Population genetic structure and hybrid zone analyses for species delimitation in Japanese toad (*Bufo japonicus*) (#87292)

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Population genetic structure and hybrid zone analyses for species delimitation in Japanese toad (*Bufo japonicus*)

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Hybridization following the second contact can produce different outcomes, depending on the extent to which genetic diversity and reproductive barriers have accumulated during isolation. Japanese toad, *B. japonicus,* is distributed on the main islands of Japan. In this study, we applied MIG-seq to achieve a fine-scale resolution on the genetic cluster in *B. j. japonicus* and *B. j. formosus*. Moreover, we elucidated hybridization patterns and gene flow degrees across the contact zones between identified clusters. Using SNPs data, we found four genetic clusters in *B. j. japonicus* and *B. j. formosus* and three contact zones of the cluster pairs of these four clusters. The oldest diverged lineage, *B. j. japonicus* and *B. j. formosus,* formed a narrow contact zone consistent with species distinctiveness. Therefore we recommend that these two subspecies be elevated to the species level. In contrast, the less diverged pairs of two clusters in *B. j. japonicus* and *B. j. formosus*, respectively, admix over a hundred kilometers, suggesting that they have not yet developed strong reproductive isolation and should be treated as conspecifics. These results contribute to resolving taxonomic confusion in Japanese toads.

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Abstract

Hybridization following the second contact can produce different outcomes, depending on the extent to which genetic diversity and reproductive barriers have accumulated during isolation. Japanese toad, *B. japonicus,* is distributed on the main islands of Japan. In this study, we applied MIG-seq to achieve a fine-scale resolution on the genetic cluster in *B. j. japonicus* and *B. j. formosus*. Moreover, we elucidated hybridization patterns and gene flow degrees across the contact zones between identified clusters. Using SNPs data, we found four genetic clusters in *B. j. japonicus* and *B. j. formosus* and three contact zones of the cluster pairs of these four clusters. The oldest diverged lineage, *B. j. japonicus* and *B. j. formosus,* formed a narrow contact zone consistent with species distinctiveness. Therefore we recommend that these two subspecies be elevated to the species level. In contrast, the less diverged pairs of two clusters in *B. j. japonicus* and *B. j. formosus*, respectively, admix over a hundred kilometers, suggesting that they have not yet developed strong reproductive isolation and should be treated as conspecifics. These results contribute to resolving taxonomic confusion in Japanese toads.

Introduction

Hybrid zones are natural laboratories offering many insights into the speciation processes, thus contributing to a better understanding of evolution (Barton & Hewitt, 1985; Hewitt, 1988; Abbott et al., 2013). Hybridization following the second contact can produce different outcomes, depending on the extent to which genetic diversity and reproductive barriers have accumulated during isolation. The results are the reduction of differentiation, and fusion of gene pools, alternatively, an increase in the strength of the genetic barrier and complete reproductive isolation (Barton & Hewitt, 1985; Wu, 2001).

Hybridization is frequent and is evolutionarily significant in amphibians. There were well-described studies for the hybrid zone in amphibians of fire-bellied toads (*Bombina bombina* and *B. variegata*; e.g., Szymura & Barton, 1986, 1991; Yanchukov et al., 2006), green toads (*Bufotes viridis* subgroups; e.g., Stöck et al., 2006; Colliard et al., 2010; Dufresnes et al., 2014) and more recently those described in the European common toads (*Bufo bufo* and *B. spinosus*; (e.g., Arntzen et al., 2016; Dufresnes et al., 2020; Riemsdijk et al., 2023). Compared to many other anuran species, the Japanese toad, *Bufo japonicus* Temminck and Schlegel, 1838, has the advantage of comprising distinct genetic lineages representing different stages of the speciation process (Fukutani et al., 2022). Moreover, several contact zones of the different genetic lineages have been recognized (Fukutani et al., 2022). For amphibian cases, the extent of natural hybridization in contact zones correlated with divergence times (Hickerson, Meyer & Moritz, 2006; Dufresnes et al., 2021).

Bufo japonicus is widely distributed in the Japanese archipelago, Honshu, Shikoku, Kyushu, and some adjacent islands. This species is divided into two subspecies, *B. j. japonicus* from western Japan and *B. j. formosus* Boulenger, 1883 from eastern Japan. These two subspecies are distributed parapatrically with the boundary in the Kinki region of the central area of Japan (Matsui & Maeda, 2018). Matsui (1984) concluded that *B. j. japonicus* and *B. j. formosus* showed climatic cline in the morphometric characters, which was insufficient to distinguish them as different species because of their identity in the fundamental patterns of modes of life. However, Dufresnes & Litvinchuk (2021) recently proposed elevating *B. j. japonicus* and *B. j. formosus* to the species level, given the Miocene split estimated by mtDNA markers. However, they refrained from taxonomic changes because mitochondrial distances might not reflect actual species distances. In the other previous studies, it was suggested that the

Kinki region might be a hybrid zone of *B. j. japonicus* and *B. j. formosus* by C-banding analysis of chromosomes (Miura, 1995).

The sympatric distribution of mitochondrial haplotypes of *B. j. japonicus* and *B. f. formosus* was also found in the Kinki region (Fukutani et al., 2022). Furthermore, several contact distributions of the genetic lineages in the two subspecies were identified. These studies indicate the necessity of revealing the degree of the hybridization between the two subspecies and other genetic lineages for taxonomic revision. The valid species should exhibit significant divergence and narrow transition zones. In contrast, insufficient diverged lineages that remained conspecific should admix freely across broad genetic areas.

In this study, we applied a multiplexed ISSR genotyping by sequencing (MIG-seq: Suyama & Matsuki, 2015) to achieve a fine-scale resolution on the genetic cluster in *B. j. japonicus* and *B. j. formosus*. Moreover, we elucidated patterns of gene flow and hybridization across the contact zone. MIG-seq has been effectively used to study molecular phylogenetic taxonomy for various taxa (see Suyama et al., 2022). Therefore, we used our results to re-examine the taxonomic status of *B. j. japonicus* and *B. f. formosus*.

Materials & Methods

Sampling and Mig-seq sequencing

A total of 155 samples of *Bufo japonicus* and 13 samples of *B. torrenticola* Matsui, 1976 were collected, covering each distribution (Table S1). The Animal Experimentation Ethics Committee in the Graduate School of Human and Environmental Studies, Kyoto University provided full approval for this research (20-A-5, 20-A-7, 22-A-2). DNA was extracted from frozen or ethanol-preserved tissue samples (e.g., muscles, livers, or skin) with Qiagen DNeasy Blood and Tissue Kit following the manufacturer's instructions.

We prepared two genomic libraries and sequenced them separately for the convenience of the experiment, and the data obtained were analyzed together as described below. Library 1 included 121 samples of *Bufo japonicus* and 13 samples of *B. torrenticola*, and library 2 had 40 samples of *B. japonicus*, with six samples of *B. japonicus* overlapping in both libraries (Table S1). The two genomic libraries were prepared following the protocol described in Matsui et al. (2019) for library 1 and the protocol described in Watanabe et al. (2020) for library 2. The libraries 1 and 2 amplicons were purified and sequenced on the Illumina MiSeq Sequencer (Illumina San Diego, CA, USA) using a MiSeq Reagent Kit v3 (150 cycles, Illumina). For the convenience of the molecular experiment, two libraries were prepared and sequenced separately, and the obtained raw sequence data were combined for subsequent data analysis.

Raw sequence reads of the Mig-seq data were deposited in the DNA Data Bank of Japan (DDBJ) Sequence Read Archive (DRA) under accession number DRA016475 (BioProject ID; PRJDB15971: BioSample ID; SAMD00622809–SAMD00622982).

The raw paired-end sequences (reads 1 and 2) were filtered by fastp version 0.23.2 (Chen et al., 2018) to trim the first 14 base sequences of read 2 and primer region of read 1 and 2 and to discard the shorter reads than 80 bp and the low-quality sequences with phred quality $Q < 30$ according to Suyama and Matsuki (2015). Then, we mapped the filtered reads to the reference sequencing because the mapping can obtain more loci than de novo analysis for the MIG-seq data (Takata et al., 2021). As the reference genome sequence for Japanese toads, we used the genome assembly of their closely related species, *B. gargarizans* (RefSeq assembly accession number: GCF_014858855.1; https://www.ncbi.nlm.nih.gov/data-hub/genome/GCF_014858855.1/). The assembly contained a total of 11 chromosome-level

contigs and unplaced scaffolds. Finally, we mapped the filtered reads to the indexed reference sequencing by bwa-mem2 version 2.2.1 (Vasimuddin et al., 2019) to make SAMfiles which were converted to BAM files and sorted with a minimum mapping quality of 20 using *samtools* version 1.15 (Li et al., 2009).

