

# Current knowledge in the Southern Hemisphere marine microbiome of eukaryotic hosts and the Strait of Magellan surface microbiome project.

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Host-microbe interactions are ubiquitous, and they play important roles in host biology, ecology, and evolution. Yet, host-microbe research has focused on inland species, whereas marine hosts and their associated microbes remain largely unexplored, especially in developing countries located in the Southern Hemisphere. Here, we review current knowledge of microbial communities associated with marine hosts in the Southern Hemisphere. There are important biases in marine host species sampled for the studies in the Southern Hemisphere, where sponges and marine mammals have received the greatest attention. Sponges associated microbes varied greatly across regions and species, nevertheless, beside taxonomic heterogeneity, the microbiome has functional consistency, whereas marine mammals' microbiome seems to be driven by geography and aging. Southern Hemisphere seabirds and macroalgae holobiont studies are also common. Seabirds associated microbes rely on amplicon studies, focused on feces. Seabird fecal microbiota is influenced by aging, sex, and species' specific factors, while macroalgae apparently actively filters the microbes that establish in its surface, in a process known as "microbial gardening". In contrast, marine invertebrates and fish microbiomes have received less attention in the Southern Hemisphere. In general, the marine holobiont in the Southern Hemisphere is characterized by few systematic and authentic holobiont studies (i.e., studies that analyze at the same time host traits [e.g., genomics, transcriptomics] and microbiome traits [e.g., 16S, metagenome]), except for some marine invertebrates and macroalgae. We believe that our project on the surface microbiome of key species in the Strait of Magellan will provide valuable information on the points listed above, which will improve the knowledge of microbial diversity in the region, as well as its current responses under the Anthropocene.

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## Abstract

Host-microbe interactions are ubiquitous, and they play important roles in host biology, ecology, and evolution. Yet, host-microbe research has focused on inland species, whereas marine hosts and their associated microbes remain largely unexplored, especially in developing countries located in the Southern Hemisphere. Here, we review current knowledge of microbial communities associated with marine hosts in the Southern Hemisphere. There are important biases in marine host species sampled for the studies in the Southern Hemisphere, where sponges and marine mammals have received the greatest attention. Sponges associated microbes varied greatly across regions and species, nevertheless, beside taxonomic heterogeneity, the microbiome has functional consistency, whereas marine mammals' microbiome seems to be driven by geography and aging. Southern Hemisphere seabirds and macroalgae holobiont studies are also common. Seabirds associated microbes rely on amplicon studies, focused on feces. Seabird fecal microbiota is influenced by aging, sex, and species' specific factors, while macroalgae apparently actively filters the microbes that establish in its surface, in a process known as "microbial gardening". In contrast, marine invertebrates and fish microbiomes have received less attention in the Southern Hemisphere. In general, the marine holobiont in the Southern Hemisphere is characterized by few systematic and authentic holobiont studies (i.e., studies that analyze at the same time host traits [e.g., genomics, transcriptomics] and microbiome traits [e.g., 16S, metagenome]), except for some marine invertebrates and macroalgae. We believe that our project on the surface microbiome of key species in the Strait of Magellan will provide valuable information on the points listed above, which will improve the knowledge of microbial diversity in the region, as well as its current responses under the Anthropocene.

## Introduction

The Southern Hemisphere, and particularly the Southern Ocean and its associated ecosystems, is characterized by its unique biodiversity (Rogers et al., 2020; Gutt et al., 2021). These important ecosystems are facing major abiotic challenges as climate change progresses. These challenges are driven primarily by ocean warming and increased UV radiation (Thompson & Solomon, 2002; Swart et al., 2018). Sea surface warming creates a stronger stratification in the water column (Pellichero et al., 2017), as well as higher variability in the duration and extent of sea ice sheet and increases in glacier melt rate in the southernmost regions (Gutt et al., 2015; Comiso et al., 2017). Additionally, ocean warming increases microplankton metabolic activity, which in turn accelerates oxygen depletion in the water column (Schmidtke et al., 2017), and decreases

ocean pH (McNeil & Matear 2008). Tragically, recent intense wildfires and volcanic eruptions in the Southern Hemisphere have increased the Antarctic ozone hole size in 2020-2021, which is expected to worsen ocean warming effects (Yook et al., 2022). On the other hand, atypical glacier melt rate are stimulating marine primary productivity, which creates complex scenarios in ice dependent species (Piñones & Fedorov 2016). For example, Antarctic krill (*Euphasia superba*), a key species in the Antarctic trophic network, requires ice in their early stages, while also forages in areas with high concentration of chlorophyll-a concentration (Kawaguchi et al., 2006). Current evidence suggests that habitat quality heterogeneity along its Antarctic distribution will produce contractions in its distribution (Atkinson et al., 2019; Veytia et al., 2020, but see Cox et al., 2018)

However, there are also some surprising results. For example, warmer temperatures coupled with low to moderate winds, increase ice melt, which in turn is expected to increase iron release (Hodson et al., 2017). Iron is a primary productivity limiting factor, so its increased availability triggers diatom growth, which in turn increases krill recruitment (Noble et al., 2013; Bertrand et al., 2007, 2015). Ultimately, krill biomass increase provides greater resources for predators, which overall increases energy transfer along the trophic network (Saba et al., 2014). As climate change progresses, it is expected an intensification in seasonality, which might intensify alterations in biological processes (e.g., bottom-up mechanisms).

For instance, in the Strait of Magellan – the southernmost continental region of South America – climate change derived effects have been recorded since the second half of the XX and early XXI century. These include, increasing sea surface temperature (Smith & Renynolds, 2004) and higher glacier melt rate (Aniya, 1999; Dixon and Ambinakudige, 2015). In the terrestrial ecosystem, warmer seasonality is expected to increase aridity in the Patagonian region, particularly in areas with herbaceous vegetation (Soto-Rogel et al., 2020). Nevertheless, the region has not experienced any formal study regarding the effect of climate change in any of its ecosystem properties (e.g. trophic network interactions, biogeochemical cycles, environmental status). This is unfortunate since the region offers an invaluable geographic position. From the marine perspective, the Strait of Magellan is uniquely influenced by the Pacific and Atlantic oceans, as well as the Cape Horn current. Additionally, glacier melt seasonal input creates local primary productivity bursts that have bottom-up effects that recruit species from higher trophic status, which overall increases the biodiversity of the region.

Despite all the previous natural history studies and museums filled with macro-organisms' diversity, eukaryotes live inside a wider microbial world. Eukaryotes' homeostasis (e.g., physiology, immunology, and metabolism) is driven or at least greatly influenced by microbes (McFall-Ngai et al., 2013; Cani et al., 2019; Peixoto et al., 2021). As a result, the "hologenome evolution theory" and "holobiont theory" emerged (Zilber-Rosenberg & Rosenberg, 2008; Bordenstein & Theis, 2015). The holobiont term per se, is not new. It was first introduced by Lynn Margulis to describe the biological unit formed between a host and a single inherited endosymbiont (Margulis L. 1991). The novelty around the holobiont concept arose as a direct consequence of the development and cost decrease of Next Sequencing Technologies, which spur host microbial communities research. This has revealed that microbes are ubiquitous in every single metazoan (Simon et al., 2019). Thus, the current holobiont concept refers to a cohesive evolutionary unit formed by the host and its associated microbes (Bordenstein & Theis, 2015; Rosenberg & Zilber-Rosenberg, 2018). In other words, a holobiont is a single ecological unit, comprised of an intricate network of mutualistic, commensalisms, and parasitic relationships between microbes and its host, that are critical for the survival of all organisms involved. Nevertheless, microbial influence in the holobiont might vary among hosts (Hammer et al., 2019), which has been interpreted as flaws in the holobiont concept (Moran & Sloan, 2015; Douglas & Werren 2016). Holobionts could be assembled by ecological, evolutive or neutral process. Hence, a first objective in holobiont research lies in determining the nature of the holobiont assembly (Theis et al., 2016).

In marine hosts – which are unlikely to be studied under experimental designs – holobiont research could be focused on the characterization of the host associated microbes in a relevant environmental and evolutionary framework (Leray et al., 2021). Up to date, metabarcoding approaches have been the most popular methods to tackle this issue. Yet, it only serves as an initial step to characterize the microbial composition, as to test the holobiont hypothesis (i.e., host and associated microbes evolve as a unit) further efforts encompassing host associated microbes' functional characterization (e.g., metagenomic or metatranscriptomic data), coupled with host molecular information (e.g., genomic or transcriptomic data), relevant environmental variables measurement, as well as seasonal replicates, must be done to test the congruence of holobiont response against seasonal fluctuations.

Once we have baseline information about bacterial composition associated to marine hosts, we should be able to recognize hosts whose microbial communities are far from the expected natural variability. Changes in the host's natural composition of microbial communities are collectively termed *dysbiosis* (Zaneveld et al., 2017). Thus, the microbiome itself could be used as a biosensor of host's status (Zolti et al., 2020; Inda & Lu 2020). In marine species, there are several examples of the interplay between host fitness and microbial symbiosis, especially in sponges (Pita et al., 2018 and references within) and algae (van der Loos et al., 2019 and references within). Dysbiosis is the microbial fingerprint underlying the disruption of the host's health and ecology, a highly relevant topic in the current anthropogenic climate change epoch.

Microbial communities routinely colonize metazoan internal (e.g., gut, oral) or external (e.g., skin) tissues (Ross et al., 2019; Diaz et al., 2021; ANID R20F0009). However, they differ in the selective pressures that influence their assemble. While internal microbial communities assembly is influenced by diet and host physiology, external microbial communities assembly is greatly influenced by environmental perturbations that impair host fitness (Byrd et al., 2018; Kuziel & Rakoff-Nahoum 2022). Therefore, we think that surface microbial communities of eukaryotes could serve as valuable tools to survey environmental status. The epidermis/outer surface of eukaryotes is considered a hostile environment, yet it is frequently colonized by microbes. These microbes must cope with constant shedding (in epidermis) molt (in feathers), intense solar radiation exposure, low temperature, pH changes, as well as antimicrobial molecules (Percival et al., 2012). Nevertheless, skin microbes play important roles in the host's health (Apprill et al., 2014), since they are the first line defense against pathogens and actively participate in the host's immune system maturation (Ross et al., 2019).

Marine eukaryotes vary greatly in the nature and complexity of their superficial tissue, so the nature of the surface/skin is a critical issue to consider when studying the marine skin microbiome of any host. For example, algae and fish both have a mucus layer in its surface/skin, but their composition and function are different (Gomez et al., 2013; van der Loos et al., 2019). In birds and mammals, the epidermis is covered by feathers and hair, respectively. Importantly seabirds and marine mammals have developed different strategies to cope with cold water, therefore their epidermis and associated elements (i.e., feathers or hair) is completely different from species inhabiting tropical and temperate latitudes (Ross et al., 2019). On the other hand, crustaceans' outer surface is solely its exoskeleton of chitin, so their adaptations to cold

temperatures are to be discovered both in their microbiome and their physiology. Even though, the unique biodiversity living in the Southern Hemisphere, few holobiont studies have been conducted in the region. Biodiversity studies are becoming increasingly important, as climate change imposes major threats in ecosystems around the globe, especially in cold environments.

Up to date, marine holobiont studies have had spatial and phylogenetic biases; on one hand, most of them have been realized in the tropics or in the Northern hemisphere (most of references within Ross et al., 2019), while, on the other hand, the most studied species have been sessile organisms, like sponges (Figure 1). Therefore, Southern Hemisphere marine host microbiome research has received less attention across the metazoan spectrum. Moreover, there are scarce microbiome studies in marine vertebrate and invertebrates. Here, we review the marine host-microbiota/microbiome interactions occurring in the Southern Hemisphere. This review will aid as a diagnosis of the field progress in the region, as well as to detect knowledge gaps and opportunities for further research. Thus, the review is intended for scientists interested in eukaryotes associated microbes, as well as anyone interested in using microbiota/microbiomes as biosensors of eukaryotes' health.

