journal of Experimental Marine Biology and Ecology*, 172(1–2). [https://doi.org/10.1016/0022-0981\(93\)90098-9](https://doi.org/10.1016/0022-0981(93)90098-9)
- Willig, M. R., & Walker, L. R. (1999). Disturbance in terrestrial ecosystems: salient themes, synthesis, and future directions. *Ecosystems of the World*, 747–768.
- Woods, D. R., & Tietjen, J. H. (1985). Horizontal and vertical distribution of meiofauna in the Venezuela Basin. *Marine Geology*, 68(1–4). [https://doi.org/10.1016/0025-3227\(85\)90014-3](https://doi.org/10.1016/0025-3227(85)90014-3)
- Young, D. K., & Richardson, M. D. (1998). Effects of waste disposal on benthic faunal succession on the abyssal seafloor. *Journal of Marine Systems*, 14(3–4), 319–336. [https://doi.org/10.1016/S0924-7963\(97\)00033-X](https://doi.org/10.1016/S0924-7963(97)00033-X)
- Zeppilli, D., Sarrazin, J., Leduc, D., Arbizu, P. M., Fontaneto, D., Fontanier, C., Gooday, A. J., Kristensen, R. M., Ivanenko, V. N., Sørensen, M. v., Vanreusel, A., Thébault, J., Mea, M., Allio, N., Andro, T., Arvigo, A., Castrec, J., Danielo, M., Foulon, V., ... Fernandes, D. (2015). Is the meiofauna a good indicator for climate change and anthropogenic impacts? *Marine Biodiversity*, 45(3). <https://doi.org/10.1007/s12526-015-0359-z>
- Zeppilli, D., Bongiorno, L., Cattaneo, A., Danovaro, R., & Santos, R. S. (2013). Meiofauna assemblages of the Condor Seamount (North-East Atlantic Ocean) and adjacent deep-sea sediments. *Deep-Sea Research Part II: Topical Studies in Oceanography*, 98(PA). <https://doi.org/10.1016/j.dsr2.2013.08.009>

Table 1 (on next page)

Multicore sampling site details, including depth ranges and number of cores used for meiofauna and sediment analyses

Time Point	Date	Voyage	Station Number	Site	Depth (m)	Latitude	Longitude	Number of cores for meiofauna	Number of cores for sediment	Reference
10 years before	November 2006	TAN0616	98	K04	1061	-42.512	173.633	2	-	This study
			105	K05	1020	-42.523	173.621	2	-	This study
6 years before	May 2010	TAN1006	6	K13	404	-42.490	173.551	3	1	Leduc et al. 2020
			4	K01	1017	-42.484	173.615	2	1	Leduc et al. 2020
			3	K02	989	-42.524	173.613	2	1	Leduc et al. 2020
			14	K03	1032	-42.504	173.619	2	1	Leduc et al. 2020
			7	K04	1061	-42.508	173.633	2	1	Leduc et al. 2020
			8	K05	1127	-42.492	173.657	2	1	Leduc et al. 2020
			2	K06	1289	-42.520	173.712	3	1	Leduc et al. 2020
			11	K07	1320	-42.524	173.736	2	1	Leduc et al. 2020
10 weeks after	February 2017	TAN1701	181	K02	1186	-42.492	173.653	2	1	This study
			182	K03	1036	-42.501	173.625	2	1	This study
10 months after	September 2017	TAN1708	130, 131	K13	422	-42.490	173.551	1	1	This study
			127	K01	994	-42.485	173.615	1	1	This study
			6	K02	1188	-42.492	173.653	1	1	This study
			12, 11	K03	1000	-42.502	173.622	1	1	This study
			16	K04	1069	-42.510	173.632	1	1	This study
			28	K05	1014	-42.524	173.613	1	1	This study
			75	K06	1230	-42.520	173.712	1	1	This study
			70	K07	1298	-42.525	173.725	1	1	This study

4 years after	October 2020	TAN2011	79	K13	425	-42.490	173.551	1	1	This study
			58	K01	1048	-42.485	173.615	1	1	This study
			38	K02	1190	-42.492	173.653	1	1	This study
			35	K03	1049	-42.502	173.622	1	1	This study
			47	K04	1068	-42.510	173.632	1	1	This study
			50	K05	1015	-42.524	173.613	1	1	This study
			86	K06	1293	-42.520	173.712	1	1	This study
			83	K07	1312	-42.525	173.725	1	1	This study

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Table 2(on next page)

Results of the main and pair-wise PERMANOVA tests for differences between time points for meiofauna community structure

		Pseudo-F/t	P(perm)	Permutations
Main		10.545	0.0001	9950
Pair-wise	Before, 10 weeks after	4.428	0.0176	66
	Before, 10 months after	3.991	0.0002	8875
	Before, 4 years after	1.293	0.1703	8874
	10 weeks after, 10 months after	0.751	0.6424	45
	10 weeks after, 4 years after	4.719	0.0207	45
	10 months after, 4 years after	3.488	0.0005	5086

Table 3(on next page)

SIMPER analysis results for the meiofauna community for each time point

SIMPER analysis results for the meiofauna community indicating average within time point community similarity and the contribution of individual taxa contributing 70% or more to within time point community similarity. Avg. Abundance is the average abundance of individuals standardized to 10 cm² of seafloor area.

Time point	Average similarity	Taxon	% Contribution	Avg. Abundance
Before (10 and 6 years)	79.44	Nematodes	71.94	2285.80
10 weeks after	74.46	Nematodes	66.08	156.75
		Copepods	15.89	15.21
10 months after	59.28	Nematodes	76.69	367.49
4 years after	81.57	Nematodes	69.32	2027.70
		Nauplii	8.98	55.20

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Table 4(on next page)

SIMPER analysis results for the nematode community for each time point

SIMPER analysis results for the nematode community indicating average within time point community similarity and the contribution of individual species contributing 70% or more to within time point community similarity. Avg. Abundance is the average abundance of individuals standardized to 10 cm² of seafloor area.

