Discovery of a peculiar insular race of *Ravenna nivea* (Nire, 1920) (Lepidoptera: Lycanidae) endemic to Yinggeling Mountain of Hainan, suggesting heterogeneous geographical history of mountain formation of the island

YU-FENG HSU1, YIK FUI PHILIP LO2, RUNG-JUEN LIN3

1Department of Life Sciences, National Taiwan Normal University, Taipei, Taiwan 116, R. O. C. E-mail: t43018@ntnu.edu.tw
2Kadoorie Conservation China, Kadoorie Farm and Botanic Garden, Lam Kam Road, Tai Po, New Territories, Hong Kong. E-mail: philiplo@kfbg.org
3RUNG-JUEN LIN

Department of Pediatrics and Medical Genetics, National Taiwan University Hospital,
Taipei 10041, Taiwan, R. O. C. E-mail: rung.juen@gmail.com

Corresponding Author:

YU-FENG HSU

88, Sec. 4, Tingzhou Road, Taipei, Taiwan, 116, Taiwan, ROC.

Email address: t43018@ntnu.edu.tw

Abstract

A peculiar population of *Ravenna nivea* (Nire, 1920) was discovered from the Yinggeling Mountain Mass of central Hainan, China. Its wing pattern and COI barcode data show considerable distinction from other geographic populations of *R. nivea*, including that of Bawangling, approximately only 40 km away and also located in Hainan. The p-distance value of the COI barcode between the Yinggeling and Bawangling populations was 1.1%, considerably higher than the value (0.6%) between Bawangling population and populations in eastern China, where the subspecific name *howarthi* Saigusa, 1993 applies. The population is regarded as a distinct subspecies *nigrolineata* Lo & Hsu, *subsp. nov*. The distinctness and high degree of COI haplotype diversity of *R. nivea* found in Hainan and Taiwan suggest continental islands may serve as glacial refugia for the butterfly and other organisms during previous glaciations, and the presence of the relict populations of montane butterflies like *R. nivea* may provide useful clues towards a better understanding of the geological history of mountain formation within islands.

Key words: Zephyrus, hairstreak, Theclinae, Theclina, butterfly, endemism, glacial relict

Introduction

The subspecies concept is one of the most controversial within Linnean taxonomy (Shiaflini, 2020). Although the controversy and utility of the concept has been continuously debated (e.g. Hillis, 2021; Queiroz, 2021; Burbrink et al., 2022), it is recognized as a legitimate species-level rank by the International Code of the Zoological Nomenclature (ICZN), currently in the 4th version. The subspecies concept is widely applied to many groups of organisms, including butterflies for which a scheme for hypothesis testing of taxonomic status of allopatric populations has been proposed (Braby et al., 2012). In this scheme, the null hypothesis of a single species (with one or more subspecies) is the default hypothesis and is rejected only if evidence from other multiple data sources (color pattern, morphology, behavior, ecology, genetics, etc.) supports the alternative hypothesis of lineage divergence and monophyly. We discovered an intriguing case in lycaenid butterflies in the genus *Ravenna*, in which a population
The genus *Ravenna* Shirayu & Yamamoto, 1956 represents a monobasic genus of hairstreak butterflies, containing the sole species *R. nivea* (Nire, 1920). This species inhabits the northern Oriental region, with four subspecies currently recognized: nominotypical *nivea* Nire, 1920 from Taiwan, *howarthi* Saigusa, 1993 from East to West China, *koiwayai* Yoshino, 1997 from Mt. Konga of Sichuan, and *miyagawai* Katayama & Saito, 2011 from Vietnam. A second species, *R. pacifica* Dubatolov & Korshunov, 1984 was described from Far East Russia but was later transferred to a newly established monobasic genus *Goldia* (Dubatolov & Korshunov, 1990). *Ravenna nivea* is a relatively large species in the tribe Theclini sensu Eliot (1973), usually characterized by the prominent sexual dimorphism of upperside wing pattern, with mostly purple with white markings in males, in contrast to white with distal dark brown margins and markings in females (Figs 1A,1B). The wing undersides lack sexual dimorphism, with ground color white, decorated with dark brown bands (Fig. 1C,1D). An interesting form, with no white marking on wing upperside of male and purplish scalings extensively overlaid on white area of wing upperside of female (Figs 1E,1H), was discovered from Yinggeling of central Hainan Island. The peculiar appearance of this form is distinct from populations found elsewhere, calling into question on its appropriate systematic status.