In this review, our main objectives are i) review the current knowledge regarding marine holobiont and microbiota/microbiome studies in the Southern Hemisphere, and ii) to describe a long-term project that will improve knowledge of microbial communities associated with selected taxa in the Strait of Magellan, Chile (ANID R20F00009). We start with a brief overview of the surface characteristics of several taxa. Later, we review holobiont interactions in a trophic level fashion, first with macroalgae, later with primary and secondary animals' consumers, and finally with predators.

## Survey methodology

We covered all marine microbe-eukaryote interaction reports in the Southern Hemisphere that we were able to find. For this, we performed a comprehensive analysis of literature of the last two decades in the following online databases: PubMed, Science Direct, Scopus and Google Scholar. The search spanned the last two decades and was concluded at 1/February/2023. Only studies in English were selected for further inspection. Several keywords were used to perform the literature search, using the following key words, in combination with the terms, -holobiont-, -

microbiota- and -microbiome-: seaweed, sponge, invertebrate, crustacean, marine vertebrate, whales, seals, seabirds. Articles found to match any of these words were examined to guarantee that their sampling was done in hosts inhabiting the Southern Hemisphere.

## Results

Overall, geographical distribution of marine host-microbial research revealed important biases. Geographical biases are reflected by sampling effort across the Southern Hemisphere, since research has been done predominantly in the Antarctic (above 60° S), especially in the Western Antarctic, and temperate latitudes (between 20 – 40° S), especially in Australia, while tropical (between 0 – 20° S) and cold (between 40 – 60° S) latitudes, as well as South America and Africa have received less attention (Fig. 2). There are also host biases, both in terms of number of studies and number of species studied. The most studied groups are marine mammals and sponges (Fig. 1), which have been studied across all latitudinal regions and continents in the Southern Hemisphere (Fig. 3). Nevertheless, when it comes to the number of species studied the pattern has some changes, while sponges remain as the best studied group with 65 studied species, the following groups are Macroalgae (24 studied species) and marine invertebrates (23 studied species) (Table 1). In general, the predominant molecular approach to study host-associated microbial communities has been with taxonomic markers (i.e., 16S), nevertheless there have been conducted some microbiome functional approaches (i.e., shotgun metagenomics), especially in Australia and Antarctica hosts, whereas South American marine hosts, have had not been studied with any functional approach to date (Fig. 4, Table 1).

## Macroalgae as an ecosystem

Marine macroalgae are important ecosystem engineers that play critical roles in primary production, biogeochemical cycles, and biodiversity recruitment in marine ecosystems (Tuya et al., 2008). Currently, there are 11,017 species of macroalgae species with cosmopolitan distribution, therefore there is a vast microbial-algae symbiotic world that remains unexplored (Guiry & Guiry 2023). Macroalgae microbiome studies in the Southern Hemisphere have received considerable attention with 24 studied species within 13 research articles (Table 1; Fig. 1). Most studies have been conducted with 16S approaches, yet there are microbiome approaches with shotgun metagenomics and DNA holobiont approaches (Table 1). Macroalgae microbiome



sampling has been conducted worldwide, but it has been done predominantly in Australia, Antarctica, and South America (Fig. 3).

Algae are an idoneous niche for aerobic and polymer degrading bacteria, since their photosynthetic activity, and their rich composition in carbon and nutrients (like agar, carrageenan, and cellulose), facilitate the establishment of aerobic and heterotrophic bacteria in their surfaces (de Oliveira et al., 2012). Microbial macroalgal epibiotic communities (biofilms) have been extensively studied in the tropics and the Northern Hemisphere (Malik et al., 2020 and references within). Nevertheless, there have been conducted several macroalgae biofilm studies in the Southern Hemisphere which have revealed Cyanobacteria, Proteobacteria, Firmicutes, Bacteroidetes and Actinobacteria as prevalent bacterial phyla (de Oliveira et al., 2012; Albakosh et al., 2016; Gaitan-Espitia & Schmid, 2020). Fungi are also important players in the macroalgae biofilm composition in the Southern Hemisphere, and different species have been documented from Antarctic seaweeds, being the most prevalent the filamentous fungus *Pseudogymnoascus pannorum*, and the yeast *Metschnikowia australis* (Loque et al., 2010; Godinho et al., 2013; Furbino et al., 2017; Ogaki et al., 2019).

There has been extensive research in the macroalgae holobiont that gives a rich body of evidence to frame the study of macroalgae biofilms. These studies have shown that bacteria and fungi inhabiting the macroalgae biofilm actively interact with its host to influence growth, development, and immune function (van der Loos et al., 2019). Valuable microbes to the algal host appear to be taxonomically restricted among bacteria at higher taxonomic levels, therefore, biofilm composition seems to be redundant at phyla or class. Nevertheless, at lower taxonomic levels (genus/ASV [Amplicon Sequence Variant]/OTU [Operational Taxonomic Unit]), there is a rich spectrum of valuable microbes (Egan et al., 2013; Hollants et al., 2013). Despite taxonomic variability in bacterial genera inhabiting its biofilms, traits among bacteria are shared, which creates biofilms with different taxonomic composition but with apparently similar functions (Egan et al., 2013).

Macroalgae seem to actively recruit its biofilm composition, a process known as “microbial gardening” (Saha & Weinberger 2019). Microbial gardening allows the recruitment of specific beneficial bacteria to the algae. In turn, the recruited microbes release antibiotics, quorum sensing inhibitors and digestive vesicles (Wiese et al., 2009; Romero et al., 2010;

Richards et al., 2017) that collectively shape the biofilm composition. Although, algal biofilm composition varies taxonomically, in space, time and host (Lachnit et al., 2011), it shares important traits related with algae morphogenesis (Wiese et al., 2009) and nutrient supplementation (Hollants et al., 2013). Overall, a “correct” microbial gardening, might confer an adequate biofilm composition, able to produce a cocktail of metabolites that provide protection to the algae from microbial pathogens, like bacteria or diatoms (Saha & Weinberger 2019), or predators like barnacle larvae or mussels (Saha et al., 2018). Interestingly, the benefit conferred to algae by the surface microbiome is not taxonomically restricted, suggesting that microbial traits underlying algae defense are shared by several bacterial taxa.

Besides, some bacteria of the algal biofilm produce antibiotic compounds, which act as a microbial filter for the establishment of environmental microbes (Albakosh et al., 2016). For example, algae’s thallus produces dimethylsulfoniopropionate (Kessler et al., 2018), which recruits *Roseovarius* bacteria. This bacterium releases specific morphogenetic compounds that enable correct algae morphogenesis. Moreover, bacterial metabolites could enhance algal performance during stress. Thallusin, a microbial derived metabolite, positively influences algal growth, cell differentiation, cell wall development, and rhizoid formation during abiotic stress (Alsufyani et al., 2020). Therefore, thallusin production might be an excellent example of a holobiont environmental stress response.

Globally, the algal holobiont will be subject to complex scenarios under climate change. Sea surface temperature, and CO<sub>2</sub> concentration increase, might have different outcomes in algal species and also affect key microbes inhabiting macroalgae biofilms, which in turn, could impair the host’s health (Gaitan-Espitia & Schmid, 2020; Marzinelli et al., 2015). Moreover, it is imperative to consider the effect on the interactions among microbes and microbe-algae. For example, algal hosts sensible to acidification might experience lower photosynthetic rate, which might hamper aerobic bacteria proliferation (Van der Loos et al., 2019). Climate change environmental driven perturbations might produce common disbiotic biofilm composition in stressed macroalgae (Marzinelli et al., 2015). Nevertheless, whether shifts in macroalgae biofilm composition associated to environmental perturbation impairs macroalgae fitness remains unexplored.

## The macroalgae holobiont in the Southern Hemisphere

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299 To the best of our knowledge the macroalgae microbiome has been addressed in 13 studies in the  
 300 Southern Hemisphere (Fig. 1). Most of these studies have been done in temperate latitudes,  
 301 particularly in Australia (Fig. 3). There are important gaps in tropical and cold latitudes, as well  
 302 as in selected regions, like South America, Africa and Indonesia (Fig. 3). Yet, we still ignore  
 303 much about South America native macroalgae microbiome, since there is only one study that  
 304 addressed this topic in a native species, *Macrocystis pyrifera* (Laminariales: Laminariaceae),  
 305 while another explored the biofilm composition of a macroalgae invasive species, *Undaria*  
 306 *pinnatifida* (Laminariales: Alariaceae) (Florez et al., 2019; Lozada et al., 2022). From these  
 307 examples, we know that macroalgae biofilm composition is different from seawater and is  
 308 influenced by seasonality and available nutrients. Interesting, invasive macroalgae in the  
 309 Southern Hemisphere, shows signs of microbial gardening, since its biofilm composition is  
 310 dominated by one bacterial taxon, *Leucothrix*, a gamma-proteobacteria (Florez et al., 2019;  
 311 Lozada et al., 2022). This could give insights about potential adaptation mechanisms of these  
 312 invasive species in the Southern Hemisphere, which might be relevant in the ecosystemic process  
 313 of the Patagonian region.

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315 Australia is the region where the macroalgae holobiont is best understood, since it has the  
 316 highest number of studies (5 studies, Table 1), and diversity of molecular approximations (i.e.,  
 317 16S, shotgun metagenomics, DNA holobiont, Table 1). Yet, most of its research effort has  
 318 focused only on one species, *Ecklonia radiata* (Laminariales, Lessoniaceae), so it is unlikely that  
 319 their macroalgae holobiont knowledge is generalizable to all macroalgae hosts in the region  
 320 (Marzinelli et al., 2015, 2018; Qiu et al., 2019; Song et al., 2021; Wood et al., 2022).  
 321 Nevertheless, it provides valuable information that gives insight into the future that might face  
 322 the macroalgae holobiont. *Ecklonia radiata* biofilm composition has a dysbiotic composition  
 323 during environmental stress (Marzinelli et al., 2015, 2018). Yet, stress microbial signatures are  
 324 not consistent across individuals, which suggest that there are unexplored meaningful covariates  
 325 (e.g., host genetic variability) that determine the final holobiont phenotype (i.e., biofilm  
 326 composition) when the algae face environmental stress (Qiu et al., 2019). Besides, horizontal  
 327 transfer of genes related with the algae niche specific environment, and stress environmental  
 328 responses between biofilm bacterial members, suggest that this process might facilitate biofilm  
 329 adaptation to environmental stress (Song et al., 2021). Interestingly, a DNA holobiont approach  
 330 in the macroalgae *Phyllospora comosa* (Fucales: Seirococcaceae) revealed that host genetic

variability has a weak relationship with microbial composition. Moreover, its biofilm composition is driven by local conditions and geography (Wood et al., 2022). Together these results suggest a complex interplay in the macroalgae holobiont, where genetic variability, biofilm composition, horizontal gene transfer and environmental conditions, are crucial players, that create a diverse array of phenotypes. Given the unavoidable environmental changes that we will face in the next decades, and the key role macroalgae play as ecosystem engineers, it is imperative to detect the factors that could promote resilience in the macroalgae holobiont against environmental stress, in order to keep the benefits they provide to marine biodiversity.

