

# New insights into the role of GSK-3 $\beta$ in the brain: From neurodegenerative disease to tumorigenesis

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Glycogen synthase kinase 3 (GSK-3) is a serine/threonine kinase widely expressed in various tissues and organs. Unlike other kinases, GSK-3 is active under resting conditions and is inactivated upon stimulation. In mammals, GSK-3 includes GSK-3  $\alpha$  and GSK-3 $\beta$  isoforms encoded by two homologous genes, namely, GSK3A and GSK3B. GSK-3 $\beta$  is essential for the control of glucose metabolism, signal transduction, and tissue homeostasis. As more than 100 known proteins have been identified as GSK-3 $\beta$  substrates, it is sometimes referred to as a moonlighting kinase. Previous studies have elucidated the regulation modes of GSK-3 $\beta$ . GSK-3 $\beta$  is involved in almost all aspects of brain functions, such as neuronal morphology, synapse formation, neuroinflammation, and neurological disorders. Recently, several comparatively specific small molecules have facilitated the chemical manipulation of this enzyme within cellular systems, leading to the discovery of novel inhibitors for GSK-3 $\beta$ . Despite these advancements, the therapeutic significance of GSK-3 $\beta$  as a drug target is still complicated by uncertainties surrounding the potential of inhibitors to stimulate tumorigenesis. This review provides a comprehensive overview of the intricate mechanisms of this enzyme and evaluates the existing evidence regarding the therapeutic potential of GSK-3 $\beta$  in brain diseases, including Alzheimer's disease, Parkinson's disease, mood disorders, and glioblastoma.

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# Abstract

Glycogen synthase kinase 3 (GSK-3) is a serine/threonine kinase widely expressed in various tissues and organs. Unlike other kinases, GSK-3 is active under resting conditions and is inactivated upon stimulation. In mammals, GSK-3 includes GSK-3 $\alpha$  and GSK-3 $\beta$  isoforms encoded by two homologous genes, namely, GSK3A and GSK3B. GSK-3 $\beta$  is essential for the control of glucose metabolism, signal transduction, and tissue homeostasis. As more than 100 known proteins have been identified as GSK-3 $\beta$  substrates, it is sometimes referred to as a moonlighting kinase. Previous studies have elucidated the regulation modes of GSK-3 $\beta$ . GSK-3 $\beta$  is involved in almost all aspects of brain functions, such as neuronal morphology, synapse formation, neuroinflammation, and neurological disorders. Recently, several comparatively specific small molecules have facilitated the chemical manipulation of this enzyme within cellular systems, leading to the discovery of novel inhibitors for GSK-3 $\beta$ . Despite these advancements, the therapeutic significance of GSK-3 $\beta$  as a drug target is still complicated by uncertainties surrounding the potential of inhibitors to stimulate tumorigenesis. This review provides a comprehensive overview of the intricate mechanisms of this enzyme and evaluates the existing evidence regarding the therapeutic potential of GSK-3 $\beta$  in brain diseases, including Alzheimer's disease, Parkinson's disease, mood disorders, and glioblastoma.

**Keywords:** Alzheimer's disease; Glioblastoma; GSK-3 $\beta$ ; Neuroinflammation; Parkinson's disease; Synaptic plasticity

# Introduction

GSK-3 is an extensively conserved serine/threonine (S/T) protein kinase that catalyses the phosphorylation of threonine or serine residues, regulating numerous cellular biological processes, including glycogen metabolism, insulin signalling, cell growth, and differentiation (Duda et al., 2020). Inhibition of glycogen synthase (GS) by GSK-3 results in reduced glycogen synthesis in the liver and muscles, accompanied by elevated blood glucose levels or hyperglycaemia (Gupte et al., 2020). For this reason, GSK-3 is closely correlated with the pathogenesis and progression of many diseases, such as diabetes, obesity, and cancer (Amar et al., 2011). GSK-3 is constitutively activated in resting cells and is frequently inhibited by growth factors [e.g., insulin, insulin-like growth factor (IGF), vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), and nerve growth factor (NGF)] and other extracellular stimuli (e.g., Wnt) (Duda et al., 2018). In humans, there are two isoforms of GSK-3, namely, GSK-3 $\alpha$  (51 kDa) and GSK-3 $\beta$  (47 kDa), which are encoded by distinct genes, GSK3A and GSK3B, respectively. The catalytic domains of GSK-3 $\alpha$  and GSK-3 $\beta$  proteins exhibit a remarkable 98% similarity, while their unique C-terminal regions diverge, showing only 36% homology (Emma et al., 2020). Although they have higher homology in the kinase domain, GSK-3 $\alpha$  and GSK-3 $\beta$  have differentiated substrate preferences, and their cellular functions are at least partially nonredundant, as demonstrated by isoform-specific gene knockout (KO) studies in mice (Gupte et al., 2022). GSK-3 $\alpha$  KO mice exhibit viability and normal developmental patterns, whereas GSK-3 $\beta$  KO leads to embryonic lethality, primarily attributed to severe liver degeneration (Hoeftlich et al., 2000). The activity of the GSK-3 $\beta$  kinase depends on phosphorylation. Phosphorylation of tyrosine 216 (T216) in GSK-3 $\beta$  and tyrosine 279 (T279) in GSK-3 $\alpha$  is needed for maximal activity, whereas phosphorylation of serine 9 (S9) in GSK-3 $\beta$  and serine 21 (S21) in GSK-3 $\alpha$  results in inhibition of the kinase (Moore et al., 2021). For example, insulin inactivates GSK-3 by phosphorylation of specific residues, namely, serine 21 (p-Ser21-GSK-3 $\alpha$ ) and serine 9 (p-Ser9-GSK-3 $\beta$ ),

in a phosphatidylinositol 3-kinase (PI3K)-dependent manner (Zakharova et al., 2019). Among the many kinases that phosphorylate GSK-3 $\beta$  and thus inhibit its activity, the best known is protein kinase B (PKB or AKT), which acts downstream of PI3K and PDK1 kinase signalling. The inhibition of GSK-3 $\beta$  through phosphorylation is not exclusive to AKT but also involves other protein kinases, including protein kinase A (PKA) and protein kinase C (PKC) (Ku et al., 2011; Moore et al., 2013). In addition, GSK-3 $\beta$  regulation is influenced by various other regulators, including the mammalian target of rapamycin (mTOR), Wnt, and p38 mitogen-activated protein kinase (p38 MAPK) signalling pathways. These pathways also hold significant importance in the regulation of GSK-3 $\beta$  (Golick et al., 2018; Pan and Valapala, 2022; Taelman et al., 2010; Thornton et al., 2008). Despite this inhibitory phosphorylation, dephosphorylation of the Ser9 phosphate group on GSK-3 $\beta$  by protein phosphatases, such as protein phosphatase 2A (PP2A) and protein phosphatase 1 (PP1), enhances GSK-3 $\beta$  activity. This phenomenon has been observed in triple-negative breast cancer and neuronal degeneration (Bennecib et al., 2000; Jian et al., 2022; Liang and Chuang, 2007). Conversely, these regulators and protein phosphatases may also function as substrates for GSK-3 $\beta$  (Hermida et al., 2017).

Over the past decades, dysregulation of GSK-3 $\beta$  has been linked to the pathogenesis of many disorders, including type 2 diabetes mellitus (T2DM), atherosclerosis, neurodegenerative diseases, and a variety of malignant tumours (Lin et al., 2020; Yang et al., 2021; Zhang et al., 2018). Because aberrant GSK-3 $\beta$  expression has been implicated in many diseases, accumulating studies have provided proof-of-concept that targeting GSK-3 $\beta$  is a promising strategy for treating various diseases. In tumours, GSK-3 $\beta$  has been demonstrated to play a paradoxical role as GSK-3 $\beta$  acts as a tumour promoter or suppressor based on the cell type and phosphorylation status (He et al., 2022; Li et al., 2017; Pecoraro et al., 2021; Zhang et al., 2020). Furthermore, increasing evidence suggests that GSK-3 $\beta$  signalling plays a critical role in driving the advancement of neurodegeneration. Therefore, inhibition of GSK-3 $\beta$  is regarded as a prospective therapeutic strategy for central nervous system (CNS)-related disorders, especially Alzheimer's disease (AD) (Eldar-Finkelman and Martinez, 2011; Lauretti et al., 2020).

In the next section, we summarize the regulatory mechanisms of GSK-3 $\beta$  action, the physiopathological functions of GSK-3 $\beta$  in the brain, the current development of GSK-3 $\beta$  inhibitors, and treatment of brain and neurological diseases. The major focus of this review is on the intricate interactions between GSK-3 $\beta$  and neuroregulation, as well as the potential of GSK-3 $\beta$  for the therapeutic intervention of brain disease.

# **Why this review is needed and who it is intended for**

The brain is the main controller of learning, memory, movement, and other behaviours. Despite intensive research, the exact mechanisms that trigger brain disorders, such as Alzheimer's disease (AD) and Parkinson's disease (PD), are still not known, and at present, there is no cure for many brain diseases. Recently, several signalling molecules, especially glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ), have been explored as possible candidate targets for the treatment of AD, PD, and mood disorders. This review discusses and summarizes the new advances in GSK-3 $\beta$  and its roles in brain physiology, neural development, neurodegenerative disorders, and neuropsychiatric disorders. Our review will appeal to researchers interested in the field of neuroscience, and it provides an overview of research on molecular targeted therapy for neurological and brain diseases. Although research on GSK-3 $\beta$  in the brain is still faced with many shortcomings and obstacles, the reported results in drug treatment, especially in GSK-3 $\beta$  inhibitors, are surprising. Importantly, these basic concepts and comprehensive knowledge are of great reference for chemists, biological scientists, pharmacists, and clinical workers as they may provide new insights into brain science and even a breakthrough in other fields.

# Search Methodology

PubMed and Web of Science were used to identify relevant articles for this review, with the most recent search conducted in 2023. The search was performed in full-text journals, focusing on the most relevant advances in GSK-3 $\beta$  functions and their role in brain physiology and pathology. We categorized keywords, synonyms, and variants into categories, and used any combination of words from those categories for our search. The following categories were used for the search: 1) GSK-3 $\beta$ , which included GSK3 $\beta$ , glycogen synthase kinase-3 $\beta$ , and glycogen synthase kinase 3; and 2) brain disease, which included central nervous system, neural, cerebral, nervous, mind, Alzheimer's disease, Parkinson's disease, Huntington's disease, amyotrophic lateral sclerosis, multiple sclerosis, bipolar disorder, schizophrenia, glioblastoma, and ischaemia–reperfusion (I/R) injury. The words were merged via the Boolean operators “AND” and “OR”. The initial search screened approximately 200 relevant articles written in English that could be useful for this review. No language restrictions were imposed.

# Regulatory Mechanisms of GSK-3 $\beta$ Action

GSK-3 $\beta$  is involved in the regulation of cell biological functions through specific substrate phosphorylation. The regulation of GSK-3 $\beta$  activity encompasses the following four pivotal mechanisms: autophosphorylation regulation of GSK-3 $\beta$ , subcellular localization of GSK-3 $\beta$ , assembly of protein complexes harbouring GSK-3 $\beta$ , and phosphorylation status of GSK-3 $\beta$  substrates (Jope and Johnson, 2004) (Figure 1). One of the clearest regulatory mechanisms is the inhibition of GSK-3 $\beta$  activity by phosphorylation of Ser9 in GSK-3 $\beta$  (corresponding to Ser21 in GSK-3 $\alpha$ ) (Patel and Werstuck, 2021). The PI3K/AKT signalling pathway is the primary regulator of GSK-3 $\beta$ , which is activated in response to insulin and several growth factors. However, AKT (PKB) and other kinases, including protein kinase A (PKA), ribosomal 70-kDa protein S6 kinase (p70S6K), and p90 ribosomal S6 kinase (p90RSK), phosphorylate the inhibitory Ser9 residue of GSK-3 $\beta$ , which results in the inactivation of GSK-3 $\beta$  (Cervello et al., 2017). In contrast, phosphorylation of Tyr216 in GSK-3 $\beta$  (corresponding to Tyr279 in GSK-3 $\alpha$ ) by Src, FYN, and PYK2 enhances the catalytic activity of GSK-3 $\beta$  kinase, which may be constitutive in resting cells, but the mechanism regulating this modification is unclear (Bhat et al., 2000; Nagini et al., 2019).

The activity of GSK-3 $\beta$  is subject to regulation through subcellular localization, particularly within the nucleus and mitochondria, where GSK-3 $\beta$  exhibits heightened activity and undergoes dynamic regulation (Bijur and Jope, 2003). When translocated into the nucleus, GSK-3 $\beta$  exhibits phosphorylation activity towards a diverse range of substrates, mostly known as transcription factors (TFs) or epigenetic regulatory factors, such as RXR $\alpha$ , p53, and KDM1A (Eom and Jope, 2009; Zhang et al., 2020; Zhou et al., 2016). The localization of GSK-3 $\beta$  within protein complexes either facilitates or impedes its activity towards specific substrates. For example, in the canonical Wnt signalling pathway, GSK-3 $\beta$  and the  $\beta$ -catenin transcriptional coactivator coexist on the Axin scaffold protein, and this colocalization guides GSK-3 $\beta$  to phosphorylate  $\beta$ -catenin, consequently triggering the degradation of  $\beta$ -catenin protein in the cytosol. Activation of the Wnt signalling pathway prevents GSK-3 $\beta$  from accessing  $\beta$ -catenin, resulting in the accumulation of active  $\beta$ -catenin (Marineau et al., 2020). The fourth mechanism refers to the effect of GSK-3 $\beta$  being regulated by the substrate phosphorylation state, which is a mechanism that indirectly regulates the substrate phosphorylation efficiency of GSK-3 $\beta$ . Most of the substrates of GSK-3 $\beta$  must be in the “primed” (sensitized) state. Under the condition that GSK-3 $\beta$  is prephosphorylated at a residue four amino acids C-terminal to the phosphorylation site by another “priming kinase” (activated kinase), GSK-3 $\beta$  further phosphorylates the “primed” substrate; that is, the regulation of substrate phosphorylation

activity by GSK-3 $\beta$  usually requires the coordination of the "priming kinase" (Duan et al., 2022; Jope et al., 2007). These complex mechanisms of GSK-3 $\beta$  regulation provide GSK-3 $\beta$  with specific control over its substrates, a particularly important ability of the enzyme that enables GSK-3 $\beta$  to phosphorylate numerous substrates and thus regulate many biological functions.

## Multifaceted Functions of GSK-3 $\beta$ in the Brain

GSK-3 $\beta$  is the most abundant in the brain and nervous system, and its expression level increases with age (Lee et al., 2006). GSK-3 $\beta$  not only mediates abnormal pathological processes but is also essential for the normal physiological function of the brain. To obtain a comprehensive understanding, we summarized the pleiotropic role of GSK-3 $\beta$  in the brain in Figure 2.

### GSK-3 $\beta$ , synaptic plasticity, and memory formation

The hippocampus is widely recognized as the central hub for learning and memory within the brain, and it plays a crucial role in synaptic plasticity, encompassing the mechanisms underlying both long-term potentiation (LTP) and long-term depression (LTD). LTP refers to the enduring enhancement of synaptic strength, while LTD represents the contrasting process (Parvez et al., 2023). GSK-3 $\beta$  exhibits high expression levels in the hippocampus of a healthy brain where it assumes a crucial function in synaptic plasticity and the formation of memories. Liu E et al. reported that mice with GSK-3 $\beta$  deletion specifically in the excitatory neurons of the dentate gyrus (DG) exhibit impairments in spatial and fear memory. Subsequent investigations have revealed that deletion of GSK-3 $\beta$  in the DG subset leads to the inhibition of synaptic transmission within the hippocampus; additionally, it results in decreased expression levels of GluN1, GluN2A, GluN2B, GluA1, PSD93, drebrin, and synaptophysin (Liu et al., 2017). Consistently, Koike R et al. demonstrated that aged GSK-3 $\beta^{+/-}$  mice manifest deficiencies in the formation of both short-term and long-term memories, thereby offering additional evidence for the essential role of GSK-3 $\beta$  in memory formation during advanced age (Koike et al., 2021). Mechanistically, Peineau S et al. reported that the activation of GSK-3 $\beta$  is enhanced during NMDA receptor-dependent LTD via activation of PP1, while the activity of GSK-3 $\beta$  is inhibited by the induction of LTP (Peineau et al., 2007). The induction of LTP in both the DG and CA1 regions of the hippocampus leads to an elevation in the inhibitory phosphorylation of GSK-3 $\beta$  at Ser9 (Cunningham et al., 2017).

