# Species divergence in valleys: the phylogeny of Phrynocephalus forsythii complex and description of a new species 

Qianru Liang and Lei Shi<br>Xinjiang Key Laboratory for Ecological Adaptation and Evolution of Extreme Environment Biology, College of Life Sciences, Xinjiang Agricultural University, Urumqi, Xinjiang, China

## ABSTRACT

Background: Geographic isolation caused by high-altitude valleys promotes the formation of geographic segregation of species, leading to species differentiation. The subgenus Oreosaura contains viviparous species from the Tibetan Plateau and the vicinity of the Tarim Basin, which can be divided into three species complexes according to their geographical distribution: Phrynocephalus vlangalii,
Phrynocephalus theobaldi, and Phrynocephalus forsythii. However, molecular data for the P. forsythii complex are limited and the diversity of this species complex has been greatly underestimated. Therefore, this study aimed to explore the species diversity of Oreosaura and species differentiation within the P. forsythii complex.
Methods: We analysed the species diversity of Oreosaura by combining previous data, constructed a phylogenetic tree of the subgenus based on cytochrome c oxidase subunit I and 16 S sequences, and estimated the divergence time.
Results: The results suggest significant genetic differences between the Tarim Basin populations and adjacent mountain valley populations of the P. forsythii complex and that the combination of deep valley landscapes in the high mountains and ice-age events have contributed to the differentiation of the viviparous toad-headed agama lizard, which is a key factor in the phylogenetics of the $P$. forsythii complex. Furthermore, we identified a population collected from Wuqia County, Xinjiang, as a new species, Phrynocephalus kangsuensis sp. nov. The results will provide data for phylogenetic studies following the P. forsythii complex and help demonstrate that valleys promote the formation of Phrynocephalus species.

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Corresponding author
Lei Shi, leis@xjau.edu.cn
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## INTRODUCTION

Geographic isolation results in an inability to exchange genes between populations and can eventually lead to population differentiation over time and high selection pressure (Coyne \& Orr, 2004; Nosil, Harmon \& Seehausen, 2009). Thus, pronounced geographical isolation promotes species differentiation (Mayr, 1970; Kadmon \& Pulliam, 1993; Coyne \& Orr, 2004; Nosil, Harmon \& Seehausen, 2009; Winger \& Bates, 2015). In addition to island habitats, high mountains and valley topography on continents geographically isolate
populations, promoting species differentiation (Wang et al., 2021, 2022; Bogdanov et al., 2023; Nazarov et al., 2023).

The subgenus of Phrynocephalus, Oreosaura, was established by Barabanov \& Ananjeva (2007) and contains the species complex of Phrynocephalus vlangalii, Phrynocephalus theobaldi, and Phrynocephalus forsythii. This subgenus contains viviparous species that live at high altitudes on the Tibetan Plateau and low altitudes in the Tarim Basin. Among them, only P. forsythii has both valleys populations living in the high mountains and valleys and plains populations distributed in the basins, rendering it ideal for studying species differentiation.

Early phylogenetic studies of Phrynocephalus found that P. forsythii was the first to diverge in the viviparous species complex, forming a sister clade with another viviparous species complex (Pang et al., 2003; Guo \& Wang, 2007; Jin \& Brown, 2013). Qi et al. (2020) conducted genetic analyses of $P$. forsythii populations distributed in the Tarim Basin and found that the geographic populations form sister clades and have a monophyletic lineage, suggesting that $P$. forsythii populations distributed around the Tarim Basin may be a circumscribed species, with the results showing no obvious genetic differentiation among these populations distributed at low altitudes; however, their study did not obtain data on populations of $P$. forsythii at high altitudes. In recent years, we collected several valleys P. forsythii populations on field trips and found significant morphological differences from plains populations. Phrynocephalus nasatus is closely related to P. forsythii and its dorsal body colour pattern is similar to that of P. forsythii (Golubev \& Dunayev, 1995; Zhao, Zhao \& Zhou, 1999). Phrynocephalus nasatus has not been reported since its discovery and description as a new species by the Zoological Museum of Moscow University in 1995 (Golubev \& Dunayev, 1995) and there have been no further studies on the holotype despite its distinctive morphological characteristics and questionable taxonomic status. Phrynocephalus nasatus has been described as a synonym of P. axillaris (Barabanov \& Ananjeva, 2007). Dunayev (2020) described the habitat, ecology, and morphological body colour of $P$. nasatus but did not make morphological comparisons with $P$. forsythii. In their analysis of Oreosaura, Solovyeva et al. (2023) found that P. nasatus was closely related to P. forsythii in phylogenetic relationships.

Solovyeva et al. (2018) combined mtDNA and nuDNA to analyse the impact of environmental change on the evolutionary and biogeographical history of Phrynocephalus and concluded that the divergence of Oreosaura was related to the timing of the major uplift of the Tibetan Plateau. Recently, Solovyeva et al. (2023) constructed a very detailed phylogenetic tree for Phrynocephalus based on cytochrome oxidase subunit I (COI) sequences, in which Oreosaura was described in detail; however, because of the limited molecular data for the P. forsythii complex, the genetic differentiation of this complex was not discussed in their study.

Thus, the present study aimed to explore the species diversity of Oreosaura and species differentiation within the P. forsythii complex based on the data of Solovyeva et al. (2023) and with additional data from valleys areas of $P$. forsythii and 16 Sequences. We explored the relationship between the phylogenetic lineages of the $P$. forsythii complex and the main
tectonically induced events in southern Xinjiang. We believe that the phylogenetic relationships of the $P$. forsythii complex can be used to evaluate the influence of paleogeographic and climatic factors.

## MATERIALS AND METHODS

## Materials

We collected specimens from three geographically segregated populations: altitude $>2,000 \mathrm{~m}$ (higher altitude populations) distributed in the valley. All the specimens ( 18 in total) we used were fixed in $95 \%$ ethanol, and were collected from three high-altitude areas (altitude > 2,000) (Table S1)—the Kangsu River (Wuqia population, altitude: 2,717 m) and Toshkan River (Akqi population, Group AKQ, altitude: $2,125 \mathrm{~m}$ ) valleys in southern Tien Shan, and the Karakash River (Hotan population, Group HT, altitude: 3,148 m) valley in the middle Kunlun Mountains in 2006 and 2009 (Fig. 1).

The liver or muscle tissue in the samples was preserved with $95 \%$ ethanol for DNA extraction. All specimens were deposited in the Animal Herbarium of the College of Life Sciences, Xinjiang Agricultural University. These experimental procedures involving animals were approved (animal protocol number: 2023013) by the Animal Welfare and Ethics Committee of Xinjiang Agricultural University, Urumqi, Xinjiang, China. We also adhered to the ARROW (Animals in Research: Reporting On Wildlife) guidelines.

Sequences available for other species of the subgenus Oreosaura were collected and downloaded from the GenBank at the National Center for Biotechnology Information and Barcode of Life Database (Table S1).

