

The dinosaurs that weren't: Osteohistology supports giant ichthyosaur affinity of enigmatic large bone segments from the European Rhaetian (#89579)

1

First revision

Guidance from your Editor

Please submit by **20 Jan 2024** for the benefit of the authors .



Structure and Criteria

Please read the 'Structure and Criteria' page for general guidance.



Raw data check

Review the raw data.



Image check

Check that figures and images have not been inappropriately manipulated.

If this article is published your review will be made public. You can choose whether to sign your review. If uploading a PDF please remove any identifiable information (if you want to remain anonymous).

Files

Download and review all files from the [materials page](#).

1 Tracked changes manuscript(s)
1 Rebuttal letter(s)
13 Figure file(s)
2 Table file(s)
2 Other file(s)



Structure and Criteria

Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

1. BASIC REPORTING
2. EXPERIMENTAL DESIGN
3. VALIDITY OF THE FINDINGS
4. General comments
5. Confidential notes to the editor

 You can also annotate this PDF and upload it as part of your review

When ready [submit online](#).

Editorial Criteria

Use these criteria points to structure your review. The full detailed editorial criteria is on your [guidance page](#).

BASIC REPORTING

-  Clear, unambiguous, professional English language used throughout.
-  Intro & background to show context. Literature well referenced & relevant.
-  Structure conforms to [Peerj standards](#), discipline norm, or improved for clarity.
-  Figures are relevant, high quality, well labelled & described.
-  Raw data supplied (see [Peerj policy](#)).

EXPERIMENTAL DESIGN

-  Original primary research within [Scope of the journal](#).
-  Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
-  Rigorous investigation performed to a high technical & ethical standard.
-  Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

-  Impact and novelty not assessed. *Meaningful* replication encouraged where rationale & benefit to literature is clearly stated.
-  All underlying data have been provided; they are robust, statistically sound, & controlled.
-  Conclusions are well stated, linked to original research question & limited to supporting results.



The best reviewers use these techniques

Tip

Example

Support criticisms with evidence from the text or from other sources

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Give specific suggestions on how to improve the manuscript

Your introduction needs more detail. I suggest that you improve the description at lines 57- 86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

Comment on language and grammar issues

The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult. I suggest you have a colleague who is proficient in English and familiar with the subject matter review your manuscript, or contact a professional editing service.

Organize by importance of the issues, and number your points

1. Your most important issue
2. The next most important item
3. ...
4. The least important points

Please provide constructive criticism, and avoid personal opinions

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

Comment on strengths (as well as weaknesses) of the manuscript

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

The dinosaurs that weren't: Osteohistology supports giant ichthyosaur affinity of enigmatic large bone segments from the European Rhaetian

Marcello Perillo ^{Corresp., 1}, P Martin Sander ^{1, 2}

¹ Section Paleontology, Institute of Geosciences, Rheinische Friedrich-Wilhelms Universität Bonn, Bonn, Germany

² The Dinosaur Institute, Natural History Museum of Los Angeles County, Los Angeles, California, United States of America

Corresponding Author: Marcello Perillo
Email address: marcelloperillo.96@gmail.com

Very large unidentified elongate and rounded fossil bone segments of uncertain origin recovered from different Rhaetian (Late Triassic) fossil localities across Europe have been puzzling the paleontological community since the second half of the 19th century. Different hypotheses have been proposed regarding the nature of these fossils: (1) giant amphibian bones, (2) dinosaurian or other archosaurian long bone shafts, and (3) giant ichthyosaurian jaw bone segments. We call the latter proposal the 'Giant Ichthyosaur Hypothesis' and test it using bone histology. In presumable ichthyosaur specimens from SW England (Lilstock), France (Autun), and indeterminate cortical fragments from Germany (Bonenburg), we found a combination of shared histological features in the periosteal cortex: an unusual woven-parallel complex of strictly longitudinal primary osteons set in a novel woven-fibered matrix type with intrinsic coarse collagen fibers (IFM), and a distinctive pattern of Haversian substitution in which secondary osteons often form within primary ones. The splenial and surangular of the holotype of the giant ichthyosaur *Shastasaurus sikanniensis* from Canada were sampled for comparison. The results of the sampling indicate a common osteohistology with the European specimens. A broad histological comparison is provided to reject alternative taxonomic affinities aside from ichthyosaurs of the very large bone segment. Most importantly, we highlight the occurrence of shared peculiar osteogenic processes in Late Triassic giant ichthyosaurs, reflecting special ossification strategies enabling fast growth and achievement of giant size and/or related to biomechanical properties akin to ossified tendons.

The dinosaurs that weren't: Osteohistology supports giant ichthyosaur affinity of enigmatic large bone segments from the European Rhaetian

Marcello Perillo^{1*}, P. Martin Sander^{1,2}

¹Section Paleontology, Institute of Geosciences, University of Bonn, 53115 Bonn, Germany

²The Dinosaur Institute, Natural History Museum of Los Angeles County, Los Angeles, California 90007, USA.

*Corresponding Author:

Email address: marcelloperillo.96@gmail.com

Current Address: Martin-Görgens Str. 43, Bonn, North Rhine-Westphalia, 53117, Germany

Abstract

Very large unidentified elongate and rounded fossil bone segments of uncertain origin recovered from different Rhaetian (Late Triassic) fossil localities across Europe have been puzzling the paleontological community since the second half of the 19th century. Different hypotheses have been proposed regarding the nature of these fossils: (1) giant amphibian bones, (2) dinosaurian or other archosaurian long bone shafts, and (3) giant ichthyosaurian jaw bone segments. We call the latter proposal the ‘Giant Ichthyosaur Hypothesis’ and test it using bone histology.

In presumable ichthyosaur specimens from SW England (Lilstock), France (Autun), and indeterminate cortical fragments from Germany (Bonenburg), we found a combination of shared histological features in the periosteal cortex: an unusual woven-parallel complex of strictly longitudinal primary osteons set in a novel woven-fibered matrix type with intrinsic coarse collagen fibers (IFM), and a distinctive pattern of Haversian substitution in which secondary osteons often form within primary ones.

The splenial and surangular of the holotype of the giant ichthyosaur *Shastasaurus sikanniensis* from Canada were sampled for comparison. The results of the sampling indicate a common osteohistology with the European specimens. A broad histological comparison is provided to reject alternative taxonomic affinities aside from ichthyosaurs of the very large bone segment. Most importantly, we highlight the occurrence of shared peculiar osteogenic processes in Late Triassic giant ichthyosaurs, reflecting special ossification strategies enabling fast growth and achievement of giant size and/or related to biomechanical properties akin to ossified tendons.

1. Introduction

The Late Triassic covers an extremely long-time span (approximately 36 Ma), encompassing two of the fundamental biological revolutions of interest to paleontology, i.e., part of the Mesozoic Marine Revolution and the End-Triassic Mass Extinction (Harper 2006; Davies *et al.* 2017). The Late Triassic also saw the rise of many tetrapod clades in the sea and on land that were to dominate the remainder of the Mesozoic (e.g., plesiosaurs and non-avian dinosaurs) or are still prominent today (e.g., mammals). Nonetheless, the complex of biotic interactions of this Mesozoic Epoch and its protagonists still needs to be fully understood (Benton 2015; Kelley & Pyenson 2015). Giant ichthyosaurs (length >12 m), prominent elements of the ecological communities of Triassic seas, are no exception due to the absence of satisfactory fossils to unravel their evolutionary history and the still obscure timing, dynamics and causes of their extinction at the end of the Triassic Period (Lomax *et al.* 2018; Sander *et al.* 2021).

1.1 Bone segments and putative giant ichthyosaurs from Europe

Large, but fragmentary bone finds from the famous Aust Cliff Rhaetic bonebeds of the Bristol area (southwestern UK) were already reported in the 19th century (Stutchbury 1850). These include what appeared to be large limb bone shafts of reptilian affinity, leading to extensive discussions in the paleontological community (Stutchbury 1850; Sanders 1876; Huene 1912; Storrs 1993, 1994; Benton & Spencer 1995; Galton 2005; Naish & Martill 2008; Redelstorff, Sander & Galton 2014; Lomax *et al.* 2018). The Aust Cliff bonebed is one of a group of similar UK and continental European bonebed-type deposits formed in the Rhaetian epicontinental sea that covered much of Western and Central Europe (Sander *et al.* 2016; Barth *et al.* 2018; Cross *et al.* 2018; Perillo & Heijne 2023) (Fig. S1). These bonebeds yield various tetrapod fossils of both terrestrial and marine origin, often showing fragmentary preservation (Storrs 1993, 1994). The proposed taxonomic affinities of the large to gigantic bone shafts, hereafter less suggestively called “bone segments”, include “labyrinthodonts” (Stutchbury 1850), dinosaurs (Sanders 1876; Reynolds 1946; Storrs 1993, 1994; Benton & Spencer 1995; Galton 2005) and unidentified archosaurs (Redelstorff, Sander & Galton 2014).

The dinosaurian origin of said bone segments (hereafter ‘Dinosaur Hypothesis’) has been supported for the last decades, with Galton (2005) discussing five of the bone segments in detail

93 and concluding that they either must represent sauropodomorph or, more likely, stegosaur long
 94 bone shaft fragments (femur, ?tibia). An inconsistency with the long bone nature of the segments
 95 would seem to be their lack of a continuous cortex and periosteal surface around their periphery.
 96 Instead, as much as two thirds of the periphery of shaft cross sections appears to consist of
 97 cancellous bone (Galton 2005, figs. 4-6). Galton (2005) had already noticed the lack of an outer
 98 bone surface in some areas. Whereas this feature could be primary, as in a jaw bone
 99 (representing a suture surface or a surface facing the Meckelian canal), it also could result from
 100 heavy abrasion, which characterizes all Aust Cliff and other bonebed material.

101 Galton's (2005) conclusion as to the stegosaurian nature of the bone segments has since
 102 been questioned by multiple workers (Maidment *et al.* 2008; Naish & Martill 2008; Sander 2013;
 103 Redelstorff, Sander & Galton 2014; Lomax *et al.* 2018) due to the lack of diagnostic
 104 morphological features and stratigraphic arguments. In particular, the largest known stegosaur
 105 already occurring in the Late Triassic would be inconsistent with the known ornithischian fossil
 106 record and result in long ghost lineages (Galton 2005; Maidment *et al.* 2008; Naish & Martill
 107 2008). Sauropods, on the other hand, would appear to be a reasonable option.

108 A histological test of sauropod affinities of the Aust Cliff bone segments was then
 109 conducted by Redelstorff, Sander & Galton (2014). Sampling two of the Aust Cliff specimens
 110 (BRSMG-Cb-3869 and BRSMG-Cb-3870, see Table 1), Redelstorff, Sander & Galton (2014)
 111 found a peculiar and previously undescribed set of histological characters (a thin cortex of
 112 fibrolamellar bone with longitudinal primary osteons and secondary osteons forming within the
 113 primary ones), inconsistent with sauropod or other sauropodomorph affinities (Redelstorff,
 114 Sander & Galton 2014). In their primary cortex, sauropodomorph long bones show a different
 115 and rather uniform histology: laminar and plexiform fibrolamellar bone and, in the case of
 116 sauropods, almost no growth marks until late in life (Sander & Klein 2005; Klein & Sander
 117 2007, 2008; Sander *et al.* 2011).

118 Following the recent find of a very large elongate and partially curved bone segment
 119 (BRSMG Cg2488, 96 cm long, Fig. S3B) in the Rhaetian of Lilstock (Lomax *et al.* 2018), also in
 120 SW England, this segment and the Aust Cliff bone segments were identified as fragments of the
 121 surangular bones derived from giant ichthyosaur jaws by Lomax *et al.* (2018). This interpretation
 122 by Lomax *et al.* (2018) was based on a morphological comparison with somewhat older giant
 123 ichthyosaurs from North America, specifically the Carnian *Shonisaurus popularis* from Nevada

(Camp 1980) and the Norian *Shastasaurus sikanniensis* (Fig. S3A) from British Columbia, Canada (Nicholls & Manabe 2004). We term this hypothesis of the affinity of the very large Aust Cliff bone segments the ‘Giant Ichthyosaur Hypothesis’.

Support for the Giant Ichthyosaur Hypothesis would seem to come from an earlier find, now lost (Fig. S3C). Huene (1912) described a 1.4 m long bone segment from Aust Cliff which he identified as the fragment of a right lower jaw of a giant ichthyosaur, including part of four elements (dentary, splenial, angular, surangular) (Fig. S3C). Huene (1912) noted that this fossil had been accessioned to the “Bristol Museum” since 1877, presumably referring to today’s Bristol City Museum and Art Gallery (BRSMG). However, Huene (1912) did not provide a specimen number, and since his 1912 study, the specimen has not been mentioned again, and it may well have been destroyed in WWII. According to Huene’s (1912) description and illustration, the specimen consists of four non-fitting parts, the penultimate of which had been sectioned transversely (Fig. S3C) at some earlier point in time before Huene’s study.

Curiously, among the putative dinosaur long bone material described by Galton (2005) from Aust Cliff, there also is a transversely sectioned specimen (BRSMG Cb3870, Fig. S2) of about the dimensions noted by Huene (1912) (Fig. S3C). Galton did not cite Huene, and there is a possibility that the two authors did study the same specimen. Arguing against the identity of the two specimens is the poor preservation of the Galton specimen (whereas Huene emphasized the good preservation of his material) and the fit with another segment (whereas Huene noted the lack of fits).

Finds similar to the Aust Cliff and Lilstock material have come from the epicontinental French Rhaetian localities of the Autun area (Fig. S1) and from southern France (Fischer *et al.* 2014; Lomax *et al.* 2018), as well as most recently, from the German locality of Bonenburg (Fig. S1) (Sander *et al.* 2016; Wintrich *et al.* 2017) and the Swiss Alps (Sander *et al.* 2022, fig. s5). Fischer *et al.* (2014) also had interpreted his material as ichthyosaurian but did not extend their considerations to the UK material and did not cite Huene (1912). Huene (1912), on the other hand, just described this one specimen from Aust Cliff and did not comment on the putative dinosaur leg bone shafts from the same locality nor on the French Rhaetian ichthyosaur material, all of which were known at the time.

153

154 1.2 The Late Triassic giant ichthyosaur record

155 Ever since the work of Charles S. Camp in the Carnian Luning Formation of Nevada,
 156 USA, in the 1950s (Camp 1980), it has been clear that Late Triassic ichthyosaurs reached body
 157 lengths of 15 m or more and must have been substantially larger than post-Triassic ichthyosaurs.
 158 Based on several partial skeletons, Camp (1908) described the new genus and species
 159 *Shonisaurus popularis* from Berlin Ichthyosaur State Park in Nevada. This material has been
 160 reevaluated several times since with regard to the size, skeletal reconstruction, taphonomy, and
 161 reproductive biology (Kosch 1990; Hogler 1992; McGowan & Motani 1999, Kelley *et al.* 2022).
 162 Even larger and more complete than any of the *S. popularis* finds is the holotype skeleton of
 163 *Shastasaurus sikanniensis* (Nicholls & Manabe 2004) from the middle Norian of British
 164 Columbia, Canada. Based on field data, this individual is estimated to have been 21 m long
 165 (Nicholls & Manabe 2004).