Memory impairment in old age is a hallmark of AD, with dementia developing in the final stages. Alzheimer's disease is marked by  $\beta$ -amyloid (A $\beta$ ) deposits and neurofibrillary tangles (NFTs). Detectable NFTs in AD-associated memory impairment and dementia are associated with synaptic and neuronal loss in diseased brains (Ashrafian et al., 2021). Prior to NFT formation, tau undergoes hyperphosphorylation due to the activation of GSK-3 $\beta$ , resulting in the formation of tau oligomers with a granular structure. The hyperphosphorylated tau is strongly linked to the loss of synapses and the impairment of memory induced by A $\beta$ . Mice overexpressing GSK-3 $\beta$  show an accumulation of hyperphosphorylated tau, a reduction in hippocampal LTP, and memory impairment in object recognition tests (Takashima, 2012). Notably, this memory deficit in mice is reversed when tau expression stops. Consequently, reducing tau levels and inhibiting GSK-3 $\beta$  each rescue memory impairment in AD models (Gómez de Barreda et al., 2010; Roberson et al., 2007). Another possible mechanism is that GSK-3 $\beta$  inactivation enables stabilization of  $\beta$ -catenin, which protects  $\beta$ -catenin from degradation mediated by the proteasome.  $\beta$ -catenin is expressed in both pre- and postsynaptic terminals, and it is responsible for cell adhesion and synaptic structure (Hui et al., 2018; Murase et al., 2002). Downregulation of the  $\beta$ -catenin signalling pathway

due to increased GSK-3 $\beta$  activity has been observed in AD brains (Vallée et al., 2018). Given the pivotal role of GSK-3 $\beta$  in the regulation of synaptic plasticity and memory formation, the reduction in GSK-3 $\beta$  activity obtained by pharmacological approaches has been extensively explored and reported in several AD models (Iqbal et al., 2023).

### **GSK-3 $\beta$ and neuroinflammation**

Inflammation has become a dominant contributor to brain disease, with principal participation from ageing and proteinopathy, which has been widely studied to understand its mechanism in neural pathology (Samim Khan et al., 2023). The initial defensive reaction of the organism in response to allergic stimuli involves immune-mediated inflammation. Although the initial response is protective, the persistent and excessive response causes pathological lesions. In the neural network, the equilibrium between inflammatory and anti-inflammatory processes is upheld through the activation of M1 and M2 microglia. M1 microglia exhibit the capacity to generate inflammatory factors primarily mediated by Toll-like receptors (TLRs), whereas M2 microglia function as reparative agents, primarily releasing IL-10 and IL-4 (Cherry et al., 2014). As a result of pathological stimuli within the brain, the innate immune response is initially engaged, leading to the subsequent transformation of microglia-mediated innate immunity into neuroinflammation. Neuroinflammation is associated with glial activation, blood–brain barrier (BBB) breakdown, proinflammatory cytokine release, and leukocyte invasion, in which the molecular mechanism generally involves the activity of NOD-like receptor family pyrin domain-containing protein 3 (NLRP3), Toll-like receptors (TLRs), and nuclear transcription factors, such as nuclear factor kappa B (NF- $\kappa$ B), nuclear factor of activated T cells (NFAT), cAMP response element-binding protein (CREB), and signal transducer and activator of transcription 3 (STAT3) (Candelario-Jalil et al., 2022; Jurcau and Simion, 2021; Zhu et al., 2021).

NLRP3 is a cytoplasmic pattern recognition receptor (PRR) that is prominently expressed in macrophages, and it serves as a crucial constituent of the inflammasome. In the past decade, NLRP3 has been shown to be involved in neuroinflammatory processes, in which the interplay between the innate immune system and the caspase-1 apoptotic protein plays a significant role (Voet et al., 2019). The assembly of the NLRP3 inflammasome results in the secretion of potent proinflammatory cytokines, namely, IL-1 $\beta$ , IL-18, and IL-33. These inflammatory factors play a crucial role in neurodegeneration. After the discovery of the regulatory role of GSK-3 $\beta$  in the brain, its participation in NLRP3-mediated inflammatory pathways has been widely explored. Agents capable of inducing the phosphorylation of GSK-3 $\beta$  to mitigate NLRP3-mediated inflammation, oxidative stress, apoptosis, and autophagy have been assessed in various models of neurological disorders (Li et al., 2021; Liu et al., 2020b; Wang et al., 2021; Wang et al., 2019). In the brain, Toll-like receptors (TLRs) mainly modulate glial and neuronal functions, as well as innate immunity and neuroinflammation, under physiological or pathological conditions (Fei et al., 2022; Mowry et al., 2021; Schilling et al., 2021). During the past decade, numerous studies have demonstrated that GSK-3 $\beta$  is a key regulator of the TLR signalling pathway via TLR3 and TLR4 (Ko and Lee, 2016). TNFR-associated factors (TRAFs) are critical for the production of inflammatory cytokines and antiviral responses in TLR3-mediated signalling pathways. Ko R et al. identified TRAF6 as a direct E3 ligase for GSK-3 $\beta$ , and they demonstrated that TRAF6-mediated GSK-3 $\beta$  ubiquitination is essential for cytokine production by promoting complexes assembled by TRIF, a TLR3 adaptor protein (Ko et al., 2015). These researchers also reported that Src phosphorylation is regulated by GSK-3 $\beta$  via TNFR-associated factor 2 (TRAF2)-mediated Src ubiquitination. Deficits of GSK-3 $\beta$  in mouse embryonic fibroblasts significantly reduce

IFN-stimulated gene expression due to a reduction in Src tyrosine phosphorylation (Ko et al., 2019). Compared to the wild type (WT) group, TLR3 deficiency significantly inhibits programmed necrosis of brain cells in neonatal mice (Zhang et al., 2022a). TLR3 serves as a vital modulator of neuronal survival and developmental neuroplasticity. TLR3-deficient mice display augmented volumes of the hippocampal CA1 and DG regions, along with heightened levels of the GluR1 AMPA receptor subunit in the CA1 region (Okun et al., 2010). In TLR4 knockout mice, stress-activated GSK-3 $\beta$  activity and the production of hippocampal cytokines and chemokines are attenuated. Similarly, administration of TDZD-8, a GSK-3 $\beta$  inhibitor, significantly reduces the expression of most hippocampal cytokines and chemokines (Cheng et al., 2016). In addition, TDZD-8 treatment downregulates TLR4 protein expression, upregulates claudin5 protein expression, and significantly improves cognitive function in aged mice (Liang et al., 2020).

Different transcription factors, such as nuclear factor kappa B (NF- $\kappa$ B), c-Jun, nuclear factor of activated T cells (NFAT), cAMP response element-binding protein (CREB), and signal transducer and activator of transcription 3 (STAT3), have been shown to be substrates for GSK-3 $\beta$  activity (Gao et al., 2017; Götschel et al., 2008; Lakshmanan et al., 2015). Furthermore, it is well documented that these GSK-3 $\beta$  substrates play important roles in the regulation of neuroinflammation (Golpich et al., 2015). Among these factors, NF- $\kappa$ B is a principal transcription factor activated by inflammatory processes leading to cytokine production. A previous study has indicated that NF- $\kappa$ B is regulated by GSK-3 $\beta$  at the level of the transcriptional complex (Hoefflich et al., 2000). In addition, GSK-3 $\beta$  directly phosphorylates the NF- $\kappa$ B protein at Ser468 in unstimulated cells, thereby controlling the basal activity of NF- $\kappa$ B (Buss et al., 2004). NF- $\kappa$ B is expressed in microglia, neurons, and astrocytes, and it plays an important role in the regulation of inflammatory intermediates during neuronal dysfunction (Singh et al., 2020). NFAT activity is needed for the A-stimulated microglial activation that occurs during Alzheimer's disease (Rojanathammanee et al., 2015). Activation of STAT3 and the c-Jun signalling pathway may contribute to neuroinflammation and cognitive impairment (Hu et al., 2021b; Li et al., 2022). However, activation of the CREB signalling pathway significantly increases the survival of neurons, improves cognitive behaviour, and promotes anti-inflammatory effects (Sharma and Singh, 2020).

### **GSK-3 $\beta$ , neuronal survival, and neurogenesis**

Neurogenesis involves cell survival, proliferation, and differentiation, which are strictly controlled by epigenetic mechanisms of gene transcription. Neurogenesis is traditionally recognized as an embryogenic phenomenon, but numerous studies have suggested that this process exists in certain areas of the brain (Espinós et al., 2022). In recent years, neurogenesis has attracted much attention, as drugs and targets promoting neurogenesis have shown beneficial results in the treatment of neurodegenerative diseases. In support of the hypothesis that GSK-3 $\beta$  is an important regulator in neurogenesis, transgenic mice overexpressing GSK-3 $\beta$  exhibit a reduction in proliferation and maturation of new functional DG neurons, as well as a severe memory impairments. This GSK-3 $\beta$ -dependent depletion of neurogenesis contributes to microglial activation and A $\beta$ -mediated neuronal death, further promoting neurodegeneration (Hernandez et al., 2013). GSK-3 $\beta$  inhibition has been shown to enhance the proliferation, migration, and differentiation of neural stem cells in vitro. Inhibition of GSK-3 $\beta$  with the small molecule, tideglusib, induces neurogenesis in the DG of the hippocampus in vivo (Morales-Garcia et al., 2012). The above findings indicate that inactivation of GSK-3 $\beta$  is beneficial to promote the survival of functional neurons in neuropathology with impaired neurogenesis.

A deficit in hippocampal neurogenesis contributes to cognitive decline in old age. Activated GSK-3 $\beta$



accelerates hippocampal neurogenesis at the early stage, while pharmacological inhibition of GSK-3 $\beta$  is efficient in preserving hippocampal neurogenesis in senescent mice (Liu et al., 2020a). Inflammation is also implicated in the neuropathology of neurodegenerative disorders, as increased levels of proinflammatory cytokines, such as interleukin-1 $\beta$  (IL-1 $\beta$ ), have been shown to be detrimental to hippocampal neurogenesis. Previous results have suggested that GSK-3 $\beta$  activation is involved in the antiproliferative and progliogenic effects of IL-1 $\beta$ , and reduced GSK-3 $\beta$  activity facilitates the restoration of hippocampal neurogenesis in neuroinflammatory conditions (Green and Nolan, 2012). Glucose is indispensable for neuronal survival, and even minor changes in the glucose supply may result in serious consequences to the normal physiology of the nervous system. Thus, glucose homeostasis needs to be precisely regulated for the normal survival of neurons. Given the pivotal roles of GSK-3 $\beta$  in insulin sensitivity, glycogen synthesis, and glucose metabolism, GSK-3 $\beta$  is also necessary for neurogenesis.

### **GSK-3 $\beta$ and neural migration**

Considerable evidence shows that GSK-3 $\beta$  directly or indirectly regulates neuronal migration in the nervous system. Several studies have investigated GSK-3 $\beta$  targeting the APC protein. Apart from its involvement in protein degradation, APC is a cytoskeletal protein that relies on microtubules and contributes to the maintenance of the polarized glial scaffold during brain development (Barth et al., 2008; Fang and Svitkina, 2022). In the absence of APC, glial cells lose their polarity and respond to extracellular polarity signals, such as neuregulin-1. Conditional gene targeting to eliminate APC further induces instability in the glial microtubule cytoskeleton, thereby indicating a significant role of APC in neuronal migration (Yokota et al., 2009). It remains unclear whether the migration defects are caused by the disruption of the glial scaffold or the absence of microtubule-associated APC in cortical neurons.

GSK-3 $\beta$  has been implicated in the regulation of neuronal migration through its influence on  $\beta$ -catenin. Mouse studies have provided evidence that genetic manipulation of  $\beta$ -catenin, either through deletion or overexpression, disrupts the development of the brain and spinal system (Zechner et al., 2003). The migration process primarily arises from aberrant neural progenitor development. In the context of a mutated nervous system, the presence of abnormally localized neurons has also been observed. Additionally, the levels of  $\beta$ -catenin in progenitor cells influence the positioning of neurons (Mutch et al., 2009). A possible explanation for the abnormal positioning of neurons is delayed determination of progenitor fate. However, studies on  $\beta$ -catenin mutations provide strong evidence for the potential involvement of GSK-3 $\beta$  in neuronal migration. Morgan-Smith M et al. demonstrated that GSK-3 $\beta$  is essential for radial migration and dendritic orientation. Interestingly, this GSK-3 $\beta$  regulation of migration in neurons is independent of  $\beta$ -catenin signalling (Morgan-Smith et al., 2014).

Researchers have also investigated the molecular regulation of GSK-3 $\beta$  in the migration of neurons. For example, DISC1, a prominent susceptibility factor for numerous mental disorders, governs the process of neuronal migration during brain development and has been implicated in the regulation of neuronal migration through its interaction with GSK-3 $\beta$ . At midembryonic stages, when neural progenitor proliferation is active, DISC1 interacts with GSK-3 $\beta$ . However, at later embryonic stages, when neuronal proliferation is occurring, DISC1 dissociates from GSK-3 $\beta$  (Ishizuka et al., 2011). LKB1, an evolutionally conserved polarity kinase also known as the upstream kinase of AMPK, is another important regulator of neuronal migration in the development of neurons. Asada N et al. reported that LKB1 phosphorylates GSK-3 $\beta$  at Ser9, thereby promoting neuronal

migration in the developing neocortex (Asada and Sanada, 2010). The delineation of the roles and underlying mechanisms of GSK-3 $\beta$  in neuronal migration holds paramount importance, as abnormalities in neuron migration are implicated in various neural disorders.

### **GSK-3 $\beta$ and cerebral ischaemia**

The brain, being the most vulnerable organ to hypoxia and ischaemia, is highly susceptible to the detrimental effects of cerebral ischaemia. This condition poses a significant threat to human life because it can lead to severe impairment of brain functions. Additionally, both cerebral ischaemia and the subsequent reperfusion phase inflict extensive damage to the brain. Numerous studies have indicated the pivotal involvement of GSK-3 $\beta$  in neuronal injury following cerebral ischaemia. Most recently, Peng S et al. observed downregulation of AKT accompanied by activation of GSK-3 $\beta$  within 12 hours in cerebral ischaemia/reperfusion, and they suggested this downregulation is mediated by mTORC instead of PI3K and PDK1 signalling (Peng et al., 2022). Downregulation of GSK-3 $\beta$  using a siRNA-mediated approach markedly attenuates neuronal damage and enhances cell viability in rat models of ischaemia/reperfusion. Conversely, upregulation of GSK-3 $\beta$  significantly exacerbates neurological impairments and inflicts damage upon cerebral cortical neurons in ischaemia/reperfusion models (Li et al., 2016). In line with the findings of Kisoh K et al., experimental models have demonstrated a substantial increase in the phosphorylation of GSK-3 $\beta$  at Ser9, as well as the phosphorylation of AKT, on the seventh day following cerebral ischaemia. Additionally, administration of a PI3K inhibitor results in a reduction in AKT activation and phosphorylation of GSK-3 $\beta$  at Ser9 in response to cerebral ischaemia (Kisoh et al., 2017). Chen BH et al. examined the expression of GSK-3 $\beta$  and p-GSK-3 $\beta$  in the gerbil hippocampal CA1 area after transient cerebral ischaemia, and they reported that p-GSK-3 $\beta$  is highly expressed in astrocytes located in the stratum oriens and radiatum. The results indicate an increase in GSK-3 $\beta$  immunoreactivity in pyramidal cells of the CA1 region at 6 h following ischaemia–reperfusion. However, GSK-3 $\beta$  levels decrease after 12 h and are scarcely detectable in CA1 pyramidal cells at 5 days postischaemia–reperfusion. Furthermore, p-GSK-3 $\beta$  is slightly decreased in CA1 pyramidal cells at 6 and 12 h but significantly increased at 1 and 2 days, and it is barely detectable in CA1 pyramidal cells at 5 days after ischaemia–reperfusion (Chen et al., 2017).