Although bacterial partners have received most of the attention in algae biofilm research, algae biofilms harbor diverse fungal communities. Fungal biofilm composition has been explored in Antarctica. Fungal epibiotic communities in Antarctic macroalgae are influenced by abiotic (i.e., dissolved oxygen and organic matter) and biotic (i.e., antifungal molecules produced by the host) factors (Ogaki et al., 2019). Moreover, macroalgae actively controls these communities, since some fungal strains associated to algae biofilms have agarolytic and carrageenolytic activity, hence, they can degrade algal biomass (Furbino et al., 2017). However, it remains unknown the interactions that might take place between bacteria and fungi inhabiting the macroalgae biofilm. Future studies should have more holistic approaches, where bacteria, fungi and environmental covariables are simultaneously considered.

In particular, the species *Macrocystis pyrifera* is a worldwide (i.e., present both in the Southern and Northern Hemisphere) distributed algae species, whose epibiotic microbial communities have been addressed across several regions in the world (Florez et al., 2019; Lin et al 2018; Weigel et al., 2019). These studies will be valuable to do *M. pyrifera* comparisons across biogeographic regions. However, little is known from the surface biofilm of *Macrocystis pyrifera* in the Strait of Magellan. In the project of the Strait of Magellan microbiome, that we will describe at the end of the paper, we are attempting to characterize the bacterial and functional traits/responses of *M. pyrifera*, in association with environmental factors at different depths. These data could improve our understanding in macroalgae microbiome response to climate change.

## **Invertebrates**

# Sponges

Currently there are 9542 sponge species around the globe, with at least 8,864 species distributed in the Southern Hemisphere (de Voogd et al., 2023; Downey et al., 2012). Sponges are the best studied holobionts in the Southern Hemisphere, both in terms of studied species (65) and number of studies (16), most likely because sponges associated microbes produce a wide array of metabolites that have biotechnological importance (Taylor et al., 2007, Fig. 1, Table 1). The predominant approach to study the sponge holobiont is 16S, nevertheless there have been conducted shotgun metagenomic approaches, which had shed light in sponges' microbiome metabolic potential (Table 1, Yang et al., 2022; Moreno-Pino et al., 2020, 2021). Sampling effort has been performed worldwide, mainly in Antarctic latitudes (tropical, temperate and cold latitudes) (Fig. 3).

Sponges are important ecosystem players that participate in several biogeochemical cycles, and provide stability to the benthos (Bell, 2008). Sponges are the first evolved metazoan, and therefore are the sister group to all animals (Wörheide et al., 2012). Their anatomy is unlike any other metazoan, but generally, it consists in several cell layers (Taylor et al., 2007). Most of the studies used in this revision use the outermost cell layer, nevertheless whenever a study sampled more cell layers, it is explicitly stated in the text. In a recent study in the tropical sponge holobiont conducted in the Northern Hemisphere, it was observed a high metabolic redundancy within the microbiomes that could help buffer the sponge from chemical and physical changes in their environment and from fluctuations in the population sizes of the individual microbial strains that make up the microbiome (Kelly et al., 2022). This is not surprising, since their early evolution occurred in a microbe dominated world in the late Precambrian (Renard et al., 2013). Sponge physiology seems to be microbe dependent and had reached different symbiotic-based solutions to environmental challenges (Thomas et al., 2016).

Current evidence suggests that the sponge microbiome is similar at the phylum level between species in the Southern Hemisphere and those in the Northern Hemisphere (Taylor et al., 2007). In the Southern Hemisphere the sponge microbiome is characterized by several phyla that construct a core including Proteobacteria, Bacteroidetes, Actinobacteria, Verrucomicrobia, Acidobacteria and Cyanobacteria (Rodríguez-Marconi et al., 2015; Matcher et al., 2017; Cárdenas et al., 2018; Savoca et al., 2019; Papale et al., 2020; Happel et al., 2022; Ruocco et al.,

2021; Yang et al., 2022). Interestingly, there is only one archaea phylum associated with sponges, Thaumarchaeota, yet its association is consistent across several species (Brochier-Armanet et al., 2008; Sacristán-Soriano et al., 2020; Moreno-Pino et al., 2020; Steinert et al., 2020).

Interestingly, some bacterial lineages associated to sponges are phylogenetic novelties, that is, their DNA sequences are new, hence, they are unable to be identified with current sequences databases knowledge. Antarctic sponges (Papale et al., 2020; Moreno-Pino 2021; Happel et al., 2022), Australian sponges (Yang et al., 2022), and South African sponges (Matcher et al., 2017) are the ones with more “unknown” bacterial partners. Noteworthy, phylogenetic novel bacteria vary in their prevalence across sponge taxa. Sponges’ genus *Sporosarcina* and *Nesterenkonia* have greater phylogenetic novelty in their microbiotas (50 % or more) while others, like *Cellulophaga algicola* tend to harbor fewer unknown microbes, therefore most of its associated microbes is recognizable (Moreno-Pino et al., 2021). Considering metabolic evidence from Northern Hemisphere tropical sponges, functional redundancy of their microbiomes might allow sponges to tolerate environmental fluctuations, which allows them to distribute over wide areas (Kelly et al., 2022). Hence, environmental variability could be an important driver of microbiota assembly in sponges of the Southern Hemisphere, both at inter- (between different species) and intraspecific (within the same species, among individuals) levels (Steinert et al., 2020).

Nevertheless, besides environmental heterogeneity, phylogenetic factors (i.e., species/genera particular affinities) also play an important role in sponges’ microbiota assembly. For example, sponges from the genus *Mycale* display a strong bacterial core among individuals and species distributed over hundreds of kilometers in the Southern Ocean (Cárdenas et al., 2018; Happel et al., 2022). Similar stable associations have been found in some sponge species from Antarctica (Steinert et al., 2019). *Mycale magellanica*, a common sponge living in the Strait of Magellan, shares up to 74% of sequences belonging to Rhodobacteriaceae and Flavobacteriaceae with individuals of *M. acerata*, a common sponge in the Western Antarctic Peninsula (Cárdenas et al., 2018). Moreover, in *M. acerata*, bacterial composition is stable among individuals distributed across the entire West Antarctic Peninsula (Happel et al., 2022).

Similar trends, with similar microbiota composition, are shown in Demospongiae and Hexactinellida sponges from Ross Sea, Antarctica. These sponges share the bacterial genera

*Erwinia*, *Methylobacterium* and *Sphingomonas* (Papale et al., 2020). In contrast, in the same region (Ross Sea), sponges' microbiotas composition has been found to be heavily influenced by environmental microbes (horizontal transmission), both in the cultivable fraction and NGS-microbiota (Savoca et al., 2019; Sacristán-Soriano et al., 2020). Overall, this highlights the differences in bacterial composition variability among sympatric sponges, which might create species or genus specific microbiota compositions.

Factors underlying microbial recruitment in sponges get more complex when we consider sponges associated Archaea. For example, in Demospongiae and Hexactinellida sponges, from the South Pacific Ocean, bacterial composition is species specific, while the Archaea composition is individual specific (Steinert et al., 2020). Thus, Archaea composition is sparse among individuals, which might suggest that Archaea are opportunistic/contingent players in the sponge holobiont, or rather, their functional benefits are widely shared among several archaea taxa. This pattern contrasts with the stable microbiota composition reported in Ross Sea sponges (Papale et al., 2020). The differentiated trends suggest that environmental fluctuation coupled with species specific filters might drive microbial composition associated with sponges.

The above examples illustrate the complex factors underlying sponge symbiotic associations with bacteria, where horizontal transmission and host specific factors appear to have a differential role among sponge species. Despite inconsistency among associated microbes' identity, it is likely that sponges associated microbes have similar functional traits. Thus, although sponges' microbiota has complex patterns, their microbiomes might have functional convergence (Cristi et al., 2022).

Nevertheless, whether sponges' microbiomes taxonomy obey contingent issues (i.e., which microbe taxa arrived first or neutral process in microbiota assembly) or indeed have a biological basis is an open question. Symbiotic interactions among sponge and its associated bacteria dynamically shape the sponge microbiota composition. On one hand, opportunistic bacteria that degrade sponge tissues, like *Bacillus*, *Micrococcus* and *Vibrio*, are common members of the sponge holobiont. On the other hand, there are antibiotic producing bacteria that regulate the former, like *Streptomyces*, *Aquimarina*, *Pseudovibrio* and *Pseudoalteromonas* (Esteves et al., 2017). Variations in quorum sensing, a microbe chemical communication system, also might play an important role in microbial recruitment. The genera *Pseudomonas*,

*Shewanella* and *Roseobacter*, common bacteria associated to sponges, produce Acyl homoserone lactone, an important chemical messenger in quorum sensing (Mangano et al., 2018). The quorum sensing activity by these bacteria might alter the community profile, by differential microbial recruitment. Quorum sensing might be an important adaptive trait in the sponge holobiont, specially under the ongoing climate derived marine changes, yet its prevalence, and more important its relevance, among sponges' microbiomes has not been thoroughly studied.

Metagenomic studies have shed light on the characteristics of the functional repertoire of sponges' microbiomes. Microbe metabolism differs throughout sponge tissue, highlighting a tissue specific microbiome metabolism (Yang et al., 2022). Several examples pinpoint to nutrient provisioning as an important trait in sponges' microbiomes. Microbial symbionts encode multiple genes related to nitrogen fixation and metabolism of nitrogen compounds, sugars derived from photosynthesis (Moreno-Pino et al., 2020), as well as vitamin B5 (Moreno-Pino et al., 2021). In consonance with antagonistic interactions among members of the sponge microbiome, antibiotic resistance, and biopolymer degradation (Moreno-Pino et al., 2021), as well as CRISPR genes, transposases, detoxification genes and restriction site modifications (Moreno-Pino et al., 2020) are common traits in sponges' microbiomes. The latter functions highlight the evolution of the microbiome within the sponge itself, since several microbes associated with sponges degrade the sponge's tissues and avoid the effect of antimicrobial compounds.

Furthermore, the high prevalence of CRISPR genes in sponges' microbiomes, suggest that their bacterial members are under constant phage attack (Moreno-Pino et al., 2020), which adds another complexity layer to the microbial interactions in the sponge microbiome. There is a high proportion of genes in the microbial communities with unknown functions, so besides phylogenetical novelty described above, there also stands out functional/metabolic microbial novelty associated with sponges (Moreno-Pino et al., 2020).

Up to date, few studies have addressed the effects that marine climate change will have on the sponge holobiont. An interesting exception is given by Kandler et al., (2018), which found that microbial communities of the tropical sponges from New Guinea, *Coelocarteria singaporensis* and *Stylissa* cf. *flabelliformis* might be tolerant to future marine pH conditions. However, it is still an unifactorial experimental approach, that does not represent a reliable test of the multifactorial climate change process. In contrast, there are other climate change driven



effects that might pose a major challenge to sponges in polar environments, like ice scour (seabed modification caused by floating icebergs), which is predicted to increase as a direct consequence of sea surface temperature rise. Ice scour damages benthic communities, specially sponges, like *Isodictya kerguelensis*. Ice scour injuries produce microbial fingerprints, that are easily identified (Rondon et al., 2020). Thus, as climate change progresses and ice scour increases, Antarctica sponges' integrity might be compromised in the next decades.

### **Non-sponge invertebrates**

Marine invertebrates comprise between 35 - 39 recognized phyla (Valentin, JW 2006; Zhang, ZQ 2013). Most iconic marine invertebrates' phyla are Mollusca (118,061 species), Echinodermata (20,550 species), Annelida (17, 426 species), Cnidaria (17,426 species), and Bryozoa (11,474 species) (Zhang, ZQ 2013). Marine invertebrates' microbiomes have received few attention, with 23 species within 10 research articles (Fig. 1, Table 1). However, this group has outstanding examples of authentic holobiont approaches (i.e., coupled measurement of host and microbe traits) (Table 1). Sampling effort has been conducted mainly in Australia, Antarctica, and Africa while there are no studies addressing any marine invertebrate microbiome in South America (Fig. 3).