Time point	Average similarity	Species	% Contribution	Avg. Abundance
Before (6 years)	46.11	<i>Hopperia beaglense</i>	12.99	178.22
		<i>Cervonema</i>		
		<i>kaikouraensis</i>	10.74	126.56
		<i>Campylaimus</i> sp. 6	8.32	96.83
		<i>Leptolaimus</i> sp. 14	8.32	78.50
		<i>Sabatieria</i> sp. A	8.22	354.57
		<i>Daptonema</i> sp. 18	7.12	83.72
		<i>Metalinhomoeus</i> sp. 1	7.12	50.84
		<i>Sabatieria</i> sp. 12	7.12	98.80
		<i>Sphaerolaimus</i> sp. 1	7.12	59.60
10 weeks after	42.77	<i>Daptonema</i> sp. 18	19.26	20.43
		<i>Sabatieria</i> sp. A	15.79	11.22
		<i>Metacyatholaimus</i> sp. 1	7.81	4.08
		<i>Cervonema</i>		
		<i>kaikouraensis</i>	7.12	1.85
		<i>Monhysteridae</i> sp. 35	7.12	1.51
		<i>Vasostoma hexodontium</i>	7.12	1.51
		<i>Daptonema</i> sp. 23	3.87	4.45
		<i>Paramonohystera</i> sp. 1	3.87	2.72
10 months after	22.35	<i>Daptonema</i> sp. 18	27.25	242.42
		<i>Sabatieria</i> sp. 12	23.03	8.41
		<i>Daptonema</i> sp. 21	14.56	18.23
		<i>Sabatieria</i> sp. A	14.56	18.23
4 years after	47.77	<i>Daptonema</i> sp. 18	30.52	1012.51
		<i>Sabatieria</i> sp. A	13.82	273.57
		<i>Microlaimus</i> sp. 34	10.27	70.90
		<i>Cervonema</i>		
		<i>kaikouraensis</i>	8.38	51.98
		<i>Daptonema</i> sp. 21	8.38	44.76

Table 5(on next page)

SIMPER analysis results for the meiofauna community between time points

SIMPER analysis results for the meiofauna community indicating average among time point community dissimilarity and the contribution of individual taxa contributing 70% of more to among time point community dissimilarity. Avg. Abundance is the average abundance of individuals standardized to 10 cm² of seafloor area.

Time points	Average dissimilarity	Taxon	% Contribution	Time 1 avg. abundance	Time 2 avg. abundance
Before (10 and 6 years), 10 weeks after	57.75	Nematodes	62.57	2285.80	156.75
		Kinorhynchs	1.76	43.69	2.46
Before (10 and 6 years), 10 months after	51.67	Nematodes	55.20	2285.80	367.49
		Kinorhynchs	10.75	43.69	0.17
		Nauplii	8.09	36.36	3.80
Before (10 and 6 years), 4 years after	20.75	Nematodes	24.56	2285.80	2027.70
		Nauplii	15.27	36.36	55.20
		Copepods	13.36	37.09	55.06
		Kinorhynchs	12.69	43.69	7.95
		Grastrotichs	6.81	2.86	1.90
10 weeks after, 10 months after	33.55	Nematodes	49.75	156.75	367.49
		Copepods	17.10	15.21	12.60
		Nauplii	11.36	2.13	3.80
10 weeks after, 4 years after	54.98	Nematodes	62.88	156.75	2027.70
		Nauplii	11.58	2.13	55.20
10 months after, 4 years after	47.95	Nematodes	55.83	367.49	2027.70
		Nauplii	11.88	3.80	55.20
		Copepods	10.14	12.60	55.06

Table 6 (on next page)

SIMPER analysis results for the nematode community between time points

SIMPER analysis results for the nematode community indicating average among time point community dissimilarity and the contribution of individual species contributing 70% of more to among time point community dissimilarity. Avg. Abundance is the average abundance of individuals standardized to 10 cm² of seafloor area.

Time points	Average dissimilarity	Species	% Contribution	Time 1 avg. abundance	Time 2 avg. abundance
Before (6 years), 10 weeks after	81.79	<i>Sabatieria</i> sp. A	7.61	354.57	11.22
		<i>Hopperia beaglense</i>	6.51	178.22	0.19
		<i>Microlaimus</i> sp. 34	6.05	133.17	1.64
		<i>Cervonema kaikouraensis</i>	5.04	126.56	1.85
		<i>Campylaimus</i> sp. 6	4.87	96.83	0.10
		<i>Sabatieria</i> sp. 12	4.58	98.80	0.55
		<i>Leptolaimus</i> sp. 14	4.37	78.50	0.10
		<i>Sphaerolaimus</i> sp. 1	3.59	59.60	0.38
		<i>Retrotheristus</i> sp. 5	3.35	58.37	0.94
		<i>Metalinhomoeus</i> sp. 1	3.13	50.84	0.94
Before (6 years), 10 months after	77.50	<i>Sabatieria</i> sp. A	7.38	354.57	18.23
		<i>Microlaimus</i> sp. 34	6.12	133.17	1.14
		<i>Hopperia beaglense</i>	6.12	178.22	2.31
		<i>Daptonema</i> sp. 18	6.01	83.72	242.42
		<i>Campylaimus</i> sp. 6	5.09	96.83	0.00
		<i>Cervonema kaikouraensis</i>	5.09	126.56	2.31
		<i>Leptolaimus</i> sp. 14	4.57	78.50	0.00
		<i>Sphaerolaimus</i> sp. 1	3.94	59.60	0.00
		<i>Sabatieria</i> sp. 12	3.56	98.80	8.41
		<i>Metalinhomoeus</i> sp. 1	3.16	50.84	0.76
Before (6 years), 4 years after	59.71	<i>Daptonema</i> sp. 18	10.51	83.72	1012.51
		<i>Microlaimus</i> sp. 34	5.32	133.17	70.90
		<i>Sabatieria</i> sp. A	5.12	354.57	273.57
		<i>Leptolaimus</i> sp. 14	4.07	78.50	0.00
		<i>Campylaimus</i> sp. 6	3.31	96.83	7.56
		<i>Endeolophos</i> sp. 3	3.03	32.38	101.20
10 weeks after, 10 months after	71.23	<i>Daptonema</i> sp. 18	15.21	20.43	242.42
		<i>Daptonema</i> sp. 21	5.31	0.19	18.23
		<i>Endeolophos</i> sp. 3	4.03	0.10	10.56
		<i>Sabatieria</i> sp. A	3.74	11.22	18.23
		<i>Daptonema</i> sp. 27	3.71	0.10	9.24
		<i>Sabatieria</i> sp. 12	3.31	0.55	8.41
		<i>Paramesonchium</i> sp. 2	3.06	0.00	4.58
		<i>Metacyatholaimus</i> sp. 1	3.04	4.08	0.00
10 weeks after, 4 years after	82.55	<i>Daptonema</i> sp. 18	16.30	20.43	1012.51
		<i>Sabatieria</i> sp. A	7.88	11.22	273.57
		<i>Endeolophos</i> sp. 3	5.62	0.10	101.20
		<i>Hopperia beaglense</i>	4.63	0.19	71.74
		<i>Microlaimus</i> sp. 34	4.19	1.64	70.90

