

Effects of drought and salt stress on seed germination and seedling growth of *Elymus nutans*

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Drought and soil salinization are global environmental issues, and *E. nutans* play an important role in vegetation restoration in arid and saline environments due to their excellent stress resistance. In the process of vegetation restoration, the stage from germination to seedling growth of forage is crucial. This experiment studied the effects of PEG-6000 simulated drought stress and NaCl simulated salinization stress on the germination of *E. nutans* seeds, and explored the growth of forage seedlings from sowing to 28 days under drought and salinization stress conditions. The results showed that although under the same environmental water potential, there were significant differences in the responses of drought stress and salinization stress to *E. nutans* seed germination, seedling growth, organic carbon, total nitrogen, and total phosphorus in the aboveground and underground parts. Using the membership function method to comprehensively evaluate the seed germination and seedling indicators of *E. nutans*, it was found that under the same environmental water potential, *E. nutans* was more severely affected by drought stress during both the seed germination and seedling growth stages. *E. nutans* showed better salt tolerance than drought resistance.

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2 *Elymus nutans*

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8 **Abstract:** Drought and soil salinization are global environmental issues, and *E. nutans* play an important role in
9 vegetation restoration in arid and saline environments due to their excellent stress resistance. In the process of
10 vegetation restoration, the stage from germination to seedling growth of forage is crucial. This experiment
11 studied the effects of PEG-6000 simulated drought stress and NaCl simulated salinization stress on the
12 germination of *E. nutans* seeds, and explored the growth of forage seedlings from sowing to 28 days under
13 drought and salinization stress conditions. The results showed that although under the same environmental water
14 potential, there were significant differences in the responses of drought stress and salinization stress to *E. nutans*
15 seed germination, seedling growth, organic carbon, total nitrogen, and total phosphorus in the aboveground and
16 underground parts. Using the membership function method to comprehensively evaluate the seed germination
17 and seedling indicators of *E.nutans*, it was found that under the same environmental water potential, *E.nutans*
18 was more severely affected by drought stress during both the seed germination and seedling growth stages.
19 *E.nutans* showed better salt tolerance than drought resistance.

20 1. Introduction

21 At present, soil salinization caused by drought, unreasonable irrigation and other factors is a global
22 environmental problem that needs to be solved urgently, and soil drought and salinization are the main non Biotic
23 stress factors affecting crop growth (Dai ,2013; Pokhrel et al., 2021). The arid zone in China accounts for 1/3 of
24 the total area, including a large amount of saline alkali soil. The important factors affecting agricultural
25 development and ecological environment protection in arid zone are drought, soil salinization, and secondary
26 salinization caused by irrigation. (Zhang et al., 2021; Zhao et al., 2022). The unique and complex and diverse
27 climate of Tibet, with high altitude, low temperature, large diurnal temperature difference; low precipitation and
28 large regional differences; strong sunlight and long daylight hours, is the main reason for its susceptibility to
29 drought and soil salinization, and the annual crop yield reduction caused by drought and water shortage or soil
30 salinization limits the development of agriculture in the region (Zhang et al. ,2021; Javed et al. ,2021; Kong et
31 al. ,2018).

32 *Elymus nutans* is a perennial herb belonging to the *Elymus* spp. in the family Gramineae, distributed in
33 Tibet, Hebei, Qinghai, Sichuan, Shaanxi, Gansu, Xinjiang, and Inner Mongolia in China, and also in Turkey,
34 Mongolia, India, and Russia (Liu et al. , 2021; Ma et al. , 2006). *E. nutans* has strong growth ability, high crude
35 protein and fat content, and good palatability. And it plays an important role in the improvement of alpine

36 grasslands and the construction of artificial grasslands. (An et al. , 2022; Sun et al. , 2022).

37 Through long-term natural selection, *E. nutans* has gradually developed resistance to cold, drought, salinity
38 and other high quality genetic characteristics, which not only have high forage value, but also can play an
39 important role in resisting wind and sand damage and maintaining water and soil (Li et al. ,2021;Luo et al.
40 ,2019).

41 Soil drought and salinization, as two major non Biotic stress factors affecting plant growth and crop yield,
42 both belong to environmental water potential stress (osmotic stress) on plants in a certain sense (Do et al. , 2022;
43 Bashir et al. , 2019). Seed germination is the beginning of plant life history and the most sensitive period to the
44 environment (especially in arid and saline environments), while in some arid and semi-arid areas seed
45 germination is subjected to both drought and salt stress (Wijewardana et al. ,2019; Silva et al. ,2018). The
46 successful establishment of vegetation communities is directly determined by the success of seedling formation
47 after seed germination, and seedlings also respond to environmental stress during their growth process (such as
48 adjusting plant height, changing root structure, etc.) (Sun et al. , 2019; Andivia et al. , 2021). Current research
49 has shown(Wang et al. , 2021) that both drought stress and salt stress can affect the germination rate of *E.nutans*
50 seeds, with higher concentrations of PEG-6000 and NaCl directly inhibiting the germination of *E.nutans* seeds.
51 Moreover, PEG-6000 and NaCl stress led to the increase of proline, malondialdehyde and soluble protein
52 content, and the increase of superoxide dismutase, peroxidase and catalase activities inhibited the growth of *E.*
53 *nutans* seedlings(Wang et al. ,2017;Song et al. , 2022). In order to study the response of *E. nutans* seed
54 germination and seedling formation to drought and salt stress, we conducted a pot experiment in the artificial
55 climate incubator of the Grassland Laboratory of Tibet Agricultural and Animal Husbandry College in Linzhi,
56 Tibet. We studied the effects of drought stress and salt stress on the germination and seedling growth of *E. nutans*
57 seeds under the same osmotic potential, and measured the germination rate, germination potential, root length,
58 bud length, and germination index of *E. nutans* seeds; Seedling growth indicators and the content of organic
59 carbon, total nitrogen, and total phosphorus in the aboveground and underground parts. Explored the similarities
60 and differences in the response of *E. nutans* to drought stress and salinization stress, comprehensively evaluated
61 the degree of drought stress and salt stress suffered by *E. nutans* seeds and seedlings, and clarified the tolerance
62 of *E. nutans* seeds and seedlings to drought and salt stress under the same environmental water potential, in order
63 to provide reference for vegetation restoration in arid and salinized areas.

64 **2.Materials and Methods**

65 **2.1 Material**

66 The experiment was completed in the Grass Science Laboratory of Tibet Agricultural and Animal
67 Husbandry College, Tibet Autonomous Region (29°39'57. 5" N ,94°20'31. 4" E). The test seeds is *Elymus*
68 *nutans*. cv. Baqing, provided by the Grass Science Laboratory of Tibet Agricultural and Animal Husbandry
69 College. The experiment commenced on October 18, 2022, and the germination experiment ended on October
70 28, totaling 10 days. And the seedling experiment ended on November 15th for a total of 28 days.

71 **2.2 Experimental design**

72 **2.2.1 Germination test**

73 PEG-6000 solution (simulated drought stress) and NaCl solution (simulated salt stress) were prepared
74 with water potential of 0 (CK), -0.04, -0.14, -0.29, -0.49, -0.73, -1.02 MPa, respectively (Michel et
75 al.,1973;Burlyn et al.,1995). The experiment used the paper germination method. After soaking the filter paper
76 in the stress solution mentioned above, it was placed in a culture dish. 50 seeds were evenly placed in each
77 culture dish as one replicate, and each treatment setting was repeated three times. To ensure a constant water
78 potential in the environment, culture dishes were placed in a 25 °C constant temperature incubator, and each dish
79 was weighed. Distilled water was added the next day to reach a constant weight. The artificial climate incubator
80 has been set with a light intensity of 1250lx, a light cycle of 12h, and a dark cycle of 12h. The experiment
81 considered root length or bud length equal to seed length as germination, and the number of sprouts was counted
82 daily and moldy seeds were promptly treated.