## DNA extraction and polymerase chain reaction (PCR) amplification

Total DNA was extracted from tissue samples using the Foregene Animal Tissue Genomic DNA Extraction Kit, according to the manufacturer's instructions. The mitochondrial COI and 16 S rRNA genes ( 16 S ) were used as DNA barcode markers and amplified by Polymerase Chain Reaction (PCR) using published primers and cycling parameters (Pavlicev \& Mayer, 2009; Nagy et al., 2012). The PCR reaction system volume was $25 \mu$ l, which contained $1 \mu \mathrm{l}$ of template DNA, $1 \mu \mathrm{l}$ each of upstream and downstream primers, $12.5 \mu \mathrm{l}$ of 2 x Taq PCR Mix, and $9.5 \mu \mathrm{l}$ of $\mathrm{ddH}_{2} \mathrm{O}$. The PCR conditions for the COI gene were as follows: $94^{\circ} \mathrm{C}, 4 \mathrm{~min} ; 9{ }^{\circ} \mathrm{C}, 30 \mathrm{~s} ; 52{ }^{\circ} \mathrm{C}, 30 \mathrm{~s} ; 72{ }^{\circ} \mathrm{C}, 50 \mathrm{~s} ; 35$ cycles; and $72{ }^{\circ} \mathrm{C}$, 10 min . The PCR conditions for the 16 S gene were as follows: $94^{\circ} \mathrm{C}, 2 \mathrm{~min} ; 95^{\circ} \mathrm{C}, 30 \mathrm{~s}$; $59{ }^{\circ} \mathrm{C}, 30 \mathrm{~s} ; 72{ }^{\circ} \mathrm{C}, 50 \mathrm{~s} ; 35$ cycles; and $72{ }^{\circ} \mathrm{C}, 7 \mathrm{~min}$. Both amplicons were stored at $4^{\circ} \mathrm{C}$ after the PCR was completed. Subsequently, gel electrophoresis with $0.5 \%$ TBE (Tris-Borate-EDTA buffer) solution and agarose was performed to confirm successful amplification. Successfully amplified PCR stock solution was sent to Sangon Biotechnology (Shanghai, China) for purification and sequencing.

## Phylogenetic analyses \& divergence time estimation

The sequencing results were comparatively spliced and manually edited using SeqMan with DNASTAR v6 (Burland, 1999). The spliced sequences were aligned using MEGA 7.0.26 (Kumar, Stecher \& Tamura, 2016) and were converted into amino acid sequences


Figure 1 Geographic characteristics and locations of the sampling sites and reference sequences of this study. Note: For names of the areas concerned, please refer to Table S1. Stars, sample sites collected in this study (high-altitude); circles, sample sites of some of the reference sequences (low-altitude).

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using protein-coding to check for stop codons. Subsequently, genetic distances ( $p$-distances) were calculated using MEGA 7.0.26. We used Partitionfinder-2.1.1 (Lanfear et al., 2017) to estimate the best model for the molecular dataset using the Akaike information criterion. The best models for COI sequences were the HKY, GTR+G, and SYM $+\mathrm{I}+\mathrm{G}$ models, the best model for 16 S sequences was the TIM+I+G model, and the best models for spliced sequences were the TRN+I+G, HKY, and GTR models. We then used MrBayes 3.2.2 (Ronquist et al., 2012) and PhyloSuite 1.2.2 (Zhang et al., 2020) to perform Bayesian inference (BI) and maximum likelihood (ML) on the three datasets. Phylogenetic trees were constructed using P. mystaceus and P. axillaris as outgroups. We used the Markov chain Monte Carlo (MCMC) theory approach in BI, running 5,000,000 generations, drawing one sample every 1,000 , running four chains per analysis, and removing the top $25 \%$ of the data with the other parameters as the default. We used the K80 (K2P) method in ML, running 1,000,000 generations with the other parameters set as their default values (Zhang et al., 2020). The final results were checked using Tracer 1.7.2 (Rambaut et al., 2018) and had effective sample size (ESS) values > 200 for all parameter values. Finally, a phylogenetic tree was plotted using ITOLV6 (Letunic \& Bork, 2021). We considered an a posteriori probability (BPP) value of $>0.99$ or maximum likelihood bootstrap value of $>95 \%$ as strong support for monophyly.

We used the mtDNA dataset (COI+16S) to estimate species divergence times within the subgenus Oreosaura using BEAST 2.6.6 (Bouckaert et al., 2014) based on the lognormal
relaxed clock assumption. We used two calibration points: (1) the Pamir-Tian Shan uplift 10 million years ago (Ma) (Charreau et al., 2009) and (2) the final uplift of the Tibetan Plateau 3.6 Ma (Cui et al., 1996). We used the GTR and Yule models and sampled every 10,000 iterations, with $25 \%$ of the initial samples discarded as aged samples. Tracer 1.7.2 was used to evaluate the estimated ESS for all the parameters.

## Species delimitation

We followed the phylogenetic species concept and used COI sequences to classify molecular operational taxonomic units (MOTUs) into putative species. To assess the number of putative species-level lineages in the subgenus Oreosaura, we referred to the results of Solovyeva et al. (2023) and used the following three automated phylogenetic species concept-based species delimitation methods to estimate the number of species in the barcode data: (1) the automatic barcode gap discovery (ABGD) (Puillandre et al., 2012), (2) assemble species by automatic partitioning (ASAP) (Puillandre, Brouillet © Achaz, 2021), and (3) a Bayesian implementation of Poisson tree processes (bPTP) (Zhang et al., 2013).

In ABGD (https://bioinfo.mnhn.fr/abi/public/abgd/abgdweb.html) and ASAP (https:// bioinfo.mnhn.fr/abi/public/asap/asapweb.html), the Kimura (K80) TS/TV (2.0) model was used, with other parameters defaulted in ABGD, 20 selected in steps and one in X (relative gap width) in ASAP, and the remaining default parameters. In bPTP (https://species.h-its. org/ptp/), the thinning selected 200 and the remaining parameters were set to default.

## Morphometry \& scale statistics

Morphometric measurements were performed on P. forsythii specimens (six in total) collected from geographic populations (Wuqia) at valleys. Seven morphometric indices were measured using 0.01 mm vernier callipers, including snout-vent length (SVL), head length (HL), maximum head width (MHW), head height (HH), rostral eye distance (RED), nasal distance (NBD), and tail length (TL). The nasal scales (NS), inter-nasal scales (IS), supralabial shields, and infralabial shields were also counted. Morphological descriptions of P. forsythii and P. nasatus from the literature (Zhao, Zhao \& Zhou, 1999; Dunayev, 2020) were also collected for comparison with the samples from this study.

## Zoobank registration

The electronic version of this article in the Portable Document Format (PDF) represents a published work according to the International Commission on Zoological Nomenclature (ICZN); hence, the new names contained in the electronic version are effectively published under that code from the electronic edition alone. This published work and the nomenclature it contains have been registered in ZooBank, an online registration system for ICZN. The ZooBank Life Science Identifiers (LSIDs) can be resolved and the associated information can be viewed through any standard web browser by appending the LSID to the prefix http://zoobank.org/. Publication LSID urn:lsid:zoobank.org:pub:812FA766-6BC6-4D39-85B9-8E2C58E06098 Phrynocephalus kangsuensis sp. nov.: urn:lsid:zoobank. org:act:53EF51A0-DEEA-4E6A-B62F-AD61AB3868E9. The online version of this work
has been archived and made available from the following digital repositories: PeerJ, PubMed Central SCIE, and CLOCKSS.

