

# Response of nine triticale genotypes to different salt concentrations at the germination and early seedling stages

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The cereal crop triticale ( $\times$  *Triticosecale* Wittmack) is developed by crossing wheat (*Triticum* spp.) and rye (*Secale cereale*) plant. Based on its growth stages, it shows differential sensitivity to salinity stress. Therefore, to improve triticale productivity, this study investigated the salinity stress tolerance of different salt-tolerant triticale genotypes aiming to cultivate them on saline soils. To this end, salinity stress impact on nine triticale genotypes i.e., Zhongsi 1084, Gannong No. 2, Gannong No. 4, Shida No. 1, C6, C16, C23, C25 and C36 at germination and early seedling stages was evaluated. Each genotype was subjected to six treatments inducing control, 40, 80, 120, 160 and 200 mM NaCl treatments to study their effect on seedling and termination traits of the nine genotypes. Compared to overall mean seedling vigor index, the seedling vigor index was higher in the genotypes Zhongsi 1084 and C6 (39% and 18.1%, respectively) and lower in Gannong No.2 (41%). Increasing NaCl concentrations negatively affected germination and seedling traits. Compared to other genotypes, Zhongsi 1084 had the highest mean of germination rate, germination vigor index, germination percentage, mean daily germination and germination energy. It also showed the lowest relative salt injury. Oppositely, the relative salt injury was higher in the genotype Shida No than those in Gannong No. 2, Gannong No. 4, Shida No. 1, C16, and C36 genotypes. All genotypes exhibited desirable mean germination time except for line C6. High significant positive correlations were observed among germination rate, germination vigor index, germination percentage, mean daily germination, seedling vigor index and root length. Principal component analysis (PCA) grouped the most desirable genotypes into two clusters. Our study indicted the importance

of these traits for salt tolerant triticale genotypes selection at the germination stage. Moreover, these primary results can be used for additional breeding programs.



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41 **Keywords:** germination rate; relative salt injury; salinity; seedling stage; seedling vigor index;  
42 triticale.

43

## 44 1. Introduction

45 Cereals are an important source of food for both human and animals consumption and  
46 nutrition (Barati and Bijanzadeh, 2021). Among cereals, triticale (*Triticosecale* Wittmack) is an  
47 important cereal crop that belongs to the grass family Poaceae and was developed by hybridizing  
48 wheat (*Triticum* spp.) and rye (*Secale cereale*) (Yang et al. 2023). Two types of triticale have

49 been developed i.e., hexaploid and octoploid (Kang et al., 2016, Alatrash et al., 2022). Triticale  
50 is rich protein (Cantale et al., 2016; Hill, 1990). Therefore, it is a good source of food and feed  
51 for cattle, particularly in the forms of grazed, stored forage, silage and green fodder (Zhao et al.,  
52 2022). Moreover, it

53         The increase in population and the reduction in arable-land area are the two major threats  
54 to agricultural sustainability (Shahbaz and Ashraf, 2013). In this context, the global population is  
55 estimated to be more than 9.8 billion in 2050 (United nations, 2023). Thus the food demand will  
56 be more than double the crop production (van Dijk et al., 2021). To this end, the use of important  
57 crop such as triticale in crop rotation will help to minimize soil pests, nutrients levels reduction  
58 through leaching and increasing crop peoduction (Cao et al., 2022). Additionally, the widespread  
59 root system of triticale contributes to the soil-particle-binding effect of the grain (Demirbas and  
60 Balkan, 2020).

61         Abiotic stresses are the most significant factors limiting crop development and  
62 productivity (Zhao et al., 2020). Among these stresses, salinity stress represents the most serious  
63 threat to agricultural production, particularly in arid and semi-arid regions where soil nutrient  
64 and organic matter levels contribute to physical instability (Zhao et al., 2020). It approximately  
65 affects one billion hectares of global land worldwide, thus affecting crop production (Saade et  
66 al., 2016). Furthermore, increasing salinity stress negatively affects all traits of plants associated  
67 with germination and early seedling growth. Salinity induced toxic ions like  $\text{Na}^+$  and  $\text{Cl}^-$  affects  
68 seed germination by changing osmotic potential, lowering water uptake, causing embryonic  
69 damage and reducing seed germination, shoot elongation, and plant growth (Farooq et al., 2015;  
70 Munns and Tester, 2008; Sosa et al., 2005). Currently, approximately 20% of the total cultivated  
71 area and 33% of irrigated agricultural regions of the world are affected by salinity. Furthermore,

72 the salinized areas are increasing at a rate of 10% annually due to several reasons, including low  
73 precipitation, high evaporation, irrigation using saline water, and poor cultural practices.  
74 Moreover, approximately 50% of arable land will probably be salinized by 2050 (Jamil et al.,  
75 2011; Barati and Bijanzadeh, 2021). In arid and semi-arid regions, salinity is one of the most  
76 important environmental factors affecting uniformity in seed germination (Deng et al., 2020).  
77 Germination is a crucial stage in the development of a plant, as it influences the early growth of  
78 the seedling and its relationship with the environment and its productivity (Mbarki et al. 2020).  
79 Salinity stress induced plant growth inhibition dependent on salt concentration and duration of  
80 exposure (Guo et al., 2022). It reduces germination rate and capacity of glycophytes (Saddiq et  
81 al., 2021). This may be explained by the increase in osmotic pressure of the soil solution (Ma et al.,  
82 2022). During germination, the effects of salinity can manifest as osmotic (reversible) and  
83 deleterious (irreversible) effects (Mbarki et al. 2020). For the vast majority of crops, seeds are  
84 the means by which sophisticated genetics are transferred to the production field. Specifically,  
85 rapid and synchronous seed germination and seedling growth are vital for the development of  
86 seedlings in the field and thus are crucial to crop production (Reed et al., 2022). Seed  
87 germination determines seedling vigor and plant growth. Therefore this stage is considered a  
88 susceptible stage for plant growth (Hakim et al., 2010). Improvement in plant growth and  
89 establishment in saline soil are dependent on the salt-tolerating ability of the cultivated  
90 genotypes in early growth stages (Keshavarizi and Mohammed, 2012).

91

92 Resilience to abiotic stresses is the driving force behind the development of high-yielding  
93 and stable triticale cultivars, which in turn led to an increase in the amount of land used for  
94 triticale farming (Zhao et al., 2020). Compared to winter cereals, triticale can outproduce on low

95 fertility soils. It has a more robust root system than wheat, barley or oats, allowing it to bond  
96 light soils and extract more nutrients (Saddiq et al., 2021). Additionally, triticale is tolerant of  
97 low pH (acidic soils), sodic soils and boron-rich soils.

98 Triticale is also a moderate halophyte with high salinity threshold and it is considered a  
99 salt-tolerant species (Grieve et al., 2012). It showed salinity tolerant even up to 10 dSm<sup>-1</sup> (Ozturk  
100 et al., 2018). The salinity threshold of triticale EC (6.1 dSm<sup>-1</sup>) is higher than that of corn (2.7  
101 dSm<sup>-1</sup>), rye (5.9 dSm<sup>-1</sup>) and wheat (4.7 dSm<sup>-1</sup>). Moreover, Kotuby-Amacher et al. (2000)  
102 reported that the salinity threshold differed among various triticale species compared to other  
103 cereals. However, the relative grain yield of triticale genotypes varies at 7.3 dSm<sup>-1</sup> soil salinity.  
104 Each unit increase in soil salinity above 7.3 dSm<sup>-1</sup> reduced triticale grain yield by 2.8%, placing  
105 triticale in the salt-tolerant category (Francois et al., 1988).

106 The establishment of salt-tolerant plants is still in its infancy and shedding the light on  
107 the of salinity tolerance mechanisms. Numerous plant species, varieties and halophytes have  
108 been studied for their salt tolerance mechanisms, which have proved to be complex (Mbarki et  
109 al. 2020). Utilizing more appropriate plant cultivars should increase productivity in salinity  
110 stressed marginal areas (Cao et al., 2022). Thus, for the future of agriculture in arid and semiarid  
111 regions, genotypes selection with higher salt tolerance has become an absolute necessity  
112 (Golebiowska-Paluch and Dyda, 2023). Although triticale is considered as salinity tolerant crop,  
113 some genotypes are less tolerant at the germination stage particulry, after the three-leaf growth  
114 stage (Francois et al., 1988). There is insufficient information in the literature on the genotypes  
115 tolerance to salinity. Therefore, we aimed to determine salt stress tolerance of nine triticale  
116 genotypes at germination and early seedling stages. The objective was to select salt-tolerant

117 genotypes that can be cultivated on saline soil or after salt irrigation. This indeed will improve  
118 crop productivity and provide traits that can be used for additional breeding programs.

119

## 120 **2. Materials and methods**

### 121 **2.1. Plant genotypes and characteristics**

122 Nine triticale genotypes were used in the current study, and their names and characteristics  
123 are listed in **Table 1**. “Zhongsi 1084”, Gannong No.2’, ‘GannongNo.4’ “Shida 1” and lines “ C6,  
124 C16, C23, C25, C36” were bred by the College of Grassland Science, Gansu Agricultural  
125 University, China, using the traditional sexual hybridization techniques and a pedigree selection  
126 method (Ramadan, et al., 2023).