83 2.2.2 Seedling test

84 The artificial climate incubator and solution preparation settings are the same as 2.2.1. The experiment
85 used sand culture as culture medium, each germination box (19×13×12 cm) was covered with 1000 g of dry sand,
86 100 ml of different gradients of PEG-6000 and NaCl solutions were added. After stirring evenly, sow at a depth
87 of 2-3 cm and a seeding rate of 5 g/m². To guarantee the water potential of the sand culture environment, the
88 distilled water was replenished three times a day, morning, noon and night, according to the evaporation in the
89 germination box, using the weighing method to ensure that the solution content in the sand culture was around
90 10 %. Replenish Hoagland Total Nutrient Solution every 7 days, 20ml each time. Shoots extending out of the
91 sand surface were considered as emergence. After all seedlings have emerged, the height of the aboveground
92 part was recorded. After 28 days, the root morphology indicators and the content of organic carbon, total
93 nitrogen, and total phosphorus in the aboveground and underground parts were measured.

94 2.3 Measurement indexes and methods

95 2.3.1 Germination test index

96 The Germination percentage, Germinating energy, root length, shoot length and other indicators related to
97 seed germination were determined and equated as follows:

98 Germination percentage (G,%) = (number of normally germinated seeds within 10 d of final
99 germination/number of seeds for testing) × 100%;

100 Germinating energy (GE, %) = (Number of seeds that germinated normally on the 5th day of germination
101 test/number of seeds for testing) × 100%;

102 Germination index (GI)= $\sum (Gt/Dt)$,Gt means the number of seeds germinated on day t and Dt means the
103 corresponding number of days to germination;

104 Vigor index (VI) = germination index × Sx, Sx is the mean shoot length;

105 Shoot length and root length: After the germination test, 10 seedlings were randomly selected from each
106 treatment petri dish and the test shoot length and root length were measured using a straightedge (1 mm).
107 Subsequent use for variance analysis.

108 2.3.2 Seedling test index determination and methods

109 10 seedlings were randomly selected from the germination boxes of each treatment, and the aboveground
110 height of 10 seedlings was measured daily. The trend of plant height change was plotted using the average value.
111 The plant growth rate was calculated every 7 days for a total of 4 cycles. The plant height growth rate $R=(L2-$
112 $L1)/7$, where L2 refers to the plant height measured on the last day of each cycle and L1 refers to the plant height
113 measured on the first day of each cycle. The total root length, surface area, volume and number of root tips were
114 determined by EPSON GT-X980 root scanner; plant height and root length were determined by straightedge.
115 After measuring the morphological indicators, *Elymus nutans* was dried in a 65 °C oven for 48 hours, and total
116 nitrogen content was determined by Kjeldahl method(Sing et al. , 2020); total phosphorus content was
117 determined by molybdenum blue ascorbic acid method(Li et al. , 2022); determination of organic carbon content
118 by Elementar vario TOC elemental analysis(Wolski et al. , 2022).

119 2.4 Data Statistics and Analysis

120 One-way analysis of the significance of seed Germination percentage, Germinating energy, root length,
121 shoot length, vigor index, and germination index under PEG-6000 versus NaCl solution stress was performed
122 using SPSS, respectively. SPSS was used to analyze seedling plant height, growth rate, total root length, root tip
123 number, root surface area, root volume, leaf root organic carbon content, root organic carbon content, leaf total
124 nitrogen content, root total nitrogen content, leaf total phosphorus content, root total phosphorus content, leaf
125 C:N, root C:N, leaf N:P, root N:P under PEG-6000 and NaCl solution stresses, respectively. The significance
126 was analyzed by one-way analysis. Principal component analysis and correlation analysis were performed using
127 Origin Pro 2021 on the environmental water potential, seedling height, total root length, root surface area, root
128 volume, leaf root organic carbon content, root organic carbon content, leaf total nitrogen content, root total
129 nitrogen content, leaf total phosphorus content, and root total phosphorus content of PEG-6000 and NaCl
130 solutions, respectively. A dose-response meta-analysis was conducted on the seed germination rate. The drought
131 and salt resistance of *Elymus nutans* seeds and seedlings were analyzed and compared using the affiliation
132 function method. The index was calculated as: $X(u) = (X-X_{min})/(X_{max}-X_{min})$, and if the index was negatively
133 correlated with drought resistance then the formula was: $X(u) = 1-(X-X_{min})/(X_{max}-X_{min})$.

134 3.Results

135 3.1 Effects of treatments on Seed germination

136 The PEG-6000 solution simulated drought stress and affected the germination rate of *E. nutans*. When
137 treated at -0.14 MPa, the germination rate of *E. nutans* seeds decreased by 15.33% compared to the control
138 group ($P<0.01$); With the decrease of environmental water potential, the germination rate of *Elymus nutans*
139 seeds under PEG-6000 solution stress showed a significant decrease trend ($P<0.05$); The lowest germination
140 rate was only 26.00% under the treatment of -1.02MPa (Figure 1: A). For germination potential, when the

141 environmental water potential was lower than -0.14 MPa, the germination potential was significantly lower
142 ($P < 0.01$) than CK, with a minimum of -1.02 MPa and a germination potential of 7.33 (Figure 1: B). In terms
143 of shoot length, with the decrease of environmental water potential, the shoot length of *E. nutans* showed a
144 significant decrease trend, and all treatments were significantly lower than CK ($P < 0.01$) (Figure 1C). In terms
145 of root length, there was no significant difference between the -0.04 and -0.14 MPa treatments and CK, while
146 the other treatments were significantly lower than CK ($P < 0.01$) (Figure 1D). The germination index showed a
147 very significant downward trend with the decrease of environmental water potential ($P < 0.01$), with the lowest
148 being -1.02 MPa treatment and the germination index being 11.67 (Figure 1E). The vitality index and
149 germination index showed a very significant downward trend with the decrease of environmental water potential
150 ($P < 0.01$), and the lowest germination index was 22.24 when treated with -1.02 MPa (Figure 1F).

151 The NaCl solution simulated salt stress and affected the germination of *E. nutans* seeds. Among them, the
152 -0.04MPa treatment had a 2.77% higher germination rate than CK treatment. As the environmental water
153 potential decreased, the germination rate also showed a decreasing trend. When the environmental water
154 potential was less than -0.49, -0.73, and -1.02MPa treatment, the seed germination rate was significantly lower
155 than CK treatment ($P < 0.01$), with the lowest being -1.02 MPa treatment, and the germination rate was 41.33%
156 (Figure 1A). In terms of germination potential, when the environmental water potential was less than -0.29 MPa,
157 it was significantly lower than CK ($P < 0.01$), with a minimum of -1.02 MPa and a germination potential of
158 11.33 (Figure 1B). For shoot length, with the decrease of environmental water potential, the shoot length of *E.*
159 *nutans* showed a significant decrease trend, and all treatments were significantly lower than CK ($P < 0.01$)
160 (Figure 1C). For root length, there was no significant difference between -0.04 and -0.14 MPa treatments and
161 CK, while the other treatments were significantly lower than CK ($P < 0.01$) (Figure 1D). The germination index
162 decreases with the decrease of environmental water potential, and is significantly lower than CK when treated
163 with -0.14 MPa ($P < 0.01$). The lowest germination index is 16.29 when treated with -1.02 MPa (Figure 1E).
164 The vitality index showed a very significant decrease with the decrease of environmental water potential ($P < 0.$
165 01), with the lowest being -1.02 MPa treatment and the vitality index being 49.71 (Figure 1F).