## RESULTS

## Sequence \& phylogenetic tree

The base sequences of two mitochondrial target fragments, 16S (475 bp) and COI sequences ( 623 bp ), and a spliced dataset ( $1,099 \mathrm{bp}$ ) were obtained by extracting and sequencing DNA from three valleys geographic populations of $P$. forsythii. The results of the COI phylogenetic tree (Fig. 2) showed that the viviparous species complex (Oreosaura) was monophyletic (BPP/BS: 1.00/100). The P. forsythii complex first differentiated in Oreosaura and was monophyletic (BPP/BS: 1.00/96), with the geographic populations of the three valleys divided into three subclades (lineages A, B, and C). The Akqi population (Lineage C) formed a sister clade to the lower altitude populations (Lineage D, containing P. nasatus), and the Wuqia population (Lineage A) formed a sister clade to the other clades. Phrynocephalus nasatus is present in the low-altitude P. forsythii (Lineage D). The 16 S phylogenetic tree (Fig. 3) shows the same topological complementary structure as the COI phylogenetic tree: Oreosaura and the P. forsythii complex are monophyletic (BPP/ BS: 1.00/100; BPP/BS: 1.00/99, respectively). The Wuqia population (Lineage A) was monophyletic (BPP/BS: 1.00/100) and formed a sister clade with the Hotan population (Lineage B). Similarly, with the same structure as the splice sequence (Fig. 4), the Wuqia population (branch A) was monophyletic in the P. forsythii complex (BPP/BS: 1.00/100).

## Genetic distance

Uncorrected $p$-distance based on COI sequences (Table 1) showed that species in the subgenus Oreosaura were genetically distant from species in other subgenera ( $P$. axillaris and P. mystaceus) by $12.5-18.5 \%$, and between species within subgenera by $0.0-11.1 \%$. In the $P$. forsythii complex, there were significant differences in genetic distances (1.4-5.2\%) between the valleys and plains geographic populations. The population from Wuqia (P. kangsuensis sp.nov.) was genetically distant from geographic populations of the other two valleys (Group HT and Group AKQ) by $5.5-5.8 \%$, and up to $5.2 \%$ from P. forsythi. No difference was noted in the genetic distance between $P$. nasatus and P. forsythii $(0.0 \%)$. In addition, uncorrected $p$-distance based on 16 S sequences showed significant differences in genetic distances between species within the subgenus Oreosaura (0.0-4.9\%). In the P. forsythii complex, genetic differences between the valleys and plains populations ranged from $2.2-0.5 \%$, with the Wuqia populations (P. kangsuensis sp.nov.) being significantly genetically different from the other populations (2.0-2.3\%).

## Species delimitation

Species delimitation of the subgenus Oreosaura was defined based on COI sequences using three species delimitation methods (ABGD, ASAP, and bPTP) (Fig. 2). All three methods defined the Wuqia population (Lineage A) and Hotan population (Lineage B) as MOTUs. ASAP considered 11 species to be present in the subgenus Oreosaura, with the Akqi population (Lineage C), low-altitude population (Lineage D), Wuqia population (Lineage


Figure 2 Phylogenetic tree of COI sequence of Oreosaura. The value on the node is BPP/BS, coloured sections are the species divisions of Solovyeva et al. (2023). The highest support values are in bold. Full-size DOI: 10.7717/peerj.17175/fig-2
A), and Hotan population (Lineage B) in the species group of $P$. forsythii considered to be three MOTUs and P. vlangalii and P. nanschanicus considered to be a MOTU. In contrast, bPTP determined 18 MOTUs.


Figure 3 Phylogenetic tree of 16S sequence of Oreosaura. The value on the node is BPP/BS. Note: QM, Qiemo; BC, Baicheng; WS, Wensu; KS, Kashgar; AKS, Aksu; RQ, Ruoqiang. The highest support values is in bold.

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## Divergence time estimates

The divergence times for subgenus Oreosaura and geographic populations of P. forsythii based on spliced sequences were estimated (Fig. 5). Oreosaura diverged at $10.43 \mathrm{Ma}(95 \%$ HPD: [10.00-11.33] Ma). The P. forsythii complex diverged at 6.73 Ma ( $95 \%$ HPD:


Figure 4 Bayesian phylogenetic trees of Oreosaura on spliced sequences (CO1 and 16S). The value on the node is BPP/BS. Note: QM, Qiemo; BC, Baicheng; AKS, Aksu. The highest support values are shown in bold.

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[4.75-8.76] Ma). The Wuqia population (P. kangsuensis sp.nov.) was the first within the complex P. forsythii, at $3.96 \mathrm{Ma}(95 \%$ HPD: [2.57-5.40] Ma), followed by the Hotan population at $2.92 \mathrm{Ma}(95 \% \mathrm{HPD}$ : [1.75-4.16] Ma). The divergence between low- and valleys populations (Akqi population) occurred at $1.59 \mathrm{Ma}(95 \%$ HPD: [ $0.88-2.35] \mathrm{Ma}$ ).

## Description of the new species

Phrynocephalus kangsuensis sp. nov.
ZooBank: lsid:zoobank.org:act:812FA766-6BC6-4D39-85B9-8E2C58E06098.
Specimens: The holotype was male (field collection no. CX09247) and was collected by Lei Shi from Kangsu Town, Wuqia County, Kizilsu-Kirghiz Autonomous Prefecture, Xinjiang. The allotype was female (CX09246) and the paratypes consisted of four males (CX09243, CX09244, CX09245, and CX09248), all of which were collected from the vicinity of Kangsu Town, Wuqia County, Xinjiang, which has an altitude of $2,300 \mathrm{~m}$ above sea level.

Table 1 Uncorrected $p$-distances (\%) between species of Oreosaura.

