

# Temporal bright light at low frequency retards lens-induced myopia in guinea pigs

Baodi Deng<sup>Equal first author, 1</sup>, Wentao Li<sup>Equal first author, 2</sup>, Ziping Chen<sup>3</sup>, Junwen Zeng<sup>1</sup>, Feng Zhao<sup>Corresp. 1</sup>

<sup>1</sup> State Key Laboratory of Ophthalmology, Zhongshan Ophthalmic Center, Sun Yat-sen University, Guangdong Provincial Key Laboratory of Ophthalmology and Visual Science, Guangzhou, China

<sup>2</sup> Huizhou Third People's Hospital, Guangzhou Medical University, Huizhou, China

<sup>3</sup> Guangdong Light Visual Health Research Institute, Guangzhou, China

Corresponding Author: Feng Zhao

Email address: zoc-zhaofeng@foxmail.com

**Purpose:** Bright light conditions are supposed to curb eye growth in animals with experimental myopia. Here we investigated the effects of temporal bright light at very low frequencies exposures on lens-induced myopia (LIM) progression. **Methods:** Myopia was induced by application of -6.00 D lenses over the right eye of guinea pigs. They were randomly divided into four groups based on exposure to different lighting conditions: constant low illumination (CLI; 300 lux), constant high illumination (CHI; 8000 lux), very low frequency light (vLFL; 300/8000 lux, 10 min/c), and low frequency light (LFL; 300/8000 lux, 20 s/c). Refraction and ocular dimensions were measured per week. Changes in ocular dimensions and refractions were analyzed by paired t-tests, and differences among the groups were analyzed by one-way ANOVA. **Results:** Significant myopic shifts in refractive error were induced in lens-treated eyes compared with contralateral eyes in all groups after 3 weeks (all  $P < 0.05$ ). Both CHI and LFL conditions exhibited a significantly less refractive shift of LIM eyes than CLI and vLFL conditions ( $P < 0.05$ ). However, only LFL conditions showed significantly less overall myopic shift and axial elongation than CLI and vLFL conditions (both  $P < 0.05$ ). The decrease in refractive error of both eyes correlated significantly with axial elongation in all groups ( $P < 0.001$ ), except contralateral eyes in the CHI group ( $P = 0.231$ ). LFL condition significantly slacked lens thickening in the contralateral eyes. **Conclusions:** Temporal bright light at low temporal frequency (0.05 Hz) appears to effectively inhibit LIM progression. Further research is needed to determine the safety and the potential mechanism of temporal bright light in myopic progression.

# Temporal bright light at low frequency retards lens-induced myopia in guinea pigs

Baodi Deng<sup>1\*</sup>, Wentao Li<sup>2\*</sup>, Ziping Chen<sup>3</sup>, Junwen Zeng<sup>1</sup>, Feng Zhao<sup>1</sup>

<sup>1</sup>State Key Laboratory of Ophthalmology, Zhongshan Ophthalmic Center, Sun Yat-sen University, Guangdong Provincial Key Laboratory of Ophthalmology and Visual Science, Guangzhou, China

<sup>2</sup>Huizhou Third People's Hospital, Guangzhou Medical University, Huizhou, China

<sup>3</sup>Guangdong Light Visual Health Research Institute, Guangzhou, China

\*These authors contributed equally to this work

Corresponding Author:

Feng Zhao<sup>1</sup>

No. 54 Xianlie South Road, Yuexiu District, Guangzhou, Guangdong Province, China

zoc-zhaofeng@foxmail.com

## Abstract

**Purpose:** Bright light conditions are supposed to curb eye growth in animals with experimental myopia. Here we investigated the effects of temporal bright light at very low frequencies exposures on lens-induced myopia (LIM) progression.

**Methods:** Myopia was induced by application of -6.00 D lenses over the right eye of guinea pigs. They were randomly divided into four groups based on exposure to different lighting conditions: constant low illumination (CLI; 300 lux), constant high illumination (CHI; 8000 lux), very low frequency light (vLFL; 300/8000 lux, 10 min/c), and low frequency light (LFL; 300/8000 lux, 20 s/c). Refraction and ocular dimensions were measured per week. Changes in ocular dimensions and refractions were analyzed by paired t-tests, and differences among the groups were analyzed by one-way ANOVA.

**Results:** Significant myopic shifts in refractive error were induced in lens-treated eyes compared with contralateral eyes in all groups after 3 weeks (all  $P < 0.05$ ). Both CHI and LFL conditions

exhibited a significantly less refractive shift of LIM eyes than CLI and vLFL conditions ( $P < 0.05$ ). However, only LFL conditions showed significantly less overall myopic shift and axial elongation than CLI and vLFL conditions (both  $P < 0.05$ ). The decrease in refractive error of both eyes correlated significantly with axial elongation in all groups ( $P < 0.001$ ), except contralateral eyes in the CHI group ( $P = 0.231$ ). LFL condition significantly slackened lens thickening in the contralateral eyes.

**Conclusions:** Temporal bright light at low temporal frequency (0.05 Hz) appears to effectively inhibit LIM progression. Further research is needed to determine the safety and the potential mechanism of temporal bright light in myopic progression.

## Introduction

Nowadays, it is recognized that outdoor activities can repress the incidence of myopia (He et al. 2015; Wu et al. 2013; Zadnik & Mutti 2019). One factor associated with this protective outdoor effect is the difference in light intensity (French et al. 2013; Lingham et al. 2020; Rose et al. 2008; Sherwin et al. 2012). However, although bright light is reported to prevent the development of form-deprivation myopia (FDM) in all species studied so far (Ashby et al. 2009; Chen et al. 2017; Lan et al. 2014; Smith et al. 2012), results on prevention of lens-induced myopia (LIM), which seemed to be a better model of human myopia (French et al. 2013), are more variable (Ashby & Schaeffel 2010; Smith et al. 2013). In addition, Biswas et al recently reported that in a lens induced hyperopia (LIH) chicken model, daily exposure to high-intensity light promotes axial shortening and hyperopia in a duration dependent manner, whereas optical refocus promotes emmetropization and slows the development of LIH (Biswas et al. 2021). One of the more surprising discoveries on this subject was the finding that daily exposure to intermittent bright light at very low frequencies (0.01 and 0.002 Hz in chicken and 4h/day intermittent bright light consisted 1h of high intensity LED light delivered every 2h in monkeys) fully suppressed FDM development (Lan et al. 2014; Ramachandran et al. 2022). These results indicated that bright light seems to indiscriminately suppress eye growth rather than suppress myopia per se, whereas changes in the visual environment (optical focus and temporal stimuli) have a stronger effect in slowing myopia development.

With respect to temporal stimuli, accumulating evidence suggests that in chicks, guinea pigs, cats and mice, stroboscopic flicker effectively induces myopia at low frequencies and prevents myopic drift at high frequencies (Cremieux et al. 1989; Di et al. 2013a; Rucker et al. 2018; Yu et al. 2011; Zhi et al. 2013). Although we still do not know what the difference between low temporal stimuli under a background with high intensity light or dim/dark light is, we do know

that the dynamic light source used in previous studies is presented as a square wave, which is unnatural and will cause dazzle reflex(Plainis et al. 2006). Another limitation of the previous experimental paradigms was that in these studies, the spectral composition of artificial light is not as well-distributed as sunlight in these studies(Li et al. 2014), which was also suggested to be an independent factor affecting myopic progression. Therefore, it is necessary to evaluate the effect of a more natural and applicable temporal bright light source in LIM for potential therapeutic application for children's myopia.

In an attempt to develop such a dynamic light source that can be applied to humans, full-spectral light with gentle changes of light intensities was applied in this study. With a well-developed visual system(Buttery et al. 1991) and rapid effective response to form-deprivation and optical defocus(Howlett & McFadden 2006; Howlett & McFadden 2009), guinea pigs have been a popular alternative for studying myopia(Howlett & McFadden 2006; Howlett & McFadden 2009; Li et al. 2014; Luo et al. 2017; Yu et al. 2021). According to previous studies, guinea pigs are born hyperopic and undergo rapid emmetropization before 3 weeks of age, which was similar to the time course for emmetropization in early childhood of humans(Zhou et al. 2006). Additionally, the temporal response and its development in the guinea pig retina is identical with those of human beings(Armitage et al. 2001; Racine et al. 2005). Therefore, we consider 1-week-old guinea pigs to be the ideal choice for research on temporal effects of juvenile myopia and by doing so validate the feasibility of this newly designed light source in controlling myopia in children. To avoid possible retinal damage by high level exposure to light(Hunter et al. 2012), we reduced the high light intensity to a less bright level at 8000 lux. Although the lowest temporal frequency used in previous research to explore the sensitivity of temporal modulation is 0.25 Hz(Swanson et al. 1987), we choose much lower frequencies here to avoid dizziness or discomfort caused by temporal light as much as possible. Here we define the two lower frequencies as the low frequency (0.05 Hz) and very low frequency (0.0016 Hz). We believe that this study makes a novel contribution to our understanding of the influence of temporal stimuli on myopia because this study was the first, to the best of our knowledge, longitudinal evaluation of the effect of mesopic light at very low frequency on the development of LIM.

## Materials and methods

### Animals housing

In the present study, male and female 1-week-old guinea pigs (*Cavia porcellus*, English short-hair stock, tricolor strain) were obtained by the Animal Experimental Centre of Zhejiang Province, China and were provided with unconstrained food and water. Two to three guinea pigs were reared in a customized cage (28.2 cm \* 38.2 cm \* 28.5 cm inside), which provides independent lighting conditions from the feeding room. Wiry bottom was applied to keep the hutch dry and ventilated with the room temperature controlled to  $22 \pm 2$  °C. To minimize potential confounders, cages of different groups were placed next to each other and female and male animals were separated by housing. The lamps were set to be on 12:12 light/dark cycle

(turned on from 8:00 AM to 8:00 PM). This experiment was carried out in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and was approved by the animal experimentation ethics committee of the Zhongshan Ophthalmic Center (approval number: 2020-095A). Before the experiment, the eyes would be checked under a slit-lamp microscope and the animals with abnormal eyes such as microphthalmia or corneal haze would be excluded. Necessary measures were used to minimize the animals suffering during the experiment. Additionally, when we observed guinea pigs suffering noticeable weight loss (rapid onset of more than 15% of body weight), weakness or dying, we would consider early termination. After the experiment, all guinea pigs were euthanized by intraperitoneal injection of excessive 1% Pentobarbital sodium (300 mg/kg) followed by cervical dislocation. Before handing over the carcass to the animal center for further disposal, we would check their breathing and heartbeat to ensure that no animals survive.

## Experimental design

Myopia was induced by application of -6.00 D lenses in the right eye of each guinea pig as described by Li(Li et al. 2014). In short, a homemade Velcro mask is glued to the face of guinea pigs. The mask has appropriate holes to expose eyes, nose, mouth and ears. Another Velcro with a round plastic frame was attached around the right eye, and the frame was glued with a negative lens (-6.00 D, PMMA, diameter 18.0 mm, optical zone: 12.0 mm, base curve: 8.0 mm). Special attention was paid to ensuring that the optical center of the lens was aligned directly in front of the center of the pupil. Those lenses were checked at least once a day to ensure that they were in the correct position and clean. If the face mask or lenses were loosened or fell off, they would be reattached at once. Additionally, once the center of the lens was found to have obvious scratches, it will be replaced immediately.