166 It is also now acknowledged that various other ichthyosaur finds from the Late Triassic
 167 must represent animals over 10 meter in length, but most giant ichthyosaurs are represented by
 168 woefully incomplete, disarticulated and fragmentary material from around the world (Camp
 169 1976; Callaway & Massare 1989; McGowan & Motani 1999; Sander *et al.* 2022; Kelley *et al.*
 170 2022) which hinders the anatomical descriptive effort. In continental Europe, the fragmentary,
 171 often reworked, and poorly understood finds attributed to giant ichthyosaurs come from late
 172 Norian to Rhaetian outcrops of France (Fischer *et al.* 2014), the eastern Swiss Alps (Sander *et al.*
 173 2022), and from a recently discovered Aust Cliff-type bonebed near the central German village
 174 of Bonenburg (Fig. S1) (Sander *et al.* 2016; Wintrich *et al.* 2017). Unlike all the other Rhaetian
 175 localities with putative giant ichthyosaurs, the Bonenburg deposit is precisely dated
 176 palynologically, ranging from late middle to early late Rhaetian in age (Schobben *et al.* 2019;
 177 Gravendyck *et al.* 2020). The Bonenburg ichthyosaur fossils include large but very short
 178 vertebral centra, a very large neural arch, and very large rib fragments (Sander *et al.* 2016). In
 179 addition, the bonebed frequently yields heavily abraded fragments of thick cortical bone up to 25
 180 cm in length (Fig. S4A, S6A), which we hypothesize to be fragments of bone segments similar to
 181 the more complete British and French specimens (Figs S2A, S3B).

182 Understanding the affinity of the fragmentary Late Triassic ichthyosaurs and of the large,
 183 more obscure fragmentary finds, is important because of the absolute size of these remains,
 184 representing records of the largest animals inhabiting the Late Triassic oceans (Lomax *et al.*
 185 2018; Sander *et al.* 2022). The fossils represent animals that far exceeded the size of any other

186 marine tetrapods except for the largest species of baleen whales and archaeocetes (Bianucci *et al.*
187 2023). The importance of these fossils also relates to the patterns of extinction at the end of the
188 Triassic, given that very large ichthyosaurs appear to have persisted to the late Rhaetian
189 (indicated by the Bonenburg finds) but are lacking in the Jurassic.

190 The lack of clear and unequivocal external morphological features in the Rhaetian
191 European bone segments due to their fragmentary and reworked nature makes alternative
192 approaches such as microstructure analysis (microanatomy and osteohistology) critically
193 important in investigating the possible affinities of these fossils. Both Galton (2005) and Lomax
194 *et al.* (2018) illustrated cross sections of UK fossils and discussed microanatomy (but not
195 histology, which is not accessible without thin-sectioning). Galton compared the midshaft
196 microanatomy of BRSMG 3869, 3870, and 4063 from Aust Cliff to that of various dinosaurs and
197 concluded that the fossils must represent stegosaurs based on the coarse cancellous bone
198 structure of the medullary region. Lomax *et al.* (2018) noted and illustrated in detail the same
199 coarse cancellous bone structure but did not use microanatomical arguments as evidence for
200 determining affinity, only cross-sectional shape. Histological analysis was already performed on
201 two Aust Cliff specimens (BRSMG-Cb-3869 and BRSMG-Cb-3870) by Redelstorff, Sander &
202 Galton (2014) (Table 1), but without considering possible ichthyosaurian affinities of the fossils.
203 Instead, Redelstorff, Sander & Galton (2014) adhered to the paradigm that the Aust Cliff
204 segments were shafts of dinosaur long bones.

205 Here we undertake a detailed and comprehensive comparison and sampling of most
206 European Rhaetian “bone segments” and putative giant ichthyosaur jaws for histological
207 analysis. The main aim of this study thus is to histologically test the Giant Ichthyosaur
208 Hypothesis by searching for shared histological characters among European material of
209 confirmed or proposed ichthyosaurian nature, on one hand, and bonafide Late Triassic giant
210 ichthyosaurs, such as *S. sikanniensis*, on the other. We also compare the “bone segments”
211 histology with other terrestrial and aquatic tetrapods that are known to have reached very large
212 body size in the Late Triassic such as sauropodomorph dinosaurs, rauisuchians, dicynodonts, and
213 plesiosaurs.

214

215 2. Materials & Methods

216

2.1 Materials

The material used in this study consists of bone histological samples taken from various specimens borrowed from multiple institutions as listed in Table 1. Abbreviations for these institutions are also listed in this table. In summary, there are eight sets of samples (Table 1). These include two samples (surangular, splenial) from the *S. sikanniensis* holotype RTMP-1994-378-0002 (Nicholls & Manabe 2004) (Fig. S3A), one sample of the Lilstock putative ichthyosaur surangular (Lomax *et al.* 2018) (Figs S2, S3B), three samples of “dinosaur bone shafts” reinterpreted as giant ichthyosaur jaw bone fragments from the Aust Cliff Rhaetic bonebed (Galton 2005; Redelstorff, Sander & Galton 2014; Lomax *et al.* 2018), two samples from a giant putative ichthyosaurian lower jaw (Fischer *et al.* 2014), identified as surangular by Lomax *et al.* (2018), from Autun, France (Figs S2, S3B), and finally 16 cortex fragments of various sizes from Bonenburg, Germany (Figs S4, S5A, S6A). For details on all of these samples, including sampling locations and methods, and their current identification, see Supplemental Article S1.

The thin sections used for the study are either in the paleohistological collections of the IGPB or with the sampled fossils (see Table 1). Note that two of the Aust Cliff thin sections were already studied by Redelstorff, Sander & Galton (2014).

2.2 Methods

2.2.1 Histological sampling

Except for the *S. sikanniensis* holotype, jaw bones and putative jaw bones were sampled by core drilling, following Sander (2000) and Stein & Sander (2009) (Table 1). The *S. sikanniensis* lower jaw was sampled with a Dremel-type cutting tool, making two parallel cuts spaced 18 mm apart (Fig. S3E) and then preparing out the sample. Complete cross sections and longitudinal sections were obtained from smaller specimens of cortical bone fragments from Bonenburg by cutting with a rock saw after embedding with a protective epoxy putty. Cores and full sections were then processed into thin sections following Lamm (2013), with slight modification of the standard technique: wet silicon carbide powder of grit sizes of 600 and 800 was used for the grinding and polishing processes.

Once covered, the thin sections were studied under a Leica DMLP polarizing light microscope in regular illumination and by using cross-polarization and circular polarization techniques. Circular polarization (Bromage *et al.* 2003) was obtained through the use of a pair of

commercially available polarizing glasses for 3D movie viewing to replace the polarizer and the analyzer of the microscope (Richtberg & Girwidz 2023). This allows observation of the thin sections in circular polarized light without the Maltese cross effect. Photomicrographs were taken using a Leica DFC420 camera (software Leica Firecam, ver. 3.1, 2007, © Leica Microsystems, Switzerland, Ltd), a Dino-Eye camera (software DinoCapture 2.0 ver 1.5.45 © 2016 AnMo Electronics Corporation), and with a Canon EOS2000D (software EOS Utility ver. 3.16.11, 2023, © Canon Europa N.V. and Canon Europe Ltd 2002-2009) mounted on the microscope.

2.2.2 Porosity quantification



Sections were scanned with a flatbed scanner or photographed under the microscope with a cell phone camera. In the latter case, successive microphotos were merged using the photomerge tool in Photoshop (Ver 20.0.4 20190227.r.76). Both scans and merged photos were transformed into binary pictures for porosity quantification (Fig. S7). Porosity quantification was executed with the software BW-counter (© Peter Göddertz, IGPB). Porosity is expressed as the percentage of white area (vascular and trabecular cavities) vs. black area (mineralized bone material).

2.2.3 Terminology, including new terminology

Histological terminology follows Buffrénil & Quilhac (2021a) for osteohistology and Buffrénil & Quilhac (2021b) for types and features of secondary osteons. These play a major role in this study. For one, there are concentric osteons, most recently discussed by Buffrénil & Quilhac (2021b). In these, a secondary osteon develops within a Haversian canal, i.e., within a preexisting secondary (not primary) osteon (Lacroix 1970). We did observe concentric osteons in this study. Concentric osteons are not to be confused with double-zoned secondary osteons (Skedros, Sorenson & Jenson 2007) where the centripetal infill of a secondary osteon happens in stages, but without intervening resorption. We did not observe such double-zoned secondary osteons in this study. However, neither of these terms describes the situation observed already by Redelstorff, Sander & Galton (2014) in the Aust Cliff material, in which a secondary osteon develops within a primary one. We refrain from erecting new terminology for this situation but use a simple descriptive approach. When the entire cortex is affected by the reuse of preexisting vascular canals by secondary osteons, we define this as “template cortex”.

Nevertheless, the histology of the giant ichthyosaur material is so unusual in other features that it does require new terminology which will be introduced in the results section. This new terminology was coined to aid in the description of a novel histology in the periosteal territory for which no proper definition was found in the literature.

Our general histological description follows the 3-Front Model of Mitchell & Sander (2014) (Fig. 2) in which the osteohistological pattern observed in an amniote cortical bone sample is conceptualized as being generated by the successive outward advance and relative speed of three fronts. These fronts are the apposition front (where bone tissue is formed), followed by the Haversian substitution front (where primary bone tissue is replaced by secondary tissue), and the resorption front (where bone tissue is resorbed to make space for bone marrow). Due to the undefined taxonomical state of the specimens and lack of clear homology in sampling location (aside for BRSMG-Cb-3869 and BRSMG-Cg-2488 R-101), the model is only used for descriptive purposes and general comparison, but not to define relative developmental stages.

3. Results

3.1 Shared histology of the British and French samples

3.1.1 General histological and microanatomical description

Laid down by the apposition front (Mitchell & Sander 2014), the outer cortex of all samples from the British and French Rhaetian is characterized by compact primary bone tissue structured by wavy growth marks parallel to the outer bone surface (Figs 1A, 2, 3A, B).

The primary periosteal bone matrix is a new matrix type, “intrinsic fiber matrix” or IFM. IFM is characterized by a network of bright anisotropic, intrinsic, mineralized fibers set in an isotropic matrix. IFM pertains to the woven-fibered type of bone matrices which are produced by static osteogenesis (Buffrenil & Quilhac 2021a). IFM probably represents the result of an intermediate type of bone deposition between woven bone and parallel fibered bone. Contrary to a normal woven-fibered bone matrix, IFM contains abundant coarse intrinsic collagen fibers, both mineralized and unmineralized, that are uniformly oriented longitudinally. Contrary to parallel fibered bone, IFM coarse fibers have different orientation showing a net or lattice-like pattern, immersed in a clearly isotropic matrix. Of particular relevance is that the fibers are intrinsic, not extrinsic (as e.g. Sharpey’s fibers).

310 Vascularization of the primary cortex is characterized by longitudinal vascular canals (Figs
311 2, 3A, B). Immature primary osteons and vascular canals open up to the outer bone surface,
312 resulting in an ornamented wavy surface (Figs 2, 3B) in thin section. This histology correlates
313 with distinctive longitudinal surface striations on the specimens (Fig. S3D), nicely illustrated by
314 Lomax *et al.* (2018, fig. 4c, 8) for the Lilstock and Aust specimens and Fischer *et al.* (2014; fig.
315 2, s5) for the French specimens.

316 Vascular canals and primary osteons are arranged in appositional circumferential rows
317 demarcated by closely spaced growth marks (GM) that vary in number (Figs 1A, 2, 3A, B).
318 Growth marks appear as depositional layers of periosteal primary bone and run around the
319 periosteal vascular canals (Figs 1A, 3A, B). The GM vary in thickness (Figs 1A, 3A vs Fig. 3B)
320 and show alternating light-dark coloration. The differential coloration seems to be related to
321 differences in intrinsic fiber density and orientation. Vascularization as observed in longitudinal
322 sections does not show anastomoses between vessels, with vessel cross sections rarely showing
323 shapes more complex than an elongated ellipsoid (Fig. 3C).

324 The Haversian substitution front is diffuse in that the outer cortex shows scattered evidence
325 of secondary remodeling through small resorption cavities and secondary osteons within primary
326 ones (Fig. 1A, D), or even mature secondary osteons (Fig. 3A). Appositional rows of primary
327 osteons may follow or precede rows of primary osteons with secondary ones within them, or
328 rows of secondary osteons may even be intercalated with rows of purely primary osteons (Figs
329 1A, D, 3E). The deep cortex, i.e., the part of the cortex that is fully within the Haversian
330 substitution front, can again be subdivided in an outer template deep cortex and an inner,
331 completely remodeled area, where none of the primary pattern of vascularity is preserved (Fig.
332 2). This situation was already described in detail by Redelstorff, Sander & Galton (2014, fig. 4).
333 The thickness of these two subzones of the deep cortex varies between samples (Fig. 2).

334 As noted above, the template deep cortex is so named because it preserves most or some of
335 the original primary vascular architecture (Figs 2, 3E). This is because of the peculiar pattern that
336 Haversian substitution is initiated from existing primary vascular canals, i.e., existing vascular
337 pathways were reused. This results in a predominance of secondary osteons within primary ones.
338 The primary osteons thus clearly influence the course of the secondary ones, even leading to
339 rows of exclusively secondary osteons forming complete Haversian tissue, templated by the
340 primary rows of osteons (Fig. 3E). The templating we observed is different from normal

341 Haversian substitution in amniotes in which the cutting cones of secondary osteons show little
342 regard for preexisting structures (Mitchell 2017).

343 Both, primary and secondary osteons, have a high number of lamellae and a small vascular
344 canal, which results in a rather low general porosity for the entirety of the sections (between 17%
345 and 13%) (Fig. S7 A-E), possibly indicating an osteosclerotic state of the cortex. 

346 Osteon cross sections vary consistently between circumferential rows, sometimes
347 horizontally flattened, sometimes more vertically (Figs 2, 3E), possibly indicating variations in
348 growth rate (Woodward 2019). Migratory and incipient osteons (Skedros *et al.* 2007; Mitchell
349 2017) are present, but secondary osteons within primary ones represent the majority of osteons in
350 the template cortex.

351 Further inward from the template cortex, the regular deep cortex can be seen as resulting
352 from complete secondary reconstruction (Fig. 2). This part of the cortex is characterized by more
353 chaotically arranged secondary osteons that have obliterated the primary vascular architecture by
354 several cycles of secondary osteon formation. The result is normal Haversian tissue which marks
355 the full effect of the Haversian substitution front (e.g. Fig. 2D).

356 The boundary of the perimedullary region, i.e., the resorption front, is also diffuse (Fig. 2).
357 Here, the deep cortex becomes more and more affected by larger resorption cavities lined only
358 by a few lamellae. Porosity in the perimedullary region is between 65 to 85% (Fig. S7 A-E). This
359 signifies an increasing imbalance between secondary bone deposition and resorption activity and
360 initiates the formation of secondary trabeculae (Fig. 2). Through the activity of the resorption
361 front, the perimedullary region is rich in resorption cavities replacing bone tissue with some
362 secondary osteons and transitioning to a medullary area of secondary trabecular bone (Fig. 2).

363 In cases, where the resorption front has overtaken the Haversian substitution front,
364 interstitial areas of primary tissue consisting of IFM are visible (Fig. 3D) between the secondary
365 trabeculae. The percentage of interstitial primary tissue decreases inwards but is patchy. Our
366 histological observations are consistent with the descriptions and illustrations of cross-sectional
367 microanatomy given by Galton (2005), Fischer *et al.* (2014), and Lomax *et al.* (2018), who all
368 note that there is only a very small open medullary cavity surrounded by an extensive zone of
369 inward-decreasing trabecular density (Galton 2005, figs 4-6; Fischer *et al.* 2014, fig. s5; Lomax
370 *et al.* 2018, fig. 6). Fischer *et al.* (2014, fig. s5) interpret this open medullary cavity as the dental
371 groove, however.