Genotyping

We prepared the following datasets: i) data from the samples of *B. japonicus* and *B. torrenticola* to explore the genetic structure of Japanese toads, and ii) Data from the samples of *B. j. japonicus* and *B. j. formosus* to investigate the degree of reproductive isolation between the two subspecies. For these two datasets, we excluded the 11 samples considered to be from the artificially introduced population. Instead, we prepared a dataset iii) including these 11 samples with dataset ii) to verify the population's genetic assignments of them.

The reference-based analysis pipeline with the *gstacks* program followed by the *populations* program in Stacks v2.60 (Rochette, Rivera-Colón & Catchen, 2019) was applied to the mapped reads of all data sets to call SNPs and genotypes. The following filters were used for the *populations* program in Stacks. First, we keep variant sites with a minimum allele count of three (--min-mac 3) to ensure an allele is in at least two diploid samples (Rochette, Rivera-Colón & Catchen, 2019). Second, we set up the maximum observed heterozygosity at 0.5 (--max-obs-het 0.50) because the heterozygosity for a biallelic SNP is expected < 0.5, and SNPs with values above this threshold may belong to paralogous loci or multilocus contigs (Hohenlohe et al., 2011; Willis et al., 2017). Third, only one random SNP per locus was extracted (--write-random-snp) to avoid any influence of linkage among SNPs on the multivariate analysis (Gargiulo, Kull & Fay, 2020). For the population designation in a populations map, we set two populations corresponding to *B. japonicus* and *B. torrenticola* for dataset i). For datasets ii) and iii), we set two populations based on the *q*-value (with *q*-value = 0.5 as a boundary) at the optimal number of clusters (*K*) = 2 of Structure analysis (Pritchard, Stephens & Donnelly, 2000) on dataset i). Finally, we processed only the loci presented in at least 80% of samples in a population (*-r* = 0.80) and that presented in two populations for all datasets (*-p* = 2). For the following stacks program, the two parameters, *-r* and *-p*, were varied, and the others were common for each analysis.

Estimation of the genetic structure

To estimate the population genetic structure of *B. japonicus* and *B. torrenticola*, we performed three different methods using the SNP genotyping information and compared grouping among these methods: discriminate analysis of principal components (DAPC; Jombart, Devillard & Balloux, 2010), Structure 2.3.4 (Pritchard, Stephens & Donnelly, 2000), and principal components analysis (PCA; Cavalli-Sforza, 1966).

DAPC was performed on dataset i) in the R package *adegenet* 2.1.8 (Jombart, 2008; Jombart, Devillard & Balloux, 2010; Jombart & Ahmed, 2011). This method maximizes the variance among groups while minimizing the variation within groups without making assumptions about the underlying population genetic model. This approach transforms multilocus genotype data using PCA to derive the uncorrelated variables that are input for discriminate analysis. First, the optimal groups were assessed using the de novo clustering method, *find.cluster*, testing *K* values from 1 to 8, and the best *K* value was chosen with the Bayesian information criterion (BIC) method. This de novo clustering method and an initial DAPC using the *dapc* function were run, after which the *optim.a.score* was used to assess the

optimal number of principal components (PCs) to retain. Once the optimal number of PCs was determined, a second DAPC analysis was conducted using this value.

The program Structure 2.3.4 (Pritchard, Stephens & Donnelly, 2000) performed the analysis by an admixture model with correlated allele frequencies based Bayesian clustering method to infer the population structure. Because excessive uneven sampling likely increases the bias on the admixture proportions of Structure analysis (Toyama, Crochet & Leblois, 2020), we reduced the sample size in Yakushima and Tanegashima from dataset i) as dataset i)-2 and conducted Structure analyses. Structure analyses were carried out for the number of clusters *K* from 1 to 8, with ten runs for each *K* value. Markov chain Monte Carlo (MCMC; Metropolis et al., 1953; Hastings, 1970) iterations of 100,000 were implemented for each run after an initial burn-in of 100,000. The parallelization of Structure 2.3.4 calculations was achieved using EasyParallel (Zhao et al., 2020) to reduce computational time. The optimal number of clusters was inferred in StructureSelector (Li & Liu, 2018) with the Delta *K* (Δ*K*; Evanno, Regnaut & Goudet, 2005), MedMeaK, MaxMeaK, MedMedK and MaxMedK (Puechmaille 2016). StructureSelector integrated the CLUMPAK program (Kopelman et al., 2015) to cluster and merge data from independent runs and generate graphical representations of the results. In Structure analysis, admixed ancestry is modeled by assuming that an individual has inherited some proportion (*q*-value) of its genome from ancestors in the population (Pritchard, Stephens & Donnelly, 2000).

PCA was performed on dataset i) using the R package *adegenet* 2.1.8 (Jombart, 2008; Jombart & Ahmed, 2011), and the first two eigenvectors were plotted in two dimensions.

Moreover, we conducted Structure analysis on dataset iii) to identify the assignment of genomic clusters for the samples from introduced populations, reducing the sample size in Yakushima and Tanegashima for the above reason as dataset iii)-2. Structure analysis was carried out for the number of clusters *K* from 1 to 6, and other parameters were the same as above.

Estimate the phylogeny

We used SNAPP 1.5.2 (Bryant et al., 2012) implemented in Beast v 2.6.7 (Bouckaert et al., 2019) to estimate the phylogenetic relationships among populations identified by our clustering. We chose four individuals for each population and applied them to the stacks program (*-r* = 1.0 and *-p* = 5). We ran SNAPP for 10,000,000 iterations with mutation rates *u* and *v* = 1.0, a gamma distribution with *alpha* = 2 and *beta* = 200 for the lambda prior, and *alpha* = 1, *beta* = 250, *kappa* = 1, and *lambda* = 10 for the *snapprior*, sampling every 1,000 steps. The convergence was examined using Tracer 1.7.2 (Rambaut et al., 2018), and the result was visualized by Densitree 2.2.7 with a burn-in of 10%. The maximum clade credibility tree with posterior probability was calculated using TreeAnnotator version 2.6.7 (Bouckaert et al., 2019). To compare, we reconstructed the mitochondrial phylogenetic tree using the mitochondrial cytochrome *b* sequences from Fukutani et al. (2021) of the same individuals used for constructing the SNP tree adding the sequence of *B. g. gargarizans* as the outgroup. RAxML version 8.2.12 (Stamatakis, 2014) was employed for 1,000 bootstrap iterations with the GTRGAMMA model to infer a maximum likelihood phylogenetic tree based on the mitochondrial sequences.

Effective estimates of migration surfaces

We visualized spatial patterns of gene flow using FEEMS (Fast Estimation of Effective Migration Surfaces; Marcus et al., 2021) to assess the genomic context and geographic location

of any historical barriers to migration in *B. japonicus*. FEEMS is an improvement of EEMS (Estimated Effective Migration Surfaces; Petkova, Novembre & Stephens, 2016) and uses a Gaussian Markov Random Field model in a penalized likelihood framework. This method uses locality information and pairwise dissimilarity matrices calculated from SNP data to identify regions where genetic similarity decays more quickly than expected under isolation by distance (Petkova, Novembre & Stephens, 2016). To estimate effective migration parameters, we used as inputs the genotype data of dataset ii), as well as the coordinate information of each individual. A polygon grid was prepared using QGIS 3.28. Finally, cross-validation was performed, and the lambda with the lowest cross-validation value was used to generate the final plot.

Hybrid zone analyses

To estimate the geographic gradient of genomic differences between adjacent clusters of *B. japonicus*, we calculated the steepness of the cline of the genetic differences. Cline analyses could explain the transition between characters of interbreeding species across the hybrid zone and contribute to understanding the mechanisms maintaining species boundaries (Barton & Hewitt, 1985). Assuming similar dispersal abilities among the individuals of each cluster, and no geographic barriers to gene flow at their transitions, wide hybrid zones for the younger pairs can be estimated if they did not yet evolve significant reproductive isolation, whereas narrow transitions for the older pairs if they represent distinct species.

We fitted clines to the nuclear ancestry (Structure *q*-value) and some mitochondrial haplogroup frequency data from our previous study (Table S1; Fukutani et al., 2022) across the geographic transition between the genetic clusters identified in our study area using the R package *hzar* version 0.2-7 (hybrid zone analysis using R; Derryberry et al., 2014). The shape of each cline is modeled by combining three equations (Szymura and Barton 1986, 1991) that describe a sigmoid shape at the center of a cline (maximum slope) and two exponential decay curves on either side of the central cline (tails).