166 3.2 Effects of treatments on plant height and root structure

167 Under simulated drought stress with PEG-6000 solution, the plant height and root structure of *E. nutans*
168 showed a decreasing trend with the decrease of environmental water potential within 7-28 days (Figure 2A). In
169 terms of plant height growth rate, CK was significantly higher in the first week than other treatments ($P < 0.01$).
170 As time increased, the growth rate of CK plant height slowed down, with the fastest growth rate being -0.49
171 MPa treatment in the second week, -1.02 MPa treatment in the third week, and -0.73 MPa treatment in the
172 fourth week (Figure 2C). For the total root length, the treatment with -0.04 MPa has the longest root length,
173 which is 64.36 cm, 11.7 cm longer than CK; With the decrease of environmental water potential, the total length
174 of the root system showed a decreasing trend, with treatments of -0.29 MPa and below significantly lower than
175 CK ($P < 0.05$). When treated with -1.02 MPa, the total length of the root system was the lowest, at 22.78 cm
176 (Figure 2E). In terms of root surface area, the treatment with -0.04 and -0.14 MPa significantly increased the
177 root surface area compared to CK ($P < 0.05$). As the environmental water potential decreased, the root surface

178 area showed a decreasing trend, with the lowest being -1.02 MPa treatment, resulting in a root surface area of
179 2.05 cm² (Figure 2F). In terms of root volume, the root volume of -0.04 and -0.14 MPa treatments was higher
180 than that of CK. When treated with -0.29, -0.49, -0.73, -1.02 MPa, the root volume showed a significant decrease
181 trend compared to CK ($P < 0.05$). The minimum root volume is 0.02 cm³ after treatment with -1.02 MPa (Figure
182 2G). In terms of the number of root tips, -0.04, -0.14, -0.29, -0.49, and -0.73 MPa treatments were significantly higher
183 than CK ($P < 0.01$), with -0.04 treatment having the highest number of 251 (Figure 2H).

184 Under simulated salinization stress with NaCl solution, the plant height of *Elymus nutans* showed a
185 decreasing trend with the decrease of environmental water potential within 7-28 days (Figure 2B). In terms of
186 plant height growth rate, CK and -0.04 MPa were significantly higher in the first week than other treatments
187 ($P < 0.01$). The plant height growth rate was the fastest in the -0.73 MPa treatment in the second and third weeks,
188 and the growth rate was the fastest in the -1.02 MPa treatment in the fourth week (Figure 2D). For the total
189 length of the root system, the treatment with -0.14 MPa had the longest root length, which was 92.36 cm,
190 showing a highly significant level of 39.7 cm longer than CK ($P < 0.01$); With the decrease of environmental
191 water potential, the total root length showed a very significant decrease trend ($P < 0.01$), with the lowest being -
192 1.02 MPa treatment, resulting in a total root length of 5.37 cm (Figure 2E). In terms of root surface area, the
193 root surface areas of -0.04, -0.14, and -0.29 MPa treatments were all greater than CK. The maximum root
194 surface area of -0.14 MPa treatment was 10.61 cm², while the minimum surface area of -1.02 MPa treatment
195 was 2.1 cm² (Figure 2F). For the root volume, when treated with -0.04, -0.14, and -0.29 MPa, it was
196 significantly higher than CK ($P < 0.05$), with a maximum root volume of 0.131 cm³ under -0.14 MPa treatment.
197 As the environmental water potential decreases, the root volume showed a downward trend, with a minimum of
198 -1.02 MPa treatment reaching 0.02 cm³ (Figure 2G). In terms of the number of root tips, -0.04, -0.14, and -0.
199 29 MPa treatments were higher than CK, with -0.14 MPa treatment being significantly higher than CK ($P < 0.01$),
200 with 134 root tips. The minimum processing time was -1.02 MPa for 50 units (Figure 2H).

201 3.3 Effects of treatments on the Content of C, N, and P in Leaves and Roots

202 Under simulated drought stress with PEG-6000 solution, the organic carbon content in leaves of CK
203 treatment was lower than that of other treatments. Except for -0.04 MPa treatment, the organic carbon content
204 was significantly higher than CK ($P < 0.05$), with the highest being -0.73 MPa treatment and the highest being
205 467.54 g/kg. The -1.02 MPa treatment decreased by 4.64 g/kg compared to -0.73 MPa treatment (Figure 3A).
206 In terms of root organic carbon content, with the decrease of environmental water potential, the root organic
207 carbon content showed a decreasing trend, and all treatments were significantly lower than CK ($P < 0.01$). The
208 root organic carbon content of CK was 411.11 g/kg. The minimum treatment pressure was -1.02 MPa, and the
209 organic carbon content was only 286.59 g/kg (Figure 3D). In terms of total nitrogen content in leaves, the

210 treatments of -0.04, -0.14, and -0.29 MPa were significantly lower than CK ($P < 0.05$), while the treatments of
211 -0.73 and -1.02 MPa were extremely significantly higher than CK ($P < 0.01$), with the lowest content of 18.58
212 g/kg in the -0.04 MPa treatment. The maximum total nitrogen content of -1.02 MPa treatment was 29.44 g/kg
213 (Figure 3B). In terms of total nitrogen content in the root system, the total nitrogen content in the root system
214 decreases with the decrease of environmental water potential. The nitrogen content in the CK root system was
215 11.21 g/kg higher than that in the treatment group, and the lowest nitrogen content in the -1.02 MPa treatment
216 was 7.07 g/kg (Figure 3E). In the total phosphorus content of leaves, with the decrease of environmental water
217 potential, the total phosphorus content of leaves showed a trend of first increasing and then decreasing. The total
218 phosphorus content of leaves in all treatments was significantly higher than CK ($P < 0.01$), with the highest
219 content of 4.06 g/kg in the -0.73 MPa treatment (Figure 3C). In terms of total phosphorus content in the root
220 system, with the decrease of environmental water potential, the total phosphorus content in the root system
221 showed a trend of first increasing and then decreasing. The highest content in the -0.29 MPa treatment was 1.
222 87 g/kg, and the -0.04, -0.14, and -0.29 MPa treatments were significantly higher than CK ($P < 0.01$). The -0.
223 49, -0.73, and -1.02 MPa treatments were all significantly lower than CK ($P < 0.01$), and the lowest was 1.50
224 g/kg in the -1.02 MPa treatment (Figure 3F).

225 Under simulated salinization stress with NaCl solution, the organic carbon content in leaves showed a trend
226 of first increasing and then decreasing. Among them, the organic carbon content in leaves under -0.04, -0.14,
227 and -0.29 MPa treatments was significantly higher than CK ($P < 0.01$), but with the decrease of environmental
228 water potential, the organic carbon content in leaves under -0.49, -0.73, and -1.02 MPa treatments was
229 significantly lower than CK ($P < 0.01$) (Figure 3A). In terms of root organic carbon, with the decrease of
230 environmental water potential, all treatments were significantly lower than CK ($P < 0.01$) and showed a very
231 significant downward trend ($P < 0.01$), with the lowest being -1.02 MPa treatment, and the organic carbon
232 content was only 288.29 g/kg (Figure 3D). In terms of total nitrogen content in leaves, with the decrease of
233 environmental water potential, the total nitrogen content in leaves of all treatments was significantly higher than
234 CK ($P < 0.01$). Among them, the -0.73 MPa treatment had the highest total nitrogen content of 30.09 g/kg, and
235 the -1.02 MPa treatment decreased by 2.01 g/kg compared to the -0.73 MPa treatment (Figure 3B). In terms of
236 root total nitrogen, the -0.04 MPa treatment had a root total nitrogen content of 13.29 g/kg, which was
237 significantly higher than the other treatments of C ($P < 0.01$). The root total nitrogen content of -0.14, -0.29 MPa,
238 -0.49, -0.73, and -1.02 MPa treatments was significantly lower than that of CK ($P < 0.05$), with the lowest
239 content of 9.15 g/kg in the -1.02 MPa treatment (Figure 3E). In the total phosphorus content of leaves, with the
240 decrease of environmental water potential, the total phosphorus content of leaves showed a trend of first
241 increasing and then decreasing. The total phosphorus content of leaves in all treatments was significantly higher

242 than CK ($P < 0.01$), with the highest content of 3.66 g/kg in the -0.29 MPa treatment (Figure 3C). In the total
243 phosphorus content of the root system, with the decrease of environmental water potential, the total phosphorus
244 content of the root system first increases and then decreases- The highest content of 0.29 MPa treatment was 1.
245 87 g/kg, and the treatments of -0.04, -0.14, and -0.29 MPa were all significantly higher than CK ($P < 0.01$).
246 The treatments of -0.49, -0.73, and -1.02 MPa were all significantly lower than CK ($P < 0.01$), and the lowest
247 was 1.53 g/kg under -1.02 MPa treatment (Figure 3F).

