

Explicating genetic architecture governing nutritional quality in pigmented rice

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Rice is one of the most important staple plant foods that provide a major source of calories and nutrients for tackling the global hunger index especially in developing countries. In terms of nutritional profile, pigmented rice grains are favoured for their nutritional and health benefits. The pigmented rice varieties are rich sources of flavonoids, anthocyanin and proanthocyanidin that can be readily incorporated into diets to help address various lifestyle diseases. However, the cultivation of pigmented rice is limited due to low productivity and unfavourable cooking qualities. With the advances in genome sequencing, molecular breeding, gene expression analysis and multi-omics approaches, various attempts have been made to explore the genetic architecture of rice grain pigmentation. In this review, we have compiled the current state of knowledge of the genetic architecture and nutritional value of pigmentation in rice based upon the available experimental evidence. Future research areas that can help to deepen our understanding and help in harnessing the economic and health benefits of pigmented rice are also explored.

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27 Abstract

28 Rice is one of the most important staple plant foods that provide a major source of calories and
29 nutrients for tackling the global hunger index especially in developing countries. In terms of
30 nutritional profile, pigmented rice grains are favoured for their nutritional and health benefits.
31 The pigmented rice varieties are rich sources of flavonoids, anthocyanin and proanthocyanidin
32 that can be readily incorporated into diets to help address various lifestyle diseases. However, the
33 cultivation of pigmented rice is limited due to low productivity and unfavourable cooking
34 qualities. With the advances in genome sequencing, molecular breeding, gene expression
35 analysis and multi-omics approaches, various attempts have been made to explore the genetic
36 architecture of rice grain pigmentation. In this review, we have compiled the current state of
37 knowledge of the genetic architecture and nutritional value of pigmentation in rice based upon

38 the available experimental evidence. Future research areas that can help to deepen our
39 understanding and help in harnessing the economic and health benefits of pigmented rice are also
40 explored.

41 **Keywords:** Pigmented rice, nutrients, secondary compounds, breeding, genetic engineering

42

43 **Introduction**

44 The sustainable food systems for ever-increasing human population include not only adequate
45 food but also adequately balanced nutrient rich food. An increased focus on foods that provide
46 adequate nutrition has resulted from a deeper understanding of the role of nutrition in maintaining
47 a healthy population, particularly in developing nations.

48 Rice is an important staple crop that feeds more than half of the world's population and is
49 being cultivated on approximately 158 million hectares of land producing around 850 million
50 tons of grains annually (*Krishnan et al., 2020*). Asia represents a major rice growing region
51 amounting for about 85% of the total production, followed by Latin America and Africa. India,
52 being the second largest producer of rice in the world contributes around 134 million metric
53 tonnes with productivity of 2.8 t/ha and in a total cultivated area of around 46 million hectares
54 (<https://www.statista.com/statistics/agriculture/farming/2022>). Rice is a good option for a healthy
55 diet as it has no cholesterol, fat, or sodium and contains eight essential amino acids in a balanced
56 proportion. Rice bran oil is rich in linoleic and oleic acid, which are essential for sustaining cell
57 membranes and nervous system functioning (*Bhat et al., 2020*). Rice grains come in a variety of
58 pigmentations, including yellow, green, brown, red, purple, and black. Pigmented rice (or
59 coloured rice) has long been considered to have nutraceutical benefits resulting in ongoing
60 production as niche rice in various parts of the world.

61 Pigmented landraces of rice have a higher content of total anthocyanin, total phenol and
62 polyphenol which signifies high antioxidant potential (*Deng et al., 2013*). Due to their rich
63 nutritional profile and high antioxidants, these rice types have the potential to boost human health
64 by addressing a variety of metabolic disorders (**Figure 1**). The consumption of coloured rice
65 reduces oxidative stress while simultaneously increasing antioxidant capacity in animal models,
66 and this may be linked to a lower risk of chronic diseases like cardiovascular disease, type 2

67 diabetes and some cancers (Wongsa, 2020). As a result of their health potential and widespread
68 demand, several nations have evaluated their coloured rice and created newer types. Varieties of
69 coloured rice, recognized for their nutritional advantages, have remained a primary crop in
70 several regions of India (**Figure 1**). Molecular breeding and biotechnology approaches are
71 regularly used for increasing the nutritional components in pigmented rice as well as in
72 transferring these quality traits to conventional white rice. However, there lies a gap between the
73 research that is available in the form of specific publications and the thoughtful compiled
74 information that is required to gain an overall understanding of the topic. Hence, the present
75 review highlights information on the important nutritional components in pigmented rice grains
76 along with the efforts made to increase the nutritional quality of both pigmented and white rice
77 through various biotechnological approaches to address nutritional food security. The present
78 review will serve as a comprehensive review for scholars, scientists and students working in the
79 area of pigmented rice.

80 **Survey Methodology**

81 For this article, we have conducted a rigorous literature review and collected information from
82 research articles, review articles, book chapters, websites (www.statista.com/;
83 www.agricoop.gov.in/) and databases (<http://rice.uga.edu/>; <https://www.gramene.org/>;
84 <https://rapdb.dna.affrc.go.jp/>). We have also used Google image for drawing a map of India. We
85 excluded the studies having abstracts available only with no full-text articles

86 **Nutritional Composition of Pigmented Rice**

87 *Protein and Amino acids*

88 Our bodies' healthy condition as well as tissue repair depends on the protein that we take from
89 foods. Since rice is a staple food for a major part of the global population so rice protein is
90 important in human nutrition. Rice protein concentrates and isolates are useful components in
91 many food applications because they offer special nutritional qualities that set them apart from
92 other cereals and legume proteins, such as a well-balanced amino acid profile and being easily
93 digestible and non-allergic (Romero, 2014; Sati and Singh, 2019). Rice proteins are also
94 excellent ingredients for baby food recipes due to their distinct nutritional and hypoallergenic
95 qualities. Rice protein hydrolysates have the potential to improve food emulsion stability and
96 serve as a natural antioxidant (Rani et al., 2018). In rice, glutelins dominates seed storage

97 proteins, making up to 60–80% of the total protein in the seed by weight. Compared to other
98 grain seeds, that contains higher prolamine content as a proportion of total protein, they make up
99 only roughly 20–30% of the protein in rice seeds (*Kawakatsu et al., 2008*).