A total of 82 1-week-old guinea pigs were used in this study, referring to previous related research(Li et al. 2014; Luo et al. 2017). They were marked with ear tags and the numbers on the ear tag were input into the excel table. Then RANDBETWEEN (1,4) functions were applied on the numbers generate the randomisation sequence. Afterwards, the numbered guinea pigs were assigned to one of the following four groups: (1) constant low illumination (CLI; n=22, 300 lux), (2) constant high illumination (CHI; n=19, 8000 lux), (3) temporal high luminance at very low frequency (vLFL, n=21, 300/8000 lux, 10 min/cycle), and (4) temporal high luminance at low frequency (LFL; n=20, 300/8000 lux, 20s/cycle).

## Lighting

Solux halogen lamps (4300k; Eiko Ltd., Shawnee, KS, USA) were used to create the dynamic simulated sunlight. The lamp was measured with a fluorospectrophotometer (HR2000; Ocean Optics, Inc., Osaka, Japan; the detection limit is 200–1100 nm) by the Department of Physics of Sun Yat-sen University in Guangzhou, China. Except for the wavelengths between 300 and 350 nm, the spectrum emitted by this lamp effectively simulates the spectral composition of sunlight(Li et al. 2014). We designed the illuminance changing from low (300 lux) to high (8000 lux) levels smoothly and automatically in a temporal wave function to achieve a single variable

(Figure 1A). To achieve the intensity of illumination and form full spectrum light needed in this study, 288 independently controlled point light sources were installed on the roof of the cage at a height of 28.5 cm from the bottom of each cage (Figure 1B). The illumination was manipulated by a function generator (Patent No.: US201916257198) linked to the lamp. Function generators were placed on the outside of the cages. According to previous studies, the temporal sensitivity function (TSF) in guinea pigs is band-pass at bright stimulus intensities (Armitage et al. 2001). Additionally, in guinea pigs, the lifetime of rod desensitization is of the order of 10 s and that of the bleached pigment is presumably 10 min for recovery from saturation (Demontis et al. 1993). Consequently, we chose 0.05 and 0.0016 Hz as the temporal frequencies to make the effective temporal bright light as comfortable as possible.

### Measurement of ocular parameters

Ocular parameters including refractive error and axial dimensions (anterior chamber depth (ACD), lens thickness (LT), vitreous chamber depth (VCD), and axial length (AL)) were measured before the experiment and once per week during the treatment.

Refractions were measured by handheld streak retinoscopy (66 Vision-Tech Co., Ltd., Suzhou, Jiangsu Province, China) with cycloplegia. 0.5% proparacaine hydrochloride (Alcaine; Alcon, Fort Worth, TX, USA) was dropped topically into the conjunctival sac of guinea pigs at first, followed by drops of 0.5% tropicamide and 0.5% phenylephrine (Mydrin-P; Santen, Osaka, Japan) every other 5 min for five times to induce cycloplegia. Results from the two independent skilled optometrists from Zhongshan Ophthalmic Center, who were blinding with regard to the treatment, were averaged. Refractive error was taken as the mean value of the refractive errors with the vertical and horizontal meridians of three repeated measurements and expressed as spherical equivalent (SE).

Ocular biometry was performed by A-scan ultrasonography with a probe of 10 MHz (KN-1800; Kangning Medical Device Co., Ltd., Wuxi, Jiangsu Province, China) as described by Li (Li et al. 2014). In order to achieve local anesthesia while using biopharostat, 0.5% proparacaine hydrochloride (Alcaine, Alcon) eye drops were applied to eyes before measurement. The ultrasonic probe was in direct contact with the corneal apex and carefully made sure that it was vertical to the corneal surface. The mean of the 10 repeated measurements was used for ocular parameters analysis. Due to the fact that the 10 MHz ultrasound probe does not allow choroidal measurements, the AL in vivo was described as the axial distance from the anterior corneal surface to the vitreo-retinal interface (Di et al. 2013b).

### Statistics

Data were represented as mean  $\pm$  SD and statistical analyses were performed with GraphPad Prism (v7.0) (GraphPad Software Inc.). Before analyzing the data, complete the normal distribution test. Paired t-tests were used to compare the relative changes between deprived eyes and non-deprived eyes within a group. Comparisons among groups were assessed by one-way ANOVA followed by Tukey's multiple comparisons test or Kruskal-Wallis test. If significant

differences were detected, post hoc range tests were performed using the Duncan test using SPSS 25 (SPSS, Chicago, IL, USA). Statistical tests were two-tailed, and p-value < 0.05 was considered statistically significant.

## Results

Ocular parameters of all guinea pigs at different time points are listed in Table 1. No significant difference was found in the parameters between the left and right eyes of the individual animals within any group prior to the treatment (all  $P > 0.05$ , see Table 1).

## Refractive errors

Although all lens-treated eyes became significantly less hyperopic than the contralateral eyes in all light conditions after 3 weeks of light exposure (Table 1), the magnitude of the response differed among the lighting conditions.

To directly compare the effect of light conditions on LIM, the overall refractive changes (change in the lens-treated eye ( $\Delta X$ ) subtract change in the contralateral eye ( $\Delta N$ ),  $\Delta X - \Delta N$ ) of the animals were compared. As shown in Figure 2A and Table 1, at the end of the experiment, refractive error in vLFL had the greatest myopic shift of  $-1.83 \pm 0.66$  D (95% confidence interval [CI]:  $-2.137, -1.522$ ;  $n=21$ ), followed by CLI ( $-1.80 \pm 0.65$  D; 95% CI:  $-2.09, -1.5$ ;  $n=22$ ), CHI ( $-1.20 \pm 0.96$  D; 95% CI:  $-1.681, -0.733$ ;  $n=19$ ), and LFL ( $-0.89 \pm 0.95$  D; 95% CI:  $-1.345, -0.432$ ;  $n=20$ ) (one-way ANOVA,  $F=6.298$ ,  $P<0.001$ ). Tukey's multiple comparisons test revealed that refractive changes in LFL had significantly less myopia shift than that of CLI ( $P=0.004$ ) and vLFL ( $P=0.003$ ). Post hoc analysis showed that CLI and vLFL belonged to one subset ( $P=0.898$ ), while CHI and LFL belonged to another subset ( $P=0.224$ ).

Given that both eyes will be affected by the lighting conditions, we further compared the changes in refraction of eyes with or without lens conditions. Consistent with the overall refractive changes, refractive error of lens-treated eyes in vLFL group had the greatest myopic shift and that in LFL group had the least at the end of the experiment (one-way ANOVA,  $F=6.298$ ,  $P<0.001$ ) (Figure 2B). Comparing different light intensities, CHI exhibited a significantly lower myopic shift than CLI ( $P=0.035$ ). Comparing different light temporal frequencies, LFL showed significantly lower myopic shift than vLFL in lens-treated eyes ( $P<0.001$ ). Post hoc analysis also showed that lens-treated eyes in CLI and vLFL groups belonged to one subset ( $P=0.665$ ), while CHI and LFL belonged to another subset ( $P=0.420$ ). Likewise, refractive error of the contralateral eyes in vLFL had the greatest myopic shift of  $-0.589 \pm 0.279$  D (95% CI:  $-0.719, -0.459$ ;  $n=21$ ), followed by CLI ( $-0.511 \pm 0.119$  D; 95% CI:  $-0.565, -0.458$ ;  $n=22$ ), LFL ( $-0.513 \pm 0.153$  D; 95% CI:  $-0.586, -0.439$ ;  $n=20$ ), and CHI ( $-0.401 \pm 0.224$  D; 95% CI:  $-0.512, -0.29$ ;  $n=19$ ) (one-way ANOVA,  $F=2.750$ ,  $P=0.048$ ) (Figure 2C). Specifically, the contralateral eyes in vLFL group exhibited a significantly more myopic shift than that in CHI group ( $P<0.001$ ).

## Ocular dimensions

All eyes elongated throughout the experiment (Table 1). As shown in Figure 3A, after 3 weeks, relative changes of axial length in LFL had the minimum elongation of  $0.38 \pm 0.12$  mm (95% CI: 0.321, 0.44;  $n=20$ ), followed by CHI ( $0.47 \pm 0.15$  mm; 95% CI: 0.395, 0.546;  $n=19$ ), CLI ( $0.47 \pm 0.16$  mm; 95% CI: 0.398, 0.547;  $n=22$ ), and vLFL ( $0.53 \pm 0.15$  mm; 95% CI: 0.464, 0.606;  $n=21$ ) (one-way ANOVA,  $F=3.488$ ,  $P=0.02$ ). Nevertheless, only LFL showed a statistically significant difference in axial elongation with vLFL ( $P=0.01$ ). The relative changes in axial length mainly came from the axial elongation in lens-treated eyes, which showed the minimum axial elongation of  $0.693 \pm 0.115$  mm (95% CI: 0.638, 0.748;  $n=20$ ) in LFL, followed by CHI ( $0.714 \pm 0.114$  mm; 95% CI: 0.657, 0.77;  $n=19$ ), CLI ( $0.776 \pm 0.123$  mm; 95% CI: 0.721, 0.832;  $n=22$ ), and vLFL ( $0.803 \pm 0.103$  mm; 95% CI: 0.755, 0.851;  $n=21$ ) (one-way ANOVA,  $F=4.001$ ,  $P=0.01$ ). No statistically significant difference in axial elongation of contralateral eyes among groups was found (one-way ANOVA,  $F=2.043$ ,  $P=0.115$ ).

During the observation period, there was no obvious change of ACD in interocular difference and lens-treated eyes (one-way ANOVA, all  $P>0.05$ , see Table 1). However, as shown in figure 3B, the ACD illustrated an increasing trend with the observation period among all groups in contralateral eyes (Friedman test,  $P<0.001$ ). On the contrary, compared with the initial time point, with the extension of treatment time, the relative changes of lens thickness ( $\Delta X - \Delta N$ ) in each groups increased gradually (one-way ANOVA, all  $P<0.01$ ), but no differences was found between the groups at any time point (all  $P>0.05$ , see Table 1). This was also true for lens-treated eyes. With respect to contralateral eyes, the lens thickened less at low temporal frequencies (One-way ANOVA,  $F=4.128$ ,  $P=0.009$ ;  $P=0.02$  for LFL with CLI,  $P=0.022$  for LFL with vLFL) (Figure 3C). VCD of all eyes also increased significantly with age (one-way ANOVA, all  $P<0.05$ ). However, changes in VCD among groups were not statistically significant (all  $P>0.05$ ) (Table 1).