We observed immatures, examined adult genitalia, and sequenced the DNA barcode (COI) of this hairstreak population to determine identity of this hairstreak. Subsequently we came to conclude that this population represents a distinct insular population of *Ravenna nivea* with unique diagnosable features. It is recognized as a new and second insular subspecies of *R. nivea* herein.

### Materials & Methods

**Acquisition of material**

Adult butterflies in question were collected from mountain slopes of Yinggeling, Hainan (109°112273-109°342063, 18°492302-19°082413), **China**. Ova were found near dormant buds of the hostplants. Larvae were provided with foliage of the hostplants. Rearing was performed using plastic containers 8 x 5.5 x 3 cm in size, with each larva kept separately.

Besides samples of *Ravenna* collected from Yinggeling, specimens of *R. nivea* from the following localities were examined and compared for morphological characters: ssp. *nivea*, ?1f1f (Taiwan); ssp. *howarthi*, 16f2f (2f2f, Guangdong, 4f, Jiangxi, 12f1f, Hainan [Bawangling], 1f1f, Fujian, 2f, Zhejiang, 3f5f, Sichuan, 7f1f, Guizhou); ssp. *miyagawai*, 4f5f (Vietnam). Of them, exemplars of taxa used for sequencing of the COI barcode are given in Table 1. COI barcodes of *Leucantigius atayalicus* and *Yamamotozephyrus kwangtangensis* were obtained from NCBI as reference outgroups (Table S1). Ssp. *koiwayai*, characterized by a reduction of tornal orange markings on hindwing undersides (Yoshino, 1997), is only known from its type locality, Mt. Konga of Sichuan in western China (Koiwaya, 2007), and was not available for the present study. The voucher specimens used for comparison listed above are currently stored in **Department of Biology, National Taiwan Normal University, Taipei (NTNU)**.
Butterfly identification was conducted based on morphological characteristics. The tissue samples were preserved in 95% ethanol and stored frozen at -20°C for further DNA extraction. Taxa listed in Table S1.

DNA extraction and sequencing

Tissue from two adult legs was digested using the DNeasy Blood & Tissue kit (Qiagen, Valencia, CA) following the manufacturer's protocol. The COI gene was amplified by PCR using a set of primers (Table S2). The amplification program was: 5 min at 94°C, 40 cycles of 30s at 94°C, 30s at 45-50°C, and 1 min at 72°C, and a final elongation step of 10 min at 72°C. The PCR products were run on 1.0% agarose gels in 1x TBE buffer to ensure correct amplification. PCR products were cleaned using a Gel/PCR DNA Fragments Extraction kit (Geneaid, Taipei, Taiwan) when only a single DNA band was visible in a gel.

Sequencing reactions were conducted using a 96-well Gel/PCR Clean Up kit (Geneaid) on an ABI3730XL DNA Analyzer (Applied Biosystems, Waltham, Massachusetts, USA). All DNA sequences have been submitted to the NCBI GenBank and accession numbers are given in Table S1.

Molecular data analyses

Molecular sequences of COI gene were checked and assembled into contigs using Sequencher 4.10 (GeneCode, Boston, USA). The software MEGA6 (Tamura et al., 2013) was used to perform sequence alignment using the MUSCLE method and sequence divergence using the Kimura2-Parameters (K2P) method. The partition schemes were determined with PartitionFinder v.2.1.1 (Lanfear et al., 2017). The data matrices were analysed using Maximum likelihood (ML) with RAxML and Bayesian inference (BI) with MrBayes on CIPRES (Miller et al., 2010). Leucantigius atayalicus and Yamamotozephyrus kwangtungensis were chosen as the outgroup because the former is considered sister to Ravenna by Hsu & Chou (2001) and the latter to be closely related to Ravenna by Koivaya (2007). The program MrBayes 3.2.6 (Ronquist et al., 2012) was used simultaneously for 5 million generations. We removed the first 25% burn-in parts and used the remainder to generate a 50% majority consensus tree.

Assessing the effective sample size (ESS), evaluating the parameters, and estimating convergence of two runs were performed with the software Tracer v.1.7 (Rambaut et al., 2018). Phylogenetic trees were read by FigTree v.1.4.3 (see http://tree.bio.ed.ac.uk/software/figtree/). The software program DnaSP 5.10.01 (Rozas et al., 2017) was used to calculate genetic parameters including the nucleotide composition, the number of polymorphic sites, and variable nucleotide positions. A haplotype network was generated via median-joining method using POPART 1.7 (Leigh & Bryant, 2015).