100 Moreover, the amount and quality of protein is much better in pigmented rice as
101 compared to non-pigmented rice. *Chen et al. (2016)* compared pigmented and white rice grains
102 at five different stages of growth and found 230 differently abundant proteins associated with
103 several metabolic activities. The pigmented grains were shown to have lower concentrations of
104 proteins involved in signalling pathways, redox homeostasis, photosynthesis, nitrogen fixation,
105 and tocopherol synthesis. In contrast, it was discovered that the pigmented grain had higher
106 levels of proteins necessary for the synthesis of sugars and flavonoids. Pigmented rices were also
107 found to have a higher concentration of proteinogenic amino acids (histidine, threonine, valine,
108 iso-leucine, methionine, phenyl alanine, lysine, proline, and tyrosine) and non-proteinogenic
109 amino acids (glutamic acid, aspartic acid, asparagine, citrulline, and GABA) as compared to non-
110 pigmented rice (*Samyot et al., 2017; Kaur et al., 2018*).

111 Genetic engineering technology has substantially helped in improving cultivated crops
112 through improvement of their essential amino acid and protein content. To raise the lysine
113 content in cereals, lysine may be added at alternative codons during translation using a
114 recombinant tRNA-lysine. Recombinant production of this tRNA in transgenic rice could greatly
115 increase the amount of lysine in the seeds (*Wu and Chen, 2003*). *Liu et al. (2016)* used an
116 endosperm-specific promoter for expression of the lysine-rich protein (LRP) gene from
117 *Psophocarpus tetragonolobus* that results in 30% more lysine in the transgenic rice seeds as
118 compared to wild-type plants. Lysine biofortification in rice seeds was also achieved by over-
119 expressing two endogenous lysine-rich histone proteins (RLRH1 and RLRH2) (*Wong et al.*
120 *2015*). Moreover, it was also found that increased lysine content also increases the threonine
121 content in the grains through direct regulation (*Das et al., 2020*). The quantity of free tryptophan
122 in the seeds increased significantly as a result of expression of an equivalent feedback-insensitive
123 α -subunit of the rice anthranilate synthase, but other important agronomical variables, including
124 spikelet fertility, yield, and germination, are negatively correlated (*Wakasa et al., 2006*).

125 *Phytosterols, Carotenoids and Vitamins*

126 Plant sterols and plant stanols are collectively referred to as "phytosterols". The natural
127 phytosterol dietary intake varies from 150 to 450 mg per day, depending on a person's eating
128 habits (*Ostlund Jr, 2002*). Phytosterols have many beneficial effects such as anti-cancer activity,
129 lowering blood levels of negative lipoproteins and cholesterol absorption (*Schaefer, 2002*;
130 *Suttuarporn et al., 2015*). The three main and prevalent phytosterols found in human diet are β -
131 sitosterol, campesterol, and stigmasterol. β -sitosterol is the most common phytosterol found in
132 commercial rice cultivars, followed by campesterol, 15-avenasterol, and stigmasterol (*Zubair et*
133 *al., 2012*). Three other sterols, including fucosterol, 24-methylene-ergosta-5-en-3b-ol, and 24-
134 methylene-ergosta-7-en-3b-ol, are also present in the bran of the black rice variety "Riceberry"
135 (*Suttiarporn et al., 2015*).

136 γ -oryzanol, is another class of phytosterols and is made up of a variety of phytosteryl
137 ferulates, notably cycloartenyl ferulate, 24-methylenecycloartanyl ferulate, β -sitosteryl ferulate,
138 and campesteryl ferulate. γ -oryzanol seems to be accumulated at a faster rate in pigmented grain
139 compared to non-pigmented grain (*Chakuton et al., 2012*). Moreover, its value is found to be
140 higher in black and purple rice as compared to red rice in Thai cultivars (*Sornkhwan et al.,*
141 *2022*).

142 The essential isoprenoid phytonutrients known as carotenoids, are produced in plastids,
143 and are known to be lacking in rice endosperm. Provitamin A, found in carotenoids, are also
144 components that lower the risk of a number of diseases like cancer, heart disease, age-related
145 muscular degeneration, immune system disorders, and certain other degenerative diseases
146 (*Perera and Yen, 2007; Bollineni et al., 2014*). Lutein and zeaxanthin account for more than 90%
147 of the carotenoids synthesised in rice but trace levels of other carotenes such as lycopenes and
148 beta-carotene are also present (*Pereira-Caro et al., 2013; Melini and Acquistucci, 2017*). Most of
149 these compounds are found in the bran, with milled rice containing little or no carotenoids
150 (*Petroni et al., 2017*). The genetically variable feature of grain carotenoid concentration is highly
151 associated with grain colour. White rice have extremely little carotenoid content, but red and
152 black grains have much more (*Ashraf et al., 2017; Petroni et al., 2017*).