## Correlation between changes in refractive error and ocular dimensions

Figure 4 shows the correlation between axial length elongation and refractive shift for the lens-treated eyes and contralateral eyes under each light regimen. Specifically, the decrease in refractive error (i.e., more myopia) of both eyes correlated significantly with the axial length elongation (contralateral eyes: CLI:  $R^2 = 0.132$ ; vLFL:  $R^2 = 0.148$ ; LFL:  $R^2 = 0.436$ ; lens-treated eyes: CLI:  $R^2 = 0.5213$ ; CHI:  $R^2 = 0.3226$ ; vLFL:  $R^2 = 0.447$ ; LFL:  $R^2 = 0.299$ ; all  $P<0.001$ ), except the contralateral eyes in CHI group ( $R^2 = 0.010$ ,  $P = 0.231$ ). These results indicated that the refraction shift was largely axial origin excluding the contralateral eyes in the CHI group. To know what correlated with the refractive shift of contralateral eyes in CHI group, we analyzed the correlation of its refractive shift with other ocular parameters. However, none of the ocular parameters correlated with refractive shift of contralateral eyes in the CHI group (All  $P>0.05$ ; data not show).

## Discussion



Here, we show that under the same housing conditions, compared with the low light conditions, the bright light conditions retard the myopic shift of LIM. The very low temporal frequency (0.0016 Hz) bright light condition produced a similar myopic shift to the low intensity illumination, while the low temporal frequency (0.05 Hz) bright light condition led to significantly less eye growth, implying a temporal sensitivity in hyperopic defocus. Additionally, LFL condition significantly slackened the thickening of lenses in contralateral eyes.

Why short outdoor time has protective effects against myopia and how myopia development is related to light parameters are two of the most studied but as yet unanswered questions in this field. Although human epidemiological studies have shown a correlation between bright light and myopia, the confounding effect of optical distance is not eliminated (Ngo et al. 2013). While in animal studies, myopia is indeed suppressed by bright light, which seems to be indiscriminate suppression of eye growth rather than suppressing myopia per se (Biswas et al. 2021; Chakraborty et al. 2020; Chen et al. 2017; Feldkaemper & Schaeffel 2013). Consistent with these studies, bright light condition at 8,000 lux effectively retarded the decrease in spherical equivalent refraction (SER) of LIM eyes. However, no significant difference was found in the overall myopia shift. One explanation was that bright light which was reported to be capable of retarding myopia development and enhanced hyperopic shifts of lens-induced myopia in guinea pigs was 10,000 lux (Li et al. 2014), while the light intensity of bright light (8,000 lux) in this study was much lower. Nevertheless, does that mean children should be exposed to continuous higher ambient light for longer periods of time? It should be noted that the refractive changes of contralateral eyes in CHI did not correlate to axial elongation. It is possible that the corneal radius of curvature was flattened under continued bright light, as was reported in chicken (Cohen et al. 2012; Li et al. 1995). Besides, recent studies suggested that a sufficient cumulative lux per day at an approximately 5000 lux light intensity with about 2.8 hours reduced  $25.5 \pm 4.5\%$  myopia risk, which was equivalent to the anti-myopic effect of the same cumulative lux at a lower outdoor light intensity with much more outdoor times (He et al. 2022). Additionally, daily exposure to intermittent bright light at very low frequencies showed to be capable of fully suppressing FDM development (Lan et al. 2014; Ramachandran et al. 2022). These evidences suggest that the overall photons arriving at eyes in necessary time, instead of continued bright light exposure, is imperative for myopic control. Nonetheless, temporal bright light at low frequencies showed stronger inhibitory effects on LIM also suggested a temporal sensitivity of bright light.

With respect to temporal stimuli, accumulating evidence suggests that in chickens, guinea pigs and mice, stroboscopic flicker effectively induces myopia at low frequencies and prevents myopic drift at high frequencies (Crewther et al. 2006; Di et al. 2013b; Schwahn & Schaeffel 1997; Yu et al. 2011). Inspired by the fact that temporal stimuli are processed by midget and parasol ganglion cells in the ON and OFF pathways within the retina (Schiller 2010), imbalance of ON and OFF retinal pathway activation is suggested to be the underlying mechanisms (Crewther & Crewther 2002; Crewther et al. 2006; Wang et al. 2019). Specifically, with accumulating evidence finding that blockade of ON pathways effectively inhibited myopia

progression, some studies suggested that ON pathways were the pro-myopic factor(Crewther & Crewther 1990; Crewther & Crewther 2002; Crewther & Crewther 2003; Smith et al. 1991). Nevertheless, other studies suggest that bright light or high frequency flicker might inhibit myopia development by stimulation of the ON pathway proposed via increasing DA pathway activation(Chen et al. 2017; Chuang & Rucker 2019). Theoretically, the temporal bright light with a smooth decline and ascent waveform used in the current study produced a strong stimulation of both ON and OFF pathways. Since we have not accomplished monitoring the b-wave (rapid ON response) and d-wave (slower OFF response) components of the flash ERG, we cannot be certain whether temporal bright stimuli interfered with emmetropization also via the imbalance of ON and OFF retinal pathway activation as it was presumed to be for stroboscopic flicker.

According to a previous study, temporal bright light with its lowest light intensity being 300 lux should activate cone-photoreceptors while rod-photoreceptors are saturated(Joesch & Meister 2016). However, a recent study found that rods do saturate at beginning, but rhodopsin bleaching allows them to escape saturation at bright conditions, with the recovery time shorter at brighter background(Kelber 2018; Tikidji-Hamburyan et al. 2017). Besides, rod activities were supposed to suppress cone flicker sensitivity and response amplitude(Alexander & Fishman 1986; Bush et al. 2019; Lankford et al. 2022; Zele et al. 2008). Additionally, several studies have found that rod activation contributed to eye growth and myopia development(Park et al. 2014; Smith et al. 2009; Smith et al. 2007). Furthermore, rod function was supposed to be the only photoreceptor defining the dopamine release light threshold which is about 400 lux for mice(Perez-Fernandez et al. 2019). Combined together, we speculated that bright temporal light at a frequency whose cycle is less than rod light response period (i.e., keep rod saturated) should be expected to retard myopia shift and eye growth, while the lower temporal frequencies failed to affect refractive development. The lifetime of rod desensitization in guinea pigs is supposed to be approximately 10 s, while the recovery of bleached pigment from saturation takes about 10 minutes(Demontis et al. 1993). In support of our hypothesis, temporal bright light at low temporal frequencies (0.05 Hz) showed significant inhibitory effects on axial elongation and the decrease in SER than CLI and vLFL group, while the vLFL condition (0.0016 Hz) showed no effect on LIM and even promoted myopic susceptibility. However, the mentioned above evidence of rod activity under bright light condition were all from mice and there are no references from previous literature about rod function under bright condition in guinea pigs far to now. Therefore, our hypothesis needs future evidence of these temporal bright lights in mouse myopic model. Additionally, since we failed to measure if the effective intervals of repeated bright light cycles were confined to the time scale of light adaptation, we cannot be certain whether this conjecture applies to all cases. Further studies are also required to clarify the effects of bright flicker on children myopia.

The exact mechanisms underlying light effects on refractive development remain elusive. A number of hypotheses have been proposed, such as the change in depth of focus, physical activity and retinal dopamine (DA) activity (for reviews see Refs. 61, 62)(Ashby et al. 2009; Ashby & Schaeffel 2010; French et al. 2013; Muralidharan et al. 2021; Troilo et al. 2019).

Among them, the involvement of retinal DA seems to be most likely. In this regard, DA synthesis and release were stimulated by light and DA concentration was down regulated in experimental myopic eyes(Boatright et al. 1989; Brainard & Morgan 1987; Dong & McReynolds 1991; Kirsch & Wagner 1989; Megaw et al. 1997; Rohrer et al. 1995; Stone et al. 1989). The antagonists of DA receptors (DR) shown to reverse the anti-myopic effect of bright light(Ashby & Schaeffel 2010; Chen et al. 2017) also favored this presumption. Besides, flicker light was shown to stimulate more retinal DA release than steady light(Kirsch & Wagner 1989; Kramer 1971; Umino et al. 1991). In addition, light with different spectral compositions also showed different efficiency in stimulating DA release(Wang et al. 2018). In particular, continuous full spectrum artificial light with no peak or valley inhibited axial elongation with higher retinal 3, 4-dihydroxyphenylacetic acid (DOPAC) /DA ratio-the metabolic efficiency of DA(Xu et al. 2023). Accordingly, it is reasonable for us to speculate that temporal bright light with full spectrum in the current study stimulated more retinal DA release which led to the inhibition of axial elongation. However, flicker light induced myopia in guinea pigs was corroborated by up regulating DA release(Luo et al. 2017). Nevertheless, a recent study in guinea pigs suggested that retinal DOPAC/DA ratio, instead of retinal DA per se, is associated with flicker-induced myopia(Tian et al. 2021). Further studies measuring levels of DOPAC/DA ratio may be helpful to better characterize the involvement of dopaminergic pathway in the temporal bright light modulation of myopia progression.

Another possible mechanism that might contribute to the anti-myopic effect of natural light is the incremental UV exposure upregulating vitamin D in circulation(Dixon et al. 2013). In favor of this mechanism, accumulating evidence showed that UV exposure was inversely associated with myopia and vitamin D level was lower in myope(Choi et al. 2014; Gao et al. 2021; Mutti & Marks 2011; Tideman et al. 2016; Yazar et al. 2014). It is further supported by the recent observation that calcipotriol supplement can effectively retard mouse FDM(Jiao et al. 2023). However, several evidences suggest that low vitamin D level is not associated with myopia(Harb & Wildsoet 2021; Li et al. 2022; Lingham et al. 2019; Specht et al. 2020; Williams et al. 2017). It should also be pointed out that the Solux halogen lamp applied in this study does not contain UVB ( $\lambda = 290\text{--}315\text{ nm}$ ) radiation, which is the only light parameter that could promote vitamin D synthesis and activation(Chan et al. 2022). Besides, our previous study comparing UV-free fluorescent lamps vs solux halogen lamps also showed no significant difference between these lamps in inhibiting LIM in guinea pigs, though with a trend of inhibiting myopia(Li et al. 2014). Nevertheless, it should be noted that the observed changes in refractive error are relatively small, most particularly for eyes raised at lower light levels. Other experiments have shown larger changes for low light stimuli with -6D lenses(Wang et al. 2014; Wu et al. 2020). Several studies suggested that violet light (VL, 360 to 400 nm) could effectively inhibit myopia development in experimental myopic mice and children(Jiang et al. 2021; Torii et al. 2017a; Torii et al. 2022; Torii et al. 2017b). Furthermore, wavelength-induced myopia was shown in guinea pigs as in chicks, mice, tree shrews, and human(Gawne et al. 2017; Jiang et al. 2021; Rohrer et al. 1992; Rucker et al. 2018; Strickland et al. 2020; Torii et al. 2017a; Torii et al. 2017b; Wang et al. 2018; Wen et al. 2022; Yu et al. 2021). Since dichromatic guinea pigs also have a violet-sensitive

pigment (peak at around 400 nm)(Parry & Bowmaker 2002), we could not deny the possibility of the VL in solux lamps producing less compensation for optical defocus. Further studies are needed to investigate the effectiveness of the spectral distribution in refractive development so as to test its necessity in indoor light design for children myopia inhibition.