Both, the largest Aust Cliff bone segments (BRSMG Cb 3869) and the Lilstock specimen (BRSMG-Cg-2488 R-101), show conspicuous cavities in the cortical bone (Figs 2A, C, 3F). There are two obvious ones in the latter and one obvious and a second possible one in the former (Figs 2A, C, S7A, C). Based on the sampling location, these open cavities represent the nutrient canals extending inwards at a low angle from the elongate foramen (Figs S2A, S3B, C) opening in caudal direction (already described by Huene 1912 and identified as part of the *fossa surangularis* by Lomax *et al.* 2018) on the bone surface. On the outward margins of the cavities (those facing the periosteal surface), both samples show primary tissue and simple vascular canals (Fig. 3F). Both BRSMG Cb 3869 and BRSMG-Cg-2488 R-101 show signs of resorption along the inner and lateral margins of the nutrient canals, indicating microanatomical drift related to the growth of the bone enclosing the canal.

383

3.1.2 Template remodeling and secondary osteons within primary ones

As noted, the distinctive template secondary remodeling is shared between all French and UK samples (Table 2). It is possible to identify secondary osteons within primary ones through the method adopted by Redelstorff, Sander & Galton (2014), i.e., focusing through the sample in normal light, using higher magnifications, a nearly closed diaphragm, and the condenser, or by observing the position of the resorption/cementing lines through the λ filter. The occurrence of multiple generations of secondary osteons within primary ones (Fig. 1A, D) tends to maintain the original periosteal appositional rows (Fig. 2) as noted above. Secondary osteons within primary ones represent the advancing Haversian substitution front and may occur quite closely to the outer bone surface in the outermost cortical layers. Whereas secondary osteons within primary ones also have been reported in mammals (e.g., Sander & Andrassy 2006 and the reference cited above), ~~in these~~, they are not such a consistent feature as observed in the template cortex of our specimens.

397

3.1.3 PIFT with longitudinal vascular canals

All UK and French Rhaetian putative jaw bone samples share the same unusual primary periosteal bone tissue: a woven-parallel complex with strictly longitudinal, highly ordered, primary osteons set in intrinsic fiber matrix (IFM). We term this woven-parallel complex 'periosteal intrinsic fiber tissue' PIFT (Table 2). PIFT is a feature at the bone tissue level of

403 integration and thus is to be used in conjunction with ‘lamellar bone tissue’, ‘parallel-fibered
404 tissue’, ‘Haversian tissue’, etc. (Buffr  nil & Quilhac 2021a, b). In PIFT, the “parallel”
405 component of the woven-parallel complex is represented by typical osteonal lamellar bone of the
406 longitudinal primary osteons, the “woven” component, building up the scaffold of the bone, is
407 IFM (Figs 1A-C, S8F). IFM is a type of woven-fibered matrix (Stein & Prondvai 2014, Buffr  nil
408 & Quilhac 2021a) because it is a combination of isotropic woven bone with coarse intrinsic
409 collagen fibers observable in cross (e.g. Fig. 1B) and longitudinal section (e.g. Fig. 1E).

410 In cross sections under cross-polarized light, IFM is easily identifiable by the presence of a
411 networks of intrinsic fibers, birefringent in cross sections (Fig. 1B), conspicuous against the dark
412 matrix of woven bone (Fig. 1B). The width and length of the intrinsic fibers is variable, and
413 strands intertwine with each other, overlapping in a fabric-weave pattern (Fig. 1 A-C). Circular
414 polarization reveals the true arrangement of these fibers to be circular and coiled (Fig. 1C). The
415 rectangular and hexagonal shape seen with crossed polarizers is thus revealed to be an artifact
416 resulting from the Maltese cross effect. The IFM shows heterogeneity in brightness. The areas of
417 denser fibers often correlate with lower brightness under cross-polarized light in both
418 longitudinal and transverse sections (Fig. 1E).

419 In longitudinal section, IFM is characterized by bundles of short parallel fibers that
420 intertwine at various angles, from acute to orthogonal (Fig. 1E). These extend across the surface
421 paralleling the direction and angles of the vascular canals (Fig. 1E). The fibers appear as black
422 strands in the tissue and show no birefringence, similar to osteocyte lacunae and canaliculi,
423 indicating a non-mineralized state of these structures (e.g., Wolf et al. 2012).

424 Osteocyte lacunae are extremely numerous in the IFM and show a wide variety of shapes,
425 from irregularly plump to discoid flattened (Fig. 1D, F). The distribution of osteocyte lacunae is
426 generally irregular with no apparent relationship to other histologic features. Lacunae are very
427 dense in some areas and almost absent in others. These dense irregular osteocyte lacunae are left
428 by multipolar static osteocytes, as is typical of a woven-fibered matrix (Stein & Prondvai 2014,
429 Buffr  nil & Quilhac 2021a). Osteocytes tend to form chaotic clusters where strands and bundles
430 of non-mineralized fibers are present (Fig. 1E). Given the variability in shape and size of both
431 the lacunae and their canaliculi (which are sometimes visible, sometimes not), the more spindle-
432 shaped osteocyte lacunae found in the IFM may represent fibrocytes. The number of osteocyte
433 lacunae is also high in the primary osteons, with a centripetal density increase.

434

435 **3.2 Histology of indeterminate cortical fragments from Bonenburg, Germany**

436 The largest cortical fragment from Bonenburg (WMNM P88133), the thin sections
437 produced from WMNM P-uncatalogued (probably derived from cranial material) and the
438 numerous smaller unidentified cortical fragments (for which no precise anatomical placement is
439 possible) share the same primary bone tissue and overall general features (Figs 4, S5, S6).
440 WMNM P88133 has a primary cortex rather similar to the previously discussed samples from
441 Europe. However, observation of histology of this and most of the other Bonenburg samples is
442 hampered by a nearly opaque outer diagenetic zone >2 mm wide (Fig. 4A, B). The remaining
443 bone tissue is very well preserved. Macroscopically, the outer bone surface bears fine
444 longitudinal striations (Fig. S4A, S5A, S6A), of the same kind noted above for the Aust and
445 Lilstock specimens (Fig. S3D).

446 As in the other specimens, vascularization is strictly longitudinal (Figs 4A, B, S5B, C,
447 S6B, C). Simple vascular canals and primary osteons are arranged in surface-parallel rows which
448 may be enhanced by GM bordering and embracing the vascular canals (Figs 4B, S6C). An
449 external fundamental system does not appear to be present. The bone matrix, in which the
450 vascular canals and primary osteons are set, is IFM, and together they form PIFT (Figs 4C, S5C,
451 S6C, D).

452 The GM show alternations of differently colored IFM but do not show an appreciable
453 pattern in spacing, while they appear to show differences in fiber density (Figs 4C, S6C, D).
454 Under cross-polarized light, it is possible to observe clearly bright coarse fibers in the paler
455 yellow areas, with a reduction of their presence corresponding with increased darkness in areas
456 of darker brown color (Figs 4B, C, S5C, D). The darkest GM seem to be made up by fewer
457 intrinsic fibers (Fig. 4C).

458 Osteocyte lacunae are numerous in the IFM with mainly plump and irregularly shaped ones
459 throughout tissue, while more flattened ones are scarcer and present only in centripetal lamellae
460 of osteons (Fig. 4D). Osteocyte lacunar density and size is greater in the primary bone matrix
461 compared to the lamellar bone of the osteons (Fig. 4D). 

462 The largest fragment (WMNM P88133) is characterized by a continuous gradient in bone
463 compactness from the inner more cancellous area to the outermost cortex (Fig. 4A), resulting
464 from the advancement of the Haversian substitution and resorption fronts. A wide and diffuse

465 Haversian substitution front is detectable toward the center of the section, evidenced by the
 466 interruption of the semicircular GM (Fig. 4B). Elsewhere in the section, secondary osteons
 467 develop preferentially within primary ones, conserving the primary arrangement of the osteon
 468 rows (Fig. 4B, C). Rarely, there are secondary and primary osteons showing an infill of
 469 centripetal layers of anisotropic woven bone (Fig. 4D) alongside osteons showing presence of
 470 intrinsic fibers similar to the ones of the surrounding IFM.

471 In the deep cortical area, there is a high amount of resorption cavities eroded into the
 472 compact bone which consist of primary bone only partially replaced by secondary remodeling.
 473 Resorption cavities are lined by lamellar bone resulting in an increasingly cancellous condition.

474 The longitudinal section (Fig. 4E) is dominated by simple longitudinal canals with a very
 475 limited degree of anastomosis. It is possible to observe diffuse strands of thin, dark fibers of
 476 variable length, distributed mainly longitudinally (Fig. 4E). Sometimes, the fibers intersect each
 477 other next to the edges of the vascular openings (Fig. 4E). The borders of secondary osteons are
 478 lined by bright lamellar bone, darker bone tissue, and by both types together in an alternating
 479 fashion.

480

481 3.3 Histology and microanatomy of *Shastasaurus sikanniensis* holotype jaw bone samples

482 Both of the jaw bone samples from the holotype of *S. sikanniensis* show poor histological
 483 preservation, which hides most of the discernable features in the areas where bone is most altered.
 484 This is particularly evident in the surangular. Poor preservation of birefringence is accompanied
 485 by a dark brown stain of the tissue, making it nearly opaque. However, with sufficiently bright
 486 illumination, the salient features, in particular the presence of IFM, can be discerned (Fig. 5B-D).
 487 Both the surangular and splenial histology are characterized by highly spongy secondary bone
 488 tissue (porosity ~ 82% and ~ 60%, respectively), dark brown in color under the crossed polarizers
 489 (Fig. 5B). Towards the outer bone surface, which appears to be compromised by preparation (see
 490 below), there are interstitial areas of primary tissue characterized by distinctive IFM, with an
 491 outwards increase in frequency. Although no obvious dense cortical bone is present, a decrease in
 492 porosity is detectable toward the outer bone surface of both samples (respectively 64% and 43%
 493 porosity) with smaller longitudinal vascular cavities and higher compactness. The vascularization,
 494 consisting of large Haversian canals and resorption cavities, is strictly longitudinal (Figs 5A, S3D,

495 S7E, F). This is also seen with the naked eye on the outer bone surface which shows regular fine
496 striations (Fig. S3E) as observed in the European specimens.

497 The secondary osteons visible in thin section have only few centripetal lamellae. On the
498 outer bone surface, the presence of osteons half cut open indicates the removal of tissue due to
499 taphonomic or diagenetic causes or harsh preparation (Figs 5A, S3D). It is not possible to
500 determine the presence of secondary osteons within primary ones in the trabecular bone. It is
501 debatable, though, whether the absence of secondary osteons within primary ones is genuine or
502 simply related to the lack of enough compact bone tissue in the sampled location of the jaw
503 bones.

504

505 4. Discussion

506

507 4.1 Rejection of the “Dinosaur Hypothesis”

508 Although we do not question the ichthyosaurian status of the Lilstock and Autun specimens
509 based on their morphology (Fisher *et al.* 2014, Lomax *et al.* 2018), the morphological information
510 provided by the more fragmentary specimens (BRSMG-Cb-3869, BRSMG-Cb-3870, BRSMG-
511 Cb-4063 from the UK and WMNM P88133 from Bonenburg, Germany) is insufficient for
512 recognizing their systematic affinities (beyond excluding certain identifications, like as
513 ichthyosaur or plesiosaur long bones). Thus, a comprehensive histological comparison was
514 needed. Based on the histological evidence obtained from sampling bonafide (i.e., *S. sikanniensis*)
515 and putative Late Triassic giant ichthyosaurs, we regard as relevant four histological features
516 (Table 2), three of which had already been noted in the histological study of two Aust Cliff
517 samples by Redelstorff, Sander & Galton (2014). IFM had not been reported by these authors, but
518 was recognized by us in the same Aust Cliff thin sections.

519 All four features (Table 2) are present in the Lilstock and Autun ichthyosaur jaw specimens
520 from the European Rhaetian. The most distinctive feature, IFM, is present in the jaw of the type
521 specimen of the best-known giant ichthyosaurs, *S. sikanniensis*. The uniqueness of IFM thus
522 provides strong support for the “Giant Ichthyosaur Hypothesis”. Redelstorff, Sander & Galton
523 (2014) concluded that the histology of the Aust Cliff bone segments did not resemble the histology
524 of any known dinosaur long bones at the time. This statement still holds true, especially since the

525 histology of virtually all dinosaur clades, and especially all large-bodied ones, is known by now.
526 We thus can confidently reject the Dinosaur Hypothesis.

527

528 4.2 Testing other possible affinities using histology

529 To test for the presence of a similar combination of features and to further test the "Giant
530 Ichthyosaur Hypothesis", we performed extensive histological comparisons, considering a "Non-
531 dinosaur Hypothesis", addressing known large or giant Late Triassic tetrapods, both terrestrial and
532 aquatic. The results of this comparison are summarized in Table 2 and discussed in more detail
533 below. We found that the unique combination of histological features of the Aust Cliff bone
534 segments and German cortical fragments, combined with their large size and shaft-like shape,
535 rules out affinities with any other Late Triassic giant tetrapod, dinosaurian or non-dinosaurian,
536 other than giant ichthyosaurs.

537 Given their large size (Fig. S3B) and thick cortex, the "mystery bones" must have come
538 from large tetrapods with body masses of several hundred kilograms. This is at the upper limit of
539 non-dinosaurian archosauromorphs and non-eutherian synapsids, sauropterygians and
540 temnospondyl amphibians. A brief review of potential tetrapod candidates is given, but none
541 offers a good fit; see Table 2 for a summary.

542

543 4.2.1 Archosauriformes

544 Among archosaurs, Crurotarsi presents Late Triassic forms with a generally S-shaped but
545 sometimes straighter femur morphology, and it is possible that giant forms would have evolved
546 straight propodial bones as seen in dicynodonts (Sulej & Niedźwiedzki 2019), although
547 convincing finds are lacking. The histology of large rauisuchians has been described in two
548 genera, *Postosuchus* (4-5 m body length) and *Batrachotomus* (6 m). In *Postosuchus* from the Late
549 Triassic of Texas, the femur shows a coexistence of lamellar-zonal tissue and a woven-parallel
550 complex with sub-plexiform to laminar organization, while the outer cortex is lamellar-zonal
551 (Ricqlès *et al.* 2003; Buffrénil *et al.* 2021). Therefore, the pattern and degree of vascularization
552 and the abundance of lamellar-zonal tissue are not compatible with our observations.

553 The histology of *Batrachotomus* (Klein *et al.* 2017) is also discussed here, despite its
554 considerably greater geologic age (Ladinian, Middle Triassic), because of its presumed
555 acquisition of gigantism through an increase in growth rate (Klein *et al.* 2017); *Batrachotomus*

thus exemplifies a hypothetical, fast-growing Rhaetian giant rauisuchian. The femur of *Batrachotomus* exhibits a highly vascularized woven-parallel complex that is more highly vascularized than that of *Postosuchus*, but the vascular organization is laminar to sub-plexiform, and secondary remodeling is rare, represented only by incipient secondary osteons (Klein *et al.* 2017). These features are significantly different from the woven-parallel complex with longitudinal osteons and the strong secondary remodeling in the deep cortex we observed. Thus, we conclude that even a plausible giant, fast-growing rauisuchian must be excluded from consideration.