### 127 **2.2. Study location**

128 The experiment was conducted at Gansu Agricultural University, P. R. China. 36° 5' 26"  
129 north, 103° 41' 41" east.

### 130 **2.3. Germination conditions**

131 The seeds of the studied genotypes were sterilized using sodium hypochlorite (1%) for 30  
132 min and washed thrice using distilled water. Next, 50 seeds of each genotype were germinated  
133 on Whatman No. 1 filter paper in 9-cm Petri dishes under the following six NaCl concentrations:  
134 control, 40 mM, 80 mM, 120 mM, 160 mM, and 200 mM. The seeds were allowed to germinate  
135 in an incubator at  $20 \pm 1^\circ\text{C}$  under a 16/8-h dark/light cycle for 7 d (Warham et al., 1995); they  
136 were irrigated and washed twice daily using their corresponding treatment solution, and the filter  
137 papers were changed once every 2 d to prevent salt accumulation. After 2 d of planting, the  
138 germinated seeds were counted; the seeds were considered to have been germinated when the  
139 emerging radicle was 1 mm in length. Germination percentage was evaluated every 24 h for 5 d.

140 **2.4. Analysis of different germination and growth parameters**

141 After 7 d of planting, shoot length (SL; cm), root length (RL; cm), shoot fresh weight  
 142 (SFW; mg), root fresh weight (RFW; mg), shoot dry weight (SDW; mg), root dry weight (RDW;  
 143 mg), and root/shoot dry weight ratio (RSR) were measured. Dry weight was measured after  
 144 drying the roots or shoots at 70°C for 72 h in an oven.

145 Germination traits were calculated as follows:

146 **Germination rate (GR)** =  $\sum_{i=1}^n S_i / D_i$  (Maguire, 1962) (1)

147  $S_i$  is the germinated seeds per total seeds,  $D_i$  represents seed numbers until  $n^{th}$  day, and  $n$  is  
 148 the number of counting.

149 **Germination vigor index (GVI)** =  $\sum_{i=1}^k n_i / t_i$  (Maguire, 1962) (2)

150  $n_i$  is the percentage of seeds germinated on the  $n^{th}$  day, and  $t_i$  is the number of days  
 151 counted from the start of the experiment ( $i$ ) to the last day on which the seeds germinated ( $k$ ).  
 152 Higher values represent a more rapid rate of germination.

153 **Germination percentage (GP%)** = (Seeds germinated/Total seeds) × 100 (Manmathan  
 154 and Lapitan, 2013). (3)

155 **Mean daily germination (MDG)** = Final germination percentage/number of days to final  
 156 germination (4)

157 **Mean germination time (MGT)** =  $\sum(T_i N_i) / \sum N_i$  (Kankarla et al., 2020) (5)

158  $N_i$  is the number of the newly germinated seeds in times of  $T_i$

159 The **energy of germination (GE)** = Percentage of the germinated seeds 4 d after  
 160 planting/Total number of seeds tested (Ruan et al., 2002). (6)

161 **Relative salt injury (RSI)** = (Germination percentage of the control – Germination  
 162 percentage of the treatment)/Germination percentage of the control (7)

163        **Seedling vigor index (SVI)** = (Average shoot length + Average root length) ×  
164 Germination percentage (Abdul-Baki and Anderson, 1973)

165        (8)

166

167

## 168        **2.5. Salinity stress tolerance**

169        As a quantitative measure, stress indices can quantify the stress responses of a crop. They  
170 are easier to use and interpret than raw data. Many indices of abiotic stress tolerance have been  
171 proposed (**Table 2**) for estimating abiotic stress tolerant genotypes using a mathematical  
172 equation that describes the relationship between growth under stress and control conditions. The  
173 abiotic stress indices are classified into two types: The first type contains indices with maximum  
174 values indicating high-stress tolerance, whereas the other type includes other indices with  
175 minimum values indicating high-stress tolerance. Using these indices, the tolerant and sensitive  
176 genotypes and their stability can be identified (Parvaze and Ahmed 2018).

177

## 178        **2.6. Statistical analysis**

179        The experiment was performed as per a factorial, completely randomized design (CRD)  
180 (where Factor-1 was genotype including nine levels, and Factor-2 was salt stress treatments  
181 including six levels) with three replicates and 50 seeds in each replicate. Two-way analysis of  
182 variance (ANOVA) was used for data analysis using SAS statistical software, version 9.2. The  
183 means were compared using Duncan's multiple range test ( $P < 0.05$ ), and correlation coefficient  
184 was calculated using SPSS version 16. Principal component analysis (PCA) was performed using

185 the statistical package PAST (Hammer et al., 2001) to visualize the differences in various stress-  
186 related traits among the nine genotypes.

187 To categorize the genotypes under both control and salinity stress treatments, cluster  
188 analysis was performed using R software version 4.1.0, 2021 (R Core Team, 2021).. Euclidian  
189 metric as a distance measure was used to measure dissimilarity among the genotypes, and  
190 Ward's algorithm (Ward, 1963) was applied for grouping the genotypes.

191 Before conducting the analysis, the data were standardized due to their different scale by  
192 subtracting the mean from each value and dividing the obtained value by the standard deviation.  
193 The cubic cluster criterion (Milligan and Cooper, 1985) was used to ensure whether clusters  
194 existed. Fuzzy C-means as a soft clustering algorithm (Bezdek, 1973, 1981) was used to detect if  
195 overlapping existed between clusters. PCA is a multi-variable statistical analysis that reduces the  
196 dimensions of high-dimension data, and fewer eigenvectors explain the multivariate data  
197 (Shlens, 2005).

198

### 199 **3. Results**

#### 200 **3.1. Genotypes differentially responded to salinity stress.**

201 To study genotype sepecific responses to salt stress treatment, we performed Analysis of  
202 variance (ANOVA). Highly significant mean squares due to the genotypes and treatments, and  
203 genotypes  $\times$  treatments were detected for all studied traits except for MGT, where the mean  
204 square was non-significant for genotypes and significant for the interaction between treatments  
205 and genotypes (**Table 3**). This result indicated high variation among the studied genotypes under  
206 different salt stress treatments.

207

### 208 3.2 Mean performance of different genotypes

209 Analysis of germination traits (**Table 4**) revealed that the triticale genotype Zhongsi 1084  
210 had the highest mean GR, GVI, GP%, MDG and GE. In contrast, the genotypes Gannong No.2  
211 and Shida No. 1 exhibited the lowest mean of GR, GVI , GP% and MDG. There were no  
212 significant difference between the two cultivars. All genotypes exhibited the highest MGT  
213 except for line C6. The lowest RSI was observed for Zhongsi 1084, whereas the highest RSI was  
214 observed in case of Shida No.1. The SVI of genotypes Zhongsi 1084 and C6 was 39% and  
215 18.1%, respectively. This was higher than the overall mean SVI, whereas that of Gannong No.2  
216 was 41% less than this mean.

217 Analysis of seedling traits revealed that the mean SL of C6 and Zhongsi 1084 was 12.4%  
218 and 9.1% higher than the overall mean of SL, respectively. Whereas the means SL of Gannong  
219 No. 2 and C16 were 16.7% and 6.1% lower than the overall mean of SL, respectively. Moreover,  
220 the mean RL of Zhongsi 1084, C6, and C23 was the highest. They recorded 17.8%, 16.2%, and  
221 11.3% higher than the overall mean value, respectively. The mean RL of genotypes C36 and  
222 Gannong No. 2 was the lowest i.e., 16.8% and 12.4% lesser than the overall mean value,  
223 respectively. The highest RSRs were observed for genotypes C23 and Gannong No. 2, whereas  
224 the lowest ratio was observed for genotype C36. The highest increase in SFW compared with the  
225 overall mean SFW was observed for genotypes C6 (15.9%) and Gannong No. 4 (12.1%).  
226 Whereas the highest decrease was observed in Gannong No. 2 (13%), C16 (10%), and C25  
227 (9.7%). For RFW, the highest mean values were exhibited by C6 and Gannong No. 4. They  
228 recorded 32.1% and 20.4% more than the general mean, respectively. Meanwhile, genotypes  
229 C25, C16 and C36 exhibited the lowest mean values i.e., they were 13.1%, 11.7%, and 11%  
230 lower than the overall mean, respectively. Genotype C6 had the highest mean SDW i.e., 12.9%

231 higher than the general mean. Meanwhile, both genotypes Gannong No. 2 and C16 had the  
232 lowest mean SDW values, exhibiting 13.1% and 8.2% decreases compared to the general mean,  
233 respectively. Moreover, both genotypes C6 and Gannong No. 4 had the highest mean RDW  
234 values. They were 24.5% and 20.5% higher than the general mean. The mean RDW values of  
235 genotypes C23, C25 and Gannong No. 2 were 13.5%, 12.6%, and 11.1% was lower than the  
236 general mean.

