

Current knowledge in the Southern Hemisphere marine microbiome of eukaryotic hosts and the Strait of Magellan surface microbiome project.

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Host-microbe interactions are ubiquitous, and they play important roles in host biology, ecology, and evolution. Yet, host-microbe research has focused on inland species, whereas marine hosts and their associated microbes remain largely unexplored, especially in developing countries located in the Southern Hemisphere. Here, we review current knowledge of microbial communities associated with marine hosts in the Southern Hemisphere. There are important biases in marine host species sampled for the studies in the Southern Hemisphere, where sponges and marine mammals have received the greatest attention. Sponges associated microbes varied greatly across regions and species, nevertheless, beside taxonomic heterogeneity, the microbiome has functional consistency, whereas marine mammals' microbiome seems to be driven by geography and aging. Southern Hemisphere seabirds and macroalgae holobiont studies are also common. Seabirds associated microbes rely on amplicon studies, focused on feces. Seabird fecal microbiota is influenced by aging, sex, and species' specific factors, while macroalgae apparently actively filters the microbes that establish in its surface, in a process known as "microbial gardening". In contrast, marine invertebrates and fish microbiomes have received less attention in the Southern Hemisphere. In general, the marine holobiont in the Southern Hemisphere is characterized by few systematic and authentic holobiont studies (i.e., studies that analyze at the same time host traits [e.g., genomics, transcriptomics] and microbiome traits [e.g., 16S, metagenome]), except for some marine invertebrates and macroalgae. We believe that our project on the surface microbiome of key species in the Strait of Magellan will provide valuable information on the points listed above, which will improve the knowledge of microbial diversity in the region, as well as its current responses under the Anthropocene.

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32 **Abstract**

33

34 Host-microbe interactions are ubiquitous, and they play important roles in host biology, ecology,
35 and evolution. Yet, host-microbe research has focused on inland species, whereas marine hosts
36 and their associated microbes remain largely unexplored, especially in developing countries
37 located in the Southern Hemisphere. Here, we review current knowledge of microbial
38 communities associated with marine hosts in the Southern Hemisphere. There are important
39 biases in marine host species sampled for the studies in the Southern Hemisphere, where sponges
40 and marine mammals have received the greatest attention. Sponges associated microbes varied
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42 has functional consistency, whereas marine mammals' microbiome seems to be driven by
43 geography and aging. Southern Hemisphere seabirds and macroalgae holobiont studies are also
44 common. Seabirds associated microbes rely on amplicon studies, focused on feces. Seabird fecal
45 microbiota is influenced by aging, sex, and species' specific factors, while macroalgae apparently
46 actively filters the microbes that establish in its surface, in a process known as "microbial
47 gardening". In contrast, marine invertebrates and fish microbiomes have received less attention in
48 the Southern Hemisphere. In general, the marine holobiont in the Southern Hemisphere is
49 characterized by few systematic and authentic holobiont studies (i.e., studies that analyze at the
50 same time host traits [e.g., genomics, transcriptomics] and microbiome traits [e.g. 16S,
51 metagenome]), except for some marine invertebrates and macroalgae. We believe that our project
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53 information on the points listed above, which will improve the knowledge of microbial diversity
54 in the region, as well as its current responses under the Anthropocene.

55

56 **Introduction**

57

58 The Southern Hemisphere, and particularly the Southern Ocean and its associated ecosystems, is
59 characterized by its unique biodiversity (Rogers et al., 2020; Gutt et al., 2021). These important
60 ecosystems are facing major abiotic challenges as climate change progresses. These challenges
61 are driven primarily by ocean warming and increased UV radiation (Thompson & Salomon,
62 2002; Swart et al., 2018). Sea surface warming creates a stronger stratification in the water
63 column (Pellichero et al., 2017), as well as higher variability in the duration and extent of sea ice
64 sheet and increases in glacier melt rate in the southernmost regions (Gutt et al., 2015; Comiso et
65 al., 2017). Additionally, ocean warming increases microplankton metabolic activity, which in
66 turn accelerates oxygen depletion in the water column (Schmidtke et al., 2017), and decreases

67 ocean pH (McNeil & Matear 2008). Tragically, recent intense wildfires and volcanic eruptions in
68 the Southern Hemisphere have increased the Antarctic ozone hole size in 2020-2021, which is
69 expected to worsen ocean warming effects (Yook et al., 2022). On the other hand, atypical
70 glacier melt rate are stimulating marine primary productivity, which creates complex scenarios in
71 ice dependent species (Piñones & Fedorov 2016). For example, Antarctic krill (*Euphasia*
72 *superba*), a key species in the Antarctic trophic network, requires ice in their early stages, while
73 also forages in areas with high concentration of chlorophyll-a concentration (Kawaguchi et al.,
74 2006). Current evidence suggests that habitat quality heterogeneity along its Antarctic
75 distribution will produce contractions in its distribution (Atkinson et al., 2019; Veytia et al., 2020,
76 but see Cox et al., 2018)

77

78 However, there are also some surprising results. For example, warmer temperatures
79 coupled with low to moderate winds, increase ice melt, which in turn is expected to increase iron
80 release (Hodson et al., 2017). Iron is a primary productivity limiting factor, so its increased
81 availability triggers diatom growth, which in turn increases krill recruitment (Noble et al., 2013;
82 Bertrand et al., 2007, 2015). Ultimately, krill biomass increase provides greater resources for
83 predators, which overall increases energy transfer along the trophic network (Saba et al., 2014).
84 As climate change progresses, it is expected an intensification in seasonality, which might
85 intensify alterations in biological processes (e.g., bottom-up mechanisms).

86

87 For instance, in the Strait of Magellan – the southernmost continental region of South
88 America – climate change derived effects have been recorded since the second half of the XX
89 and early XXI century. These include, increasing sea surface temperature (Smith & Reynolds,
90 2004) and higher glacier melt rate (Aniya, 1999; Dixon and Ambinakudige, 2015). In the
91 terrestrial ecosystem, warmer seasonality is expected to increase aridity in the Patagonian region,
92 particularly in areas with herbaceous vegetation (Soto-Rogel et al., 2020). Nevertheless, the
93 region has not experienced any formal study regarding the effect of climate change in any of its
94 ecosystem properties (e.g. trophic network interactions, biogeochemical cycles, environmental
95 status). This is unfortunate since the region offers an invaluable geographic position. From the
96 marine perspective, the Strait of Magellan is uniquely influenced by the Pacific and Atlantic
97 oceans, as well as the Cape Horn current. Additionally, glacier melt seasonal input creates local
98 primary productivity bursts that have bottom-up effects that recruit species from higher trophic
99 status, which overall increases the biodiversity of the region.

100

101 Despite all the previous natural history studies and museums filled with macro-organisms’
102 diversity, eukaryotes live inside a wider microbial world. Eukaryotes’ homeostasis (e.g.,
103 physiology, immunology, and metabolism) is driven or at least greatly influenced by microbes
104 (McFall-Ngai et al., 2013; Cani et al., 2019; Peixoto et al., 2021). As a result, the “hologenome
105 evolution theory” and “holobiont theory” emerged (Zilber-Rosenberg & Rosenberg, 2008;
106 Bordenstein & Theis, 2015). The holobiont term per se, is not new. It was first introduced by
107 Lynn Margulis to describe the biological unit formed between a host and a single inherited
108 endosymbiont (Margulis L. 1991). The novelty around the holobiont concept arose as a direct
109 consequence of the development and cost decrease of Next Sequencing Technologies, which spur
110 host microbial communities research. This has revealed that microbes are ubiquitous in every
111 single metazoan (Simon et al., 2019). Thus, the current holobiont concept refers to a cohesive
112 evolutionary unit formed by the host and its associated microbes (Bordenstein & Theis, 2015;
113 Rosenberg & Zilber-Rosenberg, 2018). In other words, a holobiont is a single ecological unit,
114 comprised of an intricated network of mutualistic, commensalisms, and parasitic relationships
115 between microbes and its host, that are critical for the survival of all organisms involved.
116 Nevertheless, microbial influence in the holobiont might vary among hosts (Hammer et al.,
117 2019), which has been interpreted as flaws in the holobiont concept (Moran & Sloan, 2015;
118 Douglas & Werren 2016). Holobionts could be assembled by ecological, evolutive or neutral
119 process. Hence, a first objective in holobiont research lies in determining the nature of the
120 holobiont assembly (Theis et al., 2016).

121

122 In marine hosts – which are unlikely to be studied under experimental designs – holobiont
123 research could be focused on the characterization of the host associated microbes in a relevant
124 environmental and evolutionary framework (Leray et al., 2021). Up to date, metabarcoding
125 approaches have been the most popular methods to tackle this issue. Yet, it only serves as an
126 initial step to characterize the microbial composition, as to test the holobiont hypothesis (i.e., host
127 and associated microbes evolve as a unit) further efforts encompassing host associated microbes’
128 functional characterization (e.g., metagenomic or metatranscriptomic data), coupled with host
129 molecular information (e.g., genomic or transcriptomic data), relevant environmental variables
130 measurement, as well as seasonal replicates, must be done to test the congruence of holobiont
131 response against seasonal fluctuations.

132

133 Once we have baseline information about bacterial composition associated to marine
134 hosts, we should be able to recognize hosts whose microbial communities are far from the
135 expected natural variability. Changes in the host's natural composition of microbial communities
136 are collectively termed *dysbiosis* (Zaneveld et al., 2017). Thus, the microbiome itself could be
137 used as a biosensor of host's status (Zolti et al., 2020; Inda & Lu 2020). In marine species, there
138 are several examples of the interplay between host fitness and microbial symbiosis, especially in
139 sponges (Pita et al., 2018 and references within) and algae (van der Loos et al., 2019 and
140 references within). Dysbiosis is the microbial fingerprint underlying the disruption of the host's
141 health and ecology, a highly relevant topic in the current anthropogenic climate change epoch.
142

143 Microbial communities routinely colonize metazoan internal (e.g., gut, oral) or external
144 (e.g., skin) tissues (Ross et al., 2019; Diaz et al., 2021; ANID R20F0009). However, they differ
145 in the selective pressures that influence their assemble. While internal microbial communities
146 assembly is influenced by diet and host physiology, external microbial communities assembly is
147 greatly influenced by environmental perturbations that impair host fitness (Byrd et al., 2018;
148 Kuziel & Rakoff-Nahoum 2022). Therefore, we think that surface microbial communities of
149 eukaryotes could serve as valuable tools to survey environmental status. The epidermis/outer
150 surface of eukaryotes is considered a hostile environment, yet it is frequently colonized by
151 microbes. These microbes must cope with constant shedding (in epidermis) molt (in feathers),
152 intense solar radiation exposure, low temperature, pH changes, as well as antimicrobial molecules
153 (Percival et al., 2012). Nevertheless, skin microbes play important roles in the host's health
154 (Apprill et al., 2014), since they are the first line defense against pathogens and actively
155 participate in the host's immune system maturation (Ross et al., 2019).
156

157 Marine eukaryotes vary greatly in the nature and complexity of their superficial tissue, so
158 the nature of the surface/skin is a critical issue to consider when studying the marine skin
159 microbiome of any host. For example, algae and fish both have a mucus layer in its surface/skin,
160 but their composition and function are different (Gomez et al., 2013; van der Loos et al., 2019).
161 In birds and mammals, the epidermis is covered by feathers and hair, respectively. Importantly
162 seabirds and marine mammals have developed different strategies to cope with cold water,
163 therefore their epidermis and associated elements (i.e., feathers or hair) is completely different
164 from species inhabiting tropical and temperate latitudes (Ross et al., 2019). On the other hand,
165 crustaceans' outer surface is solely its exoskeleton of chitin, so their adaptations to cold

166 temperatures are to be discovered both in their microbiome and their physiology. Even though,
167 the unique biodiversity living in the Southern Hemisphere, few holobiont studies have been
168 conducted in the region. Biodiversity studies are becoming increasingly important, as climate
169 change imposes major threats in ecosystems around the globe, especially in cold environments.
170

171 Up to date, marine holobiont studies have had spatial and phylogenetic biases; on one
172 hand, most of them have been realized in the tropics or in the Northern hemisphere (most of
173 references within Ross et al., 2019), while, on the other hand, the most studied species have been
174 sessile organisms, like sponges (Figure 1). Therefore, Southern Hemisphere marine host
175 microbiome research has received less attention across the metazoan spectrum. Moreover, there
176 are scarce microbiome studies in marine vertebrate and invertebrates. Here, we review the marine
177 host-microbiota/microbiome interactions occurring in the Southern Hemisphere. This review will
178 aid as a diagnosis of the field progress in the region, as well as to detect knowledge gaps and
179 opportunities for further research. Thus, the review is intended for scientists interested in
180 eukaryotes associated microbes, as well as anyone interested in using microbiota/microbiomes as
181 biosensors of eukaryotes' health.