Although much smaller, aetosaurs show some superficial similarity in histologic features (Buffrénil *et al.* 2021), but can easily be excluded. Aetosaur histology shows a general predominance of lamellar-zonal tissue or a laminar woven-parallel complex transitioning outward to poorly to non-vascularized lamellar-zonal tissue (Ricqlès *et al.* 2003; Buffrénil *et al.* 2021), a less vascularization, and less well-organized secondary remodeling (Ricqlès *et al.* 2003; Buffrénil *et al.* 2021).

Histology of the femora of large phytosaur specimens shows lamellar-zonal bone tissue (Ricqlès *et al.* 2003; Buffrénil *et al.* 2021). In addition, a gradual decrease in vascularization toward the outer cortex, a poorly vascularized lamellar-zonal outer cortex, and scattered, unorganized secondary remodeling of the cortex have been reported (Ricqlès *et al.* 2003; Buffrénil *et al.* 2021). These features are marginally consistent with those reported in this work, but more specimens would have to be added to the comparison sample. However, IFM and secondary osteons within primary osteons have not been reported for any of the Crurotarsi considered, and for this and the other reasons listed above, we have excluded them from further consideration.

4.2.2 Triassic non-mammalian synapsids

Among Triassic non-mammalian synapsids, Kanemeyeriiform dicynodonts are known to have reached large to giant sizes (3-4 m) in the Late Triassic (Benton, 2015; Sulej & Niedzwiedzki, 2019). Such animals had already been excluded by previous authors in the context of the "mystery bones" based on the morphology of the long bones (Redelstorff *et al.* 2014). The histology of the kannemeyeriiforms is somewhat closer to our samples in that they have a woven-parallel complex with longitudinal osteons often bordered by GM (Chinsamy & Rubidge, 1993; Green *et al.* 2010; Buffrénil *et al.* 2021), but the vascularization of the large *Placerias* specimens

appears to be less than that of the "mystery bones". Moreover, the avascular or nearly avascular outer cortices of lamellar bone reported for both propodials and epipodials, together with the presence of scattered and rather chaotically arranged secondary osteons (Green et al. 2010; Buffrénil et al. 2021), contrast with the outer cortical vascularization and characteristic secondary remodeling (including secondary osteons within primary osteons) observed in our specimens. Due to its large size, the largest known Kanemeyeriiform dicynodont, *Lisowicia bojani*, is more highly vascularized, but does not show the same Haversian organization in ordered periosteal rows as in our specimens (Sulej & Niedzwiedzki, 2018 Fig. s14). Finally, the presence of an IFM-like primary matrix has not been reported for *Placerias*, *Kannemeyeria*, or *Lisowicia* (Chinsamy & Rubidge, 1993; Green et al. 2010, Sulej & Niedzwiedzki, 2018).

597

4.2.3 Sauropterygians

Sauropterygians were an important component of the Late Triassic faunas, but it would be difficult to identify representatives with bones of the size range of the specimens studied. Large rib specimens of *Nothosaurus* show a tissue rich in extrinsic fibers superficially resembling IFM and longitudinal vascular canals (Klein et al. 2019, fig. 4k, n, o), but other *Nothosaurus* ribs show parallel-fibered bone tissue and radial vascularization (Klein et al. 2019, fig. 4l), so the occurrence of an IFM-like matrix and longitudinal vascularization does not appear to be consistent within the genus. The histology of plesiosaurs, including the only Triassic one, has recently been extensively studied (Wintrich et al. 2017; Sander & Wintrich, 2021), but the complete lack of dermatocranial samples (Sander & Wintrich, 2021) for comparison prevents us from testing the hypothesis of a large unknown Triassic form. The histology of plesiosaur propodials, which are dominated by radial vascularization at mid-shaft (Wintrich et al. 2017; Sander & Wintrich, 2021), is certainly not consistent with the results of this study, but interestingly, secondary remodeling in plesiosaurs appears to follow pre-existing radial canals (Sander & Wintrich, 2021, p. 449), similar to what we described as template remodeling.

613

4.2.4 Temnospondyl amphibians

Recently, the idea that many temnospondyl clades may have persisted, even with large-bodied forms, until the very end of the Triassic has been proposed (Steyer & Damiani, 2005, Sander et al. 2016; Konietzko-Meier et al. 2018). Considering also the first attribution of the Aust

618 Cliff bones to 'Labyrinthodontia' (Stutchbury, 1850), it seems appropriate to include
619 temnospondyls in our comparison.

620 Late Triassic *Metoposaurus* mandibles (Gruntmejer et al. 2021) and long bones (Konietzko-
621 Meier & Sander, 2013), as well as an indeterminate Late Triassic temnospondyl humerus
622 (Konietzko-Meier *et al.* 2018) show the diffuse presence of an IFM-like bone matrix in the bone
623 cortices, as well as a general longitudinal orientation of the vascular canals. The poor primary
624 vascularization and the rather disorganized and scattered secondary remodeling (Konietzko-Meier
625 & Sander, 2013; Konietzko-Meier et al. 2018; Gruntmejer et al. 2021) are different from our
626 specimens, but Konietzko-Meier et al. (2018) report that "in temnospondyls, the remodeling
627 process always follows the vascular pattern of the primary tissue, unlike in Amniota....".
628 Nevertheless, the differences in morphology and histology are too great between the material
629 studied here and temnospondyls to support such an affinity.

630

631 **4.3 IFM and PIFT and possible analogs**

632 Although PIFT has not been explicitly described in the literature, it is not uncommon to see
633 published micrographs seemingly showing this type of bone tissue or similar ones. A brief, but
634 probably incomplete, list of examples includes a wide variety of amniotes: the rib sample of a
635 large *Nothosaurus* specimen (Klein, Canoville & Houssaye 2019, fig. 4n, o) (Fig. S8A, B), the
636 femur of *Simosaurus* (Klein & Griebeler 2016, fig. 5) (Fig. S8H, I), various bones of the
637 thalattosaur *Askeptosaurus* (Klein *et al.* 2023) (Fig. S8D, E), the rib of the thalattosuchian
638 crocodylomorph *Metriorhynchus* (Buffrénil, Quilhac & Cubo 2021 fig. 10.2f), and the humerus of
639 the ornithomimid dinosaur *Telmatosaurus* (Buffrénil & Quilhac 2021a, fig. 8.6a). Some of the more
640 suggestive cases noted above are discussed in Supplemental Article S2 and figured in Fig. S8.

641

642 **4.4 IFM, PIFT and ossified tendons**

643 IFM remarkably resembles extrinsic fibers bundles seen in metaplastic bone tissue of
644 osteoderms (Scheyer & Sander 2004) and longitudinal fiber bundles of ossified tendons of various
645 dinosaurs (Horner, Woodward & Bailleul 2016; Surmik *et al.* 2023). However, we introduced the
646 new terms IFM and PIFT in order to set this clearly periosteal matrix and tissue apart from
647 metaplastic tissues. Nevertheless the similarity of PIFT with metaplastic bone tissue would
648 suggest a shared osteogenetic process. Horner, Woodward & Bailleul (2016) distinguish

649 metaplastic mineralized tissue from periosteal bone by the lack of true Haversian remodeling and
 650 osteocyte lacunae. However, for the purpose of our comparison, we disagree with this notion, in
 651 agreement with Organ & Adams (2005), Surmik *et al.* (2023) and with our personal observations
 652 on ossified tendons stored in the IGPB histological collection.

653 Aside from the similarity between IFM and longitudinal extrinsic fibers, it is possible to
 654 observe further similarities with ossified tendons. The longitudinal strands of unmineralized fibers
 655 are in a herringbone pattern in both cases (compare Figs 1E, 4E, S5D and, e.g., Horner *et al.* fig.
 656 2g, Surmik *et al.* fig. 2f), and there are numerous irregular, sometimes elongate cell lacunae
 657 somewhat resembling fibrocytes in shape (Horner *et al.* 2016). Finally, it is possible to see the
 658 presence of occasional centripetal coarse fibrous bone in the Haversian canals of *Meleagris*
 659 *gallopavo* tendon, figured by Adams & Organ (2005, fig. 2c, d).

660 The GM in our specimens find close similarity with the structures reported by Horner,
 661 Woodward & Bailleul (2016) as regions of varying primary orientation and density of the fibers
 662 (Horner, Woodward & Bailleul 2016 fig. 2d-f). The hypothesis proposed by Horner, Woodward &
 663 Bailleul (2016), that the variable color of similar structures is related to the density and orientation
 664 of fibers observed in longitudinal sections in ossified tendons, fits our observations (Fig. S6C, D)
 665 and explains the appearance of such marks. Contrary to what was reported by Horner, Woodward
 666 & Bailleul (2016) for ossified tendons, the GM are identifiable as classical cycles of periosteal
 667 apposition, given the clear primary origin of these structures in relation to the spatial distribution
 668 of periosteal vascular canals and nutrient foramina, and the presence of osteocyte lacunae.

669 In conclusion, our literature review (and the extensive personal knowledge of the second
 670 author) suggests that IFM is a novel matrix type that has not been reported before in the
 671 osteohistological literature. This leads to the question if a bone tissue formed of IFM may be
 672 viewed as an apomorphy of a clade of giant ichthyosaurs. This hypothesis would have to be tested
 673 by phylogenetic analysis incorporating histological characters, which may well find IFM as a non-
 674 unique synapomorphy, resulting from parallel osteogenetic processes. Alternatively, IFM could be
 675 mapped on an amniote cladogram and may show up as a synapomorphy.

676

677 4.5 Template remodeling, osteons within osteons, and unmineralized fibrous matrices

678 The phenomenon of template remodeling, secondary osteons within primary ones, and
 679 secondary osteons within secondary osteons (i.e., concentric osteons) are a distinctive feature of

680 the histology of the European samples investigated . The unifying feature of all of these types
681 of secondary remodeling is the reuse by the basic structural unit (bone remodeling unit) of
682 preexisting vascular pathways, be they primary or themselves the result of previous remodeling
683 activity. This reuse of existing pathways is unusual for secondary osteons which in dinosaurs,
684 mammals, and most other amniotes show little regard for the primary histology (Mitchell 2017).
685 We here emphasize the unusual nature of the secondary osteons within primary ones as a pattern
686 we observe. Whereas this pattern may be a special feature of the specimens investigated here, it
687 also could result from the strictly longitudinal orientation of the primary osteons or from the
688 underlying propensity of reuse of vascular pathways. Although we are currently uncertain which
689 of these mechanisms is at work, this does not diminish the systematic value of the secondary
690 osteons within the primary ones.

691 As already noted, bone remodeling involving pre-existing primary or secondary osteons has
692 been reported in various aquatic tetrapod taxa, such as in the long bones of plesiosaurs (Sander &
693 Wintrich 2021) and in temnospondyls (Konietzko-Meyer et al. 2019). Klein et al. (2015) describe
694 “secondarily widened primary osteons” (Klein et al. 2015 figs. 7, 8, s5) in various placodonts
695 (Sauropterygia). With this term, they refer to the normal transformation process of compact bone
696 to spongy bone. Specifically, these authors note that the resorption activity leading to cancellous
697 bone, i.e., the resorption front of Mitchell & Sander (2014), originates from pre-existing vascular
698 canals. In this way, there is a similarity to secondary osteons within primary osteons. The
699 difference, however, is that in our material the secondary osteons within the primary ones do not
700 lead to cancellous bone, but the tissue remains compact. The transformation to cancellous bone
701 occurs deeper in the cortex. An interesting report is that by Surmik *et al.* (2023) of what appear to
702 be secondary osteons within primary osteons in ossified tendons of ornithischian dinosaurs
703 (Surmik *et al.* 2023 fig. 2d). Although confirmation of the presence of this feature would require
704 direct observation of the Surmik *et al.* (2023) sections, this potential occurrence may be
705 informative on the underlying mechanism of vascular architecture reuse.

706 The occurrence of a common unusual feature in bone tissue formed by different processes
707 (e.g., periosteal apposition in mandibles and long bones vs. metaplastic ossification of ossified
708 tendons and osteoderms) suggests a common constraint as explanation. In the process of bone
709 resorption, osteoclasts are unable to act on the mineralized bone matrix until the organic protective
710 layer of bone lining cells is removed by cambial cells (Zylberberg 2021). It has also been

hypothesized that sites characterized by non-mineralized structures are less attractive or accessible to osteoclasts (Aaron 1980, 2012; Jones, Boyde & Ali 1984). The widespread presence of non-mineralized fibers in a bone tissue may significantly inhibit the progression of the basic structural units. The absence of unmineralized fibers in the osteonal bone matrix may thus induce primary osteons to serve as preferential "highways" for osteoclast activity, especially during the initial resorptive phase (i.e., the resorption front), thus explaining the occurrence of abundant secondary osteons within primary osteons and template remodeling.

Alternatively, a difference between the regulatory signals emanating from osteocytes in the outer cortical matrix and those in the osteonal bone matrix may be the primary driver of osteoclast regulation and attraction. Osteocyte regulatory activity is known to be influenced by mechanical loading during development and appears to vary with lacunar shape (van Oers, Wang & Bacabac 2015). Therefore, it is plausible that the numerous and highly heterogeneous lacunar spaces observed in the matrix may have played a critical additional role.

4.6 Implications of PIFT for growth rate, gigantism and feeding behavior

Several of the features we described are commonly associated with fast growth rates, the most common being a histology dominated by a woven-parallel complex (most commonly fibrolamellar bone), a high degree of vascularization, a high rate of remodeling with multiple generations of osteons, and a high number of osteocyte lacunae, both irregular and spindle-shaped (Buffrénil & Quilhac 2021b). The presence of numerous open canals in the outer cortex and a well-vascularized outer periosteal surface indicate for all bones sampled that the animals were actively growing at the time of death. The presence of unmineralized fibers in the cortex could be related to rapid mineralization of the osteoid layer laid down by the periosteum as well as to the presence of fibrocytes (Buffrénil & Quilhac et al. 2021a). The latter cell type would be rather unusual in the formation of periosteal bone, however. The occasional presence of woven bone as infill of osteons may be another feature indicating rapid bone deposition.

The similarity between the bone matrices of giant ichthyosaur mandibles, ossified tendons, and osteoderms invites speculations on the biomechanical properties of the former (as already done by Horner, Woodward & Bailleul 2016 with the hadrosaur nasal). For example, the largest bone segment from Aust Cliff has been suggested to belong to an animal in the size range of modern blue whales (Lomax *et al.* 2018). Although the feeding strategy of these giant

742 ichthyosaurs remains unknown, it is reasonable to assume that their large jaws were adapted to
 743 withstand significant stress associated with hunting and feeding underwater, similar to the feeding
 744 behavior of blue whales, which actively process thousands of liters of seawater in one gulp
 745 (Goldbogen et al. 2007). Given the high tensile strength of mineralized ossified tendons, it is
 746 possible that these large ichthyosaur jaws were selected to withstand similar stresses, either during
 747 simple opening, as in baleen whales, or during potential ramming behavior, as observed in
 748 odontocetes such as killer whales. At the same time, the high amount of unmineralized fibers in
 749 the longitudinal axis of the mandible would have provided some flexibility in different bending
 750 planes (Horner, Woodward & Bailleul 2016). The high rate of remodeling, typically associated
 751 with bones subjected to loading, is another factor supporting this hypothesis. It is possible that the
 752 presence of specialized soft tissues, such as muscle and connective tissue, likely played an
 753 important role in the development of this peculiar histology (Organ & Adams 2005; Klein,
 754 Christian & Sander 2012; Horner, Woodward & Bailleul 2016). The occurrence of specializations
 755 for buccal processing of large amounts of water (relative to body size) is not isolated within
 756 Ichthyosauromorpha (Fang et al. 2023) and is expected in the evolutionary context of achieving
 757 giant sizes in marine environments (Sander et al. 2021).