|  | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1. P. kangsuensis sp.nov. |  | 2.0 | 2.1 | 2.2 | 2.3 | - | 3.3 | - | 4.6 | - | - | - | 4.1 | 4.9 | 4.3 | 4.1 | 5.9 | 7.6 |
| 2. Group AKQ | 5.5 |  | 1.4 | 1.7 | 1.8 | - | 2.3 | - | 3.5 | - | - | - | 3.0 | 3.8 | 3.3 | 3.0 | 5.7 | 6.5 |
| 3. Group HT | 5.8 | 3.5 |  | 0.5 | 0.6 | - | 2.0 | - | 2.3 | - | - | - | 2.6 | 2.9 | 3.4 | 3.1 | 5.0 | 6.0 |
| 4. P. forsythii | 5.2 | 1.4 | 2.6 |  | 0.0 | - | 2.5 | - | 2.9 | - | - | - | 3.2 | 3.5 | 3.5 | 3.2 | 4.6 | 6.7 |
| 5. P. nasatus | 5.4 | 1.6 | 3.0 | 0.0 |  | - | 2.6 | - | 3.0 | - | - | - | 3.3 | 3.5 | 3.5 | 3.3 | 4.6 | 6.7 |
| 6. P. v. pylzowi | 8.5 | 7.2 | 8.5 | 6.9 | 7.0 |  | - | - | - | - | - | - | - | - | - | - | - | - |
| 7. P. vlangalii | 7.7 | 7.1 | 7.8 | 6.6 | 7.0 | 1.2 |  | - | 1.5 | - | - | - | 0.9 | 1.6 | 1.3 | 1.1 | 4.8 | 5.3 |
| 8. P. nanschanicus | 8.1 | 7.1 | 8.3 | 7.1 | 7.5 | 2.4 | 2.4 |  |  | - | - | - | - | - | - | - | - | - |
| 9. P. theobaldi | 9.6 | 10.0 | 10.0 | 8.9 | 9.6 | 5.9 | 6.5 | 7.0 |  | - | - | - | 2.0 | 2.0 | 3.1 | 2.8 | 5.3 | 5.3 |
| 10. P. sp. 1 | 8.0 | 7.4 | 7.4 | 6.5 | 6.6 | 5.9 | 5.2 | 5.8 | 5.9 |  | - | - | - | - | - | - | - | - |
| 11. P. Lhasaensis | 11.1 | 10.1 | 10.8 | 8.1 | 8.5 | 7.7 | 7.8 | 7.1 | 7.0 | 6.1 |  | - | - | - | - | - | - | - |
| 12. P. sp. 2 | 9.1 | 7.8 | 7.8 | 6.8 | 7.4 | 6.7 | 6.1 | 6.1 | 6.4 | 4.5 | 4.8 |  | - | - | - | - | - | - |
| 13. P. parvus | 8.7 | 9.4 | 9.4 | 8.5 | 9.0 | 6.1 | 5.5 | 6.1 | 6.9 | 5.8 | 8.1 | 6.1 |  | 1.2 | 2.3 | 2.5 | 5.4 | 5.4 |
| 14. P. erythrurus | 8.7 | 8.0 | 8.1 | 7.1 | 7.7 | 6.7 | 6.8 | 6.1 | 8.3 | 5.8 | 7.4 | 5.5 | 4.8 |  | 2.8 | 3.0 | 5.7 | 5.6 |
| 15. P. putjatai | 7.8 | 7.1 | 7.3 | 6.5 | 6.5 | 5.9 | 6.3 | 7.0 | 6.1 | 5.7 | 7.7 | 7.1 | 8.3 | 9.2 |  | 0.2 | 6.2 | 5.9 |
| 16. P. guinanensis | 7.7 | 6.8 | 7.1 | 6.0 | 5.8 | 7.2 | 7.1 | 7.9 | 7.4 | 6.1 | 8.1 | 7.1 | 9.1 | 9.4 | 0.7 |  | 5.9 | 6.2 |
| 17. P. axillaris | 13.9 | 12.8 | 13.0 | 12.5 | 13.0 | 13.9 | 13.9 | 12.5 | 17.1 | 13.9 | 15.4 | 14.3 | 13.5 | 13.5 | 13.6 | 13.2 |  | 5.4 |
| 18. P. mystaceus | 15.0 | 13.4 | 14.7 | 13.3 | 14.0 | 16.2 | 15.9 | 15.8 | 18.5 | 14.7 | 15.4 | 15.8 | 14.6 | 14.2 | 15.0 | 14.7 | 12.5 |  |

Note:
Below the diagonal is the COI sequence; 16S sequence above the diagonal.

All specimens were preserved in the Animal Herbarium of the College of Life Sciences, Xinjiang Agricultural University.
Type locality and distribution: Near Kangsu Town (39.80N, 74.73F, altitude: 2,717 m), Wuqia County, Kizilsu Kyrgyz Autonomous Prefecture, Xinjiang Uygur Autonomous Region, China.
Etymology: Type specimen locality.
Diagnosis: No axillary spots, two nasal scales, upper nasal scale bulging, nostril opening between two nasal scales; three inter-nasal scales, middle one largest, and bulging. Five to seven pairs of orange-red spots along the centre of the dorsal spine and a distinct orange-red spot on the dorsal surface of the caudal base. Dark transverse stripes on the dorsal surface of the tail, white ventral surface of the tail, and black tip.
Description of holotype (Fig. 6, Table 2): Adult males (CX09247), SVL $=38.85 \mathrm{~mm}$, $\mathrm{HL}=11.43 \mathrm{~mm}, \mathrm{HW}=11.50 \mathrm{~mm}, \mathrm{HD}=8.27 \mathrm{~mm}, \mathrm{RED}=3.61 \mathrm{~mm}, \mathrm{MB}$ (mouth breadth $)=10.56 \mathrm{~mm}, \mathrm{NBD}=1.69 \mathrm{~mm}$, LHF $($ trunk length $)=18.77 \mathrm{~mm}, \mathrm{TL}=50.12 \mathrm{~mm}$, LFL (length of forelimb) $=23.65 \mathrm{~mm}$, and LHL (length of hindlimb) $=35.30 \mathrm{~mm}$. Bluntly rounded head, with length and breadth not significantly different, and a short and blunt muzzle. One parietal eye with a translucent membrane in the centre surrounded by several irregular scales. Two nasal scales, with the upper nasal scale expanded into a kidney shape, a nostril opening between the two nasal scales, and the nostril opening slanting downwards; three inter-nasal scales, the middle one inflated and bulging and other two smaller and slightly attached to the supra-nasal scales; four scales between the nasal and


Figure 5 Divergence time estimates for Oreosaura. Note: Values represent estimated divergence time, and the blue bar represents $95 \%$ confidence interval HPD; WQ, Wuqia Group; HT, Hotan Group; AKQ, Akqi Group.

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preocular scales and 11 superciliary scales; squarer supralabial shields, with 13 on the left and 12 on the right, and three rows of scales separating the supralabial shields from the lower edge of the eye; 12 infralabial shields, flush with supralabial shields, and 39 chin scales in a longitudinal row in the throat fold.

Axillary is short and dorsal scales are mostly granular, with a few tufts of spiny scales; rhombic abdominal scales, pointed posterior spine corners without ridges, and black centre of abdomen. Short limbs; smooth dorsal scales of limbs; carinate on thigh and shin flanks; rhombic and smooth ventral scales of limbs; and ribbed sub-metacarpal scales. Relatively well-developed and unilateral toe fringe of toe IV of the hind limb. The tail is covered with granular scales, with a few prickly scales; the ventral surface of the basal end of the tail has conical scales and the caudal side is covered with prickly scales.

Grey dorsal surface, densely covered with white dots, with six pairs of orange-red spots along the centre of the dorsal spine from the neck to the base of the tail and clumps of spiny scales on the orange-red spots. No axillary spots; black dots distributed on the dorsal surface of the limbs, particularly on the palms and toes. A distinct orange-red spot on the dorsal surface of the caudal base, nine dark transverse spots on the dorsal surface of the tail


Figure 6 Pictures of Phrynocephalus kangsuensis sp. nov. (A, C, F) CX09246; (B, D, E) CX09247. Full-size DOI: 10.7717/peerj.17175/fig-6

Table 2 Some measurements (mm) and features of pholidosis of Phrynocephalus kangsuensis sp. nov.

| Numble | sex | $\mathbf{1}$ | $\mathbf{2}$ | $\mathbf{3}$ | $\mathbf{4}$ | $\mathbf{5}$ | $\mathbf{6}$ | $\mathbf{7}$ | $\mathbf{8}$ | $\mathbf{9}$ | $\mathbf{1 0}$ | $\mathbf{1 1}$ | $\mathbf{1 2}$ |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| CX09246 | F | 36.41 | 10.24 | 10.99 | 7.42 | 2.59 | 1.47 | 39.11 | 2 | 3 | $13 / 14$ | $13 / 13$ | 3 |
| CX09245 | M | 39.91 | 12.34 | 11.62 | 7.85 | 3.13 | 1.91 | 49.81 | 2 | 3 | $13 / 13$ | $12 / 13$ | 3 |
| CX09243 | M | 37.65 | 10.91 | 10.38 | 7.25 | 2.53 | 1.79 | 43.73 | 2 | 3 | $13 / 12$ | $12 / 12$ | 3 |
| CX09248 | M | 37.57 | 10.98 | 11.07 | 7.91 | 2.85 | 1.84 | 48.44 | 2 | 3 | $12 / 14$ | $12 / 13$ | 3 |
| CX09244 | M | 38.17 | 11.28 | 10.73 | 7.13 | 2.48 | 1.67 | 48.37 | 2 | 3 | $13 / 14$ | $13 / 13$ | 4 |
| CX09247 | M | 38.85 | 11.43 | 11.50 | 8.27 | 3.61 | 1.69 | 50.12 | 2 | 3 | $13 / 12$ | $12 / 12$ | 3 |