The occurrence of ischaemia–reperfusion (I/R) injury can lead to neuronal cell death, and it is closely related to oxidative stress. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a master regulator of oxidative stress correlated with brain I/R injury. Several studies have suggested that activation of Nrf2 via GSK-3 $\beta$  modulation alleviates I/R-induced apoptosis and oxidative stress in neurons (Bai et al., 2021; Duan et al., 2019; Liao et al., 2020; Xu et al., 2021). The involvement of GSK-3 $\beta$  in ischaemia–reperfusion models can be attributed to its regulation of the Nrf2/ARE pathway, resulting in a reduction in oxidative stress. Treatment with a GSK-3 $\beta$  inhibitor or GSK-3 $\beta$  siRNA has been shown to prevent neuronal injury caused by brain ischaemia through the activation of the Nrf2 signalling pathway (Li et al., 2016; Pang et al., 2016). In support of the hypothesis that GSK-3 $\beta$  is an important regulator in cerebral ischaemia, inhibition of GSK-3 $\beta$  using GSK-3 $\beta$  inhibitors enables protection of the brain from injury in various ischaemia–reperfusion models (Gao et al., 2021; Wang et al., 2017).

Altogether, GSK-3 $\beta$  is widely involved in physiological and pathological processes of the brain with sophisticated regulatory mechanisms. It is imperative for future investigations to elucidate additional facets of GSK-3 $\beta$  signalling in cerebral development that could hold therapeutic significance in the realm of brain disorders.

## GSK-3 $\beta$ and its Inhibitors in Brain Diseases

Due to its multifaceted functions in the brain, GSK-3 $\beta$  has been strongly implicated in a variety of brain diseases, including Alzheimer's disease, Parkinson's disease, mood disorders, schizophrenia, and glioblastoma (GBM). Substantial evidence suggests that small molecule inhibitors of GSK-3 $\beta$  have great value in drug development. Generally, GSK-3 $\beta$  inhibitors are classified into the following four categories: 1) cations encompassing the mood stabilizer lithium, along with other metal ions, such as zinc and copper, which exert inhibitory effects on GSK-3 $\beta$  at millimolar or submicromolar concentrations, respectively; 2) ATP competitive inhibitors; 3) allosteric non-ATP competitive inhibitors; and 4) substrate competitive inhibitors (SCIs). The representative GSK-3 $\beta$  inhibitors and their reported effects in brain diseases are summarized in Table 1.

To date, researchers have invested a great deal of effort into developing GSK-3 $\beta$  inhibitors as potential drugs, especially for the treatment of neurodegenerative and psychiatric disorders (Arciniegas Ruiz and Eldar-Finkelman, 2021). For example, AF3581 is a new class of GSK-3 $\beta$  inhibitor, and studies have shown that this compound is effective in chronic mild stress-induced depression (mimicking the low phase of bipolar disorder) and mouse aggression (mimicking the high phase), indicating the therapeutic potential of GSK-3 $\beta$  inhibitors in treating patients with bipolar disorder (Capurro et al., 2020). Previous studies have documented the efficacy of Schisandrin B stereoisomers as inhibitors of GSK-3 $\beta$ . These stereoisomers have demonstrated the ability to substantially enhance the expression of p-GSK-3 $\beta$  (Ser9) while reducing the expression of p-GSK-3 $\beta$  (Tyr216 and Tyr279). Furthermore, they have shown promising effects in mitigating cell damage induced by amyloid  $\beta$  (A $\beta$ ) and ameliorating cognitive impairment in mice with Alzheimer's disease (AD). These findings highlight the potential utility of these stereoisomers as neuroprotective agents for the treatment of Alzheimer's disease (Hu et al., 2019). After intranasal administration of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), rodents display time-dependent impairments in emotion, cognition, and motor function, thus mimicking Parkinson's disease in rodents. Lithium salt, a GSK-3 $\beta$  inhibitor, prevents olfactory discrimination and short-term memory impairment in an intranasal MPTP-induced Parkinson's disease rat model (Castro et al., 2012).

## GSK-3 $\beta$ and Alzheimer's disease

Alzheimer's disease (AD) is a neurodegenerative disease, a type of late-onset dementia, characterized by gradual loss of memory, loss of speech ability, cognitive impairment, and other mental symptoms. AD is characterized by profound alterations in the morphology of numerous neuronal somata and axon terminals. The typical neuropathological changes of AD include the formation of senile plaques, which are deposits of A $\beta$  protein in the brain, neurofibrillary tangles formed by the excessive phosphorylation of tau protein, neuronal apoptosis, and a series of inflammatory reactions (Chou et al., 2012). Neurofibrillary degeneration (NFD) is a type of intracellular damage caused by the modification of tau protein, which is a structural protein involved in stabilizing the neuronal cytoskeleton. During AD, tau protein undergoes hyperphosphorylation, which causes it to detach from microtubules and form paired pathological helical filaments, leading to progressive neurofibrillary degeneration.

GSK-3 $\beta$  is significantly expressed in the neuronal cell bodies of the brains of AD patients (Pei et al., 1997). Studies have shown that GSK-3 $\beta$  is a key enzyme for the hyperphosphorylation of tau protein. Overexpression of GSK-3 $\beta$  in the brains of transgenic mice leads to the hyperphosphorylation of tau protein, microtubule breakdown, and the appearance of neurofibrillary degeneration (Brownlee et al., 1997; Hernandez et al., 2013).

The activity of GSK-3 $\beta$  contributes to the production of A $\beta$  and A $\beta$ -mediated neuronal death. Mechanistically, tau protein is a substrate for GSK-3 $\beta$  phosphorylation and p-tau enable to form paired pathological helical filaments, leading to progressive neurofibrillary degeneration. GSK-3 $\beta$  is involved in regulating the protein hydrolysis and cleavage of amyloid precursor protein (APP), as well as the production of A $\beta$ , which is a small toxic peptide. A $\beta$  is derived from the protein hydrolysis of APP, and inhibiting GSK-3 $\beta$  reduces the production of A $\beta$  and protects neurons from the toxic effects of the peptides. In turn, abnormal aggregation of A $\beta$  increases the activity of GSK-3 $\beta$ , establishing a mechanistic link between the two major hallmarks of AD (accumulation of A $\beta$  and NFD) (Qing et al., 2008; Terwel et al., 2008). Studies have found that inhibiting the activity of GSK-3 $\beta$  reduces the production of A $\beta$  and tau phosphorylation while improving learning and memory abilities [145]. Therefore, GSK-3 $\beta$  is considered a potential target for treating Alzheimer's disease. Currently, some GSK-3 $\beta$  inhibitors have entered preclinical studies, but their efficacy and safety still need further verification (Arciniegas Ruiz and Eldar-Finkelman, 2021).

### GSK-3 $\beta$ in Parkinson's Syndrome

Parkinson's disease (PD) is a chronic neurodegenerative movement disorder, and it is the second most common neurodegenerative disease. PD is characterized by the degeneration of substantia nigra dopaminergic neurons and depletion of dopamine in the striatum, as well as the abnormal accumulation of  $\alpha$ -synuclein and synphilin-1 ( $\alpha$ -synuclein-interacting protein), resulting in pathological and clinical abnormalities (Jankovic and Tan, 2020). Dopamine helps to guide muscle activity, and when there is a significant loss of dopaminergic neurons in the substantia nigra of the brain, characteristic movement difficulties of PD, such as tremors, rigidity, bradykinesia, impaired balance, and impaired coordination, can occur (Reich and Savitt, 2019).

Numerous studies have found that the expression of GSK-3 $\beta$  is higher in the brains of patients with PD. In the striatum of PD patients after death, the levels of  $\alpha$ -synuclein and p-GSK-3 $\beta$  phosphorylated at Tyr 216 are higher, and activated GSK-3 $\beta$  hyperphosphorylates tau to produce toxic pathological forms of p-tau (Wills et al., 2010). In transgenic mouse models, the activation of GSK-3 $\beta$  depends on the presence of  $\alpha$ -synuclein, and overexpression of  $\alpha$ -synuclein is associated with increased activity of GSK-3 $\beta$ , indicating that the accumulation of  $\alpha$ -synuclein contributes to the activation and increase of GSK-3 $\beta$  in the brain of PD, leading to excessive phosphorylation of tau. Both in vitro and in vivo, GSK-3 $\beta$  is activated by MPTP in a strictly  $\alpha$ -synuclein-dependent manner and is responsible for the excessive phosphorylation of tau protein (Duka et al., 2009). Mechanistically,  $\alpha$ -synuclein is a substrate for GSK-3 $\beta$  phosphorylation and has been reported to activate NLRP3 inflammasome assembly by TLR2 stimulation. Interestingly,  $\alpha$ -synuclein reversely leads to GSK-3 $\beta$  activation, and an acidic region of  $\alpha$ -synuclein is responsible for the stimulation of GSK-3 $\beta$ -mediated tau phosphorylation (Samim Khan et al., 2023). Thus, it has been concluded that there is crosstalk between  $\alpha$ -synuclein and GSK-3 $\beta$ -mediated tau phosphorylation, resulting in PD progression. A previous study has reported that heat shock protein 70 (Hsp70) suppresses the  $\alpha$ -synuclein-induced phosphorylation of tau by GSK-3 $\beta$  through direct binding to  $\alpha$ -synuclein, suggesting that Hsp70 may act as a novel therapeutic target to counteract  $\alpha$ -synuclein-mediated tau phosphorylation in PD (Kawakami et al., 2011).

Inhibition of GSK-3 $\beta$  reduces the accumulation of  $\alpha$ -synuclein and alleviates the excessive phosphorylation of tau protein (Duka et al., 2009). Synphilin-1 is an  $\alpha$ -synuclein-interacting protein, and GSK-3 $\beta$  specifically phosphorylates synphilin-1, reducing its ubiquitination in vitro and in vivo, which reduces its degradation by proteasomes (Avraham et al., 2005). A small amount of GSK-3 $\beta$  is also detected in mitochondria, and studies

have shown that increased mitochondrial GSK-3 $\beta$  activity enhances the production of reactive oxygen species and disrupts mitochondrial morphology. Chemical inhibitors of GSK-3 $\beta$  inhibit cell apoptosis induced by rotenone, an inhibitor of complex I of the electron transport chain, and attenuate GSK-3 $\beta$ -mediated damage to mitochondria (King et al., 2008). Inhibition of GSK-3 $\beta$  activity is a key element in the treatment of PD, as GSK-3 $\beta$  appears to be a specific therapeutic target for PD due to its involvement in neuronal apoptosis, phosphorylation of tau protein, aggregation of  $\alpha$ -synuclein, aggregation of synphilin-1, and mitochondrial dysfunction (Golpich et al., 2015).

### **GSK-3 $\beta$ in Bipolar Disorder**

Bipolar disorder (BD), also known as manic-depressive illness, is a severe mental disorder. The symptoms of bipolar disorder first appear in adolescence or early adulthood, and they recur unpredictably with features of cyclic alternation between manic and depressive moods, characterized by periodic disturbances in mood, energy patterns, and behaviour (Vieta et al., 2018). Lithium and valproic acid are representative drugs and mood stabilizers used for bipolar disorder, which can alleviate mild to moderate manic episodes in BD patients (Scarselli, 2023). Both lithium and valproic acid exert therapeutic effects by inhibiting GSK-3 $\beta$  (Dandekar et al., 2018). Drugs used to treat bipolar disorder usually inhibit GSK-3 $\beta$ , indicating that GSK-3 $\beta$  plays a critical role in the therapeutic effects of bipolar disorder treatment.

GSK-3 $\beta$  is not only a target of lithium but also of other categories of mood stabilizers, antidepressants, and antipsychotic drugs (Beaulieu, 2007). In transgenic mice overexpressing GSK-3 $\beta$ , research has shown that the overexpression of GSK-3 $\beta$  leads to hyperactivity and ADHD-like symptoms, similar to human mania (Prickaerts et al., 2006). Heterozygous GSK-3 $\beta$  mice exhibit normal morphology, reduced exploratory activity, and a lithium-mimetic antidepressant-like state (O'Brien et al., 2004). Increasing evidence supports the important role of GSK-3 $\beta$  in the treatment of mania and depression (Dandekar et al., 2018). GSK-3 $\beta$  represents a promising therapeutic target for addressing depression, with demonstrated antidepressant effects observed through the administration of GSK-3 $\beta$  inhibitors in animal models (Rosa et al., 2008). Increased GSK-3 $\beta$  kinase activity has been detected in brain samples of human patients with bipolar disorder (BD), and abnormal GSK-3 $\beta$  activity occurs in patients with severe depression, indicating that abnormal GSK-3 $\beta$  activity promotes the onset of BD (Polter et al., 2010). Dysregulation of serotonin (5-HT) neurotransmission in the brain is considered the basis of bipolar disorder. In transgenic mice, the decrease in brain 5-HT levels is accompanied by the activation of GSK-3 $\beta$ , and the inactivation of GSK-3 $\beta$  alleviates abnormal behaviour caused by 5-HT deficiency. Various 5-HT drugs inhibit brain GSK-3 $\beta$  signalling, indicating that targeting the GSK-3 $\beta$  signalling pathway may provide a strategy for the treatment of certain 5-HT-related psychiatric disorders (Beaulieu et al., 2008; Latapy et al., 2012; Zheng et al., 2021).

A considerable array of GSK-3 $\beta$  inhibitors has been examined in models of bipolar disorder (BD). In mouse models of mania, administration of various GSK-3 $\beta$  inhibitors, including TDZD-8, SB-216763, SB-627772, AF3581, and indirubins, mitigates the "manic" effects, as evidenced by decreased hyperactivity and restoration of normal ambulation behaviour. GSK-3 $\beta$  inhibitors also demonstrate notable therapeutic advantages in addressing the depressive phase in animal models. Administration of tideglusib, VP2.51, and the SCI peptide yields antidepressant-like outcomes (Arciniegas Ruiz and Eldar-Finkelman, 2021).

### **GSK-3 $\beta$ and Schizophrenia**

Schizophrenia is a common debilitating neurological disorder with a multigenetic basis, and it is characterized by poor emotional response, impaired language reasoning, and significant social dysfunction (Zamanpoor, 2020). AKT1 and GSK-3 $\beta$  play critical roles in synaptic plasticity and neuronal function in the central nervous system. The AKT/GSK-3 $\beta$  signalling pathway is involved in schizophrenia, with AKT1 being a potential susceptibility gene for schizophrenia. Cognitive impairment, abnormal synaptic morphology, neuronal atrophy, and dysfunction of neurotransmitter signalling may partly be explained by reduced PI3K/AKT signalling in schizophrenia. Furthermore, lower levels of AKT may have a detrimental effect on neurodevelopment by increasing the effect of risk factors, attenuating the effect of growth factors, and reducing the response of patients to treatment of antipsychotic agents (Zheng et al., 2012). AKT1-3 are upstream inhibitors of GSK-3 $\beta$ , and Ser9 in GSK-3 $\beta$  is a phosphorylation site for AKT. Phosphorylation of GSK-3 $\beta$  (Ser9) leads to its inactivation. It has been reported that the protein level of AKT1 is reduced in patients with schizophrenia, accompanied by a decrease in phosphorylation of Ser9 in GSK-3 $\beta$  and an increase in GSK-3 $\beta$  activity (Emamian et al., 2004). GSK-3 $\beta$  has hundreds of single nucleotide polymorphisms (SNPs), and it has been reported that individuals carrying the C allele of the GSK-3 $\beta$  promoter region C are more susceptible to schizophrenia (Tang et al., 2013). In schizophrenia (SZ), carriers of the low-activity C allele variant have significantly higher brain volume in the temporal lobe, which is the brain parenchymal region with the most consistent morphological abnormalities in schizophrenia. The neuropathological process in this area develops rapidly at the onset of the disease, suggesting that carrying the low-activity mutant C allele gene protects the brain from related neuropathological damage. GSK-3 $\beta$  is an important factor that affects the neuropathology of schizophrenia (Benedetti et al., 2010).