		<i>Daptonema</i> sp. 21	3.69	0.19	44.76
		<i>Cervonema kaikouraensis</i>	3.42	1.85	51.98
		<i>Dichromadora</i> sp. 7	3.37	0.76	44.89
		<i>Comesomatidae</i> sp. 6	3.25	0.00	27.77
		<i>Sabatieria</i> sp. 12	3.22	0.55	27.77
		<i>Chromadora</i> sp. 1	3.13	0.10	32.72
10 months after,					
4 years after	69.10	<i>Daptonema</i> sp. 18	13.03	242.42	1012.51
		<i>Sabatieria</i> sp. A	8.56	18.23	273.57
		<i>Microlaimus</i> sp. 34	4.83	1.14	70.90
		<i>Hopperia beaglense</i>	4.68	2.31	71.74
		<i>Endeolophos</i> sp. 3	4.57	10.56	101.20
		<i>Dichromadora</i> sp. 7	3.95	0.38	44.89
		<i>Cervonema kaikouraensis</i>	3.89	2.31	51.98
		<i>Chromadora</i> sp. 1	3.78	0.00	32.72
		<i>Comesomatidae</i> sp. 6	3.70	0.00	27.77
		<i>Sabatieria</i> sp. 12	3.60	8.41	27.77

Table 7 (on next page)

DISTLM results for the marginal and sequential tests for meiofauna and nematode community

DISTLM results for the marginal and sequential tests for meiofauna and nematode community structure relationships with environmental variables before and after a turbidity flow in Kaikōura Canyon. AIC = Akaike Information Criterion, SS = sum of squares, Pseudo-F = multivariate analogue Fisher's F test, P = p-value (significant values (<0.05) are in bold), and Prop. = indicates the proportion of variation explained by each variable.

Variable	Test	AIC	SS (trace)	Pseudo-F	P	Prop.
Meiofauna	Marginal					
Depth (m)		-	800.13	1.002	0.365	0.046
TOM%		-	2363.60	3.264	0.037	0.135
Chl <i>a</i> (µg/g)		-	653.15	0.811	0.437	0.037
Chl <i>a</i> : Phaeo		-	625.69	0.775	0.468	0.036
C : N (molar)		-	541.77	0.668	0.528	0.031
PN%		-	2137.20	2.908	0.049	0.122
Skewness (F&W phi)		-	1400.30	1.818	0.143	0.080
<16 micron		-	1771.50	2.355	0.085	0.101
	Sequential					
TOM%		153.36	2363.60	3.264	0.038	0.135
Chl <i>a</i> : Phaeo		152.85	1573.90	2.309	0.084	0.090
Skewness (F&W phi)		152.82	1153.30	1.756	0.155	0.066
Nematodes	Marginal					
Depth (m)		-	1533.00	0.563	0.913	0.086
TOM%		-	3561.90	1.493	0.132	0.199
Chl <i>a</i> (µg/g)		-	4149.30	1.813	0.073	0.232
Chl <i>a</i> : Phaeo		-	1914.60	0.720	0.740	0.107
C : N (molar)		-	5862.10	2.926	0.018	0.328
PN %		-	2827.20	1.127	0.304	0.158
Skewness (F&W phi)		-	2173.50	0.830	0.596	0.122
<16 micron		-	3575.30	1.500	0.129	0.200
	Sequential					
C : N (molar)		62.518	5862.1	2.926	0.016	0.328
Chl <i>a</i> (µg/g)		62.245	2973	1.643	0.063	0.166
TOM%		61.589	2555.6	1.575	0.124	0.143
Chl <i>a</i> : Phaeo		60.595	2026.4	1.362	0.301	0.113
Skewness (F&W phi)		58.388	1825.4	1.384	0.336	0.102
PN %		56.28	1059.6	0.671	0.605	0.059

Figure 1

Map of sampling locations

Location of sampling sites in Kaikōura Canyon overlaid on canyon flushing-induced bathymetric changes. A) Magnitude of erosion and deposition (seafloor change) within Kaikōura Canyon caused by the canyon flushing triggered by the Kaikōura Earthquake, measured by the differencing the pre- and post-earthquake bathymetry data sets (Mountjoy et al., 2018) . B) Location of the time-series of multicorer sampling sites (yellow circles = sampled in 2010, late 2017, and 2020; purple circles = sampled in 2006 in addition to other time points; green circles = sampled in early 2017 in addition to other time points) within the head of Kaikōura Canyon. Inset shows the location of Kaikōura Canyon (star) relative to New Zealand. Some of the red (erosional) banding evident along the bottom reach of Kaikōura Canyon is an artefact of higher levels of uncertainty in bathymetric differencing for overlapping multibeam coverages (for more detail see Mountjoy et al., 2018). Figure replicated from Bigham et al. (2023b).