153 Vitamins constitute an important component of a balanced diet. Both tocopherols and
154 tocotrienols (enriched sources of vitamin E) are found in rice grains (*Zubair et al., 2012*). The
155 most prevalent tocotrienols in rice are the β - and γ -tocotrienols (*Irakli et al., 2016*). According to
156 *Gunaratne et al. (2013)*, red rice grains contain higher total levels of tocopherol and tocotrienol

157 compared to white rice varieties. The quantity of tocopherol in the grain is, however,
158 dramatically decreased by the processes of dehulling and milling (*Zubair et al., 2012*). Rice is
159 deficient in vitamins A, C and D. However, brown rice has been found to contain significant
160 amounts of vitamin B complex {thiamin (B1), riboflavin (B2), niacin (B3), pantothenic acid
161 (B5), pyridoxine (B6), biotin (B7), folate (B9)} and E (α -tocopherol) (*Juliano, 1985; Samyor et*
162 *al., 2017*). Through molecular biology and genomic techniques, tremendous progress has been
163 achieved in the genetic engineering of carotenoid production in plants during the past few
164 decades. The whole collection of carotenoid biosynthesis pathway genes and related enzymes
165 have been described. Metabolic engineering of the carotenoid biosynthesis pathway was carried
166 out by marker assisted backcross breeding of two genes- phytoene synthase (*Zmpsy1*) from *Zea*
167 *mays* and carotene desaturase (*Crtl*) from common soil bacterium *Pantoea ananatis* to create
168 second generation transgenic rice, which accumulates phytoene, a crucial provitamin A
169 intermediate (*Mallikarjuna et al., 2021; Biswas et al., 2021*). Due to their instability and
170 degradation over time in long-term storage, folates in rice grain are less available. By genetic
171 engineering a folate-binding protein, which increases the stability of folates by attaching to it
172 over prolonged storage periods, biofortified high-folate rice grains were created that had 150
173 times more folate than wild rice (*Blancquaert et al., 2015*).

174 The mutant TNG71-GE brown rice variety was found to be richer in total tocopherol and
175 tocotrienol than the wild-type. As a result, this mutant TNG71-GE rice variety could be used to
176 produce a crop with high vitamin E content (*Jeng et al., 2012*). Later, it was also found that a
177 single-point mutation of the giant embryo gene (*GE*) in Chao2-10 rice led to the development of
178 a new mutant known as Shangshida No. 5. When compared to Chao2-10, Shangshida No. 5
179 brown rice contains more total vitamin E and α -tocopherol (*Wang et al., 2013*). The overall
180 amount of tocochromanols in rice was very slightly enhanced in transgenic rice created by
181 constitutive overexpression of the *Arabidopsis thaliana* r-hydroxyphenylpyruvate dioxygenase
182 (*HPPD*) gene (*Farre et al., 2012*). The elite Japanese rice cultivar Wuyujing 3 (WY3) provides
183 transgenic brown rice with increased quantities of α -tocotrienol by both constitutive and
184 endosperm-specific over expression of the *Arabidopsis g-TMT (AtTMT)* gene (*Zhang et al.,*
185 *2013*).

186 *Flavonoids*

187 Plants contain large amounts of flavonoids, which are secondary metabolites and play a
188 significant role in plant development, pigmentation, UV protection, as well as in safeguarding
189 the interaction with microorganisms (*Samanta et al., 2011*). Coloured flavonoids (flavanols,
190 isoflavonoids, and flavones) are the pigments responsible for the colour of leaves, fruits, and
191 flowers (*Yang et al., 2022*). Flavonoids play a crucial role in floral colours and fragrance, fruit
192 pollinator attraction, and fruit dispersion (*Panche et al., 2016*). Due to the presence of
193 flavonoids, terpenoids, steroids, and alkaloids, pigmented rice exhibits cytotoxic, anti-tumor,
194 anti-inflammatory, antioxidant, and neuroprotective activities (*Goufo and Trindade, 2014*). In a
195 recent study on the total flavonoid content (TFC) of rice, it was found that black and red rice had
196 a higher TFC when compared to white rice varieties (*Chen et al., 2022*). Based on the amount of
197 aromatic compounds, flavonoids can be classified into a wide range of groups such as chalcones,
198 proanthocyanidins, anthocyanins, flavones, flavonols, flavanones, and flavanonols (*Mbanjo et*
199 *al., 2020*). However, proanthocyanidins and anthocyanins are the two primary flavonoids present
200 in pigmented rice. Anthocyanins are responsible for the purple to blue coloration of such grains
201 (*Zhang et al., 2015*). Consuming foods high in these substances may reduce inflammation and
202 lower the risk of developing type-2 diabetes, cancer, and heart disease (*Rengasamy et al., 2019*).
203 Eating foods high in anthocyanins on a regular basis also enhanced memory and overall brain
204 health (*Henriques et al., 2020*).

205 Red and white rice grains lack anthocyanin (*Xionsiyee et al., 2018*), although some red
206 and brown rice accessions have low levels (*Ghasemzadeh et al., 2018*). The enzyme anthocyanin
207 reductase converts unstable anthocyanidins into the colourless flavan-3-ols epiafzelechin,
208 epicatechin, and epigallocatechin, and when these molecules are glycosylated, a broad variety of
209 unique compounds are produced (*Kim et al., 2015*). After examining the transcription of eight
210 flavonoid biosynthesis genes in various rice cultivars, it was found that pigmented variants had
211 stronger expression of genes than non-pigmented forms (*Mbanjo et al., 2020*). At least two
212 chalcone synthetase-encoding genes *CHS2* on chromosome 7 and *CHS1* on chromosome 11,
213 help the production of flavanones in rice (*Cheng et al., 2014*). Proanthocyanidins are produced
214 by three flavone 3-hydroxylases: *F3H-1* (on chromosome 4), *F3H-2* (on chromosome 10), and
215 *F3H-3* (on chromosome 4) (*Park et al., 2016*). An effective method for researching genetic
216 variants and genetic engineering in plants is the recombinant technology (*Mackon et al., 2021*).
217 Understanding the function of several genes associated with anthocyanin biosynthesis and the

218 development of anthocyanin in the endosperm has become possible due to analysis of the
219 anthocyanin colouring mechanism in rice. The regulator of a rice prolamin gene was used to
220 insert the maize *CI/R-S* regulatory genes into the white rice japonica cultivar Hwa-Young that
221 resulted in the production of a wide range of flavonoid compounds (Mackon *et al.*, 2021).