### **GSK-3 $\beta$ and Tumour Therapy**

GSK-3 $\beta$  is associated with the occurrence and development of tumours. Given its ability to negatively regulate oncogenic proteins, such as MYC and  $\beta$ -catenin, GSK-3 $\beta$  likely plays a suppressive role in tumourigenesis. Nevertheless, several investigations have demonstrated that GSK-3 $\beta$  may exhibit a positive regulatory role in tumourigenesis within human ovarian, liver, colon, and pancreatic carcinomas (Luo, 2009). Whether GSK-3 $\beta$  plays a tumour suppressor role or serves as a tumour promoter depends on the cell type and cellular context (Takahashi-Yanaga, 2013). Glioblastoma (GBM) is the most common primary intracranial tumour, originating from glial cells and accounting for approximately 81% of intracranial tumours (Xu et al., 2020). The expression levels of GSK-3 $\beta$  and Tyr216-phosphorylated GSK-3 $\beta$  are higher in GBM tissues than in nontumor brain tissues. GSK-3 $\beta$  in GBM often plays a pro-cancer role. Inhibiting the activity or expression of GSK-3 $\beta$  in vitro reduces the survival and proliferation of glioblastoma cells, as well as induces apoptosis of glioblastoma cells and delays the growth of mouse neuroblastoma tumours (Dickey et al., 2011). The phosphorylation of GSK-3 $\beta$  at Ser9 results in the inhibition of GSK-3 $\beta$ , which subsequently leads to the activation of glycogen synthase, thereby facilitating glycogen synthesis. Glioblastoma cells exhibit notably elevated levels of glycogen, and the accumulation of glycogen serves as a promoter for the growth of GBM (Majewska and Szeliga, 2017). In the clinical setting, it has been observed that the levels of GSK-3 $\beta$  and its phosphorylated form (Tyr216) are elevated in tumours compared to normal tissues. Significant inhibition of GSK-3 $\beta$  has been shown to reduce the survival and proliferation of glioblastoma cells, and this effect is accompanied by an upregulation of p53 and p21 expression (Miyashita et al., 2009). In addition, Kotliarova S et al. reported that inhibition of GSK-3 $\beta$  activity results in c-MYC activation, leading to the induction of glioma

cell death (Kotliarova et al., 2008).

Of note, the role of GSK-3 $\beta$  in glioblastoma tumours is controversial (Figure 3). Li Y et al. demonstrated that GSK-3 $\beta$  is highly expressed and activated during differentiation in sensitive C6 malignant glioma cells. Interference of GSK-3 $\beta$  activity with GSK-3 $\beta$  inhibitors or siRNA potently suppresses differentiation in sensitive C6 cells. Conversely, overexpression of a constitutively active form of GSK-3 $\beta$  (GSK-3 $\beta$ -S9A) mutant in differentiation-resistant U251 glioma cells restores their differentiation abilities (Li et al., 2010). Zhao P et al. demonstrated that the levels of p-GSK-3 $\beta$  (Ser9) are significantly upregulated in glioma tissues compared to normal tissues. Ectopic expression of GSK-3 $\beta$  in glioma cells significantly inhibits tumour growth, which is accompanied by a decrease in  $\beta$ -catenin expression and downregulation of p-mTOR and p-p70S6K1 (Zhao et al., 2015). Consistently, inactivation or degradation of GSK-3 $\beta$  promotes glioblastoma invasion and epithelial-mesenchymal transition (EMT) in glioblastoma cells (Li et al., 2019; Yang et al., 2019). Furthermore, CHIR99021, a GSK-3 $\beta$  inhibitor, greatly increases glioma stem-like cell (GSLC) properties in patient-derived glioma samples (Yang et al., 2020). Nevertheless, increasing evidence also suggests that GSK-3 $\beta$  is an important molecule that leads to the malignant phenotype of GBM. AZD2858 is an effective adjunct to glioma radiotherapy at clinical doses. Most recently, studies have shown that GSK-3 $\beta$  inhibition by AZD2858 promotes glioma cell death by disrupting centrosome function and inducing mitotic defects (Brüning-Richardson et al., 2021).

## Conclusions and Perspectives

In the present review, the possible underlying mechanisms that connect GSK-3 $\beta$  signalling with the physiopathology of the brain are discussed. From our perspective, the function of GSK-3 $\beta$  in brain pathophysiology is complicated and not entirely understood. GSK-3 $\beta$  is implicated in a variety of psychiatric and neurological disorders, emphasizing both its importance and complexity within the brain. It is meaningful to elucidate the basic biological mechanisms underlying GSK-3 $\beta$  in the brain because the increased understanding of the relationship between GSK-3 $\beta$  and the neural system greatly impacts the progress in treating several neurological diseases. Notably, there is a different opinion in the field regarding the effects of GSK-3 $\beta$  on cognition in the brain. For example, studies have shown that GSK-3 $\beta$  activity in the amygdala and hippocampus is needed for memory reconsolidation (Hong et al., 2012; Xie et al., 2022). With an increase in total GSK-3 $\beta$  and phosphorylated GSK-3 $\beta$  (Tyr216) in adult mice after exercise, Zang J et al. observed that there is a significant enhancement in adult neurogenesis and cognitive functions (Zang et al., 2017). In old mice, GSK-3 $\beta$  is needed for memory formation (Koike et al., 2021). These findings suggest that inhibition of GSK-3 $\beta$  in neurological disorders may bring potential risk of cognitive impairment. To some degree, these results also possibly explain the poor efficacy of GSK-3 $\beta$  inhibitors in preserving memory capacity in AD patients.

GSK-3 $\beta$  has emerged as a pivotal molecule with multifaceted roles in various cellular processes, extending from glycogen metabolism to critical signalling pathways, especially those involving  $\beta$ -catenin and NF- $\kappa$ B. The intricate regulatory mechanisms of GSK-3 $\beta$ , particularly through phosphorylation events, underscore its significance in both normal cellular homeostasis and pathological conditions. In the context of glioblastoma (GBM), the aberrant activation of the GSK3 $\beta$ / $\beta$ -catenin pathway and the consequential modulation of key proteins, such as c-Myc and c-Jun, highlight the potential of GSK-3 $\beta$  as a therapeutic target. However, the role of GSK-3 $\beta$  in glioblastoma tumours is still controversial. Considering the carcinogenic risk of GSK-3 $\beta$  in gliomas, GSK-3 $\beta$  inhibitors should be used with caution as a treatment for glioblastoma. It is best to utilize an effective GSK-3 $\beta$  inhibitor that does not cause any carcinogenic risk while having an ideal anticancer effect

through the appropriate dose range. Overall, the profound influence of GSK-3 $\beta$  on GBM malignancy, especially its implications in GSC survival, warrants further investigation to elucidate its complete role and pave the way for targeted therapeutic strategies in GBM management.

Comprehensive investigations are imperative to elucidate the precise signalling mechanisms regulated by GSK-3 $\beta$  and the mechanistic contributions of GSK-3 $\beta$  in the development of brain diseases. Urgent efforts are required to expedite advancements in substantiating the advantages of GSK-3 $\beta$  interventions and addressing the challenges associated with their implementation as a therapeutic strategy for brain diseases. Despite encouraging outcomes observed in preclinical studies utilizing GSK-3 $\beta$  inhibitors, caution must be exercised prior to their utilization, necessitating further examinations to assess both their beneficial effects and potential side effects during application. With the rapid development of technologies that allow elucidation of the characteristics and pattern of GSK-3 $\beta$  in brain diseases, the time has come to build on these findings to dissect the complexity of organisms and exploit the optimal therapy method.

While many studies have contributed to the understanding of how GSK-3 $\beta$  is involved in brain physiology and pathology, the recent development of conditional gene editing, molecular imaging, and optogenetic tools allows a more precise determination of the functional significance of these findings and application of them to improve the understanding and treatment of neurological disorders. Combination therapeutic strategies utilizing GSK-3 $\beta$  inhibitors hold great promise for the treatment of Alzheimer's disease (AD), Parkinson's disease (PD), and bipolar disorder (BD). The activity of GSK-3 $\beta$  is regulated by diverse stimuli, which can lead to its activation or inactivation. This phenomenon is intricately associated with the complex characteristics and genetic heterogeneity of brain disorders, particularly in the realm of brain tumours. Personalized genomics and diverse omics methodologies hold promise in identifying patients who may derive therapeutic benefits from GSK-3 $\beta$  inhibitor treatment. Ultimately, this knowledge can guide clinical practitioners in selecting the most appropriate therapy for each individual patient.

## Author contributions

Conceptualization, S.Z. and S.L.; literature analysis, S.Z. and S.L.; writing—original draft preparation, S.L., P.W. and J.G.; writing—review and editing, S.Z. and P.W.; funding acquisition, S.Z. and J.G. All authors have read and agreed to the published version of the manuscript.

## Conflicts of Interest

The authors declare no conflict of interest.

## References

- Aguilar-Morante, D., J. A. Morales-Garcia, M. Sanz-SanCristobal, M. A. Garcia-Cabezas, A. Santos and A. Perez-Castillo 2010. Inhibition of glioblastoma growth by the thiadiazolidinone compound TDZD-8. *PLoS One* 5: e13879. doi: 10.1371/journal.pone.0013879
- Al-Zaidi, A. S., N. Mohtarrudin, J. Chupri and M. Hassan 2021. Differential time course of glycogen synthase kinase-3 inhibition in experimental autoimmune encephalomyelitis. *Malays J Pathol* 43: 413-424.



Amar, S., R. H. Belmaker and G. Agam 2011. The possible involvement of glycogen synthase kinase-3 (GSK-3) in diabetes, cancer and central nervous system diseases. *Curr Pharm Des* 17: 2264-2277. doi: 10.2174/138161211797052484

Arciniegas Ruiz, S. M. and H. Eldar-Finkelman 2021. Glycogen Synthase Kinase-3 Inhibitors: Preclinical and Clinical Focus on CNS-A Decade Onward. *Front Mol Neurosci* 14: 792364. doi: 10.3389/fnmol.2021.792364

Asada, N. and K. Sanada 2010. LKB1-mediated spatial control of GSK3 $\beta$  and adenomatous polyposis coli contributes to centrosomal forward movement and neuronal migration in the developing neocortex. *J Neurosci* 30: 8852-8865. doi: 10.1523/jneurosci.6140-09.2010

Ashrafian, H., E. H. Zadeh and R. H. Khan 2021. Review on Alzheimer's disease: Inhibition of amyloid beta and tau tangle formation. *Int J Biol Macromol* 167: 382-394. doi: 10.1016/j.ijbiomac.2020.11.192

Avraham, E., R. Szargel, A. Eyal, R. Rott and S. Engelender 2005. Glycogen synthase kinase 3 $\beta$  modulates synphilin-1 ubiquitylation and cellular inclusion formation by SIAH: implications for proteasomal function and Lewy body formation. *J Biol Chem* 280: 42877-42886. doi: 10.1074/jbc.M505608200

Bahmad, H. F., R. M. Chalhoub, H. Harati, J. Bou-Gharios, S. Assi, F. Ballout, A. Monzer, H. Msheik, T. Araji, M. K. Elajami, P. Ghanem, F. Chamaa, H. Kadara, T. Abou-Antoun, G. Daoud, Y. Fares and W. Abou-Kheir 2021. Tideglusib attenuates growth of neuroblastoma cancer stem/progenitor cells in vitro and in vivo by specifically targeting GSK-3 $\beta$ . *Pharmacol Rep* 73: 211-226. doi: 10.1007/s43440-020-00162-7

Bai, J., P. Jia, Y. Zhang, K. Wang and G. Wu 2021. Paraoxonase 2 protects against oxygen-glucose deprivation/reoxygenation-induced neuronal injury by enhancing Nrf2 activation via GSK-3 $\beta$  modulation. *Hum Exp Toxicol* 40: 1342-1354. doi: 10.1177/0960327121996032

Barth, A. I., H. Y. Caro-Gonzalez and W. J. Nelson 2008. Role of adenomatous polyposis coli (APC) and microtubules in directional cell migration and neuronal polarization. *Semin Cell Dev Biol* 19: 245-251. doi: 10.1016/j.semcdb.2008.02.003

Beaulieu, J. M. 2007. Not only lithium: regulation of glycogen synthase kinase-3 by antipsychotics and serotonergic drugs. *Int J Neuropsychopharmacol* 10: 3-6. doi: 10.1017/s1461145706006857

Beaulieu, J. M., X. Zhang, R. M. Rodriguiz, T. D. Sotnikova, M. J. Cools, W. C. Wetsel, R. R. Gainetdinov and M. G. Caron 2008. Role of GSK3  $\beta$  in behavioral abnormalities induced by serotonin deficiency. *Proc Natl Acad Sci U S A* 105: 1333-1338. doi: 10.1073/pnas.0711496105

Benedetti, F., S. Poletti, D. Radaelli, A. Bernasconi, R. Cavallaro, A. Falini, C. Lorenzi, A. Pirovano, S. Dall'aspezia, C. Locatelli, G. Scotti and E. Smeraldi 2010. Temporal lobe grey matter volume in schizophrenia is associated with a genetic polymorphism influencing glycogen synthase kinase 3- $\beta$  activity. *Genes Brain Behav* 9: 365-371. doi: 10.1111/j.1601-183X.2010.00566.x

Bennecib, M., C. X. Gong, I. Grundke-Iqbal and K. Iqbal 2000. Role of protein phosphatase-2A and -1 in the regulation of GSK-3, cdk5 and cdc2 and the phosphorylation of tau in rat forebrain. *FEBS Lett* 485: 87-93. doi: 10.1016/S0014-5793(00)02203-1

Beurel, E., O. Kaidanovich-Beilin, W. I. Yeh, L. Song, V. Palomo, S. M. Michalek, J. R. Woodgett, L. E. Harrington, H. Eldar-Finkelman, A. Martinez and R. S. Jope 2013. Regulation of Th1 cells and experimental autoimmune encephalomyelitis by glycogen synthase kinase-3. *J Immunol* 190: 5000-5011. doi: 10.4049/jimmunol.1203057

Bhat, R. V., J. Shanley, M. P. Correll, W. E. Fieles, R. A. Keith, C. W. Scott and C. M. Lee 2000. Regulation and localization of tyrosine216 phosphorylation of glycogen synthase kinase-3 $\beta$  in cellular and animal models of neuronal degeneration. *Proc Natl Acad Sci U S A* 97: 11074-11079. doi: 10.1073/pnas.190297597

640 Bijur, G. N. and R. S. Jope 2003. Glycogen synthase kinase-3 beta is highly activated in nuclei and mitochondria.  
641 Neuroreport 14: 2415-2419. doi: 10.1097/00001756-200312190-00025

642 Brownlees, J., N. G. Irving, J. P. Brion, B. J. Gibb, U. Wagner, J. Woodgett and C. C. Miller 1997. Tau phosphorylation  
643 in transgenic mice expressing glycogen synthase kinase-3beta transgenes. Neuroreport 8: 3251-3255. doi:  
644 10.1097/00001756-199710200-00013

645 Brüning-Richardson, A., G. C. Shaw, D. Tams, T. Brend, H. Sangane, S. T. Barry, G. Hamm, R. J. A. Goodwin, J. G.  
646 Swales, H. King, L. Steele, R. Morton, A. Widyadari, T. A. Ward, F. Esteves, M. Boissinot, G. Mavria, A. Droop, S. E.  
647 Lawler and S. C. Short 2021. GSK-3 Inhibition Is Cytotoxic in Glioma Stem Cells through Centrosome Destabilization  
648 and Enhances the Effect of Radiotherapy in Orthotopic Models. Cancers (Basel) 13. doi: 10.3390/cancers13235939

649 Buss, H., A. Dörrie, M. L. Schmitz, R. Frank, M. Livingstone, K. Resch and M. Kracht 2004. Phosphorylation of serine  
650 468 by GSK-3beta negatively regulates basal p65 NF-kappaB activity. J Biol Chem 279: 49571-49574. doi:  
651 10.1074/jbc.C400442200

652 Candelario-Jalil, E., R. M. Dijkhuizen and T. Magnus 2022. Neuroinflammation, Stroke, Blood-Brain Barrier  
653 Dysfunction, and Imaging Modalities. Stroke 53: 1473-1486. doi: 10.1161/strokeaha.122.036946