Material depository

Primary types are deposited in the following collections: Institute of Zoology, Academia Sinica, Beijing (IOZ), Kadoorie Farm and Botanic Garden, Hong Kong (KFBG), and Department of Biology, National Taiwan Normal University, Taipei (NTNU).

Terminology for description

Measurements are defined and abbreviated as follows: forewing length (FL) and antennal length (AL). Terminology of wing patterns follows that of Nijhout (1991).

Nomenclature

The electronic version of this article in Portable Document Format (PDF) will represent a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new
names contained in the electronic version are effectively published under that Code from the electronic
edition alone. This published work and the nomenclatural acts it contains have been registered in
ZooBank, the online registration system for the ICZN. The ZooBank LSID (Life Science Identifiers) can
be resolved and the associated information viewed through any standard web browser by appending the
LSID to the prefix http://zoobank.org/. The LSID for this publication is:
urn:lsid:zoobank.org:act:
Ravenna
described
render
conceivable
Fig.
etc.)
Taxonomic
high,
POPART
Fujian
Vietnam.
only
bp)
S3
The
genetic
Results
 supports
if
ZooBank,
edition
and
CLOCKSS.
be
Zoobank.org:pub:43494A4-F758-4C1D-A332-B2D2E8906908. The online version of this work
is archived and available from the following digital repositories: PeerJ, PubMed Central SCIE and
CLOCKSS.

Results
Genetic divergence
The genetic divergence of COI barcode found in the samples used in the present study is shown in Tables
S3 and S4. The phylogenetic tree based on BI analysis is given in Fig. 2. DNA barcode sequences (1034
bp) were obtained from 80 samples (Table S1). A total of 37 variable sites were detected, with 24
haplotypes defined. Two out of 37 polymorphic sites were singleton variable sites, one from Yinggeling
and one from Zhejiang. The variation of mtDNA revealed the presence of 24 haplotypes, seven found
only in Taiwan, three restricted to Yinggeling, seven to Bawangling, four to Zhejiang, and two to
Vietnam. Samples from Guizhou and Guangdong shared the same haplotype, and those from Jiangxi and
Fujian had the same haplotype. Haplotype networks were constructed using the TCS analysis in the
POPART software (Fig. 3). The overall haplotype diversity (Hd) was 0.899 and nucleotide diversity (A)
was 0.012. The values of haplotype diversity and nucleotide diversity that were present are shown in
Table S3. The haplotype diversities of Ravenna nivea from Yinggeling and Bawangling within Hainan are
high, suggesting that the genetic variance is associated with geographic distribution and isolation.

Taxonomic decision
The pairwise genetic divergence of the COI barcode was 0.1-1.7% between Ravenna samples examined
from different sites and 1.1-1.5% between the Yinggeling sample and samples from the other localities
(Table S4), both of which are lower than the 3% COI genetic divergence for species discrimination of
Lepidoptera suggested by [Hebert et al. (2004)]. According to Braby et al. (2012) scheme for testing
hyothesis of taxonomic status of allopatric populations, the null hypothesis of a single species is rejected
only if evidence from other multiple data sources (color pattern, morphology, behavior, ecology, genetics,
etc.) supports the alternative. The morphology of immature of the Yinggeling (Fig. 4) is
indistinguishable from that of R. nivea from the other localities (see p. 481 in Igarashi, 1997; p. 33 in Uchida1999
Fig. 45 in Koswaya 2007; P. 223 in Lu & Chen, 2014) in morphology and biology. There is also no
conceivable diagnosis present in morphology of genitalia of the Yinggeling samples (Figs. 5, 8A) from
those of the other subspecies (Figs. 6, 7, 8B, 8C). The morphology, biology, and COI barcodes thus
render support to place the Yinggeling Ravenna sample within R. nivea, but deserving a subspecies status
with its well-differentiated wing patterns and genetic make-up. Accordingly, a new subspecies is
described herein.