### 237 **3.3. Differential effect of salt treatments on germination**

238         The GR of different triticale genotypes under varying salt concentrations were 1.42–4.4%  
239 (**Table 5**). The highest GR was observed in the control and 40 mM NaCl treated groups. GR  
240 gradually also reduced with increasing NaCl concentrations. GR reduced by 41% and 67.9% in  
241 the 80 mM and 200 mM NaCl treated groups, respectively. GVI was significantly different under  
242 different salinity levels, whereas mean values of different treatments were 10.28–33.54. The  
243 highest value was observed in the 40 mM NaCl treated group i.e. the lowest value was observed  
244 in the 200 mM NaCl group. Moreover, significant differences were not observed between the  
245 control and 40 mM NaCl groups. The highest reduction in GVI was observed, where 60%, 62%,  
246 and 69.2 were observed under 120, 160, and 200 mM NaCl treatments, respectively. However,  
247 the significant differences were not observed in GP % between control and 40 mM NaCl treated  
248 groups. In contrast, significant differences in GP% were observed as NaCl concentration  
249 increased from 80 mM to 200 mM and the GP% reduced by 39.8% in the 80 mM NaCl group.  
250 The highest GP% (88.04%) was observed for the 40 mM NaCl treated group, whereas the lowest  
251 GP% (28.29%) was observed in the 200 mM NaCl treated group. The highest MDG was  
252 observed in both control and 40 mM NaCl groups (12.50 and 12.58, respectively) and the lowest  
253 value was observed in the 200 mM NaCl treated group. The reduction % in MDG increased from

254 39.8% to 67.4% as NaCl concentration increased from 80 mM to 200 mM. The number of days  
255 required for germination increased from 2.48 day in the control group to 4.09 day in the 120 mM  
256 NaCl treated group. In groups treated with NaCl concentration >120 mM, the number of days for  
257 germination gradually decreased with increasing NaCl concentrations. However, significant  
258 differences were not observed among 40, 80, 160, and 200 mM NaCl groups. GE decreased from  
259 48.76% in the control group to 35.96% in the 120 mM NaCl group. However, in groups with  
260 NaCl concentration > 120 mM, GE gradually increased and it was 51.35% at 200 mM NaCl.  
261 However, significant differences in GE were not observed between the control, and 160 mM and  
262 200 mM NaCl treated groups. The RSI was negative in the 40 mM NaCl treated group and  
263 increased significantly with increasing salt concentration. It increased from 39.82% under 80  
264 mM NaCl treatment to 67.44% under 200 mM NaCl treatment. SVI decreased with increasing  
265 salt concentrations, where SVI was reduced by 27.2% in the 40 mM NaCl treated group and by  
266 95.6% in the 200 mM NaCl treated group.

267

268 Both SL and RL reduced significantly with increasing salt stress (**Table 5**). The highest  
269 mean values were observed in the control group, whereas the lowest mean values were recorded  
270 in the 200 mM NaCl group. Mean SL varied from 9.83 cm to 1.77 cm, and the reduction in SL  
271 ranged from 27.2% (40 mM NaCl) to 82% (200 mM NaCl). Mean RL varied from 6.57 cm to  
272 0.48 cm, and the reduction in RL ranged from 32.4% (40 mM NaCl) to 92.7% (200 mM NaCl).  
273 RSR gradually decreased from 0.67 in the control group to 0.3 in the 200 mM NaCl treated  
274 group; however, significant differences were not observed between the 120 and 160 mM NaCl  
275 treated groups. In the 200 mM NaCl group, >50% reduction in RSR was observed compared to  
276 that of the control group. SFW and SDW were significantly affected by salt stress. Compared to

277 that of the control group, reduction in SFW and SDW was 13.6–75.4% and 10.3–68.1% under  
278 increased NaCl concentration from 40 mM to 200 mM. Moreover, RFW and RDW were  
279 significantly reduced by salinity, where increasing NaCl concentration increased from 40 mM to  
280 200 mM reduced the RFW and RDW values by 18.4–69% and 14.5–55.6%, respectively.

281

### 282 **3.4 Interaction effects**

283 The mean performance of the different triticale genotypes under salt stress (**Figures 1**  
284 **A,B** and **2 A,B**). The highest GR, GVI, and GP% were observed for Zhongsi 1084 under 40–200  
285 mM NaCl treatments, whereas the lowest values were observed for Shida No.1 under 80–200  
286 mM NaCl treatments. Zhongsi 1084 exhibited the best MDG under 40–200 mM NaCl  
287 treatments, whereas Shida No. 1 was the most affected under high salt concentrations (120–200  
288 mM NaCl). MGT was 2.01–3.41, 2.7–3.31, 2.58–3.96, 3.23–4.59, 2.68–3.53 and 2.59–3.37 days  
289 for control, and 40 mM, 80 mM, 120 mM, 160 mM, and 200 Mm NaCl treated groups,  
290 respectively. The lowest number of days under control and 120 Mm NaCl treatments was  
291 observed in genotype C6. Gannong No. 4 exhibited the best GE under control treatment  
292 (55.97%), Zhongsi 1084 under 40 and 120 mM NaCl treatments (52.84 and 48.03%,  
293 respectively), C6 under 80 mM treatment (46.06%) and Shida No. 1 under 160 mM and 200 mM  
294 NaCl treatments (56.5 and 57.5%, respectively). RSI increased with increasing salt  
295 concentrations. The lowest percentage of injury was observed in Zhongsi 1084 i.e., 10.23, 24.18,  
296 25.36 and 38% under 80, 120, 160, and 200 mM NaCl treatments, respectively. Meanwhile, the  
297 highest percentage of injury was observed in Shida No. 1 (57.57, 82.17, 87.38, and 87.21%  
298 under 80, 120, 160, and 200 mM NaCl treatments, respectively). For SVI, the most desirable  
299 genotypes were Zhongsi 1084 and Gannong No. 4 under control treatment; Zhongsi 1084 and C6

300 under 40 mM, 120, and 200 mM NaCl treatments; Zhongsi 1084 and C23 under 80 mM NaCl  
301 treatments; and both Zhongsi 1084 and C25 under 160 mM NaCl treatments. In contrast, Shida  
302 No. 1 was the most affected genotype under high salt concentrations.

303 Furthermore, Zhongsi 1084 had the highest mean SL under control and 40 mM NaCl  
304 treatments, but the lowest mean SL under 160 and 200 mM NaCl treatments. C6 had the mean  
305 highest SL under 80 and 120 mM NaCl treatments. C16 also had the highest mean SL under 160  
306 and 200 mM NaCl treatments. The lowest mean SL under control and 40 mM, 80 mM, and 120  
307 mM NaCl treatments were exhibited by Gannong No. 2. The mean RL was 8.57–5.13 and 5.45–  
308 3.61 cm for control and 40 mM NaCl groups, respectively. It gradually decreased to 0.97–0.62  
309 and 0.61–0.29 cm for 160 mM and 200 Mm NaCl treated groups, respectively. The mean RSR  
310 decreased with increasing salt concentrations. The mean ratios ranged from 0.79 to 0.53 under  
311 control and from 0.42 to 0.20 under 200 mM NaCl treatment. Shida No. 1 had the highest RSRs  
312 under 160 and 200 mM NaCl treatments, whereas C16 had the lowest ratios. Both C6 and Shida  
313 No. 1 were the best genotypes as per their mean SFW under 0, 40 and 80 mM NaCl treatments,  
314 whereas both Gannong No. 4 and C6 were the best genotypes under 120, 160, 200 mM NaCl  
315 treatments as per their mean SFW. C6 and Gannong No. 4 were the best genotypes as per mean  
316 RFW under 0-120 mM NaCl treatments, whereas C6 and Gannong No. 2 were the best under  
317 160 and 200 mM NaCl treatments. Furthermore, the highest mean SDW was observed for Shida  
318 No. 1 and C6 under 0-80 mM NaCL treatments, for C6 under 120 mM and 160 mM NaCl  
319 treatments and for Gannong No. 4 under 200 mM NaCl treatment. In contrast, the lowest mean  
320 SDW under high salt concentrations was exhibited by Zhongsi 1084 and C36. Gannong No. 4  
321 and C6 were the most desirable genotypes under 0-120 mM salt treatments for RDW. It was also  
322 reported that different salinity concentrations caused considerable effects on GP%, GR, total dry

323 weight and all seedling traits in all studied genotypes. Similar results for the interaction between  
324 salt stress and genotypes have been reported by Kandil et al. (2012).