Overall, marine invertebrate microbiome is characterized by several bacterial phyla, where Proteobacteria, Bacteroidetes, Verrucomicrobia, Tenericutes and Actinobacteria are the most abundant (Webster & Bourne, 2007; Murray et al., 2016, 2020; Unzueta-Martínez et al., 2022) Antarctic corals and snails have stable microbiotas among individuals. The Antarctic soft coral, *Alcyonium antarcticum*, has a core microbiota composed by Proteobacteria, Bacteroidetes, Firmicutes, Actinomycetales, Planctomycetes, Chlorobi and sulfate reductor bacteria (Webster and Bourne, 2007). A similar trend is observed in the Antarctic snail *Synoicum adareanum*. This invertebrate's microbiota is characterized by a high prevalence among individuals of Proteobacteria, Verrucomicrobia, Actinobacteria, Nitrospirae and Bacteroidetes (Murray et al., 2020). In other cases, microbes associated to invertebrates display low diversity, like the ice-adhered anemone *Edwardsiella andrillae*, an endemic anemone of Ross Sea living in the sea ice-water interphase, where the main phyla are Proteobacteria and Tenericutes. Interestingly, most of its sequences display recent diversification branching, which suggests that its associated bacteria are evolutionary recent (phylogenetically new) (Murray et al., 2016).

On the other hand, invertebrates with long and complex life cycles, also have a complex pattern in their associated microbes along its life stages. In the Sydney rock oyster (*Saccostrea glomerata*), bacterial composition is driven by life history characteristics (Unzueta-Martínez et al., 2022). For example, environmental bacteria are a major source of bacterial composition in swimming larvae stages, thus, the microbiota of these stages is characterized by common marine free-living bacteria. On the contrary, sessile stages, like pre adult stage, adult and gametes have distinct microbiota profiles. Overall, the bacterial composition across life stages in the Sydney rock oyster is patchy, which suggests that the majority of oyster associated microbes are either opportunistic or commensals, with few relevance to the oyster. Nevertheless, the genus *Nautella* (Rhodobacterales) was consistently present across stages, increasing its abundance notably in the last stages.

To the best of our knowledge, the only crustaceans in the Southern Hemisphere studied to date are lobsters, krill and copepods. Overall, the bacterial microbiota of crustaceans in the Southern Hemisphere is dominated by Campilobacterota, Tenericutes, Actinobacteria, Firmicutes, Bacteroidetes and Proteobacteria (Clarke et al., 2019; Ooi et al., 2019; Clarke et al., 2021; Oh et al., 2021; Zhang et al., 2021).

Microcrustaceans are important trophic links between primary producers (e.g., diatoms), primary, and secondary predators (e.g., seabirds and fish, respectively). Antarctic krill (*Euphasia superba*) is the trophic basis in the Southern Ocean ecosystem. Despite its inherent exposure to marine bacteria, Antarctic krill hosts unique bacteria phyla in its body; furthermore, its epibiotic associated bacteria differentiates as geographic distance increases. Hence, distance, rather than environmental heterogeneity drives epibiotic bacteria composition in krill (Clarke et al., 2019; Clarke et al., 2021). The major bacterial players in their chitin surface are Campilobacterota and Tenericutes, while in the stomach and intestinal gland were Actinobacteria and Firmicutes (Clarke et al., 2019). Interestingly, *Colwellia* bacteria is a prevalent member of epibiotic microbiota in Antarctic krill swarms at local and regional scales (Clarke et al., 2021). Its persistent association, through thousands of kilometers, might suggest an important role to krill health.

Sea temperature increase might disrupt psychrophilic bacteria associated with Antarctic crustaceans. In the Antarctic copepod *Tigriopus kingsejongensis*, 15 °C temperature treatment has profound effects in its fecal microbiome. Temperature increase diminishes abundance of psychrophilic bacteria (e.g., *Colwellia*), whereas it facilitates the increase in opportunistic pathogens (e.g., *Vibrio*) and virulence genes (Oh et al., 2021). As sea surface temperature increases, it is probable that the microcrustacean holobiont will face major shifts in its associated bacteria, on one hand the increase of bacteria that prefer higher temperatures (e.g., *Vibrio*), and on the other hand, the decrease of psychrophilic bacteria.

Natural life history events, like molting and temperature, have important effects in crustacean's microbiota composition (Ooi et al., 2019; Zhang et al., 2021). Molting is a critical process in crustaceans that allows them growing and sexual maturation. Evidence from the chinese mud crab, *Scylla paramamosain*, suggests that molting represents a bottleneck to most of its associated microbes in gills and midgut. Nevertheless, hemolymph bacteria, *Halomonas* and *Shewanella*, prevail despite molting (Zhang et al., 2021). Noteworthy, the abundance of this bacteria, had a high correlation with the expression of crab antimicrobial gene expression. These results suggest the presence of highly adapted bacteria to the complex life cycle of the mud crabs. Yet, it is uncertain to which degree, molting could alter microbiome traits in the crab holobiont. Sea temperature increase might impose microbe related burdens to crustaceans, as exemplified by the spiny lobster, *Panulirus ornatus*, in Australia (Ooi et al., 2019). In the spiny lobster, temperature increase has a direct relationship with juvenile mortality. As temperature increases, so does bacteria metabolism, which burdens the lobster immune system by bacteria infiltration and subsequent uncontrolled proliferation in the hemolymph.

Interestingly, there have been conducted holobiont approaches in snails in deep hydrothermal vents and several Lucanidae species (Mollusca) across the world, where host DNA and microbial DNA have been simultaneously addressed (Lan et al., 2021; Osvatic et al., 2023). These results have highlighted the effect of ecological niche and host-microbe metabolic complementarity in microbiome assembly. Interestingly, sulfur oxidizing bacteria is present in phylogenetical and geographical distant species of Lucanidae Mollusca (Lan et al., 2021; Osvatic et al., 2023). The above examples highlight the relevance of coupling holobiont approaches with relevant ecological data to address the meaningfulness of the interactions among hosts and its associated microbes.

In the Strait of Magellan study that we will describe at the end of the article, we are including two crustacean species, the centolla, *Lithodes santolla* and the channel sprawns, *Munida gregaria*. The former, is an economically important species that spend most of its life in the sea floor, while the latter is a key species in trophic energy transfer, since it is simultaneously an important plankton consumer and is eaten by several predators (e.g., Magellanic penguin and sea lion). In our holobiont study in the Magallanes region, we expect that the centolla will present fewer signals related to the UV light and heat stress than any of the other species that have a more ample niche in the water column or in the surface as is the case of sea lions and penguins.

## Vertebrates

### Fish

There are more than 20,000 species of marine fish around the globe (CML, visited 2023). Fish are important links between trophic basal levels and higher ones, yet the fish holobiont is the least studied in the Southern Hemisphere with 9 studied species within 4 research articles (Fig. 1). All fish studies have been conducted with 16S metabarcoding approaches (Table 1). Fish microbiome sampling effort in the Southern Hemisphere has been predominantly done template (Australia) and Antarctic latitudes (Antarctica) (Fig 2). Hence, there are important gaps in tropical, temperate and cold latitudes, specifically in South America, Africa, and Indonesia (Fig. 3). Importantly, we still ignore much about native fish species microbiome, since most studies have been done in commercially important species. Most fish microbiome studies have been performed with the objective of testing the usefulness of microbial taxa as biosensors of the fish health. Fish skin has a mucous layer over its epidermis that serves as an additional barrier between the environment and the host's skin. The mucous layer consists of immunogenic compounds that play important roles in innate and adaptive immunity (Gomez et al., 2013). Thus, bacteria inhabiting fish skin, might be commensals in healthy individuals or opportunistic/pathogenic in fish with compromised health.

Current evidence of fish bacterial microbiota in the Southern Hemisphere, has shown that the most prevalent bacteria phyla associated to fish are Actinobacteria, Firmicutes, Proteobacteria, Tenericutes, and Bacteroidetes (Song et al., 2016; Minich et al., 2020; Legrand et

al., 2018; Heindler et al., 2018). In the southern Bluefin Tuna, in Portland Australia, *Thunnus maccoyii*, captivity and antiparasitic administration (i.e., praziquantel) have important effects in fish microbiota composition. Healthy fish microbiota, without praziquantel, is dominated by Mycoplasmataceae in its skin, and *Pseudomonas*, *Acinetobacter*, *Brevundimonas*, and *Delfita* in its gut (Minich et al., 2020). In the Yellowtail Kingfish (*Seriola lalandi*), in temperate and southern waters of Australia, early enteritis produces microbial fingerprints in its skin microbiota. Early enteritis is associated with greater abundance of *Loktanelia*, *Marivita*, and *Planktomarina*, *Simplicispira*, and *Litoricola*, as well as decreased diversity in the microbial community (Legrand et al., 2018).

Fish microbiota also has been addressed under a natural history framework. In four species of wild Antarctic fish (*Trematomus bernacchii* (family Notothenioidei), *Chionodraco hamatus*, (family Channichthyidae), *Gymnodraco acuticeps* (family Bathydraconidae), and *Pagothenia borchgrevinki* (family Notothenioidei), gut microbiota has a stable composition among several species (up to 50% sequences are shared among individuals) (Song et al., 2016). This suggests the presence of a core intestinal microbiome in Antarctic fish, despite differences in environment and diet, which might play important roles in fish health.

Interestingly, fish gut microbiota could serve as a biological prognosis of anthropogenic impact in marine environments, when it is possible to compare historical and contemporary samples. In the Antarctic fish *Trematomus* spp. historical samples (museum samples with 100 years old, fixed with formalin and embedded in paraffin) have revealed notable shifts in gut microbiota composition. Contemporary fish gut microbiota was characterized by Chlamydia, Firmicutes, Cyanobacteria and Mycoplasma. In contrast, historical fish gut microbiota was dominated by Proteobacteria. Despite the richer appearance of phyla in contemporary fish, OTU richness and Shannon index diversities were higher in ancient fish (Heindler et al., 2018). These results attempt to elucidate the relationship between fishing practices and fish gut microbiota, under a historical context. Before global fishing practices, fish were able to have a consistent diet that produce redundant gut communities at the phylum level. In contrast, fishing practices disrupted prey availability, which forced fish to become more opportunistic in their feeding, producing gut communities with wider phylogenetic representation, albeit less diversity (Heindler et al., 2018). Whether the shift from more diverse gut microbiotas enclosed in one phylum, to less

diverse communities with members spanning several phyla has impacts in fish (and any marine host's) health, is a deep open question.

In the Strait of Magallanes project that we describe at the end of the paper, we are studying two fish species, the sardine *Sprattus fuegensis* and the farm salmon, *Salmo salar*. We think that the skin microbiome comparison between a wild native species (i.e., sardine) and an introduced species raised in captivity (i.e., salmon) will provide important clues about local marine conditions as well as the effect of captivity.

## Seabirds

Seabirds are important components of the Southern Hemisphere (Woehler et al., 2001). Currently, there are 350 species around the globe, with at least 61 species endemic to the Southern Hemisphere (Croxall et al., 2012). Seabirds' microbiome has received substantial attention in the Southern Hemisphere, with 22 studied species within 16 research articles (Fig. 1, Table 1). Nevertheless, most of these studies are restricted to few penguin species (Table 1) and in most cases have been done with fecal samples (but see Leclaire et al., 2019). However, this is a global trend, since there are only two studies in the Northern Hemisphere addressing feather bacterial communities, most likely because these are challenging species to sample (Pearce et al., 2017; Leclaire et al., 2019). Seabird microbiome sampling effort in the Southern Hemisphere has been performed in most latitudinal regions, temperate (Brazil & Australia), cold (Argentina and Kerguelen) and Antarctic latitudes (Antarctic islands and Western Antarctic Peninsula (Fig. 3). Seabird microbiome has been predominantly conducted with taxonomic marker approaches (i.e., 16S), hence their associated microbes have been predominantly addressed at the bacterial community level. Nevertheless, these studies have been conducted with metatranscriptomics and RNA viromics surveys (Marcelino et al., 2019; Wille et al., 2020).