248 3.4 Effects of treatments on C: N, N: P of seedlings

249 Under simulated drought stress with PEG-6000 solution, in terms of leaf C: N, with the decrease of
250 environmental water potential, leaf C: N showed a trend of first increasing and then decreasing. Among them,
251 the maximum C: N value under -0.04 MPa treatment was 21.87, and the minimum value under -1.02 MPa
252 treatment was 15.72. Moreover, under -1.02 MPa treatment, it was significantly lower than CK ($P < 0.01$) (Figure
253 4A). In terms of root C: N, the root C: N of each treatment was significantly higher than that of CK ($P < 0.05$),
254 with the highest being 42.76 in the -0.04 MPa treatment (Figure 4C). In terms of leaf N: P, the N: P values of
255 all treatments were significantly lower than those of CK ($P < 0.01$), with the lowest N: P value of 5.94 in the -0.
256 73 MPa treatment (Figure 4B). In terms of root N: P, the N: P values of each treatment were significantly lower
257 than those of CK ($P < 0.01$), with the lowest being -0.29 MPa treatment, 4.34 (Figure 4D).

258 Under simulated salinization stress with NaCl solution, with the decrease of environmental water potential,
259 the C: N value of leaves showed a trend of first increasing and then decreasing. Among them, the maximum C:
260 N value under -0.14 MPa treatment was 19.86, and the minimum value under -1.02 MPa treatment was 12.72.
261 Among them, the values under -0.29, -0.49, -0.73, and -1.02 MPa treatment were significantly lower than CK
262 ($P < 0.01$) (Figure 4A). In terms of root C: N, the root C: N of each treatment was significantly lower than that
263 of CK ($P < 0.01$), with the lowest being 27.23 in the -0.04 MPa treatment (Figure 4C). In terms of leaf N: P, the
264 N: P values of -0.04, -0.14, -0.29, and -0.49 MPa treatments were significantly lower than CK ($P < 0.01$), while
265 the N: P values of -0.73 and -1.02 MPa treatments were slightly higher than CK, with the highest being 10.23
266 under -1.02 MPa treatment (Figure 4B). In terms of root N: P, the N: P value of -0.04 MPa treatment was 7.33,
267 which was significantly higher than CK ($P < 0.01$), while the N: P values of other treatments were significantly
268 lower than CK ($P < 0.01$). Among them, the lowest N: P value of -0.29 MPa treatment was 5.66 (Figure 4D).

269

270 3.5 Comprehensive evaluation

271 As shown in Figure 5A, the environmental water potential of PEG-6000 solution showed a linear
272 relationship with germination rate ($p = 0.000$). The model formula was Probit (p) = $0.670 + 1.442 * \text{environmental}$
273 $\text{water potential (PEG-6000)}$, and the LD50 value was -0.465 (95% CI: 0.425, 0.505). According to (Figure 5B,
274 C), it can be seen that under PEG-6000 stress, environmental water potential showed a highly significant negative

275 correlation with leaf organic carbon content, leaf total nitrogen content, and leaf total phosphorus content ($P < 0.01$). The environmental water potential showed a highly significant positive correlation with plant height, total
276 root length, root surface area, root volume, root organic carbon, root total nitrogen content, and root total
277 phosphorus content ($P < 0.01$). There was a highly significant negative correlation between plant height and leaf
278 organic carbon content, leaf total nitrogen content, and leaf total phosphorus content ($P < 0.01$). And there was a
279 highly significant positive correlation between plant height and total root length, root surface area, root volume,
280 root organic carbon, root total nitrogen content, and root total phosphorus content ($P < 0.01$). The total root
281 length, root surface area, and root volume showed a highly significant negative correlation with leaf organic
282 carbon content, leaf total nitrogen content, and leaf total phosphorus content ($P < 0.01$); The total root length,
283 root surface area, root volume showed a highly significant positive correlation with plant height, root organic
284 carbon, root total nitrogen content, and root total phosphorus content ($P < 0.01$).

285
286 As shown in Figure 5D, there was a linear relationship between the environmental water potential of NaCl
287 solution and germination rate ($p = 0.000$), with Probit (p) = $0.961 + 1.131 * \text{environmental water potential (NaCl)}$,
288 corresponding to an LD_{50} value of -0.850 MPa (95% CI: $0.770, 0.929$). Under the simulated salinization stress
289 of NaCl solution (Figure 5E, F), the environmental water potential showed a negative correlation with the total
290 nitrogen content and total phosphorus content of leaves, among which it reached a very significant level with
291 the total nitrogen content of leaves ($P < 0.01$); The environmental water potential showed a highly significant
292 positive correlation with leaf organic carbon content, total root length, root surface area, root volume, plant
293 height, root organic carbon content, root total nitrogen content, and root total phosphorus content ($P < 0.01$).
294 There was a highly significant negative correlation between plant height and total nitrogen content in leaves
295 ($P < 0.01$). And there was a highly significant positive correlation between plant height and total root length, root
296 surface area, root volume, leaf organic carbon, root organic carbon, root total nitrogen content, and root total
297 phosphorus content ($P < 0.01$). The total root length, root surface area, root volume, and total nitrogen content in
298 leaves showed a highly significant negative correlation ($P < 0.01$); The total root length, root surface area, root
299 volume showed a significant positive correlation with plant height, leaf organic carbon content, root organic
300 carbon, root total nitrogen content, and root total phosphorus content ($P < 0.05$).

301 4 Discussion

302 4.1 Effect of drought stress and salinity stress on seed germination of *E. nutans*

303 Seed germination is the starting point of plant life history, and water is one of the necessary conditions for
304 seed germination. Water deficiency will affect the activity of internal enzymes, cell division, and other
305 physiological metabolic processes in seeds. At the same time, water deficiency may cause seeds to lose vitality
306 due to not being able to absorb enough water at once, thereby affecting seed germination. (Costa et al., 2021;
307 Atia et al., 2011). This experiment showed that water deficit caused by PEG-6000 simulated drought stress and
308 NaCl simulated salinization stress can affect the germination of *E. nutans* seeds (Figure 1A). Under -0.04 MPa
309 treatment, NaCl solution stress promoted *E. nutans* seed germination, which was 2.77% higher than CK. PEG-
310 6000 solution stress inhibited *E. nutans* seed germination; Moreover, under the lowest environmental water
311 potential of -1.02 MPa in this experiment, the germination rate of NaCl solution treatment still exceeded 40%,
312 while the germination rate of PEG-6000 solution treatment was only 26%. According to the dose-response

313 analysis results, the LD_{50} values under PEG-6000 and NaCl stress were -0.465 and -0.850MPa, respectively.
314 This may be due to the different expression of ABA signals in drought and salt stress during the germination
315 process of *E. nutans* seeds, or because *E. nutans* has relatively high salt tolerance, and the PEG-6000 solution
316 stress treatment has a more direct impact on the seed germination process. (He et al., 2022; Li et al., 2020;
317 Nakashima et al., 2013). And under all treatments of the two solutions, GE, GI, and VI were lower than CK
318 (Figure 1B, E, F), which may be related to the response of seed enzyme activity to water deficiency. Although
319 both solutions reduced GE, GI, and VI under stress, under the same environmental water potential treatment, the
320 GE, GI, and VI of NaCl solution were higher than those of PEG-6000 solution treatment. This may be due to the
321 PEG-6000 solution treatment affecting the process of seed germination, causing the seeds to not fully absorb
322 water and resulting in slow germination speed. (Dietz et al., 2021; Pan et al., 2021). The study by Wang (Wang
323 et al., 2017) showed that with the increase of PEG-6000 and NaCl solution concentration, the germination index
324 of *Elymus nutans* seeds showed a downward trend, which is basically consistent with this study.