### 325 **3.5 Phenotypic correlation**

326 Phenotypic correlation coefficients among the studied traits (**Table 6**). The highest  
327 positive correlation ( $r = 1.00$ ) was observed between GP% and MDG. High significant positive  
328 correlations were observed among GR, GVI, GP%, MDG, SVI, and RL. Significant positive  
329 correlations were also observed among RL, SFW, RFW and RDW. SVI was significantly  
330 positively correlated with RL. GVI was significantly positively correlated with GE and SL.  
331 Significant positive correlations were observed between GE, SVI, and SL, and between SL and  
332 RL. Positive but non-significant correlations were observed between germination traits GR, GVI,  
333 GP%, MGT, GE and SVI and seedling traits RSR, SFW, RFW, SDW and RDW. In contrast,  
334 highly significant negative correlations were observed between RSI and GR, GVI, GP%, and  
335 MDG. Significant negative correlations were also observed between MGT and RDW and  
336 between RSI and both SVI and RL.

337

### 338 **3.6 PCA**

339 In the current study, PCA classified the nine genotypes into four clusters based on their  
340 mean performance under different NaCl treatments (**Figure 3**). The first cluster was found in the  
341 1<sup>st</sup> quadrant, which included triticale genotypes C6 and Gannong No. 4. Both genotypes scored  
342 the highest values for the seedling traits SFW, RFW, SDW, RDW and high values for RL, SVI,  
343 MDG and GVI. The second cluster was found in the 2<sup>nd</sup> quadrant and included the genotypes  
344 Zhongsi 1084, C23, and C25. These genotypes had high mean GR, GVI, GP%, MDG, SVI, SL  
345 and RL and low RSI. The third cluster was found in the 3<sup>rd</sup> quadrant and included Gannong No .

346 2 and C16 genotypes, whereas the fourth cluster was found in the 4<sup>th</sup> quadrant and included both  
347 Shida No. 1 and C36. The genotypes in the third and the fourth clusters had the lowest mean GR,  
348 GVI, GP%, MDG, SVI, and RL. These results suggested considerable variability for salt  
349 tolerance in the studied triticale genotypes.

350 ts in **Tables 7 and 8** reveal that Gannong No. 4 was the most tolerant genotype with an  
351 average rank (AR) equal to 2.12 (**Figure 4**). However, Zhongsi 1084 was the least tolerant  
352 genotype (AR = 8.04). Both Gannong No. 2 and C25 were moderately tolerant as their ARs were  
353 4.29 and 4.62, respectively. Higher AR suggested the lower tolerance of the genotype (**Table 8**).

### 354 **3.8 Cluster analysis**

355 SFW and RFW were used to construct a distance matrix and to generate a tanglegram  
356 exhibiting dissimilarity among all genotypes under control and the treatment with the highest salt  
357 concentration (200 mM) (**Figure 5**). The fuzzy C-means method elucidated that low overlap  
358 existed between clusters, thus hard clustering methods were applied to construct the tanglegram  
359 (**Figure 5**). Six hard clustering methods were compared using an agglomerative coefficient to  
360 choose the most accurate method for clustering the data, which were average, generalized  
361 average, single, and weighted.

362

363 The valued of agglomerative coefficients were 0.76, 0.81, 0.53, 0.77, 0.85, and 0.88  
364 respectively, under control treatment, whereas under 200 mM NaCl treatment, they were 0.68,  
365 0.72, 0.55, 0.73, 0.77, and 0.81 respectively. These results reveal that Ward's method had the  
366 highest coefficient compared to those of the other five methods under control and 200 mM NaCl  
367 treatments. Therefore, Ward's method was chosen to conduct cluster analysis. To identify the  
368 optimum number of clusters in the data, 30 internal validation indices were selected and screened

369 (Charrad et al., 2014). As shown in **Figure 5**, all genotypes were separated into two clusters  
370 under control and 200 mM NaCl treatment groups (**Table 9**). The structure of the clusters  
371 changed markedly when the genotypes were subjected to 200 mM NaCl treatment except for  
372 genotypes Gannong No. 4 and C6, which migrated from cluster 1 under control to cluster 2 under  
373 the saline treatment because they were more tolerant than the other members of their cluster.

374 Heatmaps elucidate the relationship between the genotypes and the studied traits based on  
375 standardized (scaled) data using a color scale under control and 200 mM NaCl treatments  
376 (**Figures 6 and 7**). Before drawing the heatmap, the data were standardized by subtracting the  
377 mean from each value and dividing the obtained value by the standard deviation. Genotype C6  
378 had the highest mean SFW and SDW in the control group, whereas genotype Gannong No. 4 had  
379 the highest mean SFW and SDW under the highest salinity treatment (200 mM). These results  
380 demonstrated that Gannong No. 4 was the most tolerant genotype. The lowest mean SFW and  
381 SDW under control treatment were observed in C16, whereas Zhongsi 1084 exhibited the lowest  
382 mean SFW and C26 had the lowest mean SDW under 200 mM NaCl treatment. Moreover, GP%  
383 of genotypes Gannong No. 4 and Gannong No. 2 was the highest and the lowest, respectively,  
384 under control treatment.

385 In contrast, the genotypes Zhongsi 1084 and Shida No. 1 were the highest and the lowest,  
386 respectively, under 200 mM. The genotype Zhongsi 1084 had higher values of germination traits  
387 under the highest salinity treatment. However, it had the lowest mean SFW, RFW, SL, and RSI.  
388 Gannong No. 4 had higher values of germination traits under control treatment. The heatmap  
389 does not reveal any association between germination traits and the tolerance indices of the  
390 genotypes, except for MGT, which was negatively associated with the tolerance of the  
391 genotypes.

392

393

## 394 4. Discussion

395 These results elucidated that the response to salinity differed among the studied triticale

396 genotypes. Genotypes Zhongsi 1084, C6, C23, and C25 showed the highest salinity stress

397 tolerance based on germination traits. Meanwhile, C6 and Gannong No. 4 were the best

398 genotypes based on their seedling traits. In contrast, the germination traits of Gannong No. 2

399 and Shida No. 1 genotypes were the most affected by salinity stress, whereas the seedling

400 traits of Gannong No. 2 were the most affected by salinity stress. These results indicated that

401 the effect of salinity on triticale at germination and early seedling stage varied between

402 different genotypes. The effect of soil salinity on plants is associated with their growth stage

403 (Shannon, 1997). Seed germination and seedling establishment are the most salt-sensitive

404 stages of plants (Ashraf and Foolad, 2005). Atak et al. (2006) also reported that the delay in

405 germination was mainly due to high  $\text{Na}^+$  accumulation in the seeds and not due to osmotic

406 stress. (Kandil et al., 2012) investigated the impact of salt stress under different salinity

407 levels on eleven bread wheat varieties (*Triticum aestivum* L.) and reported that wheat

408 cultivars significantly varied in the means of the final GP%, GR, SVI, SL, RL, SFW, RFW,

409 SDW, and RDW.

410 This was in accordance with the results reported by Akgun et al. (2011). They studied the

411 effects of different salt concentrations ( $\text{EC} = 3.9, 6.1, 8.3, 10.5, 14.9, 19.3, 25.0 \text{ dSm}^{-1}$ ) on

412 germination and seedling traits of triticale and reported that GR, SL, RL and dry weights of

413 the green parts and roots considerably decreased with increasing salt concentrations. Kandil

414 et al. (2012) and Atri et al., (2018) reported that with increasing salt concentrations, the

415 average values of germination and seedling growth traits gradually reduced. Francois et al.  
416 (1988) reported that at soil salinity of up to 11.6 dSm<sup>-1</sup>, significant effect on final GP% of  
417 triticale was not observed. However, at >6.0 dSm<sup>-1</sup> salinity, delay in seed germination was  
418 observed. They also reported that the final germination rate was reduced by 17% upon  
419 increasing salinity levels up to 20.5 dSm<sup>-1</sup>.

420 In accordance with these results, Alom et al. (2016) reported that the salt tolerance index  
421 for seedling dry weight of wheat genotypes irrigated with saline water (15 dSm<sup>-1</sup>) was positively  
422 correlated with salt tolerance indices GR, GVI, SL, and RL suggesting their role as selection  
423 criteria. Aflaki et al. (2017) investigated the effect of salinity on germination of different  
424 genotypes of wheat and found that MDG exhibited the highest correlation with GP%. In a  
425 previous study, PCA classified different genotypes of wheat and soy beans into three groups, i.e.,  
426 salt tolerant, moderately salt tolerant, and salt susceptible, based on the performance of these  
427 genotypes under different salt concentrations at the early seedling stage (Saboora et al., 2006;  
428 Shelke et al., 2017).

429

## 430 **Conclusions**

431 In the present study, the mean performance of most traits gradually decreased with  
432 increasing salt concentration. Mean germination time increased upon increasing NaCl  
433 concentration to 120 mM, then decreased with increasing NaCl concentrations. Non-significant  
434 differences were observed under control and 40 mM treatments germination rate, germination  
435 vigor index, germination percentage and mean daily germination. Genotype Zhongsi 1084  
436 exhibited the best performance for treatments germination rate, germination vigor index,  
437 germination percentage, mean daily germination, Mean germination time (days), Germination

438 energy, Seedling vigor index and Root length (cm). Line C6 and genotype Gannong No.4  
439 resulted in best performance for shoot length (cm), root length (cm)shoot fresh weight (mg),  
440 root fresh weight (mg), shoot dry weight (mg) and root dry weight (mg). Highly significant  
441 positive correlations were observed between germination rate and (the germination vigor index,  
442 germination percentage, mean daily germination). Also, highly significant positive correlations  
443 were observed between the germination vigor index and (germination percentage, mean daily  
444 germination). Moreover highly significant positive correlations were observed between  
445 germination percentage and mean daily germination . PCA divide the studied genotypes into four  
446 clusters. The most desirable genotypes were gathered into clusters 1 and 2, whereas other  
447 genotypes were grouped into clusters 3 and 4.