654 Capurro, V., M. Lanfranco, M. Summa, P. F. Porceddu, M. Ciampoli, N. Margaroli, L. Durando, B. Garrone, R. Ombrato,  
655 S. Tongiani and A. Reggiani 2020. The mood stabilizing properties of AF3581, a novel potent GSK-3β inhibitor. Biomed  
656 Pharmacother 128: 110249. doi: 10.1016/j.biopha.2020.110249

657 Castro, A. A., K. Ghisoni, A. Latini, J. Quevedo, C. I. Tasca and R. D. Prediger 2012. Lithium and valproate prevent  
658 olfactory discrimination and short-term memory impairments in the intranasal 1-methyl-4-phenyl-1,2,3,6-  
659 tetrahydropyridine (MPTP) rat model of Parkinson's disease. Behav Brain Res 229: 208-215. doi:  
660 10.1016/j.bbr.2012.01.016

661 Cervello, M., G. Augello, A. Cusimano, M. R. Emma, D. Balasus, A. Azzolina, J. A. McCubrey and G. Montalto 2017.  
662 Pivotal roles of glycogen synthase-3 in hepatocellular carcinoma. Adv Biol Regul 65: 59-76. doi:  
663 10.1016/j.jbior.2017.06.002

664 Chen, B. H., J. H. Ahn, J. H. Park, B. N. Shin, Y. L. Lee, I. J. Kang, S. Hong, Y. H. Kim, Y. H. Jeon, I. H. Kim, J. H. Cho, T. K.  
665 Lee, J. C. Lee, M. H. Won, J. H. Cho and J. B. Moon 2017. Transient Cerebral Ischemia Alters GSK-3β and p-GSK-3β  
666 Immunoreactivity in Pyramidal Neurons and Induces p-GSK-3β Expression in Astrocytes in the Gerbil Hippocampal  
667 CA1 Area. Neurochem Res 42: 2305-2313. doi: 10.1007/s11064-017-2245-5

668 Cheng, Y., M. Pardo, R. S. Armini, A. Martinez, H. Mouhsine, J. F. Zagury, R. S. Jope and E. Beurel 2016. Stress-induced  
669 neuroinflammation is mediated by GSK3-dependent TLR4 signaling that promotes susceptibility to depression-like  
670 behavior. Brain Behav Immun 53: 207-222. doi: 10.1016/j.bbi.2015.12.012

671 Cherry, J. D., J. A. Olschowka and M. K. O'Banion 2014. Neuroinflammation and M2 microglia: the good, the bad, and  
672 the inflamed. J Neuroinflammation 11: 98. doi: 10.1186/1742-2094-11-98

673 Chou, C. H., A. K. Chou, C. C. Lin, W. J. Chen, C. C. Wei, M. C. Yang, C. M. Hsu, F. W. Lung, J. K. Loh, S. L. Howng and  
674 Y. R. Hong 2012. GSK3β regulates Bcl2L12 and Bcl2L12A anti-apoptosis signaling in glioblastoma and is inhibited by  
675 LiCl. Cell Cycle 11: 532-542. doi: 10.4161/cc.11.3.19051

676 Cunningham, L. A., J. Newville, L. Li, P. Tapia, A. M. Allan and C. F. Valenzuela 2017. Prenatal Alcohol Exposure Leads  
677 to Enhanced Serine 9 Phosphorylation of Glycogen Synthase Kinase-3β (GSK-3 β ) in the Hippocampal Dentate Gyrus  
678 of Adult Mouse. Alcohol Clin Exp Res 41: 1907-1916. doi: 10.1111/acer.13489

679 Dandekar, M. P., S. S. Valvassori, G. C. Dal-Pont and J. Quevedo 2018. Glycogen Synthase Kinase-3β as a Putative  
680 Therapeutic Target for Bipolar Disorder. Curr Drug Metab 19: 663-673. doi: 10.2174/1389200219666171227203737

del Ser, T., K. C. Steinwachs, H. J. Gertz, M. V. Andrés, B. Gómez-Carrillo, M. Medina, J. A. Vericat, P. Redondo, D. Fleet and T. León 2013. Treatment of Alzheimer's disease with the GSK-3 inhibitor tideglusib: a pilot study. *J Alzheimers Dis* 33: 205-215. doi: 10.3233/jad-2012-120805

Duan, J., J. Cui, Z. Yang, C. Guo, J. Cao, M. Xi, Y. Weng, Y. Yin, Y. Wang, G. Wei, B. Qiao and A. Wen 2019. Neuroprotective effect of Apelin 13 on ischemic stroke by activating AMPK/GSK-3 $\beta$ /Nrf2 signaling. *J Neuroinflammation* 16: 24. doi: 10.1186/s12974-019-1406-7

Duan, X., D. M. Norris, S. J. Humphrey, P. Yang, K. C. Cooke, W. P. Bultitude, B. L. Parker, O. J. Conway, J. G. Burchfield, J. R. Krycer, F. M. Brodsky, D. E. James and D. J. Fazakerley 2022. Trafficking regulator of GLUT4-1 (TRARG1) is a GSK3 substrate. *Biochem J* 479: 1237-1256. doi: 10.1042/bcj20220153

Duda, P., S. M. Akula, S. L. Abrams, L. S. Steelman, A. M. Martelli, L. Cocco, S. Ratti, S. Candido, M. Libra, G. Montalto, M. Cervello, A. Gizak, D. Rakus and J. A. McCubrey 2020. Targeting GSK3 and Associated Signaling Pathways Involved in Cancer. *Cells* 9. doi: 10.3390/cells9051110

Duda, P., J. Wiśniewski, T. Wójtowicz, O. Wójcicka, M. Jaśkiewicz, D. Drulis-Fajdasz, D. Rakus, J. A. McCubrey and A. Gizak 2018. Targeting GSK3 signaling as a potential therapy of neurodegenerative diseases and aging. *Expert Opin Ther Targets* 22: 833-848. doi: 10.1080/14728222.2018.1526925

Duka, T., V. Duka, J. N. Joyce and A. Sidhu 2009. Alpha-Synuclein contributes to GSK-3 $\beta$ -catalyzed Tau phosphorylation in Parkinson's disease models. *FASEB J* 23: 2820-2830. doi: 10.1096/fj.08-120410

Eldar-Finkelman, H. and A. Martinez 2011. GSK-3 Inhibitors: Preclinical and Clinical Focus on CNS. *Front Mol Neurosci* 4: 32. doi: 10.3389/fnmol.2011.00032

Eldar-Finkelman, H. and A. M. VanHook 2016. Science Signaling Podcast for 15 November 2016: A new type of kinase inhibitor. *Sci Signal* 9: c22. doi: 10.1126/scisignal.aal2621

Emamian, E. S., D. Hall, M. J. Birnbaum, M. Karayiorgou and J. A. Gogos 2004. Convergent evidence for impaired AKT1-GSK3 $\beta$  signaling in schizophrenia. *Nat Genet* 36: 131-137. doi: 10.1038/ng1296

Emma, M. R., G. Augello, A. Cusimano, A. Azzolina, G. Montalto, J. A. McCubrey and M. Cervello 2020. GSK-3 in liver diseases: Friend or foe? *Biochim Biophys Acta Mol Cell Res* 1867: 118743. doi: 10.1016/j.bbamcr.2020.118743

Eom, T. Y. and R. S. Jope 2009. GSK3  $\beta$  N-terminus binding to p53 promotes its acetylation. *Mol Cancer* 8: 14. doi: 10.1186/1476-4598-8-14

Espinós, A., E. Fernández-Ortuño, E. Negri and V. Borrell 2022. Evolution of genetic mechanisms regulating cortical neurogenesis. *Dev Neurobiol* 82: 428-453. doi: 10.1002/dneu.22891

Fang, X. and T. M. Svitkina 2022. Adenomatous Polyposis Coli (APC) in cell migration. *Eur J Cell Biol* 101: 151228. doi: 10.1016/j.ejcb.2022.151228

Fei, X., Y. N. Dou, W. Lv, B. Ding, J. Wei, X. Wu, X. He, Z. Fei and F. Fei 2022. TLR4 Deletion Improves Cognitive Brain Function and Structure in Aged Mice. *Neuroscience* 492: 1-17. doi: 10.1016/j.neuroscience.2022.04.007

Gao, L., L. Yang and H. Cui 2021. GSK-3 $\beta$  inhibitor TWS119 alleviates hypoxic-ischemic brain damage via a crosstalk with Wnt and Notch signaling pathways in neonatal rats. *Brain Res* 1768: 147588. doi: 10.1016/j.brainres.2021.147588

Gao, S., S. Li, X. Duan, Z. Gu, Z. Ma, X. Yuan, X. Feng and H. Wang 2017. Inhibition of glycogen synthase kinase 3  $\beta$  (GSK3 $\beta$ ) suppresses the progression of esophageal squamous cell carcinoma by modifying STAT3 activity. *Mol Carcinog* 56: 2301-2316. doi: 10.1002/mc.22685

Golick, L., Y. Han, Y. Kim and S. W. Park 2018. BRD7 regulates the insulin-signaling pathway by increasing phosphorylation of GSK3 $\beta$ . *Cell Mol Life Sci* 75: 1857-1869. doi: 10.1007/s00018-017-2711-x

722 Golpich, M., E. Amini, F. Hemmati, N. M. Ibrahim, B. Rahmani, Z. Mohamed, A. A. Raymond, L. Dargahi, R. Ghasemi  
723 and A. Ahmadiani 2015. Glycogen synthase kinase-3 beta (GSK-3 $\beta$ ) signaling: Implications for Parkinson's disease.  
724 *Pharmacol Res* 97: 16-26. doi: 10.1016/j.phrs.2015.03.010

725 Gómez de Barreda, E., M. Pérez, P. Gómez Ramos, J. de Cristobal, P. Martín-Maestro, A. Morán, H. N. Dawson, M. P.  
726 Vitek, J. J. Lucas, F. Hernández and J. Avila 2010. Tau-knockout mice show reduced GSK3-induced hippocampal  
727 degeneration and learning deficits. *Neurobiol Dis* 37: 622-629. doi: 10.1016/j.nbd.2009.11.017

728 Götschel, F., C. Kern, S. Lang, T. Sparna, C. Markmann, J. Schwager, S. McNelly, F. von Weizsäcker, S. Laufer, A. Hecht  
729 and I. Merfort 2008. Inhibition of GSK3 differentially modulates NF-kappaB, CREB, AP-1 and beta-catenin signaling  
730 in hepatocytes, but fails to promote TNF-alpha-induced apoptosis. *Exp Cell Res* 314: 1351-1366. doi:  
731 10.1016/j.yexcr.2007.12.015

732 Green, H. F. and Y. M. Nolan 2012. Unlocking mechanisms in interleukin-1 $\beta$ -induced changes in hippocampal  
733 neurogenesis--a role for GSK-3 $\beta$  and TLX. *Transl Psychiatry* 2: e194. doi: 10.1038/tp.2012.117

734 Gupte, M., S. Tousif, J. J. Lemon, A. Toro Cora, P. Umbarkar and H. Lal 2022. Isoform-Specific Role of GSK-3 in High  
735 Fat Diet Induced Obesity and Glucose Intolerance. *Cells* 11. doi: 10.3390/cells11030559

736 Gupte, M., P. Umbarkar, A. P. Singh, Q. Zhang, S. Tousif and H. Lal 2020. Deletion of Cardiomyocyte Glycogen  
737 Synthase Kinase-3 Beta (GSK-3 $\beta$ ) Improves Systemic Glucose Tolerance with Maintained Heart Function in  
738 Established Obesity. *Cells* 9. doi: 10.3390/cells9051120

739 He, L., J. Endress, S. Cho, Z. Li, Y. Zheng, J. M. Asara and J. Blenis 2022. Suppression of nuclear GSK3 signaling  
740 promotes serine/one-carbon metabolism and confers metabolic vulnerability in lung cancer cells. *Sci Adv* 8:  
741 eabm8786. doi: 10.1126/sciadv.abm8786

742 Hermida, M. A., J. Dinesh Kumar and N. R. Leslie 2017. GSK3 and its interactions with the PI3K/AKT/mTOR signalling  
743 network. *Adv Biol Regul* 65: 5-15. doi: 10.1016/j.jbior.2017.06.003

744 Hernandez, F., J. J. Lucas and J. Avila 2013. GSK3 and tau: two convergence points in Alzheimer's disease. *J Alzheimers*  
745 *Dis* 33 Suppl 1: S141-144. doi: 10.3233/jad-2012-129025

746 Hoeflich, K. P., J. Luo, E. A. Rubie, M. S. Tsao, O. Jin and J. R. Woodgett 2000. Requirement for glycogen synthase  
747 kinase-3beta in cell survival and NF-kappaB activation. *Nature* 406: 86-90. doi: 10.1038/35017574

748 Hong, J. G., D. H. Kim, C. H. Lee, S. J. Park, J. M. Kim, M. Cai, D. S. Jang and J. H. Ryu 2012. GSK-3 $\beta$  activity in the  
749 hippocampus is required for memory retrieval. *Neurobiol Learn Mem* 98: 122-129. doi: 10.1016/j.nlm.2012.07.003

750 Hu, D., X. Sun, A. Magpusao, Y. Fedorov, M. Thompson, B. Wang, K. Lundberg, D. J. Adams and X. Qi 2021a. Small-  
751 molecule suppression of calpastatin degradation reduces neuropathology in models of Huntington's disease. *Nat*  
752 *Commun* 12: 5305. doi: 10.1038/s41467-021-25651-y

753 Hu, S., M. Hu, J. Liu, B. Zhang, Z. Zhang, F. H. Zhou, L. Wang and J. Dong 2020. Phosphorylation of Tau and  $\alpha$ -Synuclein  
754 Induced Neurodegeneration in MPTP Mouse Model of Parkinson's Disease. *Neuropsychiatr Dis Treat* 16: 651-663.  
755 doi: 10.2147/ndt.S235562

756 Hu, X. L., C. Guo, J. Q. Hou, J. H. Feng, X. Q. Zhang, F. Xiong, W. C. Ye and H. Wang 2019. Stereoisomers of Schisandrin  
757 B Are Potent ATP Competitive GSK-3 $\beta$  Inhibitors with Neuroprotective Effects against Alzheimer's Disease:  
758 Stereochemistry and Biological Activity. *ACS Chem Neurosci* 10: 996-1007. doi: 10.1021/acscchemneuro.8b00252

759 Hu, Y., X. Zhang, J. Zhang, X. Xia, H. Li, C. Qiu, Y. Liao, H. Chen, Z. He, Z. Song and W. Zhou 2021b. Activated STAT3  
760 signaling pathway by ligature-induced periodontitis could contribute to neuroinflammation and cognitive  
761 impairment in rats. *J Neuroinflammation* 18: 80. doi: 10.1186/s12974-021-02071-9

762 Huang, S., H. Wang, E. Turlova, A. Abussaud, X. Ji, L. R. Britto, S. P. Miller, A. Martinez, H. S. Sun and Z. P. Feng 2017.