222 *Phenolics*

223 Polyphenols constitute the most common secondary metabolites of plants and seem to be largely
224 indigenous to the plant kingdom (Dai *et al.*, 2010). They are essential for the plant's
225 development, fertilization, and defence against viruses, parasites, and environmental conditions
226 including light, cold, pollutants, and also impact the plant's colour (Kabera *et al.*, 2014). Phenolic
227 compounds benefit humans by reducing the risk of contracting chronic diseases, have a high
228 antioxidant property and make significant contributions to the prevention of many oxidative
229 stress-related diseases, including malignancy. Furthermore, there has been a lot of focus on
230 identifying and synthesising phenolic compounds or extracts from diverse plants in the realms of
231 health care and medicine (Dai *et al.*, 2010).

232 Despite differences in the content of the grains, the amount of phenolic acid in brown,
233 red, and black rice was found to be nearly the same. The different phenolic acids found in these
234 pigmented rice varieties include ferulic acid, p-cumaric, sinapic, ferulic, and hydroxybenzoic
235 acid. While ferulic, protocatechuic, and p-cumaric acids constituted the most prevalent common
236 cell wall-bound phenolic acids, sinapic, ferulic (28%) and vanillic acids constituted the most
237 significant soluble phenolic acids in black rice (Blandino *et al.*, 2022). Black rice was found to
238 possess higher protocatechuic and vanillic acids than brown rice (Zaupa *et al.*, 2015; Shao *et al.*,
239 2018). Using UV spectroscopy examination, the total phenolic content (TPC) of white, red and
240 black rice varieties was determined. White rice types had TPCs much lower than those of black
241 and red rice varieties (Chen *et al.*, 2022). The genes involved in the biosynthesis of polyphenols
242 interrelate with each other and have particular functions in the control of the polyphenols levels
243 in rice grain (Galland *et al.*, 2014). *OsCHS*, *OsCHI*, *OsF3H*, *OsF3'H*, *OsDFR*, and *OsANS* genes
244 from brown rice have the ability to change the yellow seed coat of *Arabidopsis thaliana* to
245 purplish. A yeast two-hybrid study revealed that, *OsCHS1*, *OsF3H*, *OsF3'H*, *OsDFR*, and
246 *OsANSI* interact with one another directly (Shih *et al.*, 2008). The rice mutant *Rcrd* turns red

247 when *DFR* is introduced, demonstrating that *Rd* encodes the dihydroflavonol 4-reductase
248 (*Furukawa et al., 2007*).

249 *Polysaccharides*

250 Starch is the most abundant component which constitutes approximately 90% of rice grain. It is a
251 polyglucan of two polymers, amylose (linear) and amylopectin (highly branched) with α -1 \rightarrow 4-
252 linked linear glucans and α -1 \rightarrow 6-linked branches. Starch biosynthesis is a highly regulated
253 process that requires synchronized activities between various enzymes such as starch synthase
254 (SS), ADP-glucose pyro-phosphorylase (AGPase), starch branching enzyme (SBE) and de-
255 branching enzyme (DBE). Studies have reported that starch synthesis in all higher plants and
256 green algae and it is observed in higher plants, that the enzymes have undergone multi-sequential
257 changes throughout the process of evolution (*Qu et al., 2018*). Conserved mutations in the
258 enzymes- starch synthase SSIIIa and branching enzyme IIB (BEIIB) are helpful in breeding of
259 highly resistant starch with more health benefits (*Bao, 2019*). Several efforts have been made to
260 alter the expression and activities of starch biosynthetic enzyme by using various genetic as well
261 as molecular approaches. Transgenic rice plants have been beneficial for evaluating different
262 functions of the genes responsible as rice is easily transformed and a single DNA construct can
263 be used to produce a variety of transformed lines having different expression levels.

264 *Amylose and Amylopectin content modification by alteration of single gene function*

265 The modification of amylose content has been one among the most significant breeding
266 objectives because it affects gelatinization and cooking qualities. A lower amylose content rice is
267 usually selected as the milled grains are more appetizing than those with higher content
268 (*Denardin et al., 2012*). Amylose is synthesized by the granule-bound starch synthases (GBSS)
269 (**Figure 2**). The amylose content in Japonica type rice (GBSSIb) in *ss3a* mutant was found
270 elevated when it replaced indica type rice (GBSSIa) (*Crofts et al., 2012*). The regulation of
271 *OsGBSSI* expression has been established in rice endosperm at the transcriptional as well as post
272 transcriptional levels and that accordingly differentiates these two varieties of rice-indica and
273 japonica in terms of amylose contents (*Liu et al., 2013*). The amylopectin content and its
274 structure may be directly altered by the isozymes involved in its biosynthesis and the extent of
275 such modification depends upon the isozyme's specificity. The change in amylopectin structure
276 for example alters the starch to be highly resistant for gelatinization because of more and long

277 double helices (*Miura et al., 2021*). There is also change in the X-ray diffraction patterns from
278 wild-type to mutant-type because the amylopectin has longer external chains. Similarly the
279 impact on amylopectin structure was seen when branching enzyme (*BEIIb*) activity in japonica
280 rice was down regulated by the RNAi approach (*Tsuiki et al., 2016; Zhang et al., 2022*). The
281 increase in amylopectin longer chains leads to an increase in the resistant starch content rather
282 than an increase in amylose content (*Tsuiki et al., 2016*).

283 *Modification of Amylose and Amylopectin Content by Alteration in Multiple Gene Functions*

284 Multiple gene functions leads to intense modifications in the starch biosynthesis and thus, the
285 phenotypes of these mutants exhibit variations. The changes are more extreme than predicted by
286 the single gene mutants and the plant usually doesn't survive. For example, the triple mutants
287 (*ss1/ss2a/ss3a*) of japonica rice type referred as null mutants becomes sterile which indicates the
288 functional properties of each isozyme and importance of the presence of at least one of these
289 (*SSI, SSIIa, and SSIIIa*) for starch biosynthesis in rice endosperm (*Fujita et al., 2011*). Other
290 indirect effects are also observed such as the ADP glucose concentrations were found enhanced
291 in the *ss3a* mutant as well as in *ss3a/ss4b* mutants when compared with wild type causing an
292 increase in the Amylose content (*Fujita et al., 2007*). Similarly, the *BEIIb* and *BEI* gene
293 inhibition results in the accumulation of amylose which leads to the different physico-chemical
294 properties in the rice endosperm providing highly resistant starch. DNA editing with
295 CRISPR/Cas9 was found to be beneficial for specific editing of the *BEIIb* gene in rice for
296 improved starch structure (*Baysal et al., 2016*).