We reduced two-dimensional space (latitude and longitude) into a single-dimensional distance from the hybrid zone for *hzar*. The probable center line of admixture was estimated using R package *tess3r* version 1.1 (Caye, 2016, 2018), considered the baseline for *hzar*, and calculated the minimum distances from the baseline to individuals in QGIS 3.28. We assigned a positive or negative sign to these distances depending on individual orientation to the baseline of admixture.

Admixture proportions inferred by the Structure program (Pritchard, Stephens & Donnelly, 2000) have often been used to fit a geographic consensus cline, from which the width of the hybrid zone is estimated (e.g., Tominaga et al., 2018; Dufresnes et al., 2020b). As before, to avoid bias on the admixture proportions of Structure, we also reduced the sample size in Yakushima and Tanegashima from dataset ii).

We performed the stacks program on this subset setting four populations based on the result of DAPC on dataset ii), with *-r* = 0.80 and *-p* = 4, and conducted Structure analysis using the parameters same as above. This subset was divided into three sub-datasets i), ii), and iii), based on the *q*-value at *K* = 4 with some samples overlapped. Each sub-dataset contained individuals of two pure clusters, considering *q*-value > 0.90 as pure individuals and admixtured individuals between pure clusters. We applied the stacks program for each sub-datasets, setting three populations (two pure and one admixtured population) with *-r* = 0.80 and *-p* = 3, and conducted Structure analysis. The *q*-values on *K* = 2 for each sub-dataset were used to perform *hzar*. In addition to the three sub-datasets, we prepared sub-datasets iii)-2, which is the data

excluding samples in Shikou and Seto Inland Sea (see discussion), and performed the analysis in the same way as for the other sub-dataset.

We tested 15 different models for each cline, plus a null model with no cline, running the MCMC algorithm and using the Akaike information criterion score corrected for a small sample size (AICc; Anderson & Burnham, 2002).

These 15 models combined three trait intervals and five fitting tails. The three possible combinations of trait intervals were used to scale clines by the minimum (p_{min}) and maximum (p_{max}) values in the cline: no scale (fixed to $p_{min} = 0$ and $p_{max} = 1$), observed values (fixed to $p_{min} =$ minimum observed mean values, $p_{max} =$ maximum observed mean values), and estimated values (p_{min} and p_{max} as the free parameter). The five possible combinations of fitting tails represent the cline shapes: no tails, right tail only, left tail only, symmetrical tails, mirror tails, and both tails estimated separately.

The MCMC was performed for each model with the default values of 100,000 generations, each with a randomly selected seed and 10% of steps discarded as a burn-in. After each run, we compared the model performance using the AICc. The model with the lowest AICc score was chosen as the best-fit model to infer cline widths and centers along with a 95% confidence interval. The stability and convergence of the cline parameters of the best-fit model were assessed by visualizing MCMC traces. We plotted the maximum-likelihood clines and 95% credible cline region for the best-fit model.

Introgression

We assigned individuals in each contact zone to hybrid classes to estimate whether gene flow is an ongoing or historic admixture. First, we temporarily designated individuals with q-values > 0.98 for the $K = 2$ in Structure analysis on sub-datasets iv), v), and vi) as parental individuals for each cluster following Scordato et al. (2017). We identified ancestry-informative markers by calculating AMOVA F_{ST} for SNP between pairs of parental clusters using the stacks program on the sub-datasets, setting three populations (two parental and one admixtured population) and -*r* = 0.80 and -*p* = 3. The diagnostic loci, $F_{ST} = 1$, were selected as ancestry-informative markers segregating between each pair of parental clusters.

We used the R package INTROGRESS version 1.2.3 (Gompert & Buerkle, 2010) to calculate maximum-likelihood estimates of the hybrid index for each individual and the average heterozygosity of each individual across informative loci. We compared genomic hybrid indices with heterozygosity to determine the individual hybrid classes. Pure individuals were defined by a hybrid index of 0 or 1 because only the loci fixed in parental individuals with $F_{ST} = 1$ were used. The first-generation hybrid (F1) have an expected hybrid index of 0.5 and heterozygosity of 1.0. We considered individuals with intermediate hybrid indices (> 0.25 and < 0.75) and high heterozygosity (≥ 0.5) as recent-generation hybrids, with intermediate hybrid indices (> 0.25 and < 0.75) and low heterozygosity (< 0.5) as later-generation hybrids, and with low hybrid indices (≤ 0.25 or ≥ 0.75) as backcross to one or the other parental type following previous studies (Milne & Abbott, 2008; Scordato et al., 2017; Slager et al., 2020).

Estimation of the migration rates

Lastly, recent migration rates between parental and hybrid populations were calculated using the Bayesian inference approach by BayesAss3-SNPs v 1.1 (Wilson & Rannala, 2003; Mussmann et al., 2019). Using the sub-dataset i), ii), and iii) after applying for the stacks program with each setting three populations (two parental and one admixtured population) and -*r* = 0.80 and -*p* = 3,

BA3-SNPs -autotune v2.1.2 (Mussmann et al., 2019) was performed with the default parameters to find the mixing parameters for BA3-SNPs. BayesAss3-SNPs was conducted with ten million generations sampling every 100 generations using the predefined mixing parameters. The first one million generations were discarded as a burn-in, and the chain convergence was assessed in Tracer v 1.7.2 (Rambaut et al., 2018).

All analyses by R were conducted in R studio version 2022.07.2.576 (Rstudio Team 2022) using R version 4.2.2 (R Core Team 2022).

Results

Mig-seq analyses data

A total of 46,889,160 clean reads in 168 samples passed the quality filtering, with the average percentage of reads that passed filtering for each sample being 77.6%. Among them, 17,644,888 reads were successfully mapped to the reference genome of *B. gargarizans* in the reference-mapping approach with an average mapping quality of 27.2%.

Population structure

A total of 839 variants were identified in dataset i) of 157 samples of *B. japonicus* and *B. torrenticola*.

We retained keeping all the information (157 PCs) for the initial DAPC on dataset i). After running the initial steps, the first 21 PCs were retained following the result of the *optim.a.score* function (Fig. S1A). The BIC plot in DAPC displayed the lowest value at $K = 4$ and 5 (Fig. S1B), and both clearly identified three clusters corresponding to *B. j. formosus*, *B. j. japonicus,* and *B. torrenticola*. The results of $K = 4$ identified two subclusters within *B. j. japonicus*. In addition to that, two subclusters within *B. j. formosus* were recognized for $K = 5$, but these defined subclusters within *B. j. japonicus* and *B. j. formosus* had substantially overlapped plotts between subclusters (Fig. 1A).

A total of 570 variants were identified in dataset i)-2 of 131 samples. A Structure analysis on dataset i)-2 supported two peaks for ΔK estimation, $K = 2$ and 5 (Fig. S2A), and the number of K estimated from MedMeaK, MaxMeaK, MedMedK, and MaxMedK values was 5 (Fig. S2B). Therefore $K = 5$ can be the valid cluster number in our result, resulting in a similar grouping pattern to the DAPC (Fig. 1B). The five genetic clusters identified by DAPC and Structure analyses corresponded to northern *B. j. formosus* (NF), southern *B. j. formosus* (SF), eastern *B. j. japonicus* (EJ), western *B. j. japonicus* (WJ) and *B. torrenticola*. The cluster assignments for individuals by DAPC are shown in Table S1.

The Structure bar plot revealed that *B. torrenticola* has rare admixtures with *B. japonicus*, three samples of *B. torrenticola* had the *q*-values from 0.85 to 0.9, and one sample of *B. j. formosus* had a *q*-value of 0.09 admixed with *B. torrenticola*. These samples are likely hybrid individuals based on the *q*-value threshold following Vähä & Primmer (2006). Therefore, the one admixed sample of *B. j. formosus* was excluded from the following analysis on the *B. japonicus*.

Structure assignments also revealed hybridization between each adjacent cluster of *B. japonicus* (Fig. 1B). The proportion assignment for each cluster of *B. japonicus* changed in steps. High levels of continuous admixture were indicated across the geographic transition between NF and SF and between EJ and WJ, while the hybrid individuals were limitedly confined at the boundary between SF and EJ.

In PCA, the first PC axis explained 25.1% of the genomic covariance and separated the two subspecies, *B. j. formosus* and *B. j. japonicus* (Fig. 1C). By the second PC axis, *B.*

torrenticola was clearly split from *B. japonicus*. In addition, the second axis separated two continuous clusters within *B. j. japonicus*.