763 GSK-3 $\beta$  inhibitor TDZD-8 reduces neonatal hypoxic-ischemic brain injury in mice. *CNS Neurosci Ther* 23: 405-415.  
764 doi: 10.1111/cns.12683

765 Hui, J., J. Zhang, M. Pu, X. Zhou, L. Dong, X. Mao, G. Shi, J. Zou, J. Wu, D. Jiang and G. Xi 2018. Modulation of GSK-  
766 3 $\beta$ / $\beta$ -Catenin Signaling Contributes to Learning and Memory Impairment in a Rat Model of Depression. *Int J*  
767 *Neuropsychopharmacol* 21: 858-870. doi: 10.1093/ijnp/pyy040

768 Iqbal, D., M. T. Rehman, M. F. Alajmi, M. Alsaweed, Q. M. S. Jamal, S. M. Alasiry, A. B. Albaker, M. Hamed, M. Kamal  
769 and H. M. Albadrani 2023. Multitargeted Virtual Screening and Molecular Simulation of Natural Product-like  
770 Compounds against GSK3 $\beta$ , NMDA-Receptor, and BACE-1 for the Management of Alzheimer's Disease.  
771 *Pharmaceuticals (Basel)* 16. doi: 10.3390/ph16040622

772 Ishizuka, K., A. Kamiya, E. C. Oh, H. Kanki, S. Seshadri, J. F. Robinson, H. Murdoch, A. J. Dunlop, K. Kubo, K. Furukori,  
773 B. Huang, M. Zeledon, A. Hayashi-Takagi, H. Okano, K. Nakajima, M. D. Houslay, N. Katsanis and A. Sawa 2011. DISC1-  
774 dependent switch from progenitor proliferation to migration in the developing cortex. *Nature* 473: 92-96. doi:  
775 10.1038/nature09859

776 Jankovic, J. and E. K. Tan 2020. Parkinson's disease: etiopathogenesis and treatment. *J Neurol Neurosurg Psychiatry*  
777 91: 795-808. doi: 10.1136/jnnp-2019-322338

778 Jian, Y., L. Kong, H. Xu, Y. Shi, X. Huang, W. Zhong, S. Huang, Y. Li, D. Shi, Y. Xiao, M. Yang, S. Li, X. Chen, Y. Ouyang,  
779 Y. Hu, X. Chen, L. Song, R. Ye and W. Wei 2022. Protein phosphatase 1 regulatory inhibitor subunit 14C promotes  
780 triple-negative breast cancer progression via sustaining inactive glycogen synthase kinase 3 beta. *Clin Transl Med*  
781 12: e725. doi: 10.1002/ctm2.725

782 Jiang, M., X. Zhang, X. Yan, S. Mizutani, H. Kashiwazaki, J. Ni and Z. Wu 2021. GSK3 $\beta$  is involved in promoting  
783 Alzheimer's disease pathologies following chronic systemic exposure to *Porphyromonas gingivalis*  
784 lipopolysaccharide in amyloid precursor protein(NL-F/NL-F) knock-in mice. *Brain Behav Immun* 98: 1-12. doi:  
785 10.1016/j.bbi.2021.08.213

786 Jope, R. S. and G. V. Johnson 2004. The glamour and gloom of glycogen synthase kinase-3. *Trends Biochem Sci* 29:  
787 95-102. doi: 10.1016/j.tibs.2003.12.004

788 Jope, R. S., C. J. Yuskaitis and E. Beurel 2007. Glycogen synthase kinase-3 (GSK3): inflammation, diseases, and  
789 therapeutics. *Neurochem Res* 32: 577-595. doi: 10.1007/s11064-006-9128-5

790 Jurcau, A. and A. Simion 2021. Neuroinflammation in Cerebral Ischemia and Ischemia/Reperfusion Injuries: From  
791 Pathophysiology to Therapeutic Strategies. *Int J Mol Sci* 23. doi: 10.3390/ijms23010014

792 Kalinichev, M. and L. A. Dawson 2011. Evidence for antimanic efficacy of glycogen synthase kinase-3 (GSK3) inhibitors  
793 in a strain-specific model of acute mania. *Int J Neuropsychopharmacol* 14: 1051-1067. doi:  
794 10.1017/s1461145710001495

795 Kawakami, F., M. Suzuki, N. Shimada, G. Kagiya, E. Ohta, K. Tamura, H. Maruyama and T. Ichikawa 2011. Stimulatory  
796 effect of  $\alpha$ -synuclein on the tau-phosphorylation by GSK-3 $\beta$ . *Febs j* 278: 4895-4904. doi: 10.1111/j.1742-  
797 4658.2011.08389.x

798 King, T. D., B. Clodfelder-Miller, K. A. Barksdale and G. N. Bijur 2008. Unregulated mitochondrial GSK3beta activity  
799 results in NADH: ubiquinone oxidoreductase deficiency. *Neurotox Res* 14: 367-382. doi: 10.1007/bf03033861

800 Kisoh, K., H. Hayashi, T. Itoh, M. Asada, M. Arai, B. Yuan, K. Tanonaka and N. Takagi 2017. Involvement of GSK-3 $\beta$   
801 Phosphorylation Through PI3-K/Akt in Cerebral Ischemia-Induced Neurogenesis in Rats. *Mol Neurobiol* 54: 7917-  
802 7927. doi: 10.1007/s12035-016-0290-8

803 Ko, R. and S. Y. Lee 2016. Glycogen synthase kinase 3 $\beta$  in Toll-like receptor signaling. *BMB Rep* 49: 305-310. doi:

10.5483/bmbrep.2016.49.6.059

Ko, R., H. Park, N. Lee, J. Seo, W. Jeong and S. Y. Lee 2019. Glycogen Synthase Kinase 3 $\beta$  Regulates Antiviral Responses of TLR3 via TRAF2-Src Axis. *J Immunol* 203: 2990-2999. doi: 10.4049/jimmunol.1900685

Ko, R., J. H. Park, H. Ha, Y. Choi and S. Y. Lee 2015. Glycogen synthase kinase 3 $\beta$  ubiquitination by TRAF6 regulates TLR3-mediated pro-inflammatory cytokine production. *Nat Commun* 6: 6765. doi: 10.1038/ncomms7765

Koehler, D., Z. A. Shah and F. E. Williams 2019. The GSK3 $\beta$  inhibitor, TDZD-8, rescues cognition in a zebrafish model of okadaic acid-induced Alzheimer's disease. *Neurochem Int* 122: 31-37. doi: 10.1016/j.neuint.2018.10.022

Koike, R., Y. Takaichi, Y. Soeda and A. Takashima 2021. Memory formation in old age requires GSK-3 $\beta$ . *Aging Brain* 1: 100022. doi: 10.1016/j.nbas.2021.100022

Kotliarova, S., S. Pastorino, L. C. Kovell, Y. Kotliarov, H. Song, W. Zhang, R. Bailey, D. Maric, J. C. Zenklusen, J. Lee and H. A. Fine 2008. Glycogen synthase kinase-3 inhibition induces glioma cell death through c-MYC, nuclear factor-kappaB, and glucose regulation. *Cancer Res* 68: 6643-6651. doi: 10.1158/0008-5472.Can-08-0850

Ku, B. M., Y. K. Lee, J. Y. Jeong, J. Ryu, J. Choi, J. S. Kim, Y. W. Cho, G. S. Roh, H. J. Kim, G. J. Cho, W. S. Choi and S. S. Kang 2011. Caffeine inhibits cell proliferation and regulates PKA/GSK3 $\beta$  pathways in U87MG human glioma cells. *Mol Cells* 31: 275-279. doi: 10.1007/s10059-011-0027-5

Lakshmanan, J., B. Zhang, I. C. Nweze, Y. Du and B. G. Harbrecht 2015. Glycogen synthase kinase 3 regulates IL-1 $\beta$  mediated iNOS expression in hepatocytes by down-regulating c-Jun. *J Cell Biochem* 116: 133-141. doi: 10.1002/jcb.24951

Latapy, C., V. Rioux, M. J. Guitton and J. M. Beaulieu 2012. Selective deletion of forebrain glycogen synthase kinase 3 $\beta$  reveals a central role in serotonin-sensitive anxiety and social behaviour. *Philos Trans R Soc Lond B Biol Sci* 367: 2460-2474. doi: 10.1098/rstb.2012.0094

Lauretti, E., O. Dincer and D. Praticò 2020. Glycogen synthase kinase-3 signaling in Alzheimer's disease. *Biochim Biophys Acta Mol Cell Res* 1867: 118664. doi: 10.1016/j.bbamcr.2020.118664

Lee, S. J., Y. H. Chung, K. M. Joo, H. C. Lim, G. S. Jeon, D. Kim, W. B. Lee, Y. S. Kim and C. I. Cha 2006. Age-related changes in glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) immunoreactivity in the central nervous system of rats. *Neurosci Lett* 409: 134-139. doi: 10.1016/j.neulet.2006.09.026

Leucht, S., W. Kissling and J. McGrath 2007. Lithium for schizophrenia. *Cochrane Database Syst Rev*: Cd003834. doi: 10.1002/14651858.CD003834.pub2

Li, C. H., C. W. Liu, C. H. Tsai, Y. J. Peng, Y. H. Yang, P. L. Liao, C. C. Lee, Y. W. Cheng and J. J. Kang 2017. Cytoplasmic aryl hydrocarbon receptor regulates glycogen synthase kinase 3  $\beta$ , accelerates vimentin degradation, and suppresses epithelial-mesenchymal transition in non-small cell lung cancer cells. *Arch Toxicol* 91: 2165-2178. doi: 10.1007/s00204-016-1870-0

Li, H., J. Li, G. Zhang, Q. Da, L. Chen, S. Yu, Q. Zhou, Z. Weng, Z. Xin, L. Shi, L. Ma, A. Huang, S. Qi and Y. Lu 2019. HMGB1-Induced p62 Overexpression Promotes Snail-Mediated Epithelial-Mesenchymal Transition in Glioblastoma Cells via the Degradation of GSK-3 $\beta$ . *Theranostics* 9: 1909-1922. doi: 10.7150/thno.30578

Li, Q., J. Wu, L. Huang, B. Zhao and Q. Li 2021. Ephedrine ameliorates cerebral ischemia injury via inhibiting NOD-like receptor pyrin domain 3 inflammasome activation through the Akt/GSK3 $\beta$ /NRF2 pathway. *Hum Exp Toxicol* 40: S540-S552. doi: 10.1177/096032712111052981

Li, Y., H. Fan, M. Ni, W. Zhang, F. Fang, J. Sun, P. Lyu and P. Ma 2022. Etanercept Reduces Neuron Injury and Neuroinflammation via Inactivating c-Jun N-terminal Kinase and Nuclear Factor- $\kappa$ B Pathways in Alzheimer's Disease: An In Vitro and In Vivo Investigation. *Neuroscience* 484: 140-150. doi: 10.1016/j.neuroscience.2021.11.001

Li, Y., H. Lu, Y. Huang, R. Xiao, X. Cai, S. He and G. Yan 2010. Glycogen synthase kinases-3 $\beta$  controls differentiation of malignant glioma cells. *Int J Cancer* 127: 1271-1282. doi: 10.1002/ijc.25020

Li, Y., J. Zhu, Y. Liu, X. Chen, S. Lei, L. Li, B. Jiang, L. Tan, S. Yu and Y. Zhao 2016. Glycogen Synthase Kinase 3 $\beta$  Influences Injury Following Cerebral Ischemia/Reperfusion in Rats. *Int J Biol Sci* 12: 518-531. doi: 10.7150/ijbs.13918

Liang, M. H. and D. M. Chuang 2007. Regulation and function of glycogen synthase kinase-3 isoforms in neuronal survival. *J Biol Chem* 282: 3904-3917. doi: 10.1074/jbc.M605178200

Liang, T., H. Ju, Y. Zhou, Y. Yang, Y. Shi and H. Fang 2020. Inhibition of glycogen synthase kinase 3 $\beta$  improves cognitive function in aged mice by upregulating claudin presences in cerebral endothelial cells. *Acta Biochim Biophys Sin (Shanghai)* 52: 363-370. doi: 10.1093/abbs/gmaa002

Liao, S., J. Wu, R. Liu, S. Wang, J. Luo, Y. Yang, Y. Qin, T. Li, X. Zheng, J. Song, X. Zhao, C. Xiao, Y. Zhang, L. Bian, P. Jia, Y. Bai and X. Zheng 2020. A novel compound DBZ ameliorates neuroinflammation in LPS-stimulated microglia and ischemic stroke rats: Role of Akt(Ser473)/GSK3 $\beta$ (Ser9)-mediated Nrf2 activation. *Redox Biol* 36: 101644. doi: 10.1016/j.redox.2020.101644

Lin, J., T. Song, C. Li and W. Mao 2020. GSK-3 $\beta$  in DNA repair, apoptosis, and resistance of chemotherapy, radiotherapy of cancer. *Biochim Biophys Acta Mol Cell Res* 1867: 118659. doi: 10.1016/j.bbamcr.2020.118659

Liu, E., A. J. Xie, Q. Zhou, M. Li, S. Zhang, S. Li, W. Wang, X. Wang, Q. Wang and J. Z. Wang 2017. GSK-3 $\beta$  deletion in dentate gyrus excitatory neuron impairs synaptic plasticity and memory. *Sci Rep* 7: 5781. doi: 10.1038/s41598-017-06173-4

Liu, F., N. Tian, H. Q. Zhang, S. H. Li, Q. Z. Zhou, Y. Yang, J. Zheng and J. Z. Wang 2020a. GSK-3 $\beta$  activation accelerates early-stage consumption of Hippocampal Neurogenesis in senescent mice. *Theranostics* 10: 9674-9685. doi: 10.7150/thno.43829

Liu, H., X. Wu, J. Luo, L. Zhao, X. Li, H. Guo, H. Bai, W. Cui, W. Guo, D. Feng and Y. Qu 2020b. Adiponectin peptide alleviates oxidative stress and NLRP3 inflammasome activation after cerebral ischemia-reperfusion injury by regulating AMPK/GSK-3 $\beta$ . *Exp Neurol* 329: 113302. doi: 10.1016/j.expneurol.2020.113302

Luo, J. 2009. Glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) in tumorigenesis and cancer chemotherapy. *Cancer Lett* 273: 194-200. doi: 10.1016/j.canlet.2008.05.045

Majewska, E. and M. Szeliga 2017. AKT/GSK3 $\beta$  Signaling in Glioblastoma. *Neurochem Res* 42: 918-924. doi: 10.1007/s11064-016-2044-4

Marineau, A., K. A. Khan and M. J. Servant 2020. Roles of GSK-3 and  $\beta$ -Catenin in Antiviral Innate Immune Sensing of Nucleic Acids. *Cells* 9. doi: 10.3390/cells9040897

Martínez-González, L., C. Gonzalo-Consuegra, M. Gómez-Almería, G. Porras, E. de Lago, Á Martín-Requero and A. Martínez 2021. Tideglusib, a Non-ATP Competitive Inhibitor of GSK-3 $\beta$  as a Drug Candidate for the Treatment of Amyotrophic Lateral Sclerosis. *Int J Mol Sci* 22. doi: 10.3390/ijms22168975

Miyashita, K., K. Kawakami, M. Nakada, W. Mai, A. Shakoori, H. Fujisawa, Y. Hayashi, J. Hamada and T. Minamoto 2009. Potential therapeutic effect of glycogen synthase kinase 3 $\beta$  inhibition against human glioblastoma. *Clin Cancer Res* 15: 887-897. doi: 10.1158/1078-0432.Ccr-08-0760

Moore, S. F., E. O. Agbani, A. Wersäll, A. W. Poole, C. M. Williams, X. Zhao, Y. Li, J. L. Hutchinson, R. W. Hunter and I. Hers 2021. Opposing Roles of GSK3 $\alpha$  and GSK3 $\beta$  Phosphorylation in Platelet Function and Thrombosis. *Int J Mol Sci* 22. doi: 10.3390/ijms221910656

Moore, S. F., M. T. van den Bosch, R. W. Hunter, K. Sakamoto, A. W. Poole and I. Hers 2013. Dual regulation of glycogen synthase kinase 3 (GSK3) $\alpha/\beta$  by protein kinase C (PKC) $\alpha$  and Akt promotes thrombin-mediated integrin

886  $\alpha$ IIb $\beta$ 3 activation and granule secretion in platelets. *J Biol Chem* 288: 3918-3928. doi: 10.1074/jbc.M112.429936

887 Morales-Garcia, J. A., R. Luna-Medina, S. Alonso-Gil, M. Sanz-Sancristobal, V. Palomo, C. Gil, A. Santos, A. Martinez

888 and A. Perez-Castillo 2012. Glycogen synthase kinase 3 inhibition promotes adult hippocampal neurogenesis in vitro

889 and in vivo. *ACS Chem Neurosci* 3: 963-971. doi: 10.1021/cn300110c

890 Moretti, D. V. 2015. Are there treatments for atypical parkinsonism? An update on actual options. *Rev Neurosci* 26:

891 547-553. doi: 10.1515/revneuro-2015-0008

892 Morgan-Smith, M., Y. Wu, X. Zhu, J. Pringle and W. D. Snider 2014. GSK-3 signaling in developing cortical neurons is

893 essential for radial migration and dendritic orientation. *Elife* 3: e02663. doi: 10.7554/eLife.02663

894 Mowry, F. E., S. C. Peadar, J. E. Stern and V. C. Biancardi 2021. TLR4 and AT1R mediate blood-brain barrier disruption,

895 neuroinflammation, and autonomic dysfunction in spontaneously hypertensive rats. *Pharmacol Res* 174: 105877.