One unexpected finding was that thickening of crystalline lenses on the contralateral eyes in the LFL group was less than that in other lighting conditions. According to previous studies, the lenticular thickness of guinea pigs increased rapidly from birth to 5 weeks of age under laboratory lighting conditions, which mainly determines the increase of the axial length(Zhou et al. 2006). The initial increase in lens thickness after visual experience was also reported in tree shrew(Norton & McBrien 1992) and marmoset(Graham & Judge 1999), which differed from children and primates whose crystalline lenses thinned throughout infancy and childhood(Mutti et al. 2005; Mutti et al. 2018; Qiao-Grider et al. 2007). The increase in lens thickness, assuming no change in lens curvature, would tend to increase the lens power, which was suggested to be compensated for by the steepening of the cornea, leading to the continued decline of hyperopia toward emmetropization(Zhou et al. 2006). In addition, the acceleration of decline in lens thickness in children is related to myopia onset and progression(Lu et al. 2021). Given that at the end of the experiment the guinea pigs used in this study were 4 weeks of age and lenses thickening occurred synchronously with increasing vitreous chamber depth, we considered the tardier increase in lens thickness in the LFL group as the reflection of slowing the overall eye growth. Considering that crystalline is essential to the focusing power of the vertebrate eye lens(Roskamp et al. 2020), how flickering light affects lenticular thickness and refractive power still needs further studies.

A limitation in the current study was that the 10 MHz ultrasound probe used in the current study does not allow choroidal measurements. According to previous studies, experimental myopia in guinea pigs shows significant changes in choroidal thickness(Yang et al. 2023; Zhang et al. 2019). The insensitive ultrasound may compromise the accuracy of axial length measurements and account for our failure to detect significant axial differences between groups. However, the results of refraction and ocular dimensions in the present study still provide support for the inhibitory effect of bright light on LIM. In this respect, since light levels affect axial responses to lenses and occluders in guinea pigs(Li et al. 2014; Zhang & Qu 2019), in the current study, a group with the same mean illuminance as the modulated light sources might be an alternative control. However, the brightest light in the current study did not show significant influence on the axial length compared with the low light group. The impact of mean illuminance on refractive outcomes across light modulated conditions is likely to be minimal. One more shortage in the current study is that the duration of treatment is short (3W), which might also be the reason that the axial differences between groups are not captured. Longer-term studies are needed in future studies to provide a better understanding of the prolonged effects.

## Conclusion

Collectively, the results of this preliminary study suggest that temporal modulation of Solux

halogen lamps at low temporal frequency (0.05 Hz) could be an effective way of inhibiting LIM progression. Nonetheless, the application of these findings to humans is limited by the fact that, different from humans, guinea pigs are dichromatic and have no fovea(Do-Nascimento et al. 1991; Rohlich et al. 1994). Future studies are required to investigate how temporal stimuli affect the refractive shift and eye growth. Nevertheless, this study is helpful in understanding the effect of light environment and temporal stimuli on myopia, which may help the development of novel and effective treatment options for slowing myopia progression in children.

## Funding

This work was supported by Science and Technology Planning Project of Guangdong Province (2016A040403016); Science and Technology Project of Guangzhou (202102020886); China Educational Equipment Industry Association (CEFR20001K1). The founders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. No funding or sponsorship was received for the publication of this article.

## Disclosures

All authors declare no conflicts of interest.

## Reference

- Alexander KR, and Fishman GA. 1986. Rod influence on cone flicker detection: variation with retinal eccentricity. *Vision Res* 26:827-834.DOI 10.1016/0042-6989(86)90141-0
- Armitage JA, Bui BV, Gibson R, and Vingrys AJ. 2001. Postnatal development of flicker sensitivity in guinea pigs. *Clin Exp Optom* 84:270-275.DOI 10.1111/j.1444-0938.2001.tb05037.x
- Ashby R, Ohlendorf A, and Schaeffel F. 2009. The effect of ambient illuminance on the development of deprivation myopia in chicks. *Invest Ophthalmol Vis Sci* 50:5348-5354.DOI 10.1167/iops.09-3419
- Ashby RS, and Schaeffel F. 2010. The effect of bright light on lens compensation in chicks. *Invest Ophthalmol Vis Sci* 51:5247-5253.DOI 10.1167/iops.09-4689
- Biswas S, Ramachandran MA, Barathi V, Low WYS, Busoy JF, Milea D, Brennan NA, and Najjar R. 2021. The drive for myopia control: interactions between bright light and optical refocus. *INVESTIGATIVE OPHTHALMOLOGY & VISUAL SCIENCE* 62.DOI
- Boatright JH, Hoel MJ, and Iuvone PM. 1989. Stimulation of endogenous dopamine release and metabolism in amphibian retina by light- and K<sup>+</sup>-evoked depolarization. *Brain Res* 482:164-168.DOI 10.1016/0006-8993(89)90555-6
- Brainard GC, and Morgan WW. 1987. Light-induced stimulation of retinal dopamine: a dose-response relationship.

- 460 *Brain Res* 424:199-203.DOI 10.1016/0006-8993(87)91211-x
- 461 Bush RA, Tanikawa A, Zeng Y, and Sieving PA. 2019. Cone ERG Changes During Light Adaptation in Two All-  
462 Cone Mutant Mice: Implications for Rod-Cone Pathway Interactions. *Invest Ophthalmol Vis Sci* 60:3680-  
463 3688.DOI 10.1167/iov.19-27242
- 464 Buttery RG, Hinrichsen CF, Weller WL, and Haight JR. 1991. How thick should a retina be? A comparative study  
465 of mammalian species with and without intraretinal vasculature. *Vision Res* 31:169-187.DOI 10.1016/0042-  
466 6989(91)90110-q
- 467 Chakraborty R, Ostrin LA, Benavente-Perez A, and Verkicharla PK. 2020. Optical mechanisms regulating  
468 emmetropisation and refractive errors: evidence from animal models. *Clin Exp Optom* 103:55-67.DOI  
469 10.1111/cxo.12991
- 470 Chan HN, Zhang XJ, Ling XT, Bui CH, Wang YM, Ip P, Chu WK, Chen LJ, Tham CC, Yam JC, and Pang CP.  
471 2022. Vitamin D and Ocular Diseases: A Systematic Review. *Int J Mol Sci* 23.DOI 10.3390/ijms23084226
- 472 Chen S, Zhi Z, Ruan Q, Liu Q, Li F, Wan F, Reinach PS, Chen J, Qu J, and Zhou X. 2017. Bright Light Suppresses  
473 Form-Deprivation Myopia Development With Activation of Dopamine D1 Receptor Signaling in the ON  
474 Pathway in Retina. *Invest Ophthalmol Vis Sci* 58:2306-2316.DOI 10.1167/iov.16-20402
- 475 Choi JA, Han K, Park YM, and La TY. 2014. Low serum 25-hydroxyvitamin D is associated with myopia in Korean  
476 adolescents. *Invest Ophthalmol Vis Sci* 55:2041-2047.DOI 10.1167/IOVS.13-12853
- 477 Chuang KK, and Rucker FJ. 2019. The role of dopamine in eye growth responses to color and luminance flicker in  
478 chicks. *Exp Eye Res* 189:107822.DOI 10.1016/j.exer.2019.107822
- 479 Cohen Y, Peleg E, Belkin M, Polat U, and Solomon AS. 2012. Ambient illuminance, retinal dopamine release and  
480 refractive development in chicks. *Exp Eye Res* 103:33-40.DOI 10.1016/j.exer.2012.08.004
- 481 Cremieux J, Orban GA, Duysens J, Amblard B, and Kennedy H. 1989. Experimental myopia in cats reared in  
482 stroboscopic illumination. *Vision Res* 29:1033-1036.DOI 10.1016/0042-6989(89)90117-x
- 483 Crewther DP, and Crewther SG. 1990. Pharmacological modification of eye growth in normally reared and visually  
484 deprived chicks. *Curr Eye Res* 9:733-740.DOI 10.3109/02713689008999568
- 485 Crewther DP, and Crewther SG. 2002. Refractive compensation to optical defocus depends on the temporal profile  
486 of luminance modulation of the environment. *Neuroreport* 13:1029-1032.DOI 10.1097/00001756-  
487 200206120-00010
- 488 Crewther SG, Barutcu A, Murphy MJ, and Crewther DP. 2006. Low frequency temporal modulation of light  
489 promotes a myopic shift in refractive compensation to all spectacle lenses. *Exp Eye Res* 83:322-328.DOI  
490 10.1016/j.exer.2005.12.016
- 491 Crewther SG, and Crewther DP. 2003. Inhibition of retinal ON/OFF systems differentially affects refractive  
492 compensation to defocus. *Neuroreport* 14:1233-1237.DOI 10.1097/00001756-200307010-00009
- 493 Demontis GC, Bisti S, and Cervetto L. 1993. Light sensitivity, adaptation and saturation in mammalian rods. *Prog*  
494 *Brain Res* 95:15-24.DOI 10.1016/s0079-6123(08)60353-2
- 495 Di Y, Liu R, Chu RY, Zhou XT, and Zhou XD. 2013a. Myopia induced by flickering light in guinea pigs: a detailed  
496 assessment on susceptibility of different frequencies. *Int J Ophthalmol* 6:115-119.DOI 10.3980/j.issn.2222-  
497 3959.2013.02.01
- 498 Di Y, Lu N, Li B, Liu R, Chu RY, Zhou XT, and Zhou XD. 2013b. Effects of chronic exposure to 0.5 Hz and 5 Hz  
499 flickering illumination on the eye growth of guinea pigs. *Curr Eye Res* 38:1182-1190.DOI  
500 10.3109/02713683.2013.807931