Note:
$\quad$ F, female; M, male; 1, SVL; 2, HL; 3, HW; 4, HD; 5, RED; 6, NBD; 7, TL; 8, NS; 9, IS; 10, supralabial shields (left/right); 11, infralabial shields (left/right); 12, number of scales between lower border of eye and row of supralabial shields.
derived from the end of the tail, ventral surface white, faintly reddish, and the tip of the tail black.
Variation among Paratypes (Table 2): SVL $=36.41-39.91 \mathrm{~mm} ; \mathrm{HL}=10.24-12.43 \mathrm{~mm}$; $\mathrm{HW}=10.38-11.62 \mathrm{~mm} ; \mathrm{HD}=7.13-7.91 \mathrm{~mm} ; \mathrm{RED}=2.48-3.13 \mathrm{~mm} ; \mathrm{MB}=9.09-10.63$ $\mathrm{mm} ;$ NBD $=1.47-1.91 \mathrm{~mm} ;$ LHF $=16.85-17.77 \mathrm{~mm} ; \mathrm{TL}=43.73-49.81 \mathrm{~mm}$; LFL $=20.12-22.16 \mathrm{~mm}$; and $\mathrm{LHL}=29.72-32.70 \mathrm{~mm}$. There were $5-7$ pairs of orange spots


Figure 7 Type locality of Phrynocephalus kangsuensis sp. nov.
Full-size
DOI: 10.7717/peerj.17175/fig-7
on the back. Adult female (CX09246) had a white ventral surface without black spots and 3-5 scales between the nasal and preocular scales, 11-12 superciliary scales, 12-14 supralabial scales, 12-13 infralabial scales, 3-4 scales between the infraocular margin and supralabial scales, and 6-9 dark transverse spots on the dorsal surface of the tail.
Comparison with other species: This species exhibits morphological differences
compared with those of $P$. forsythii and P. nasatus. The relatively widely spaced snout and distinctly inflated and raised inter-nasal scales of $P$. kangsuensis sp. nov. are different from those of the other two species. Second, the pattern of spots on the dorsal surface and the transverse stripe on the dorsal surface of the tail are important features for distinguishing the three species. Phrynocephalus kangsuensis sp. nov. and P. nasatus have outwards-facing nostrils with a relatively wide nasal spacing. In addition, P. nasatus has 5-6 small inter-nasal scales, while $P$. kangsuensis sp. nov. has three inter-nasal scales but the middle one is significantly larger and raised. In contrast, $P$. forsythii has a relatively small nasal spacing, with 3-4 inter-nasal scales of equal size. Phrynocephalus forsythii and P. nasatus have distinct 4-5 pairs of black dorsal blotches, whereas P. kangsuensis sp. nov. has 5-7 pairs of orange-red dorsal blotches. The dorsal surface of the tail of $P$. kangsuensis sp. nov. has 6-9 dark transverse stripes, while that of $P$. nasatus has distinct 8-9 black transverse stripes and $P$. forsythii has no dark transverse stripes.

Ecology and Environment (Fig. 7): This species lives in valleys and is a typical desert animal. The habitat is located in desert flatlands between high mountains. It is a typical arid Gobi environment with a substrate of sandy gravel covered with short grasses.

## DISCUSSION

## Phylogenetic trees

Viviparous species in Phrynocephalus are considered to be the subgenus Oreosaura (Barabanov \& Ananjeva, 2007), which includes Phrynocephalus spp. distributed at high altitudes. Based on geographic distribution and morphological differences, Oreosaura can be divided into three species: P. forsythii (P. forsythii and P. nasatus), P. vlangalii (P. vlangalii, P. guinanensis, P. putjatia, and P. nanschanicus), and P. theobaldi (P. theobaldi, P. erythrurus, P. lhasaensis, and P. parvus) (Solovyeva et al., 2023).

In the viviparous species group, only P. forsythii was distributed on the periphery of the Tarim Basin at lower elevations in southern Xinjiang. As the P. forsythia complex contains the only viviparous Phrynocephalus species distributed at low altitudes, it has received constant attention from researchers (Qi et al., 2019; Chen, Li \& Guo, 2019; Qi et al., 2020, 2021, 2023). Based on the phylogenetic tree constructed by Solovyeva et al. (2023) for Phrynocephalus using COI sequences, we added molecular data from three valleys geographic populations of $P$. forsythii to explore their taxonomic position in the subgenus Oreosaura. Our results showed a topology similar to that reported by Solovyeva et al. (2023). Within the P. forsythii complex, the three valleys geographic populations (groups $\mathrm{WQ}, \mathrm{HT}$, and AKQ ) formed three clades with strong support and sister clades to the reference sequences from lower altitudes (Qiemo, Ruoqiang, Kashgar, and Aksu regions). Simultaneously, P. nasatus, a lizard collected from Baicheng County, was embedded in P. forsythia, and its status needs to be further studied. Second, we added 16 S sequences and analyses of spliced sequences from the two datasets as a supplement and the results were consistent with the phylogenetic tree of the COI sequences, with the three valleys populations forming strongly supported clades. Genetic differentiation among several populations in the $\sim 3,000 \mathrm{~km}$ ring of the Tarim Basin was extremely low, with significant differences from populations in the closer high-altitude alpine valleys. Geographic populations distributed at valleys were genetically differentiated from populations at plains altitudes.

## Genetic distances

Solovyeva et al. (2023) analysed the genetic distances of COI sequences of all species of Phrynocephalus and found that the genetic distances between the P. forsythii complex and other species in the subgenera were $7.18-8.75 \%$ and with $P$. axillaris and $P$. mystaceus were $10.59 \%$ and $12.42 \%$, respectively, while the genetic distances between different subgenera exceeding $10 \%$. This was based on the results of Solovyeva et al. (2023) and their criteria for genetic differences within species (subgenus $>10 \%$, interspecies $>5 \%$ and intraspecies $<3 \%$ ). Among them, the genetic distance between the Wuqia population (Phrynocephalus kangsuensis sp. nov.) and other populations within the P. forsythii species group reached interspecific differences (>5\%), and that between other populations within the $P$. forsythii
complex (Group HT, Group AKQ, and low-altitude populations) was in line with the results of Qi et al. (2019) for various geographic P. forsythii populations (0.2-5.1\%). We calculated and analysed the genetic distances of the 16 S rRNA sequences and found that they were lower than those of the COI sequences; however, the results also showed significant genetic differences between the Wuqia population and other populations of the P. forsythia complex.

## Species delimitation

The three species delimitations used in this study yielded different results, with ABGD and ASAP delimiting the most similar MOTUs and bPTP delineating additional species. Solovyeva et al. (2023) used five methods of species delimitation to define species delimitation in Phrynocephalus. They concluded that the ASAP algorithm best reflected the current classification of Phrynocephalus and that bPTP grossly overestimated the number of species. Some studies on species delineation recognised that ABGD and ASAP are more conservative and accurate in delineating species, whereas bPTP suffers from over delineation (Blair \& Bryson, 2017; Goulpeau et al., 2022; Guo \& Kong, 2022; Ranasinghe et al., 2022). In the present study, the results of the ABGD and ASAP analyses were consistent with those of Solovyeva et al. (2023). Therefore, the results of our delimitation exhibited a high degree of confidence that the Wuqia and Hotan populations in the P. forsythii complex may be hidden species.