758

759 5. Conclusions

760 Paleohistology can be a powerful tool for determining the taxonomic affinity of
 761 fragmentary bone specimens, as has been demonstrated in dinosaur studies previously (e.g.,
 762 Garilli et al. 2009; Hurum et al. 2006). However, paleohistology can also be used to show that
 763 dinosaur-sized fragmentary bones do not belong to dinosaurs at all. Our study does just that,
 764 ruling out Sauropodomorpha and Stegosauria as possible sources of the mysterious large bone
 765 segments and fragments found in the European Rhaetian, thus rejecting the Dinosaur Hypothesis
 766 and instead supporting the Giant Ichthyosaur Hypothesis laid out by Lomax et al. (2018).

767 There are four distinctive histologic features common to the very large indeterminate bone
 768 segments and cortical fragments from the European Rhaetian: 1) IFM, 2) strictly longitudinal
 769 vascular architecture in the primary cortex, 3) closely spaced skeletal growth marks structuring
 770 primary osteons and vascular canals, and 4) abundance of secondary osteons within primary
 771 osteons. While IFM as a type of woven-fibered matrix and secondary osteons within primary
 772 osteons have rarely been observed in amniotes, the combination of all four features is unique to

773 the material sampled here, and even small fragments of bone cortex, e.g. from Bonenburg,
 774 Germany, are diagnostic. The same four histologic features are present in giant ichthyosaur jaw
 775 bones from the Rhaetian of the UK (Lilstock) and France (Autun). Two of the features, the
 776 unique IFM and the strictly longitudinal vascular architecture, is also seen in the jaw bones of the
 777 giant ichthyosaur *Shastasaurus sikanniensis* from the middle Norian of Canada. The four
 778 features in combination are absent in dinosaur histological samples, and two, IFM and secondary
 779 osteons within primary osteons (as a pervasive pattern), are not known from dinosaur histology.
 780 Similarly, we reject any affinities with hypothetical giant Crurotarsi, Kannemeyeriiformes, and
 781 Plesiosauria. We note some similarities with other secondarily aquatic tetrapods
 782 (Temnospondyli, thalattosaurs and possibly large nothosaurs), but these groups are also rejected
 783 due to significant size and morphological differences.

784 The histology reported here is thus that can be used to reliably identify cortical bone
 785 segments as belonging to giant ichthyosaurs, overcoming the problem of scarce morphological
 786 evidence. We conclude that the large bone segments from Aust Cliff are indeed fragments of
 787 giant ichthyosaur jaws, as are the cortical fragments from Bonenburg. WMNM P88133 and
 788 WMNM P-uncatalogued indicate animals comparable in size to the British and French
 789 mandibular fragments, suggesting the potential for similar discoveries of very large-bodied
 790 ichthyosaurs in the Exter Formation of northern Germany.

791 The common occurrence of a unique bone matrix type, IFM, in several giant Late Triassic
 792 ichthyosaurs indicates a shared ossification strategy in their lower jaws. IFM appears to be
 793 associated with closely spaced growth marks that show rhythmic changes in bone formation and
 794 template remodeling produced by the reuse of existing vascular architecture by the basic
 795 structural unit during remodeling. These features may be apomorphic for a clade of giant
 796 ichthyosaurs and/or related to specific biomechanical properties of their mandibles. More
 797 comparative histological samples of ichthyosaurs and more complete specimens are needed to
 798 confirm these hypotheses.

799 Finally, our study shows that there still are novel bone matrix and bone tissue types to be
 800 discovered that are restricted to a specific extinct clade. IFM and PIFT are apparently extinct,
 801 and future work must address the evolutionary, phylogenetic, and developmental dynamics
 802 associated with the nature of IFM and its possible unrecognized presence in modern animals and
 803 the fossil record, and the reasons for its strong resemblance to the products of metaplastic

804 ossification of extrinsic fibers, despite IFM being composed of intrinsic fibers in the periosteal
805 territory.

806

807 6. Acknowledgements

808 The authors are deeply indebted to Deborah Hutchinson and Roger Vaughan (BRSMG),
809 Valentin Fischer (University of Liège, Belgium), Brandon Strelitzky and Don Brinkman (Royal
810 Tyrrell Museum of Paleontology, Drumheller, Alberta, Canada), and Achim Schwerman
811 (Westphalian Museum of Natural History, Münster, Germany) for access, permission to
812 sample, and assistance with photographs and information about the specimens in their care.
813 Olaf Dülfer and Pia Schucht (University of Bonn, Germany) are thanked for assistance and
814 preparation of the thin sections. René-Paul Eustache (Combon, France) helped us to modify the
815 Leica polarizing microscope for circular polarization. We thank Dorota Konietzko-Meier and
816 Sudipta Kalita (University of Bonn, Germany), Peter Galton (University of Bridgeport,
817 Connecticut, USA), Dean Lomax (University of Manchester, UK), and Paul de la Salle (The
818 Etches Collection, Wareham, UK) for information and discussions. Many thanks to Andrzej
819 Wolniewicz (Polish Academy of Sciences, Warsaw, Poland) for pointing out the von Huene
820 study (1912). Finally, we would like to thank the editor, Mark Young, and the three reviewers
821 (Megan Withney and Christopher Griffin and an anonymous reviewer) for their work and
822 useful comments that greatly improved this manuscript.

823

824 7. References

825

826 Aaron J. E. 1980. Demineralization of bone in vivo and in vitro. *Metab. Bone Dis. Relat. Res.* 2S,
827 117– 125.

828 Aaron J. E. 2012. Periosteal sharpey's fibers: A novel bone matrix regulatory system. *Frontiers in*
829 *Endocrinology* 3,1– 10. DOI 10.3389/fendo.2012.00098.

830 Adams J.S. and Organ C. L. 2005. Histologic determination of ontogenetic patterns and processes
831 in hadrosaurian ossified tendons. *Journal of Vertebrate Paleontology*, 25(3), 614– 622.

832 Barth G., Pieńkowski G., Zimmermann J., Franz M. and Kuhlmann G. 2018. Palaeogeographical
833 evolution of the Lower Jurassic: High-resolution biostratigraphy and sequence stratigraphy
834 in the Central European Basin. *Geological Society Special Publication*, 469, 341– 369.

835Benton M. J. 2015. *Vertebrate Palaeontology* (4th ed.). Wiley-Blackwell Ltd., Chichester, 506 pp.

836Benton M. J. and Spencer P. S. 1995. *Fossil Reptiles of Great Britain*, Springer Netherlands,
837 Dordrecht, 386 pp.

838Bianucci G., Lambert O., Urbina M., Merella M., Collareta A., Bennion R., Salas-Gismondi R.,
839 Benites-Palomino R., Post K., de Muizon C., Bosio C., Di Celma C., Malinverno E.,
840 Pierantoni P.P., Villa I. M. & Amson E. 2023. A heavyweight early whale pushes the
841 boundaries of vertebrate morphology. *Nature*. <https://doi.org/10.1038/s41586-023-06381-1>

842Bromage T. G., Goldman H. M., McFarlin S. C., Warshaw J., Boyde A. and Riggs C. M. 2003.
843 Circularly polarized light standards for investigations of collagen fiber orientation in bone.
844 *The Anatomical Record (Part B: New Anat.)*, 274B, 157– 168.

845Buffrénil V. de and Quihac A. 2021a. Bone Tissue Types: A Brief Account of Currently Used
846 Categories. 183–188. In de Buffrénil V., de Ricqlès A.J., Zylberberg L. and Padian K. (Eds.).
847 (2021). *Vertebrate Skeletal Histology and Paleohistology* (1st ed.). Boca Raton: CRC Press.
848 838 pp. <https://doi.org/10.1201/9781351189590>

849Buffrénil V. de and Quihac A. 2021b. Bone Remodeling. 229– 241. In de Buffrénil V., de Ricqlès
850 A.J., Zylberberg L. and Padian K. (Eds.). (2021). *Vertebrate Skeletal Histology and*
851 *Paleohistology* (1st ed.). Boca Raton: CRC Press. 838 pp.
852 <https://doi.org/10.1201/9781351189590>

853Buffrénil V. de, Quihac A. and Cubo J. 2021. Accretion Rate and Histological Features of Bone.
854 221– 227. In de Buffrénil V., de Ricqlès A.J., Zylberberg L. and Padian K. (Eds.). (2021).
855 *Vertebrate Skeletal Histology and Paleohistology* (1st ed.). Boca Raton: CRC Press. 838 pp.
856 <https://doi.org/10.1201/9781351189590>

857Buffrénil V. de, Zylberberg L. 2021. Remarks on Metaplastic Processes in the Skeleton. 247– 254.
858 In de Buffrénil V., de Ricqlès A.J., Zylberberg L. and Padian K. (Eds.). (2021). *Vertebrate*
859 *Skeletal Histology and Paleohistology* (1st ed.). Boca Raton: CRC Press. 838 pp.
860 <https://doi.org/10.1201/9781351189590>

861Callaway J. M., and J. A. Massare. 1989. *Shastasaurus altispinus* (Ichthyosauria, Shastasauridae)
862 from the Upper Triassic of the El Antimonio District, Northwestern Sonora, Mexico. *Journal*
863 *of Paleontology*, 63, 930– 939.

864Camp C. L. 1976. Vorläufige Mitteilung über große Ichthyosaurier aus der oberen Trias von
865 Nevada. *Sitzungsberichte der Akademie der Wissenschaften, mathematisch-*
866 *naturwissenschaftliche Klasse*, 185, 125– 134.

867Camp C. L. 1980. Large ichthyosaurs from the Upper Triassic of Nevada. *Palaeontographica, AI*,
868 170, 139– 200.

869Chen X-h., Motani R., Cheng L., Jiang D-y. and Rieppel O. 2014. The enigmatic marine reptile
870 *Nanchangosaurus* from the Lower Triassic of Hubei, China and the phylogenetic affinities of
871 *Hupehsuchia*. *PLoS ONE* 9:e102361. 10.1371/journal.pone.0102361

872Chinsamy A. and Rubidge B.S. 1993. Dicynodont (Therapsida) bone histology: phylogenetic and
873 physiological implications. *Palaeontologia Africana*, 30, 97– 102.

874Cross S. R. R., Ivanovski N., Duffin C. J., Hildebrandt C., Parker A. and Benton M. J. 2018.
875 Microvertebrates from the basal Rhaetian Bone Bed (latest Triassic) at Aust Cliff, S.W.
876 England. *Proceedings of the Geologists' Association*, 129, 635– 653.

877Davies J., Marzoli A., Bertrand H., Youbi N., Ernesto M. and Schaltegger U. 2017 End-Triassic
878 mass extinction started by intrusive CAMP activity. *Nat Commun* 8, 15596.
879 <https://doi.org/10.1038/ncomms15596>

880Fang ZC., Li JL., Yan CB., Zou YR. Tian L., Zhao B. Benton M. J., Cheng L. and Lai XL. 2023.
881 First filter feeding in the Early Triassic: cranial morphological convergence between
882 *Hupehsuchus* and baleen whales. *BMC Ecol Evo*, 23, 36. [https://doi.org/10.1186/s12862-023-](https://doi.org/10.1186/s12862-023-02143-9)
883 02143-9

884Fischer V., Cappetta H., Vincent P., Garcia G., Goolaerts S., Martin J. E., Roggero D. and Valentin
885 X. 2014. Ichthyosaurs from the French Rhaetian indicate a severe turnover across the
886 Triassic–Jurassic boundary. *Naturwissenschaften*, 101, 1027– 1040.

887Galton, P. M. 2005. Bones of large dinosaurs (Prosauropoda and Stegosauria) from the Rhaetic
888 Bone Bed (Upper Triassic) of Aust Cliff, southwest England. *Revue de Paleobiologie*, 24, 51–
889 74.

890Garilli V., Klein N., Buffetaut, E., Sander P. M., Pollina F., Galletti L., Cillari A. and Guzzetta D.
891 2009. First dinosaur bone from Sicily identified by histology and its paleobiogeographical
892 implications. *Neues Jahrbuch für Geologie und Paläontologie - Abhandlungen*, 252, 207–
893 216.

894Goldbogen J.A., Calambokidis J., Oleson E., Potvin J., Pyenson N.D., Schorr G., Shadwick R.E.
895 2011. Mechanics, hydrodynamics and energetics of blue whale lunge feeding: efficiency
896 dependence on krill density. *Journal of Experimental Biology*, 214, 131– 146.
897 doi:10.1242/jeb.048157

898Gravendyck J., Schobben M., Bachelier J. B., and Kürschner W. M. 2020. Macroecological patterns
899 of the terrestrial vegetation history during the end-Triassic biotic crisis in the central European
900 Basin: A palynological study of the Bonenburg section (NW-Germany) and its supra-regional
901 implications. *Global and Planetary Change*, 194, 103286.

902Green J. L., Schweitzer M. H. and Lamm E. 2010. Limb bone histology and growth in *Placerias*
903 *hesternus* (Therapsida: Amonodontia) from the Upper Triassic of North America.
904 *Palaeontology*, 53(2), 347– 364.

905Gruntmeijer K., Konietzko-Meier D., Bodzioch A. 2016. Cranial bone histology of *Metoposaurus*
906 *krasiejowensis* (Amphibia, Temnospondyli) from the Late Triassic of Poland. *PeerJ*, 4,e2685;
907 DOI 10.7717/peerj.2685.

908Gruntmeijer K., Bodzioch A., Konietzko-Meier D. 2021. Mandible histology in *Metoposaurus*
909 *krasiejowensis* (Temnospondyli, Stereospondyli) from the Upper Triassic of Poland. *PeerJ*,
910 9,e12218; DOI 10.7717/peerj.12218.

911Harper E.M. 2006. Dissecting post-Palaeozoic arms races. *Palaeogeography, Palaeoclimatology*,
912 *Palaeoecology* 232, 322–343.

913Hogler J. A. 1992. Taphonomy and paleoecology of *Shonisaurus popularis* (Reptilia:
914 Ichthyosauria). *Palaios*, 7,108– 117.

915Horner J., Woodward H. and Bailleul A. 2016. Mineralized tissues in dinosaurs interpreted as
916 having formed through metaplasia: A preliminary evaluation. *C.R. Palevol*, 15, 176– 196;
917 DOI 10.1016/j.crpv.2015.01.006.

918Houssaye A., Lindgren J., Pellegrini R., Lee A.H., Germain D. and Polcyn M.J., 2013.
919 Microanatomical and Histological Features in the Long Bones of Mosasaurine Mosasaurs
920 (Reptilia, Squamata) – Implications for Aquatic Adaptation and Growth Rates. *PLoS ONE*
921 8(10): e76741. doi:10.1371/journal.pone.0076741

922Huene, F. v. 1912. Der Unterkiefer eines riesigen Ichthyosauriers aus dem englischen Rhät.
923 *Zentralblatt für Mineralogie, Geologie und Paläontologie*, 1912, 61– 63.

924 Hurum J.H., Bergan M., Müller R., Nystuen J.P. and Klein N. 2006. A Late Triassic dinosaur bone,
925 offshore Norway. *Norwegian Journal of Geology*, 86, 117– 123.