297 *Iron, Zinc and Micronutrients*

298 Rice grain has trace amounts of several vital micronutrients, including zinc (Zn), magnesium
299 (Mg), iron (Fe), copper (Cu), potassium (K), manganese (Mn) and calcium (Ca) (*Shao et al.,*
300 *2018*). However, the coloured rice possess greater amounts of Zn, Fe, and Mn when compared
301 with white rice (*Hurtada et al., 2018; Shao et al., 2018*). Brown rice can deliver up to 75% of the
302 optimal regular intake of zinc, copper, and iron while white rice only provides 37% (*Hashmi and*
303 *Tianlin, 2016*). Red rice from North-East India has been observed to contain highest content of
304 micronutrients (Al, K, Zn and Ca) than white rice (*Samyov et al., 2016*). A recent survey found
305 that two black rice variants from Korea, Heukjinjubyeo and Heukgwangbyeo possess greater
306 levels of Ca and K in comparison to a white variety (*Hiemori et al., 2009*). It has been also

307 observed that the ratio of micronutrients between coloured and non-coloured rice also differs.
308 The important nutrients in polished rice samples were found to be in the order of
309 $K > Na > Mg > Zn > Fe > Mn > Cu > Cr$, while in contrary to Brown rice variants, the order is:
310 $K > Mg > Na > Mn > Zn > Fe > Cr > Cu$ (Hashmi and Tianlin, 2016). A survey was done in North East
311 India about the mineral content in black, red and white rice varieties and a complementary
312 investigation was conducted between mineral elements (Zn, Fe, Ca, Ni and Mn). The mineral
313 elements like Fe, Ca, Ni, Mn, Zn in various varieties of pigmented rice were present in highest
314 quantity than the white rice varieties (Singh et al., 2022). The growth conditions also affect the
315 Zn and Fe concentrations in the purple rice varieties. Wetland conditions were found to be more
316 advantageous than aerobic culture for producing purple rice with vivid coloration and greater
317 Zn (30 mg kg⁻¹) and Fe contents (15 mg kg⁻¹) (Jaksomsak et al., 2021).

318

319 *β-carotene*

320 Rice endosperm is not able to synthesize beta-carotene which acts as a precursor for vitamin A
321 synthesis leading to deficiency in vitamin A especially in the developing countries where rice is
322 the staple food and is the only way for fulfilling this nutritional requirement (Das et al., 2020).
323 Vitamin A or β -carotene is a most important micronutrient for proper vision and development in
324 humans and prevents many diseases such as night blindness, xerophthalmia and keratomalacia
325 (Klemm et al., 2010). As rice endosperm is lacking β -carotene so it was necessary to make rice
326 more nutritious by making it able to synthesize β -carotene (Das et al., 2020). In this context, an
327 experiment was designed by Burkhardt et al. (1997) in which they found, in rice endosperm,
328 geranyl geranyl diphosphate (GGPP), an important precursor for β -carotene biosynthesis making
329 rice endosperm capable of β -carotene synthesis. The Rice Taipei 309 (japonica variety) was
330 transformed with phytoene synthase from *Narcissus pseudonarcissus* by using micro projectile
331 bombardment leading to β -carotene synthesis (Burkhardt et al., 1997). In another study the β -
332 carotene content of Golden rice was enhanced ~23-fold by replacing the daffodil psy gene with
333 its homolog from maize and was named as Golden rice 2 (GR2) (Mallikarjuna et al., 2021)
334 which is now a good source of vitamin A and might become a part of many breeding programs in
335 Asia. Furthermore, β -carotene can be converted into the derivative astaxanthin which is keto-
336 carotenoid and red in color with high antioxidant activity but most higher plants are unable to
337 produce astaxanthin (Ha et al., 2019). Researchers genetically engineered *sPaCrtI* (phytoene

338 desaturase), *sZmPSYI* (phytoene synthase), *sHpBHY* (b-carotene hydroxylase), and *sCrBKT* (b-
339 carotene ketolase) genes to initiate the astaxanthin biosynthetic pathway to produce endospermic
340 astaxanthin in rice grains (Zhu et al., 2018). Tian et al. (2019) reported that bioengineering of
341 three chemically synthesized genes i.e., *tHMG1*, *ZmPSYI*, and *PaCRTI* in rice increased the
342 endospermic carotenoid biosynthesis through the mevalonate route. This engineering of
343 astaxanthin biosynthesis in rice endosperm converts Golden rice to aSTARice (Zhu et al., 2018).
344 These improved rice genotypes contain one more gene for β -carotene hydroxylase that produces
345 a red coloured Astaxanthin Rice (aSTARice). Therefore, biofortification of rice through
346 metabolic engineering could prove rice as a health promoting food and can be processed to
347 produce dietary supplements (Zhu et al., 2018).