## Divergence date estimation

In Oreosaura, the P. forsythii complex diverged first, followed by the $P$. vlangalii complex and P. theobaldi complex, which may be related to the evolutionary relationships of P. forsythii and the timing of its geographic occurrence (Guo \& Wang, 2007; Jin \& Brown, 2013; Solovyeva et al., 2018, 2023). Guo \& Wang (2007) suggested that the viviparous species group diverged at $9.12 \pm 1.44 \mathrm{Ma}$; Jin $\Leftarrow$ Brown (2013) suggested that the oviparous and viviparous species groups diverged at 9.73 Ma . The results of the divergence time estimation in this study were similar to those of two previous (Guo \& Wang, 2007; Jin \& Brown, 2013). Solovyeva et al. (2018) suggested that the formation of the subgenus Oreosaura was associated with the rapid uplift of the Qinghai-Tibetan Plateau (QTP) and the associated evolution of the East Asian monsoon climate and that the group of viviparous species lived on the QTP from $13.5-10.0 \mathrm{Ma}$, with differentiation beginning at 3.8 Ma . The timing of the divergence of the viviparous species group was closely related to the uplift of the QTP (Macey et al., 2018) and the formation of mountain basins on the QTP at 5.7 Ma may have contributed to significant climate change (Li et al., 1996). In addition, the rapid uplift of the Tibetan region at approximately 5 Ma (Shackleton \& Chengfa, 1988) may have led to the differentiation of $P$. theobaldi complex and $P$. vlangalii complex distributed on the QTP, with the uplift of the Himalayas and the Kunlun Mountains contributing to the formation of species in both species complexes. In their biogeographic study of the P. theobaldi complex, Jin, Liu \& Brown, 2017 suggested that the first species divergence of $P$. theobaldi occurred during a possible period of tectonic uplift in the Himalayan region (3.6-7.4 Ma). Jin, Brown \& Liu, 2008 suggested that the
divergence of the $P$. vlangalii complex was caused by three major periods of plateau uplift in the QTP ( $3.4 \mathrm{Ma}, 2.5 \mathrm{Ma}$, and 1.7 Ma ), and the results of the present study support these inferences.

Solovyeva et al. (2018) suggested that the progression of aridification, basin reduction, orogeny, and Upper Pleistocene climatic oscillations in the late Miocene may have contributed to the further diversification of Phrynocephalus. The divergence of the P. forsythii complex is closely related to the uplift of the QTP, Kunlun Mountains, and Tian Shan Mountains. Guo \& Wang (2007) suggested that the P. forsythii complex diverged at $4.35 \pm 0.65 \mathrm{Ma}$, while Jin \& Brown (2013) suggested that an earlier collapse of the interior of the QTP, which impacted the environment north of the QTP, led to the separation of the P. forsythii complex from the other two in the viviparous species group at 5.04 Ma . The present study suggests that the P. forsythii complex segregated at $6.73 \mathrm{Ma}(95 \%$ confidence interval: $4.75-8.76 \mathrm{Ma}$ ), which may be related to climate change in the Tarry Basin during the period of $\sim 5-7 \mathrm{Ma}$ (Sun et al., 2017).

Phrynocephalus kangsuensis sp. nov. is found in the northwestern part of the QTP, bordered by the southern foothills of the Tien Shan and the Western Kunlun and Pamir Plateau knots. It inhabits habitats in the Kangsu Valley, which are different from populations living in the lower-altitude plains. The major uplift of the northwest part of the QTP at 4.5 Ma (Zheng et al., 2000) and its mutual collision with the Pamir Plateau led to the formation of mountains in the NW part of the QTP, which formed a distinct high and low relief with the Tarim Basin; this may have led to the separation of P. kangsuensis sp. nov. from other geographic populations.

The Hotan population is located on the northern slopes of the Kunlun Mountains in a high-altitude montane habitat (Karakash Valley). The estimated divergence time for this population is $2.92 \mathrm{Ma}(95 \%$ HPD: $1.75-4.16 \mathrm{Ma}$ ), which may be related to the rapid uplift of the Middle Kunlun area on the northern margin of the QTP since the late Pliocene (4.2-3.9 Ma). The rapid uplift of the Kunlun region led to the vertical geographic isolation of this population from lower-altitude populations, which ultimately led to population differentiation over time.

The Central Kunlun area on the northern margin of the QTP also experienced an uplift in the middle Early Pleistocene ( $1.66 \pm 0.31 \mathrm{Ma}$ ) (Bai et al., 2003). Second, the uplift of the Tien Shan Mountains during the Early Pleistocene formed the present-day alpine topography (Sun, Zhu \& Bowler, 2004), which coincides with the timing of species divergence in the AKQ population (Toshkan Valley) and is situated in high-altitude mountainous terrain. The Quaternary climate history of the QTP is characterised by four major ice ages, the most extensive of which was the Naynayxungla Ice Age ( $0.50-0.72 \mathrm{Ma}$ ) (Zheng, Xu \& Shen, 2002). At the time, many large ice caps, glacial complexes, and grand canyon glaciers formed, which may have facilitated further differentiation of the different alpine valley P. forsythii complexes. Qi et al. (2021) analysed low-altitude populations in the $P$. forsythii complex and concluded that, in addition to the uplift of the plateau, the rivers and oases of the Tarim Basin and climatic variations, had a marked effect on population differentiation. Since the late Pliocene, Increased aridity led to changes in the vegetation of the Tarim Basin and rainfall and snowmelt on mountaintops have caused
oases to appear in some of the foothill plains, allowing low-altitude populations of P. forsythii to form a ring around the Tarim Basin (Qi et al., 2020).

Since we did not collect samples from lower altitudes. A limitation of this study is that there was not a large amount of molecular data on P. forsythii from low altitudes. Only three of the most representative methods of species delimitation were used. In future studies, more populations of $P$. forsythii from plains could be added and more methods of species delimitation could be used to define the species of the P. forsythii complex to allow the specific effects of geological history on P. forsythii complex to be studied in further detail.

## CONCLUSIONS

Here, we constructed a COI phylogenetic tree of Phrynocephalus based on Solovyeva et al. (2023) with additional data. In addition, we constructed a phylogenetic tree of 16 S sequences to explore species affinities and divergence events among Oreosaura, including within the species complex, as well as between three populations of $P$. forsythii from high-altitude geographic populations.

The results indicate that Oreosaura is a strongly supported monophyletic lineage that diverged at $10.43 \mathrm{Ma}(95 \%$ HPD: [10.00-11.33] Ma), while the P. forsythii complex diverged at $6.73 \mathrm{Ma}(95 \%$ HPD: [4.75-8.76] Ma). The results confirmed that high-altitude valleys promoted the geographic isolation of the $P$. forsythii complex, ultimately leading to species divergence and promoting the formation of new species. The Wuqia population is geographically isolated from the other populations, which leads to genetic differentiation. The molecular results indicated that the Wuqia population is a strongly supported monophyletic lineage with an earlier estimated divergence time than the other populations, at 3.96 Ma ( $95 \% \mathrm{HPD}$ : [2.57-5.40] Ma). In addition, this population showed significant differences in morphology.

This study combined molecular and morphological aspects into an integrated taxonomic approach and identified the Wuqia population as a new species, Phrynocephalus kangsuensis sp. nov.