448

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#### 450 **Author Contributions**

451 All authors have contributed equally to the research and analysis of the various results sections  
452 within the review. All have corrected and modified the different versions of the manuscript as  
453 prepared by the corresponding and senior authors. All authors read and approved the final  
454 manuscript.

#### 455 **Conflict of Interest Statement**

456 The authors declare that the research was conducted in the absence of any commercial or  
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#### 458 **Data Availability Statement**

459 The authors confirm that the data supporting the findings of this study are available within the  
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**Table 1** (on next page)

List of genotypes and names, of triticale investigated in this study

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<b>Number</b>	<b>Genotype names</b>
1	Zhongsì 1084 (Chinese Triticale cultivar)
2	Gannong No. 2 (Chinese Triticale cultivar)
3	Gannong No. 4 (Chinese Triticale cultivar)
4	Shida No. 1 (Chinese Triticale cultivar)
5	C6 (Triticale line bred by GASU)
6	C16 (Triticale line bred by GASU)
7	C23 (Triticale line bred by GASU)
8	C25 (Triticale line bred by GASU)
9	C36 (Triticale line bred by GASU)

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GASU: Gansu Agricultural University of P.R. China

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**Table 2** (on next page)

Abiotic stress screening indices

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Index	Formula	Reference
Indices with maximum values corresponding to more tolerant		
Mean productivity (MP)	$(Y_S + Y_{NS})/2$	Rosielle and Hamblin (1981)
Geometric mean productivity (GMP)	$(Y_{NS})^{(1/2)} \times Y_S$	Fernandez (1992)
Harmonic mean (HM)	$2 \times (Y_S \times Y_{NS})/(Y_S + Y_{NS})$	Bidinger et al. (1987)
Stress Tolerance Index (STI)	$(Y_S \times Y_{NS})/(Y_{NS-m})^2$	Fernandez (1992)
Yield index (YI)	$Y_S/Y_{S-m}$	Gavuzzi et al. (1997)
Modified stress tolerance index-I (MSTI1)	$((Y_{NS})^2/(Y_{NS-m})^2) \times ((Y_S \times Y_{NS})/(Y_{NS-m})^2)$	Farshadfar and Sutka (2003)
Modified stress tolerance index- II (MSTI2)	$((Y_S)^2/(Y_{S-m})^2) \times ((Y_S \times Y_{NS})/(Y_{NS-m})^2)$	Farshadfar and Sutka (2003)
Yield stability index (YSI)	$Y_S/Y_{NS}$	Bousslama and Schapaugh (1984)
Relative stress index (RSI)	$(Y_S/Y_{NS})/(Y_{S-m}/Y_{NS-m})$	Fischer and Wood (1979)
Drought index (DI)	$(Y_S \times (Y_S/Y_{NS}))/Y_{S-m}$	Bidinger et al. (1987)
Stress/non-stress productivity index (SNPI)	$((Y_{NS}+Y_S)/(Y_{NS}-Y_S))^{(1/3)} \times (Y_{NS} \times Y_S \times Y_S)^{(1/3)}$	Moosavi et al. (2008)
Relative efficiency index (REI)	$(Y_S \times Y_{NS})/(Y_{S-m} \times Y_{NS-m})$	Ramirez-Vallejo and Kelly (1998)
Mean relative performance (MRP)	$(Y_S/Y_{S-m}) + (Y_{NS}/Y_{NS-m})$	Ramirez-Vallejo and Kelly (1998)
Golden mean (Gm)	$(Y_{NS} + Y_S) / (Y_{NS} - Y_S)$	Moradi et al. (2012)
Indices with minimum values corresponding to more tolerant genotype		
Tolerance index (TOL)	$Y_{NS} - Y_S$	Rosielle and Hamblin (1981)
Stress susceptibility Index (SSI)	$(1 - (Y_S/Y_{NS})) / (1 - (Y_{S-m}/Y_{NS-m}))$	Schnieder et al. (1997)
Stress susceptibility percentage index (SSPI)	$(Y_{NS} - Y_S) / (2 \times Y_{NS-m})$	Moosavi et al. (2008)
Yield reduction (YR)	$1 - (Y_S/Y_{NS})$	Choukan et al. (2006)
Abiotic stress tolerance index (ATI)	$((Y_{NS} - Y_S) / (Y_{NS-m}/Y_{S-m})) \times (Y_{NS} \times Y_S)^{(1/2)}$	Moosavi et al. (2008)
Mean productivity index (MPI)	$(Y_{NS} - Y_S) / 2$	Rosielle and Hamblin (1981)
Schnieder's stress susceptibility index (SSSI)	$1 - (Y_S/Y_{NS}) - (1 - (Y_{S-m}/Y_{NS-m}))$	Schnieder et al. (1997)
Sensitivity drought index (SDI)	$(Y_{NS} - Y_S) / Y_{NS}$	Farshadfar and Javadina (2011)

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**Table 3** (on next page)

Mean square estimates for the parameters of triticale genotypes under different salt treatments

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Source of Variance	Treatment	Genotype	Treatment × Genotype	Error
degree of freedom	5	8	40	108
Germination rate	47.51**	8.59**	0.365**	0.06
Germination vigor index	2891.46**	470.32**	18.675**	4.16
Germination percentage (%)	18758.28**	3197.63**	170.011**	29.56
Mean daily germination	382.77**	65.25**	3.475**	0.60
Mean germination time (d)	7.67**	0.48 <sup>ns</sup>	0.413*	0.27
Germination energy (%)	1164.17**	168.13**	55.973**	22.18
Relative salt injury	25066.84**	2360.53**	243.20**	43.22
Seedling vigor index	881.77**	30.83**	4.481**	0.58
Shoot length (cm)	268.10**	3.44**	1.43**	0.29
Root length (cm)	152.64**	1.90**	0.67**	0.18
Root/shoot ratio	0.54**	0.02**	0.01**	0.01
Shoot fresh weight (mg)	382627.79**	11844.21**	2341.06**	911.49
Root fresh weight (mg)	88069.30**	6957.79**	1506.71**	515.80
Shoot dry weight (mg)	5271.16**	171.06**	38.94**	13.39
Root dry weight (mg)	1217.71**	143.46**	18.28**	7.11

5        \*\*: highly significant differences at 0.01 level; \*: significant differences at the 0.05 level; and ns: no  
6        significant differences

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**Table 4**(on next page)

The overall mean performance of different studied triticale genotypes under six salt treatments

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Genotypes		Zhongs	Gannon	Gannon	Shida	C6	C16	C23	C25	C36	Mean
Traits		1084	g No.2	g No.4	No.1						
Germination traits	Germination rate	3.93a	1.85e	2.89c	1.91e	3.26b	2.40d	3.14b	3.19b	2.33d	2.77
	Germin. vigor index	28.83a	13.76f	22.02d	14.64f	24.98b	17.50e	23.30cd	23.92bc	17.44e	20.71
	Germin. (%)	79.15a	38.49f	56.65c	39.06f	63.32b	49.52d	63.10b	62.64b	45.74e	55.3
	Mean daily germin.	11.31a	5.50f	8.09c	5.58f	9.05b	7.07d	9.01b	8.95b	6.53e	7.9
	Mean germin. time (days)	3.14ab	3.26a	2.94ab	3.2ab	2.85b	3.29a	3.28a	2.96ab	3.05ab	3.11
	Germin. energy	49.26a	42.66b	43.78b	45.43b	49.09a	39.42c	44.62b	46.17ab	44.93b	45.04
	Relative salt injury	0.19 g	0.50 c	0.51 c	0.64 a	0.31 f	0.47 cd	0.38 e	0.43 d	0.58 b	0.45
	Seedling vigor index	7.57a	3.22e	6.03bc	4.68d	6.44b	4.59d	6.13bc	5.85c	4.51d	5.45
	Shoot length (cm)	5.42ab	4.14f	5.27abc	5.16bcd	5.58a	4.67e	4.97cde	4.80de	4.85de	4.99
	Root length (cm)	3.11a	2.31de	2.86ab	2.55cd	3.07ab	2.46cde	2.94ab	2.75bc	2.20e	2.69
	Root / shoot ratio	0.48ab	0.51a	0.47ab	0.48ab	0.45b	0.45b	0.51a	0.49ab	0.40c	0.47
	Shoot fresh weight (mg)	258.99c	223.25e	287.44ab	271.74bc	297.25a	230.84de	251.52cd	231.62de	255.94c	256.51
	Root fresh weight (mg)	122.03bc	114.36bc	148.09a	124.95b	162.56a	108.62bc	111.40bc	105.83c	109.49bc	123.04
	Shoot dry weight (mg)	33.53b	29.65c	37.18a	37.78a	38.52a	31.32bc	33.33b	31.86bc	33.99b	34.13
	Root dry weight (mg)	20.57b	17.32c	23.48a	20.25b	24.26a	17.55c	16.85c	17.04c	18.10c	19.49