348 *Folate*

349 Folates are a group of water-soluble B vitamins (B9), derived from most reduced folate form
350 known as tetrahydrofolate (THF) contains three building blocks—the pteridine, *p*-aminobenzoate
351 (*p*-ABA) and glutamate moieties (Re'beille et al., 2006). Living organisms use folates as C1
352 donors and acceptors and are mainly involved in the biosynthesis and metabolism of nucleotides,
353 amino acids and vitamin B5 (Blancquaert et al., 2010). However, only plants and micro-
354 organisms can synthesize THF and its derivatives by de novo pathways. Therefore, humans are
355 dependent on food to meet their daily requirement of folates needed to regulate many
356 physiological and molecular processes (Blancquaert et al., 2014) for their survival. Many plants
357 like vegetables, pulses and fruits are loaded with folates but most staple crops such as rice which
358 is consumed by 1/2 of world's population; contain low levels of folate leads to folate deficiency
359 especially in developing countries (Dong et al., 2014). To eradicate folate deficiency worldwide
360 by biofortification of rice through metabolic engineering is a promising and cost-effective
361 approach. Moreover, the concept of enhancing folate content by over expressing the folate
362 biosynthesis genes/ metabolic engineering has been carried out in rice (Strobbe and Van Der
363 Straeten, 2017) and is an interesting target for improvement (Re'beille et al., 2006). The folate
364 biosynthesis pathway in plants is a multi-step process occurs in three different subcellular
365 compartments involves the conversion of chorismate (2 step process) by the action of ADC
366 synthase (ADCS) into *p*-aminobenzoate (*p*ABA) in plastids. Genes involved in folate
367 biosynthesis such as *ADCS*, *GTPCHI*, *FPGS* and folate binding proteins (*FBP*) originated from
368 different organisms has been genetically engineered and over expressed in rice to produce higher

369 content of folates up to 100 fold (*Malik and Maqbool., 2020*) but further effective biofortification
370 strategy is needed. The lists of genes engineered in rice as a single or in combination by different
371 researchers to increase content of folates till now are mentioned in **Table 1**.

372 **Pigmentation in Rice**

373 Pigmented rice grains contain high levels of flavonoids, which are biosynthesized by two genes
374 (*CHS1* and *CHS2*) encoding chalcone synthetase located on chromosomes 11 and 7, respectively
375 (*Cheng et al., 2014; Lei et al., 2009*). Similarly, three flavone 3-hydroxylases, *F3H1*, *F3H2*, and
376 *F3H3*, contribute to proanthocyanidin production in red rice grains (*Park et al., 2016*) and the
377 two anthocyanin synthases are important for anthocyanins synthesis, *ANS1* and *ANS2* (*Shih et*
378 *al., 2008*).

379 *Red and White Pigmentation of Rice grain*

380 Rice grain colour was a major target during domestication and white rice was mostly selected,
381 while most wild type rice is red. The colour is determined by the functional activities of different
382 transcription factors. Two complementary genes, *Rc* and *Rd* (located on chromosomes 7 and 1,
383 respectively), encode a basic helix-loop-helix (bHLH) transcription factor and are responsible for
384 the red pericarp. *RcRd* genotypes produce red rice grain, while *Rc**rd* genotypes produce brown
385 rice grain (*Furukawa et al., 2007*). The *Rc* gene is also involved in rice grain dormancy and
386 shattering. White variants have a loss-of-function mutation in the *Rc* allele (*Gross et al., 2010*).

387 *Purple Rice Pigmentation*

388 The purple-pericarp formation in rice is determined by the gene *Kala4/OsB2/Pb* which produces
389 anthocyanins (*Rahman et al., 2013*). Pigmentation variation is under polygenic control (*Ham et*
390 *al., 2015*). The *Pl* locus on chromosome 4 has three alleles (*Plw*, *Pli*, and *Plj*), each responsible
391 for a different type of pigmentation. The wild type (*Plw*) produces anthocyanin in the aerial parts
392 of the rice plant. *Pl* locus possesses the two genes- *OSB-1&2* encodes a helix loop helix
393 transcription factor (*Sakamoto et al., 2001*). Purple colour is characterized by the two dominant
394 genes *Pb* and *Pp* (*Ham et al., 2015*). A comparative genomics study in different rice lines by
395 *Lachagari et al. (2019)* revealed important allelic variants in genes related to flavonoid
396 synthesis, cytokinin glucosides, and betanidin degradation, and purple rice pigmentation. *Black*
397 *Rice Pigmentation*

398 The *Kala1*, *Kala3*, and *Kala4* loci located on chromosomes 1, 3, and 4 express black pericarp
399 traits in rice (Maeda et al., 2014). The pigments, mostly observed in the aleurone layers of black
400 rice, are a mixture of anthocyanins and range from black to dark purple. Variations in the *Kala4*
401 promoter sequence mostly result in black rice grain phenotypes. *Kala4* encodes a Helix Loop
402 Helix transcription factor which relates to *OSB2* gene responsible for the synthesis of
403 anthocyanins (Sakulsingharoj et al., 2014). Genetic studies, including QTL mapping and GWAS
404 analysis, have been used to understand the cause of rice pigmentation. Nine QTLs were reported
405 for flour pigmentation in an inbred line population (Tan et al., 2001), and four QTLs were
406 observed for red pigmentation (Dong et al., 2008). Twenty-one QTLs for variations in the
407 composition and content of proanthocyanidins and anthocyanins were identified in a study by Xu
408 et al. (2017). Twenty-five marker trait associations for grain pigmentation were identified using
409 GWAS (Shao et al., 2011). GWAS has been more useful than QTL mapping in determining the
410 cause of rice pigmentation (Korte and Farlow, 2013). With the advance in GWAS, a total of 763
411 SNPs associated with pericarp pigmentation were reported by Yang et al. (2018) and some
412 specific SNPs were also identified associated with Rc (Butardo et al., 2017).