The results will provide data for phylogenetic studies following the P. forsythii complex and data to confirm that valleys promote the formation of species differentiation.

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## ADDITIONAL INFORMATION AND DECLARATIONS

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## Competing Interests

The authors declare that they have no competing interests.

## Author Contributions

- Qianru Liang conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Lei Shi conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.


## Animal Ethics

The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):

All experimental procedures involving animals were approved (animal protocol number: 2023013) by the Animal Welfare and Ethics Committee of Xinjiang Agricultural University, Urumqi, Xinjiang, China.

## DNA Deposition

The following information was supplied regarding the deposition of DNA sequences:
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## New Species Registration

The following information was supplied regarding the registration of a newly described species:

Publication LSID: urn:lsid:zoobank.org:pub:812FA766-6BC6-4D39-85B9-

## 8E2C58E06098

Phrynocephalus kangsuensis sp. nov. phrynocephalus kangsuensis sp. nov. Qianru \& Shi, LSID: urn:lsid:zoobank.org:act:53EF51A0-DEEA-4E6A-B62F-AD61AB3868E9

## Supplemental Information

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## REFERENCES

Bai DY, Meng DB, Liu YR, Ma TQ. 2003. Apatite fission-track records of the tectonic uplift of the central segment of the Kunlun Mountains on the northern margin of the Qinghai-Tibet Plateau. China Geology in English 30:240-246.
Barabanov AV, Ananjeva NB. 2007. Catalogue of the available scientific species-group names for lizards of the genus Phrynocephalus Kaup, 1825 (Reptilia, Sauria, Agamidae). Zootaxa 1399(1):1-56 DOI 10.11646/zootaxa.1399.1.1.
Blair C, Bryson RW Jr. 2017. Cryptic diversity and discordance in single-locus species delimitation methods within horned lizards (Phrynosomatidae: Phrynosoma). Molecular Ecology Resources 17(6):1168-1182 DOI 10.1111/1755-0998.12658.
Bogdanov A, Tambovtseva V, Matveevsky S, Bakloushinskaya I. 2023. Speciation on the roof of the world: parallel fast evolution of cryptic mole vole species in the Pamir-Alay-Tien shan region. Life 13(8):1751 DOI 10.3390/life13081751.
Bouckaert R, Heled J, Kühnert D, Vaughan T, Wu CH, Xie D, Suchard MA, Rambaut A, Drummond AJ. 2014. BEAST 2: a software platform for Bayesian evolutionary analysis. PLOS Computational Biology 10(4):e1003537 DOI 10.1371/journal.pcbi. 1003537.
Burland TG. 1999. DNAStar's Lasergene sequence analysis software. Bioinformatics Methods and Protocols 132:71-91 DOI 10.1385/1-59259-192-2:71.
Charreau J, Chen Y, Gilder S, Barrier L, Dominguez S, Augier R, Sen S, Avouac J, Gallaud A, Graveleau F, Wang Q. 2009. Neogene uplift of the Tian Shan Mountains observed in the magnetic record of the Jingou River section (Northwest China). Tectonics 28(2):1791 DOI 10.1029/2007TC002137.
Chen D, Li J, Guo X. 2019. Next-generation sequencing yields a nearly complete mitochondrial genome of the Forsyth's toad-headed agama, Phrynocephalus forsythii (Reptilia, Squamata, Agamidae). Mitochondrial DNA Part B 4(1):817-819 DOI 10.1080/23802359.2019.1574681.
Coyne JA, Orr HA. 2004. Speciation. Vol. 276. Sunderland, MA: Sinauer Associates, 281.
Cui ZJ, Gao QZ, Liu GN, Pan BT, Chen HL. 1996. Planation surfaces, palaeokarst and uplift of Xizang (Tibet) Plateau. Science in China (Earth Sciences) 4:391-400.
Dunayev EA. 2020. History of study, taxonomy, distribution, and ecology of Phrynocephalus nasatus Golubev ET Dunayev, 1995 (Reptilia: Agamidae). Russian Journal of Herpetology 27(2):87-96 DOI 10.30906/1026-2296-2020-27-2-87-96.
Golubev ML, Dunayev EA. 1995. Phrynocephalus nasatus (Reptilia, Agamidae), a new species of toad agama from Western China. Russian Journal of Herpetology 2:5-9.
Goulpeau A, Penel B, Maggia ME, Marchán DF, Steinke D, Hedde M, Decaëns T. 2022. OTU delimitation with earthworm DNA barcodes: a comparison of methods. Diversity 14(10):866 DOI 10.3390/d14100866.
Guo B, Kong L. 2022. Comparing the efficiency of single-locus species delimitation methods within Trochoidea (Gastropoda: Vetigastropoda). Genes 13(12):2273 DOI 10.3390/genes13122273.
Guo X, Wang Y. 2007. Partitioned Bayesian analyses, dispersal-vicariance analysis, and the biogeography of Chinese toad-headed lizards (Agamidae: Phrynocephalus): a re-evaluation. Molecular Phylogenetics and Evolution 45(2):643-662 DOI 10.1016/j.ympev.2007.06.013.
Jin YT, Brown RP. 2013. Species history and divergence times of viviparous and oviparous Chinese toad-headed sand lizards (Phrynocephalus) on the Qinghai-Tibetan Plateau. Molecular Phylogenetics and Evolution 68(2):259-268 DOI 10.1016/j.ympev.2013.03.022.

Jin YT, Brown RP, Liu NF. 2008. Cladogenesis and phylogeography of the lizard Phrynocephalus vlangalii (Agamidae) on the Tibetan Plateau. Molecular Ecology 17(8):1971-1982 DOI 10.1111/j.1365-294X.2008.03721.x.
Jin Y, Liu N, Brown RP. 2017. The geography and timing of genetic divergence in the lizard Phrynocephalus theobaldi on the Qinghai-Tibetan Plateau. Scientific Reports 7:2281 DOI 10.1038/s41598-017-02674-4.