182

183 In this review, our main objectives are i) review the current knowledge regarding marine
184 holobiont and microbiota/microbiome studies in the Southern Hemisphere, and ii) to describe a
185 long-term project that will improve knowledge of microbial communities associated with selected
186 taxa in the Strait of Magellan, Chile (ANID R20F00009). We start with a brief overview of the
187 surface characteristics of several taxa. Later, we review holobiont interactions in a trophic level
188 fashion, first with macroalgae, later with primary and secondary animals' consumers, and finally
189 with predators.

190

191 **Survey methodology**

192

193 We covered all marine microbe-eukaryote interaction reports in the Southern Hemisphere that we
194 were able to find. For this, we performed a comprehensive analysis of literature of the last two
195 decades in the following online databases: PubMed, Science Direct, Scopus and Google Scholar.
196 The search spanned the last two decades and was concluded at 1/February/2023. Only studies in
197 English were selected for further inspection. Several keywords were used to perform the
198 literature search, using the following key words, in combination with the terms, -holobiont-, -

199 microbiota- and -microbiome-: seaweed, sponge, invertebrate, crustacean, marine vertebrate,
200 whales, seals, seabirds. Articles found to match any of these words were examined to guarantee
201 that their sampling was done in hosts inhabiting the Southern Hemisphere.

202

203 **Results**

204

205 Overall, geographical distribution of marine host-microbial research revealed important biases.
206 Geographical biases are reflected by sampling effort across the Southern Hemisphere, since
207 research has been done predominantly in the Antarctic (above 60° S), especially in the Western
208 Antarctic, and temperate latitudes (between 20 – 40° S), especially in Australia, while tropical
209 (between 0 – 20° S) and cold (between 40 – 60° S) latitudes, as well as South America and Africa
210 have received less attention (Fig. 2). There are also host biases, both in terms of number of
211 studies and number of species studied. The most studied groups are marine mammals and
212 sponges (Fig. 1), which have been studied across all latitudinal regions and continents in the
213 Southern Hemisphere (Fig. 3). Nevertheless, when it comes to the number of species studied the
214 pattern has some changes, while sponges remain as the best studied group with 65 studied
215 species, the following groups are Macroalgae (24 studied species) and marine invertebrates (23
216 studied species) (Table 1). In general, the predominant molecular approach to study host-
217 associated microbial communities has been with taxonomic markers (i.e., 16S), nevertheless there
218 have been conducted some microbiome functional approaches (i.e., shotgun metagenomics),
219 especially in Australia and Antarctica hosts, whereas South American marine hosts, have had not
220 been studied with any functional approach to date (Fig. 4, Table 1).

221

222 **Macroalgae as an ecosystem**

223

224 Marine macroalgae are important ecosystem engineers that play critical roles in primary
225 production, biogeochemical cycles, and biodiversity recruitment in marine ecosystems (Tuya et
226 al., 2008. Currently, there are 11,017 species of macroalgae species with cosmopolitan
227 distribution, therefore there is a vast microbial-algae symbiotic world that remains unexplored
228 (Guiry & Guiry 2023). Macroalgae microbiome studies in the Southern Hemisphere have
229 received considerable attention with 24 studied species within 13 research articles (Table 1; Fig.
230 1). Most studies have been conducted with 16S approaches, yet there are microbiome approaches
231 with shotgun metagenomics and DNA holobiont approaches (Table 1). Macroalgae microbiome

232 sampling has been conducted worldwide, but it has been done predominantly in Australia,
233 Antarctica, and South America (Fig. 3).

234

235 Algae are an idoneous niche for aerobic and polymer degrading bacteria, since their
236 photosynthetic activity, and their rich composition in carbon and nutrients (like agar,
237 carrageenan, and cellulose), facilitate the establishment of aerobic and heterotrophic bacteria in
238 their surfaces (de Oliveira et al., 2012). Microbial macroalgal epibiotic communities (biofilms)
239 have been extensively studied in the tropics and the Northern Hemisphere (Malik et al., 2020 and
240 references within). Nevertheless, there have been conducted several macroalgae biofilm studies
241 in the Southern Hemisphere which have revealed Cyanobacteria, Proteobacteria, Firmicutes,
242 Bacteroidetes and Actinobacteria as prevalent bacterial phyla (de Oliveira et al., 2012; Albakosh
243 et al., 2016; Gaitan-Espitia & Schmid, 2020). Fungi are also important players in the macroalgae
244 biofilm composition in the Southern Hemisphere, and different species have been documented
245 from Antarctic seaweeds, being the most prevalent the filamentous fungus *Pseudogymnoascus*
246 *pannorum*, and the yeast *Metschnikowia australis* (Loque et al., 2010; Godinho et al., 2013;
247 Furbino et al., 2017; Ogaki et al., 2019).

248

249 There has been extensive research in the macroalgae holobiont that gives a rich body of
250 evidence to frame the study of macroalgae biofilms. These studies have shown that bacteria and
251 fungi inhabiting the macroalgae biofilm actively interact with its host to influence growth,
252 development, and immune function (van der Loos et al., 2019). Valuable microbes to the algal
253 host appear to be taxonomically restricted among bacteria at higher taxonomic levels, therefore,
254 biofilm composition seems to be redundant at phyla or class. Nevertheless, at lower taxonomic
255 levels (genus/ASV [Amplicon Sequence Variant]/OTU [Operational Taxonomic Unit]), there is a
256 rich spectrum of valuable microbes (Egan et al., 2013; Hollants et al., 2013). Despite taxonomic
257 variability in bacterial genera inhabiting its biofilms, traits among bacteria are shared, which
258 creates biofilms with different taxonomic composition but with apparently similar functions
259 (Egan et al., 2013).

260

261 Macroalgae seem to actively recruit its biofilm composition, a process known as
262 “microbial gardening” (Saha & Weinberger 2019). Microbial gardening allows the recruitment of
263 specific beneficial bacteria to the algae. In turn, the recruited microbes release antibiotics,
264 quorum sensing inhibitors and digestive vesicles (Wiese et al., 2009; Romero et al., 2010;

265 Richards et al., 2017) that collectively shape the biofilm composition. Although, algal biofilm
266 composition varies taxonomically, in space, time and host (Lachnit et al., 2011), it shares
267 important traits related with algae morphogenesis (Wiese et al., 2009) and nutrient
268 supplementation (Hollants et al., 2013). Overall, a “correct” microbial gardening, might confer
269 an adequate biofilm composition, able to produce a cocktail of metabolites that provide
270 protection to the algae from microbial pathogens, like bacteria or diatoms (Saha & Weinberger
271 2019), or predators like barnacle larvae or mussels (Saha et al., 2018). Interestingly, the benefit
272 conferred to algae by the surface microbiome is not taxonomically restricted, suggesting that
273 microbial traits underlying algae defense are shared by several bacterial taxa.

274

275 Besides, some bacteria of the algal biofilm produce antibiotic compounds, which act as a
276 microbial filter for the establishment of environmental microbes (Albakosh et al., 2016). For
277 example, algae’s thallus produces dimethylsulfoniopropionate (Kessler et al., 2018), which
278 recruits *Roseovarius* bacteria. This bacterium releases specific morphogenetic compounds that
279 enable correct algae morphogenesis. Moreover, bacterial metabolites could enhance algal
280 performance during stress. Thallusin, a microbial derived metabolite, positively influences algal
281 growth, cell differentiation, cell wall development, and rhizoid formation during abiotic stress
282 (Alsufyani et al., 2020). Therefore, thallusin production might be an excellent example of a
283 holobiont environmental stress response.

284

285 Globally, the algal holobiont will be subject to complex scenarios under climate change.
286 Sea surface temperature, and CO₂ concentration increase, might have different outcomes in algal
287 species and also affect key microbes inhabiting macroalgae biofilms, which in turn, could impair
288 the host’s health (Gaitan-Espitia & Schmid, 2020; Marzinelli et al., 2015). Moreover, it is
289 imperative to consider the effect on the interactions among microbes and microbe-algae. For
290 example, algal hosts sensible to acidification might experience lower photosynthetic rate, which
291 might hamper aerobic bacteria proliferation (Van der Loos et al., 2019). Climate change
292 environmental driven perturbations might produce common disbiotic biofilm composition in
293 stressed macroalgae (Marzinelli et al., 2015). Nevertheless, whether shifts in macroalgae biofilm
294 composition associated to environmental perturbation impairs macroalgae fitness remains
295 unexplored.

296

297 **The macroalgae holobiont in the Southern Hemisphere**

298

299 To the best of our knowledge the macroalgae microbiome has been addressed in 13 studies in the
300 Southern Hemisphere (Fig. 1). Most of these studies have been done in temperate latitudes,
301 particularly in Australia (Fig. 3). There are important gaps in tropical and cold latitudes, as well
302 as in selected regions, like South America, Africa and Indonesia (Fig. 3). Yet, we still ignore
303 much about South America native macroalgae microbiome, since there is only one study that
304 addressed this topic in a native species, *Macrocystis pyrifera* (Laminariales: Laminariaceae),
305 while another explored the biofilm composition of a macroalgae invasive species, *Undaria*
306 *pinnatifida* (Laminariales: Alariaceae) (Florez et al., 2019; Lozada et al., 2022). From these
307 examples, we know that macroalgae biofilm composition is different from seawater and is
308 influenced by seasonality and available nutrients. Interesting, invasive macroalgae in the
309 Southern Hemisphere, shows signs of microbial gardening, since its biofilm composition is
310 dominated by one bacterial taxon, *Leucothrix*, a gamma-proteobacteria (Florez et al., 2019;
311 Lozada et al., 2022). This could give insights about potential adaptation mechanisms of these
312 invasive species in the Southern Hemisphere, which might be relevant in the ecosystemic process
313 of the Patagonian region.

314

315 Australia is the region where the macroalgae holobiont is best understood, since it has the
316 highest number of studies (5 studies, Table 1), and diversity of molecular approximations (i.e.,
317 16S, shotgun metagenomics, DNA holobiont, Table 1). Yet, most of its research effort has
318 focused only on one species, *Ecklonia radiata* (Laminariales, Lessoniaceae), so it is unlikely that
319 their macroalgae holobiont knowledge is generalizable to all macroalgae hosts in the region
320 (Marzinelli et al., 2015, 2018; Qiu et al., 2019; Song et al., 2021; Wood et al., 2022).
321 Nevertheless, it provides valuable information that gives insight into the future that might face
322 the macroalgae holobiont. *Ecklonia radiata* biofilm composition has a dysbiotic composition
323 during environmental stress (Marzinelli et al., 2015, 2018). Yet, stress microbial signatures are
324 not consistent across individuals, which suggest that there are unexplored meaningful covariates
325 (e.g., host genetic variability) that determine the final holobiont phenotype (i.e., biofilm
326 composition) when the algae face environmental stress (Qiu et al., 2019). Besides, horizontal
327 transfer of genes related with the algae niche specific environment, and stress environmental
328 responses between biofilm bacterial members, suggest that this process might facilitate biofilm
329 adaptation to environmental stress (Song et al., 2021). Interestingly, a DNA holobiont approach
330 in the macroalgae *Phyllospora comosa* (Fucales: Seirococcaceae) revealed that host genetic

331 variability has a weak relationship with microbial composition. Moreover, its biofilm
332 composition is driven by local conditions and geography (Wood et al., 2022). Together these
333 results suggest a complex interplay in the macroalgae holobiont, where genetic variability,
334 biofilm composition, horizontal gene transfer and environmental conditions, are crucial players,
335 that create a diverse array of phenotypes. Given the unavoidable environmental changes that we
336 will face in the next decades, and the key role macroalgae play as ecosystem engineers, it is
337 imperative to detect the factors that could promote resilience in the macroalgae holobiont against
338 environmental stress, in order to keep the benefits they provide to marine biodiversity.