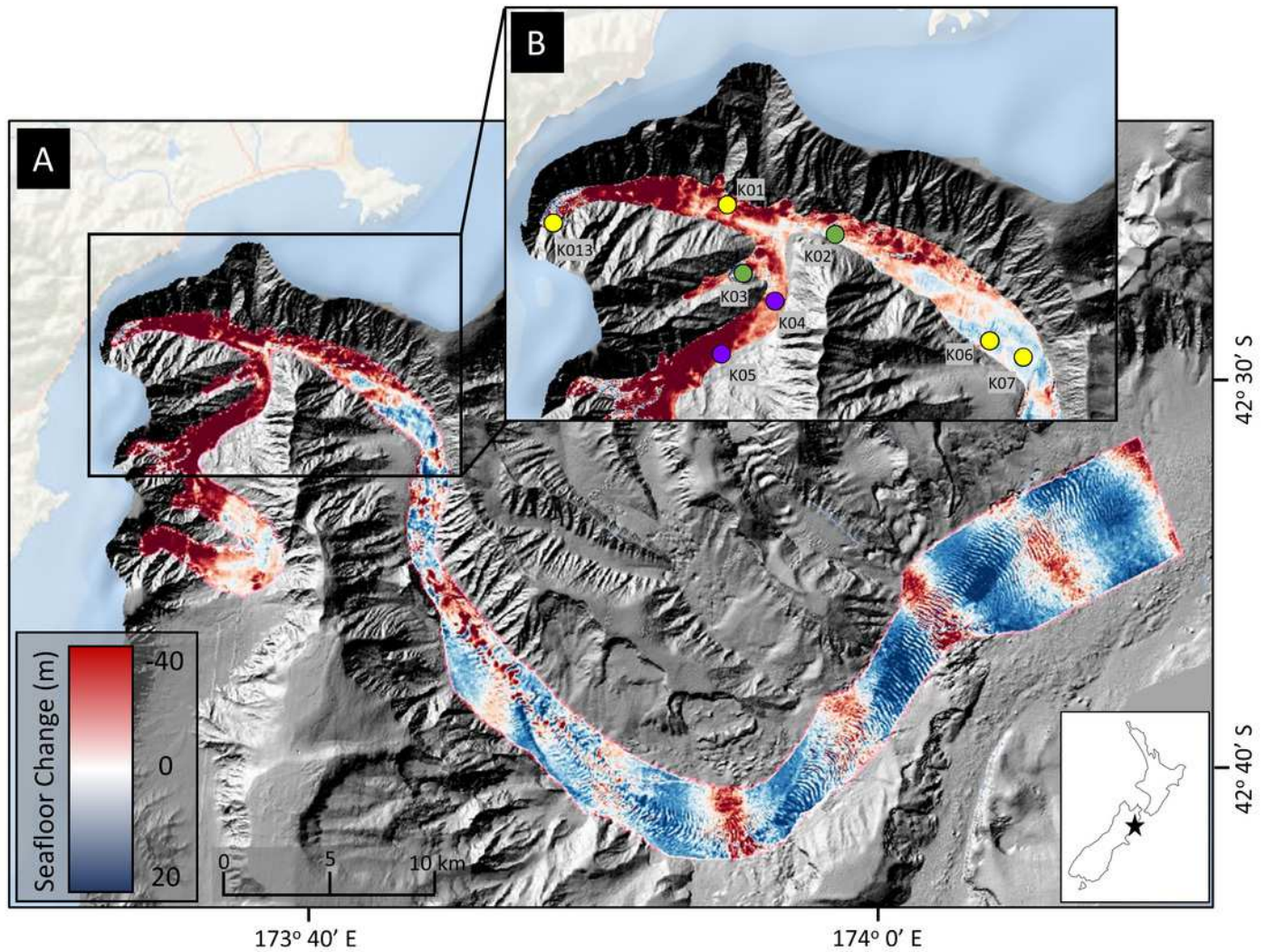


Figure 2

Non-metric multidimensional scaling (nMDS) plots of meiofauna and nematode community structure

Non-metric multidimensional scaling (nMDS) plots of community structure: A) meiofauna, B) meiofauna centroids, C) nematodes, and D) nematode centroids at 10 and 6 years before the turbidity flow and at 10 weeks, 10 months, and 4 years after the disturbance in Kaikōura Canyon. Similarities were calculated from zero adjusted, square root transformed fauna abundances for both community levels. All stress values are below 0.2, indicating that the plots are acceptable representations of the similarity patterns.