Kadmon R, Pulliam HR. 1993. Island biogeography: effect of geographical isolation on species composition. Ecology 74(4):977-981 DOI 10.2307/1940467.
Kumar S, Stecher G, Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular Biology and Evolution 33(7):1870-1874 DOI 10.1093/molbev/msw054.
Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. 2017. PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. Molecular Biology and Evolution 34:772-773 DOI 10.1093/molbev/msw260.
Letunic I, Bork P. 2021. Interactive tree of life (iTOL) v5: an online tool for phylogenetic tree display and annotation. Nucleic Acids Research 49(W1):W293-W296 DOI 10.1093/nar/gkab301.
Li JJ, Fang XM, Ma HZ, Pan BT, Chen HL. 1996. Geomorphological and environmental evolution in the upper reaches of the Yellow River during the Late Cenozoic. Science in China, Series D: Earth Sciences 39:380-390.
Macey JR, Schulte JAI, Ananjeva NB, Van Dyke ETV, Wang Y, Orlov N, Shafiei S, Robinson MD, Dujsebayeva T, Freund GS, Fischer CM, Liu D, Papenfuss TJ. 2018. A molecular phylogenetic hypothesis for the Asian agamid lizard genus Phrynocephalus reveals discrete biogeographic clades implicated by plate tectonics. Zootaxa 4467(1):1-81 DOI 10.11646/zootaxa.4467.1.1.
Mayr E. 1970. Populations, species, and evolution: an abridgment of animal species and evolution. Vol. 19. Harvard University Press.
Nagy ZT, Sonet G, Glaw F, Vences M. 2012. First large-scale DNA barcoding assessment of reptiles in the biodiversity hotspot of madagascar, based on newly designed COI primers. PLOS ONE 7(3):e34506 DOI 10.1371/journal.pone.0034506.
Nazarov RA, Abduraupov TV, Shepelya EY, Gritsina MA, Melnikov DA, Buehler MD, Lapin JD, Poyarkov NA, Grismer JL. 2023. The Fergana Valley is an isolate of biodiversity: a discussion of the endemic herpetofauna and description of two new species of Alsophylax (Sauria: Gekkonidae) from Eastern Uzbekistan. Animals 13(15):2516 DOI 10.3390/ani13152516.
Nosil P, Harmon LJ, Seehausen O. 2009. Ecological explanations for (incomplete) speciation. Trends in Ecology and Evolution 24(3):145-156 DOI 10.1016/j.tree.2008.10.011.
Pang J, Wang Y, Zhong Y, Rus Hoelzel AR, Papenfuss TJ, Zeng X, Ananjeva NB, Zhang YP. 2003. A phylogeny of Chinese species in the genus Phrynocephalus (Agamidae) inferred from mitochondrial DNA sequences. Molecular Phylogenetics and Evolution 27(3):398-409 DOI 10.1016/S1055-7903(03)00019-8.
Pavlicev M, Mayer W. 2009. Fast radiation of the subfamily Lacertinae (Reptilia: Lacertidae): history or methodical artefact? Molecular Phylogenetics and Evolution 52(3):727-734 DOI 10.1016/j.ympev.2009.04.020.
Puillandre N, Brouillet S, Achaz G. 2021. ASAP. ASAP: assemble species by automatic partitioning. Molecular Ecology Resources 21(2):609-620 DOI 10.1111/1755-0998.13281.

Puillandre N, Lambert A, Brouillet S, Achaz GJME. 2012. ABGD, automatic barcode gap discovery for primary species delimitation. Molecular Ecology 21(8):1864-1877 DOI 10.1111/j.1365-294X.2011.05239.x.
Qi Y, Ding L, Zhao Y, Niu C, Wang X, Zhao W. 2020. Toad-headed lizard Phrynocephalus forsythii (Squamata, Agamidae) as a potential ring species inferred from population genetic differentiation. Asian Herpetological Research 11:312-319D DOI 10.16373/j.cnki.ahr. 200001.
Qi Y, Zhao W, Huang Y, Wang X, Zhao Y. 2019. Correlation between climatic factors and genetic diversity of Phrynocephalus forsythii. Asian Herpetological Research 10:270-275A DOI 10.16373/j.cnki.ahr. 190028.
Qi Y, Zhao W, Li Y, Zhao Y. 2021. Environmental and geological changes in the Tarim Basin promoted the phylogeographic formation of Phrynocephalus forsythii (Squamata: Agamidae). Gene 768(14):145264 DOI 10.1016/j.gene.2020.145264.
Qi Y, Zhao W, Zhao Y, Niu C, Cao S, Zhang Y. 2023. Chromosome-level genome assembly of Phrynocephalus forsythii using third-generation DNA sequencing and Hi-C analysis. DNA Research 30(2):dsad003 DOI 10.1093/dnares/dsad003.
Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. Systematic Biology 67(5):901-904 DOI 10.1093/sysbio/syy032.
Ranasinghe UGSL, Eberle J, Thormann J, Bohacz C, Benjamin SP, Ahrens D. 2022. Multiple species delimitation approaches with COI barcodes poorly fit each other and morphospecies an integrative taxonomy case of Sri Lankan Sericini chafers (Coleoptera: Scarabaeidae). Ecology and Evolution 12(5):e8942 DOI 10.1002/ece3.8942.
Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3):539-542 DOI 10.1093/sysbio/sys029.
Shackleton RM, Chengfa C. 1988. Cenozoic uplift and deformation of the Tibetan Plateau: the geomorphological evidence. Philosophical Transactions of the Royal Society of London, Series A: Mathematical and Physical Sciences 327:365-377 DOI 10.1098/rsta.1988.0134.
Solovyeva EN, Dunayev EA, Nazarov RA, Bondarenko DA, Poyarkov NA. 2023. COI-barcoding and species delimitation assessment of toad-headed Agamas of the genus Phrynocephalus (Agamidae, Squamata) reveal unrecognized diversity in Central Eurasia. Diversity 15(2):149 DOI 10.3390/d15020149.
Solovyeva EN, Lebedev VS, Dunayev EA, Nazarov RA, Bannikova AA, Che J, Murphy RW, Poyarkov NA. 2018. Cenozoic aridization in Central Eurasia shaped diversification of toad-headed agamas (Phrynocephalus; Agamidae, Reptilia). PeerJ 6(6608):e4543 DOI 10.7717/peerj. 4543.
Sun J, Liu W, Liu Z, Deng T, Windley BF, Fu B. 2017. Extreme aridification since the beginning of the Pliocene in the Tarim Basin, western China. Palaeogeography, Palaeoclimatology, Palaeoecology 485(B7):189-200 DOI 10.1016/j.palaeo.2017.06.012.
Sun J, Zhu R, Bowler J. 2004. Timing of the Tianshan Mountains uplift constrained by magnetostratigraphic analysis of molasse deposits. Earth and Planetary Science Letters 219(3-4):239-253 DOI 10.1016/S0012-821X(04)00008-1.
Wang K, Gao W, Wu J, Dong W, Feng X, Shen W, Jin J, Shi X, Qi Y, Siler CD, Che J. 2021. Two new species of Diploderma Hallowell, 1861 (Reptilia: Squamata: Agamidae) from the Hengduan Mountain Region in China and Rediscovery of D. brevicaudum (Manthey, Wolfgang, Hou, Wang, 2012). Zootaxa 4941(1):143-158 DOI 10.11646/zootaxa.4941.1.1.

Wang K, Zhou XH, Liu D, Li Y, Yao Z, He WM, Liu Y. 2022. The uplift of the Hengduan Mountains contributed to the speciation of three Rhododendron species. Global Ecology and Conservation 35:e02085 DOI 10.1016/j.gecco.2022.e02085.
Winger BM, Bates JM. 2015. The tempo of trait divergence in geographic isolation: avian speciation across the Marañon Valley of Peru. Evolution 69(3):772-787 DOI 10.1111/evo.12607.
Zhang D, Gao F, Jakovlić I, Zou H, Zhang J, Li WX, Wang GT. 2020. PhyloSuite: an integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. Molecular Ecology Resources 20(1):348-355 DOI 10.1111/1755-0998.13096.
Zhang J, Kapli P, Pavlidis P, Stamatakis A. 2013. A general species delimitation method with applications to phylogenetic placements. Bioinformatics 29:2869-2876 DOI 10.1093/bioinformatics/btt499.
Zhao EM, Zhao KT, Zhou KY. 1999. Fauna Sinica Reptilia vol. 2 Squamata. Lacertilia. Beijing: Beijing Science Press.
Zheng H, Powell CM, An Z, Zhou J, Dong G. 2000. Pliocene uplift of the northern Tibetan Plateau. Geology 28:715-718 DOI 10.1130/0091-7613(2000)28<715:PUOTNT>2.0.CO;2.
Zheng B, Xu Q, Shen Y. 2002. The relationship between climate change and quaternary glacial cycles on the Qinghai-Tibetan Plateau: review and speculation. Quaternary International 97-98:93-101 DOI 10.1016/S1040-6182(02)00054-X.