Systematics
Ravenna nivea ngiannmoiae Lo & Hsu, subsp. nov.
(Figs. 1E-1H, 5, 8A)

Paratypes. 5f: same locality as for holotype, 1000-1400 m, 13. V. 2014, Coll. Y. F. Lo (4f, KFBG; 1f, NTNU); 1f3f, same collecting data as for holotype, emgd. 26. II. - 7. III. 2015 (HSU 15A29)(1f dissected, genitalia preparation YFH 1565, IOZ; 1f, NTNU; 2f, KFBG) (DNA voucher YFH 170160).

1f, same locality as for holotype, 1200-1300m, 30. I. 2015, reared from O. fleuryi, emgd. 9. III. 2015, Coll. Y. F. Hsu & Y. F. Lo (HSU 15A34) (NTNU); 2f3f, same locality as for holotype, 1200m, 31. I. 2016, reared from O. fleuryi, emgd. 7-27. III. 2016 (HSU 15A40) (1f dissected, genitalia preparation YFH 15611, NTNU; 1f, NTNU; 1f2f, KFBG); 1f1f, same locality as for holotype, 1500m, 31. I. 2016, reared from O. fleuryi, emgd. 24. III/19. IV. 2016 (HSU 15A50) (NTNU).

Description. Male (Figs. 1E, 1F): FL 19.2-22.0 mm (mean, 20.4 ± 1.1 mm, n = 5); AL 8.3-8.7 mm (mean, 8.6 ± 0.2 mm, n = 4). Head: Hairy, vertex, frons dark brown, with mesal white patch on vertex. Eye semi-oval, sparsely covered with short, buff setae. Narrow white rim surrounding eye. Labelal palpus porrect, with 3rd segment pointed downwards, covered with white scalings and hairs, dark brown apically. Maxillary palpus reduced, inconspicuous. Proboscis uncaged. Antenna smoothly scaled, naked at terminal end of nodum, dark brown with lateral white dots on each flagellomere. Thorax: Dark brown dorsad, white ventrad. Legs with tarsus of foreleg segmented; white, with brown bands. Forewing: Generally triangular in shape, with apex slightly obtuse; termen, costa slightly convex; dorsum straight. Ground color of upperside purple, with narrow, dark brown margin along termen. Fringe with outer cilia white, inner cilia brown. Ground color of underside white. Discal spot as hollow brown bar. Distal band of central symmetry system represented as a pair of well-separated brown lines, approximate in cell Cu1. Submarginal band as a series of faint, brown spots; element g (sensu Nijhout, 1991) as a broken brown band. Fringe white, prominent. Hindwing: Contour of wing slightly produced at distal end of Cu1; Cu2 bearing long, tail-like projection. Ground color purple, with brown area along costa; faint, narrow, white margin along termen. Fringe white. Ground color of underside white. Discal spot inconspicuous. Distal band of central symmetry system represented as a pair of brown lines, approximate posteriad, dislocated proximally as tick-shaped lines in cell Cu1, retracing into straight lines in cell 1A+2A. Proximal band of central symmetry as a broken, brown line. Submarginal band and element g similar to those of forewing, but a black, rounded spot crowned with orange in cell Cu1, and at tornus. Fringe white, prominent. Abdomen: Brown dorsally, white ventrally. Genitalia (Figs. 15-17): Ring-shaped sclerites of 9+10 segments with posterior end quadricuncate. Tegumen broad, most prominent, short, conical, setose; Lines forming a pair of elongate, lateral processes with pointed distal tip, slightly down-curved. Vinculum narrow. Brachia (falces) up-curved, ox horn-like, distal end pointed. Saccus longer than length of tegumen, with anterior end obtuse. Valva elongate, narrowed posteriorly with distal end rounded, small teeth present along inner margin. Juxta broad ventrally, with two arms extending dorsal, forming opposite pointed tips. Phallos with phallobase approximately as long as aedeagus. Aedeagus with dorsal opening at distal end, blunt at caudal tip.

Female (Figs. 1G, 1H). FL 19.0-21.7 mm (mean, 20.4 ± 1.1 mm, n = 13); AL 7.0-8.8 mm (mean, 8.1 ± 0.6 mm, n = 13). Body, wing patterns of underside as described for male except ground color darker. Wing upperside mostly white with extensive purple scalings proximally. Prominent dark brown scalings covering forewing apex, along costa and termen, and along termen of hindwing. Prominent dark brown bar present at distal end of discoidal cell of forewing; faint, narrow, white margin also present.
along termen of hindwing. *Genitalia* (Fig. 8A): Corpus bursae oval, elongate. Ductus bursae short, thick, sclerized and enlarged posteriorly, with large ostium bursae. Sterigma with lamella antevaginalis forming a pair of sclerotized, lateral patches; lamella postvaginalis as broad sclerotized wall, with posterior margin produced, blunt. A small, detached transverse, rectangular, sclerotized band posterior to lamella postvaginalis. Posterior apophyses slender, much longer than anal papillae, enlarged and flattened basally. Anal papillae as weakly sclerotized band, setose.