339

340 Although bacterial partners have received most of the attention in algae biofilm research,
341 algae biofilms harbor diverse fungal communities. Fungal biofilm composition has been explored
342 in Antarctica. Fungal epibiotic communities in Antarctic macroalgae are influenced by abiotic
343 (i.e., dissolved oxygen and organic matter) and biotic (i.e., antifungal molecules produced by the
344 host) factors (Ogaki et al., 2019). Moreover, macroalgae actively controls these communities,
345 since some fungal strains associated to algae biofilms have agarolytic and carrageenolytic
346 activity, hence, they can degrade algal biomass (Furbino et al., 2017). However, it remains
347 unknown the interactions that might take place between bacteria and fungi inhabiting the
348 macroalgae biofilm. Future studies should have more holistic approaches, where bacteria, fungi
349 and environmental covariables are simultaneously considered.

350

351 In particular, the species *Macrocystis pyrifera* is a worldwide (i.e., present both in the
352 Southern and Northern Hemisphere) distributed algae species, whose epibiotic microbial
353 communities have been addressed across several regions in the world (Florez et al., 2019; Lin et
354 al 2018; Weigel et al., 2019). These studies will be valuable to do *M. pyrifera* comparisons across
355 biogeographic regions. However, little is known from the surface biofilm of *Macrocystis pyrifera*
356 in the Strait of Magellan. In the project of the Strait of Magellan microbiome, that we will
357 describe at the end of the paper, we are attempting to characterize the bacterial and functional
358 traits/responses of *M. pyrifera*, in association with environmental factors at different depths.
359 These data could improve our understanding in macroalgae microbiome response to climate
360 change.

361

362 **Invertebrates**

363

364 **Sponges**

365

366 Currently there are 9542 sponge species around the globe, with at least 8,864 species distributed
367 in the Southern Hemisphere (de Voogd et al., 2023; Downey et al., 2012). Sponges are the best
368 studied holobionts in the Southern Hemisphere, both in terms of studied species (65) and number
369 of studies (16), most likely because sponges associated microbes produce a wide array of
370 metabolites that have biotechnological importance (Taylor et al., 2007, Fig. 1, Table 1). The
371 predominant approach to study the sponge holobiont is 16S, nevertheless there have been
372 conducted shotgun metagenomic approaches, which had shed light in sponges' microbiome
373 metabolic potential (Table 1, Yang et al., 2022; Moreno-Pino et al., 2020, 2021). Sampling effort
374 has been performed worldwide, mainly in Antarctic latitudes (tropical, temperate and cold
375 latitudes) (Fig. 3).

376

377 Sponges are important ecosystem players that participate in several biogeochemical
378 cycles, and provide stability to the benthos (Bell, 2008). Sponges are the first evolved metazoan,
379 and therefore are the sister group to all animals (Wörheide et al., 2012). Their anatomy is unlike
380 any other metazoan, but generally, it consists in several cell layers (Taylor et al., 2007). Most of
381 the studies used in this revision use the outermost cell layer, nevertheless whenever a study
382 sampled more cell layers, it is explicitly stated in the text. In a recent study in the tropical sponge
383 holobiont conducted in the Northern Hemisphere, it was observed a high metabolic redundancy
384 within the microbiomes that could help buffer the sponge from chemical and physical changes in
385 their environment and from fluctuations in the population sizes of the individual microbial strains
386 that make up the microbiome (Kelly et al., 2022). This is not surprising, since their early
387 evolution occurred in a microbe dominated world in the late Precambrian (Renard et al., 2013).
388 Sponge physiology seems to be microbe dependent and had reached different symbiotic-based
389 solutions to environmental challenges (Thomas et al., 2016).

390

391 Current evidence suggests that the sponge microbiome is similar at the phylum level
392 between species in the Southern Hemisphere and those in the Northern Hemisphere (Taylor et al.,
393 2007). In the Southern Hemisphere the sponge microbiome is characterized by several phyla that
394 construct a core including Proteobacteria, Bacteroidetes, Actinobacteria, Verrucomicrobia,
395 Acidobacteria and Cyanobacteria (Rodríguez-Marconi et al., 2015; Matcher et al., 2017;
396 Cárdenas et al., 2018; Savoca et al., 2019; Papale et al., 2020; Happel et al., 2022; Ruocco et al.,

397 2021; Yang et al., 2022). Interestingly, there is only one archaea phylum associated with sponges,
398 Thaumarchaeota, yet its association is consistent across several species (Brochier-Armanet et al.,
399 2008; Sacristán-Soriano et al., 2020; Moreno-Pino et al., 2020; Steinert et al., 2020).

400

401 Interestingly, some bacterial lineages associated to sponges are phylogenetic novelties,
402 that is, their DNA sequences are new, hence, they are unable to be identified with current
403 sequences databases knowledge. Antarctic sponges (Papale et al., 2020; Moreno-Pino 2021;
404 Happel et al., 2022), Australian sponges (Yang et al., 2022), and South African sponges (Matcher
405 et al., 2017) are the ones with more “unknown” bacterial partners. Noteworthy, phylogenetic
406 novel bacteria vary in their prevalence across sponge taxa. Sponges’ genus *Sporosarcina* and
407 *Nesterenkonia* have greater phylogenetic novelty in their microbiotas (50 % or more) while
408 others, like *Cellulophaga algicola* tend to harbor fewer unknown microbes, therefore most of its
409 associated microbes is recognizable (Moreno-Pino et al., 2021). Considering metabolic evidence
410 from Northern Hemisphere tropical sponges, functional redundancy of their microbiomes might
411 allow sponges to tolerate environmental fluctuations, which allows them to distribute over wide
412 areas (Kelly et al., 2022). Hence, environmental variability could be an important driver of
413 microbiota assembly in sponges of the Southern Hemisphere, both at inter- (between different
414 species) and intraspecific (within the same species, among individuals) levels (Steinert et al.,
415 2020).

416

417 Nevertheless, besides environmental heterogeneity, phylogenetic factors (i.e.,
418 species/genera particular affinities) also play an important role in sponges’ microbiota assembly.
419 For example, sponges from the genus *Mycale* display a strong bacterial core among individuals
420 and species distributed over hundreds of kilometers in the Southern Ocean (Cárdenas et al., 2018;
421 Happel et al., 2022). Similar stable associations have been found in some sponge species from
422 Antarctica (Steinert et al., 2019). *Mycale magellanica*, a common sponge living in the Strait of
423 Magellan, shares up to 74% of sequences belonging to Rhodobacteriaceae and Flavobacteriaceae
424 with individuals of *M. acerata*, a common sponge in the Western Antarctic Peninsula (Cárdenas
425 et al., 2018). Moreover, in *M. acerata*, bacterial composition is stable among individuals
426 distributed across the entire West Antarctic Peninsula (Happel et al., 2022).

427

428 Similar trends, with similar microbiota composition, are shown in Demospongiae and
429 Hexactinellida sponges from Ross Sea, Antarctica. These sponges share the bacterial genera

430 *Erwinia*, *Methylobacterium* and *Sphingomonas* (Papale et al., 2020). In contrast, in the same
431 region (Ross Sea), sponges' microbiotas composition has been found to be heavily influenced by
432 environmental microbes (horizontal transmission), both in the cultivable fraction and NGS-
433 microbiota (Savoca et al., 2019; Sacristán-Soriano et al., 2020). Overall, this highlights the
434 differences in bacterial composition variability among sympatric sponges, which might create
435 species or genus specific microbiota compositions.

436

437 Factors underlying microbial recruitment in sponges get more complex when we consider
438 sponges associated Archaea. For example, in Demospongiae and Hexactinellida sponges, from
439 the South Pacific Ocean, bacterial composition is species specific, while the Archaea composition
440 is individual specific (Steinert et al., 2020). Thus, Archaea composition is sparse among
441 individuals, which might suggest that Archaea are opportunistic/contingent players in the sponge
442 holobiont, or rather, their functional benefits are widely shared among several archaea taxa. This
443 pattern contrasts with the stable microbiota composition reported in Ross Sea sponges (Papale et
444 al., 2020). The differentiated trends suggest that environmental fluctuation coupled with species
445 specific filters might drive microbial composition associated with sponges.

446

447 The above examples illustrate the complex factors underlying sponge symbiotic
448 associations with bacteria, where horizontal transmission and host specific factors appear to have
449 a differential role among sponge species. Despite inconsistency among associated microbes'
450 identity, it is likely that sponges associated microbes have similar functional traits. Thus,
451 although sponges' microbiota has complex patterns, their microbiomes might have functional
452 convergence (Cristi et al., 2022).

453

454 Nevertheless, whether sponges' microbiomes taxonomy obey contingent issues (i.e.,
455 which microbe taxa arrived first or neutral process in microbiota assembly) or indeed have a
456 biological basis is an open question. Symbiotic interactions among sponge and its associated
457 bacteria dynamically shape the sponge microbiota composition. On one hand, opportunistic
458 bacteria that degrade sponge tissues, like *Bacillus*, *Micrococcus* and *Vibrio*, are common
459 members of the sponge holobiont. On the other hand, there are antibiotic producing bacteria that
460 regulate the former, like *Streptomyces*, *Aquimarina*, *Pseudovibrio* and *Pseudoalteromonas*
461 (Esteves et al., 2017). Variations in quorum sensing, a microbe chemical communication system,
462 also might play an important role in microbial recruitment. The genera *Pseudomonas*,

463 *Shewanella* and *Roseobacter*, common bacteria associated to sponges, produce Acyl homoserone
464 lacton, an important chemical messenger in quorum sensing (Mangano et al., 2018). The quorum
465 sensing activity by these bacteria might alter the community profile, by differential microbial
466 recruitment. Quorum sensing might be an important adaptive trait in the sponge holobiont,
467 specially under the ongoing climate derived marine changes, yet its prevalence, and more
468 important its relevance, among sponges' microbiomes has not been thoroughly studied.
469

470 Metagenomic studies have shed light on the characteristics of the functional repertoire of
471 sponges' microbiomes. Microbe metabolism differs throughout sponge tissue, highlighting a
472 tissue specific microbiome metabolism (Yang et al., 2022). Several examples pinpoint to nutrient
473 provisioning as an important trait in sponges' microbiomes. Microbial symbionts encode multiple
474 genes related to nitrogen fixation and metabolism of nitrogen compounds, sugars derived from
475 photosynthesis (Moreno-Pino et al., 2020), as well as vitamin B5 (Moreno-Pino et al., 2021). In
476 consonance with antagonistic interactions among members of the sponge microbiome, antibiotic
477 resistance, and biopolymer degradation (Moreno-Pino et al., 2021), as well as CRISPR genes,
478 transposases, detoxification genes and restriction site modifications (Moreno-Pino et al., 2020)
479 are common traits in sponges' microbiomes. The latter functions highlight the evolution of the
480 microbiome within the sponge itself, since several microbes associated with sponges degrade the
481 sponge's tissues and avoid the effect of antimicrobial compounds.
482

483 Furthermore, the high prevalence of CRISPR genes in sponges' microbiomes, suggest that
484 their bacterial members are under constant phage attack (Moreno-Pino et al., 2020), which adds
485 another complexity layer to the microbial interactions in the sponge microbiome. There is a high
486 proportion of genes in the microbial communities with unknown functions, so besides
487 phylogenetical novelty described above, there also stands out functional/metabolic microbial
488 novelty associated with sponges (Moreno-Pino et al., 2020).
489

490 Up to date, few studies have addressed the effects that marine climate change will have on
491 the sponge holobiont. An interesting exception is given by Kandler et al., (2018), which found
492 that microbial communities of the tropical sponges from New Guinea, *Coelocarteria*
493 *singaporensis* and *Stylissa* cf. *flabelliformi* might be tolerant to future marine pH conditions.
494 However, it is still an unifactorial experimental approach, that does not represent a reliable test of
495 the multifactorial climate change process. In contrast, there are other climate change driven

496 effects that might pose a major challenge to sponges in polar environments, like ice scour (seabed
497 modification caused by floating icebergs), which is predicted to increase as a direct consequence
498 of sea surface temperature rise. Ice scour damages benthic communities, specially sponges, like
499 *Isodictya kerguelenensis*. Ice scour injuries produce microbial fingerprints, that are easily
500 identified (Rondon et al., 2020). Thus, as climate change progresses and ice scour increases,
501 Antarctica sponges' integrity might be compromised in the next decades.