Overall, the main bacterial phyla of penguins' fecal microbiota in the Southern Hemisphere are Firmicutes, Proteobacteria, Actinobacteria and Bacteroidetes (Potti et al., 2002; Barbosa et al., 2016; Dewar et al., 2013, 2014, 2017; Yew et al., 2017; Lee et al., 2019; Tian et al., 2021a, Tian et al., 2021b). Whereas the main bacterial phyla of seabird plumage microbiota in the Southern Hemisphere, is characterized by Actinobacteria, Proteobacteria, Firmicutes, and Acidobacteria (Leclaire et al., 2019).

Aging is an important driver of fecal microbiota composition, both in wild and captive penguins (Barbosa et al., 2016; Dewar et al., 2017; Tian et al., 2021a). In wild chinstrap penguin chicks (*Pygoscelis antarctica*) fecal microbiota is dominated by Firmicutes, specially by Clostridiales, *Leuconostoc* and *Fusobacterium*. In contrast, adults' fecal microbiota is dominated by Proteobacteria and Bacteroidetes, specially by Neisserales, Fusobacteriales and Campylobacteriales, yet there is high variability among individuals (Barbosa et al., 2016; Lee et al., 2019). In contrast, in captive chinstrap penguins in the Dalian Sun Asia Aquarium, China, aging only changes relative abundances of the main constituents of fecal microbiotas. Chicks' fecal microbiota was dominated by *Acinetobacter*, while *Pasteurella* was the dominant bacteria in senior penguins (between 22-28 years old); the fecal composition was further completed with *Clostridium*, and *Fusobacterium* in chicks and adult penguins (Tian et al., 2021a). Furthermore, these compositional shifts followed predicted functionality shifts. In general, predicted functionality reaches its maximum diversity in adults, while it starts to decline in senior penguins (after 22 years old) (Tian et al., 2021a).

Support for age of the host as an important driver of fecal microbiota composition comes from the little blue penguin (*Eudyptula minor*), at the Phillip Island Nature Parks, Australia, where fecal microbiota between chicks and adults differs (Dewar et al., 2017). Differences in fecal microbiota composition might be explained by the kind of food that chicks and adults ate. Chicks eat regurgitated food, which might not require a robust microbial metabolic repertoire to aid in digestion; in contrast, adults eat raw food that might contain recalcitrant chemicals, like domoic acid in fish (Lefebvre et al., 2002) or fluoride, from krill (Yoshitomi et al., 2012).

Besides community compositional comparisons, 16S surveys have been used to understand changes in penguins' fecal microbiome predicted metabolic functions. For example, in captive gentoo penguins (*Pygoscelis papua*) in the Dalian Sun Asia Aquarium, China, sex apparently influences fecal microbiome predicted metabolism (Tian et al., 2021b). Males' fecal microbiota is enriched in carbohydrate metabolism, putatively driven by Lachnospiraceae family, whereas females had a fecal microbiota enriched in protein metabolism, putatively driven by Fusobacteriaceae family (Tian et al., 2021b). Microbiome predicted functions from 16S data are constrained by the number of available microbial genomes sequenced (Douglas et al., 2020) and information on its "optimal performance" is limited to human samples, and decreases sharply in environmental samples (Sun et al., 2020). Therefore, functional microbiome studies (i.e.,

metagenomics or metatranscriptomics) coupled with experimental/culture assays are needed to validate the metabolic functions that have been attributed to penguin fecal microbiota.

Fecal microbiota comparisons between several penguin species have shed light about penguin species specific factors influencing fecal microbiota composition. For example, Dewar et al., (2013) compared the fecal microbiota composition of four penguin species: macaroni penguins (*Eudyptes chrysolophus*), king penguins (*Aptenodytes patagonicus*) and gentoo penguins from two sites, Bird Island in South Georgia and Baie du Marine, Possession Island Crozet Archipelago, and little blue penguins from Phillip Island, Australia. This study revealed interesting patterns in penguin fecal microbiota at phyla level, where the dominant bacterial phylum of each penguin species was as follows: Firmicutes in macaroni penguins, Actinobacteria in king penguins, and Proteobacteria in gentoo and little blue penguins (Dewar et al., 2013). Fecal microbiota divergence among penguin species might be explained by trophic niche differences. Additionally, hormone profiles might also have specific effects in gut microbiota composition. However, differences could also be attributed to geographical factors. Systematic studies, where multiple colonies of each species with an adequate sample size are needed to verify to what extent these results reflect penguin fecal microbiota in these species.

Penguin's gastrointestinal tract has several differences along its structure. Given that fecal microbiota represents the last section of the gastrointestinal tract, it is unlikely that it is a representative sample of penguins' gastrointestinal microbial diversity. Indeed, stomach microbiota studies conducted in Adélie (*Pygoscelis adeliae*) and chinstrap penguins in Signy Island, South Orkney Islands, Antarctica have revealed bacterial community structure between the stomach and fecal microbiota. Stomach microbiota of Adélie and chinstrap penguins is characterized at the phyla level by Firmicutes, Proteobacteria, Fusobacteria, and Tenericutes. Common genera in both species are: *Cetobacterium*, *Psychrobacter*, *Chelonobacter*, *Clostridium* (family: Clostridiaceae), *Mycoplasma* and *Ornithobacterium* (Yew et al., 2017). These stomach bacterial communities differed from those reported from the fecal microbiota of these species, where Actinobacteria and Firmicutes were the dominant phyla in Adélie penguin fecal microbiota (Banks et al., 2009), while chinstrap fecal microbiota is characterized by Proteobacteria and Bacteroidetes (Barbosa et al., 2016; Tian et al., 2021a). Stomach bacterial differentiation might be explained in part, by the presence of sphenicins, special compounds in the penguin stomach that prevent bacteria from digesting food (Thouzeau et al., 2003).



758

759 Fasting is a critical event inherent to penguins' life history. Penguins experience fast when  
760 they rear their chicks, and when they molt. Fasting has species-specific compositional changes in  
761 the fecal microbiota of penguins (Dewar et al., 2014; Lee et al., 2019). This could be explained  
762 by penguin species differences in fast length and their fecal associated microbes. In king Penguin,  
763 fasting increases the relative abundance of Proteobacteria, Firmicutes, Actinobacteria,  
764 *Fusobacteria* and *Bacteroidetes*. In contrast, in little blue penguin, all phyla but Proteobacteria  
765 decreased its relative abundance as fasting progressed (Dewar et al., 2014). *Fusobacteria* is a  
766 butyrate producer bacterium. Butyrate is a known anti-inflammatory agent (Canani et al., 2011).  
767 Furthermore, butyrate administration in chickens has immune system functional improvement,  
768 which improves health and decreases pathogen incidence (Panda et al., 2009). The enrichment of  
769 *Fusobacteria* in king penguin fasting, suggest that this bacterium could play an important role in  
770 metabolic homeostasis. Further evidence supporting the effect of fasting in penguins comes from  
771 gentoo and chinstrap penguins in different trophic status (i.e., feeding season or fasting). In  
772 feeding chinstrap and gentoo penguins, *Fusobacteria* and Proteobacteria were the dominant  
773 bacteria, while in molting (fasting) birds, these phyla decreased while Firmicutes increased its  
774 relative abundance. Nevertheless, the magnitude in the compositional changes of its fecal  
775 microbiota composition differed among species. While shifts in chinstrap penguins were subtle  
776 (i.e., not supported statistically), shifts in gentoo penguin were major (i.e., statistically supported)  
777 (Lee et al., 2019).

778

779 It is worth mentioning, that penguin and Antarctic birds' fecal microbiota studies have shed  
780 light in the widespread occurrence of genera close to known pathogens. Those genera include  
781 *Campylobacter*, *Yersinia*, *Salmonella*, and *Escherichia* (Barbosa et al., 2009), yet, it is uncertain  
782 to which degree they affect penguins' health, but they might have a parasitic basis. Support of  
783 this, comes from Magellan penguins from Peninsula Valdez, Argentina where in penguin chicks,  
784 *Corynebacterium*, a dominant bacterium in fecal samples appears to divert resources from the  
785 chick, impairing its growth., but administration of a wide spectrum antibiotic, reduced  
786 *Corynebacterium* abundances, reversing growth halting (Potti et al., 2002).

787

788 On the other hand, the presence of potential pathogens in Antarctic animals can be a  
789 misinterpretation of what "is normal" and what is human related (Souza et al., 1999). The same  
790 may occurs with virus associated with Antarctic fauna. For example, chinstrap, Adélie, and

gentoo penguins harbor a great diversity of virus in their cloaca and of their ectoparasitic mites (Wille et al., 2020). These results highlight the uniqueness fauna living in the Antarctic biome. Moreover, it calls for further microbiome research in these remote places, in order to elucidate their relationship with worldwide fauna.

Seabird plumage microbiota has been poorly studied in the Southern Hemisphere, yet, we have some insights from the blue petrel, *Halobaena caerulea*, whose plumage microbiota composition is highly variable among body sites (Leclaire et al., 2019). Furthermore, some bacteria show a positive correlation with MHC (Major histocompatibility index), which suggests that plumage bacteria are influenced by MHC allele diversity in this seabird.

Interestingly, a metatranscriptomic approach revealed a high incidence of antibiotic resistant genes in several seabirds from Australia with different trophic ecologies (Marcelino et al., 2019). Synanthropic (living near human settlement) species with filter eating habits, like several Australian ducks (*Anas* spp. and *Tadorna tadornoides*), had the highest diversity of antibiotic resistant genes, whereas avocets (*Recurvirostra novaehollandiae*) and gentoo penguins, which live in remote areas and prey invertebrates and fish, respectively, had the fewest. However, the presence of antibiotic resistant genes is not a surprise since this seems to be a very ancient strategy in microbial communities (Souza et al., 1999). Noteworthy, although the cloacal microbiome of gentoo penguins had the less diversity of antibiotic resistant genes, it displayed resistance against unique drugs, like Macrolides, Lincosamide, and Streptogramin (Marcelino et al., 2019).

In a similar fashion, kelp gulls (*Larus dominicanus*) and Magellan penguins from Brazil were assessed by qPCR to evaluate the diversity of antibiotic genes they harbor (Ewbank et al., 2021). Ecological strategies (synanthropic / remote, migratory / non, opportunistic / specialized feeding) might have a strong association with antibiotic gene resistance transmission, with those related to anthropocentric activities having the greatest diversity in antibiotic resistance genes. As expected, kelp gull, a synanthropic species, has the greatest diversity in antibiotic resistance genes. Its antibiotic resistant gene pool has resistance against eight drugs: tetracycline, aminoglycosides, sulfonamides, chloramphenicols, macrolides, quinolones, betalactams, polymyxins. On the contrary, in the Magellan penguin, a migratory, non-synanthropic, specialized feeder (it preys mainly fish and squid) their antibiotic resistance genes diversity was

lower, with specific resistance against two drugs, tetracycline and quinolone (Ewbank et al., 2021).