926 Jones S.J., Boyde A. and Ali N.N. (1984) The resorption of biological and non-biological substrates
927 by cultured avian and mammalian osteoclasts. *Anat Embryol*, 170, 247– 256.

928 Kelley N. P. and Pyenson N. D. 2015. Evolutionary innovation and ecology in marine tetrapods
929 from the Triassic to the Anthropocene. *Science*, 348, aaa3716; DOI 10.1126/science.aaa3716.

930 Kelley N. P., Irmis R. B., Depolo P. E., Noble P. J., Montague-Judd D., Little H., Blundell J.,
931 Rasmussen C., Percival L. M. E., Mather T. A. and Pyenson N. D. 2022. Grouping behavior
932 in a Triassic marine apex predator. *Current Biology*, 32, 5398– 5405.e3.

933 Klein N. and Sander P. M. 2007. Bone histology and growth of the prosauropod *Plateosaurus*
934 *engelhardti* MEYER, 1837 from the Norian bonebeds of Trossingen (Germany) and Frick
935 (Switzerland). *Special Papers in Palaeontology*, 77, 169– 206.

936 Klein N. and Sander P. M. 2008. Ontogenetic stages in the long bone histology of sauropod
937 dinosaurs. *Paleobiology*, 34, 247– 263.

938 Klein N. and Griebeler E.M. 2016. Bone histology, microanatomy, and growth of the nothosauroid
939 *Simosaurus gaillardoti* (Sauropterygia) from the Upper Muschelkalk of southern
940 Germany/Baden-Württemberg. *Comptes Rendus Palevol*, 15, 142– 162.
941 <https://doi.org/10.1016/j.crpv.2015.02.009>

942 Klein N., Christian A. and Sander P. M. 2012. Histology shows that elongated neck ribs in sauropod
943 dinosaurs are ossified tendons. *Biology Letters*, 8 1032– 1035.

944 Klein N., Houssaye A., Neenan J.M. and Scheyer T.M. 2015. Long bone histology and
945 microanatomy of Placodontia (Diapsida: Sauropterygia). *Contributions to Zoology*, 84 (1), 59-
946 84.

947 Klein N., Canoville A. and Houssaye A. 2019. Microstructure of vertebrae, ribs, and Gastralia of
948 Triassic sauroptrygians—New insights into the microanatomical processes involved in
949 aquatic adaptations of marine reptiles. *Anatomical Record*, 302, 1770– 1791.

950 Klein N., Sander P. M., Liu J., Druckenmiller P., Metz E. T., Kelley N. P. and Scheyer T. M. 2023.
951 Comparative bone histology of two thalattosaurians (Diapsida: Thalattosauria): *Askeptosaurus*
952 *italicus* from the Alpine Triassic (Middle Triassic) and a Thalattosauroida indet. from the
953 Carnian of Oregon (Late Triassic). *Swiss Journal of Palaeontology*, 142, 15
954 <https://doi.org/10.1186/s13358-023-00277-3>.

955 Konietzko-Meier D. and Sander P. M. 2013. Histology of long bones of *Metoposaurus diagnosticus*
956 *krasiejowensis* (Temnospondyli) from the Late Triassic of Krasiejów (Opole, Silesia Region).
957 *Journal of Vertebrate Paleontology*, 33, 1003– 1018.

958 Konietzko-Meier D., Werner J. D., Wintrich T. and Sander P. M. 2018. A large temnospondyl
959 humerus from the Rhaetian (Late Triassic) of Bonenburg (Westphalia, Germany) and its
960 implications for temnospondyl extinction. *Journal of Iberian Geology*, 45, 287– 300.

961 Kosch B. F. 1990. A Revision of the skeletal reconstruction of *Shonisaurus popularis* (Reptilia :
962 Ichthyosauria). *Journal of Vertebrate Paleontology*, 10, 512– 514.

963 Lacroix P. 1970. Recherches sur le remaniement interne des os. *Arch. Biol. (Liège)*, 81, 275– 304.

964 Lamm E.T. 2013. Preparation and sectioning of specimens; pp. 55–160. In K. Padian, and E.-T.
965 Lamm (Eds.). *Bone Histology of Fossil Tetrapods. Advancing Methods, Analysis, and*
966 *Interpretation*. University of California Press, Berkeley. 298 pp.

967 Lomax D., De la Salle R.P., Massare J. A. and Gallois R. 2018. A giant Late Triassic ichthyosaur
968 from the UK and a reinterpretation of the Aust Cliff ‘dinosaurian’ bones. *PLoS ONE*, 13, 1–
969 16.

970 Liebe L. and Hurum J.H. 2012. Gross internal structure and microstructure of plesiosaur limb bones
971 from the Late Jurassic, central Spitsbergen. *Norwegian Journal of Geology*, 92, 285-309.
972 ISSN 029-196X.

973 Maidment S. C., Norman D. B., Barrett P. M. and Upchurch P. 2008. Systematics and phylogeny of
974 Stegosauria (Dinosauria: Ornithischia). *Journal of Systematic Paleontology*, 6, 367– 407.

975 McGowan C. and Motani R. 1999. A reinterpretation of the Upper Triassic ichthyosaur
976 *Shonisaurus*. *Journal of Vertebrate Paleontology*, 19, 42– 49.

977 Mitchell J. 2017. *Cortical Bone Remodeling in Amniota - a Functional, Evolutionary and*
978 *Comparative Perspective of Secondary Osteons*. PhD Dissertation, University of Bonn, Bonn,
979 Germany. 225 pp.

980 Mitchell J. and Sander P. M. 2014. The three-front model: a developmental explanation of long
981 bone diaphyseal histology of Sauropoda. *Biological Journal of the Linnean Society* 112, 765–
982 781.

983 Naish D. and Martill D. M. 2008. Dinosaurs of Great Britain and the role of the Geological Society
984 of London in their discovery: Ornithischia. *Journal of the Geological Society* 165, 613– 623.

985 Nicholls E. L. and Manabe M. 2004. Giant ichthyosaurs of the Triassic-A new species of
 986 *Shonisaurus* from the Pardonet Formation (Norian, Late Triassic) of British Columbia.
 987 *Journal of Vertebrate Paleontology* 24, 838– 849.

988 Organ C. and Adams J. 2005. The histology of ossified tendons in dinosaurs. *Journal of Vertebrate*
 989 *Paleontology*, 25(3), 602– 613.

990 Padian K. and Woodward H. 2021. Archosauromorpha: Avemetatarsalia – dinosaurs and their
 991 relatives. 511– 549. In de Buffrénil V., de Ricqlès A.J., Zylberberg L., & Padian K. (Eds.).
 992 (2021). *Vertebrate Skeletal Histology and Paleohistology* (1st ed.). Boca Raton: CRC Press.
 993 838 pp. DOI 10.1201/9781351189590

994 Perillo M. and Heijne J. 2023. Storms and bones: evidence for palaeocurrents in the Rhaetian
 995 bonebeds of Bonenburg (Germany).. In Alba D.M., Marigó J., Nacarino-Meneses, C., Villa A.
 996 (Eds.). Book of Abstracts of the 20th Annual Conference of the European Association of
 997 Vertebrate Palaeontologists, 26th June – 1st July 2023. *Palaeovertebrata*, Special Volume 1-
 998 2023: 208. DOI:10.18563/pv.eavp2023

999 Redelstorff R. and Sander P. M. 2009. Long and girdle bone histology of *Stegosaurus*: implications
 1000 for growth and life history. *Journal of Vertebrate Paleontology*, 29, 1087– 1099.

1001 Redelstorff R., Sander P. M. and Galton P. M. 2014. Unique bone histology in partial large bone
 1002 shafts from Upper Triassic of Aust Cliff, England: An early independent experiment in
 1003 gigantism. *Acta Palaeontologica Polonica* 59, 607– 615.

1004 Reynolds S. H. 1946. The Aust section. *Cottswold Naturalists' Field Club, Proceedings*, 29, 29–
 1005 39.

1006 Ricqlès A. de, Padian K. and Horner J. R. 2003. On the bone histology of some Triassic
 1007 pseudosuchian archosaurs and related taxa. *Ann. de Paléont.*, 89, 67– 101.

1008 Ricqlès A. de, Buffrénil V. de, Laurin M. 2021. Archosauromorpha: from early diapsids to
 1009 archosaurs. 467– 483. In de Buffrénil V., de Ricqlès A. J., Zylberberg L., & Padian K. (Eds.).
 1010 (2021). *Vertebrate Skeletal Histology and Paleohistology* (1st ed.). Boca Raton: CRC Press.
 1011 838 pp. DOI 10.1201/9781351189590

1012 Richtberg & Girwidz 2023

1013 Sander P. M. 2000. Long bone histology of the Tendaguru sauropods: Implications for growth and
 1014 biology. *Paleobiology*, 26, 466– 488.

- 1015Sander P. M. 2013. An evolutionary cascade model for sauropod dinosaur gigantism - overview,
1016 update and tests. *PLoS ONE*, 8,e78573 (23 pages).
- 1017Sander P. M. and Andrassy P. 2006. Lines of arrested growth and long bone histology in
1018 Pleistocene large mammals from Germany: What do they tell us about dinosaur physiology?
1019 *Palaeontographica A*, 277, 143– 159.
- 1020Sander P. M. and Klein N. 2005. Developmental plasticity in the life history of a prosauropod
1021 dinosaur. *Science*, 310, 1800– 1802.
- 1022Sander P. M., Klein N., Stein K. and Wings O. 2011. Sauropod bone histology and implications for
1023 sauropod biology. 276– 302. In Klein N., Remes K., Gee C. T. and Sander P. M. (eds).
1024 *Biology of the Sauropod Dinosaurs. Understanding the Life of Giants*. Indiana University
1025 Press, Bloomington, 344 pp.
- 1026Sander P. M., Griebeler E. M., Klein N., Juarbe J. V., Wintrich T., Revell L. J. and Schmitz L.
1027 2021. Early giant reveals faster evolution of large body size in ichthyosaurs than in cetaceans.
1028 *Science*, 374:eabf5787 (15 pages).
- 1029Sander P. M., Romero Pérez De Villar P., Furrer H., and Wintrich T. 2022. Giant Late Triassic
1030 ichthyosaurs from the Kössen Formation of the Swiss Alps and their paleobiological
1031 implications. *Journal of Vertebrate Paleontology*, 42, e2046017.
- 1032Sander P. M., Wintrich T., Schwermann A. H. and Kindlimann R. 2016. Die paläontologische
1033 Grabung in der Rhät-Lias-Tongrube der Fa. Lücking bei Warburg-Bonenburg (Kr. Höxter) im
1034 Frühjahr 2015. *Geologie und Paläontologie in Westfalen* 88, 11– 37.
- 1035Sander P. M. and Wintrich T. 2021. Sauropterygia: Histology of Plesiosauria. 444– 455. In de
1036 Buffrénil V., de Ricqlès A.J., Zylberberg L., & Padian K. (Eds.). (2021). *Vertebrate Skeletal*
1037 *Histology and Paleohistology* (1st ed.). Boca Raton: CRC Press. 838 pp.
1038 <https://doi.org/10.1201/9781351189590>
- 1039Sanders W. 1876. On certain large bones in Rhaetic beds at Aust Cliff, near Bristol. *Annual Report*
1040 *of the Association for the Advancement of Science, Transactions of the Sections 1875*, 45, 88–
1041 81
- 1042Scheyer T. and Sander P. M. 2004. Histology of ankylosaur osteoderms: implications for
1043 systematics and function. *Journal of Vertebrate Paleontology*, 24, 874– 893.
- 1044Scheyer T. M., Syromyatnikova E. V. and Danilov I. G. 2017. Turtle shell bone and osteoderm
1045 histology of Mesozoic and Cenozoic stem-trionychian Adocidae and Nanhsiungchelyidae

- 1046 (Cryptodira: Adocusia) from Central Asia, Mongolia, and North America. *Fossil Record*, 20,
1047 69– 85.
- 1048Schobben M., Gravendyck J., Mangels F., Struck U., Bussert R., Kürschner W. M., Korn D., Sander
1049 P. M., and Aberhan M. 2019. A comparative study of total organic carbon- $\delta^{13}\text{C}$ signatures in
1050 the Triassic-Jurassic transitional beds of the central European basin and western Tethys shelf
1051 seas. *Newsletters on Stratigraphy*, 52, 461– 486.
- 1052Skedros J. G., Sorenson S. M. and Jenson N. H. 2007. Are distributions of secondary osteon
1053 variants useful for interpreting load history in mammalian bones? *Cells Tissues Organs*, 185,
1054 285– 307.
- 1055Stein K. and Sander P. M. 2009. Histological core drilling: a less destructive method for studying
1056 bone histology. In Brown M.A., Kane J.F., and Parker W.G., (Eds.). *Methods in fossil*
1057 *preparation. Proceedings of the First Annual Fossil Preparation and Collections Symposium*,
1058 pp. 69-80.
- 1059Stein K. and Prondvai E. 2014. Rethinking the nature of fibrolamellar bone: an integrative
1060 biological revision of sauropod plexiform bone formation. *Biological Reviews of the*
1061 *Cambridge Philosophical Society*, 89, 24– 47.
- 1062Storrs G. W. 1993. Terrestrial components of the Rhaetian (uppermost Triassic) Westbury
1063 Formation of southwest Britain. In Spencer G. Lucas and Michael Morales (eds.), *The*
1064 *Nonmarine Triassic, Transactions of the International Symposium and Field Trip on the*
1065 *Nonmarine Triassic, New Mexico Museum of Natural History & Science Bulletin*, 3, 447–
1066 451.
- 1067Storrs G. W. 1994. Fossil vertebrate faunas of the British Rhaetian (latest Triassic). *Zoological*
1068 *Journal of the Linnean Society of London*, 112, 217– 259.
- 1069Stutchbury S. 1850. On a large cylindrical bone found by Mr. Thompson in the “Bone-bed” of Aust
1070 Cliff, on the Severn. *Annual Report of the Association for the Advancement of Science*,
1071 *Transactions of the Sections*, 19, 67.
- 1072Sulej T., Niedźwiedzki G. 2019. An elephant-sized Late Triassic synapsid with erect limbs. *Science*,
1073 363, 78– 80.
- 1074Surmik D., Słowiak-Morkovina J., Szczygielski T., Wojtyniak M., Środek D., Dulski M., Balin K.,
1075 Krzykowski T. and Pawlicki R. 2023. The first record of fossilized soft parts in ossified

1076 tendons and implications for the understanding of tendon mineralization. *Zoological Journal*
 1077 *of the Linnean Society*, XX, 1–20.

1078 van Oers R.F.M., Wang H. and Bacabac R.G. 2015 Osteocyte Shape and Mechanical Loading. *Curr*
 1079 *Osteoporos Rep*, 13, 61–66. <https://doi.org/10.1007/s11914-015-0256-1>

1080 Vickaryous M. K., Meldrum G. and Russell A. P. 2015. Armored geckos: A histological
 1081 investigation of osteoderm development in *Tarentola* (Phyllodactylidae) and *Gekko*
 1082 (Gekkonidae) with comments on their regeneration and inferred function. *Journal of*
 1083 *Morphology*, 276, 1345–1357.

1084 Wintrich T., Hayashi S., Houssaye A., Nakajima Y. and Sander P. M. 2017. A Triassic
 1085 plesiosaurian skeleton and bone histology inform on evolution of a unique body plan.
 1086 *Sciences Advances*, 3, e1701144, 1–11.