- Dixon KM, Tongkao-On W, Sequeira VB, Carter SE, Song EJ, Rybchyn MS, Gordon-Thomson C, and Mason RS. 2013. Vitamin D and death by sunshine. *Int J Mol Sci* 14:1964-1977.DOI 10.3390/ijms14011964
- Do-Nascimento JL, Do-Nascimento RS, Damasceno BA, and Silveira LC. 1991. The neurons of the retinal ganglion cell layer of the guinea pig: quantitative analysis of their distribution and size. *Braz J Med Biol Res* 24:199-214.DOI
- Dong CJ, and McReynolds JS. 1991. The relationship between light, dopamine release and horizontal cell coupling in the mudpuppy retina. *J Physiol* 440:291-309.DOI 10.1113/jphysiol.1991.sp018709
- Feldkaemper M, and Schaeffel F. 2013. An updated view on the role of dopamine in myopia. *Exp Eye Res* 114:106-119.DOI 10.1016/j.exer.2013.02.007
- French AN, Ashby RS, Morgan IG, and Rose KA. 2013. Time outdoors and the prevention of myopia. *Exp Eye Res* 114:58-68.DOI 10.1016/j.exer.2013.04.018 S0014-4835(13)00106-1 [pii]
- Gao F, Li P, Liu YQ, and Chen Y. 2021. Association study of the serum 25(OH)D concentration and myopia in Chinese children. *Medicine (Baltimore)* 100:e26570.DOI 10.1097/MD.00000000000026570
- Gawne TJ, Siegwart JT, Jr., Ward AH, and Norton TT. 2017. The wavelength composition and temporal modulation of ambient lighting strongly affect refractive development in young tree shrews. *Exp Eye Res* 155:75-84.DOI 10.1016/j.exer.2016.12.004
- Graham B, and Judge SJ. 1999. Normal development of refractive state and ocular component dimensions in the marmoset (*Callithrix jacchus*). *Vision Res* 39:177-187.DOI 10.1016/s0042-6989(98)00188-6
- Harb EN, and Wildsoet CF. 2021. Nutritional Factors and Myopia: An Analysis of National Health and Nutrition Examination Survey Data. *Optom Vis Sci* 98:458-468.DOI 10.1097/OPX.0000000000001694
- He M, Xiang F, Zeng Y, Mai J, Chen Q, Zhang J, Smith W, Rose K, and Morgan IG. 2015. Effect of Time Spent Outdoors at School on the Development of Myopia Among Children in China: A Randomized Clinical Trial. *JAMA* 314:1142-1148.DOI 10.1001/jama.2015.10803
- He X, Sankaridurg P, Wang J, Chen J, Naduvilath T, He M, Zhu Z, Li W, Morgan IG, Xiong S, Zhu J, Zou H, Rose KA, Zhang B, Weng R, Resnikoff S, and Xu X. 2022. Time Outdoors in Reducing Myopia: A School-Based Cluster Randomized Trial with Objective Monitoring of Outdoor Time and Light Intensity. *Ophthalmology* 129:1245-1254.DOI 10.1016/j.ophtha.2022.06.024
- Howlett MH, and McFadden SA. 2006. Form-deprivation myopia in the guinea pig (*Cavia porcellus*). *Vision Res* 46:267-283.DOI 10.1016/j.visres.2005.06.036
- Howlett MH, and McFadden SA. 2009. Spectacle lens compensation in the pigmented guinea pig. *Vision Res* 49:219-227.DOI 10.1016/j.visres.2008.10.008
- Hunter JJ, Morgan JJ, Merigan WH, Sliney DH, Sparrow JR, and Williams DR. 2012. The susceptibility of the retina to photochemical damage from visible light. *Prog Retin Eye Res* 31:28-42.DOI 10.1016/j.preteyeres.2011.11.001
- Jiang X, Pardue MT, Mori K, Ikeda SI, Torii H, D'Souza S, Lang RA, Kurihara T, and Tsubota K. 2021. Violet light suppresses lens-induced myopia via neuropsin (OPN5) in mice. *Proc Natl Acad Sci U S A* 118.DOI 10.1073/pnas.2018840118
- Jiao S, Reinach PS, Huang C, Yu L, Zhuang H, Ran H, Zhao F, Srinivasalu N, Qu J, and Zhou X. 2023. Calcipotriol Attenuates Form Deprivation Myopia Through a Signaling Pathway Parallel to TGF-beta2-Induced Increases in Collagen Expression. *Invest Ophthalmol Vis Sci* 64:2.DOI 10.1167/iovs.64.2.2
- Joesch M, and Meister M. 2016. A neuronal circuit for colour vision based on rod-cone opponency. *Nature* 532:236-

239.DOI 10.1038/nature17158

Kelber A. 2018. Vision: Rods See in Bright Light. *Curr Biol* 28:R364-R366.DOI 10.1016/j.cub.2018.02.062

Kirsch M, and Wagner HJ. 1989. Release pattern of endogenous dopamine in teleost retinæ during light adaptation and pharmacological stimulation. *Vision Res* 29:147-154.DOI 10.1016/0042-6989(89)90120-x

Kramer SG. 1971. Dopamine: A retinal neurotransmitter. I. Retinal uptake, storage, and light-stimulated release of H3-dopamine in vivo. *Invest Ophthalmol* 10:438-452.DOI

Lan W, Feldkaemper M, and Schaeffel F. 2014. Intermittent episodes of bright light suppress myopia in the chicken more than continuous bright light. *PLoS One* 9:e110906.DOI 10.1371/journal.pone.0110906

Lankford CK, Umino Y, Poria D, Kefalov V, Solessio E, and Baker SA. 2022. Cone-Driven Retinal Responses Are Shaped by Rod But Not Cone HCN1. *J Neurosci* 42:4231-4249.DOI 10.1523/JNEUROSCI.2271-21.2022

Li T, Troilo D, Glasser A, and Howland HC. 1995. Constant light produces severe corneal flattening and hyperopia in chickens. *Vision Res* 35:1203-1209.DOI 10.1016/0042-6989(94)00231-a

Li W, Lan W, Yang S, Liao Y, Xu Q, Lin L, and Yang Z. 2014. The effect of spectral property and intensity of light on natural refractive development and compensation to negative lenses in guinea pigs. *Invest Ophthalmol Vis Sci* 55:6324-6332.DOI 10.1167/iovs.13-13802

Li X, Lin H, Jiang L, Chen X, Chen J, and Lu F. 2022. Low Serum Vitamin D Is Not Correlated With Myopia in Chinese Children and Adolescents. *Front Med (Lausanne)* 9:809787.DOI 10.3389/fmed.2022.809787

Lingham G, Mackey DA, Lucas R, and Yazar S. 2020. How does spending time outdoors protect against myopia? A review. *Br J Ophthalmol* 104:593-599.DOI 10.1136/bjophthalmol-2019-314675 bjophthalmol-2019-314675 [pii]

Lingham G, Yazar S, Lucas RM, Walsh JP, Zhu K, Hunter M, Lim EM, Cooke BR, and Mackey DA. 2019. Low 25-Hydroxyvitamin D Concentration Is Not Associated With Refractive Error in Middle-Aged and Older Western Australian Adults. *Transl Vis Sci Technol* 8:13.DOI 10.1167/tvst.8.1.13

Lu T, Song J, Wu Q, Jiang W, Tian Q, Zhang X, Xu J, Wu J, Hu Y, Sun W, and Bi H. 2021. Refractive lens power and lens thickness in children (6-16 years old). *Sci Rep* 11:19284.DOI 10.1038/s41598-021-98817-9

Luo X, Li B, Li T, Di Y, Zheng C, Ji S, Ma Y, Zhu J, Chen X, and Zhou X. 2017. Myopia induced by flickering light in guinea pig eyes is associated with increased rather than decreased dopamine release. *Mol Vis* 23:666-679.DOI

Megaw PL, Morgan IG, and Boelen MK. 1997. Dopaminergic behaviour in chicken retina and the effect of form deprivation. *Aust N Z J Ophthalmol* 25 Suppl 1:S76-78.DOI 10.1111/j.1442-9071.1997.tb01764.x

Muralidharan AR, Lanca C, Biswas S, Barathi VA, Wan Yu Shermaine L, Seang-Mei S, Milea D, and Najjar RP. 2021. Light and myopia: from epidemiological studies to neurobiological mechanisms. *Ther Adv Ophthalmol* 13:25158414211059246.DOI 10.1177/25158414211059246

Mutti DO, and Marks AR. 2011. Blood levels of vitamin D in teens and young adults with myopia. *Optom Vis Sci* 88:377-382.DOI 10.1097/OPX.0b013e31820b0385

Mutti DO, Mitchell GL, Jones LA, Friedman NE, Frane SL, Lin WK, Moeschberger ML, and Zadnik K. 2005. Axial growth and changes in lenticular and corneal power during emmetropization in infants. *Invest Ophthalmol Vis Sci* 46:3074-3080.DOI 10.1167/iovs.04-1040

Mutti DO, Sinnott LT, Lynn Mitchell G, Jordan LA, Friedman NE, Frane SL, and Lin WK. 2018. Ocular Component Development during Infancy and Early Childhood. *Optom Vis Sci* 95:976-985.DOI 10.1097/OPX.0000000000001296



- Ngo C, Saw SM, Dharani R, and Flitcroft I. 2013. Does sunlight (bright lights) explain the protective effects of outdoor activity against myopia? *Ophthalmic Physiol Opt* 33:368-372.DOI 10.1111/opo.12051
- Norton TT, and McBrien NA. 1992. Normal development of refractive state and ocular component dimensions in the tree shrew (*Tupaia belangeri*). *Vision Res* 32:833-842.DOI 10.1016/0042-6989(92)90026-f
- Park H, Jabbar SB, Tan CC, Sidhu CS, Abey J, Aseem F, Schmid G, Iuvone PM, and Pardue MT. 2014. Visually-driven ocular growth in mice requires functional rod photoreceptors. *Invest Ophthalmol Vis Sci* 55:6272-6279.DOI 10.1167/iov.14-14648
- Parry JW, and Bowmaker JK. 2002. Visual pigment coexpression in Guinea pig cones: a microspectrophotometric study. *Invest Ophthalmol Vis Sci* 43:1662-1665.DOI
- Perez-Fernandez V, Milosavljevic N, Allen AE, Vessey KA, Jobling AI, Fletcher EL, Breen PP, Morley JW, and Cameron MA. 2019. Rod Photoreceptor Activation Alone Defines the Release of Dopamine in the Retina. *Curr Biol* 29:763-774 e765.DOI 10.1016/j.cub.2019.01.042
- Plainis S, Murray IJ, and Carden D. 2006. The dazzle reflex: electrophysiological signals from ocular muscles reveal strong binocular summation effects. *Ophthalmic Physiol Opt* 26:318-325.DOI 10.1111/j.1475-1313.2006.00350.x
- Qiao-Grider Y, Hung LF, Kee CS, Ramamirtham R, and Smith EL, 3rd. 2007. Normal ocular development in young rhesus monkeys (*Macaca mulatta*). *Vision Res* 47:1424-1444.DOI 10.1016/j.visres.2007.01.025
- Racine J, Joly S, Rufiange M, Rosolen S, Casanova C, and Lachapelle P. 2005. The photopic ERG of the albino guinea pig (*Cavia porcellus*): a model of the human photopic ERG. *Doc Ophthalmol* 110:67-77.DOI 10.1007/s10633-005-7345-x
- Ramachandran MA, Chong LY, Tan RKY, Barathi VA, Hung LF, Tan BY, Schmetterer L, Yen JCK, Milea D, Saw SM, and Najjar R. 2022. Intermittent exposure to bright light can prevent form-deprivation myopia in a monkey model. *INVESTIGATIVE OPHTHALMOLOGY & VISUAL SCIENCE* 63.DOI
- Rohlich P, van Veen T, and Szel A. 1994. Two different visual pigments in one retinal cone cell. *Neuron* 13:1159-1166.DOI 10.1016/0896-6273(94)90053-1
- Rohrer B, Iuvone PM, and Stell WK. 1995. Stimulation of dopaminergic amacrine cells by stroboscopic illumination or fibroblast growth factor (bFGF, FGF-2) injections: possible roles in prevention of form-deprivation myopia in the chick. *Brain Res* 686:169-181.DOI 10.1016/0006-8993(95)00370-6
- Rohrer B, Schaeffel F, and Zrenner E. 1992. Longitudinal chromatic aberration and emmetropization: results from the chicken eye. *J Physiol* 449:363-376.DOI 10.1113/jphysiol.1992.sp019090
- Rose KA, Morgan IG, Ip J, Kifley A, Huynh S, Smith W, and Mitchell P. 2008. Outdoor activity reduces the prevalence of myopia in children. *Ophthalmology* 115:1279-1285.DOI 10.1016/j.ophtha.2007.12.019 S0161-6420(07)01364-4 [pii]
- Roskamp KW, Paulson CN, Brubaker WD, and Martin RW. 2020. Function and Aggregation in Structural Eye Lens Crystallins. *Acc Chem Res* 53:863-874.DOI 10.1021/acs.accounts.0c00014
- Rucker F, Britton S, and Taylor C. 2018. Color and Temporal Frequency Sensitive Eye Growth in Chick. *Invest Ophthalmol Vis Sci* 59:6003-6013.DOI 10.1167/iov.18-25322
- Schiller PH. 2010. Parallel information processing channels created in the retina. *Proc Natl Acad Sci U S A* 107:17087-17094.DOI 10.1073/pnas.1011782107
- Schwahn HN, and Schaeffel F. 1997. Flicker parameters are different for suppression of myopia and hyperopia. *Vision Res* 37:2661-2673.DOI 10.1016/s0042-6989(97)00114-4