**Immatures.** Egg (Fig. 4A) 0.82 ± 0.03 mm in diameter, 0.49 ± 0.02 mm in height (*n* = 15), hemispherical but compressed, white, surface with sculpture forming framework with regularly arranged, stub-like processes. Larvae (Figs. 4C, 4D) with four instars. Head brown, glossy. Body onisiform, with surface bearing transparent setae; T1 shield as a short, transverse band, slightly produced laterally; anal lobe semi-circular, with posterior margin rounded. Ground color of body green tinged with yellow. Red markings present in late instars, laterally and around medial and caudal portion dorsally. White chevrons present subdorsally. Spiracles white, surrounded by brown peritremes. Full-grown larva (Fig. 4D) reaching 18 mm in body length. Pupae reaching 12 mm in length, of typical lycaenid form, ground color pale brown, decorated with yellow chevrons and dark brown, subdorsal bands; spiracles cream white.

**Diagnosis.** The taxon *ngiunmoiae* ssp. nov. can be distinguished from other subspecies of *R. nivea* by the following external features: 1) in the males, prominent white scalings are absent on wing undersides in *ngiunmoiae* (Fig. 5), whereas present in the other subspecies (Fig. 1, 2) in the females, extensive purple scalings are present on wing undersides in *ngiunmoiae* (Fig. 5), whereas weakly-developed brown scalings are found proximally in the other subspecies (Fig. 3).

**Hostplant.** *Quercus hui* and *Q. fleuryi* (Fagaceae).

**Distribution.** Known from montane zone of Mt. Yinggeling, central Hainan approximately 1000–1500m in elevation.

**Phenology.** Univoltine. Adults were collected in May, suggesting occurrence of adult imago may be in early summer. Overwintering as egg near bases of dormant buds on the hostplants.

**Eymology.** The subspecific name *ngiunmoiae* refers to the pronunciation of the name of the late grandmother in the Hakka dialect, of the first author of the new subspecies, YFP Lo, for her warm and full support to his scientific work on butterflies and other animals.

**Discussion**

The most interesting finding about *Ravena nivea* *ngiunmoiae* ssp. nov. is that its type locality Yinggeling is only approximately 40 km distant from Bawangling, where another population of *R. nivea* occurs, with wing patterns indistinguishable from continental race *ssp. howarthi*. It appears that the p-distance of the COI barcode between samples from Bawangling and localities from southern China (Jiangxi, Zhejiang and Fujian) was 0.6%, evidently lower than the value between samples of Yinggeling and Bawangling (1.1%) (Table S4). A phylogenetic analysis also suggested that the population of *R. nivea* from Bawangling is more closely related to those of southern China than to the population of Yinggeling (*ngiunmoiae* ssp. nov.) (Fig. 2). These results suggest that the population of Yinggeling may have been separated from that of Bawangling for a long time, and they may have been established independently from different continental sources, instead of differentiated from a common ancestral source. Geological evidence has shown that Hainan was part of continental China until the early Quaternary, but separated when the Qiongzhou Strait formed, and then subsequently re-connected and re-disconnected due to the arrival and departure of
glacial periods (Yan, 2006; Zhao et al., 2007). Zhu (2016) conducted a biogeographical study based on
taxonomic and geological evidence suggesting that Hainan was closer to Guangxi and Vietnam in the Eocene.

Nevertheless, the formation history of mountain ranges within Hainan has yet to be established, thus the
pathways of immigration for Yinggeling and Bawangling remain unsolved. The fact that the values of
haplotype diversity of Ravena nivea found in Taiwan and Yinggeling and Bawangling of Hainan are
higher than those in the continental populations (Table S4) suggest islands may serve as glacial refugia
for this montane butterfly during glacial periods. Although the value of subspecies concept has been
debated, and some authors downplay the utility of subspecies (Burbrink et al., 2022), the case found in the
Ravena butterfly demonstrates that the subspecies concept provides clues towards the nature of
heterogeneous geological history of mountain formation within Hainan island.