502

503 **Non-sponge invertebrates**

504

505 Marine invertebrates comprise between 35 - 39 recognized phyla (Valentin, JW 2006; Zhang, ZQ
506 2013). Most iconic marine invertebrates' phyla are Mollusca (118,061 species), Echinodermata
507 (20,550 species), Annelida (17, 426 species), Cnidaria (17,426 species), and Bryozoa (11,474
508 species) (Zhang, ZQ 2013). Marine invertebrates' microbiomes have received few attention, with
509 23 species within 10 research articles (Fig. 1, Table 1). However, this group has outstanding
510 examples of authentic holobiont approaches (i.e., coupled measurement of host and microbe
511 traits) (Table 1). Sampling effort has been conducted mainly in Australia, Antarctica, and Africa
512 while there are no studies addressing any marine invertebrate microbiome in South America (Fig.
513 3).

514

515 Overall, marine invertebrate microbiome is characterized by several bacterial phyla,
516 where Proteobacteria, Bacteroidetes, Verrucomicrobia, Tenericutes and Actinobacteria are the
517 most abundant (Webster & Bourne, 2007; Murray et al., 2016, 2020; Unzueta-Martínez et al.,
518 2022) Antarctic corals and snails have stable microbiotas among individuals. The Antarctic soft
519 coral, *Alcyonium antarcticum*, has a core microbiota composed by Proteobacteria, Bacteroidetes,
520 Firmicutes, Actinomycetales, Planctomycetes, Chlorobi and sulfate reductor bacteria (Webster
521 and Bourne, 2007). A similar trend is observed in the Antarctic snail *Synoicum adareanum*. This
522 invertebrate's microbiota is characterized by a high prevalence among individuals of
523 Proteobacteria, Verrucomicrobia, Actinobacteria, Nitrospirae and Bacteroidetes (Murray et al.,
524 2020). In other cases, microbes associated to invertebrates display low diversity, like the ice-
525 adhered anemone *Edwardsiella andrillae*, an endemic anemone of Ross Sea living in the sea ice-
526 water interphase, where the main phyla are Proteobacteria and Tenericutes. Interestingly, most of
527 its sequences display recent diversification branching, which suggests that its associated bacteria
528 are evolutionary recent (phylogenetically new) (Murray et al., 2016).

529

530 On the other hand, invertebrates with long and complex life cycles, also have a complex
531 pattern in their associated microbes along its life stages. In the Sydney rock oyster (*Saccostrea*
532 *glomerata*), bacterial composition is driven by life history characteristics (Unzueta-Martínez et
533 al., 2022). For example, environmental bacteria are a major source of bacterial composition in
534 swimming larvae stages, thus, the microbiota of these stages is characterized by common marine
535 free-living bacteria. On the contrary, sessile stages, like pre adult stage, adult and gametes have
536 distinct microbiota profiles. Overall, the bacterial composition across life stages in the Sydney
537 rock oyster is patchy, which suggests that the majority of oyster associated microbes are either
538 opportunistic or commensals, with few relevance to the oyster. Nevertheless, the genus *Nautella*
539 (*Rhodobacterales*) was consistently present across stages, increasing its abundance notably in the
540 last stages.

541

542 To the best of our knowledge, the only crustaceans in the Southern Hemisphere studied to
543 date are lobsters, krill and copepods. Overall, the bacterial microbiota of crustaceans in the
544 Southern Hemisphere is dominated by Campilobacterota, Tenericutes, Actinobacteria,
545 Firmicutes, Bacteroidetes and Proteobacteria (Clarke et al., 2019; Ooi et al., 2019; Clarke et al.,
546 2021; Oh et al., 2021; Zhang et al., 2021).

547

548 Microcrustaceans are important trophic links between primary producers (e.g., diatoms),
549 primary, and secondary predators (e.g., seabirds and fish, respectively). Antarctic krill (*Euphasia*
550 *superba*) is the trophic basis in the Southern Ocean ecosystem. Despite its inherent exposure to
551 marine bacteria, Antarctic krill hosts unique bacteria phyla in its body; furthermore, its epibiotic
552 associated bacteria differentiates as geographic distance increases. Hence, distance, rather than
553 environmental heterogeneity drives epibiotic bacteria composition in krill (Clarke et al., 2019;
554 Clarke et al., 2021). The major bacterial players in their chitin surface are Campilobacterota and
555 Tenericutes, while in the stomach and intestinal gland were Actinobacteria and Firmicutes
556 (Clarke et al., 2019). Interestingly, *Colwellia* bacteria is a prevalent member of epibiotic
557 microbiota in Antarctic krill swarms at local and regional scales (Clarke et al., 2021). Its
558 persistent association, through thousands of kilometers, might suggest an important role to krill
559 health.

560

561 Sea temperature increase might disrupt psychrophilic bacteria associated with Antarctic
562 crustaceans. In the Antarctic copepod *Tigriopus kingsejongensis*, 15 °C temperature treatment
563 has profound effects in its fecal microbiome. Temperature increase diminishes abundance of
564 psychrophilic bacteria (e.g., *Colwellia*), whereas it facilitates the increase in opportunistic
565 pathogens (e.g., *Vibrio*) and virulence genes (Oh et al., 2021). As sea surface temperature
566 increases, it is probable that the microcrustacean holobiont will face major shifts in its associated
567 bacteria, on one hand the increase of bacteria that prefer higher temperatures (e.g., *Vibrio*), and
568 on the other hand, the decrease of psychrophilic bacteria.

569

570 Natural life history events, like molting and temperature, have important effects in
571 crustacean's microbiota composition (Ooi et al., 2019; Zhang et al., 2021). Molting is a critical
572 process in crustaceans that allows them growing and sexual maturation. Evidence from the
573 chinese mud crab, *Scylla paramamosain*, suggests that molting represents a bottleneck to most of
574 its associated microbes in gills and midgut. Nevertheless, hemolymph bacteria,
575 *Halomonas* and *Shewanella*, prevail despite molting (Zhang et al., 2021). Noteworthy, the
576 abundance of this bacteria, had a high correlation with the expression of crab antimicrobial gene
577 expression. These results suggest the presence of highly adapted bacteria to the complex life
578 cycle of the mud crabs. Yet, it is uncertain to which degree, molting could alter microbiome traits
579 in the crab holobiont. Sea temperature increase might impose microbe related burdens to
580 crustaceans, as exemplified by the spiny lobster, *Panulirus ornatus*, in Australia (Ooi et al.,
581 2019). In the spiny lobster, temperature increase has a direct relationship with juvenile mortality.
582 As temperature increases, so does bacteria metabolism, which burdens the lobster immune
583 system by bacteria infiltration and subsequent uncontrolled proliferation in the hemolymph.

584

585 Interestingly, there have been conducted holobiont approaches in snails in deep
586 hydrothermal vents and several Lucanidae species (Mollusca) across the world, where host DNA
587 and microbial DNA have been simultaneously addressed (Lan et al., 2021; Osvatic et al., 2023).
588 These results have highlighted the effect of ecological niche and host-microbe metabolic
589 complementarity in microbiome assembly. Interestingly, sulfur oxidizing bacteria is present in
590 phylogenetical and geographical distant species of Lucanidae Mollusca (Lan et al., 2021; Osvatic
591 et al., 2023). The above examples highlight the relevance of coupling holobiont approaches with
592 relevant ecological data to address the meaningfulness of the interactions among hosts and its
593 associated microbes.

594

595 In the Strait of Magellan study that we will describe at the end of the article, we are
596 including two crustacean species, the centolla, *Lithodes santolla* and the channel sprawns,
597 *Munida gregaria*. The former, is an economically important species that spend most of its life in
598 the sea floor, while the latter is a key species in trophic energy transfer, since it is simultaneously
599 an important plankton consumer and is eaten by several predators (e.g., Magellanic penguin and
600 sea lion). In our holobiont study in the Magallanes region, we expect that the centolla will present
601 fewer signals related to the UV light and heat stress than any of the other species that have a more
602 ample niche in the water column or in the surface as is the case of sea lions and penguins.

603

604 **Vertebrates**

605

606 **Fish**

607

608 There are more than 20,000 species of marine fish around the globe (CML, visited 2023).
609 Fish are important links between trophic basal levels and higher ones, yet the fish holobiont is the
610 least studied in the Southern Hemisphere with 9 studied species within 4 research articles (Fig.
611 1). All fish studies have been conducted with 16S metabarcoding approaches (Table 1). Fish
612 microbiome sampling effort in the Southern Hemisphere has been predominantly done template
613 (Australia) and Antarctic latitudes (Antarctica) (Fig 2). Hence, there are important gaps in
614 tropical, temperate and cold latitudes, specifically in South America, Africa, and Indonesia (Fig.
615 3). Importantly, we still ignore much about native fish species microbiome, since most studies
616 have been done in commercially important species. Most fish microbiome studies have been
617 performed with the objective of testing the usefulness of microbial taxa as biosensors of the fish
618 health. Fish skin has a mucous layer over its epidermis that serves as an additional barrier
619 between the environment and the host's skin. The mucous layer consists of immunogenic
620 compounds that play important roles in innate and adaptive immunity (Gomez et al., 2013). Thus,
621 bacteria inhabiting fish skin, might be commensals in healthy individuals or
622 opportunistic/pathogenic in fish with compromised health.

623

624 Current evidence of fish bacterial microbiota in the Southern Hemisphere, has shown that
625 the most prevalent bacteria phyla associated to fish are Actinobacteria, Firmicutes,
626 Proteobacteria, Tenericutes, and Bacteroidetes (Song et al., 2016; Minich et al., 2020; Legrand et

627 al., 2018; Heindler et al., 2018). In the southern Bluefin Tuna, in Portland Australia, *Thunnus*
628 *maccoyii*, captivity and antiparasitic administration (i.e., praziquantel) have important effects in
629 fish microbiota composition. Healthy fish microbiota, without praziquantel, is dominated by
630 Mycoplasmataceae in its skin, and *Pseudomonas*, *Acinetobacter*, *Brevundimonas*, and *Delfita* in
631 its gut (Minich et al., 2020). In the Yellowtail Kingfish (*Seriola lalandi*), in temperate and
632 southern waters of Australia, early enteritis produces microbial fingerprints in its skin microbiota.
633 Early enteritis is associated with greater abundance of *Loktanelia*, *Marivita*, and *Planktomarina*,
634 *Simplicispira*, and *Litoricola*, as well as decreased diversity in the microbial community (Legrand
635 et al., 2018).

636

637 Fish microbiota also has been addressed under a natural history framework. In four
638 species of wild Antarctic fish (*Trematomus bernacchii* (family Notothenioidei), *Chionodraco*
639 *hamatus*, (family Channichthyidae), *Gymnodraco acuticeps* (family Bathydraconidae), and
640 *Pagothenia borchgrevinki* (family Notothenioidei), gut microbiota has a stable composition
641 among several species (up to 50% sequences are shared among individuals) (Song et al., 2016).
642 This suggests the presence of a core intestinal microbiome in Antarctic fish, despite differences in
643 environment and diet, which might play important roles in fish health.

644

645 Interestingly, fish gut microbiota could serve as a biological prognosis of anthropogenic
646 impact in marine environments, when it is possible to compare historical and contemporary
647 samples. In the Antarctic fish *Trematomus* spp. historical samples (museum samples with 100
648 years old, fixed with formalin and embedded in paraffin) have revealed notable shifts in gut
649 microbiota composition. Contemporary fish gut microbiota was characterized by Chlamydia,
650 Firmicutes, Cyanobacteria and Mycoplasma. In contrast, historical fish gut microbiota was
651 dominated by Proteobacteria. Despite the richer appearance of phyla in contemporary fish, OTU
652 richness and Shannon index diversities were higher in ancient fish (Heindler et al., 2018). These
653 results attempt to elucidate the relationship between fishing practices and fish gut microbiota,
654 under a historical context. Before global fishing practices, fish were able to have a consistent diet
655 that produce redundant gut communities at the phylum level. In contrast, fishing practices
656 disrupted prey availability, which forced fish to become more opportunistic in their feeding,
657 producing gut communities with wider phylogenetic representation, albeit less diversity (Heindler
658 et al., 2018). Whether the shift from more diverse gut microbiotas enclosed in one phylum, to less

659 diverse communities with members spanning several phyla has impacts in fish (and any marine
660 host's) health, is a deep open question.