- Sherwin JC, Hewitt AW, Coroneo MT, Kearns LS, Griffiths LR, and Mackey DA. 2012. The association between time spent outdoors and myopia using a novel biomarker of outdoor light exposure. *Invest Ophthalmol Vis Sci* 53:4363-4370.DOI 10.1167/iovs.11-8677
- Smith EL, 3rd, Fox DA, and Duncan GC. 1991. Refractive-error changes in kitten eyes produced by chronic on-channel blockade. *Vision Res* 31:833-844.DOI 10.1016/0042-6989(91)90150-4
- Smith EL, 3rd, Hung LF, Arumugam B, and Huang J. 2013. Negative lens-induced myopia in infant monkeys: effects of high ambient lighting. *Invest Ophthalmol Vis Sci* 54:2959-2969.DOI 10.1167/iovs.13-11713
- Smith EL, 3rd, Hung LF, and Huang J. 2009. Relative peripheral hyperopic defocus alters central refractive development in infant monkeys. *Vision Res* 49:2386-2392.DOI 10.1016/j.visres.2009.07.011
- Smith EL, 3rd, Hung LF, and Huang J. 2012. Protective effects of high ambient lighting on the development of form-deprivation myopia in rhesus monkeys. *Invest Ophthalmol Vis Sci* 53:421-428.DOI 10.1167/iovs.11-8652
- Smith EL, 3rd, Ramamirtham R, Qiao-Grider Y, Hung LF, Huang J, Kee CS, Coats D, and Paysse E. 2007. Effects of foveal ablation on emmetropization and form-deprivation myopia. *Invest Ophthalmol Vis Sci* 48:3914-3922.DOI 10.1167/iovs.06-1264
- Specht IO, Jacobsen N, Frederiksen P, and Heitmann BL. 2020. Neonatal vitamin D status and myopia in young adult men. *Acta Ophthalmol* 98:500-505.DOI 10.1111/aos.14349
- Stone RA, Lin T, Laties AM, and Iuvone PM. 1989. Retinal dopamine and form-deprivation myopia. *Proc Natl Acad Sci U S A* 86:704-706.DOI 10.1073/pnas.86.2.704
- Strickland R, Landis EG, and Pardue MT. 2020. Short-Wavelength (Violet) Light Protects Mice From Myopia Through Cone Signaling. *Invest Ophthalmol Vis Sci* 61:13.DOI 10.1167/iovs.61.2.13
- Swanson WH, Ueno T, Smith VC, and Pokorny J. 1987. Temporal modulation sensitivity and pulse-detection thresholds for chromatic and luminance perturbations. *J Opt Soc Am A* 4:1992-2005.DOI 10.1364/josaa.4.001992
- Tian T, Zou L, Wang S, Liu R, and Liu H. 2021. The Role of Dopamine in Emmetropization Modulated by Wavelength and Temporal Frequency in Guinea Pigs. *Invest Ophthalmol Vis Sci* 62:20.DOI 10.1167/iovs.62.12.20
- Tideman JW, Polling JR, Voortman T, Jaddoe VW, Uitterlinden AG, Hofman A, Vingerling JR, Franco OH, and Klaver CC. 2016. Low serum vitamin D is associated with axial length and risk of myopia in young children. *Eur J Epidemiol* 31:491-499.DOI 10.1007/s10654-016-0128-8
- Tikidji-Hamburyan A, Reinhard K, Storch R, Dietter J, Seitter H, Davis KE, Idrees S, Mutter M, Walmsley L, Bedford RA, Ueffing M, Ala-Laurila P, Brown TM, Lucas RJ, and Munch TA. 2017. Rods progressively escape saturation to drive visual responses in daylight conditions. *Nat Commun* 8:1813.DOI 10.1038/s41467-017-01816-6
- Torii H, Kurihara T, Seko Y, Negishi K, Ohnuma K, Inaba T, Kawashima M, Jiang X, Kondo S, Miyauchi M, Miwa Y, Katada Y, Mori K, Kato K, Tsubota K, Goto H, Oda M, Hatori M, and Tsubota K. 2017a. Violet Light Exposure Can Be a Preventive Strategy Against Myopia Progression. *EBioMedicine* 15:210-219.DOI 10.1016/j.ebiom.2016.12.007
- Torii H, Mori K, Okano T, Kondo S, Yang HY, Yotsukura E, Hanyuda A, Ogawa M, Negishi K, Kurihara T, and Tsubota K. 2022. Short-Term Exposure to Violet Light Emitted from Eyeglass Frames in Myopic Children: A Randomized Pilot Clinical Trial. *J Clin Med* 11.DOI 10.3390/jcm11206000

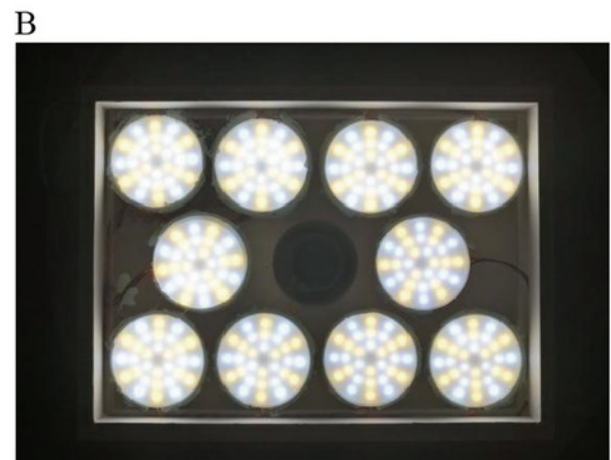
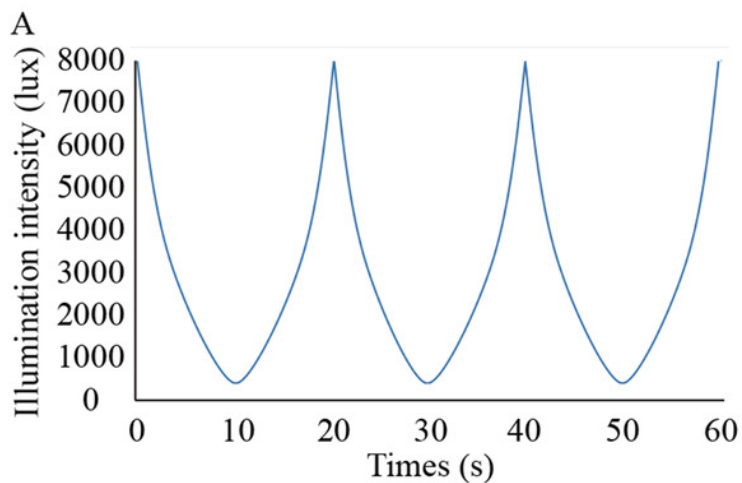
- Torii H, Ohnuma K, Kurihara T, Tsubota K, and Negishi K. 2017b. Violet Light Transmission is Related to Myopia Progression in Adult High Myopia. *Sci Rep* 7:14523.DOI 10.1038/s41598-017-09388-7
- Troilo D, Smith EL, 3rd, Nickla DL, Ashby R, Tkatchenko AV, Ostrin LA, Gawne TJ, Pardue MT, Summers JA, Kee CS, Schroedl F, Wahl S, and Jones L. 2019. IMI - Report on Experimental Models of Emmetropization and Myopia. *Invest Ophthalmol Vis Sci* 60:M31-M88.DOI 10.1167/iovs.18-25967
- Umino O, Lee Y, and Dowling JE. 1991. Effects of light stimuli on the release of dopamine from interplexiform cells in the white perch retina. *Vis Neurosci* 7:451-458.DOI 10.1017/s0952523800009743
- Wang M, Aleman AC, and Schaeffel F. 2019. Probing the Potency of Artificial Dynamic ON or OFF Stimuli to Inhibit Myopia Development. *Invest Ophthalmol Vis Sci* 60:2599-2611.DOI 10.1167/iovs.18-26471
- Wang M, Schaeffel F, Jiang B, and Feldkaemper M. 2018. Effects of Light of Different Spectral Composition on Refractive Development and Retinal Dopamine in Chicks. *Invest Ophthalmol Vis Sci* 59:4413-4424.DOI 10.1167/iovs.18-23880
- Wang S, Liu S, Mao J, and Wen D. 2014. Effect of retinoic acid on the tight junctions of the retinal pigment epithelium-choroid complex of guinea pigs with lens-induced myopia in vivo. *Int J Mol Med* 33:825-832.DOI 10.3892/ijmm.2014.1651
- Wen Y, Dai B, Zhang X, Zhu H, Xie C, Xia J, Sun Y, Zhu M, Tong J, and Shen Y. 2022. Retinal Transcriptomics Analysis Reveals the Underlying Mechanism of Disturbed Emmetropization Induced by Wavelength Defocus. *Curr Eye Res* 47:908-917.DOI 10.1080/02713683.2022.2048395
- Williams KM, Bentham GC, Young IS, McGinty A, McKay GJ, Hogg R, Hammond CJ, Chakravarthy U, Rahu M, Seland J, Soubrane G, Tomazzoli L, Topouzis F, and Fletcher AE. 2017. Association Between Myopia, Ultraviolet B Radiation Exposure, Serum Vitamin D Concentrations, and Genetic Polymorphisms in Vitamin D Metabolic Pathways in a Multicountry European Study. *JAMA Ophthalmol* 135:47-53.DOI 10.1001/jamaophthalmol.2016.4752
- Wu PC, Tsai CL, Wu HL, Yang YH, and Kuo HK. 2013. Outdoor activity during class recess reduces myopia onset and progression in school children. *Ophthalmology* 120:1080-1085.DOI 10.1016/j.ophtha.2012.11.009
- Wu S, Guo D, Wei H, Yin X, Zhang L, Guo B, Xu F, Hao Y, Jiang W, and Bi H. 2020. Disrupted potassium ion homeostasis in ciliary muscle in negative lens-induced myopia in Guinea pigs. *Arch Biochem Biophys* 688:108403.DOI 10.1016/j.abb.2020.108403
- Xu X, Shi J, Zhang C, Shi L, Bai Y, Shi W, and Wang Y. 2023. Effects of artificial light with different spectral composition on eye axial growth in juvenile guinea pigs. *Eur J Histochem* 67.DOI 10.4081/ejh.2023.3634
- Yang Y, Chen M, Yao X, Wang J, Shi J, Wang Y, Tian J, Zhou X, Qu J, and Zhang S. 2023. Choroidal blood perfusion could predict the sensitivity of myopia formation in Guinea pigs. *Exp Eye Res* 232:109509.DOI 10.1016/j.exer.2023.109509
- Yazar S, Hewitt AW, Black LJ, McKnight CM, Mountain JA, Sherwin JC, Oddy WH, Coroneo MT, Lucas RM, and Mackey DA. 2014. Myopia is associated with lower vitamin D status in young adults. *Invest Ophthalmol Vis Sci* 55:4552-4559.DOI 10.1167/iovs.14-14589
- Yu M, Liu W, Wang B, and Dai J. 2021. Short Wavelength (Blue) Light Is Protective for Lens-Induced Myopia in Guinea Pigs Potentially Through a Retinoic Acid-Related Mechanism. *Invest Ophthalmol Vis Sci* 62:21.DOI 10.1167/iovs.62.1.21
- Yu Y, Chen H, Tuo J, and Zhu Y. 2011. Effects of flickering light on refraction and changes in eye axial length of C57BL/6 mice. *Ophthalmic Res* 46:80-87.DOI 10.1159/000323179