2 Values followed by the different letter(s) are significantly different from each other by Duncan's  
3 multiple range test at 5% level of probability

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**Table 5** (on next page)

The overall mean performance of the six salt treatments

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Treatments	Control	40 mM	80 mM	120 mM	160 mM	200 mM	Mean
Germination traits							
Germination rate	4.40a	4.40a	2.60b	2.04c	1.74d	1.42e	2.77
Germin. vigor index	33.39a	33.54a	19.65b	14.69c	12.71d	10.28e	20.71
Germin. (%)	87.50a	88.04a	52.66b	39.63c	35.47d	28.49e	55.30
Mean daily germin.	12.50a	12.58a	7.52b	5.66c	5.07d	4.07e	7.90
Mean germin. time (days)	2.48c	2.94b	3.15b	4.09a	3.03b	2.96b	3.11
Germin. energy (%)	48.76ab	47.61b	37.51c	35.96c	49.05ab	51.35a	45.04
Relative salt injury	0.00e	-0.62e	39.82d	54.71c	59.47b	67.44a	36.80
Seedling vigor index	14.46a	10.53b	3.96c	2.04d	1.06e	0.64f	5.45
Shoot length (cm)	9.83a	7.42b	5.14c	3.60d	2.15e	1.77f	4.99
Root length (cm)	6.57a	4.44b	2.32c	1.50d	0.85e	0.48f	2.69
Root / shoot ratio	0.67a	0.60b	0.45c	0.40d	0.40d	0.30e	0.47
Shoot fresh weight (mg)	411.88a	355.74b	298.72c	233.47d	130.17e	101.20f	255.20
Root fresh weight (mg)	208.23a	169.93b	128.82c	89.86d	69.42e	64.56e	121.80
Shoot dry weight (mg)	51.09a	45.82b	40.50c	30.28d	20.75e	16.32f	34.13
Root dry weight (mg)	29.27a	25.04b	20.89c	15.54d	13.20e	13.01e	19.49

Values followed by the different letter(s) are significantly different from each other by Duncan's multiple range test at 5% level of probability

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**Table 6** (on next page)

Phenotypic correlation coefficients among the studied traits

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Traits	GR	GVI	GP	MDG	MGT	GE	RSI	SVI	SL	RL	RSR	SFW	RFW	SDW
GVI	0.997**													
GP	0.996**	0.988**												
MDG	0.996**	0.988**	1.000**											
MGT	-0.4	-0.462	-0.333	-0.333										
GEN	0.652	0.681*	0.600	0.600	-0.569									
RSI	-0.881**	-0.864**	-0.900**	-0.900**	0.200	-0.566								
SVI	0.952**	0.960**	0.944**	0.944**	-0.432	0.700*	-0.769*							
SL	0.642	0.677*	0.614	0.614	-0.567	0.708*	-0.432	0.823**						
RL	0.868**	0.886**	0.864**	0.864**	-0.382	0.648	-0.771*	0.928**	0.777*					
RSR	0.203	0.200	0.240	0.240	0.330	0.060	-0.272	0.200	-0.100	0.430				
SFW	0.292	0.348	0.246	0.246	-0.627	0.542	-0.106	0.506	0.869**	0.536	-0.219			
RFW	0.251	0.308	0.209	0.209	-0.654	0.445	-0.218	0.388	0.694*	0.533	-0.104	0.883**		
SDW	0.156	0.214	0.109	0.109	-0.572	0.492	0.065	0.409	0.835**	0.445	-0.199	0.963**	0.798**	
RDW	0.303	0.354	0.264	0.265	-0.672*	0.488	-0.212	0.467	0.782*	0.535	-0.191	0.913**	0.962**	0.837**

4 Where: GR, germination rat; GVI, germination vigor index; GP, germination percentage; MDG, mean daily  
5 germination; MGT, mean germination time; GE, germination energy; RSI, relative salt injury; SVI, seedling vigor  
6 index; SL, shoot length; RL, root length; RSR, root/shoot ratio; SFW, shoot fresh weight; RFW, root fresh weight;  
7 SDW, shoot dry weight, RDW, root dry weight; \*\*, highly significant differences exited at the 0.01 level; \* ,  
8 significant differences exited at the 0.05 level.

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**Table 7** (on next page)

Values of 22 abiotic stress indices based on shoot fresh weight under stress (Ys) and control (Yc) treatments

	Zhongs 1084	Gannong No.2	Gannong No.4	Shida No.1	C6	C16	C23	C25	C36
Yield under normal condition (Yns)	457.67	342.25	433.00	480.33	488.33	324.67	385.00	373.33	422.33
Yield under stress condition (Ys)	70.00	101.87	144.00	88.33	121.17	117.80	93.34	97.97	76.33
Mean productivity (MP)	263.83	222.06	288.50	284.33	304.75	221.23	239.17	235.65	249.33
Geometric mean productivity (GMP)	1497.52	1884.53	2996.45	1935.96	2677.57	2122.58	1831.40	1892.90	1568.71
Harmonic mean (HM)	121.43	157.00	216.12	149.22	194.16	172.88	150.25	155.21	129.30
Stress Tolerance Index (STI)	0.19	0.21	0.37	0.25	0.35	0.23	0.21	0.22	0.19
Yield index (YI)	0.69	1.01	1.42	0.87	1.20	1.16	0.92	0.97	0.75
Modified stress tolerance index-I (MSTI1)	0.23	0.14	0.41	0.34	0.49	0.14	0.19	0.18	0.20
Modified stress tolerance index- II (MSTI2)	0.09	0.21	0.74	0.19	0.50	0.31	0.18	0.20	0.11
Yield stability index (YSI)	0.15	0.30	0.33	0.18	0.25	0.36	0.24	0.26	0.18
Relative stress index (RSI)	0.62	1.21	1.35	0.75	1.01	1.48	0.99	1.07	0.74
Drought index (DI)	0.11	0.30	0.47	0.16	0.30	0.42	0.22	0.25	0.14

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Cont.

	Zhongs 1084	Gannong No.2	Gannong No.4	Shida No.1	C6	C16	C23	C25	C36
stress/non-stress productivity index (SNPI)	145.06	187.21	261.72	175.84	228.31	212.80	176.52	183.04	152.50
relative efficiency index (REI)	0.77	0.84	1.50	1.02	1.42	0.92	0.86	0.88	0.77
mean relative performance (MRP)	1.80	1.84	2.47	2.04	2.38	1.95	1.86	1.87	1.78
golden mean (GM)	1.36	1.85	2.00	1.45	1.66	2.14	1.64	1.71	1.44
tolerance index (TOL)	387.67	240.38	289.00	392.00	367.17	206.87	291.66	275.37	346.00
stress susceptibility Index (SSI)	1.12	0.93	0.88	1.08	1.00	0.84	1.00	0.98	1.09
stress susceptibility percentage index (SSPI)	0.47	0.29	0.35	0.48	0.45	0.25	0.35	0.33	0.42
yield reduction (YR)	0.85	0.70	0.67	0.82	0.75	0.64	0.76	0.74	0.82
abiotic stress tolerance index (ATI)	17048.80	11028.18	17731.07	19839.54	21944.46	9940.17	13584.70	12939.30	15264.15
mean productivity index (MPI)	193.83	120.19	144.50	196.00	183.58	103.43	145.83	137.68	173.00
Schnieder's stress susceptibility index (SSSI)	0.09	-0.05	-0.09	0.06	0.00	-0.12	0.00	-0.02	0.06
sensitivity drought index (SDI)	0.85	0.70	0.67	0.82	0.75	0.64	0.76	0.74	0.82

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**Table 8**(on next page)

Rank of genotypes by 22 abiotic stress indices and shoot fresh weight under stress (Ys) and control (Yc) treatments as well as their average rank (AR).

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	Zhongs 1084	Gannong No.2	Gannong No.4	Shida No.1	C6	C16	C23	C25	C36
Yield under normal condition (Yns)	3	8	4	2	1	9	6	7	5
Yield under stress condition (Ys)	9	4	1	7	2	3	6	5	8
Mean productivity (MP)	4	8	2	3	1	9	6	7	5
Geometric mean productivity (GMP)	9	6	1	4	2	3	7	5	8
Harmonic mean (HM)	9	4	1	7	2	3	6	5	8
Stress Tolerance Index (STI)	9	7	1	3	2	4	6	5	8
Yield index (YI)	9	4	1	7	2	3	6	5	8
Modified stress tolerance index-I (MSTI1)	4	8	2	3	1	9	6	7	5
Modified stress tolerance index- II (MSTI2)	9	4	1	6	2	3	7	5	8
Yield stability index (YSI)	9	3	2	7	5	1	6	4	8
Relative stress index (RSI)	9	3	2	7	5	1	6	4	8
Drought index (DI)	9	3	1	7	4	2	6	5	8

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Cont.