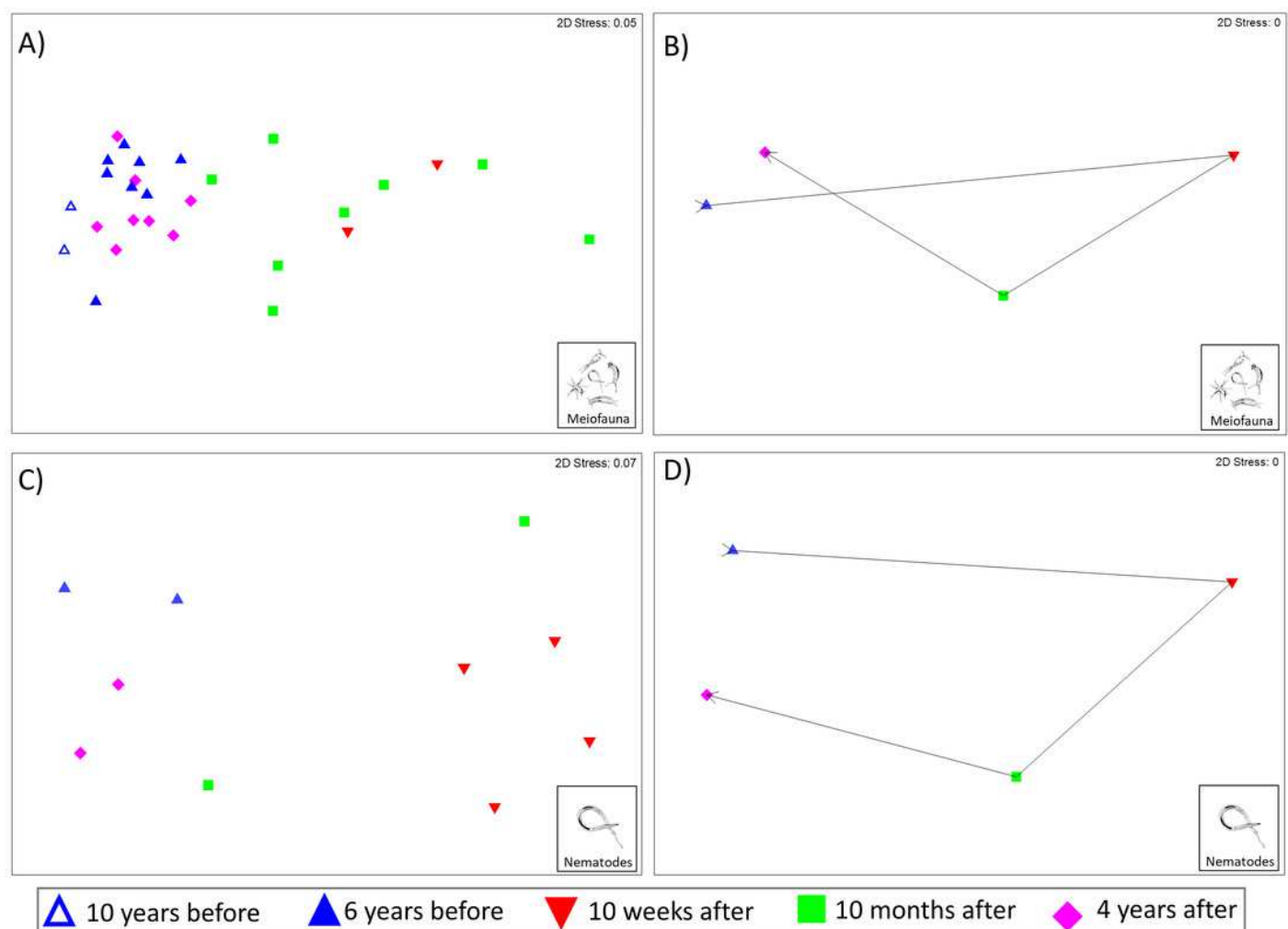


Figure 3

Distance-based redundancy analysis (dbRDA) plots for meiofauna and nematodes

Distance-based redundancy analysis (dbRDA) plot visualising in two-dimensions the relationships between variation in community structure for A) meiofauna and B) nematodes (6 years before, and 10 weeks, 10 months, and 4 years after the turbidity flow event in Kaikōura Canyon) and environmental variables examined by the DISTLM analysis. Only variables with a Spearman rank correlation greater than 0.2 are displayed. Vector lengths are proportional to their contribution to the overall variation.

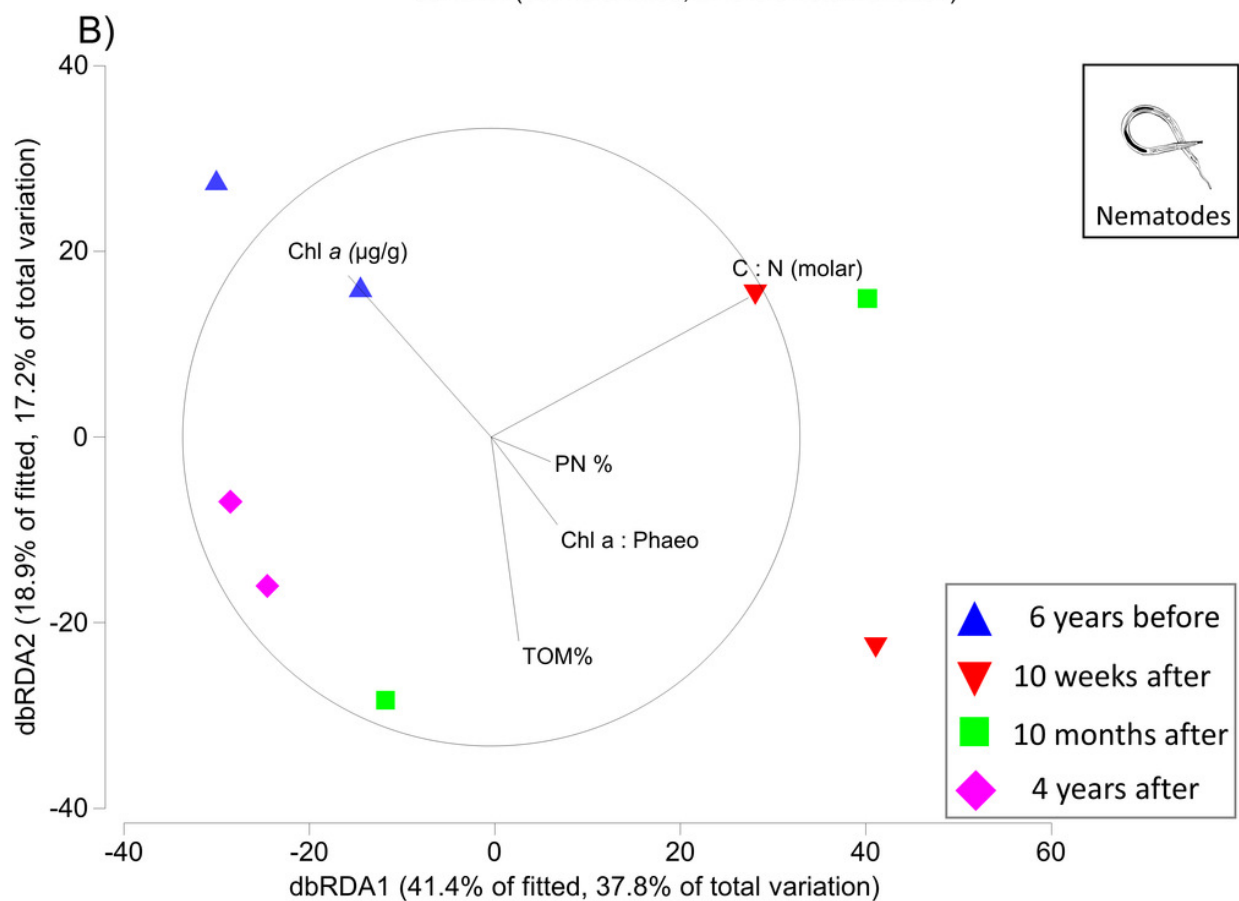
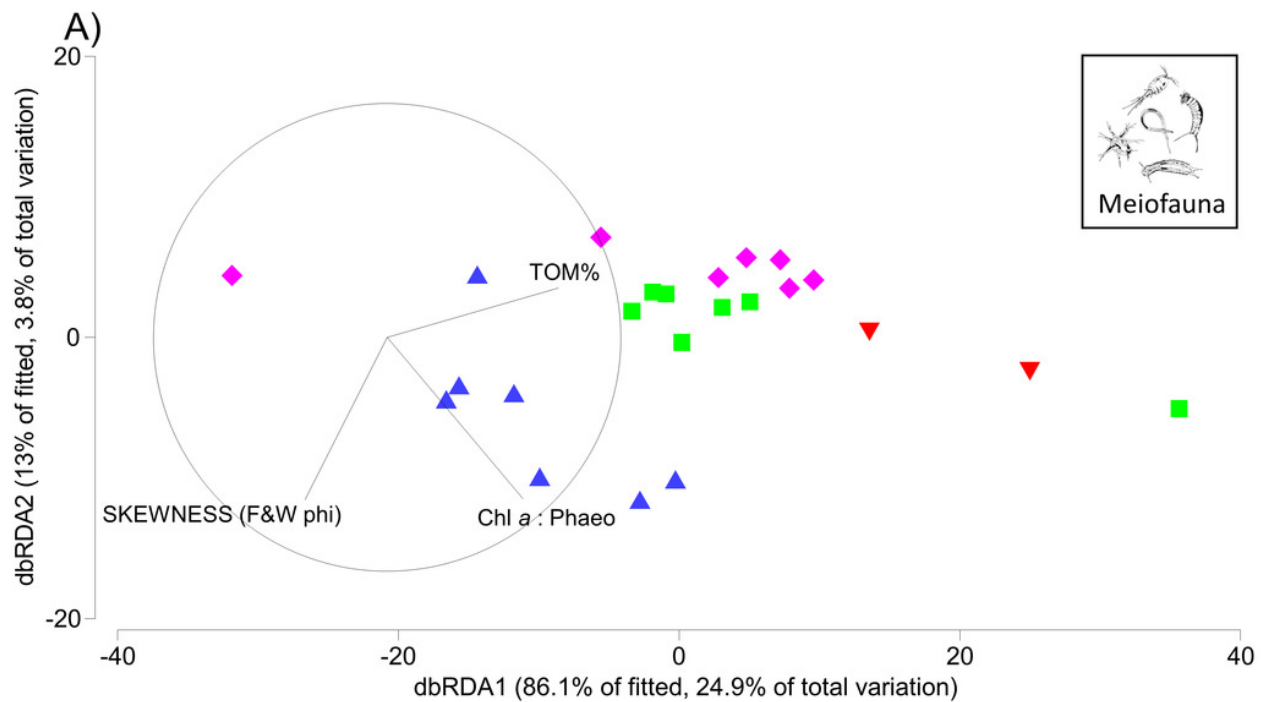