325 Under the same environmental water stress of the 2 solutions in this study (Fig. 1C, D), *E. nutans* bud length
326 showed a significant decrease, and each treatment was highly significant lower than the CK treatment ($P < 0.01$),
327 but the decrease in bud length was lower under the NaCl solution treatment than the PEG-6000 solution
328 treatment; The bud length of PEG-6000 treatment was higher than that of NaCl treatment when the ambient
329 water potential was -0.04, -0.14 and -0.29 MPa treatments; NaCl bud length exceeded that of PEG-6000 when
330 the ambient water potential was -0.49, 0.73, and -1.02 MPa treatments. PEG-6000 and NaCl solution
331 treatments also reduced root length, and root length was greater under all PEG-6000 treatments than NaCl
332 treatments. This is due to the fact that under PEG-6000 solution stress *E. nutans* safeguards root water uptake
333 by increasing root biomass, whereas NaCl solution causes direct damage to plant root cells inhibiting plant root
334 growth (Dietz et al., 2021; Pan et al., 2021). It has been shown (Song et al., 2000; Wang et al., 2021) that the
335 root length and bud length of *E. nutans* seeds show a decreasing trend when stressed by PEG-6000 and NaCl
336 solutions during germination, which is basically consistent with the present study. Moreover, the analysis of seed
337 germination indexes under PEG-6000 and NaCl stresses by the affiliation function method revealed that (Table
338 1), the inhibition of *E. nutans* seed germination by PEG-6000 solution stress under the same environmental water
339 potential was stronger than that by NaCl solution stress.

340 **4.2 Effects of drought stress and salinity stress on *E.nutans* plant height, total root length, root surface** 341 **area, and root volume**

342 The most intuitive response of plants to water deficiency is the morphological indicators of the aboveground
343 and underground parts. Water deficiency can lead to shorter plants, thinner stems and leaves, and also affect the
344 total length, surface area, volume, and other aspects of the root system. (Ulrich et al., 2022; Dell'Aversana et

345 al.,2021). This experimental study showed that with the decrease of environmental water potential, the stress of
346 two different solutions would reduce the plant height of *E.nutans*. Among them, PEG-6000 solution showed a
347 significant decrease in plant height under -0. 04, -0. 14, and -0. 29 MPa treatment, while NaCl solution did not
348 show this situation. However, the two solutions showed exactly the same decrease trend under -0. 49 MPa
349 treatment. And with the increase of sowing time, the growth rate of *E. nutans* plant height showed a decreasing
350 trend under CK treatment. However, with the decrease of environmental water potential, the growth rate of plant
351 height under both solution stresses showed an increase in low environmental water potential over time, while a
352 decrease in high environmental water potential over time. The reason may be that during the first two weeks
353 after seed germination under high environmental water potential, the growth rate of plant height is faster than
354 under low concentration environmental water potential stress. As time goes on, the growth rate of plant height
355 under low concentration environmental water potential gradually increases, which is due to the slowing down of
356 plant height growth under high concentration environmental water potential. This is basically consistent with
357 Ling's (Ling's et al. ,2020) research on rice seedlings under salt stress.

358 When lacking water, plants compensate for the decrease in root absorption area by enhancing root vitality
359 and maintaining a higher root biomass, thereby maintaining a higher root water absorption capacity. The total
360 length, surface area, volume, and number of root tips of the root system have a significant impact on the water
361 absorption capacity of the plant root system. (Bloom ,2015;Fowda et al.,2022;Lambers et al.,2015). This
362 experimental study showed that with the decrease of environmental water potential, the total root length of *E.*
363 *nutans* showed a trend of first increasing and then decreasing. Among them, under the treatment of PEG-6000
364 solution, the total root length of -0. 04Mpa treatment was the longest, while under the treatment of NaCl solution,
365 the total root length of -0. 14MPa treatment was the longest. Moreover, under the treatment of -0. 04, -0. 14, -0.
366 29, and -0. 49, the total root length of NaCl solution treatment was greater than that of PEG-6000 solution
367 treatment, but under the treatment of -0. 73 and -1. 02 MPa, the total root length of NaCl was significantly
368 reduced, treatment below PEG-6000. The surface area, volume, and total length of the root system showed
369 identical trends, and the maximum area and volume of PEG-6000 and NaCl solutions were also -0. 04Mpa and
370 -0. 14MPa treatments, respectively. However, in terms of surface area and volume, NaCl treatments were higher
371 than PEG-6000. The number of root tips also showed a trend of first increasing and then decreasing. The PEG-
372 6000 solution with -0. 04 MPa treatment had significantly higher root tip numbers than other treatments ($P<0.$
373 01), and the NaCl solution with -0. 14 MPa treatment was also significantly higher than other treatments ($P<0.$
374 01). However, the PEG-6000 solution had a relatively gentle decrease in root tip numbers, while the NaCl
375 solution had a significant decrease in root tip numbers, which was lower than the CK treatment at -0. 73 and -1.
376 02 MPa treatments. Related studies have shown (Hazman et al., 2018; Wang et al., 2018) that a small amount of

377 water deficiency promotes plant root growth, resulting in an increase in the total length, surface area, volume,
378 and number of root tips of plant roots. However, as the environmental water potential decreases, the degree of
379 drought and salt damage to the roots increases, leading to root growth inhibition, which is basically consistent
380 with the results of this study.

381 **4.3 Effects of drought stress and salinity stress on the organic carbon, total nitrogen and total** 382 **phosphorus contents of *E.nutans* leaves and roots**

383 When facing drought and salt stress, the organic carbon content of plant leaves and roots is affected to adapt
384 to the impact of environmental water potential changes on plant growth. (Ulrich et al. ,2022;Dell'Aversana et al.
385 ,2021). This study indicated that the organic carbon, total nitrogen, and total phosphorus contents of *E. nutans*
386 leaves were all affected by the decrease in environmental water potential. In terms of organic carbon content,
387 PEG-6000 solution showed a gradual upward trend under stress, and the highest content was observed when
388 treated with -0. 73 MPa. Under NaCl solution stress, the content showed a trend of first increasing and then
389 decreasing, with the highest content observed under -0. 14 MPa treatment. The organic carbon content in the
390 root system of *E. nutans* was directly affected by environmental water potential. With the decrease of
391 environmental water potential, both PEG-6000 solution and NaCl solution showed a significant decrease trend
392 ($P<0. 01$), and except for the -0. 04MPa treatment, the organic carbon content in the root system under NaCl
393 solution stress was higher than that under PEG-6000 solution stress. As the degree of water deficiency increases,
394 the organic carbon content in the roots of *E. nutans* showed a decreasing trend, which is different from the results
395 of Dell'Aversana (Dell'Aversana et al. ,2021) under salt stress in barley and may be due to different research
396 subjects.