- Zadnik K, and Mutti DO. 2019. Outdoor Activity Protects Against Childhood Myopia-Let the Sun Shine In. *JAMA Pediatr* 173:415-416.DOI 10.1001/jamapediatrics.2019.0278
- Zele AJ, Cao D, and Pokorny J. 2008. Rod-cone interactions and the temporal impulse response of the cone pathway. *Vision Res* 48:2593-2598.DOI 10.1016/j.visres.2008.04.003
- Zhang L, and Qu X. 2019. The Effects of High Lighting on the Development of Form-Deprivation Myopia in Guinea Pigs. *Invest Ophthalmol Vis Sci* 60:4319-4327.DOI 10.1167/iovs.18-25258
- Zhang S, Zhang G, Zhou X, Xu R, Wang S, Guan Z, Lu J, Srinivasalu N, Shen M, Jin Z, Qu J, and Zhou X. 2019. Changes in Choroidal Thickness and Choroidal Blood Perfusion in Guinea Pig Myopia. *Invest Ophthalmol Vis Sci* 60:3074-3083.DOI 10.1167/iovs.18-26397
- Zhi Z, Pan M, Xie R, Xiong S, Zhou X, and Qu J. 2013. The effect of temporal and spatial stimuli on the refractive status of guinea pigs following natural emmetropization. *Invest Ophthalmol Vis Sci* 54:890-897.DOI 10.1167/iovs.11-8064
- Zhou X, Qu J, Xie R, Wang R, Jiang L, Zhao H, Wen J, and Lu F. 2006. Normal development of refractive state and ocular dimensions in guinea pigs. *Vision Res* 46:2815-2823.DOI 10.1016/j.visres.2006.01.027

# Figure 1

Lighting conditions used in the study

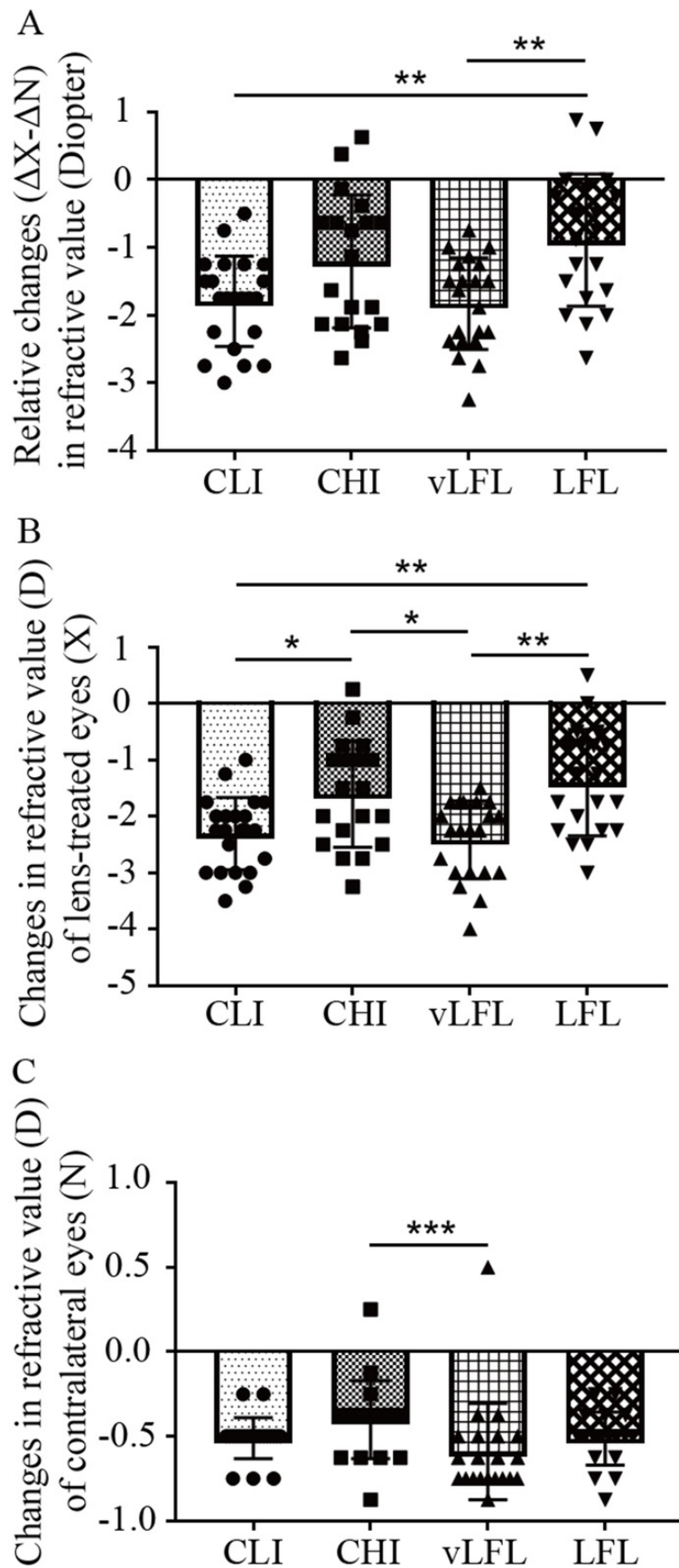
(A) Waveforms of flicker used in the experiment changed smoothly; (B) Arrangement of lamps on top of cages.



# Figure 2

Comparison of the changes of refractive error among the groups at the end of experiment.

(A) Guinea pigs exposed to CLI and vLFL demonstrated a significant reduction in the average refractive shift (OD-OS) compared to the CHI and LFL groups. The refractive shift of lens-treated eyes (B) and the contralateral eyes (C) showed different changes. CLI, constant low illumination (n=22); CHI, constant high illumination (n=19); vLFL, temporal bright light at very low frequency (n=21); LFL, temporal bright light at low frequency (n=20); Data are presented as mean  $\pm$  SD. \*P<0.05, \*\*P<0.01, \*\*\*P<0.001; Error bars:  $\pm$  SEM.

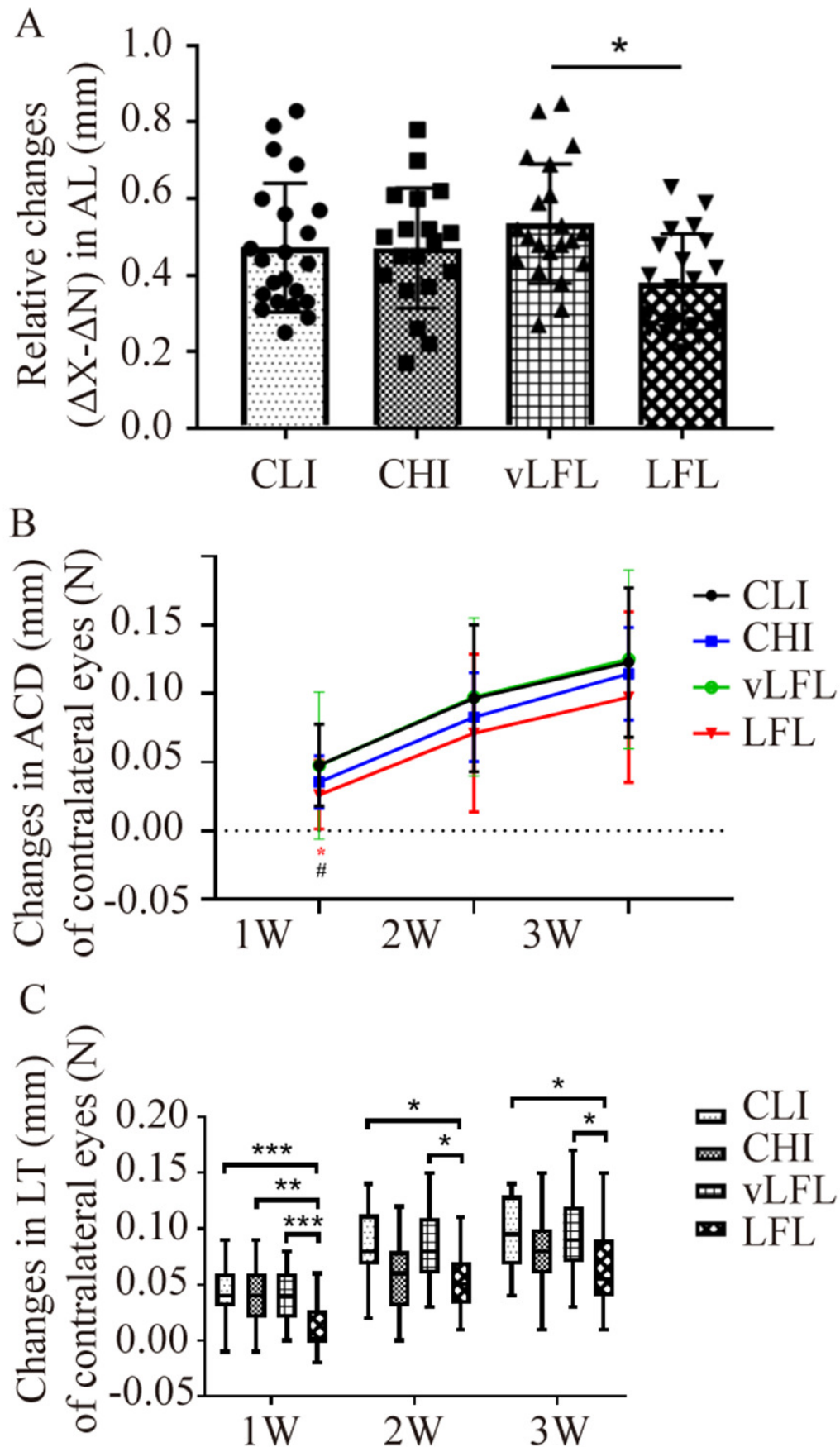


# Figure 3

The effects of four light conditions on ocular biometry.