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	Zhongs 1084	Gannong No.2	Gannong No.4	Shida No.1	C6	C16	C23	C25	C36
stress/non-stress productivity index (SNPI)	9	4	1	7	2	3	6	5	8
relative efficiency index (REI)	9	7	1	3	2	4	6	5	8
mean relative performance (MRP)	8	7	1	3	2	4	6	5	9
golden mean (GM)	9	3	2	7	5	1	6	4	8
tolerance index (TOL)	8	2	4	9	7	1	5	3	6
stress susceptibility Index (SSI)	9	3	2	7	5	1	6	4	8
stress susceptibility percentage index (SSPI)	8	2	4	9	7	1	5	3	6
yield reduction (YR)	9	3	2	7	5	1	6	4	8
abiotic stress tolerance index (ATI)	6	2	7	8	9	1	4	3	5
mean productivity index (MPI)	8	2	4	9	7	1	5	3	6
Schnieder's stress susceptibility index (SSSI)	9	3	2	7	5	1	6	4	8
sensitivity drought index (SDI)	9	3	2	7	5	1	6	4	8
AR	8.04	4.29	2.12	6.08	3.75	2.92	5.88	4.63	7.25

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**Table 9** (on next page)

Average of the studied traits for the two clusters under normal and water stress conditions

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2 **Table 9.** Average of the studied traits for the two clusters under normal and water stress

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conditions

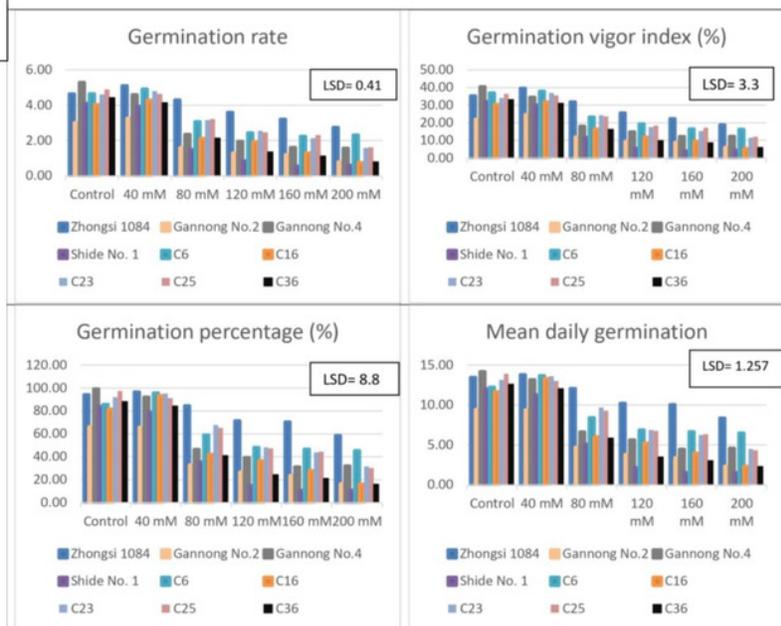
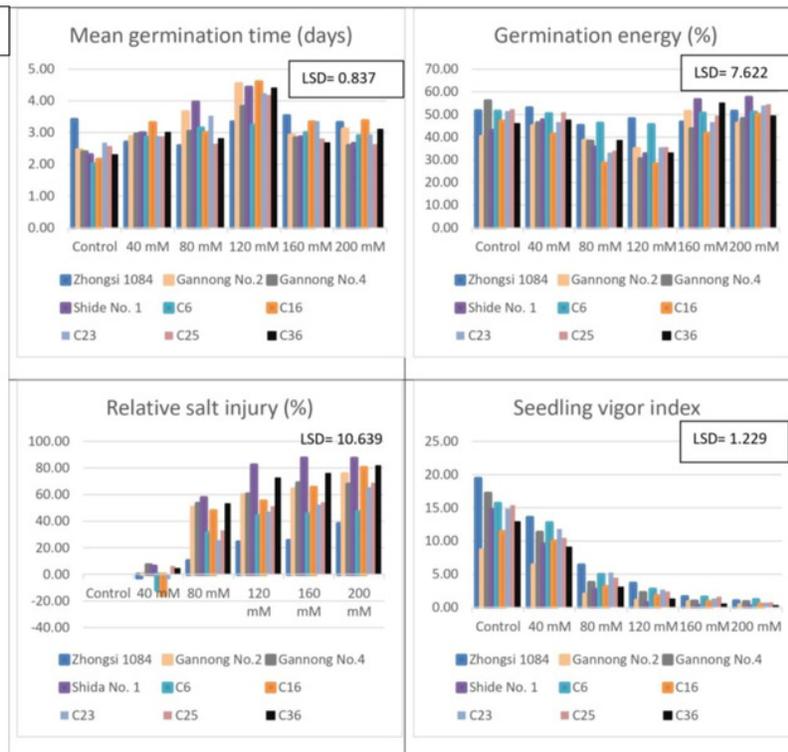
Treatment Group	Control		200 mM	
	1	2	1	2
Germination rate (GR)	4.28	4.65	1.35	1.54
Germination vigor index (GVI)	32.15	35.86	9.79	11.26
Germination percentage (GP)	86.89	88.71	27.17	31.13
Mean daily germination (MDG)	12.41	12.67	3.88	4.45
Mean germination time (MGT)	2.62	2.19	2.96	2.95
Germination energy (GE)	47.42	51.46	52.17	49.70
Relative salt injury (RSI)	0.00	0.00	69.46	65.02
Seedling vigor index (SVI)	14.32	14.73	0.55	0.81
Shoot length (SL)	9.84	9.80	1.56	2.19
Root length (RL)	6.49	6.73	0.48	0.48
Root/shoot ratio (RSR)	0.66	0.69	0.33	0.22
Shoot fresh weight (SFW)	410.15	415.33	87.97	127.66
Shoot fresh weight (RFW)	188.29	248.11	63.06	67.56
Shoot dry weight (SDW)	51.37	50.53	14.31	20.34
Root dry weight (RDW)	27.59	32.63	12.24	14.54

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# Figure 1

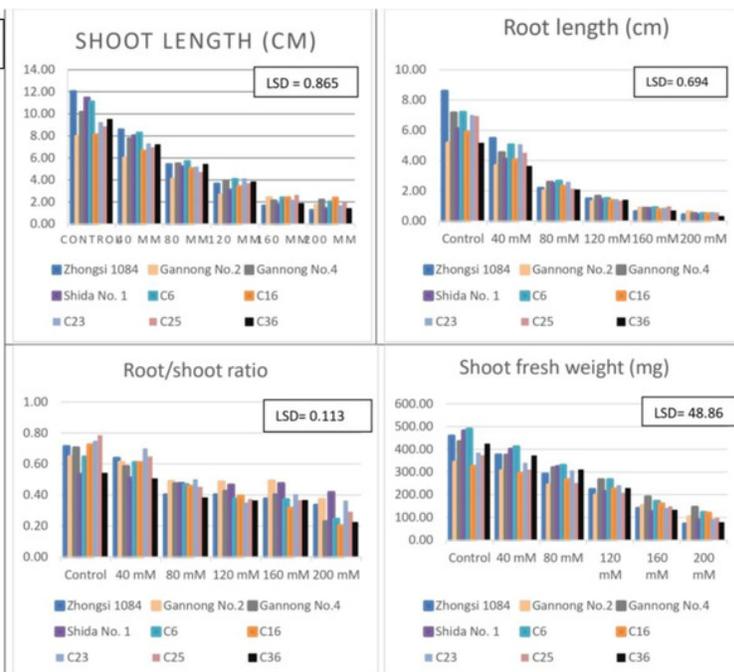
Mean performance of germination traits as affected by the interaction between genotypes and salt treatments (mM NaCl)

**1 A****1 B**

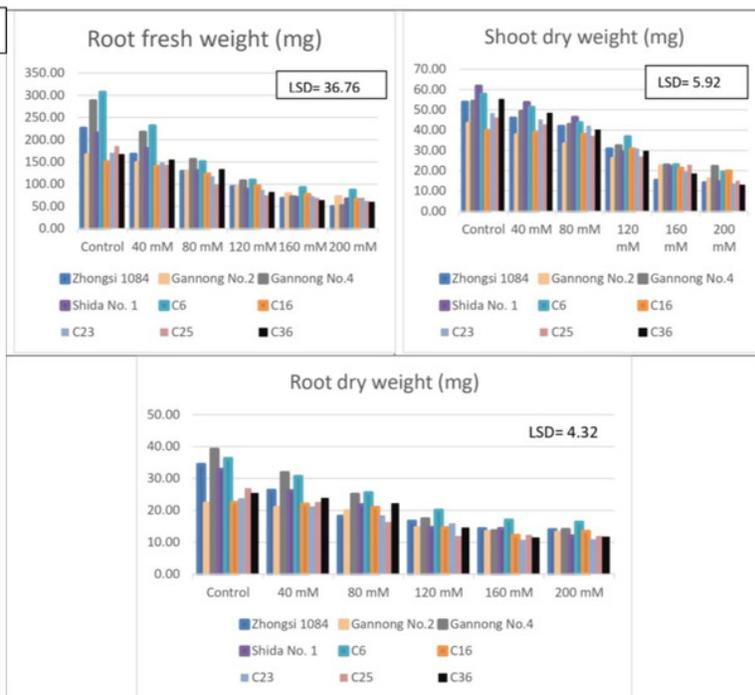
## Figure 2

Mean performance of seedling traits as affected by the interaction between genotypes and salt treatments (mM NaCl)