397 Nitrogen (N) and phosphorus (P) are both basic macronutrients that limit plant growth and primary
398 productivity in different terrestrial ecosystems (Bloom,2015;Fowda et al. ,2022;Lambers et al. ,2015). In
399 terms of leaves, the effects of PEG-6000 and NaCl stress on the total nitrogen content of *E. nutans* showed
400 completely different trends. Under salt stress, the total nitrogen content of leaves was higher than CK, but under
401 drought stress, it showed a trend of first decreasing and then increasing. Both solutions of stress increased the
402 total phosphorus content in *E. nutans* leaves, and both showed a trend of first increasing and then decreasing.
403 Both solutions of stress increased the total phosphorus content in *E. nutans* leaves, and both showed a trend of
404 first increasing and then decreasing. In terms of roots, under PEG-6000 stress, the total nitrogen content of *E.*
405 *nutans'* roots was lower than that of CK, while under the treatment of -0. 04MPa in NaCl solution, the total
406 nitrogen content of *E. nutans'* roots was higher than that of CK, and under the same environmental water
407 potential, the total nitrogen content of *E. nutans'* roots was higher than that under PEG-6000 solution stress. The
408 decrease in environmental water potential had little impact on the total phosphorus content in roots, and both
409 solutions showed a trend of first increasing and then decreasing under stress. The trend of nitrogen content
410 changes in plants in this experiment is basically consistent with Zhao's study on *E. nutans* under drought
411 stress(Zhao et al. ,2020;). The changes in phosphorus content are consistent with some of Liang's research results,
412 which may be due to different experimental environments and methods(Liang et al. ,2022).

413 **4.4 Effects of C, N and P contents of above and below ground parts on the growth of *E.nutans***

414 C and N are the two most fundamental elements for plant growth and development, and their mutual
415 coupling makes C:N an important indicator for exploring plant element allocation and adaptation strategies (He

416 et al. ,2019; Wang et al. ,2015). For example, high C:N plants have high nitrogen use efficiency; while low C:N
417 apoplankton has the quality of fast decomposition. This experiment showed that water deficit affected C:N of *E.*
418 *nutans* leaves and roots, basically showing a rise followed by a fall, and that C:N was higher for PEG-6000
419 simulated drought stress than for NaCl simulated salinity stress under the same environmental water potential
420 treatment. It may indicate that water potential stress had an effect on the nitrogen utilization of *E. nutans* and
421 showed a trend of first had and then decreasing, and NaCl solution stress at the same ambient water potential
422 had a greater effect on nitrogen utilization, which is close to part of the findings of Wan (Wan et al. ,2022).

423 The balance of the coupling of N and P has a great impact on the structure and function of plants from the
424 molecular level to the biome level or different biological tissues (Niu et al. ,2019; Liang et al. ,2022). This
425 experiment showed that water deficit affected the nitrogen and phosphorus balance of *E. nutans* leaves and roots,
426 and showed a trend of first will then increase in the leaves nitrogen and phosphorus ratio; Both leaves and roots
427 N:P were higher in NaCl solution treatment than in PEG-6000 solution treatment under the same ambient water
428 potential treatment. It may be suggested by the N:P aspect that NaCl solution stress has a greater effect on the
429 biological organization and function of *E. nutans*.

430 **4.5 Effects of C, N and P contents of above and below ground parts on the growth of *E.nutans***

431 Drought stress and salt stress alter the C, N, and P contents of plant leaves, and the changes in plant leaf
432 and root morphology are particularly pronounced (Zhou et al. , 2022; Dibar et al. , 2020).
433 The results of the correlation analysis showed (Fig. 5C, F) that the growth of plant leaves and roots under drought
434 stress would be hindered by the reduction of leaf organic carbon, total nitrogen and total phosphorus contents.
435 The main factor leading to the stunted growth of plant leaves and roots under salinity stress is the total nitrogen
436 content of the leaves. The main factor affecting plant growth under the 2 solution stresses is the change of
437 elements in the leaves, so it may be possible to improve the resistance of forage seedlings to drought stress and
438 salt stress by foliar spraying of fertilizers (Gao et al.,2022; Shabbir et al.,2015). The results of the analysis of the
439 growth indicators of *E. nutans* seedlings using the affiliation function method showed (Table 2) that under the
440 same environmental water stress, *E. nutans* seedlings were subjected to salinity stress to a lesser extent than
441 drought stress and showed some salt tolerance.

442 **5 Conclusions**

443 This experiment unified the effects of PEG-6000 and NaCl solutions on water deficit in forage by adjusting
444 the osmotic potential of the solution, and explored the effects of drought stress and salinization stress on the
445 growth of *E. nutans* seedlings under the same environmental water potential stress. The results indicated that
446 although under the same environmental water potential, there were significant differences in the responses of
447 drought stress and salinization stress to the germination, seedling growth, and organic carbon, total nitrogen, and
448 total phosphorus in the aboveground and underground parts of *E. nutans*. A comprehensive assessment of *E.*
449 *nutans* seed germination and seedling indicators using the affiliation function method revealed that *E. nutans*

450 was more severely affected by drought stress at both the seed germination and seedling growth stages, showing
451 results of better salt tolerance than drought tolerance.

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Figure 1

Figure 1: Effects of Two Solutions on Seed Germination

Two types of solution stress affect *E. nutans* seed germination rate (A), germination potential (B), bud length (C), root length (D), germination index (E), and vitality index (F). Different lowercase letters indicate significant differences in the same solution under different treatments ($P < 0.05$), different capital letters indicate that the differences in the same solution under different treatments reach a highly significant level ($P < 0.01$).

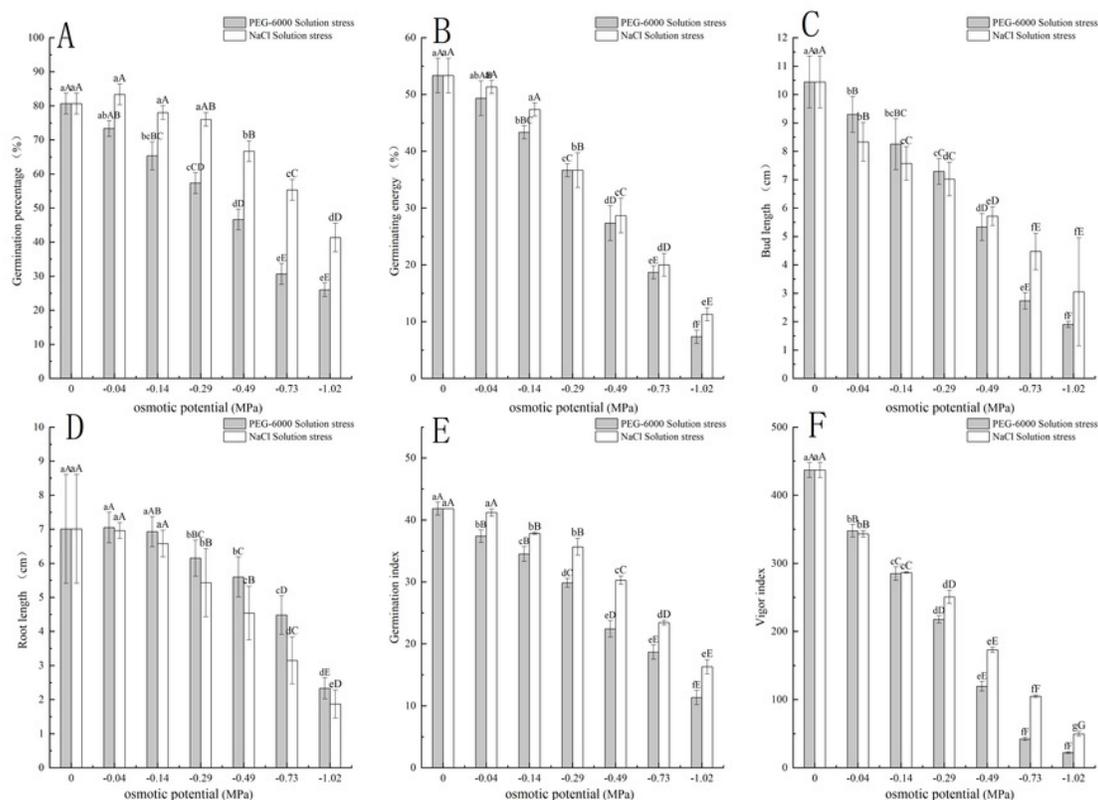


Figure 2

Figure 2 Effects of processing on the growth of aboveground and underground parts

Changes in plant height of *E.nutans* under PEG-6000 and NaCl solution stress (A, B), plant height trends (C, D), total root length (E), root surface area (F), root volume (G), and root tip number (H) on nutans. Different lowercase letters indicate significant differences in the same solution under PEG different the MPa treatments ($P < 0.05$), different capital letters indicate that the differences in the same solution under different treatments reach a highly significant level ($P < 0.01$).