Figure 4

Scatter plots of key environmental factors

Scatter plots of the most important environmental factors identified by the DISTLM analysis for structuring meiofauna and nematode communities before and after a turbidity flow in Kaikōura Canyon. A) The percent total organic matter (% TOM), B) nitrogen (%N C) the ratio of molar carbon (C) to nitrogen (N), D) Chl *a* ($\text{mg g}^{-1}_{\text{sediment}}$), E) ratio of Chl *a* to phaeopigments, F) the skewness of grain size, and G) the percent of grains less than 16 μm . Each dot represents a single core.

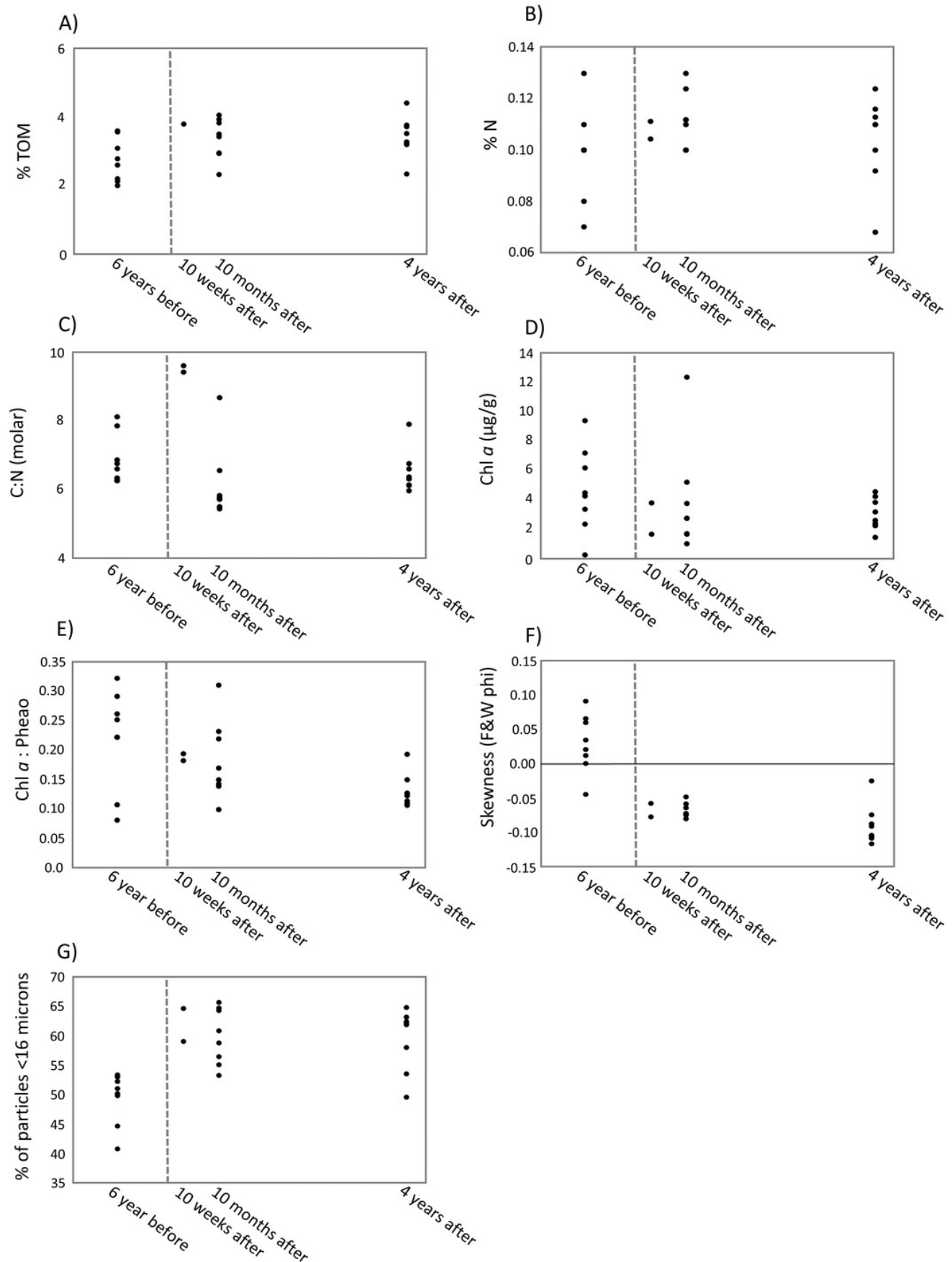


Figure 5

Plots predicting time to recovery for meiofauna and nematodes

Plots showing three hypothetical models of population growth (linear, exponential, and logistic) used to predict the time to community recovery (indicated by the grey area on the plot; the minimum threshold of 79% or 46% similarity is the within-group similarity of the pre-turbidity community structure) for: A) the meiofauna and B) nematode communities in Kaikōura Canyon.