(A) At the end of treatment, LFL exposure significantly reined in the axial elongation (OD-OS) than vLFL condition. (B) Changes in anterior chamber depth of contralateral eyes were growing with age. (C) Changes in lens thickness of contralateral eyes was less in LFL group. CLI, constant low illumination (n=22); CHI, constant high illumination (n=19); vLFL, temporal bright light at very low frequency (n=21); LFL, temporal bright light at low frequency (n=20); Data are presented as mean  $\pm$  SD. \* (black)  $P<0.05$ , \*\* $P<0.01$ , \*\*\* $P<0.001$ ; \* (red)  $P<0.05$ , LFL versus CLI group; #  $P<0.05$ ; LFL versus vLFL group; Error bars:  $\pm$  SEM.

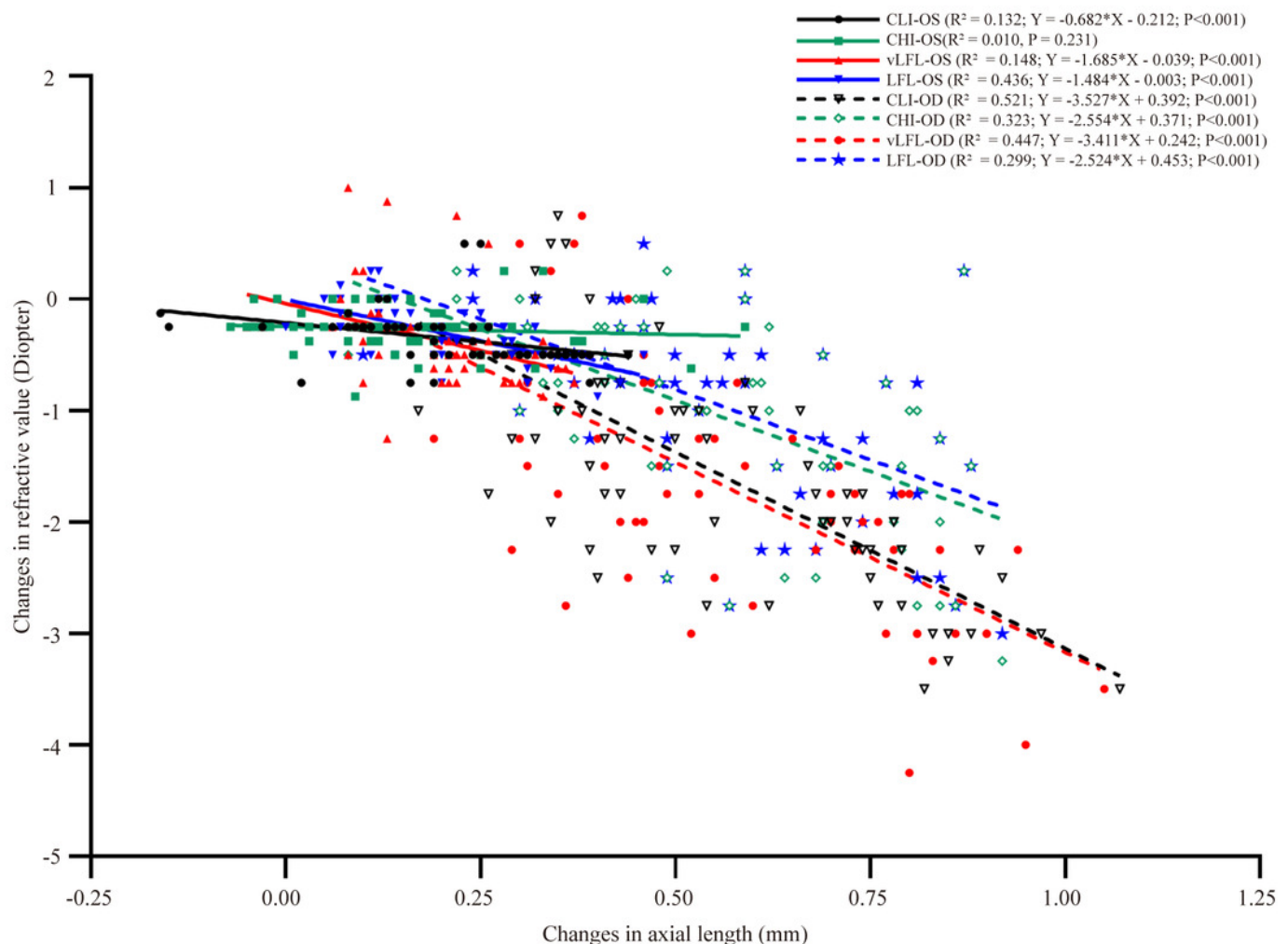




# Figure 4

Correlations between changes in axial length and refractive error.

All eyes, including the contralateral and lens-treated eyes, showed a significant correlation between changes in axial length and refractive error, except the contralateral eyes in CHI group. Solid line represented the data from the contralateral eyes (i.e., OS) of all guinea pigs; long dash line represented the data from the lens-treated eyes (i.e., OD) of all guinea pigs.



# **Table 1**(on next page)

Biometric results (mean  $\pm$  SD) of ocular parameters and changes at different time points.

CLI, constant low illumination; CHI, constant high illumination; vLFL, very low frequency cycles of dynamic light; LFL, low frequency cycles of dynamic light; ACD, anterior chamber depth; LT, lens thickness; VCD, vitreous chamber depth; AL, axial length. Data are presented as mean  $\pm$  SD.

Paradigms	Groups	Time Points	Refractive Error, D	ACD, mm	LT, mm	VCD, mm	AL, mm
Without lenses	CLI	Baseline	3.60±0.58	1.10±0.05	2.49±0.14	3.46±0.16	7.24±0.14
		First Week	3.39±0.57	1.11±0.05	2.53±0.14	3.52±0.15	7.33±0.14
		Second week	3.28±0.54	1.13±0.05	2.57±0.14	3.57±0.16	7.45±0.14
		Third Week	3.09±0.55	1.14±0.05	2.58±0.14	3.61±0.16	7.55±0.16
		Change	-0.51±0.05	0.05±0.02	0.10±0.03	0.14±0.05	0.30±0.08
	CHI	Baseline	3.64±0.62	1.09±0.06	2.47±0.14	3.43±0.20	7.25±0.13
		First Week	3.50±0.65	1.10±0.06	2.51±0.15	3.48±0.18	7.34±0.16
		Second week	3.39±0.65	1.12±0.06	2.53±0.15	3.53±0.17	7.40±0.17
		Third Week	3.24±0.62	1.13±0.06	2.55±0.15	3.57±0.18	7.49±0.17
		Change	-0.40±0.23	0.05±0.01	0.08±0.03	0.14±0.05	0.24±0.15
	vLFL	Baseline	3.95±0.67	1.11±0.05	2.51±0.14	3.44±0.19	7.26±0.09
		First Week	3.84±0.79	1.13±0.06	2.55±0.13	3.49±0.20	7.36±0.08
		Second week	3.56±0.60	1.15±0.06	2.59±0.13	3.55±0.20	7.48±0.08
		Third Week	3.36±0.60	1.16±0.06	2.61±0.12	3.59±0.20	7.53±0.08
		Change	-0.59±0.29	0.05±0.03	0.10±0.04	0.14±0.05	0.27±0.07
	LFL	Baseline	3.76±0.62	1.11±0.05	2.50±0.12	3.44±0.20	7.28±0.09
		First Week	3.69±0.54	1.12±0.06	2.51±0.13	3.48±0.20	7.38±0.10
		Second week	3.41±0.63	1.14±0.06	2.55±0.12	3.53±0.20	7.50±0.12
		Third Week	3.25±0.60	1.15±0.06	2.56±0.12	3.57±0.20	7.60±0.12
		Change	-0.51±0.16	0.04±0.02	0.07±0.03	0.13±0.04	0.31±0.08
With -6D lenses	CLI	Baseline	3.58±0.41	1.09±0.04	2.49±0.14	3.43±0.16	7.22±0.1
		First Week	2.89±0.62	1.1±0.04	2.62±0.13	3.58±0.17	7.61±0.1
		Second week	1.78±0.52	1.11±0.05	2.7±0.13	3.73±0.21	7.74±0.1
		Third Week	1.27±0.48	1.09±0.05	2.83±0.16	4.07±0.2	7.99±0.09
		Change	-2.31±0.64	-0.01±0.07	0.35±0.24	0.64±0.27	0.78±0.13

CHI	Baseline	3.54±0.53	1.08±0.05	2.45±0.14	3.39±0.18	7.22±0.09
	First Week	2.96±0.39	1.1±0.04	2.61±0.14	3.54±0.25	7.65±0.15
	Second week	2.55±0.77	1.09±0.05	2.71±0.12	3.56±0.19	7.76±0.14
	Third Week	1.93±0.9	1.11±0.06	2.85±0.17	3.88±0.24	7.94±0.1
	Change	-1.61±0.94	0.02±0.07	0.4±0.2	0.48±0.25	0.71±0.12
vLFL	Baseline	3.67±0.58	1.09±0.05	2.48±0.14	3.42±0.19	7.19±0.07
	First Week	2.8±0.67	1.11±0.04	2.61±0.13	3.56±0.17	7.58±0.09
	Second week	1.77±0.58	1.11±0.05	2.69±0.14	3.64±0.22	7.74±0.1
	Third Week	1.25±0.47	1.09±0.05	2.83±0.16	3.97±0.18	8±0.08
	Change	-2.42±0.69	0±0.07	0.35±0.24	0.55±0.3	0.8±0.11
LFL	Baseline	3.55±0.52	1.11±0.05	2.48±0.14	3.43±0.15	7.22±0.08
	First Week	2.96±0.38	1.09±0.05	2.6±0.13	3.56±0.25	7.65±0.14
	Second week	2.68±0.76	1.09±0.05	2.71±0.13	3.65±0.17	7.76±0.13
	Third Week	2.15±0.89	1.1±0.04	2.84±0.17	3.87±0.22	7.91±0.09
	Change	-1.4±0.94	-0.01±0.06	0.36±0.25	0.44±0.24	0.69±0.12