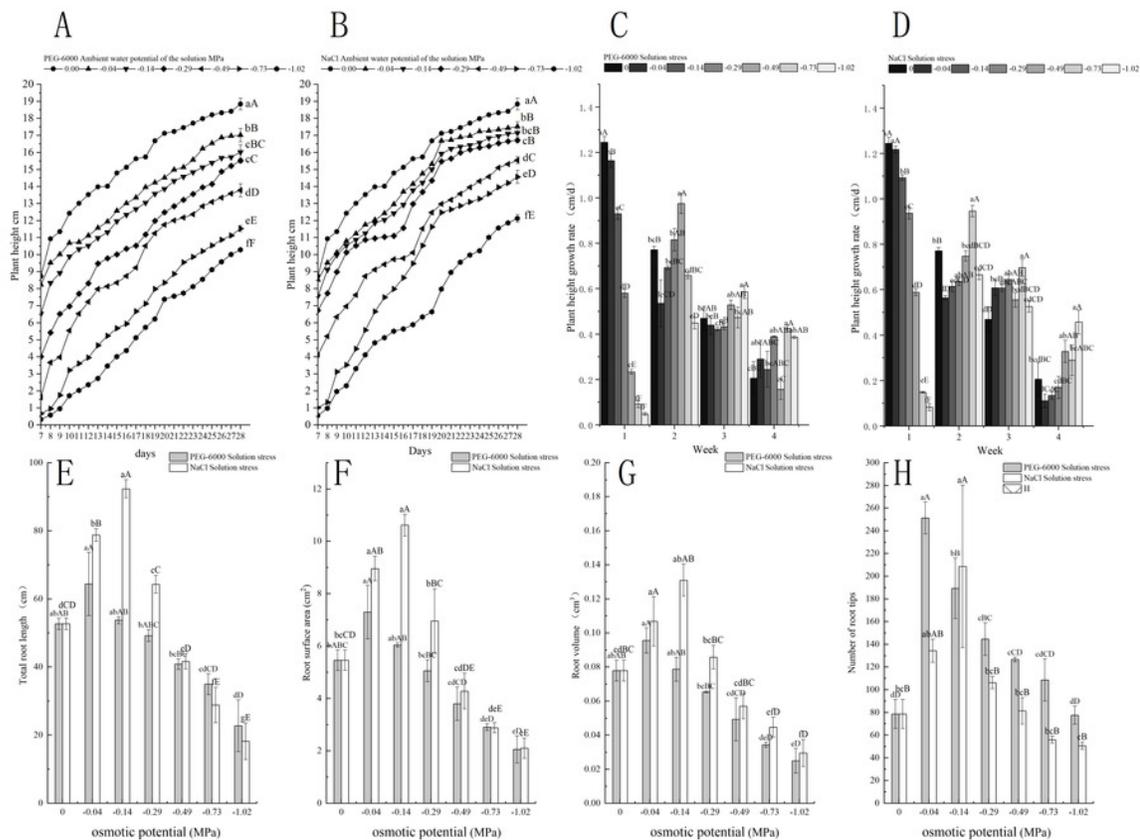


Figure 3

Figure 3 Effect of processing on C, N, and P content in the aboveground and underground parts

Two types of solution stress affect *E. nutans* leaf organic carbon (A), leaf total nitrogen (B), leaf total phosphorus (C), root organic carbon (D), root total nitrogen (E), and root total phosphorus (F) contents on nutans. Different lowercase letters indicate significant differences in the same solution under different treatments ($P < 0.05$), different capital letters indicate that the differences in the same solution under different treatments reach a highly significant level ($P < 0.01$).

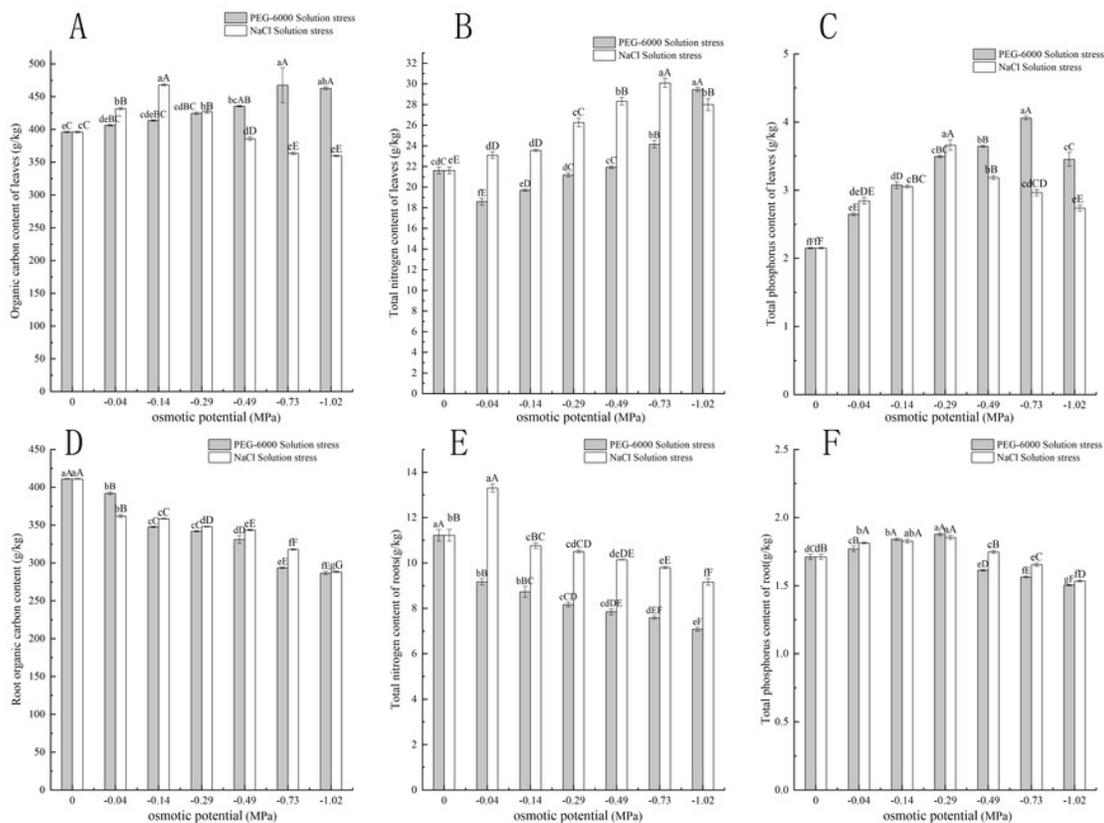


Figure 4

Figure 4: Effects of Two Solutions on C: N and N: P in Leaves and Roots

Two types of solution stress on *E. nutans* leaves C: N (A), leaf N: P (B), root C: N (C), and root N: P (D). Different lowercase letters indicate significant differences in the same solution under different treatments ($P < 0.05$), different capital letters indicate that the differences in the same solution under different treatments reach a highly significant level ($P < 0.01$).