413 **Marker Trait Association Studies**

414 Quantitative trait locus (QTL) is a part of DNA that affects quantitative trait and QTL mapping is
415 a powerful and effective approach to analyse the chromosomal regions controlling quantitative
416 traits for the marker-assisted selection (MAS) strategy in rice (Hu et al., 2021; Islam et al.,
417 2020). Moreover, nutrient biofortification of rice by this method has been proved as a sustainable
418 strategy to overcome mineral deficiencies (Majumder et al., 2019). Mineral accumulation in
419 grain being a complex process and highly influenced by environmental factors has made
420 breeding and early-generation phenotypic-based selections of biofortified rice varieties slow and
421 less effective (Sharma et al., 2020). However, mapping major-effect QTLs by understanding
422 genetics of grain mineral elements at the molecular level would be helpful for the rapid
423 development of nutrient biofortification of rice varieties using marker-assisted breeding (MAB)
424 (Swamy et al., 2018). To date, different mapping populations derived from biparental inter- or
425 intra-subspecific and interspecific crosses have been used for identification of a large number of
426 QTLs in rice genome which are mainly associated with mineral contents (Wang et al., 2020) and
427 would be helpful in nutrient biofortification of rice varieties (Hu et al., 2021). In milled rice, 20
428 QTLs has been identified for P, K, Mg, Ca, Zn, Mn, and Cu contents (Yu et al., 2015) and in

429 another study 51 QTLs in brown rice and 61 QTLs in rice straw was identified for different
430 minerals (*Wang et al., 2020*) by using the RILs from the intra-subspecific cross Zhenshan
431 97/Miyang 46. *Garcia-Oliveira et al. (2009)* used introgression lines (ILs) from an interspecific
432 cross of cultivar “Teqing” and Yunnan wild rice (*O. rufipogon*) and reported 31 QTLs in brown
433 rice for P, K, Mg, Ca, Fe, Zn, Mn, and Cu contents. Some other researchers found 134 QTLs in
434 brown rice for 16 minerals content by using both RILs and backcross introgression lines (BILs)
435 from an inter-subspecific cross Lemont/Teqing (*Zhang et al., 2014*). *Descalsota-Empleo et al.*
436 *(2019)* used two sets of doubled haploid (DH) lines from two inter-subspecific crosses
437 IR64/IR69428 and BR29/IR75862 and reported 50 QTLs in milled rice for 13 minerals (P, K,
438 Na, Mg, and Ca). Furthermore, *Du et al. (2013)* showed influence of environmental factors in the
439 detection of QTL for grain mineral contents. In this study they selected brown rice grown in two
440 different ecological environments namely Lingshui and Hangzhou and mapped 23 and 9 QTLs
441 for seven mineral contents respectively and reported only two QTLs for the Mg content were
442 found in both the environments simultaneously. Several other reports based on QTLs for mineral
443 accumulation in rices have been detailed in **Table 2**.

444 **Conclusion and Future Prospectus**

445 Population growth and adverse global climatic changes negatively affect our food and
446 nutritional securities which has resulted in hunger and malnutrition in our society. Rice
447 being a staple food for half of the world can be a source of energy for our generations
448 only if it is fortified with balanced nutrients. Pigmented rice as a food contain many bioactive
449 compounds that display significant potential concerning a range of beneficial health effects
450 like anti-cancerous, anti-allergic, anti-aging, anti-diabetic, and anti-obesity properties and
451 include many medicinal properties like treating ulcer, fracture, burns, skin lesions, and many
452 more. The extent of diseases faced today may be reduced significantly by simply replacing
453 white rice with pigmented rice in our day to day diet. The present limitations of low
454 productivity and palatability in pigmented rice can be solved by framing efficient breeding
455 strategies along with use of multiomics approaches. Development of highly palatable and high
456 yielding coloured rices will have a great impact in tackling various malnutrition concerns
457 observed in rice eating countries and this can have great implications in attaining nutritional
458 security.

459

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471 Authors declare no potential conflict of interest for this article.

472 **Author contributions**

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475 proof read the manuscript.
- 476 • Uneeb Urwat: performed the experiment, contributed in preparing first draft of the article,
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- 478 • Asmat Farooq: performed the experiment, contributed in preparing first draft, analysed the
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- 480 • Mohammad Maqbool Pakhtoon: performed the experiment, contributed in preparing first
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- 486 • Aneesa Batool: performed the experiment, contributed in preparing first draft of manuscript
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842 **Figure legends**

843 **Figure 1.** The importance of pigmented rice in India with respect to the (a) varieties cultivated in
844 different parts; (b) elemental content in different coloured-rice and (c) different nutraceutical
845 benefits of pigmented rice.

846 **Figure 2.** Genes Regulating Starch Biosynthesis in the endosperm through various
847 transcriptional factors, different abbreviations indicates amylose content (AC), eating & cooking
848 quality (ECQ), nutritional quality (NQ) and appearance quality(AQ), granule-bound starch
849 synthase (GBSS), soluble starch synthase (SS) and starch branching enzymes (SBE).

850 **Table legends**

851 **Table 1.** Genes engineered in rice seed for different nutrients and metabolites

852 **Table 2.** QTL studies showing different markers trait associations for nutrient enhancement in
853 rice

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Table 1 (on next page)

Table 1

Genes engineered in rice seed for different nutrients and metabolites

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2 **Table 1.** Genes engineered in rice seed for different nutrients and metabolites

Nutrient/ Metabolite	Gene Origin	Engineering Approach	Genes Transformed	Expression Details/ Fold change in expression	References
Folate	<i>Arabidopsis thaliana</i>	Biosynthesis Single-gene	ADCS	(1/6)-fold upregulated	<i>Strobbe and Van DerStraeten., 2017</i>
	<i>Arabidopsis thaliana</i>	Biosynthesis Single	GTPCHI	6.1-fold	
	<i>Arabidopsis thaliana</i>	Biosynthesis Single	HPPK/DHPS	1.4-fold	
	<i>Triticum aestivum</i>	Biosynthesis Single	DHFS	1.27-fold	
	<i>Arabidopsis thaliana</i>	Biosynthesis Multi-gene	GTPCHI + other biosynthesis genes	6.1-fold	
	<i>Arabidopsis thaliana</i>	Biosynthesis Multi-gene	GTPCHI + ADCS	100-fold	
	<i>Arabidopsis thaliana</i>	Polyglutamylolation	FPGS	1.45-fold	
	<i>Oryza sativa</i>	Polyglutamylolation	FPGS	4.7-fold	
	<i>Arabidopsis thaliana</i>	Polyglutamylolation	GTPCHI + ADCS + FPGS	100-fold	
	<i>Bos taurus</i>	Folate binding proteins	FBP	6.2-fold	
	<i>Rattus norvegicus</i>	Folate binding proteins	GNMT	8.8-fold	
Arabidopsis thaliana (G + A) Bostaurus (FBP)	Folate binding proteins	GTPCHI + ADCS + FBP	150-fold		
Lysine	<i>Arabidopsis thaliana</i>	Manipulation of lysine content	DHDPS	Feedback inhibition	<i>Das et al. 2020</i>