Artificially introduced population

A total of 718 variants were identified in the 128 samples of dataset iii)-2, *B. japonicus,* including the 11 samples from the artificially introduced populations in Hokkaido, Izu Islands, and the Kanto region. Two individuals in Hokkaido (Asahikawa and Hakodate) had an admixture, mainly two clusters of NF and SF, similar to those in Niijima and Kouzushima (Fig. S3). Individuals in Tokyo and Kanagawa prefectures had admixtured four clusters of NF, SF, EJ, and WJ. The individual in Oshima had three clusters of SF, EJ, and WJ, and one in Hachijojima had SF and EJ clusters.

Species tree

The phylogeny based on five clusters recovered all mitochondrial splits (Fukutani et al., 2022) but the most recent one, northern and southern Tohoku region (Fig. 2). However, the topology based on SNP conflicted with the mitochondrial tree for the clades in western Japan. The EJ and WJ clusters were derived as sister lineages, while mitochondrial topology showed that the EJ and *B. torrenticola* were sister lineages leading to the paraphyly of *B. j. japonicus*.

Effective estimates of migration surfaces

A total of 783 variants were identified in the dataset ii) 143 samples of *B. j. japonicus* and *B. j. formosus*. The estimated effective migration rates ~~evidenced~~ low migration rates between *B. j. formosus* and *B. j. japonicus* despite the absence of any geographic barrier that limits gene flow between subspecies (Fig. 3). Among *B. j. japonicus*, low migration rates were detected between Chugoku and Shikoku vs. Kyushu, and Kyushu vs. Yakushima, likely due to the presence of straits. In contrast, high migration rates were detected within them. On the other hand, among *B. j. formosus*, low migration rates were widely identified from Tohoku to Chubu, likely due to less interaction between regions than among *B. j. japonicus*.

Hybrid zone analyses

Each sub-dataset consisted of cluster pairs, sub-datasets i) NF–SF of 47 samples, ii) SF–EJ of 47 samples, and iii) EJ–WJ of 59 samples. The geographic distribution of each cluster detected by *tess3r* on each sub-dataset ($K = 2$) was not different from that of Structure analyses by SNP data. The baselines across the three contact zones are shown in Fig. 4. There were two best-supported models in *hzar* with the lowest AICc score. One model (model 1) was that scaling was fixed to the minimum value of 0 and maximum value of 1, and no exponential tails were desired for SNP data of sub-dataset SF–EJ and mtDNA data of EJ–WJ. The other model (model 2) was that the scaling was fixed to the minimum and maximum observed mean values, and no exponential tails were desired for SNP data of sub-datasets NF–SF and EJ–WJ and mtDNA data of SF–EJ and NF–SF.

Based on the SNP data, the cline width decreased from NF–SF 170 [CI (95% confidence interval): 82–362] km to EJ–WJ 162 (CI: 63–330) km, SF–EJ 29 (CI: 24–76) km (Fig. 5). The estimated center based on SNP data as the distance from the baseline were 0.6 (CI: -9.5–12) km for SF–EJ, 5.4 (CI: -42–58) km for NF–SF, and 7.0 (CI: -40–56) km for EJ–WJ.

Based on the mtDNA data, the cline width decreased from NF–SF 86 [CI (95% confidence interval): 35–223] km to EJ–WJ 75 (CI: 31–212) km, SF–EJ 39 (CI: 18–106)

km(Fig. 5). The estimated center based on mtDNA data as the distance from the baseline were 0.3 (CI: -12–15) km for SF–EJ, 23 (CI: -12–64) km for NF–SF, and -33 (CI: -75–-6.2) km for EJ–WJ.

In addition, for the sub-datasets iii)-2 EJ–WJ excluding Shikoku and Seto Inland Sea, model 2 was selected for SNP and mtDNA data. Based on the SNP data, the width was 99 (CI: 33–301) km, and the distance from the baseline was -1.2 (CI: -38–53) km, and based on the mtDNA data, the width was 79 (CI: 32–245) km and the distance from the baseline was -33 (CI: -76–-2.0) km (Fig. 5).

Introgression

We identified loci that were informative for assigning hybrid classes for each sub-dataset. There were 40 loci with $F_{ST} > 1.0$ between parental SF and EJ, and six loci for NF and SF pair and EJ and WJ pair. Comparing individual hybrid indices and average heterozygosity using these differentiated loci revealed that none of the pairs contained F1 individuals (Fig. 6). The recent-generation hybrids with high heterozygosity were detected only in NF–SF contact zone, confirming ongoing gene flow. The later-generation hybrids were detected in all contact zones, and the hybrid individuals with intermediated hybrid index values and heterozygosity of zero were identified in NF–SF and EJ–WJ contact zones, suggesting the old-origin hybrids survived. The backcrossed individuals with both parental populations were identified in NF–SF and SF–EJ contact zones, while those with one parental population were in EJ–WJ contact zone.

BayesAss directional migration

The mixing parameters for migration rates (-*m*), allele frequencies (-*a*) and inbreeding coefficients (-*f*) were determined by BA3-SNPs-autotune for each subdataset, sub-dataset NF–SF, -*m* =0.2125, -*a* =0.55, -*f* =0.1; SF–EJ, -*m* = 0.2125,-*a* =0.55, -*f* =0.1281; EJ–WJ, -*m* = 0.1563, -*a* =0.325, -*f* =0.1.

All estimated migration rates between populations are shown in Table 1. For the sub-dataset NF–SF, the self-recruitment estimate of the parental population of SF was high at > 95%, while that of the parental population of NF and hybrid population are shown to be slightly low (90–95%). The northward migration rates through the hybrid zone, from the parental SF to the hybrid (5.9%) and from the hybrid to the parental NF (3.6%), were higher than the opposite direction migration rates, from the parental NF to the hybrid (1.5%) and from the hybrid to the parental SF (1.7%).

For the sub-dataset SF–EJ, the self-recruitments within both parental populations were estimated to be high at > 95%. In contrast, the hybrid population had low self-recruitment rates at 76.2%. Correspondingly, the outward migration rates from the hybrid population into parental populations were relatively low (2.0% to parental SF and 1.3% to parental EJ efflux). In contrast, the migration rates into hybrid populations were high (16.7% from parental SF and 7.1% from parental EJ influx).

For the sub-dataset EJ–WJ, the self-recruitment of both parental populations was high at > 95%, and that of the hybrid population was intermediate value at 80.1%. The estimations of migration rates from hybrid into both parental populations were relatively low (1.3% to parental EJ and 1.4% to parental WJ efflux), while the migration rates into hybrid populations were high (7.3% from parental EJ and 12.6% from parental WJ influx). The migration rates among each parental population were estimated to be very low, ranging from 1.3% to 2.6%.

Discussion

Genetic clustering and phylogeny

The previous study showed that Japanese toads diverged into six mitochondrial lineages from the late Miocene to the middle Pleistocene (Igawa et al., 2006; Fukutani et al., 2022). Especially the two subspecies, *B. j. japonicus* and *B. j. formosus,* were recommended to elevate to species level given their Miocene split. However, these previous studies were insufficient for the taxonomic conclusion because they were based on only mitochondrial analyses (Dufresnes & Litvinchuk, 2021). Given the contact distribution of the two subspecies (Fukutani et al., 2022) and the possible presence of a hybrid zone between them (Miura, 1995), identifying the status of the hybrid zone is necessary for the systematic study of Japanese toads. First, we used SNP markers with samples covering virtually the complete distribution ranges of *B. j. japonicus*, *B. j. formosus*, and *B. torrenticola* and presented the clustering and phylogenetic relationship between the identified clusters. Second, this study presents a fine-scale analysis of gene flow across secondary contact zones of *B. j. formosus* and *B. j. japonicus*.

The consensus across independent methods suggests that $K = 5$ best describes the population structure of *B. japonicus* and *B. torrenticola*. This SNP clustering was roughly concordant with the five main mitochondrial clades in Fukutani et al. (2022), except for the lesser diverged mitochondrial clade in the Tohoku region. However, the topology of SNP was discordant with the mitochondrial phylogenetic topology. The SNP phylogenetic tree showed EJ and WJ as sister clades and supported the monophyly of *B. j. japonicus*. However, in the mitochondrial phylogenetic tree, *B. j. japonicus* was paraphyletic since *B. torrenticola* and WJ were identified as sister clades with a high node support (Fig. 2). One explanation is that *B. torrenticola* and the ancestor of EJ and WJ all diverged simultaneously. Alternatively, the discordance may stem from ancestral mitochondrial introgression between *B. torrenticola* and WJ after they diverged. These hypotheses could be tested explicitly in future phylogenetic studies.

The hybrid zone between *B. j. japonicus* and *B. j. formosus*

We found mitochondrial, and SNP marker cline positions and shapes vary for three contact zone and different patterns of gene flow. First, the hybrid zone between *B. j. formosus* and *B. j. japonicus* showed a sharp genetic transition, with concordant and coincided clines between mtDNA and SNP (Fig. 5).