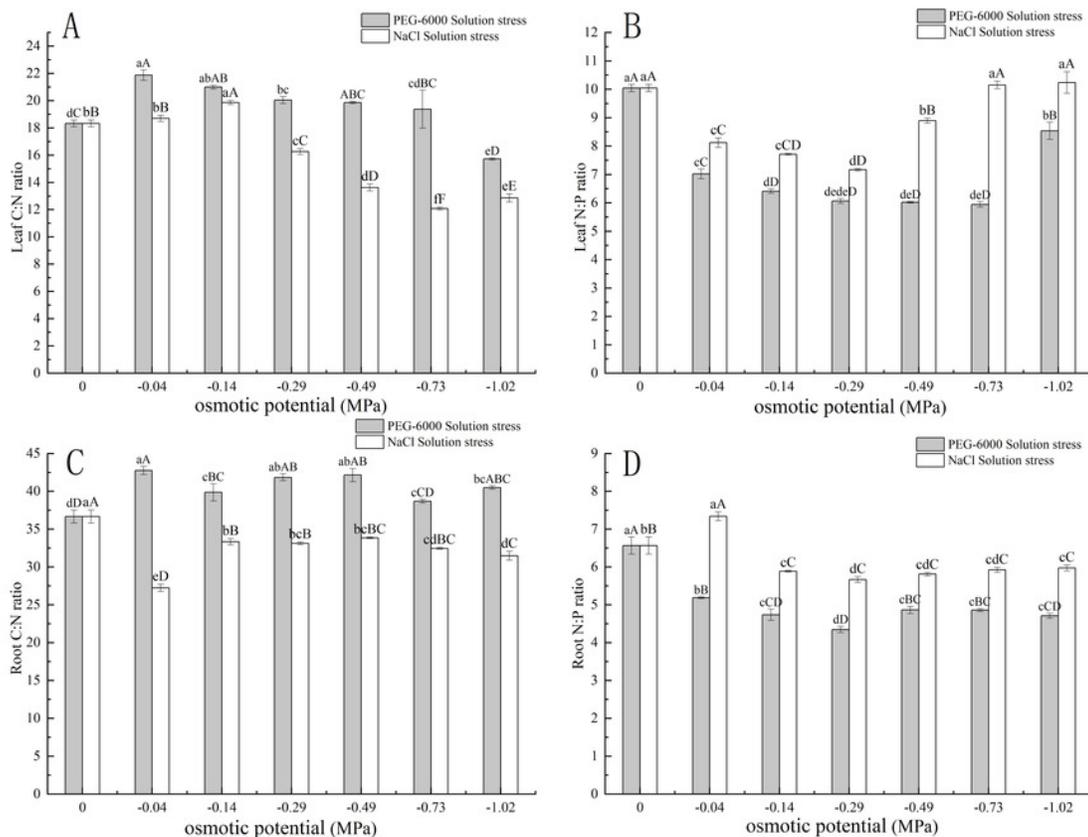


Figure 5

Figure 5 Comprehensive Analysis

Figure 5 Dose response analysis of PEG-6000 stress on seed germination (A); Principal Component Analysis of PEG-6000 on Seedling Growth (B); Factor analysis of PEG-6000 on seedling growth (C). Dose response analysis of NaCl stress on seed germination (D); Principal component analysis of NaCl on seedling growth (E); Factor analysis of NaCl on seedling growth (F). (EWP, environmental water potential; RL, total root length; RSA, root surface area; RV, root volume; RNT, number of root tips; H, plant height; LN, total nitrogen content in leaves; LP, total phosphorus content in leaves; LC, organic carbon content in leaves; RN, total nitrogen content in roots; RP, total phosphorus content in roots; RC, organic carbon content in roots). $**P < 0.01$, $*P < 0.05$.

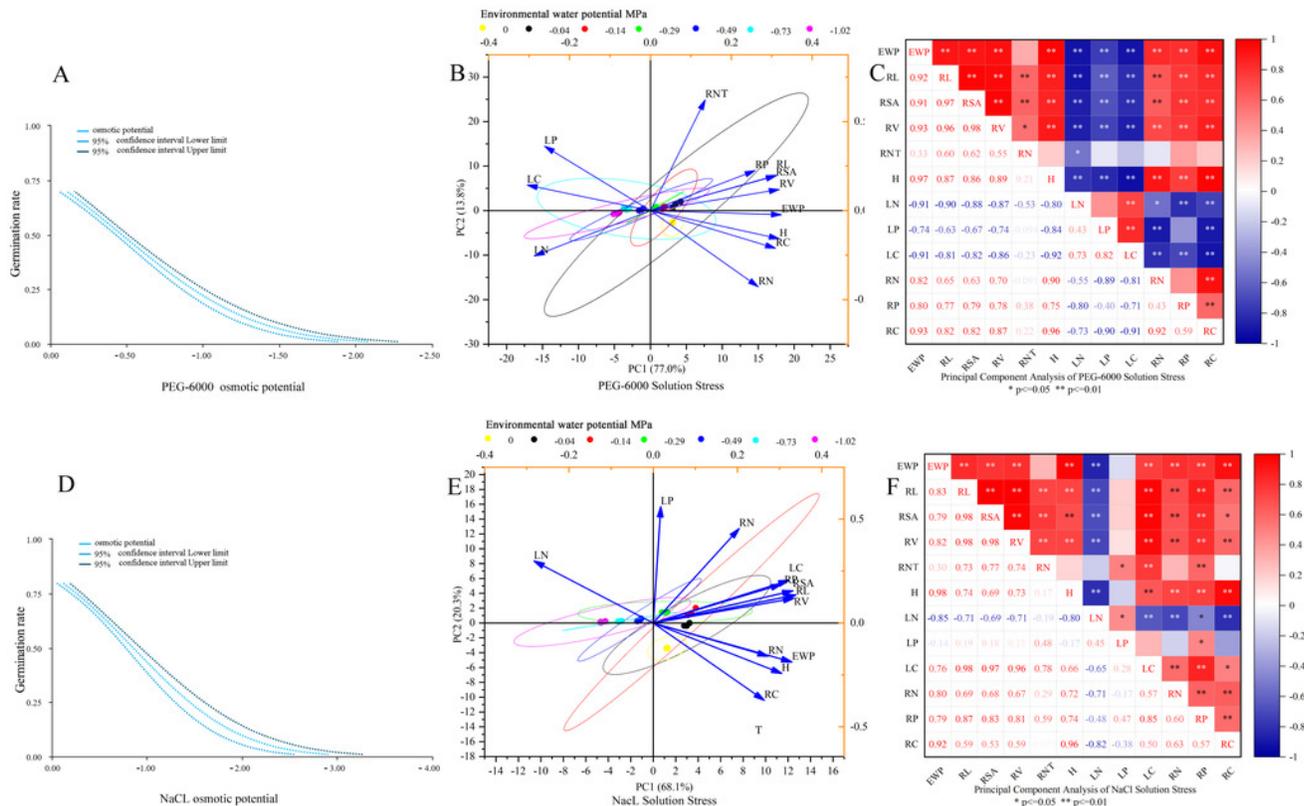


Table 1 (on next page)

table 1

Table 1 Affiliation function values and ranking of seed germination data

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| Solution | membership function value | | | | | | average value | sort |
|----------|---------------------------|------|------------|-------------|-------------------|-------------|---------------|------|
| | G | GE | bud length | root length | Germination index | Vigor index | | |
| PEG-6000 | 0.50 | 0.52 | 0.52 | 0.44 | 0.63 | 0.59 | 0.52 | 2 |
| NaCl | 0.64 | 0.57 | 0.62 | 0.46 | 0.62 | 0.46 | 0.56 | 1 |

1

Table 2 (on next page)

table 2

Table 2 Seedling growth data affiliation function values and ranking

Table 2 Seedling growth data affiliation function values and ranking

| Solution | membership function value | | | | | | | | | | | average value | sort |
|----------|---------------------------|------|------|------|------|------|------|------|------|------|------|---------------|------|
| | RL | RSA | RV | RNT | H | LN | LP | LC | RN | RP | RC | | |
| PEG-600 | 0.51 | 0.45 | 0.49 | 0.32 | 0.49 | 0.35 | 0.55 | 0.33 | 0.34 | 0.50 | 0.46 | 0.43 | 2 |
| NaCl | 0.50 | 0.45 | 0.46 | 0.31 | 0.56 | 0.48 | 0.50 | 0.45 | 0.36 | 0.59 | 0.48 | 0.46 | 1 |

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