1087 Woodward H. 2019. *Maiasaura* (dinosauria: Hadrosauridae) tibia osteohistology reveals non-
 1088 annual cortical vascular rings in young of the year. *Frontiers in Earth Science*, 7, 50. doi:
 1089 10.3389/feart.2019.00050

1090 Zylberberg L. 2021. Bone cells and organic matrix. 85–103. In de Buffrénil V., de Ricqlès A.J.,
 1091 Zylberberg L. and Padian K. (Eds.). (2021). *Vertebrate Skeletal Histology and Paleohistology*
 1092 (1st ed.). Boca Raton: CRC Press. 838 pp. <https://doi.org/10.1201/9781351189590>

Figure 1

Main histological features of the giant ichthyosaurs lower jaws

(A) BRSMG-Cg-2488 R 101 seen in cross-polarized light (left) and with a lambda filter added (right). The specimen shows a regular arrangement of rows of primary osteons with secondary osteons within, separated by thin periosteal GM (white arrows), and a high number of osteocyte lacunae. (B) Polarized light view of BRSMG-Cg-2488 R 101 showing the grid pattern of periosteal intrinsic fibers that characterizes the intrinsic fiber matrix (IFM). (C) BRSMG-Cg-2488 R 101 in circular polarized light revealing the seemingly helicoidal arrangement of the periosteal structural fibers and their interconnection within osteonal lamellar bone (top left). (D) Normal light view of the cross section of KULEuven PVL-1964 showing two primary osteons. The right one (dotted line) shows a secondary osteon within the primary one. (E) Longitudinal section of KULEuven PVL-1964 showing strands of unmineralized fibers (dark) running longitudinally in a herringbone pattern (green arrows) in normal light (left) and in polarized light with lambda filter (right). (F) KULEuven PVL-1964 in normal light showing the irregular shape of osteocyte lacunae and the unmineralized fibers (green arrows). *Abbreviations:* Lb, lamellar bone; Po, primary osteon, IFM, intrinsic fiber matrix; Rl, resorption line; Vc, vascular canal. Scale bars equal 100 μm (A, B, D, E) or 50 μm (C, F).

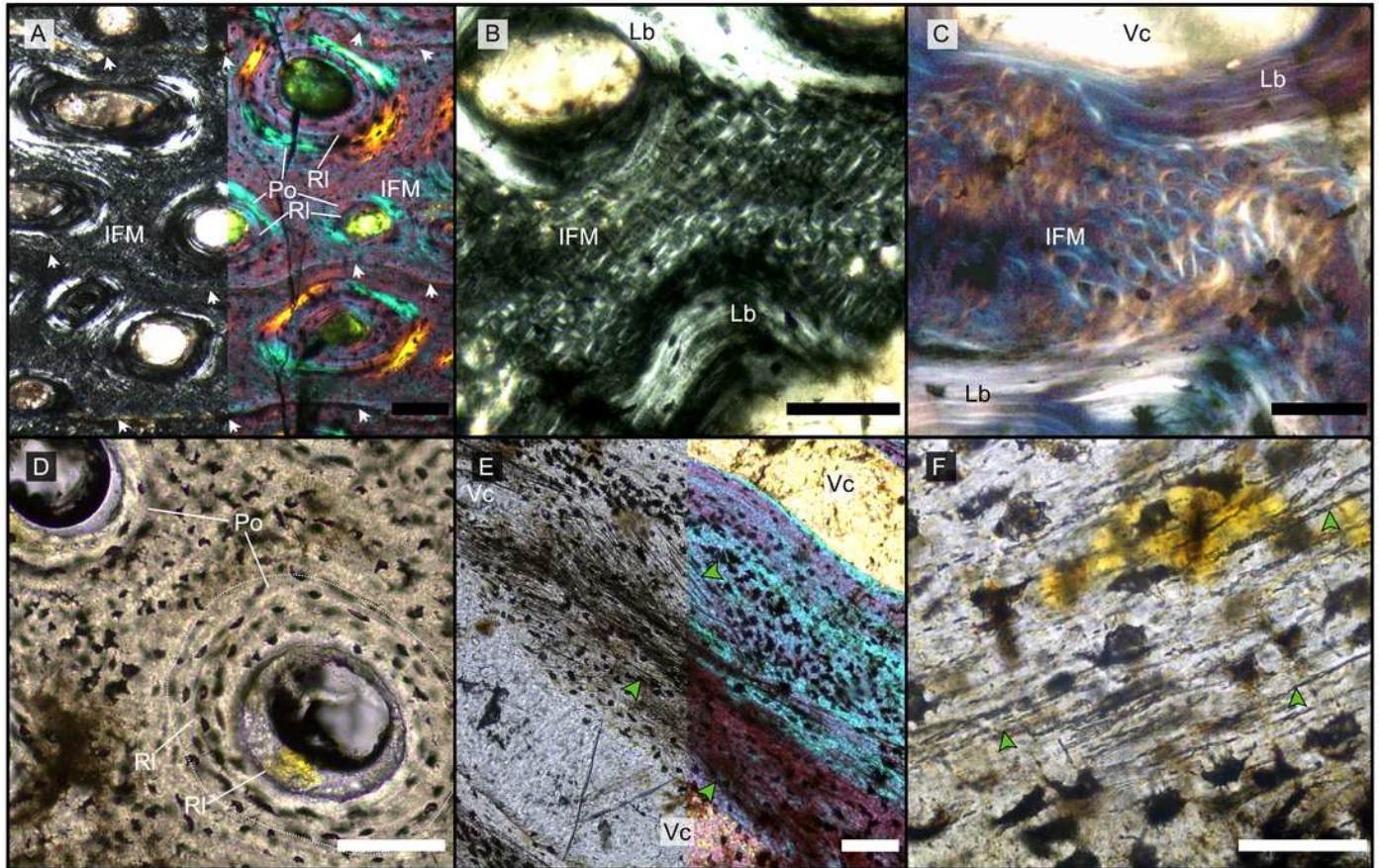


Figure 2

Overview of composite micrographs of selected thin sections

The resorption front is indicated by a blue hatched line, a black dotted line indicates the boundary between rDC and tDC. (A) BRSMG-Cb-3869, from Aust Cliff. (B) BRSMG-Cb-3870, from Aust Cliff. (C) BRSMG-Cg-2488 R 101, from Lilstock. (D) BRSMG-Cb-4063, from Aust Cliff. (E) KULEuven PVL-1964, from Cuers. Squares indicate the position of related figures (Figs. 4B, E, D). *Abbreviations:* DC, deep cortex; OC, outer cortex; rDC, regular deep cortex; RF, resorption front; tb, trabecular bone; tDC, template deep cortex. All scale bars equal 2 mm.

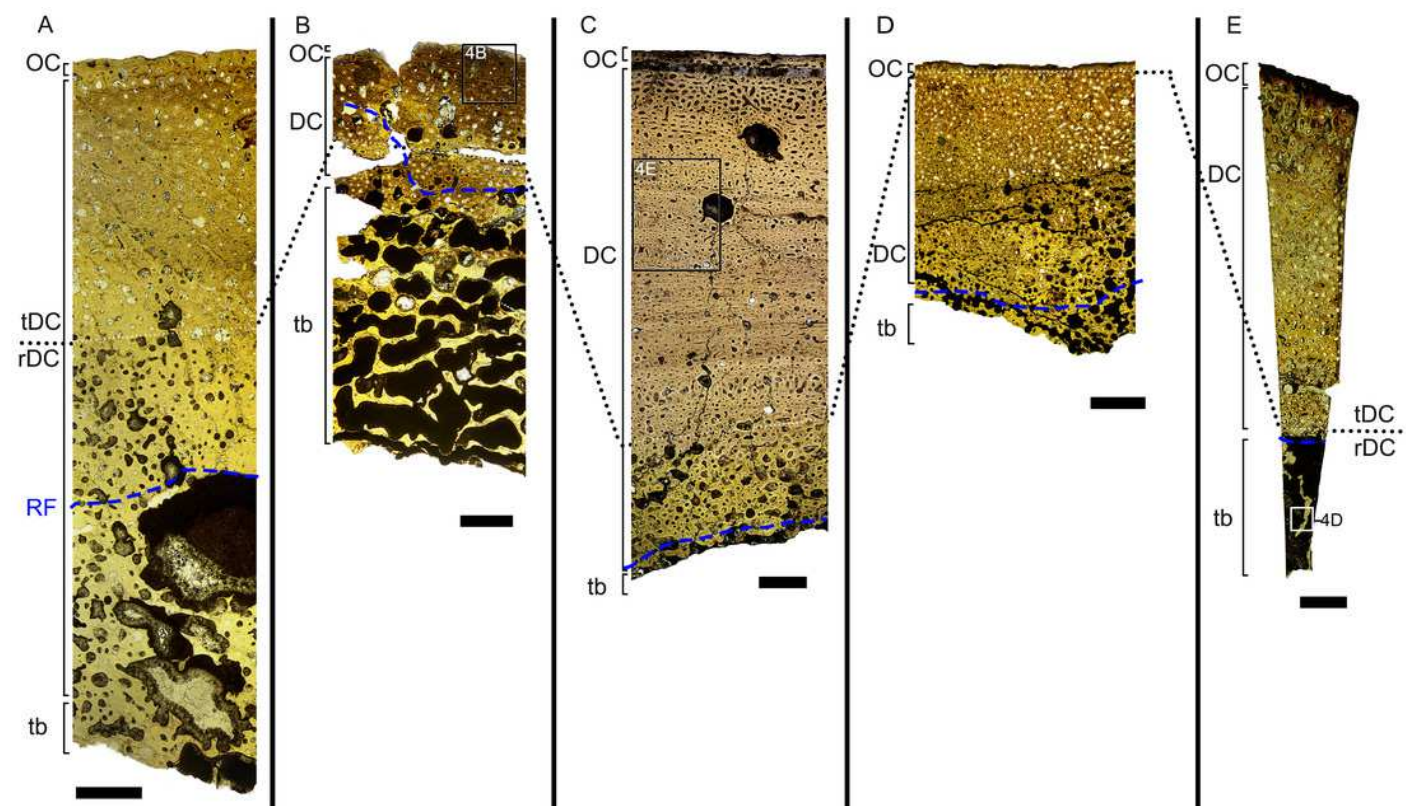


Figure 3

Features characterizing the areas identified as outer cortex, trabecular bone and deep cortex.

(A) Outer cortex of BRSMG-Cb-4063 in normal light (left) and in cross-polarized light with lambda filter added (right) showing primary tissue and growth marks (white arrows). Secondary osteons are present on the outer edge of the bone and may interrupt the continuity of the GM. The outer surface also shows diagenetic damage leading to the opening up of a secondary osteon. (B) BRSMG-Cb-3870 showing GM (white arrows) and a vascular canal open to the outer bone surface. (C) Longitudinal section of KULEuven PVL-1964 in cross-polarized light (left) and with a lambda filter added (right) revealing longitudinal vascularization. (D) Detail of trabecular bone of KULEuven PVL-1964 showing primary IFM and secondary lamellar bone in cross-polarized light. (E) BRSMG-Cg-2488 R 101 showing a template cortex characterized by parallel rows of primary and secondary osteons (white and purple narrow arrows) bordered by successive GM. Note the steep downturning of the rows in the vicinity of the nutrient canal. (F) Nutrient canal of BRSMG-Cb-3869 in normal light showing the presence of primary simple vascular canals and resorption cavities on the outer edge of the canal. *Abbreviations:* Lb, lamellar bone; NC, nutrient canal; oc, open vascular canal; IFM, intrinsic fiber matrix; so, secondary osteon. Scale bars equal 100 μm (A, C, D), 500 μm (B) or 2 mm (E, F).

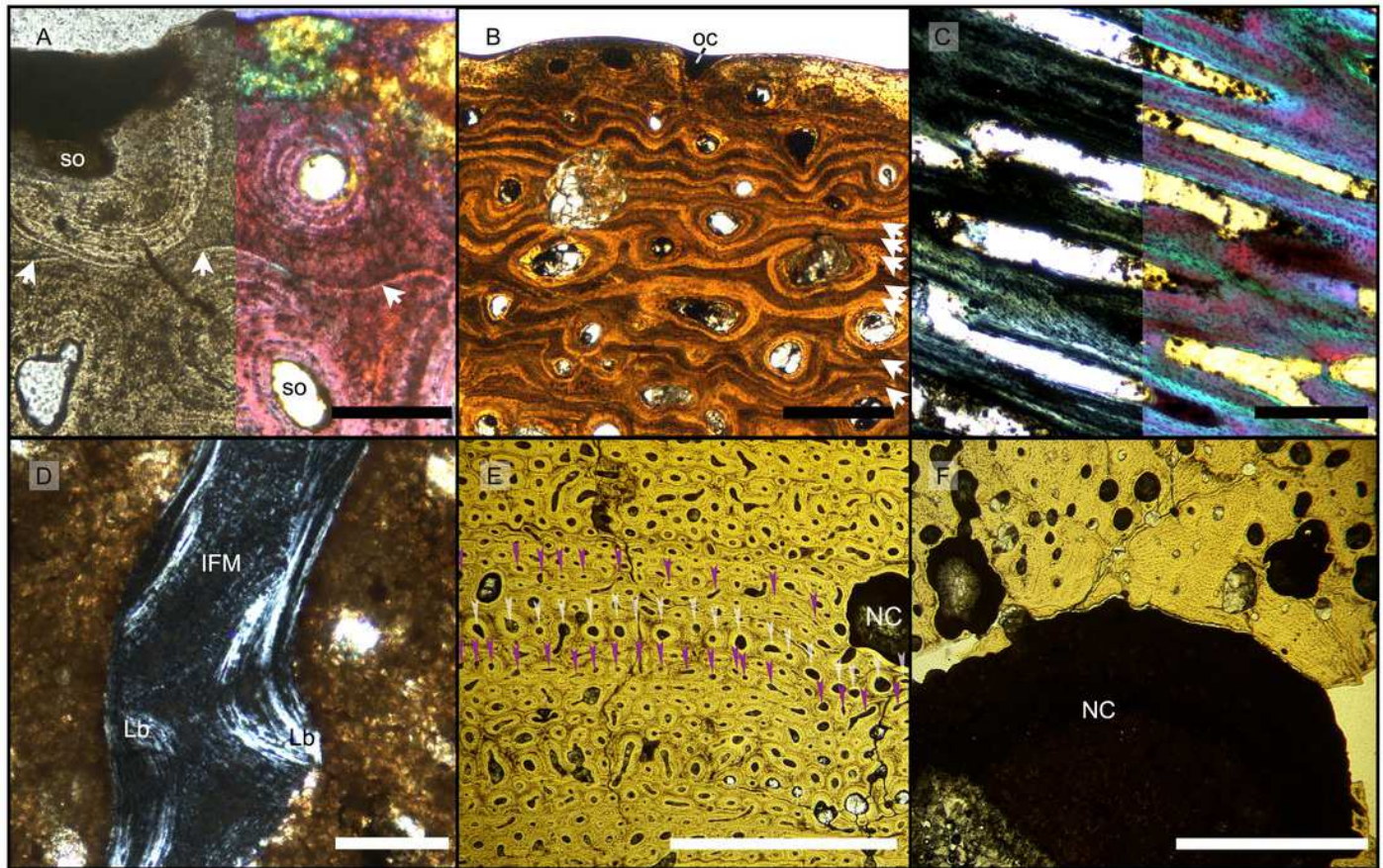


Figure 4

Overview of WMNM P88133, the largest cortical bone fragment WMNM P88133 from the late Middle Rhaetian of Bonenburg, Germany.