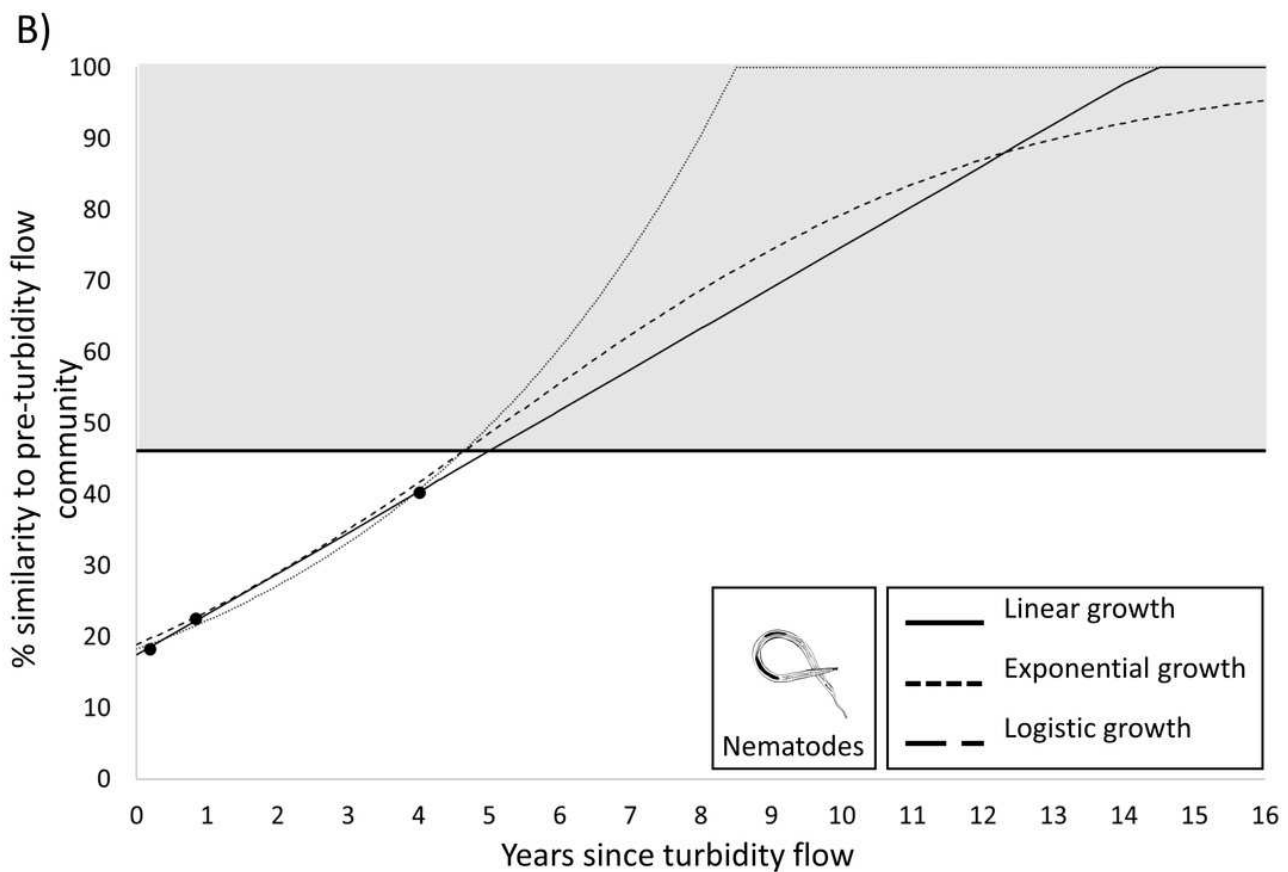
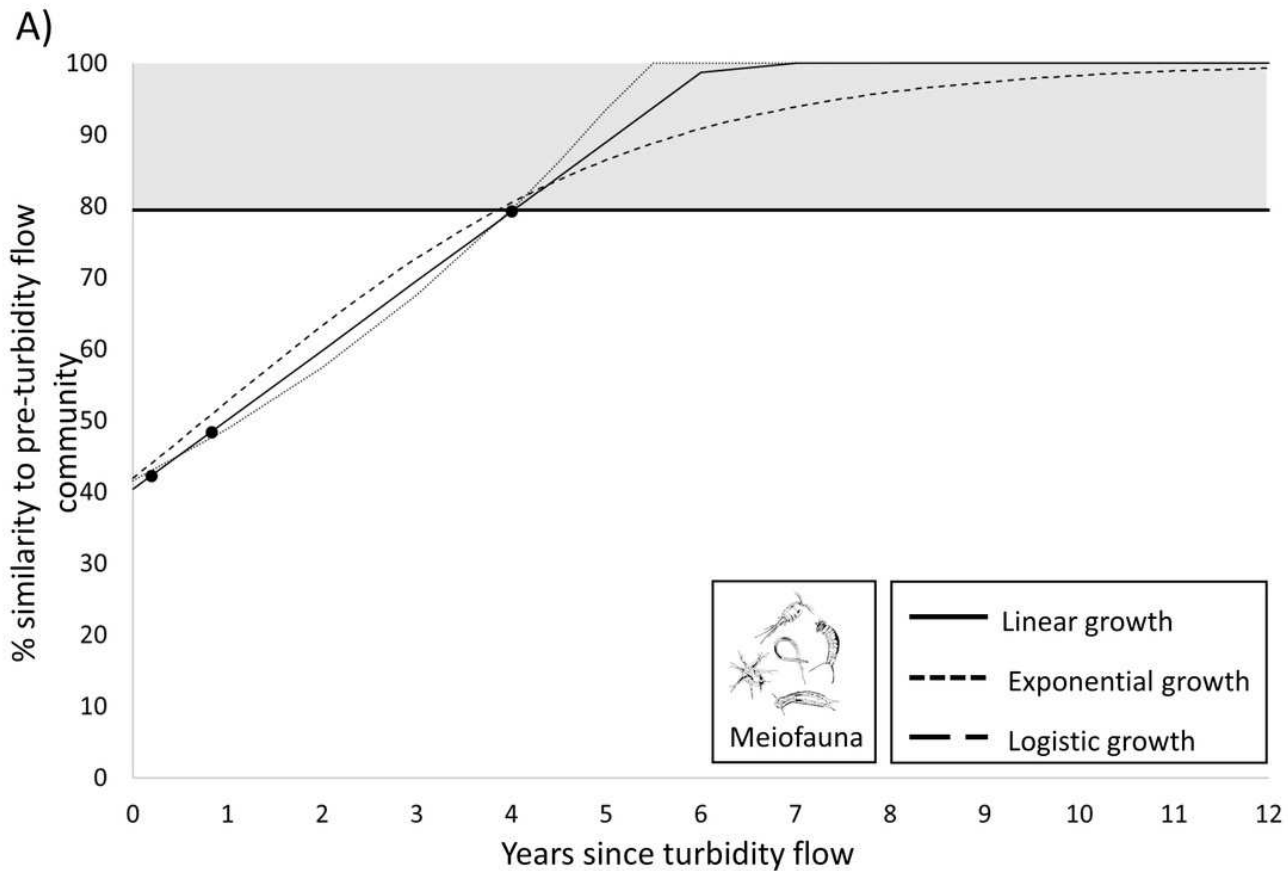