661

662 In the Strait of Magallanes project that we describe at the end of the paper, we are
663 studying two fish species, the sardine *Sprattus fuegensis* and the farm salmon, *Salmo salar*. We
664 think that the skin microbiome comparison between a wild native species (i.e., sardine) and an
665 introduced species raised in captivity (i.e., salmon) will provide important clues about local
666 marine conditions as well as the effect of captivity.

667

668 **Seabirds**

669

670 Seabirds are important components of the Southern Hemisphere (Woehler et al., 2001).
671 Currently, there are 350 species around the globe, with at least 61 species endemic to the
672 Southern Hemisphere (Croxall et al., 2012). Seabirds' microbiome has received substantial
673 attention in the Southern Hemisphere, with 22 studied species within 16 research articles (Fig. 1,
674 Table 1). Nevertheless, most of these studies are restricted to few penguin species (Table 1) and
675 in most cases have been done with fecal samples (but see Leclaire et al., 2019). However, this is a
676 global trend, since there are only two studies in the Northern Hemisphere addressing feather
677 bacterial communities, most likely because these are challenging species to sample (Pearce et al.,
678 2017; Leclaire et al., 2019). Seabird microbiome sampling effort in the Southern Hemisphere has
679 been performed in most latitudinal regions, temperate (Brazil & Australia), cold (Argentina and
680 Kerguelen) and Antarctic latitudes (Antarctic islands and Western Antarctic Peninsula (Fig. 3).
681 Seabird microbiome has been predominantly conducted with taxonomic marker approaches (i.e.,
682 16S), hence their associated microbes have been predominantly addressed at the bacterial
683 community level. Nevertheless, these studies have been conducted with metatranscriptomics and
684 RNA viromics surveys (Marcelino et al., 2019; Wille et al., 2020).

685 Overall, the main bacterial phyla of penguins' fecal microbiota in the Southern
686 Hemisphere are Firmicutes, Proteobacteria, Actinobacteria and Bacteroidetes (Potti et al., 2002;
687 Barbosa et al., 2016; Dewar et al., 2013, 2014, 2017; Yew et al., 2017; Lee et al., 2019; Tian et
688 al., 2021a, Tian et al., 2021b). Whereas the main bacterial phyla of seabird plumage microbiota in
689 the Southern Hemisphere, is characterized by Actinobacteria, Proteobacteria, Firmicutes, and
690 Acidobacteria (Leclaire et al., 2019).

691

692 Aging is an important driver of fecal microbiota composition, both in wild and captive
693 penguins (Barbosa et al., 2016; Dewar et al., 2017; Tian et al., 2021a). In wild chinstrap penguin
694 chicks (*Pygoscelis antarctica*) fecal microbiota is dominated by Firmicutes, specially by
695 Clostridiales, *Leuconostoc* and *Fusobacterium*. In contrast, adults' fecal microbiota is dominated
696 by Proteobacteria and Bacteroidetes, specially by Neisserales, Fusobacteriales and
697 Campylobacteriales, yet there is high variability among individuals (Barbosa et al., 2016; Lee et
698 al., 2019). In contrast, in captive chinstrap penguins in the Dalian Sun Asia Aquarium, China,
699 aging only changes relative abundances of the main constituents of fecal microbiotas. Chicks'
700 fecal microbiota was dominated by *Acinetobacter*, while *Pasteurella* was the dominant bacteria
701 in senior penguins (between 22-28 years old); the fecal composition was further completed with
702 *Clostridium*, and *Fusobacterium* in chicks and adult penguins (Tian et al., 2021a). Furthermore,
703 these compositional shifts followed predicted functionality shifts. In general, predicted
704 functionality reaches its maximum diversity in adults, while it starts to decline in senior penguins
705 (after 22 years old) (Tian et al., 2021a).

706

707 Support for age of the host as an important driver of fecal microbiota composition comes
708 from the little blue penguin (*Eudyptula minor*), at the Phillip Island Nature Parks, Australia,
709 where fecal microbiota between chicks and adults differs (Dewar et al., 2017). Differences in
710 fecal microbiota composition might be explained by the kind of food that chicks and adults ate.
711 Chicks eat regurgitated food, which might not require a robust microbial metabolic repertoire to
712 aid in digestion; in contrast, adults eat raw food that might contain recalcitrant chemicals, like
713 domoic acid in fish (Lefebvre et al., 2002) or fluoride, from krill (Yoshitomi et al., 2012).

714

715 Besides community compositional comparisons, 16S surveys have been used to
716 understand changes in penguins' fecal microbiome predicted metabolic functions. For example,
717 in captive gentoo penguins (*Pygoscelis papua*) in the Dalian Sun Asia Aquarium, China, sex
718 apparently influences fecal microbiome predicted metabolism (Tian et al., 2021b). Males' fecal
719 microbiota is enriched in carbohydrate metabolism, putatively driven by Lachnospiraceae family,
720 whereas females had a fecal microbiota enriched in protein metabolism, putatively driven by
721 Fusobacteriaceae family (Tian et al., 2021b). Microbiome predicted functions from 16S data are
722 constrained by the number of available microbial genomes sequenced (Douglas et al., 2020) and
723 information on its "optimal performance" is limited to human samples, and decreases sharply in
724 environmental samples (Sun et al., 2020). Therefore, functional microbiome studies (i.e.,

725 metagenomics or metatranscriptomics) coupled with experimental/culture assays are needed to
726 validate the metabolic functions that have been attributed to penguin fecal microbiota.

727

728 Fecal microbiota comparisons between several penguin species have shed light about
729 penguin species specific factors influencing fecal microbiota composition. For example, Dewar et
730 al., (2013) compared the fecal microbiota composition of four penguin species: macaroni
731 penguins (*Eudyptes chrysolophus*), king penguins (*Aptenodytes patagonicus*) and gentoo
732 penguins from two sites, Bird Island in South Georgia and Baie du Marine, Possession Island
733 Crozet Archipelago, and little blue penguins from Phillip Island, Australia. This study revealed
734 interesting patterns in penguin fecal microbiota at phyla level, where the dominant bacterial
735 phylum of each penguin species was as follows: Firmicutes in macaroni penguins, Actinobacteria
736 in king penguins, and Proteobacteria in gentoo and little blue penguins (Dewar et al., 2013). Fecal
737 microbiota divergence among penguin species might be explained by trophic niche differences.
738 Additionally, hormone profiles might also have specific effects in gut microbiota composition.
739 However, differences could also be attributed to geographical factors. Systematic studies, where
740 multiple colonies of each species with an adequate sample size are needed to verify to what
741 extent these results reflect penguin fecal microbiota in these species.

742

743 Penguin's gastrointestinal tract has several differences along its structure. Given that fecal
744 microbiota represents the last section of the gastrointestinal tract, it is unlikely that it is a
745 representative sample of penguins' gastrointestinal microbial diversity. Indeed, stomach
746 microbiota studies conducted in Adélie (*Pygoscelis adeliae*) and chinstrap penguins in Signy
747 Island, South Orkney Islands, Antarctica have revealed bacterial community structure between
748 the stomach and fecal microbiota. Stomach microbiota of Adélie and chinstrap penguins is
749 characterized at the phyla level by Firmicutes, Proteobacteria, Fusobacteria, and Tenericutes.
750 Common genera in both species are: *Cetobacterium*, *Psychrobacter*, *Chelonobacter*, *Clostridium*
751 (family: Clostridiaceae), *Mycoplasma* and *Ornithobacterium* (Yew et al., 2017). These stomach
752 bacterial communities differed from those reported from the fecal microbiota of these species,
753 where Actinobacteria and Firmicutes were the dominant phyla in Adélie penguin fecal microbiota
754 (Banks et al., 2009), while chinstrap fecal microbiota is characterized by Proteobacteria and
755 Bacteroidetes (Barbosa et al., 2016; Tian et al., 2021a). Stomach bacterial differentiation might
756 be explained in part, by the presence of sphenicins, special compounds in the penguin stomach
757 that prevent bacteria from digesting food (Thouzeau et al., 2003).

758

759 Fasting is a critical event inherent to penguins' life history. Penguins experience fast when
760 they rear their chicks, and when they molt. Fasting has species-specific compositional changes in
761 the fecal microbiota of penguins (Dewar et al., 2014; Lee et al., 2019). This could be explained
762 by penguin species differences in fast length and their fecal associated microbes. In king Penguin,
763 fasting increases the relative abundance of Proteobacteria, Firmicutes, Actinobacteria,
764 *Fusobacteria* and *Bacteroidetes*. In contrast, in little blue penguin, all phyla but Proteobacteria
765 decreased its relative abundance as fasting progressed (Dewar et al., 2014). *Fusobacteria* is a
766 butyrate producer bacterium. Butyrate is a known anti-inflammatory agent (Canani et al., 2011).
767 Furthermore, butyrate administration in chickens has immune system functional improvement,
768 which improves health and decreases pathogen incidence (Panda et al., 2009). The enrichment of
769 *Fusobacteria* in king penguin fasting, suggest that this bacterium could play an important role in
770 metabolic homeostasis. Further evidence supporting the effect of fasting in penguins comes from
771 gentoo and chinstrap penguins in different trophic status (i.e., feeding season or fasting). In
772 feeding chinstrap and gentoo penguins, *Fusobacteria* and Proteobacteria were the dominant
773 bacteria, while in molting (fasting) birds, these phyla decreased while Firmicutes increased its
774 relative abundance. Nevertheless, the magnitude in the compositional changes of its fecal
775 microbiota composition differed among species. While shifts in chinstrap penguins were subtle
776 (i.e., not supported statistically), shifts in gentoo penguin were major (i.e., statistically supported)
777 (Lee et al., 2019).

778

779 It is worth mentioning, that penguin and Antarctic birds' fecal microbiota studies have shed
780 light in the widespread occurrence of genera close to known pathogens. Those genera include
781 *Campylobacter*, *Yersinia*, *Salmonella*, and *Escherichia* (Barbosa et al., 2009), yet, it is uncertain
782 to which degree they affect penguins' health, but they might have a parasitic basis. Support of
783 this, comes from Magellan penguins from Peninsula Valdez, Argentina where in penguin chicks,
784 *Corynebacterium*, a dominant bacterium in fecal samples appears to divert resources from the
785 chick, impairing its growth., but administration of a wide spectrum antibiotic, reduced
786 *Corynebacterium* abundances, reversing growth halting (Potti et al., 2002).

787

788 On the other hand, the presence of potential pathogens in Antarctic animals can be a
789 misinterpretation of what "is normal" and what is human related (Souza et al., 1999). The same
790 may occurs with virus associated with Antarctic fauna. For example, chinstrap, Adélie, and

791 gentoo penguins harbor a great diversity of virus in their cloaca and of their ectoparasitic mites
792 (Wille et al., 2020). These results highlight the uniqueness fauna living in the Antarctic biome.
793 Moreover, it calls for further microbiome research in these remote places, in order to elucidate
794 their relationship with worldwide fauna.

795

796 Seabird plumage microbiota has been poorly studied in the Southern Hemisphere, yet, we have
797 some insights from the blue petrel, *Halobaena caerulea*, whose plumage microbiota composition
798 is highly variable among body sites (Leclaire et al., 2019). Furthermore, some bacteria show a
799 positive correlation with MHC (Major histocompatibility index), which suggests that plumage
800 bacteria are influenced by MHC allele diversity in this seabird.

801

802 Interestingly, a metatranscriptomic approach revealed a high incidence of antibiotic resistant
803 genes in several seabirds from Australia with different trophic ecologies (Marcelino et al., 2019).
804 Synanthropic (living near human settlement) species with filter eating habits, like several
805 Australian ducks (*Anas* spp. and *Tadorna tadornoides*), had the highest diversity of antibiotic
806 resistant genes, whereas avocets (*Recurvirostra novaehollandiae*) and gentoo penguins, which
807 live in remote areas and prey invertebrates and fish, respectively, had the fewest. However, the
808 presence of antibiotic resistant genes is not a surprise since this seems to be a very ancient
809 strategy in microbial communities (Souza et al., 1999). Noteworthy, although the cloacal
810 microbiome of gentoo penguins had the less diversity of antibiotic resistant genes, it displayed
811 resistance against unique drugs, like Macrolides, Lincosamide, and Streptogramin (Marcelino et
812 al., 2019).