(A) Cross section showing a dark diagenetic seam staining the outer bone surface and the resorption front (blue dotted line). Note the low curvature of the outer bone surface and the great thickness of the cortex, suggesting that the fragment derives from a very large bone. (B) Overview of the external cortex showing the characteristic, strictly longitudinal vascular canals arranged in circumferential rows, vascular canals open to the outer bone surface (partially hidden by the dark seam), and the numerous secondary osteons inside the primary ones and the concentric secondary osteons. The obliteration of the multiple parallel rows of GM (white arrows) reveals the border between rDC and tDC (white dotted line). The tDC is characterized by essentially Haversian tissue. (C) Detail of the tDC, showing secondary osteons and IFM (left half of image cross-crossed polarized light, right half circular polarized light). The intrinsic fibers form parallel GM of alternating colors (white arrows). (D) Secondary osteon successively filled in by lamellar bone followed by woven or poorly mineralized bone (left side of image cross-polarized light with lambda filter, right side cross-polarized light only). (E) Longitudinal section seen in cross-polarized light with a lambda filter showing unmineralized fiber strands (dark, green arrows). *Abbreviations:* Cl, cementing line; HT Haversian tissue; IFM, intrinsic fiber matrix; Lb, lamellar bone; oc: open vascular canal; rDC, regular deep cortex; RF, resorption front; tDC, templating deep cortex; So: secondary osteon; Vc, vascular canal; Wb, woven bone. Scale bars: 2 cm (A), 1 mm (B), 100 μ m (C-E).

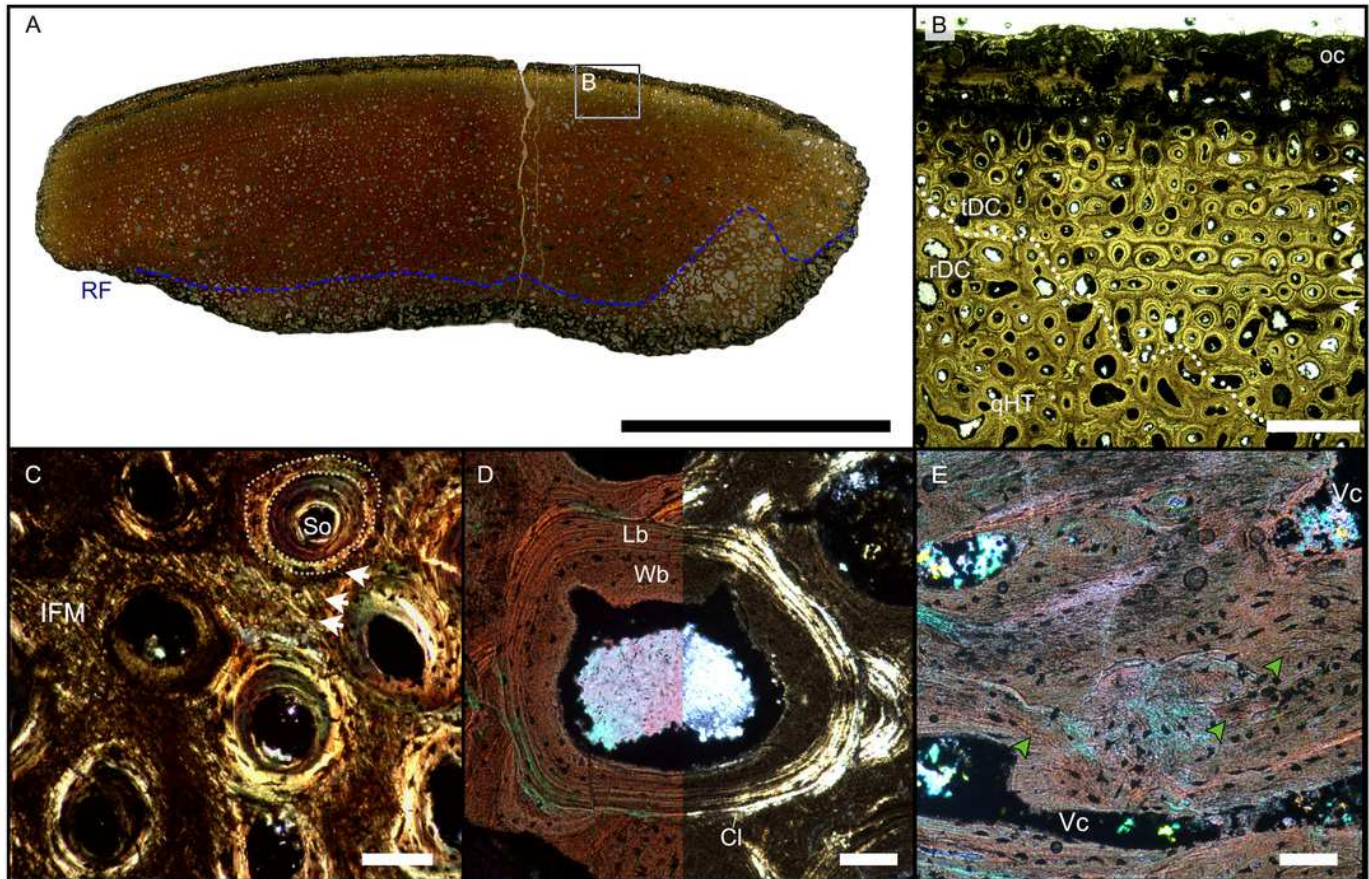


Figure 5

Histology of the sample from the splenial of the *Shastasaurus sikanniensis* type specimen RTMP-1994-378-0002 from the middle Norian of British Columbia, Canada.

(A) Cross section of the splenial section (dorsal at top), the highly cancellous structure is evident, as well as the lack of a dense outer cortex, caused by taphonomic processes. (B) Close-up view of area indicated in (A). Primary cortex with IFM is preserved interstitially between secondary trabeculae. Left half of the image is in cross-polarized light, right half in normal light. Note the dark stain of the bone tissue in the normal-light image. Post-mortem, pre-burial erosion of the bone surface is evident from the truncation of the bone structure and covered by opaque sediment. (C-D) Close-up showing IFM in cross-polarized light (C) and in circular polarized light (D). Note the helical arrangement of the fibers around a dark core. *Abbreviations:* IFM, intrinsic fiber matrix; RC, resorption cavity. Scale bars represent: 5 mm (A), 100 μ m (B), 50 μ m (C, D).

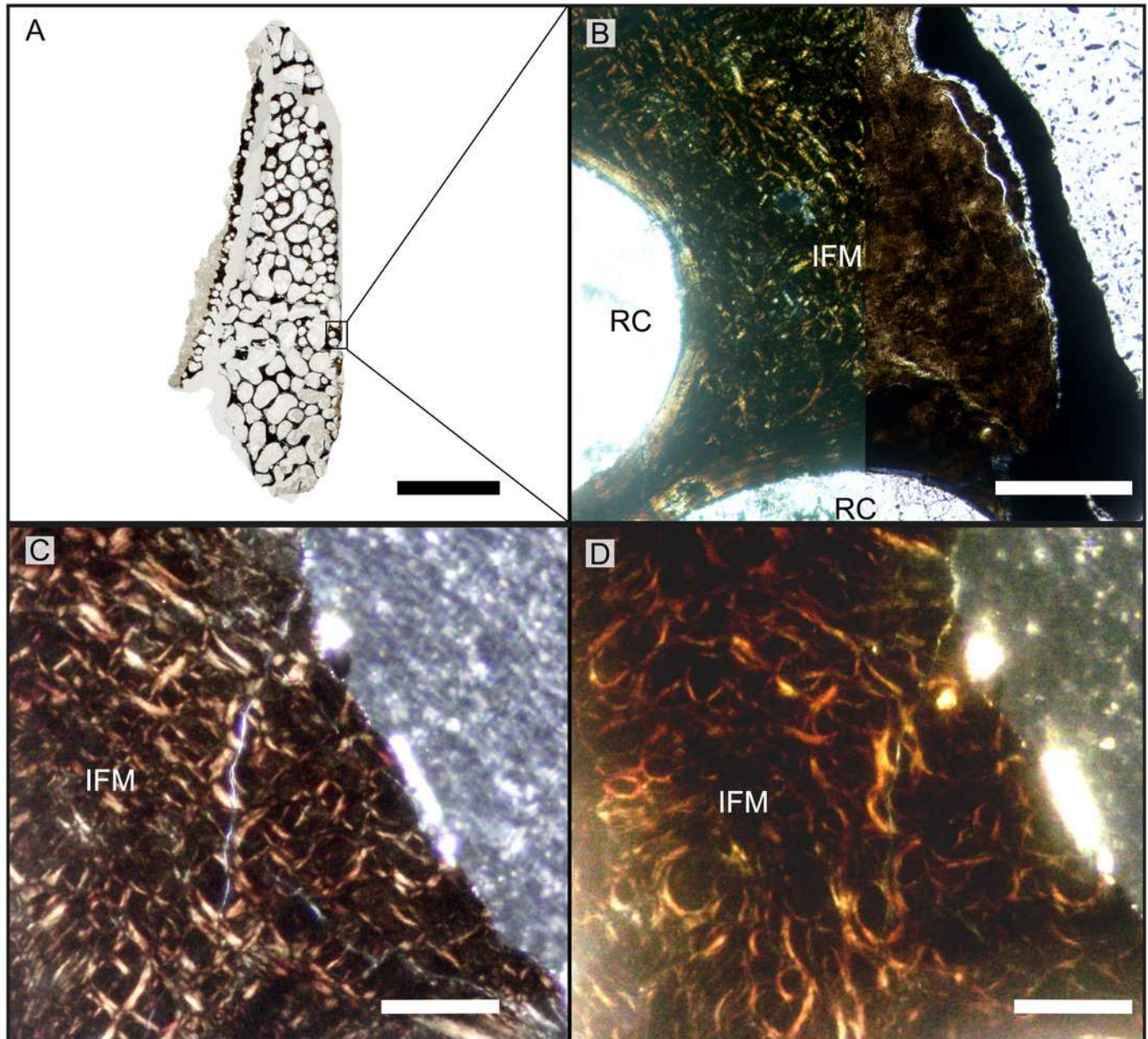


Table 1(on next page)

List of specimens used in this study.

1

Spec. No.	Locality	Age	Strat. Unit	Anatomy	Taxon	Reference	Samples	Sampling method	Plane of section	Thin section repository	Remarks
RTMP-1994-378-0002	Sikanni Chief River, British Columbia, Lillooet, UK	middle Norian Rhaetian	Pardonet Formation	surangular, splenial	<i>S. sikanniensis</i>	Nicholls & Manabe 2004	2	cut	cross	IGPB	holotype
BRSMG-Cg-2488 R 101			Top of Westbury Mudstone Formation	surangular	Shastasauridae indet.	Lomax et al. 2018	1	core	cross	BRSMG	
BRSMG-Cb-3869, 3870, 4063	Aust Cliff, UK	Rhaetian	Rhaetic bonebed at base of Westbury Mudstone	surangular	Shastasauridae indet.	Galton 2005; Redelstorff et al. 2014; Lomax et al. 2018	3	core	cross	BRSMG	
KULeuven PVL-1964	Autun, France	Rhaetian	Grès à Avicula contorta, Grès Blonds Formation	surangular	Shastasauridae indet.	Fischer et al. 2014, Lomax et al. 2018	2	core, cut	cross and long	IGPB	
WMNM P-uncatalogued	Bonenburg, Germany	late middle Rhaetian	Exter Formation	cortical fragment	Tetrapoda indet.	Sander et al. 2016	1	cut	cross	IGPB	
WMNM P88130,...,P88144	Bonenburg, Germany	late middle Rhaetian	Exter Formation	15 cortical fragments	Tetrapoda indet.	Sander et al. 2016	14	cut	cross and long	IGPB	

Table 2 (on next page)

Comparison of the osteohistological features across our study sample and other Late Triassic taxa from the literature.

The results here summarized are based on literature research and, when available, on authors observation of the samples in the GIPB histology collection. *Abbreviations*: CPF, coarse parallel fibered bone; GM, growth marks; IFM, intrinsic fiber matrix; ISFs, interwoven structural fibers; LAGs, Lines of arrested growth; WPC, woven-parallel complex; PIFT, periosteal intrinsic fiber tissue.

Groups considered in this study	Source of histological samples	Main bone organization	Vascularization rate	Vascular organization	Cyclical structures	Periosteal remodeling strategy	Relative remodeling rate	Abundant concentric osteons	Main references
Lilstock ichthyosaur	Lower jaws	WPC with IFM (PIFT)	High	Longitudinal	GM	Template + diffused	High	Yes	This study
Autun ichthyosaur	Lower jaws	WPC with IFM (PIFT)	High	Longitudinal	GM	Template + diffused	High	Yes	This study
Aust Cliff bone segments	Lower jaws(?)	WPC with IFM (PIFT)	High	Longitudinal	GM	Template + diffused	High	Yes	Redelstorff, Sander & Galton (2014), this study
<i>S. sikanniensis</i>	Splenal and surangular	WPC with IFM (PIFT)	High	Longitudinal	Not preserved	Not preserved	Not preserved	Not preserved	This study
Bonenburg cortical fragments	Unidentified cortices	WPC with IFM (PIFT)	High	Longitudinal	GM	Template + diffused	High	Yes	This study
Sauropodomorpha	Long bones	WPC (fibrolamellar)	Moderate to high	Plexiform/laminar	LAGs	Organized front	Moderate to high	Not observed or reported	Klein & Sander (2007); Mitchell & Sander (2014)
Stegosauria	Long bones	WPC	Moderate	Longitudinal	LAGs	Scattered front	Moderate to high	Not reported	Redelstorff & Sander (2009); Padian & Woodward (2021)
Rauisuchia - Slow growth	Long bones	Lamellar-Zonal + WPC	Low	Laminar/subplexiform	Annuli+LAGs	Scattered	Low	Not reported	de Ricqlès <i>et al.</i> (2003); de Buffrénil <i>et al.</i> (2021)
Rauisuchia - Fast growth	Long bones	WPC	Moderate to high	Laminar/subplexiform	Annuli	Scattered	Low	Not reported	Klein et al. (2017); de Buffrénil et al. (2021)
Phytosauria	Long bones	Lamellar-Zonal	Low	Longitudinal	Annuli+LAGs	Scattered front	Low	Not reported	de Ricqlès, Padian & Horner (2003); ; Ricqlès, Buffrénil & Laurin (2021)
Dicynodontia	Long bones	WPC	Moderate to high	Longitudinal	GM	Scattered and unorganized		Not reported	Chinsamy & Rubidge (1993); Green, Schweitzer & Lamm (2010)
Plesiosauria	Long bones	WPC	Moderate to high	Longitudinal+radial	GM	Template + front	High	Yes (?)	Wintrich <i>et al.</i> (2017); Sander & Wintrich (2021)
large Nothosauria	Ribs	WPC+CPF	Moderate to high	Longitudinal+radial	LAGs	Absence	Low	Not reported	Klein, Canoville & Houssaye. (2019)
Temnospondyli	Lower jaws, long bones	Lamellar-Zonal+HSFs	Low to moderate	Longitudinal+plexiform	Annuli+LAGs	Template	Low to moderate	Not reported	Konietzko-Meier <i>et al.</i> (2018); Gruntmejer, Bodzioch & Konietzko-Meier (2021)