2 A

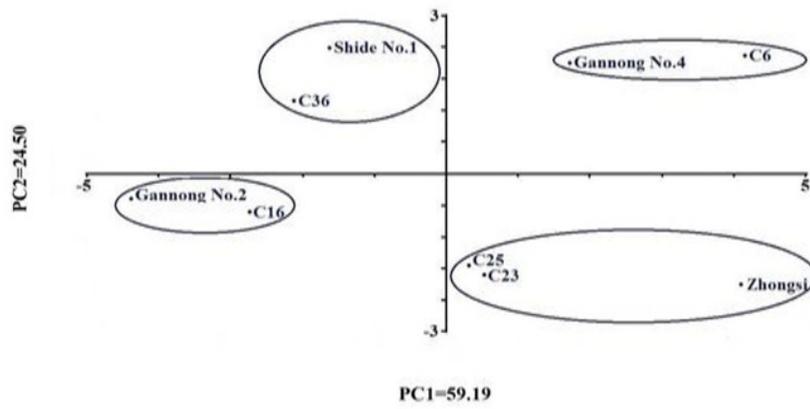


2 B



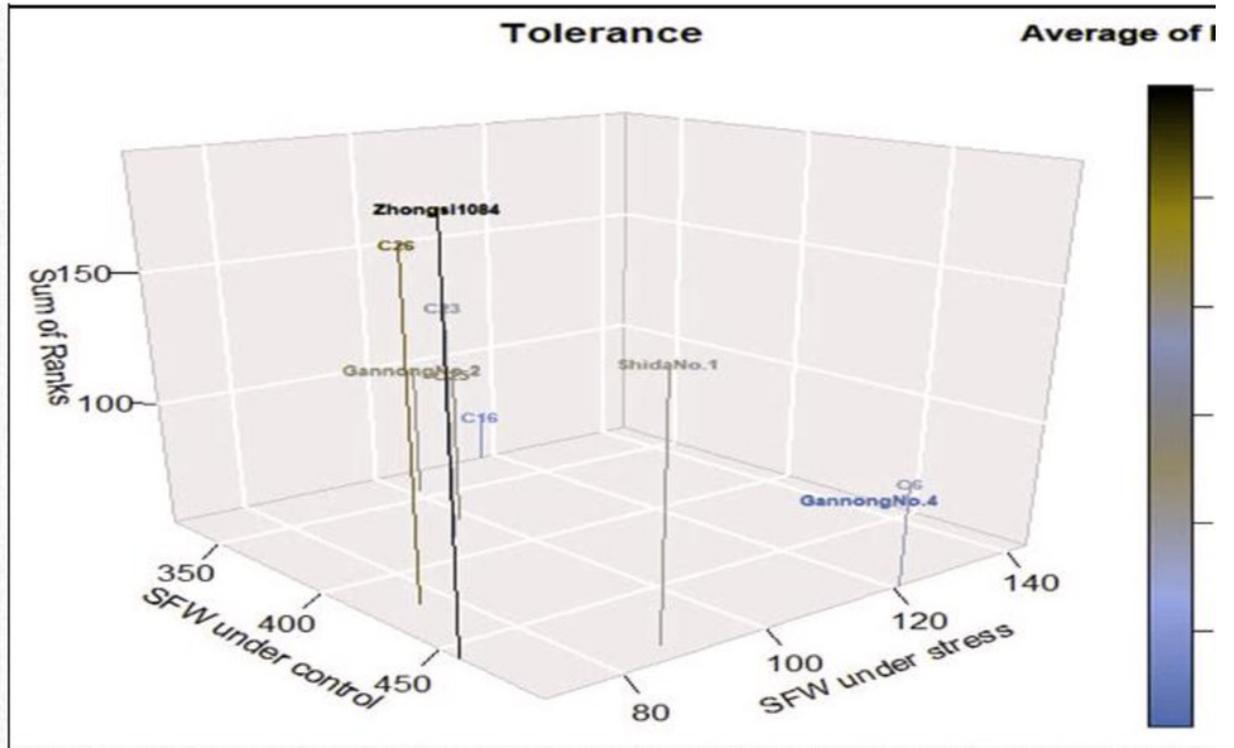
## Figure 3

Two-dimensional ordination of the nine Triticale genotypes investigated in this study based on their overall mean performance under salt treatments



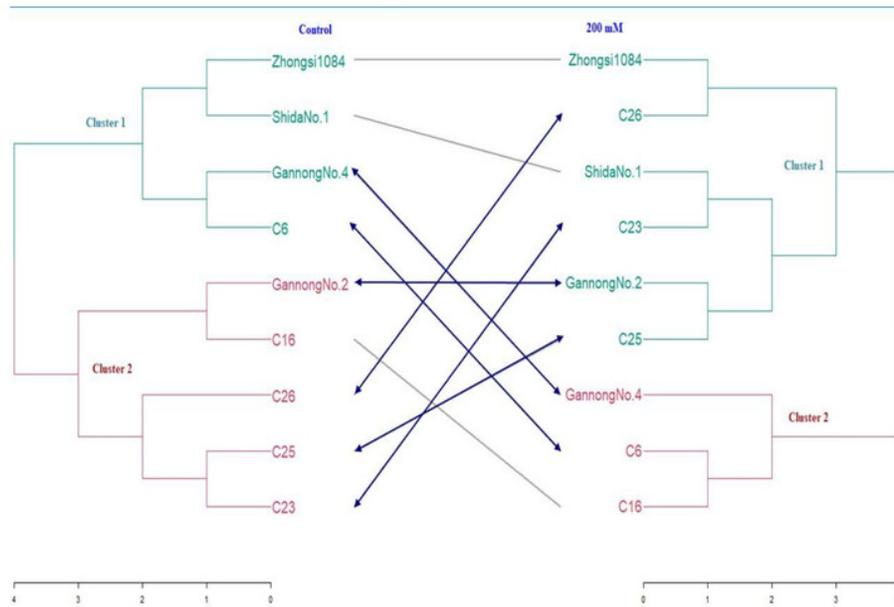
## Figure 4

Tolerance of genotypes according to the average rank of 22 abiotic stress indices  
(Lower average rank indicates higher tolerance)



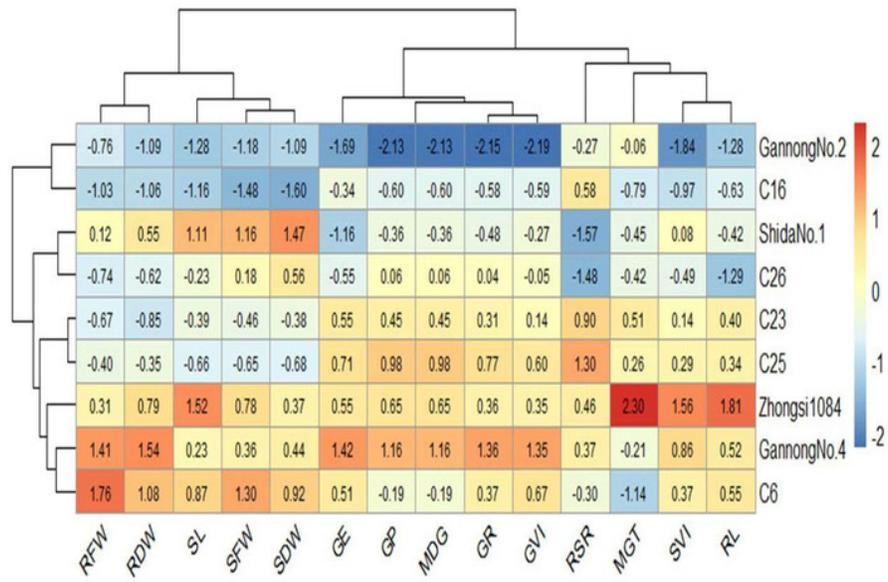
## Figure 5

Tanglegram showing results of cluster analysis based on Euclidian coefficient and Ward method under normal and water stress conditions.



## Figure 6

Heatmap of the relationship between genotypes and the studied traits under control treatment



## Figure 7

Heatmap of the relationship between genotypes and the studied traits under 200 mM NaCl treatment