The cline width depends, in part, on whether the hybrid zone is structured primarily by selection or by a neutral process (Mallet et al., 1990). The cline width without any form of selection can be calculated using the diffusion approximation from Barton & Gale (1993), $w = 2.51\sigma\sqrt{T}$, where w, cline width, T, number of generations since secondary contact, and σ, average lifetime dispersal. While the lifetime dispersal distance for *B. japonicus* is unknown, the maximum dispersal distance recorded for native *B. j. formosus* between the breeding pond and the summer home range is 0.26 km, and the generation time is three years (Kusano, Maruyama & Kaneko, 1995). At a dispersal of 0.78 km per generation, cline width exceeds the 29.4 km width of the hybrid zone in ca. 677 years of unrestricted diffusion. Considering their paleo distribution, they contacted with expansion after the last glacial period at the latest (Fukutani et al., 2022). Therefore, the contact between the subspecies is arguably much older than 677 years ago. The cline width may have been kept narrow over a long time despite no geographic barriers to dispersal, presumably through selection against hybrids, suggesting that the two subspecies form a tension zone (Key, 1968) in the Kinki region. In addition, all hybrid individuals were

classified by INTROGRESS as layer-generation hybrids or backcrosses, suggesting the relatively ancient origin of their contact.

Taxonomic revision of *B. japonicus*

Our result showed that hybridization persists over time as parentals move into the hybrid zone (Table. 1), while introgression is limited by negative selection against hybrids, allowing species to maintain their genetic distinctiveness (Barton and Hewitt 1985). These results thus call for a taxonomic revision of *B. japonicus*. Therefore, we now consider the eastern Japanese common toad *Bufo ~~japonicus~~ formosus* Boulenger, 1883 as a distinct species previously considered a subspecies of the western Japanese common toad *Bufo japonicus* Temminck and Schlegel, 1838. However, the previous study did not find intermediate forms in the Kinki region, and the morphological boundary extended more westerly to the Chugoku region (Matsui, 1984). The discordant patterns in morphological and genetic markers can be due to various factors worth further exploration.

Speciation with gene flow is common in anurans (Dufresnes et al. 2021). For example, the study on two European *Bufo* species, *B. bufo* and *B. spinosus*, which diverged in the Late Miocene, showed limited gene flow across a narrow hybrid zone (about 30 km width) in the northwest of France despite no barriers to dispersal (Arntzen et al., 2016). Despite the presence of a hybrid zone for *B. formosus* and *B. japonicus*, the identity of the parental species is distinctive and seems to have been unaffected. These two species could be considered to remain in partial reproductive isolation over a long period (Servedio and Hermisson, 2020). Cline coupling may have progressed further toward reproductive isolation after secondary contact, and it could still be ongoing throughout the hybrid zone (Harrison & Larson, 2014; Butlin & Smadja, 2018).

We also found that the geographic location of the hybrid zone between the two species is likely environment-independent. The ecological niche modeling in Fukutani et al. (2022) showed that environmental conditions are suitable for both species across the hybrid zone identified in this study, suggesting that environment-associated selection may not act directly to keep the hybrid zone. It is known that ~~the~~ many anuran speciation processes initiate through a gradual accumulation of multiple barrier loci scattered across the genome, which reduces hybrid fitness by intrinsic postzygotic isolation (Dufresnes et al., 2021). Similarly, for *B. formosus* and *B. japonicus*, it is possible that many genomic regions ~~experience~~ local barriers to gene flow and maintain steep and coincide clines rather than exogenous selection. We could identify the genomic mechanism that induced the speciation in future studies.

The hybrid zone within *B. japonicus*

Given the refugia distributions ~~in~~ Fukutani et al. (2022), the mitochondrial boundary of EJ and WJ may have been maintained at the western edge of the Chugoku region from the last glacial period to the present. Therefore, EJ and WJ likely shared refugia during the glacial period, resulting in admixture. The admixed individuals may have spread to Shikoku and surrounding islands through the Seto Inland Sea, which covered a terrestrial and freshwater environment due to the lower sea level during the glacial period until 13,000 years ago between the western part of Chugoku and Shikoku regions (Yashima, 1994).

While the strait formation between the Chugoku and Kyushu region was 8,000 years ago (Yashima, 1994), later than between the Chugoku and Shikoku region, the admixtured individuals were identified in Shikoku but not in Kyushu, possibly suggesting the asymmetric

introgression. Furthermore, the asymmetric introgression may have resulted in discordance in mtDNA and nuclear cline position between EJ and WJ, where the mitochondrial cline center is shifted about 40 km west compared to the nuclear cline center, with partially overlapping confidence intervals. The discongruity of clines inferred from different sets of molecular markers is a common phenomenon of terrestrial vertebrate hybrid zones, including amphibians (e.g., Dufresnes et al., 2014; Arntzen et al., 2017; Sequeira et al., 2020). Prezygotic or postzygotic effects could explain the discordance in mtDNA and nuclear cline position. For prezygotic effects, sex-biased asymmetries (Toews & Brelsford, 2012) and an environmental gradient acting on mtDNA (Cheviron & Brumfield, 2009), and for the postzygotic effects, Haldane's rule (Haldane, 1922; Orr, 1997) and Dobzhansky–Muller incompatibilities (Dobzhansky, 1936; Muller, 1942) can produce discordance of mtDNA and nuclear cline. Future field and genomic studies could test these hypotheses, revealing the factors that caused admixed individuals to spread mainly to the east of the mtDNA boundary at the time of secondary contact during the glacial period.

The width of the cline, including Shikoku, is wide, and using the above formula, it takes 20,490 years to reach 161.8 km, while the width of the cline, not including Shikoku, is 88.6 km, reaching 6,144 years, suggesting that selection may not act in the Shikoku region, but in the Chugoku region. Furthermore, the range of present suitable habitats for EJ and WJ in Fukutani et al. (2022) was almost consistent with the actual distribution boundaries within the Chugoku region, indicating the possibility of exogenous environmental factors. However, ~~the~~ morphological differences between EJ and WJ ~~did~~ not identify in a previous study (Matsui, 1984). Moreover, the distribution of the ~~admixture~~ individual in the Shikoku region suggests that EJ and WJ are the same species, notwithstanding the different degrees of admixture on the transect in the Chugoku and Shikoku region.

The toad population in Yakushima was once considered a different subspecies (Okada, 1928) but is now recognized to be the same species based on morphology (Matsui 1984). ~~It supports a~~ morphological ~~classification that they~~ are not monophyletic based on the mitochondrial phylogeny in ~~the~~ previous study (Fukutani et al., 2022) and not ~~clustered in~~ a single cluster in this study. There might have been interbreeding between Kyushu, Yakushima, and Tanegashima populations when the straits between Yakushima, Tanegashima~~,~~ and Kagoshima were terrestrial during the glacial period (Ikehara, 1992). The geographic isolation after the last glacial period could have led to the deviation from isolation by distance.

The hybrid zone within *B. formosus*

We identified that the hybrid zone between NF and SF was the widest in our study, which is an expected result because of their recent evolutionary histories. The widespread gene flow and recent hybridization could indicate the absence of endogenous reproductive barriers between NF and SF. Furthermore, the mtDNA and SNPs clines between NF and SF had an almost concordant center, which could also suggest ~~no~~ selection (Toews & Brelsford, 2012). In contrast, the SNP clines wider than the mitochondrial one across the transition between NF and SF~~, which~~ is ~~the~~ general ~~case~~ due to a lower effective population size of mitochondrial DNA than nuclear markers (Toews & Brelsford, 2012).

~~It can be calculated to be~~ 2,2619 years to reach the 170 km width of the SNP cline between NF and SF without selection, suggesting a prominent role for neutral processes. According to our previously predicted distributions during the glacial period, NF and SF could have shared their refugia around the southern Tohoku to northern Kanto region. The expansion

of distribution after the last glacial period probably led to widespread hybridization. An expansive hybrid zone consisting of late-generation hybrids and backcrosses is consistent overall with a prolonged period of neutral expansion. Though we did not find any asymmetry for the hybrid class assignment in the triangle plots, the results of the direction of migration were predominantly from SF to NF across the hybrid population. Consequently, this hybrid zone probably leads to forming ~~the~~ hybrid swarm in NF populations in the future rather than ~~the~~ hybrid zone movement.

Conclusion

In summary, we have presented the three hybrid zones, which were different in clines shapes. The populations with greater divergence had a sharper hybrid zone. These results in Japanese toads ~~were~~ consistent with other studies on anuran species (e.g., Dufresnes et al., 2018, 2020c). Especially the most deeply diverged populations, *B. japonicus* and *B. formosus,* had a quite sharp cline, suggesting the presence of strong selection (Mallet et al., 1990). This result contributes to resolv~~ing~~ taxonomic confusion in Japanese toads. In addition, these results provide insights into the role of the hybrid zone ~~o~~n speciation and the processes that create and maintain biodiversity.