896 doi: 10.1016/j.phrs.2021.105877

897 Murase, S., E. Mosser and E. M. Schuman 2002. Depolarization drives beta-Catenin into neuronal spines promoting

898 changes in synaptic structure and function. *Neuron* 35: 91-105. doi: 10.1016/s0896-6273(02)00764-x

899 Mutch, C. A., N. Funatsu, E. S. Monuki and A. Chenn 2009. Beta-catenin signaling levels in progenitors influence the

900 laminar cell fates of projection neurons. *J Neurosci* 29: 13710-13719. doi: 10.1523/jneurosci.3022-09.2009

901 Nagini, S., J. Sophia and R. Mishra 2019. Glycogen synthase kinases: Moonlighting proteins with theranostic potential

902 in cancer. *Semin Cancer Biol* 56: 25-36. doi: 10.1016/j.semcancer.2017.12.010

903 O'Brien, W. T., A. D. Harper, F. Jové, J. R. Woodgett, S. Maretto, S. Piccolo and P. S. Klein 2004. Glycogen synthase

904 kinase-3 $\beta$  haploinsufficiency mimics the behavioral and molecular effects of lithium. *J Neurosci* 24: 6791-6798.

905 doi: 10.1523/jneurosci.4753-03.2004

906 Ochoa, E. L. M. 2022. Lithium as a Neuroprotective Agent for Bipolar Disorder: An Overview. *Cell Mol Neurobiol* 42:

907 85-97. doi: 10.1007/s10571-021-01129-9

908 Okun, E., K. Griffioen, B. Barak, N. J. Roberts, K. Castro, M. A. Pita, A. Cheng, M. R. Mughal, R. Wan, U. Ashery and M.

909 P. Mattson 2010. Toll-like receptor 3 inhibits memory retention and constrains adult hippocampal neurogenesis.

910 *Proc Natl Acad Sci U S A* 107: 15625-15630. doi: 10.1073/pnas.1005807107

911 Palomo, V., D. I. Perez, C. Perez, J. A. Morales-Garcia, I. Soteras, S. Alonso-Gil, A. Encinas, A. Castro, N. E. Campillo,

912 A. Perez-Castillo, C. Gil and A. Martinez 2012. 5-imino-1,2,4-thiadiazoles: first small molecules as substrate

913 competitive inhibitors of glycogen synthase kinase 3. *J Med Chem* 55: 1645-1661. doi: 10.1021/jm201463v

914 Pan, H. Y. and M. Valapala 2022. Regulation of Autophagy by the Glycogen Synthase Kinase-3 (GSK-3) Signaling

915 Pathway. *Int J Mol Sci* 23. doi: 10.3390/ijms23031709

916 Pang, T., Y. J. Wang, Y. X. Gao, Y. Xu, Q. Li, Y. B. Zhou, L. Xu, Z. J. Huang, H. Liao, L. Y. Zhang, J. R. Gao, Q. Ye and J. Li

917 2016. A novel GSK-3 $\beta$  inhibitor YQ138 prevents neuronal injury induced by glutamate and brain ischemia through

918 activation of the Nrf2 signaling pathway. *Acta Pharmacol Sin* 37: 741-752. doi: 10.1038/aps.2016.3

919 Parvez, S., B. Ramachandran, M. Kaushik, H. Tabassum and J. U. Frey 2023. Long-term depression induction and

920 maintenance across regions of the apical branch of CA1 dendrites. *Hippocampus* 33: 1058-1066. doi:

921 10.1002/hipo.23553

922 Patel, S. and G. H. Werstuck 2021. Macrophage Function and the Role of GSK3. *Int J Mol Sci* 22. doi:

923 10.3390/ijms22042206

924 Pecoraro, C., B. Faggion, B. Balboni, D. Carbone, G. J. Peters, P. Diana, Y. G. Assaraf and E. Giovannetti 2021. GSK3 $\beta$

925 as a novel promising target to overcome chemoresistance in pancreatic cancer. *Drug Resist Updat* 58: 100779. doi:

926 10.1016/j.drug.2021.100779



Pei, J. J., T. Tanaka, Y. C. Tung, E. Braak, K. Iqbal and I. Grundke-Iqbal 1997. Distribution, levels, and activity of glycogen synthase kinase-3 in the Alzheimer disease brain. *J Neuropathol Exp Neurol* 56: 70-78. doi: 10.1097/00005072-199701000-00007

Peineau, S., C. Taghibiglou, C. Bradley, T. P. Wong, L. Liu, J. Lu, E. Lo, D. Wu, E. Saule, T. Bouchet, P. Matthews, J. T. Isaac, Z. A. Bortolotto, Y. T. Wang and G. L. Collingridge 2007. LTP inhibits LTD in the hippocampus via regulation of GSK3 $\beta$ . *Neuron* 53: 703-717. doi: 10.1016/j.neuron.2007.01.029

Peng, S., J. H. Gu, C. L. Dai, K. Iqbal, F. Liu and C. X. Gong 2022. AKT/GSK-3 $\beta$  signaling is altered through downregulation of mTOR during cerebral Ischemia/Reperfusion injury. *Mol Biol Rep* 49: 3955-3964. doi: 10.1007/s11033-022-07247-x

Polter, A., E. Beurel, S. Yang, R. Garner, L. Song, C. A. Miller, J. D. Sweatt, L. McMahon, A. A. Bartolucci, X. Li and R. S. Jope 2010. Deficiency in the inhibitory serine-phosphorylation of glycogen synthase kinase-3 increases sensitivity to mood disturbances. *Neuropsychopharmacology* 35: 1761-1774. doi: 10.1038/npp.2010.43

Prickaerts, J., D. Moechars, K. Cryns, I. Lenaerts, H. van Craenendonck, I. Goris, G. Daneels, J. A. Bouwknecht and T. Steckler 2006. Transgenic mice overexpressing glycogen synthase kinase 3 $\beta$ : a putative model of hyperactivity and mania. *J Neurosci* 26: 9022-9029. doi: 10.1523/jneurosci.5216-05.2006

Qing, H., G. He, P. T. Ly, C. J. Fox, M. Staufenbiel, F. Cai, Z. Zhang, S. Wei, X. Sun, C. H. Chen, W. Zhou, K. Wang and W. Song 2008. Valproic acid inhibits A $\beta$  production, neuritic plaque formation, and behavioral deficits in Alzheimer's disease mouse models. *J Exp Med* 205: 2781-2789. doi: 10.1084/jem.20081588

Redondo, M., V. Palomo, J. Brea, D. I. Pérez, R. Martín-Álvarez, C. Pérez, N. Paúl-Fernández, S. Conde, M. I. Cadavid, M. I. Loza, G. Mengod, A. Martínez, C. Gil and N. E. Campillo 2012. Identification in silico and experimental validation of novel phosphodiesterase 7 inhibitors with efficacy in experimental autoimmune encephalomyelitis mice. *ACS Chem Neurosci* 3: 793-803. doi: 10.1021/cn300105c

Reich, S. G. and J. M. Savitt 2019. Parkinson's Disease. *Med Clin North Am* 103: 337-350. doi: 10.1016/j.mcna.2018.10.014

Rippin, I., K. Bonder, S. Joseph, A. Sarsor, L. Vaks and H. Eldar-Finkelman 2021. Inhibition of GSK-3 ameliorates the pathogenesis of Huntington's disease. *Neurobiol Dis* 154: 105336. doi: 10.1016/j.nbd.2021.105336

Roberson, E. D., K. Scarce-Levie, J. J. Palop, F. Yan, I. H. Cheng, T. Wu, H. Gerstein, G. Q. Yu and L. Mucke 2007. Reducing endogenous tau ameliorates amyloid  $\beta$ -induced deficits in an Alzheimer's disease mouse model. *Science* 316: 750-754. doi: 10.1126/science.1141736

Rojanathammanee, L., A. M. Floden, G. D. Manocha and C. K. Combs 2015. Attenuation of microglial activation in a mouse model of Alzheimer's disease via NFAT inhibition. *J Neuroinflammation* 12: 42. doi: 10.1186/s12974-015-0255-2

Rosa, A. O., M. P. Kaster, R. W. Binfaré, S. Morales, E. Martín-Aparicio, M. L. Navarro-Rico, A. Martinez, M. Medina, A. G. García, M. G. López and A. L. Rodrigues 2008. Antidepressant-like effect of the novel thiadiazolidinone NP031115 in mice. *Prog Neuropsychopharmacol Biol Psychiatry* 32: 1549-1556. doi: 10.1016/j.pnpbp.2008.05.020

Samim Khan, S., S. Janrao, S. Srivastava, S. Bala Singh, L. Vora and D. Kumar Khatri 2023. GSK-3 $\beta$ : An exuberating neuroinflammatory mediator in Parkinson's disease. *Biochem Pharmacol* 210: 115496. doi: 10.1016/j.bcp.2023.115496

Scarselli, M. 2023. Lithium and Valproic Acid in Bipolar Disorders and Beyond. *Curr Neuropharmacol* 21: 890. doi: 10.2174/1570159x2104230307123319

Schilling, S., B. Chausse, H. O. Dikmen, F. Almouhanna, J. O. Hollnagel, A. Lewen and O. Kann 2021. TLR2- and TLR3-

968 activated microglia induce different levels of neuronal network dysfunction in a context-dependent manner. *Brain*  
 969 *Behav Immun* 96: 80-91. doi: 10.1016/j.bbi.2021.05.013

970 Sharma, V. K. and T. G. Singh 2020. CREB: A Multifaceted Target for Alzheimer's Disease. *Curr Alzheimer Res* 17:  
 971 1280-1293. doi: 10.2174/1567205018666210218152253

972 Singh, S. S., S. N. Rai, H. Birla, W. Zahra, A. S. Rathore and S. P. Singh 2020. NF- $\kappa$ B-Mediated Neuroinflammation in  
 973 Parkinson's Disease and Potential Therapeutic Effect of Polyphenols. *Neurotox Res* 37: 491-507. doi:  
 974 10.1007/s12640-019-00147-2

975 Smith, A. R., J. Mill and K. Lunnon 2020. The molecular etiology of Alzheimer's disease. *Brain Pathol* 30: 964-965. doi:  
 976 10.1111/bpa.12879

977 Taelman, V. F., R. Dobrowolski, J. L. Plouhinec, L. C. Fuentealba, P. P. Vorwald, I. Gumper, D. D. Sabatini and E. M. De  
 978 Robertis 2010. Wnt signaling requires sequestration of glycogen synthase kinase 3 inside multivesicular endosomes.  
 979 *Cell* 143: 1136-1148. doi: 10.1016/j.cell.2010.11.034

980 Takahashi-Yanaga, F. 2013. Activator or inhibitor? GSK-3 as a new drug target. *Biochem Pharmacol* 86: 191-199. doi:  
 981 10.1016/j.bcp.2013.04.022

982 Takashima, A. 2012. GSK-3 $\beta$  and memory formation. *Front Mol Neurosci* 5: 47. doi: 10.3389/fnmol.2012.00047

983 Tang, H., N. Shen, H. Jin, D. Liu, X. Miao and L. Q. Zhu 2013. GSK-3 $\beta$  polymorphism discriminates bipolar disorder and  
 984 schizophrenia: a systematic meta-analysis. *Mol Neurobiol* 48: 404-411. doi: 10.1007/s12035-013-8414-x

985 Terwel, D., D. Muyliaert, I. Dewachter, P. Borghgraef, S. Croes, H. Devijver and F. Van Leuven 2008. Amyloid activates  
 986 GSK-3 $\beta$  to aggravate neuronal tauopathy in bigenic mice. *Am J Pathol* 172: 786-798. doi:  
 987 10.2353/ajpath.2008.070904

988 Thornton, T. M., G. Pedraza-Alva, B. Deng, C. D. Wood, A. Aronshtam, J. L. Clements, G. Sabio, R. J. Davis, D. E.  
 989 Matthews, B. Doble and M. Rincon 2008. Phosphorylation by p38 MAPK as an alternative pathway for GSK3 $\beta$   
 990 inactivation. *Science* 320: 667-670. doi: 10.1126/science.1156037

991 Vallée, A., Y. Lecarpentier, R. Guillevin and J. N. Vallée 2018. Thermodynamics in Neurodegenerative Diseases:  
 992 Interplay Between Canonical WNT/Beta-Catenin Pathway-PPAR Gamma, Energy Metabolism and Circadian Rhythms.  
 993 *Neuromolecular Med* 20: 174-204. doi: 10.1007/s12017-018-8486-x

994 Vieta, E., M. Berk, T. G. Schulze, A. F. Carvalho, T. Suppes, J. R. Calabrese, K. Gao, K. W. Miskowiak and I. Grande  
 995 2018. Bipolar disorders. *Nat Rev Dis Primers* 4: 18008. doi: 10.1038/nrdp.2018.8

996 Voet, S., S. Srinivasan, M. Lamkanfi and G. van Loo 2019. Inflammasomes in neuroinflammatory and  
 997 neurodegenerative diseases. *EMBO Mol Med* 11. doi: 10.15252/emmm.201810248

998 Wang, F., L. Wang, G. Sui, C. Yang, M. Guo, X. Xiong, Z. Chen and P. Lei 2021. IGF-1 Alleviates Mitochondrial Apoptosis  
 999 through the GSK3 $\beta$ /NF- $\kappa$ B/NLRP3 Signaling Pathway in LPS-Treated PC-12 Cells. *J Mol Neurosci* 71: 1320-1328. doi:  
 1000 10.1007/s12031-020-01759-6

1001 Wang, H., S. Huang, K. Yan, X. Fang, A. Abussaud, A. Martinez, H. S. Sun and Z. P. Feng 2016. Tideglusib, a chemical  
 1002 inhibitor of GSK3 $\beta$ , attenuates hypoxic-ischemic brain injury in neonatal mice. *Biochim Biophys Acta* 1860: 2076-  
 1003 2085. doi: 10.1016/j.bbagen.2016.06.027

1004 Wang, W., M. Li, Y. Wang, Z. Wang, W. Zhang, F. Guan, Q. Chen and J. Wang 2017. GSK-3 $\beta$  as a target for protection  
 1005 against transient cerebral ischemia. *Int J Med Sci* 14: 333-339. doi: 10.7150/ijms.17514

1006 Wang, Y., C. Meng, J. Zhang, J. Wu and J. Zhao 2019. Inhibition of GSK-3 $\beta$  alleviates cerebral ischemia/reperfusion  
 1007 injury in rats by suppressing NLRP3 inflammasome activation through autophagy. *Int Immunopharmacol* 68: 234-  
 1008 241. doi: 10.1016/j.intimp.2018.12.042

Willi, R., A. Harmer, S. Giovanoli and U. Meyer 2013. Altered GSK3 $\beta$  signaling in an infection-based mouse model of developmental neuropsychiatric disease. *Neuropharmacology* 73: 56-65. doi: 10.1016/j.neuropharm.2013.05.012

Wills, J., J. Jones, T. Haggerty, V. Duka, J. N. Joyce and A. Sidhu 2010. Elevated tauopathy and alpha-synuclein pathology in postmortem Parkinson's disease brains with and without dementia. *Exp Neurol* 225: 210-218. doi: 10.1016/j.expneurol.2010.06.017

Xie, Y., Y. Zhang, T. Hu, Z. Zhao, Q. Liu and H. Li 2022. Inhibition of Glycogen Synthase Kinase 3 $\beta$  Activity in the Basolateral Amygdala Disrupts Reconsolidation and Attenuates Heroin Relapse. *Front Mol Neurosci* 15: 932939. doi: 10.3389/fnmol.2022.932939

Xiong, Y. S., D. L. Wang, L. Tan, X. Wang, L. M. Chen, C. X. Gong, J. Z. Wang and L. Q. Zhu 2013. Inhibition of glycogen synthase kinase-3 reverses tau hyperphosphorylation induced by Pin1 down-regulation. *CNS Neurol Disord Drug Targets* 12: 436-443. doi: 10.2174/1871527311312030016