Figure 6

Plot of juvenile nematode percentages through time

Plot showing the average percentage of juvenile nematodes from sites K2 and K3 at each time point. The dashed line indicates when the turbidity flow in Kaikōura Canyon occurred.

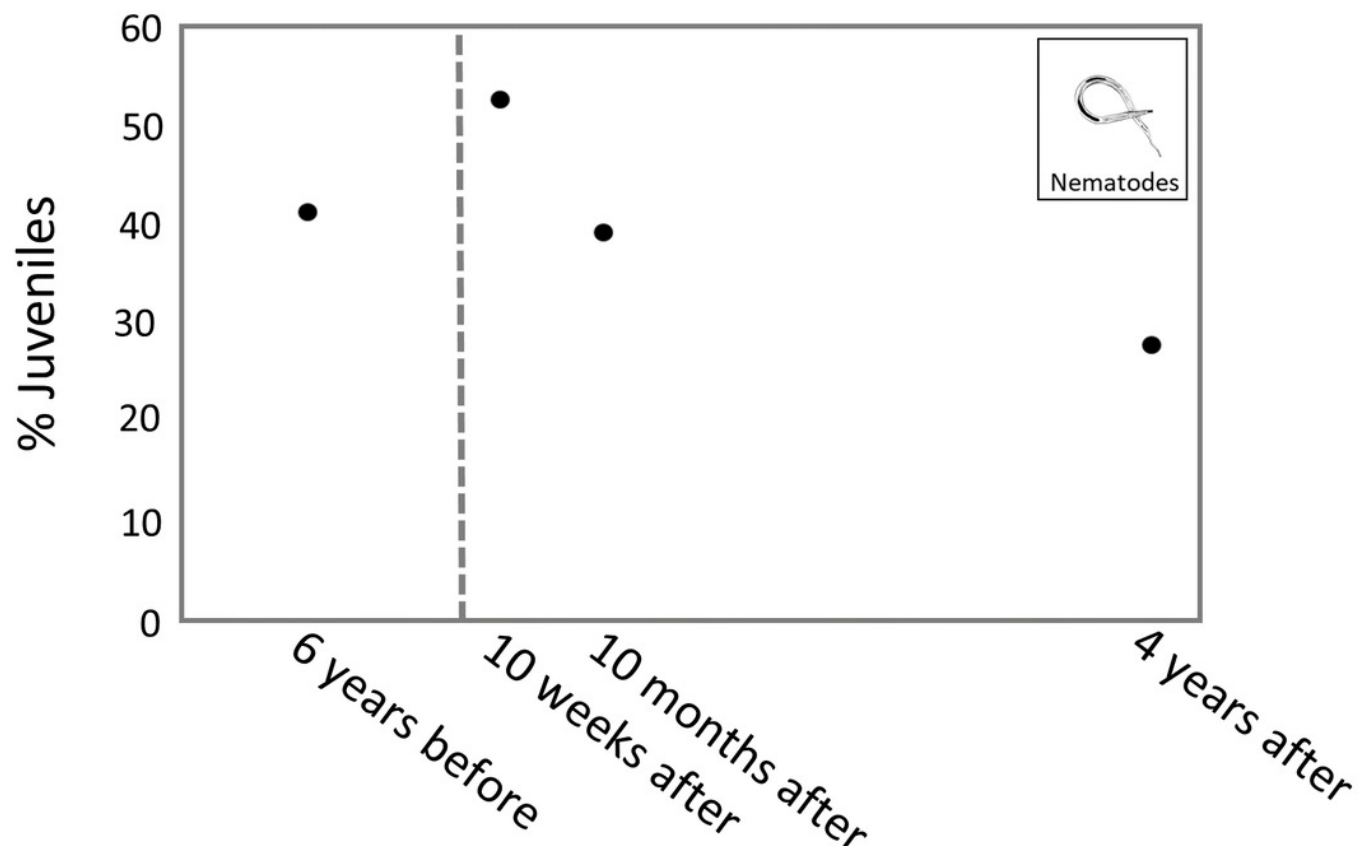


Figure 7

Illustrated schematic showing the changes in the meiofauna community through time

Schematic illustration showing of the relative abundances of the key taxa identified by the meiofauna SIMPER analysis that characterised the changes in the meiofauna community before and after the turbidity flow in Kaikōura Canyon. Solid arrows connect time points. One individual represents an average abundance of 1-10 ind./10 cm², two individuals represent an average abundance of 10-100 ind./10 cm², three individuals represent an average abundance of 100-1000 ind./10 cm², four individuals represent an average abundance of 1000-2000 ind./10 cm², and five individuals represents 2000+ ind./10 cm².