813

814 In a similar fashion, kelp gulls (*Larus dominicanus*) and Magellan penguins from Brazil
815 were assessed by qPCR to evaluate the diversity of antibiotic genes they harbor (Ewbank et al.,
816 2021). Ecological strategies (synanthropic / remote, migratory / non, opportunistic / specialized
817 feeding) might have a strong association with antibiotic gene resistance transmission, with those
818 related to anthropocentric activities having the greatest diversity in antibiotic resistance genes. As
819 expected, kelp gull, a synanthropic species, has the greatest diversity in antibiotic resistance
820 genes. Its antibiotic resistant gene pool has resistance against eight drugs: tetracycline,
821 aminoglycosides, sulfonamides, chloramphenicols, macrolides, quinolones, betalactams,
822 polymyxins. On the contrary, in the Magellan penguin, a migratory, non-synanthropic,
823 specialized feeder (it preys mainly fish and squid) their antibiotic resistance genes diversity was

824 lower, with specific resistance against two drugs, tetracycline and quinolone (Ewbank et al.,
825 2021).

826

827 In our project on the surface microbiome of key species in the Strait of Magellan, we are
828 planning to study the feather microbiome in Magellan and King penguins. We are interested in
829 testing geographical, phenological, and developmental effects in feather microbiome of these
830 species. These works will provide original baseline knowledge regarding penguin feather
831 associated bacteria.

832

833 **Marine mammals**

834

835 Marine mammals comprise several species that collectively are considered as “marine sentinels”,
836 since their population trends could give valuable insights about the status of the marine
837 ecosystem (Moore, SE, 2008). Currently, there are 115 species distributed around the globe
838 (Kaschner et al., 2011). Up to one third of them are cooccur between the template – cold latitudes
839 (i.e., 20-50) of the Southern Hemisphere. Important hotspots of marine mammals’ biodiversity:
840 New Zealand, Sub-Antarctic and Southeastern Pacific islands, and offshore waters along the
841 coasts of southern South America (Kaschner et al., 2011). Marine mammals’ microbiome has
842 received the greatest attention in the Southern Hemisphere, with 15 studied species within 17
843 research articles (Fig. 1, Table 1). Nevertheless, most studies have been conducted in one species
844 (humpback whale) (Table 1). In most cases, these studies have been conducted with 16S
845 approaches, but there is a pinniped shotgun metagenome (Table 1, Smith et al., 2013).
846 Interestingly, marine mammals’ microbiome research has been done worldwide, with sampling
847 effort across all latitude regimes (tropical, template, cold and Antarctic), specifically in template
848 latitudes (i.e., Australia), followed by Antarctica and South America (Fig. 3).

849

850 We still lack a more comprehensive view of cetaceans’ microbiome in the Southern
851 Hemisphere, since most of current studies have been done in one species over different
852 geographic locations (i.e., humpback whale) (Table 1). In contrast with cetacean microbiome
853 knowledge in the Northern Hemisphere where several species have been addressed (Sanders et
854 al., 2015; Van Cise et al., 2020; Apprill et al., 2020; Miller et al., 2020). While this gives us depth
855 in understanding humpback whale microbiome across geographic regions, it also pinpoints to the

856 great void that remains in the rest of cetaceans, like whales and dolphins. Besides humpback
857 whale, there have been conducted several studies in pinnipeds (Table 1).

858

859 In particular, marine mammals' microbiome research has been focused in whale skin and
860 blow microbiota studies. In the Southern Hemisphere skin microbiological surveys have been
861 done in rorquals (Balaenopteridae, species *Megaptera novaeangliae* [humpback whale],
862 *Balaenoptera musculus*, and *B. physalus*) as well as killer whales (*Orcinus orca*). Overall, the
863 main bacterial elements in the whale skin microbiota are Proteobacteria, Bacteroidetes,
864 Actinobacteria, and Firmicutes (Apprill et al., 2014; Pirotta et al., 2017; Bierlich et al., 2018;
865 Hooper et al., 2019; Vendl et al., 2019; Vendl et al., 2020; Toro et al., 2021).

866

867 Humpback whales are the best studied cetaceans in the Southern Hemisphere. Studies
868 from humpback whales in the South Pacific (Samoa islands), Chilean coasts and Antarctic
869 regions have allowed the detection of a skin core microbiota, characterized by *Tenacibaculum*
870 and *Psychrobacter* (Apprill et al., 2014; Bierlich et al., 2018; Toro et al., 2021). Rorquals
871 (*Megaptera novaeangliae*, *Balaenoptera musculus*, and *B. physalus*) skin microbiota along
872 several points in Chilean coasts have idiosyncratic and species-specific trends, that is, whales
873 have a unique skin microbial composition at the individual and the interspecific level. However,
874 their skin microbiota alpha diversity was similar in compositional terms (i.e., Shannon diversity),
875 but was slightly different in phylogenetic terms (i.e. Faith phylogenetic diversity) (Toro et al.,
876 2021).

877

878 Skin microbiota of humpback whales foraging in Antarctica and the Strait of Magellan are
879 enriched in *Psychrobacter* bacteria. Changes in sea surface temperature, as well as shifts towards
880 northern areas, are associated with decreases in *Psychrobacter* relative abundance. This pattern
881 suggests that sea temperature is an important driver in humpback whale skin microbiota assembly
882 (Bierlich et al., 2018; Toro et al., 2021).

883

884 Blow microbiota of Australian humpback whales has been surveyed to address whales'
885 health. In contrast with skin microbiota, blow microbiota is sparse among individuals, without
886 any discernable core. The most abundant microbes in the blow were *Tenacibaculum*,
887 *Pseudomonas Leptotrichia* and *Corynebacteria*. Additionally, some individuals had potential
888 respiratory pathogens in their blow microbiota, like *Balneatrix*, *Clostridia*, *Bacilli*,

889 *Staphylococcus* and *Streptococcus* (Pirota et al., 2017; Vendl et al., 2019). Furthermore, whale
890 blow associated bacteria harbors significant phylogenetic novelty, since half sequences in some
891 individuals were only able to identify at class level (Vendl et al., 2019). Whale feeding
892 phenology is an important factor that could underlie the sparsity of blow microbiota structure.
893 While feeding blow microbiota has a core microbiota composed of *Arcobacter*,
894 *Corynebacterium*, *Enhydrobacter*, *Helcococcus* and *Tenacibaculum*, albeit in very low
895 abundance, 1.5% or less. In contrast, during migration, when whales are fasting, blow microbiota
896 among individuals becomes highly variable, with no discernible core (Vendl et al., 2020a).
897 Besides phenology or health status, whale blow microbiota is also influenced by the sociality
898 degree of the studied species. Whale species with gregarious habits (e.g., humpback whale), have
899 higher diversity and a great microbial core in their blow microbiota, in contrast with more
900 solitary whales (Vendl et al., 2020b). Likely, this could reflect the horizontal transmission of
901 blow microbes among contiguous whales, where the microbes exhaled by one individual are
902 inhaled by another, and so on. Nevertheless, more studies are needed to address if this pattern
903 emerges because of horizontal transmission or could reflect common health status among whales'
904 groups.

905

906 In the case of Killer whales, the Antarctic ecotypes have been found to harbor a distinct
907 skin microbiota from ecotypes in the northern hemisphere. Differences are driven by
908 *Tenacibaculum dicentrarchi* bacteria, diatoms, and several algae-associated bacteria (Hooper et
909 al., 2019). More systematic studies of whales associated bacteria, and ideally of an “authentic”
910 holobiont approaches (i.e., describing whale genetic traits, as well as their microbiome and
911 transcriptome) to better understand the whale holobiont.

912

913 In our project on the surface microbiome of key species in the Strait of Magellan (ANID
914 R20F0009), we are including a holobiont approach to study the humpback whales of the area. In
915 particular, we are sampling individuals that migrate to the Strait of Magellan to feed in the austral
916 summer season.

917

918 Pinnipeds are apex predators whose health might inform about marine ecosystem conditions
919 (Moore, SE 2008). Sampling pinnipeds is challenging due to the remote location of their colonies
920 and proclivity to escape from humans, hence, the most feasible samples to study them, are the
921 feces they leave in rocks. Seal fecal microbiota apparently is driven by several host factors,

922 including feeding, geographic distribution, ontogeny, trophic niche differences, anatomy, and
923 physiology, explaining, at least in part, the great fecal microbiota composition variability among
924 species, where there is no detectable fecal core microbiota. Nevertheless, it is uncertain if the
925 taxonomic differences are congruent with microbiome metabolic traits.

926

927 Overall, the main bacterial components of seal fecal microbiota are Firmicutes,
928 Fusobacteria, Bacteroidetes, Proteobacteria, and Actinobacteria (Nelson et al., 2013; Delpont et
929 al., 2016; Grosser et al., 2019; Kim et al., 2020; Toro-Valdivieso et al., 2021). Nevertheless, fecal
930 microbiota from pinnipeds has been a challenging task as there seems to be a high proportion of
931 phylogenetic novelty that interferes with taxonomic identification (i.e., at the family or genus
932 level) and proper analysis of the divergence among communities (Toro-Valdivieso et al., 2021).
933 For example, across pinniped species, fecal microbiota composition at phylum level seems to be
934 identical (Nelson et al., 2013; Kim et al., 2020) but diverges at lower taxonomic levels. For
935 instance, in the southern elephant (*Mirounga leonina*) and Weddell seals (*Leptonychotes*
936 *weddelli*) at the phylum level, fecal microbiota is dominated by Firmicutes, yet at the family level
937 Ruminococcaceae and Acidaminococcaceae, respectively, drive differences in the fecal
938 microbiota composition (Kim et al., 2020). Life history characteristics create complex patterns
939 that influence fecal microbiota, as shown with the fecal microbiota of southern elephant seals and
940 leopard seals (*Hydrurga leptonyx*). In these species, fecal microbiota is shaped by the
941 simultaneous effect of species, age and sex, creating complex fecal microbiota patterns with no
942 discernable trend (Nelson et al., 2013). In the Australian fur seal (*Arctocephalus pusillus*) aging
943 from pups to adults produces a successional pattern in fecal microbiota composition. Adult fecal
944 microbiota had unique bacterial taxa, dominated by *Clostridium*, *Lactobacillus* and
945 *Enterococcus*. The diet shift from milk with high fat-protein in pups to a marine raw diet in
946 adults, is thought to underlie the fecal microbiota diversification (Smith et al., 2013).

947

948 A fecal microbiota survey in the Australian sea lion (*Neophoca cinerea*) showed a core fecal
949 microbiota at family level, composed of Clostridiaceae bacteria. This core arises only in wild
950 seals, which suggests that natural diet might represent a cohesive driver in fecal microbiota
951 composition. Instead, captive animals lacked Clostridiaceae bacteria in their fecal microbiota.
952 Moreover, seals from wild colonies had more fecal microbial diversity than captive colonies,
953 especially those with high densities (Delpont et al., 2016). At the functional level, the Australian
954 sea lion fecal microbiome is enriched in carbohydrate metabolism, nitrogen biosynthetic and

955 nutrient transport pathways, as well as virulence genes. The fecal microbiome composition
956 appears to play an important role in fat – nutrient storage, key issues in marine mammals’
957 survival in polar ecosystems (Lavery et al., 2012). This could pinpoint to a fat storage mechanism
958 driven, or at least with high influence, of gut microbes. Nevertheless, a higher sample and more
959 phylogenetic inclusivity (i.e., more seal and sea lion’s species) is needed to validate the
960 prevalence of this microbiome traits among populations and pinniped species. Interestingly,
961 Clostridiaceae bacteria has a high prevalence among Australian seal species fecal microbiota
962 (Lavery et al., 2012; Smith et al., 2013; Delpont et al., 2016), which might suggest an important
963 role for these bacteria in seals digestion. Nevertheless, it is a cautionary interpretation since gut
964 microbiota composition varies along the gastrointestinal tract. Hence, fecal microbiota reflects
965 only a portion of the gut microbiome.

966

967 In the case of skin microbiota, there is one example from the Antarctic fur seal
968 (*Arctocephalus gazella*) (Grosser et al., 2019). In this fur seal, colony density drives skin
969 microbiota structure, rather than genetic similarity. High density colony had fewer alpha diversity
970 in its skin microbiota. This suggests that stress associated with overcrowding could scale to skin
971 microbiota composition, diminishing the richness and abundance of bacterial taxa. Nevertheless,
972 overcrowding, also might facilitate horizontal transmission since fur seals are in close contact.
973 This could allow fast transmission of opportunistic microbes, capable of dominating the
974 community. Most likely, the combination of both explanations might aid in illuminating the
975 pattern; stress produced by overcrowding, coupled with increased transmission of fast
976 reproducing microbes, shape the skin microbiota of colonies with high density. (Grosser et al.,
977 2019).