Xu, B., J. Xu, N. Cai, M. Li, L. Liu, Y. Qin, X. Li and H. Wang 2021. Roflumilast prevents ischemic stroke-induced neuronal damage by restricting GSK3 $\beta$ -mediated oxidative stress and IRE1 $\alpha$ /TRAF2/JNK pathway. *Free Radic Biol Med* 163: 281-296. doi: 10.1016/j.freeradbiomed.2020.12.018

Yang, L., L. Chen, Y. Fang and S. Ma 2021. Downregulation of GSK-3 $\beta$  Expression via Ultrasound-Targeted Microbubble Destruction Enhances Atherosclerotic Plaque Stability in New Zealand Rabbits. *Ultrasound Med Biol* 47: 710-722. doi: 10.1016/j.ultrasmedbio.2020.11.002

Yang, W., P. F. Wu, J. X. Ma, M. J. Liao, X. H. Wang, L. S. Xu, M. H. Xu and L. Yi 2019. Sortilin promotes glioblastoma invasion and mesenchymal transition through GSK-3 $\beta$ / $\beta$ -catenin/twist pathway. *Cell Death Dis* 10: 208. doi: 10.1038/s41419-019-1449-9

Yang, Y., Q. Q. Wang, O. Bozinov, R. X. Xu, Y. L. Sun and S. S. Wang 2020. GSK-3 inhibitor CHIR99021 enriches glioma stem-like cells. *Oncol Rep* 43: 1479-1490. doi: 10.3892/or.2020.7525

Yokota, Y., W. Y. Kim, Y. Chen, X. Wang, A. Stanco, Y. Komuro, W. Snider and E. S. Anton 2009. The adenomatous polyposis coli protein is an essential regulator of radial glial polarity and construction of the cerebral cortex. *Neuron* 61: 42-56. doi: 10.1016/j.neuron.2008.10.053

Young, W. 2009. Review of lithium effects on brain and blood. *Cell Transplant* 18: 951-975. doi: 10.3727/096368909x471251

Zakharova, I. O., T. V. Sokolova, L. V. Bayunova, Zorina, II, M. P. Rychkova, A. O. Shpakov and N. F. Avrova 2019. The Protective Effect of Insulin on Rat Cortical Neurons in Oxidative Stress and Its Dependence on the Modulation of Akt, GSK-3 $\beta$ , ERK1/2, and AMPK Activities. *Int J Mol Sci* 20. doi: 10.3390/ijms20153702

Zamanpoor, M. 2020. Schizophrenia in a genomic era: a review from the pathogenesis, genetic and environmental etiology to diagnosis and treatment insights. *Psychiatr Genet* 30: 1-9. doi: 10.1097/ypg.0000000000000245

Zang, J., Y. Liu, W. Li, D. Xiao, Y. Zhang, Y. Luo, W. Liang, F. Liu and W. Wei 2017. Voluntary exercise increases adult hippocampal neurogenesis by increasing GSK-3 $\beta$  activity in mice. *Neuroscience* 354: 122-135. doi: 10.1016/j.neuroscience.2017.04.024

Zechner, D., Y. Fujita, J. Hülken, T. Müller, I. Walther, M. M. Taketo, E. B. Crenshaw, 3rd, W. Birchmeier and C. Birchmeier 2003.  $\beta$ -Catenin signals regulate cell growth and the balance between progenitor cell expansion and differentiation in the nervous system. *Dev Biol* 258: 406-418. doi: 10.1016/s0012-1606(03)00123-4

Zhang, Q., Y. Li, J. Yu, C. Yin, J. Guo, J. Zhao and Q. Wang 2022a. TLR3 deletion inhibits programmed necrosis of brain cells in neonatal mice with sevoflurane-induced cognitive dysfunction. *Aging (Albany NY)* 14: 4714-4727. doi: 10.18632/aging.204092

1050 Zhang, S., W. Gao, J. Tang, H. Zhang, Y. Zhou, J. Liu, K. Chen, F. Liu, W. Li, S. K. Y. To, A. S. T. Wong, X. K. Zhang, H.  
1051 Zhou and J. Z. Zeng 2020. The Roles of GSK-3 $\beta$  in Regulation of Retinoid Signaling and Sorafenib Treatment Response  
1052 in Hepatocellular Carcinoma. *Theranostics* 10: 1230-1244. doi: 10.7150/thno.38711

1053 Zhang, X., X. Heng, T. Li, L. Li, D. Yang, X. Zhang, Y. Du, R. S. Doody and W. Le 2011. Long-term treatment with lithium  
1054 alleviates memory deficits and reduces amyloid- $\beta$  production in an aged Alzheimer's disease transgenic mouse  
1055 model. *J Alzheimers Dis* 24: 739-749. doi: 10.3233/jad-2011-101875

1056 Zhang, X. M., M. Yin and M. H. Zhang 2014. Cell-based assays for Parkinson's disease using differentiated human  
1057 LUHMES cells. *Acta Pharmacol Sin* 35: 945-956. doi: 10.1038/aps.2014.36

1058 Zhang, Y., N. Q. Huang, F. Yan, H. Jin, S. Y. Zhou, J. S. Shi and F. Jin 2018. Diabetes mellitus and Alzheimer's disease:  
1059 GSK-3 $\beta$  as a potential link. *Behav Brain Res* 339: 57-65. doi: 10.1016/j.bbr.2017.11.015

1060 Zhang, Y., Z. Wu, Z. Huang, Y. Liu, X. Chen, X. Zhao, H. He and Y. Deng 2022b. GSK-3 $\beta$  inhibition elicits a  
1061 neuroprotection by restoring lysosomal dysfunction in neurons via facilitation of TFEB nuclear translocation after  
1062 ischemic stroke. *Brain Res* 1778: 147768. doi: 10.1016/j.brainres.2021.147768

1063 Zhao, P., Q. Li, Z. Shi, C. Li, L. Wang, X. Liu, C. Jiang, X. Qian, Y. You, N. Liu, L. Z. Liu, L. Ding and B. H. Jiang 2015. GSK-3 $\beta$   
1064 regulates tumor growth and angiogenesis in human glioma cells. *Oncotarget* 6: 31901-31915. doi:  
1065 10.18632/oncotarget.5043

1066 Zheng, W., H. Wang, Z. Zeng, J. Lin, P. J. Little, L. K. Srivastava and R. Quirion 2012. The possible role of the Akt  
1067 signaling pathway in schizophrenia. *Brain Res* 1470: 145-158. doi: 10.1016/j.brainres.2012.06.032

1068 Zheng, Z., X. Shi, Y. Xiang, A. Zhang and Y. Fang 2021. Involvement of 5-Hydroxytryptamine Receptor 2A in the  
1069 Pathophysiology of Medication-Overuse Headache. *J Pain Res* 14: 453-461. doi: 10.2147/jpr.S283734

1070 Zhou, A., K. Lin, S. Zhang, Y. Chen, N. Zhang, J. Xue, Z. Wang, K. D. Aldape, K. Xie, J. R. Woodgett and S. Huang 2016.  
1071 Nuclear GSK3 $\beta$  promotes tumorigenesis by phosphorylating KDM1A and inducing its deubiquitylation by USP22. *Nat*  
1072 *Cell Biol* 18: 954-966. doi: 10.1038/ncb3396

1073 Zhu, H., Z. Jian, Y. Zhong, Y. Ye, Y. Zhang, X. Hu, B. Pu, L. Gu and X. Xiong 2021. Janus Kinase Inhibition Ameliorates  
1074 Ischemic Stroke Injury and Neuroinflammation Through Reducing NLRP3 Inflammasome Activation via JAK2/STAT3  
1075 Pathway Inhibition. *Front Immunol* 12: 714943. doi: 10.3389/fimmu.2021.714943

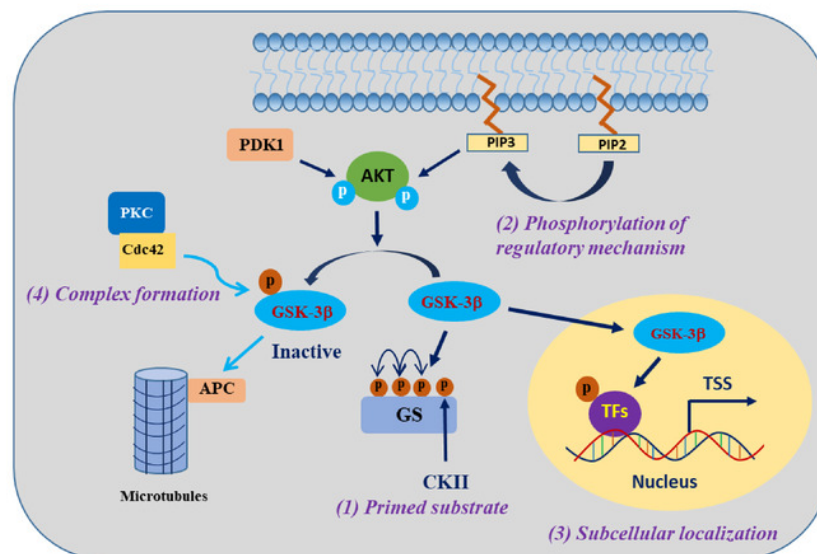
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# Figure 1

Regulatory mechanisms governing the activities of GSK-3 $\beta$ .

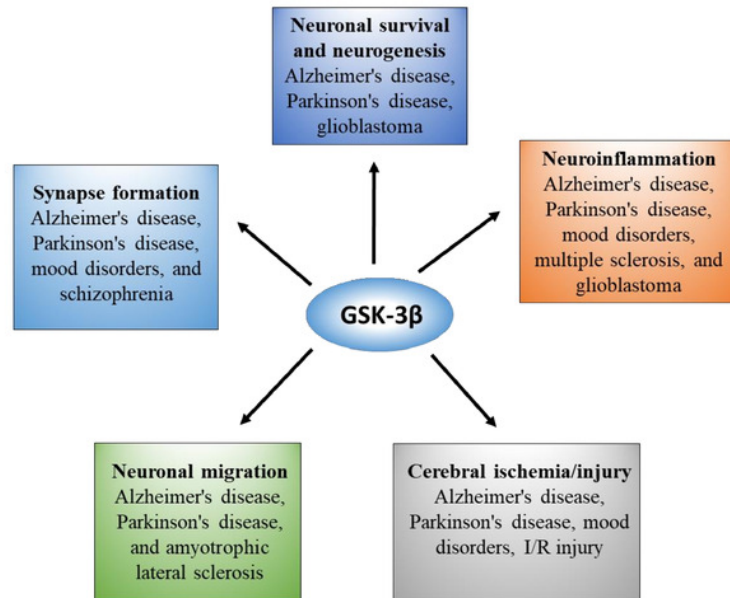
GSK-3 $\beta$  phosphorylation of substrates is synergistically regulated by four mechanisms.

Modified from Jope RS et al., 2007. Copyright 2007, Springer Nature.



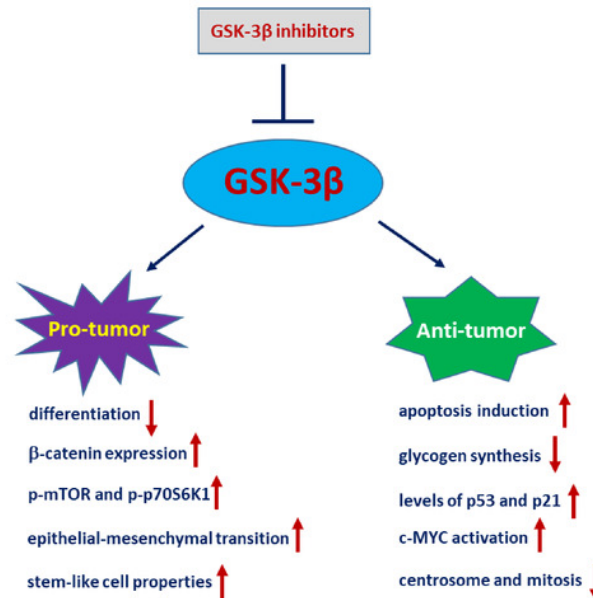
# Figure 2

Overview of GSK-3 $\beta$  functions and correlated diseases in the brain.



# Figure 3

GSK-3 $\beta$  plays a paradoxical role in glioblastoma.

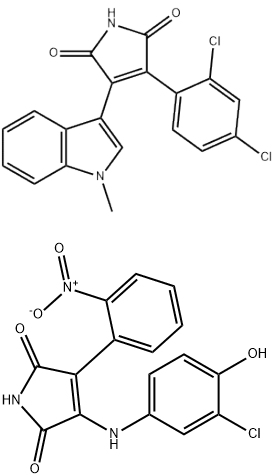
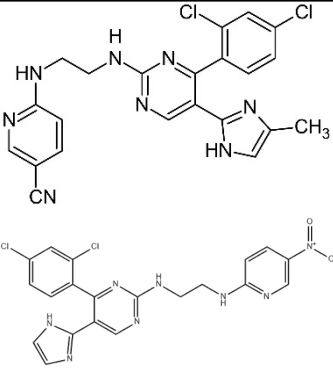
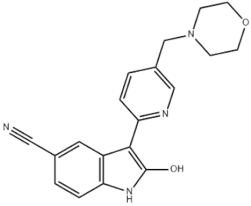
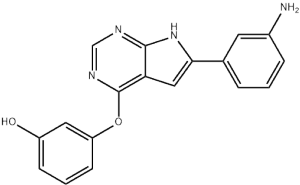


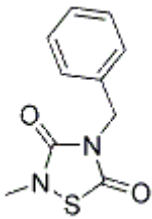
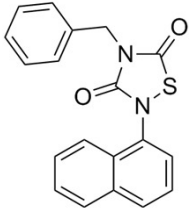
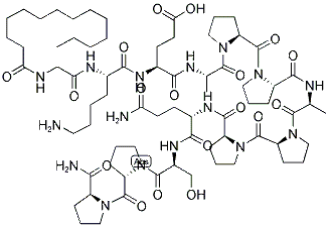
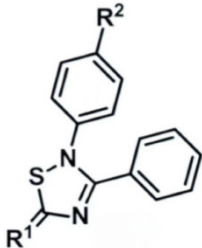
# **Table 1**(on next page)

Representative GSK-3 $\beta$  inhibitors and its usage in brain disease models.

Abbreviations of disease: AD: Alzheimer's disease; PD: Parkinson's disease; HD: Huntington's disease; ALS: Amyotrophic Lateral Sclerosis; MS: Multiple Sclerosis; BD: Bipolar Disorder; SZ: Schizophrenia; GBM: Glioblastoma; I/R injury: Ischemia-reperfusion (I/R) injury.



Type	Typical compounds	Structures	Applications in brain disease
ATP competitive	SB-216763, SB-415286		AD, PD, BD, I/R injury (Kalinichev and Dawson, 2011; Wang et al., 2019; Xiong et al., 2013; Zhang et al., 2014)
	CHIR99021, CHIR98023		HD, GBM, I/R injury (Hu et al., 2021a; Yang et al., 2020; Zhang et al., 2022b)
	AZD1080		PD (Hu et al., 2020)
	TWS119		AD, I/R injury (Gao et al., 2021; Jiang et al., 2021)

Non-ATP competitive	TDZD-8		AD, PD, BD, GBM, MS, SZ, I/R injury (Aguilar-Morante et al., 2010; Beurel et al., 2013; Duka et al., 2009; Huang et al., 2017; Kalinichev and Dawson, 2011; Koehler et al., 2019; Willi et al., 2013)
	Tideglusib		AD, PD, ALS, BD, GBM, I/R injury (Al-Zaidi et al., 2021; Bahmad et al., 2021; del Ser et al., 2013; Martínez-González et al., 2021; Moretti, 2015; Wang et al., 2016)
Substrate competitive	L803mts		AD, MS, HD (Beurel et al., 2013; Eldar-Finkelman and VanHook, 2016; Rippin et al., 2021)
	ITDZs		MS (Palomo et al., 2012; Redondo et al., 2012)

Cations	Lithium	Li	AD, PD, ALS, MS, BD, GBM, SZ, I/R injury (Leucht et al., 2007; Ochoa, 2022; Smith et al., 2020; Young, 2009)
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