Conclusions

The subspecies concept has been controversial and criticized for its utility in Linnaean taxonomy, but it
remains a valid species-group name category in the International Code of the Zoological Nomenclature
(ICZN), currently in the 4th version. The discovery of a population of the lycaenid butterfly Ravena
nivea inhabiting Yinggeling on Hainan island, unique in genetic make-up and wing pattern, suggests that
the subspecies concept may help bring attention to this local population with unique history. Without
adoption of the subspecies concept, the significance of such a population to conservation may be
overlooked.

Acknowledgements

We are grateful to Mr. Trevor Padgett (OZ & Tunghai University) for reading and
improving the manuscript of this article. Ge-xia Qiao (Chinese Academy of Science), Alexander
Monastyrskiy (Vietnam National Museum of Nature, Vietnam Academy of Science and Technology),
Takashi Hasegawa and Akiko Kubota (both Asian Insect Research Society), Jianqing Zhu (Shanghai
Zoo), Ju-ling Chen (Guangdong Insect Kingdom Co.) kindly provided materials used in the present
study. Jia-Yuan Liang and Chi-Wei Huang (NTNU) prepared the plates.

Hainan Wildlife Conservation Bureau, Yinggeling National Nature Reserve and Bawangling National
Nature Reserve granted permission to conduct the survey and provided essential assistance with field
work.

References

Bouckaert, R., Vaughan, T.G., Barido-Sottani, J., Duchêne, S., Fourment, M., Gavryushkina, A.,
Drummond, A.J., 2019. BEAST 2.5: An advanced software platform for Bayesian evolutionary

Braby, M.F., Eastwood, R., Murray, N., 2012. The subspecies concept in butterflies: has its application
in taxonomy and conservation biology outlived its usefulness? Biological Journal of the Linnean

Burbrink, F.T., Crother, B.J., Murray, C.M., Smith, B.T.,

Commented [A6]: Not appeared in the text

Deleted: Academia Sinica

Deleted: National Taiwan Normal University

Deleted: …R… Vaughan… T… Barido-Sottani…. J… Duchêne… S…. Fourment…. M… Gavryushkina… A…

Deleted: …rummond… A… (…019)

Deleted: …M…… Eastwood… R… Murray… N… (…012)…

Deleted: …F….. Crother… B… Murray… C… M… Brian Tilston ….math… B…


Figure 1
Adults of *Ravenna nivea*. (A) *ssp. howarthi* Saigusa, male, upperside, Bawangling, Hainan. (B) Underside. (C) Same, female, upperside. (D) Underside. (E) *ssp. ngiunmoiae* Lo & Hsu, subsp. nov., paratype, male, upperside, Yinggeling, Hainan. (F) Underside. (G) Same, holotype, female, upperside. (H) Underside.

Figure 2
Phylogenetic construction for samples of *Ravenna nivea* from various localities based on the COI barcode resulting from BI analysis.

Figure 3
Haplotype networks with haplotype distribution for *Ravenna*. Colors indicate geographic regions. The size of the circle represents the frequency of each haplotype.

Figure 4
Immatures of *Ravenna nivea ngiunmoiae* Lo & Hsu, subsp. nov.,
(A) Egg, dorsal view. (B) Egg, lateral view. (C) Larva in 3rd instar. (D) Larva in 4th (final) instar.

Figure 5
Male genitalia of *Ravenna nivea ngiunmoiae* Lo & Hsu, subsp. nov. (Yinggeling, Hainan).
(A) Dorsal view of ring (tegumen + vinculum). (B) Caudal view of right valva. (C) Ventral view of phallus.

Figure 6
Male genitalia of *Ravenna nivea howarthi* Koiwaya (Bawangling, Hainan).
(A) Dorsal view of ring (tegumen + vinculum). (B) Caudal view of right valva. (C) Ventral view of phallus.

Figure 7
Male genitalia of *Ravenna nivea nivea* Nire (Taiwan).
Figure 8

Female genitalia of *Ravenna nivea*.

(A) ssp. *ngiummoae* Lo & Hsu, **subsp. nov.** (Yinggeling, Hainan). (B) ssp. *howarthi* Koiwaya (Guizhou). (C) ssp. *nivea* Nire (Taiwan)

(A) Dorsal view of *ring* (tegumen+vinculum). (B) Caudal view of right valva. (C) Ventral view of phallus.