978 In our Strait of Magellan project (ANID R20F0009), we are studying the skin surface
979 microbiome of colonies of the South American sea lion (*Otaria byronia*), a keystone species in
980 the sub-Antarctic ecosystem.

981

982 **Discussion and perspectives**

983

984 **Microbiomes in the southern ecosystems and climate change context.**

985

986 In the southern ecosystems, including Antarctica, it has been predicted that higher temperature
987 will increase coastal ice-free areas, sea-ice loss, glacial retreat, ocean acidification and ocean

988 warming (Morley et al., 2020), affecting marine biota in all trophic level. Lower trophic levels
989 are expected to move south, depending on their tolerance of warming ocean condition and
990 productivity, meanwhile, ocean acidification will impact mainly over crustaceans and calcifying
991 organisms. Marine animals, mammals, and seabirds are expected to move to alternative locations
992 for food and breeding, for survivor and adaptation (Constable et al., 2014). However, the impact
993 of climate change over organisms and their associated microbiota as a whole—i.e., the holobiont
994 – has been poorly studied across the metazoan spectrum, with notable exceptions in some
995 invertebrates and macroalgae (Table 1). Yet, we still lack ecosystem holistic approaches (i.e.
996 including abiotic variables or ideally holobiont experimental assays where important abiotic
997 parameters, like temperature and pH are controlled), that could give us insight about marine
998 holobiont adaptability potential to the environmental changes that soon will face as a
999 consequence of anthropogenic climate change.

1000

1001 We think that the relation between the holobiont and their ecosystems is bidirectional,
1002 where the environmental changes. will affect the host and their associated microbes, meanwhile
1003 changes in the host like abundance, behavior, feeding, molting and reproduction; and their
1004 microbiome like diversity, taxonomic composition, and nutrient recycling, will impact over the
1005 environment. For example, soils impacted by penguins and pinnipeds presented high amount of
1006 nutrients such carbon, nitrogen and phosphorous (Ugolini, FC 1972; Tatur et al., 1990). Also,
1007 marine animals' impact over greenhouse gases emissions, penguins and pinniped settlement had
1008 been founded a hotspot of CO₂, methane (CH₄) and nitrous oxide (N₂O) emissions (Zhu et al.,
1009 2008; 2009). In these processes animals had a huge impact over the coastal sediments where they
1010 colonized through their feces, eggs, prey, carcasses, among others (Guo et al., 2018; Almela et
1011 al., 2022; Ramírez-Fernández et al., 2019). Feces can directly impact over soil microbiome,
1012 seeding gut microbes from marine animals, and indirectly because the high amount of nutrients
1013 that they transport from marine to terrestrial ecosystems (Guo et al., 2018). At functional level,
1014 marine animals increased soil microbial communities related with denitrification pathways
1015 (Ramírez-Fernández et al., 2021), and other nitrogen pathways involved in N₂O emissions.

1016

1017 Climate change studies should take in account the impact of environmental changes in the
1018 host and their microbiome. In our project on the surface microbiome of key species in the Strait
1019 of Magellan, we will measure environmental factors such as water temperature, UV radiation,
1020 chlorophyll content, oxygen levels, pH, salinity and nutrient content to correlates holobiont

1021 abundance, diversity, and distribution, focusing on the skin microbiome due their direct contact
1022 with the environmental parameters. The equilibrium between the holobiont and the ecosystems
1023 will allow their protection and conservation (Carthey et al., 2020).

1024

1025 **Surface microbiome of key species in the Strait of Magellan, an integrative holobiont**
1026 **project**

1027

1028 To increase the number of microbial biodiversity studies in the southern hemisphere, emerges our
1029 microbiome project of key species in the Strait of Magellan in Chile (Fig. 5). Our project will
1030 generate new baseline data for almost all the species that will be sampled (except for humpback
1031 whale skin, which has been extensively studied). Hence our project, has two stages, first baseline
1032 data generation for all considered species. Second, microbiome functional data (i.e.,
1033 metagenomics and metatranscriptomics) as well as ecosystem data integration, where host
1034 microbiome data from multiple years along environmental variables will be integrated. We think
1035 this approach will give valuable insights about host-microbiome responses to ongoing
1036 Anthropocene derived climate change. Moreover, it will generate host microbiome data framed
1037 under an ecosystemic approach, which will be valuable for further comparisons in next years,
1038 where environmental variables likely would have changed.

1039

1040 We consider that microbes are an excellent biodiversity study target, since they can show
1041 fast evolutionary responses to environmental alterations, and they have enormous metabolic and
1042 genetic diversity. We are focusing on surface microbes to test their value as biosensors of climate
1043 change effects in key hosts in the Strait of Magellan. Our project attempts to encompass trophic-
1044 level inclusivity, through sampling hosts in different (yet related) trophic levels, and an
1045 introduced species (i.e., farm salmon). We will sample a primary producer (i.e., *Macrocystis*
1046 *pyrifera*, huiro/kelp), primary consumers (i.e., two crustacean species *Lithodes santolla*, centolla
1047 and *Munida gregaria*, channel prawns), secondary predators (i.e. *Sprattus fuegensis*, Fuegian
1048 sprat; *Spheniscus magellanicus*, Magellan penguin, *Aptenodytes patagonicus*, king penguin and
1049 *Megaptera novaeangliae*, humpback whale) and an apex predator (i.e. *Otaria byronia*, South
1050 American sea lion).

1051

1052 Our project will generate original baseline knowledge of bacterial communities associated
1053 to the surface of some taxa (like penguins' feathers, sea lion fur, fish scales, or crustacea shell).

1054 Furthermore, as we plan to perform multiannual samplings, the project will bring the opportunity
1055 to dilucidate if there are microbial signatures (both at the community and genomic level)
1056 associated to seasonal variation and/or to environmental variables. Our main objective is to test if
1057 there is a core surface microbiome among marine hosts sharing the same environment (i.e., in
1058 particular in Coastal Marine Protected Area “Francisco Coloane”). Alternatively, we are
1059 interested in testing the existence of a core microbiome at different levels. For example, at host
1060 complexity (i.e., a core microbiome for invertebrates, another for mammals, etc.); trophic level
1061 (i.e., a core microbiome for primary consumers, another for primary predators, etc.); or
1062 alternatively a species-specific surface microbiome.

1063

1064 In general, the marine holobiont in the Southern Hemisphere is characterized by few
1065 systematic and authentic holobiont studies (i.e., studies that analyze at the same time host traits
1066 [e.g., genomics, transcriptomics] and microbiome traits [e.g., 16S, metagenome], see Table 1 for
1067 holobiont studies examples). Besides there is a great bias in sampled hosts. For instance, sponges
1068 have been thoroughly sampled, but there is a great gap in marine invertebrate and vertebrate
1069 hosts. We highlight the case of marine invertebrates, which although have a relatively high
1070 number of studied species (23), it remains a tiny fraction of its huge diversity.

1071

1072 Holobiont studies are dominated by microbiota approaches, where the focus is centered in
1073 taxonomic patterns across ecological/life story conditions. These studies have highlighted the
1074 prevalence of Proteobacteria among a wide spectrum of hosts, while specific host-phyla
1075 associations complete the bacterial community. Noteworthy, microbiota approaches have been
1076 centered in the bacterial fraction, completely ignoring the potential role of fungi, virus and
1077 archaea in the marine host holobiont. Notable exceptions where these groups have been
1078 addressed are macroalgae (fungal communities), penguins (viromics) and sponges (archaea).
1079 Nevertheless, there is no study that addresses the interactions among all these groups
1080 simultaneously, likely because technical and economic challenges (i.e., computational resources,
1081 computational skills).

1082

1083 Microbiota studies are important and economically feasible explorations, yet detailed
1084 microbiome studies including metagenomics and metatranscriptomics studies are needed to
1085 inclusively address the microbe community (fungi, eukaryotes, virus, bacteria, and archaea) as
1086 well as its functional potential along with its host. Moreover, functional holobiont data (i.e., host

1087 genomics & metagenomics/metatranscriptomics) coupled with environmental data, might provide
1088 valuable insights about the influence of ecosystemic status in host associated microbiome, which
1089 in turn could be tested for the plausibility of the microbiome to reflect host stress.

1090

1091 Microbiome studies might help in elucidating whether patchy/heterogenous distribution in
1092 microbe taxonomic profiles has different functional potential, or rather, if distinct taxonomic
1093 profiles have convergent/redundant functional profiles. Additionally, microbiome studies might
1094 help to elucidate if there is a microbiome functional profile associated with eukaryote hosts, or
1095 even among distinct hosts if there are core functions among their microbiomes. Finally, sampling
1096 must span as many individuals as possible, as well as geographic and seasonal
1097 (longitudinal/annual) representability to address whether patchy distributions among associated
1098 microbes are a natural feature of marine holobionts or a consequence of low sampling. Such a
1099 systems biology approach might bring further understanding in the complex interplay among
1100 microbes and its hosts, as well as the impact it might carry for them. We think that our project on
1101 the surface microbiome of key species in the Strait of Magellan will provide valuable information
1102 on the points mentioned above, which will contribute to the knowledge of microbial diversity in
1103 the region, as well as their current responses under Anthropocene-derived climate change.

1104

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1116

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Table 1 (on next page)

Table 1. Summary of marine host microbiome research sampling effort in the Southern hemisphere

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Macroalgae				
Number of species sampled	Species repetitively studied (number of studies)	Predominant molecular approach	Functional microbiome/holobiont approaches	Regions repetitively sampled (number of studies)
24	<i>Ecklonia radiata</i> (4)	16S	DNA holobiont (Wood et al., 2022) Shotgun metagenomics (Song et al., 2018)	Antarctica (3) Australia (5)
Sponges				
65	<i>Mycale acerate</i> (4)	16s	Shotgun metagenomics (Moreno-Pino et al., 2020, 2021, Yang et al., 2022)	Antarctica (13)
Marine invertebrates				
23	<i>Euphasia superba</i> (2)	16S	Shotgun metagenomics (Ooh et al., 2022) DNA & RNA holobiont (Lan et al., 2021; Osvatic et al. 2023)	Antarctica (4) Australia (3)
Fish				
9	NA	16S	NA	Australia (2)
Seabirds				
22	<i>Aptenodytes patagonicus</i> (2) <i>Eudyptula minor</i> (2) <i>Pygoscelis adeliae</i> (3) <i>Pygoscelis antarcticus</i> (4) <i>Pygoscelis papua</i> (4) <i>Spheniscus magellanicus</i> (2)	16S	Metatranscriptomics (Marcelino et al., 2019)	Antarctica (5) Australia (4) Bird Island, South Georgia (2)
Marine mammals				

15	<i>Megaptera novaeanglia</i> (6) <i>Balaenoptera musculus</i> (2) <i>Neophoca cinerea</i> (2) <i>Mirounga leonine</i> (2)	16S	Shotgun metagenomics (Lavery et al., 2012)	Antarctica (4) Australia (6) Chile (2)
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Figure 1

Marine microbiome studies in the Southern Hemisphere

Figure 1. Total marine microbiome studies in the Southern Hemisphere reported in this review (75). The most studied groups are marine mammals (17 studies), followed by sponges (16), seabirds (15), macroalgae (13), invertebrates (10), and fish (4).