In our project on the surface microbiome of key species in the Strait of Magellan, we are planning to study the feather microbiome in Magellan and King penguins. We are interested in testing geographical, phenological, and developmental effects in feather microbiome of these species. These works will provide original baseline knowledge regarding penguin feather associated bacteria.

### **Marine mammals**

Marine mammals comprise several species that collectively are considered as “marine sentinels”, since their population trends could give valuable insights about the status of the marine ecosystem (Moore, SE, 2008). Currently, there are 115 species distributed around the globe (Kaschner et al., 2011). Up to one third of them are cooccur between the template – cold latitudes (i.e., 20-50) of the Southern Hemisphere. Important hotspots of marine mammals’ biodiversity: New Zealand, Sub-Antarctic and Southeastern Pacific islands, and offshore waters along the coasts of southern South America (Kaschner et al., 2011). Marine mammals’ microbiome has received the greatest attention in the Southern Hemisphere, with 15 studied species within 17 research articles (Fig. 1, Table 1). Nevertheless, most studies have been conducted in one species (humpback whale) (Table 1). In most cases, these studies have been conducted with 16S approaches, but there is a pinniped shotgun metagenome (Table 1, Smith et al., 2013). Interestingly, marine mammals’ microbiome research has been done worldwide, with sampling effort across all latitude regimes (tropical, template, cold and Antarctic), specifically in template latitudes (i.e., Australia), followed by Antarctica and South America (Fig. 3).

We still lack a more comprehensive view of cetaceans’ microbiome in the Southern Hemisphere, since most of current studies have been done in one species over different geographic locations (i.e., humpback whale) (Table 1). In contrast with cetacean microbiome knowledge in the Northern Hemisphere where several species have been addressed (Sanders et al., 2015; Van Cise et al., 2020; Apprill et al., 2020; Miller et al., 2020). While this gives us depth in understanding humpback whale microbiome across geographic regions, it also pinpoints to the

great void that remains in the rest of cetaceans, like whales and dolphins. Besides humpback whale, there have been conducted several studies in pinnipeds (Table 1).

In particular, marine mammals' microbiome research has been focused in whale skin and blow microbiota studies. In the Southern Hemisphere skin microbiological surveys have been done in rorquals (Balaenopteridae, species *Megaptera novaeangliae* [humpback whale], *Balaenoptera musculus*, and *B. physalus*) as well as killer whales (*Orcinus orca*). Overall, the main bacterial elements in the whale skin microbiota are Proteobacteria, Bacteroidetes, Actinobacteria, and Firmicutes (Apprill et al., 2014; Pirotta et al., 2017; Bierlich et al., 2018; Hooper et al., 2019; Vendl et al., 2019; Vendl et al., 2020; Toro et al., 2021).

Humpback whales are the best studied cetaceans in the Southern Hemisphere. Studies from humpback whales in the South Pacific (Samoa islands), Chilean coasts and Antarctic regions have allowed the detection of a skin core microbiota, characterized by *Tenacibaculum* and *Psychrobacter* (Apprill et al., 2014; Bierlich et al., 2018; Toro et al., 2021). Rorquals (*Megaptera novaeangliae*, *Balaenoptera musculus*, and *B. physalus*) skin microbiota along several points in Chilean coasts have idiosyncratic and species-specific trends, that is, whales have a unique skin microbial composition at the individual and the interspecific level. However, their skin microbiota alpha diversity was similar in compositional terms (i.e., Shannon diversity), but was slightly different in phylogenetic terms (i.e. Faith phylogenetic diversity) (Toro et al., 2021).

Skin microbiota of humpback whales foraging in Antarctica and the Strait of Magellan are enriched in *Psychrobacter* bacteria. Changes in sea surface temperature, as well as shifts towards northern areas, are associated with decreases in *Psychrobacter* relative abundance. This pattern suggests that sea temperature is an important driver in humpback whale skin microbiota assembly (Bierlich et al., 2018; Toro et al., 2021).

Blow microbiota of Australian humpback whales has been surveyed to address whales' health. In contrast with skin microbiota, blow microbiota is sparse among individuals, without any discernable core. The most abundant microbes in the blow were *Tenacibaculum*, *Pseudomonas* *Leptotrichia* and *Corynebacteria*. Additionally, some individuals had potential respiratory pathogens in their blow microbiota, like *Balneatrix*, *Clostridia*, *Bacilli*,

*Staphylococcus* and *Streptococcus* (Pirota et al., 2017; Vendl et al., 2019). Furthermore, whale blow associated bacteria harbors significant phylogenetic novelty, since half sequences in some individuals were only able to identify at class level (Vendl et al., 2019). Whale feeding phenology is an important factor that could underlie the sparsity of blow microbiota structure. While feeding blow microbiota has a core microbiota composed of *Arcobacter*, *Corynebacterium*, *Enhydrobacter*, *Helcococcus* and *Tenacibaculum*, albeit in very low abundance, 1.5% or less. In contrast, during migration, when whales are fasting, blow microbiota among individuals becomes highly variable, with no discernible core (Vendl et al., 2020a). Besides phenology or health status, whale blow microbiota is also influenced by the sociality degree of the studied species. Whale species with gregarious habits (e.g., humpback whale), have higher diversity and a great microbial core in their blow microbiota, in contrast with more solitary whales (Vendl et al., 2020b). Likely, this could reflect the horizontal transmission of blow microbes among contiguous whales, where the microbes exhaled by one individual are inhaled by another, and so on. Nevertheless, more studies are needed to address if this pattern emerges because of horizontal transmission or could reflect common health status among whales' groups.

In the case of Killer whales, the Antarctic ecotypes have been found to harbor a distinct skin microbiota from ecotypes in the northern hemisphere. Differences are driven by *Tenacibaculum dicentrarchi* bacteria, diatoms, and several algae-associated bacteria (Hooper et al., 2019). More systematic studies of whales associated bacteria, and ideally of an “authentic” holobiont approaches (i.e., describing whale genetic traits, as well as their microbiome and transcriptome) to better understand the whale holobiont.

In our project on the surface microbiome of key species in the Strait of Magellan (ANID R20F0009), we are including a holobiont approach to study the humpback whales of the area. In particular, we are sampling individuals that migrate to the Strait of Magellan to feed in the austral summer season.

Pinnipeds are apex predators whose health might inform about marine ecosystem conditions (Moore, SE 2008). Sampling pinnipeds is challenging due to the remote location of their colonies and proclivity to escape from humans, hence, the most feasible samples to study them, are the feces they leave in rocks. Seal fecal microbiota apparently is driven by several host factors,

including feeding, geographic distribution, ontogeny, trophic niche differences, anatomy, and physiology, explaining, at least in part, the great fecal microbiota composition variability among species, where there is no detectable fecal core microbiota. Nevertheless, it is uncertain if the taxonomic differences are congruent with microbiome metabolic traits.

Overall, the main bacterial components of seal fecal microbiota are Firmicutes, Fusobacteria, Bacteroidetes, Proteobacteria, and Actinobacteria (Nelson et al., 2013; Delpont et al., 2016; Grosser et al., 2019; Kim et al., 2020; Toro-Valdivieso et al., 2021). Nevertheless, fecal microbiota from pinnipeds has been a challenging task as there seems to be a high proportion of phylogenetic novelty that interferes with taxonomic identification (i.e., at the family or genus level) and proper analysis of the divergence among communities (Toro-Valdivieso et al., 2021). For example, across pinniped species, fecal microbiota composition at phylum level seems to be identical (Nelson et al., 2013; Kim et al., 2020) but diverges at lower taxonomic levels. For instance, in the southern elephant (*Mirounga leonina*) and Weddell seals (*Leptonychotes weddelli*) at the phylum level, fecal microbiota is dominated by Firmicutes, yet at the family level Ruminococcaceae and Acidaminococcaceae, respectively, drive differences in the fecal microbiota composition (Kim et al., 2020). Life history characteristics create complex patterns that influence fecal microbiota, as shown with the fecal microbiota of southern elephant seals and leopard seals (*Hydrurga leptonyx*). In these species, fecal microbiota is shaped by the simultaneous effect of species, age and sex, creating complex fecal microbiota patterns with no discernable trend (Nelson et al., 2013). In the Australian fur seal (*Arctocephalus pusillus*) aging from pups to adults produces a successional pattern in fecal microbiota composition. Adult fecal microbiota had unique bacterial taxa, dominated by *Clostridium*, *Lactobacillus* and *Enterococcus*. The diet shift from milk with high fat-protein in pups to a marine raw diet in adults, is thought to underlie the fecal microbiota diversification (Smith et al., 2013).

A fecal microbiota survey in the Australian sea lion (*Neophoca cinerea*) showed a core fecal microbiota at family level, composed of Clostridiaceae bacteria. This core arises only in wild seals, which suggests that natural diet might represent a cohesive driver in fecal microbiota composition. Instead, captive animals lacked Clostridiaceae bacteria in their fecal microbiota. Moreover, seals from wild colonies had more fecal microbial diversity than captive colonies, especially those with high densities (Delpont et al., 2016). At the functional level, the Australian sea lion fecal microbiome is enriched in carbohydrate metabolism, nitrogen biosynthetic and

nutrient transport pathways, as well as virulence genes. The fecal microbiome composition appears to play an important role in fat – nutrient storage, key issues in marine mammals' survival in polar ecosystems (Lavery et al., 2012). This could pinpoint to a fat storage mechanism driven, or at least with high influence, of gut microbes. Nevertheless, a higher sample and more phylogenetic inclusivity (i.e., more seal and sea lion's species) is needed to validate the prevalence of this microbiome traits among populations and pinniped species. Interestingly, Clostridiaceae bacteria has a high prevalence among Australian seal species fecal microbiota (Lavery et al., 2012; Smith et al., 2013; Delport et al., 2016), which might suggest an important role for these bacteria in seals digestion. Nevertheless, it is a cautionary interpretation since gut microbiota composition varies along the gastrointestinal tract. Hence, fecal microbiota reflects only a portion of the gut microbiome.

In the case of skin microbiota, there is one example from the Antarctic fur seal (*Arctocephalus gazella*) (Grosser et al., 2019). In this fur seal, colony density drives skin microbiota structure, rather than genetic similarity. High density colony had fewer alpha diversity in its skin microbiota. This suggests that stress associated with overcrowding could scale to skin microbiota composition, diminishing the richness and abundance of bacterial taxa. Nevertheless, overcrowding, also might facilitate horizontal transmission since fur seals are in close contact. This could allow fast transmission of opportunistic microbes, capable of dominating the community. Most likely, the combination of both explanations might aid in illuminating the pattern; stress produced by overcrowding, coupled with increased transmission of fast reproducing microbes, shape the skin microbiota of colonies with high density. (Grosser et al., 2019).

In our Strait of Magellan project (ANID R20F0009), we are studying the skin surface microbiome of colonies of the South American sea lion (*Otaria byronia*), a keystone species in the sub-Antarctic ecosystem.

## Discussion and perspectives

### Microbiomes in the southern ecosystems and climate change context.

In the southern ecosystems, including Antarctica, it has been predicted that higher temperature will increase coastal ice-free areas, sea-ice loss, glacial retreat, ocean acidification and ocean

warming (Morley et al., 2020), affecting marine biota in all trophic level. Lower trophic levels are expected to move south, depending on their tolerance of warming ocean condition and productivity, meanwhile, ocean acidification will impact mainly over crustaceans and calcifying organisms. Marine animals, mammals, and seabirds are expected to move to alternative locations for food and breeding, for survivor and adaptation (Constable et al., 2014). However, the impact of climate change over organisms and their associated microbiota as a whole—i.e., the holobiont – has been poorly studied across the metazoan spectrum, with notable exceptions in some invertebrates and macroalgae (Table 1). Yet, we still lack ecosystem holistic approaches (i.e. including abiotic variables or ideally holobiont experimental assays where important abiotic parameters, like temperature and pH are controlled), that could give us insight about marine holobiont adaptability potential to the environmental changes that soon will face as a consequence of anthropogenic climate change.