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Figure 1

Population structure using (A) DAPC, (B) Structure, and (C) PCA based on SNPs datasets, dataset i) for DAPC and PCA, and dataset i)-2 for Structure.

The four different genetic clusters, northern *B. j. formosus* (NF), southern *B. j. formosus* (SF), eastern *B. j. japonicus* (EJ), western *B. j. japonicus* (WJ), are displayed with *B. torrenticola*. (A) DAPC plot shows the best fit for $K = 5$ clusters. The axes represent the first two linear discriminants (LD), and the dots represent individuals colored by their groups in DAPC. (B) Structure bar plots show individual ancestry to the five clusters ($K = 5$). (C) PC1 and PC2 are plotted. Each dot corresponds to an individual colored according to their genetic cluster found in DAPC. The first axis distinguishes *B. j. formosus* and *B. j. japonicus*, and the second axis distinguishes *B. japonicus* and *B. torrenticola* and reflects intraspecific structure within *B. japonicus*.

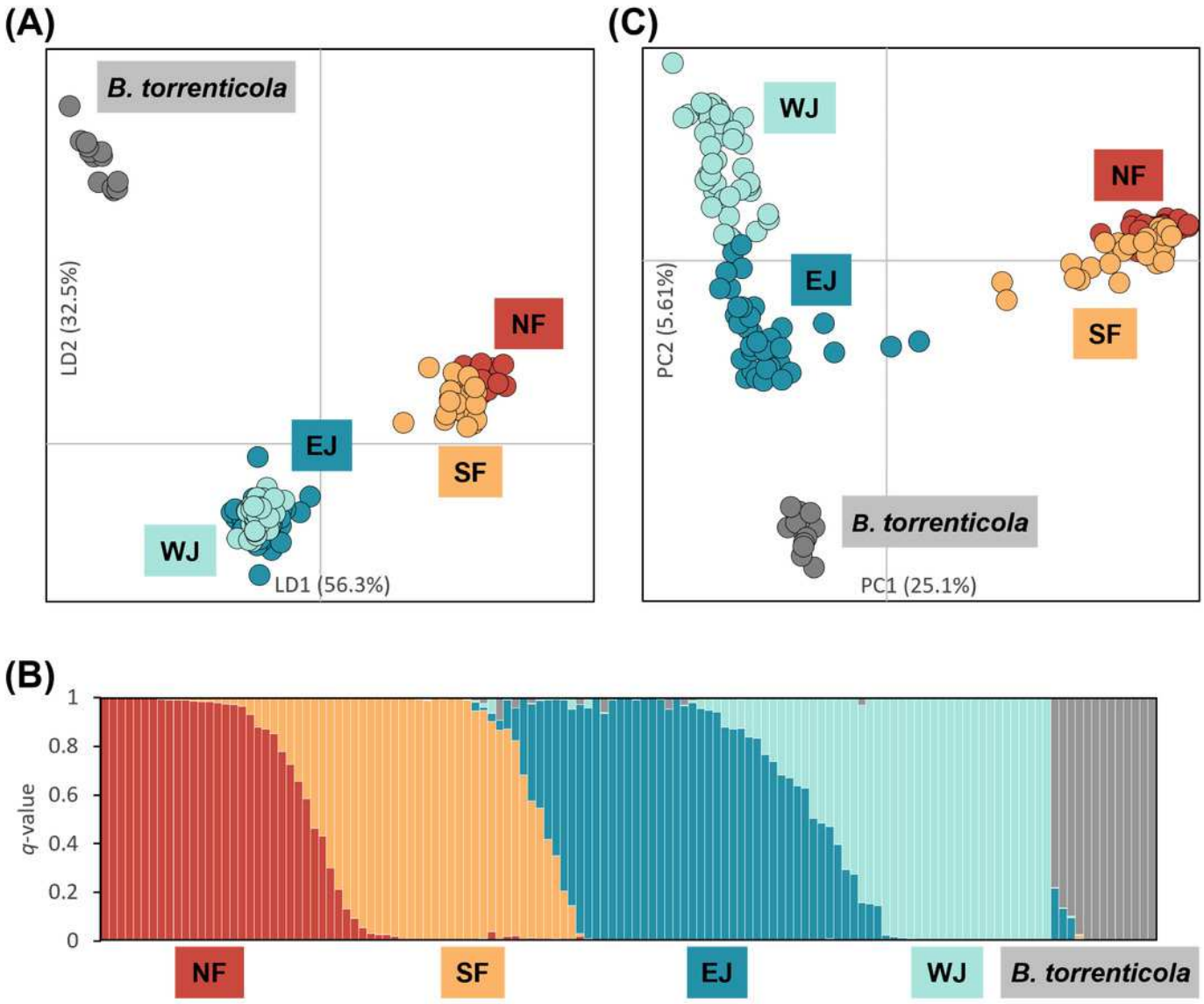
(A)
B. torrenticola
NF
EJ
SF
WJ
LD2 (32.5%)
LD1 (56.3%)
(C)
WJ
NF
EJ
SF
B. torrenticola
PC2 (5.61%)
PC1 (25.1%)
(B)
q-value
1
0.8
0.6
0.4
0.2
0
NF
SF
EJ
WJ
B. torrenticola

Figure 2

(A) Densitree diagram representing the species tree obtained from SNAPP using SNPs.
(B) The phylogenetic tree using mitochondrial cytochrome *b* sequences.

(A) All nodes were supported by posterior probabilities of 1.0. (B) Asterisks on each node indicate bootstrap supports are more than 85%.

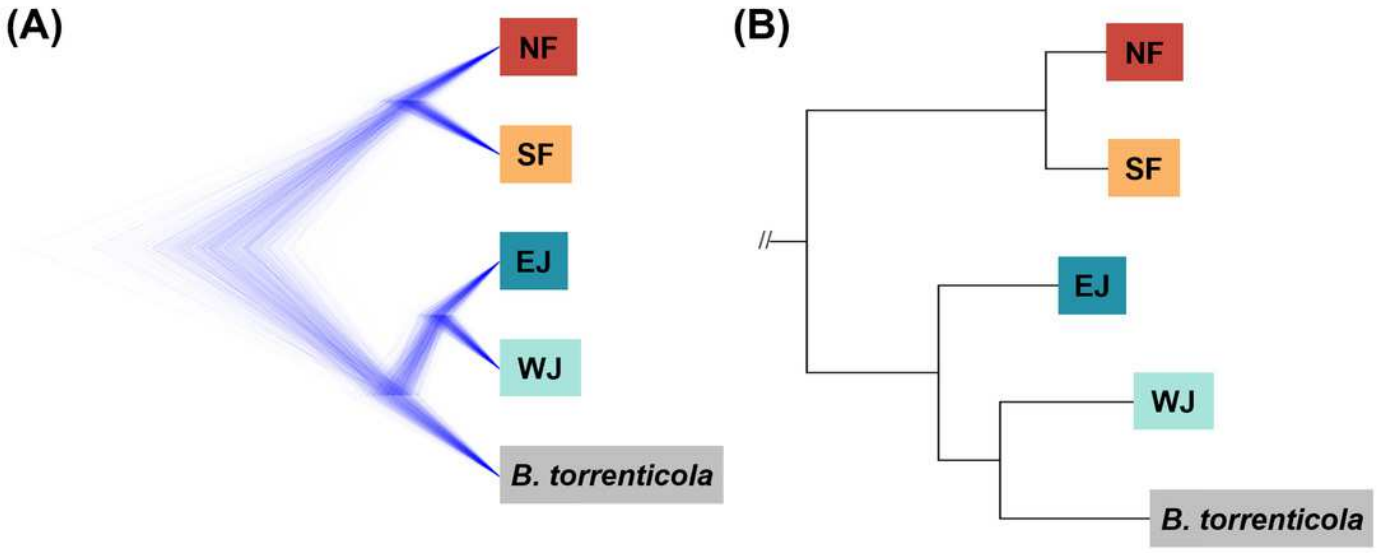

Figure 3

Effective migration rates for the lowest cross-validation lambda estimated by FEEMS (Fast Estimation of Effective Migration Surfaces) using dataset ii).

The figure shows the fitted parameters in the log scale, with lower effective migration shown in orange and higher effective migration shown in blue. Dots represent individuals.