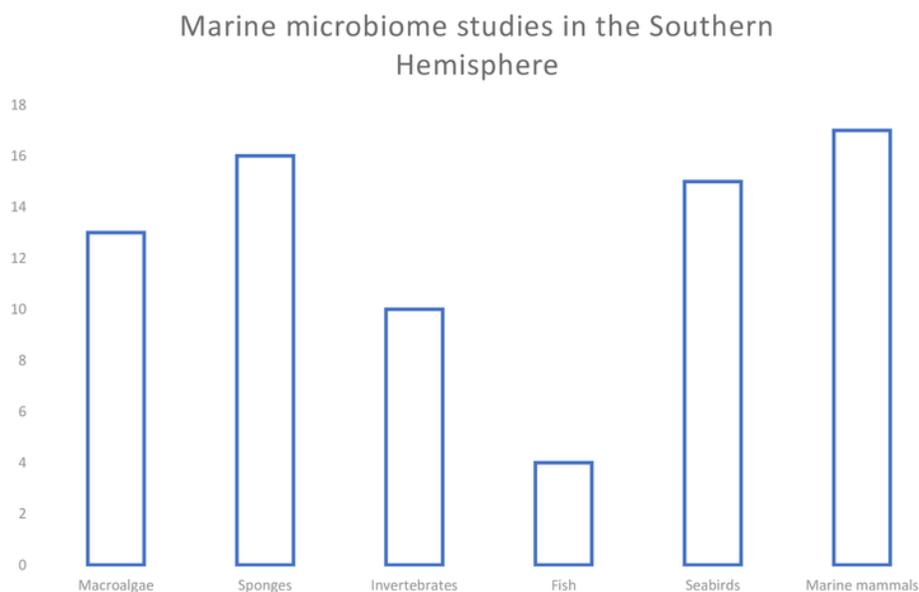


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Figure 2

Latitudinal distribution of microbiome studies in the Southern Hemisphere

Figure 2. Southern Hemisphere map latitudinal distribution of host microbial communities sampling (A) Worldwide Southern Hemisphere. Insets with highly sampled regions. (B) Western Antarctic Peninsula. (C) Australia.

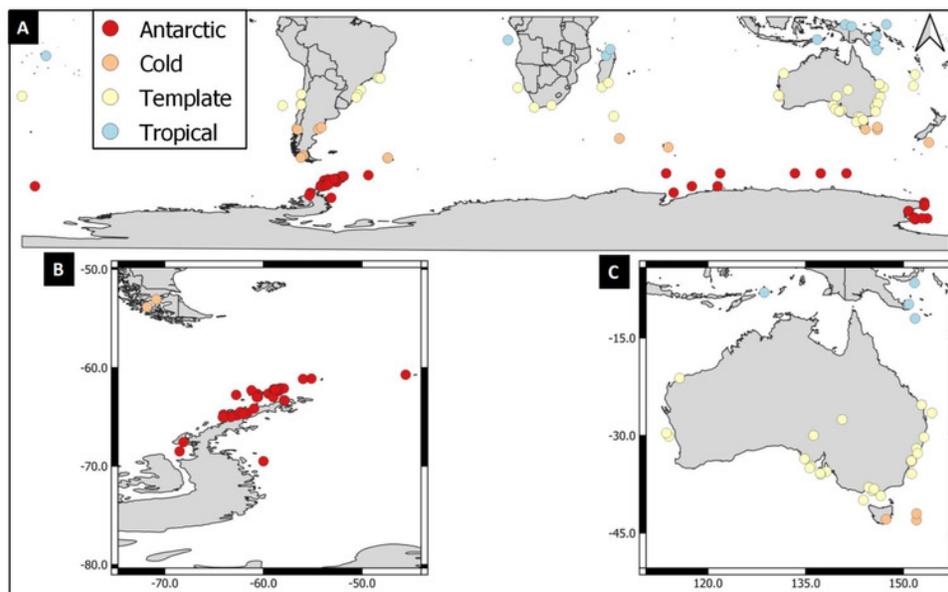


Figure 2. Southern Hemisphere map latitudinal distribution of host microbial communities sampling, A) worldwide Southern Hemisphere. Insets with highly sampled regions, B) Western Antarctic Peninsula, C) Australia.

Figure 3

Host microbiome sampling distribution in the Southern Hemisphere

Figure 3. Southern Hemisphere map of studied hosts distribution. (A) worldwide Southern Hemisphere. Insets with highly sampled regions. (B) Western Antarctic Peninsula. (C) Australia.

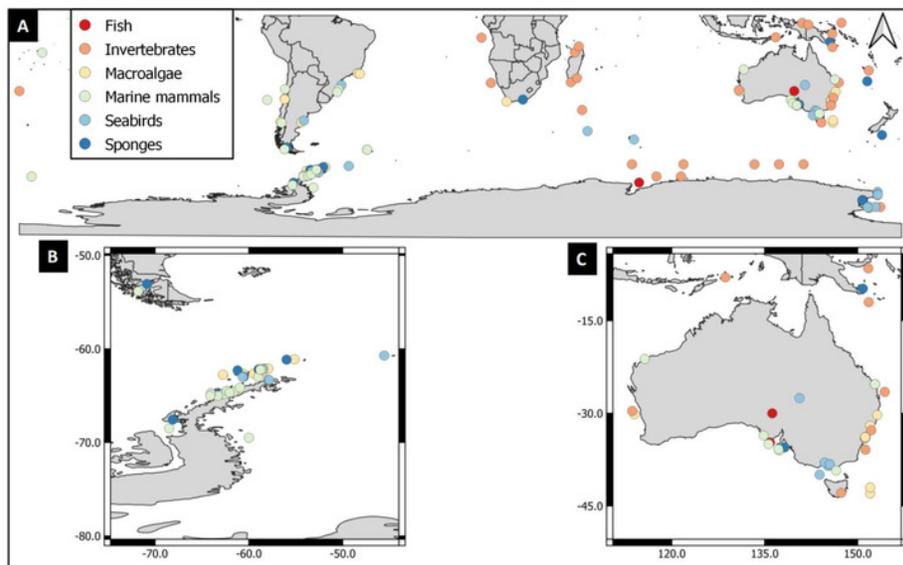


Figure 3. Southern Hemisphere map of studied hosts distribution, A) worldwide Southern Hemisphere. Insets with highly sampled regions, B) Western Antarctic Peninsula, C) Australia.

Figure 4

Southern Hemisphere distribution of molecular approaches to study marine host microbiomes

Figure 4. Southern Hemisphere map of molecular approaches used to study host microbial communities. (A) worldwide Southern Hemisphere. Insets with highly sampled regions, (B) Western Antarctic Peninsula. (C) Australia.

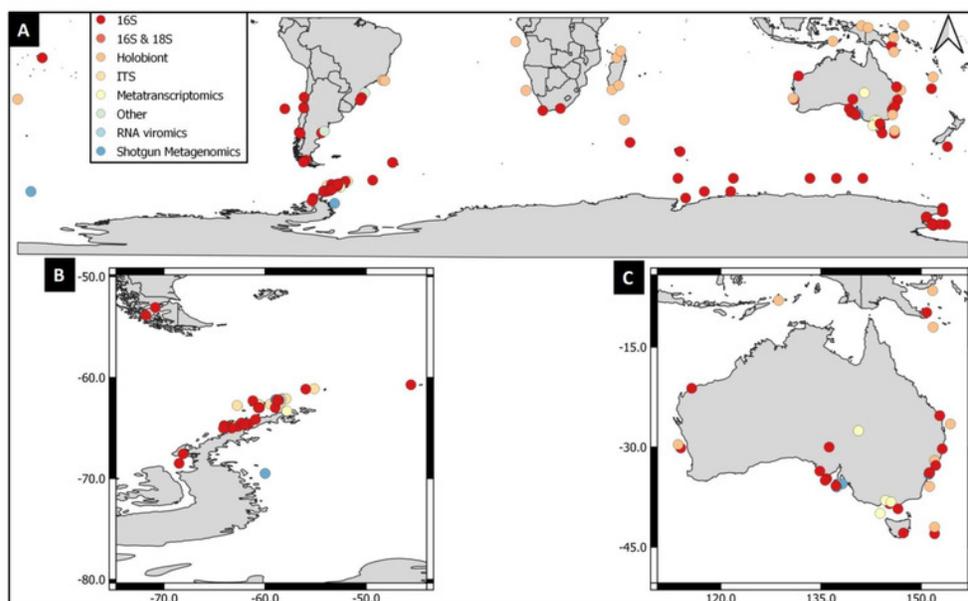


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Figure 5(on next page)

Summary of the Strait of Magellan surface microbiome of key taxa project

Figure 5. Target species and main areas of field sampling of the microbiome project. (A) Target species of the microbiome project are (from top left to right): kelp / huiro (*Macrocystis pyrifera*), Fuegian sprat (*Sprattus fuegensis*), channel prawns (*Munida gregaria*), centolla (*Lithodes santolla*), Magellanic penguin (*Spheniscus magellanicus*), King penguin (*Aptenodytes patagonicus*), Humpback whale (*Megaptera novaeangliae*), South american sea lion (*Otaria byronia*), Atlantic salmon (*Salmo salar*). (B) Main areas across the Strait of Magellan that field work is taking place. The lower right side of Figures 5 (A-B) shows photographs of Carlos III Island, Contramaestre Island, King Penguin Reserve and Tuckers Islands.

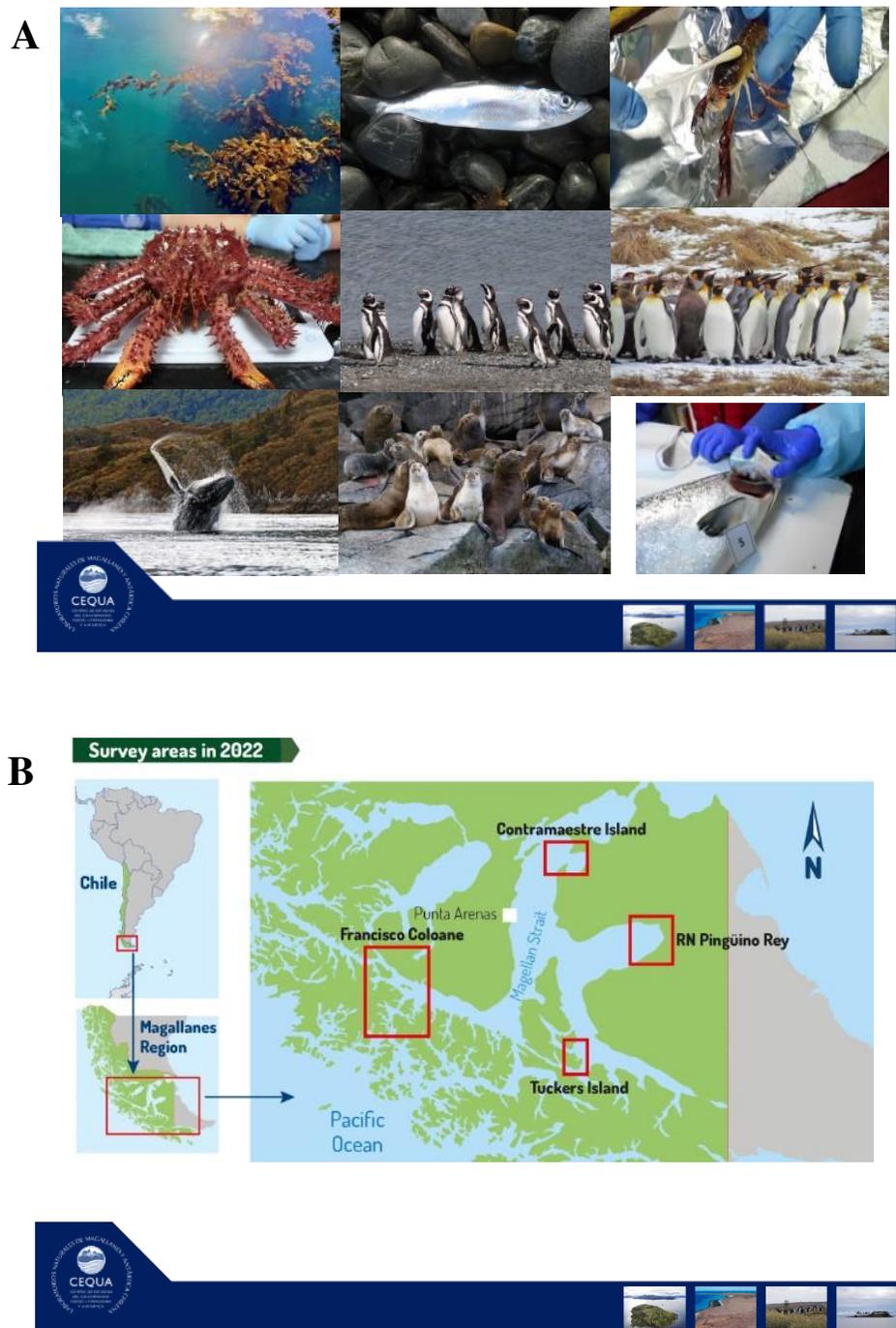


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