We think that the relation between the holobiont and their ecosystems is bidirectional, where the environmental changes. will affect the host and their associated microbes, meanwhile changes in the host like abundance, behavior, feeding, molting and reproduction; and their microbiome like diversity, taxonomic composition, and nutrient recycling, will impact over the environment. For example, soils impacted by penguins and pinnipeds presented high amount of nutrients such carbon, nitrogen and phosphorous (Ugolini, FC 1972; Tatur et al., 1990). Also, marine animals' impact over greenhouse gases emissions, penguins and pinniped settlement had been founded a hotspot of CO<sub>2</sub>, methane (CH<sub>4</sub>) and nitrous oxide (N<sub>2</sub>O) emissions (Zhu et al., 2008; 2009). In these processes animals had a huge impact over the coastal sediments where they colonized through their feces, eggs, prey, carcasses, among others (Guo et al., 2018; Almela et al., 2022; Ramírez-Fernández et al., 2019). Feces can directly impact over soil microbiome, seeding gut microbes from marine animals, and indirectly because the high amount of nutrients that they transport from marine to terrestrial ecosystems (Guo et al., 2018). At functional level, marine animals increased soil microbial communities related with denitrification pathways (Ramírez-Fernández et al., 2021), and other nitrogen pathways involved in N<sub>2</sub>O emissions.

Climate change studies should take in account the impact of environmental changes in the host and their microbiome. In our project on the surface microbiome of key species in the Strait of Magellan, we will measure environmental factors such as water temperature, UV radiation, chlorophyll content, oxygen levels, pH, salinity and nutrient content to correlates holobiont



abundance, diversity, and distribution, focusing on the skin microbiome due their direct contact with the environmental parameters. The equilibrium between the holobiont and the ecosystems will allow their protection and conservation (Carthey et al., 2020).

# **Surface microbiome of key species in the Strait of Magellan, an integrative holobiont project**

To increase the number of microbial biodiversity studies in the southern hemisphere, emerges our microbiome project of key species in the Strait of Magellan in Chile (Fig. 5). Our project will generate new baseline data for almost all the species that will be sampled (except for humpback whale skin, which has been extensively studied). Hence our project, has two stages, first baseline data generation for all considered species. Second, microbiome functional data (i.e., metagenomics and metatranscriptomics) as well as ecosystem data integration, where host microbiome data from multiple years along environmental variables will be integrated. We think this approach will give valuable insights about host-microbiome responses to ongoing Anthropocene derived climate change. Moreover, it will generate host microbiome data framed under an ecosystemic approach, which will be valuable for further comparisons in next years, where environmental variables likely would have changed.

We consider that microbes are an excellent biodiversity study target, since they can show fast evolutionary responses to environmental alterations, and they have enormous metabolic and genetic diversity. We are focusing on surface microbes to test their value as biosensors of climate change effects in key hosts in the Strait of Magellan. Our project attempts to encompass trophic-level inclusivity, through sampling hosts in different (yet related) trophic levels, and an introduced species (i.e., farm salmon). We will sample a primary producer (i.e., *Macrocystis pyrifera*, huiro/kelp), primary consumers (i.e., two crustacean species *Lithodes santolla*, centolla and *Munida gregaria*, channel prawns), secondary predators (i.e. *Sprattus fuegensis*, Fuegian sprat; *Spheniscus magellanicus*, Magellan penguin, *Aptenodytes patagonicus*, king penguin and *Megaptera novaeangliae*, humpback whale) and an apex predator (i.e. *Otaria byronia*, South American sea lion).

Our project will generate original baseline knowledge of bacterial communities associated to the surface of some taxa (like penguins' feathers, sea lion fur, fish scales, or crustacea shell).

Furthermore, as we plan to perform multiannual samplings, the project will bring the opportunity to elucidate if there are microbial signatures (both at the community and genomic level) associated to seasonal variation and/or to environmental variables. Our main objective is to test if there is a core surface microbiome among marine hosts sharing the same environment (i.e., in particular in Coastal Marine Protected Area “Francisco Coloane”). Alternatively, we are interested in testing the existence of a core microbiome at different levels. For example, at host complexity (i.e., a core microbiome for invertebrates, another for mammals, etc.); trophic level (i.e., a core microbiome for primary consumers, another for primary predators, etc.); or alternatively a species-specific surface microbiome.

In general, the marine holobiont in the Southern Hemisphere is characterized by few systematic and authentic holobiont studies (i.e., studies that analyze at the same time host traits [e.g., genomics, transcriptomics] and microbiome traits [e.g., 16S, metagenome], see Table 1 for holobiont studies examples). Besides there is a great bias in sampled hosts. For instance, sponges have been thoroughly sampled, but there is a great gap in marine invertebrate and vertebrate hosts. We highlight the case of marine invertebrates, which although have a relatively high number of studied species (23), it remains a tiny fraction of its huge diversity.

Holobiont studies are dominated by microbiota approaches, where the focus is centered in taxonomic patterns across ecological/life story conditions. These studies have highlighted the prevalence of Proteobacteria among a wide spectrum of hosts, while specific host-phyla associations complete the bacterial community. Noteworthy, microbiota approaches have been centered in the bacterial fraction, completely ignoring the potential role of fungi, virus and archaea in the marine host holobiont. Notable exceptions where these groups have been addressed are macroalgae (fungal communities), penguins (viromics) and sponges (archaea). Nevertheless, there is no study that addresses the interactions among all these groups simultaneously, likely because technical and economic challenges (i.e., computational resources, computational skills).

Microbiota studies are important and economically feasible explorations, yet detailed microbiome studies including metagenomics and metatranscriptomics studies are needed to inclusively address the microbe community (fungi, eukaryotes, virus, bacteria, and archaea) as well as its functional potential along with its host. Moreover, functional holobiont data (i.e., host

genomics & metagenomics/metatranscriptomics) coupled with environmental data, might provide valuable insights about the influence of ecosystemic status in host associated microbiome, which in turn could be tested for the plausibility of the microbiome to reflect host stress.

Microbiome studies might help in elucidating whether patchy/heterogenous distribution in microbe taxonomic profiles has different functional potential, or rather, if distinct taxonomic profiles have convergent/redundant functional profiles. Additionally, microbiome studies might help to elucidate if there is a microbiome functional profile associated with eukaryote hosts, or even among distinct hosts if there are core functions among their microbiomes. Finally, sampling must span as many individuals as possible, as well as geographic and seasonal (longitudinal/annual) representability to address whether patchy distributions among associated microbes are a natural feature of marine holobionts or a consequence of low sampling. Such a systems biology approach might bring further understanding in the complex interplay among microbes and its hosts, as well as the impact it might carry for them. We think that our project on the surface microbiome of key species in the Strait of Magellan will provide valuable information on the points mentioned above, which will contribute to the knowledge of microbial diversity in the region, as well as their current responses under Anthropocene-derived climate change.

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**Table 1** (on next page)

Table 1. Summary of marine host microbiome research sampling effort in the Southern hemisphere

1 Table 1. Summary of marine host microbiome research sampling effort in the Southern hemisphere

2

| Macroalgae                |   |                                |  |   |
|---------------------------|---|--------------------------------|--|---|
| Number of species sampled | Species repetitively studied (number of studies)  | Predominant molecular approach | Functional microbiome/holobiont approaches   | Regions repetitively sampled (number of studies)                  |
| 24                        | <i>Ecklonia radiata</i> (4)   | 16S                            | DNA holobiont (Wood et al., 2022)<br>Shotgun metagenomics (Song et al., 2018)                          | Antarctica (3)<br>Australia (5)                                   |
| Sponges                   |   |                                |  |   |
| 65                        | <i>Mycale acerata</i> (4)   | 16s                            | Shotgun metagenomics (Moreno-Pino et al., 2020, 2021, Yang et al., 2022)                               | Antarctica (13)   |
| Marine invertebrates      |   |                                |  |   |
| 23                        | <i>Euphasia superba</i> (2)   | 16S                            | Shotgun metagenomics (Ooh et al., 2022)<br>DNA & RNA holobiont (Lan et al., 2021; Osvatic et al. 2023) | Antarctica (4)<br>Australia (3)                                   |
| Fish                      |   |                                |  |   |
| 9                         | NA  | 16S                            | NA   | Australia (2)   |
| Seabirds                  |   |                                |  |   |
| 22                        | <i>Aptenodytes patagonicus</i> (2)<br><i>Eudyptula minor</i> (2)<br><i>Pygoscelis adeliae</i> (3)<br><i>Pygoscelis antarcticus</i> (4)<br><i>Pygoscelis papua</i> (4)<br><i>Spheniscus magellanicus</i> (2) | 16S                            | Metatranscriptomics (Marcelino et al., 2019)   | Antarctica (5)<br>Australia (4)<br>Bird Island, South Georgia (2) |
| Marine mammals            |   |                                |  |   |

|    |                                  |     |                       |                |
|----|----------------------------------|-----|-----------------------|----------------|
| 15 | <i>Megaptera novaeanglia</i> (6) | 16S | Shotgun metagenomics  | Antarctica (4) |
|    | <i>Balaenoptera musculus</i> (2) |     | (Lavery et al., 2012) | Australia (6)  |
|    | <i>Neophoca cinerea</i> (2)      |     |                       | Chile (2)      |
|    | <i>Mirounga leonine</i> (2)      |     |                       |                |

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# Figure 1

## Marine microbiome studies in the Southern Hemisphere

Figure 1. Total marine microbiome studies in the Southern Hemisphere reported in this review (75). The most studied groups are marine mammals (17 studies), followed by sponges (16), seabirds (15), macroalgae (13), invertebrates (10), and fish (4).

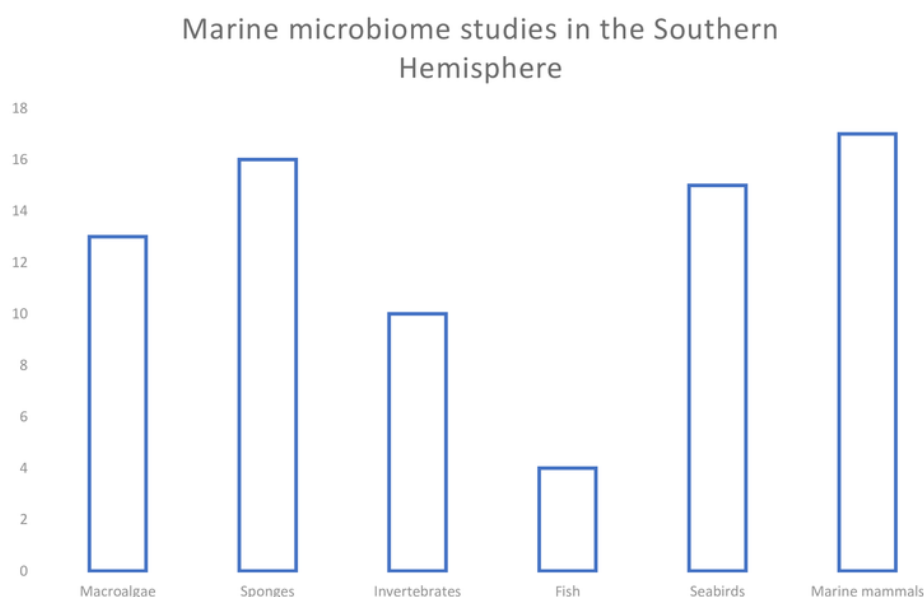


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# Figure 2

Latitudinal distribution of microbiome studies in the Southern Hemisphere

Figure 2. Southern Hemisphere map latitudinal distribution of host microbial communities sampling (A) Worldwide Southern Hemisphere. Insets with highly sampled regions. (B) Western Antarctic Peninsula. (C) Australia.