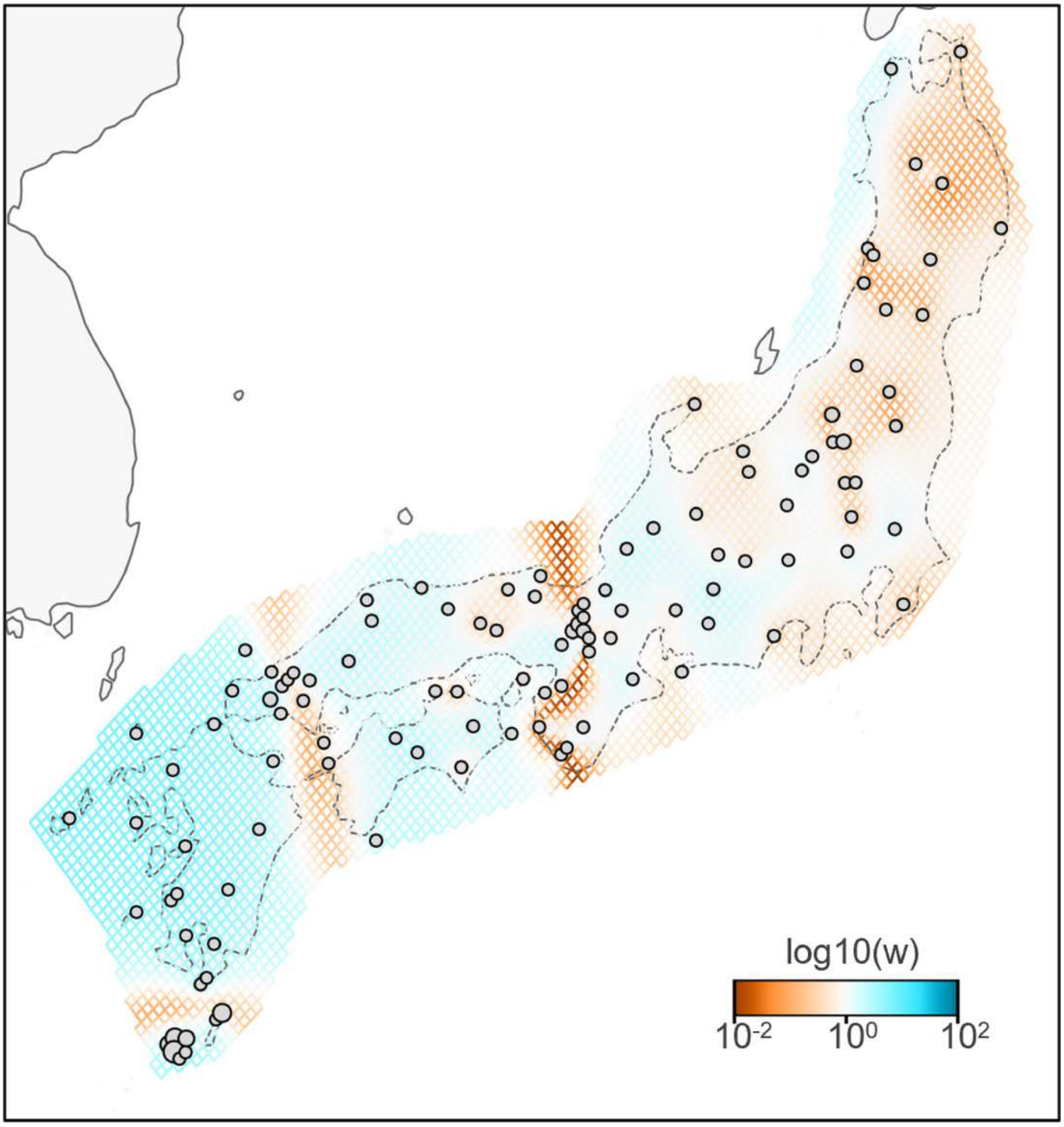
log10(w)
10-2
100
102

Figure 4

Maps showing sampling localities with pie charts for three different contact zone of sub-datasets, (A) SF–EJ, (B) NF–SF, and (C) EJ–WJ.

Pie charts show the *q*-values inferred by the Structure program for each individual. The dotted lines indicate the baselines used for *hzar*.

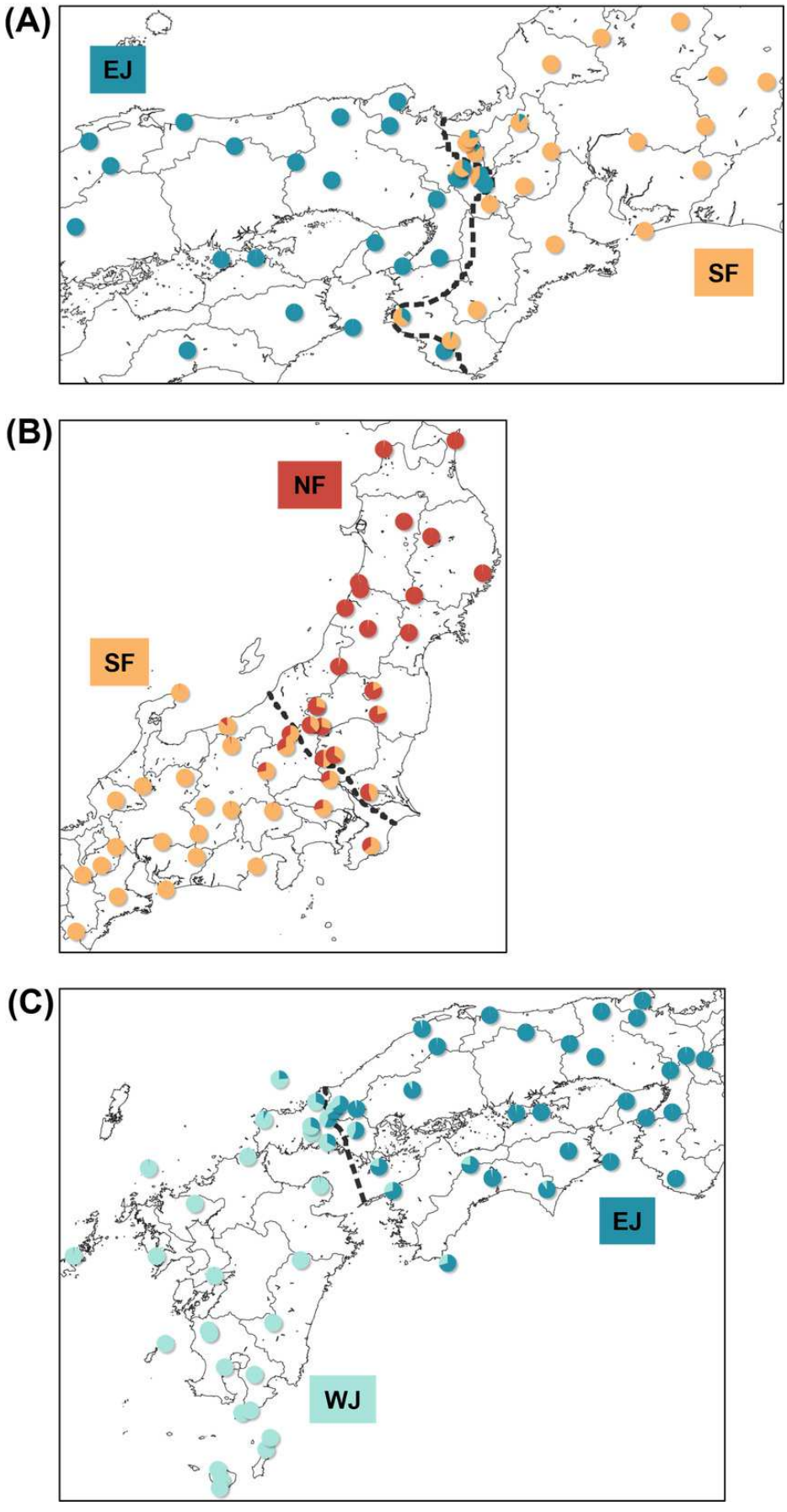
(A)
EJ
SF
(B)
NF
SF
(C)
EJ
WJ

Figure 5

The maximum-likelihood clines fitting on nuclear genomic average ancestry and mitochondrial allele frequencies along three different transect of sub-datasets, (A) SF–EJ, (B) NF–SF, and (C) EJ–WJ.

The grey areas show the 95% credible cline region. The x-axis represents distances (km) from the baselines shown in Fig. 4. Crosses indicate the observed values for individuals.