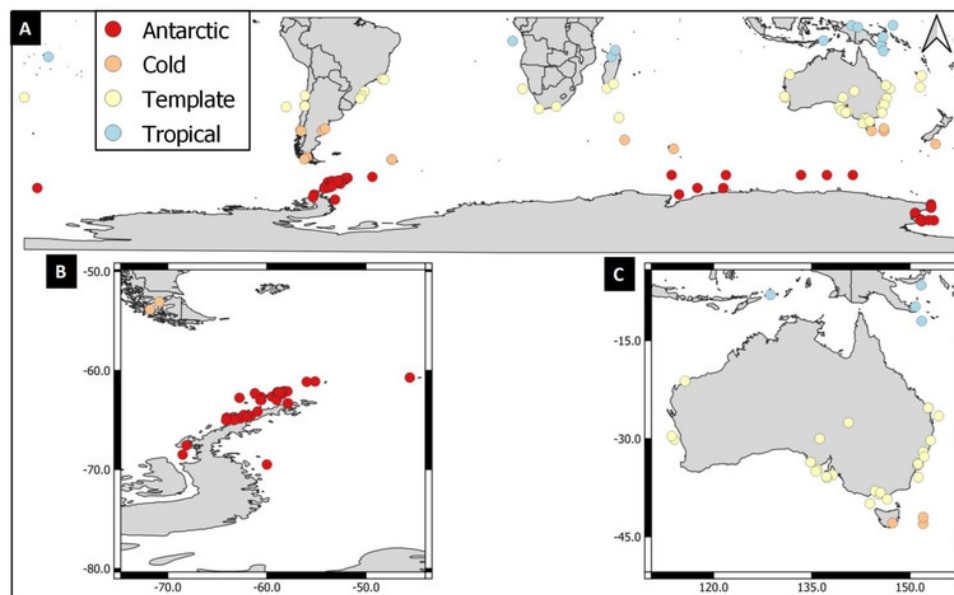


Figure 2. Southern Hemisphere map latitudinal distribution of host microbial communities sampling, A) worldwide Southern Hemisphere. Insets with highly sampled regions, B) Western Antarctic Peninsula, C) Australia.



# Figure 3

Host microbiome sampling distribution in the Southern Hemisphere

Figure 3. Southern Hemisphere map of studied hosts distribution. (A) worldwide Southern Hemisphere. Insets with highly sampled regions. (B) Western Antarctic Peninsula. (C) Australia.

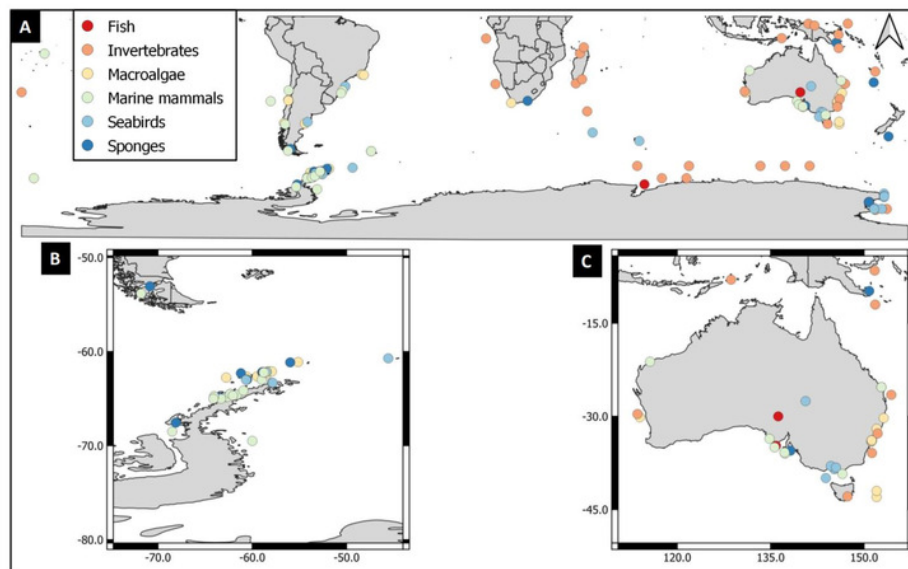


Figure 3. Southern Hemisphere map of studied hosts distribution, A) worldwide Southern Hemisphere. Insets with highly sampled regions, B) Western Antarctic Peninsula, C) Australia.

# Figure 4

Southern Hemisphere distribution of molecular approaches to study marine host microbiomes

Figure 4. Southern Hemisphere map of molecular approaches used to study host microbial communities. (A) worldwide Southern Hemisphere. Insets with highly sampled regions. (B) Western Antarctic Peninsula. (C) Australia.

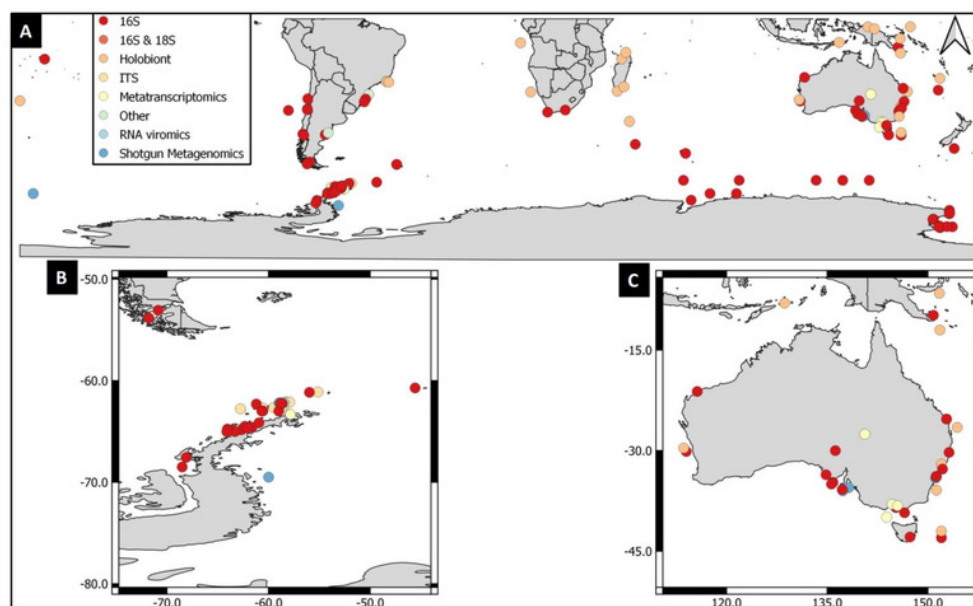


Figure 4. Figure 3. Southern Hemisphere map of molecular approaches used to study host microbial communities, A) worldwide Southern Hemisphere. Insets with highly sampled regions, B) Western Antarctic Peninsula, C) Australia.

# Figure 5(on next page)

Summary of the Strait of Magellan surface microbiome of key taxa project

Figure 5. Target species and main areas of field sampling of the microbiome project. (A) Target species of the microbiome project are (from top left to right): kelp / huiro (*Macrocystis pyrifera*), Fuegian sprat (*Sprattus fuegensis*), channel prawns (*Munida gregaria*), centolla (*Lithodes santolla*), Magellanic penguin (*Spheniscus magellanicus*), King penguin (*Aptenodytes patagonicus*), Humpback whale (*Megaptera novaeangliae*), South american sea lion (*Otaria byronia*), Atlantic salmon (*Salmo salar*). (B) Main areas across the Strait of Magellan that field work is taking place. The lower right side of Figures 5 (A-B) shows photographs of Carlos III Island, Contramaestre Island, King Penguin Reserve and Tuckers Islands.

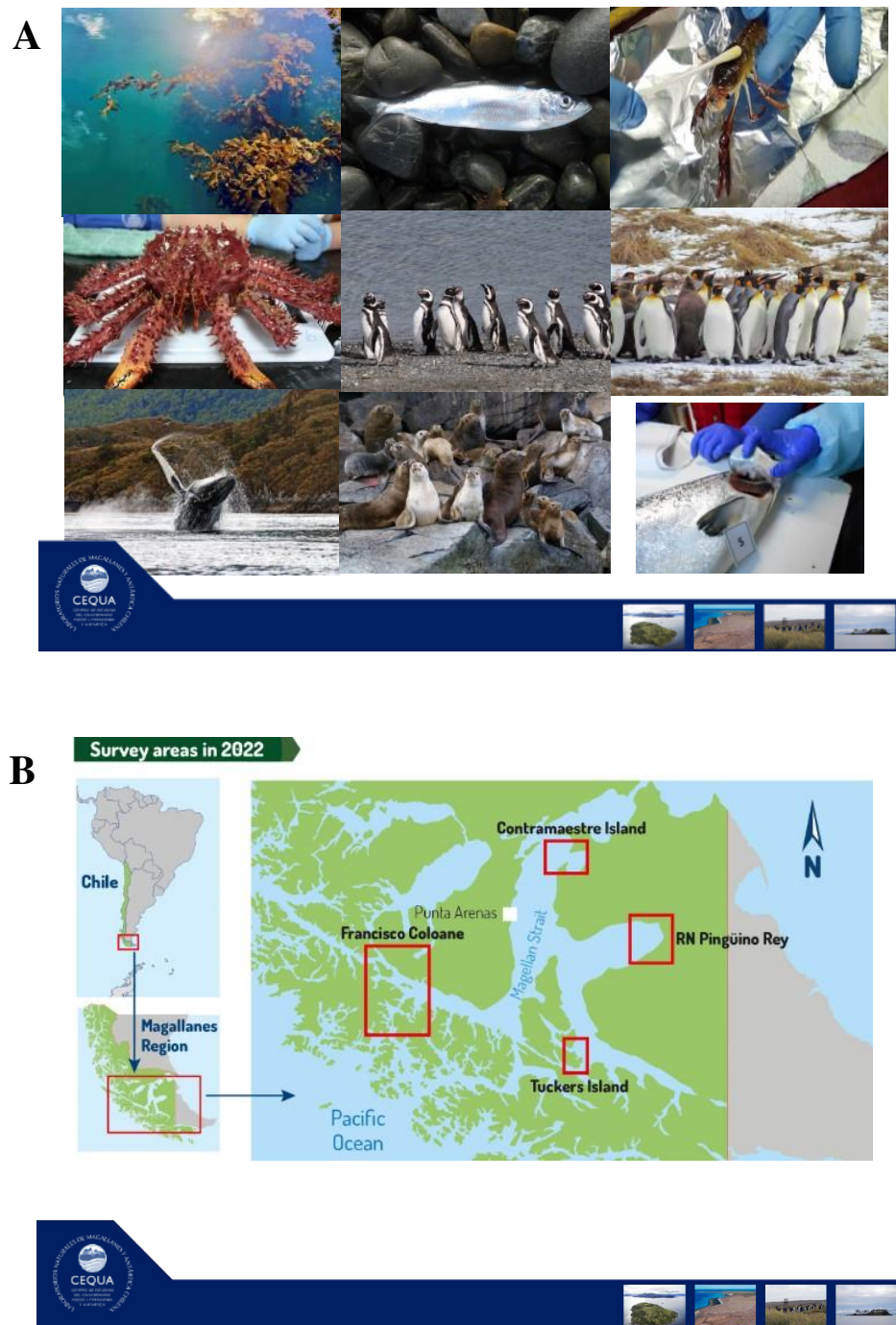


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