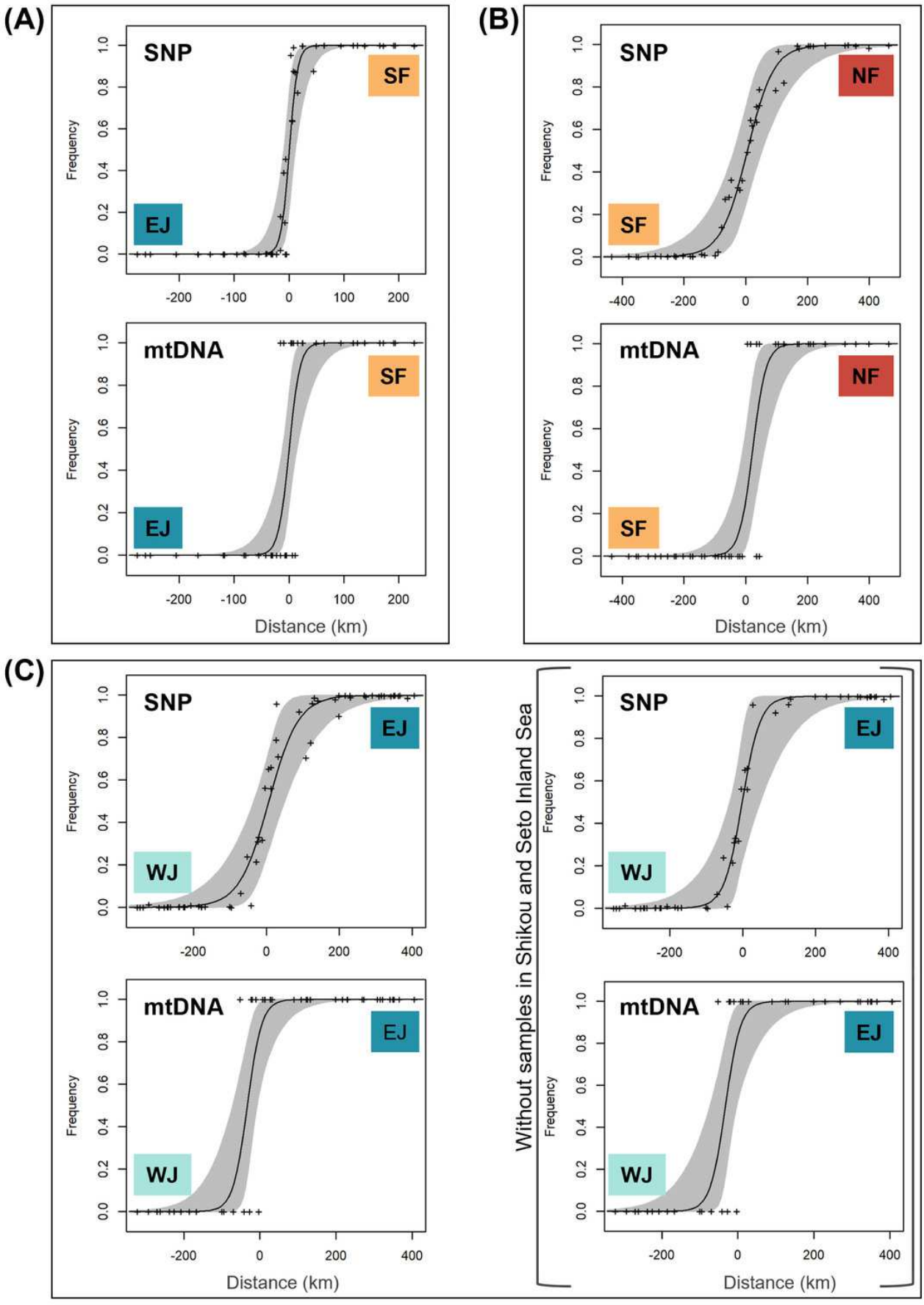
(A)
SNP
SF
EJ
mtDNA
SF
EJ
Frequency
Distance (km)
(B)
SNP
NF
SF
mtDNA
NF
SF
Frequency
Distance (km)
(C)
SNP
EJ
WJ
mtDNA
EJ
WJ
Frequency
Distance (km)
Without samples in Shikou and Seto Inland Sea
SNP
EJ
WJ
mtDNA
EJ
WJ
Frequency
Distance (km)

Figure 6

Triangle plots of the hybrid index versus heterozygosity of individuals based on selected ancestry-informative SNP markers (*F*st = 1) for sub-datasets, (A) SF–EJ, (B) NF–SF, and (C) EJ–WJ.

Individual with intermediate hybrid indices (> 0.25 and < 0.75) and high heterozygosity (≥ 0.5) was considered as recent-generation hybrid (a gray square), and those with intermediate hybrid indices (> 0.25 and < 0.75) and low heterozygosity (< 0.5) as later-generation hybrids (gray triangles). Those with low hybrid indices (≤ 0.25 or ≥ 0.75) were considered as backcross to one or the other parental type (triangles colored by parental assignments). Each colored circle indicates the pure individuals.

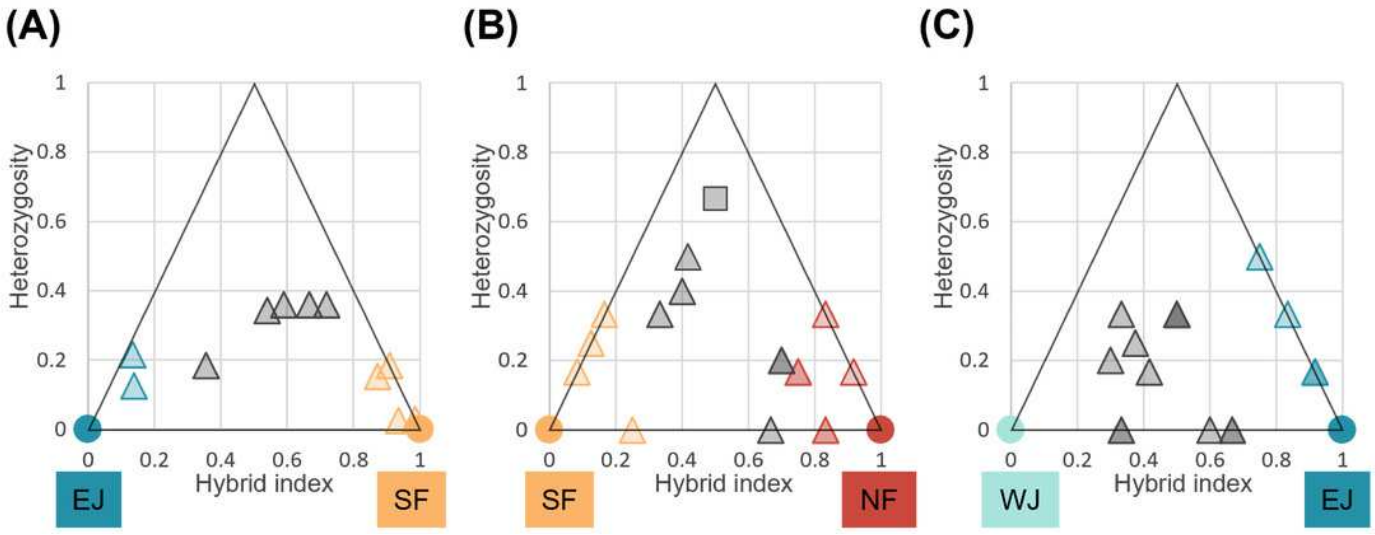

Table 1(on next page)

Estimates of migrants from BayesAss3-SNPs analyses between population clusters, (A) SF–EJ, (B) NF–SF, and (C) EJ–WJ.

The row headers represent the populations into where the individuals migrated, and the column headers represent the populations from where the migrant derived. Standard deviations of the values are given in parentheses.

(A)

		Migration from		
		parental NF	Hybrid	parental SF
Migration into	parental NF	0.9407 (0.0349)	0.0355 (0.0290)	0.0238 (0.0221)
	Hybrid	0.0152 (0.0145)	0.9262 (0.0293)	0.0586 (0.0269)
	parental SF	0.0167 (0.0158)	0.0167 (0.0158)	0.9667 (0.0218)

(B)

		Migration from		
		parental SF	Hybrid	parental EJ
Migration into	parental SF	0.9607 (0.0253)	0.0196 (0.0185)	0.0196 (0.0185)
	Hybrid	0.1667 (0.0431)	0.7619 (0.0389)	0.0714 (0.0353)
	parental EJ	0.0133 (0.0128)	0.0133 (0.0128)	0.9733 (0.0178)

(C)

		Migration from		
		parental EJ	Hybrid	parental WJ
Migration into	parental EJ	0.9683 (0.0209)	0.0133 (0.0151)	0.0159 (0.0152)
	Hybrid	0.0725 (0.0280)	0.8013 (0.0340)	0.1262 (0.0337)
	parental WJ	0.0139 (0.0133)	0.0139 (0.0133)	0.9722 (0.0184)

1