Carotenoids	<i>Zea mays</i>	Agrobacterium mediated DNA transfer	<i>OsLCYB</i>	1.9-fold upregulated	<i>Tian et al. 2019</i>
Carotenoids	<i>Pantoea ananatis</i>	Agrobacterium mediated DNA transfer	<i>OsBCH2</i>	1.1-fold upregulated	<i>Tian et al. 2019</i>
	<i>Saccharomyces cerevisiae</i>	Agrobacterium mediated DNA transfer	<i>OsPDS</i>	1.6-fold upregulated	<i>Tian et al. 2019</i>
Vitamin A	<i>Narcissus pseudonarcissus</i>	Transformation by microprojectile bombardment	<i>pCPsyH</i>	Expression observed in only the transformed plants.	<i>Burkhardt et al. 1997</i>
Vitamin E	<i>Oryza sativa</i>	Single point mutation	Giant embryo gene (ge)	2.2-fold upregulated	<i>Wang et al. 2013</i>
	<i>Arabidopsis thaliana</i>	Transformation by particle bombardment	<i>PDS1</i>	upregulated	<i>Farre et al. 2012</i>
Starch	<i>Solanum tuberosum</i>	Agrobacterium-mediated transformation	StGWD1	9-fold higher 6-phospho (6-P) monoesters and double amounts of 3-phospho (3-P) monoesters.	<i>Chen et al. 2017</i>
	<i>Zea mays</i>	Agrobacterium-mediated transformation	OsSUS1-6	Upregulated starch accumulation for improved grain filling.	<i>Fan et al. 2019</i>

	<i>Oryza sativa</i>	Map-based cloning- Mutation of T- DNA(gamma- radiated hybrid-rice)	SSIII and Waxy (Wx)	High in resistant starch (RS)	<i>Zhou et al. 2016</i>
	<i>Arabidopsis thaliana</i>	RNA interference approach	Starch Excess 4 (SEX4)	improved bioethanol yield, with a 50% increase in ethanol production	<i>Huang et al. 2020</i>
	<i>Oryza sativa</i>	RNA interference approach (antisenseWx gene)	Wx	amylose content in transgenic caryopsis was downregulated	<i>Chen et al. 2006;</i> <i>Khandagale et al.</i> <i>2018</i>
	<i>Thermoan aerobacter ethanolicus</i>	Agrobacterium tumefaciens mediated transfer	APU (amylopullulanase)	Reduction of amylose, altered starch properties	<i>Chiang et al. 2005</i>
Nitrogen	<i>Arabidopsis thaliana</i>	Constitutive overexpression	MYB12, MYC, WD40	Upregulated	<i>Lee et al. 2016</i>
Phosphorus	<i>Arabidopsis thaliana</i>	Constitutive overexpression	OsMYB3R-2	Upregulated	<i>Huang et al. 2018</i>

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Table 2 (on next page)

Table 2

QTL studies showing different markers trait associations for nutrient enhancement in rice

1 **Table 2.** QTL studies showing different markers trait associations for nutrient enhancement in rice

Grain Traits	QTL	Trait marker	References
Zinc (Zn)	<i>qZN-5, qZN-7, qSZn2, qSZn12, qZn7, qZn3.1, qZn7.1, qZn7.2, qZn7.3, qZn12.1, qZn4, qZn6, qZn1-1, qZn12-1, qZn5-1, qZn8-1, qZn12-1, qZn2.1, qZn2.1, qZn3.1, qZn6.1, qZn6.2, qZn8.1, qZn11.1, qZn12.1, qZn12.2, qZn2.2, qZn8.3, qZn12.3, qZn3.1, qZn7, qZn8.3, qZn3-1, qZn1.1, qZn5.1, qZn9.1, qZn12.1, qZn1.1, qZn6.1, qZn6.2, qZn2-1, qZn2-2, qZn5, qZn10, qZn2.1, qZn3.1, qZn5.1, qZn5.2, qZn7.1, qZn9.1, qZn11.1, qZn 1.1, qZn2.1, qZn3.1, qZn3.2, qZn5.1, qZn6.1, qZn8.1, qZn8.2, qZn9.1, qZn12.1, QTL.Zn.4, QTL.Zn.5</i>	RMID2009463, RM2147095, RM2785595, RM6047367, RMID6006214, RM8832534, RMID11000778, RM12985052, RM13057679, RM34-RM237, RM7-RM517, RZ398-RM204 RM501-OsZip2, RM152, RM25-R1629, RM235-RM17, RM260-RM7102, RM551, RM413	<i>Das et al. 2020; Islam et al. 2020; Thangadurai et al. 2020; Swamy et al. 2018</i>
Iron (Fe)	<i>qFE-1, qFE-9, qGFe4, qSFe1, qSFe12, qFe1, qFe3, qFe6, qFe2-1, qFe9-1, qFe4.1, qFe3.3, qFe7.3, Fe8.1, qFe12.2, qFe3-1, qFe9.1, qFe12.1, qFe1.1, qFe1.2, qFe6.1, qFe6.2, qFe5, qK6.1, qFe2.2, qFe3.1, qFe4.1., qFe6.1, qFe8.1, qFe11.2, qFe11.3, qFe12.1,</i>	RM4743351, RM574-RM122, RM234-RM248, RM137-RM325A, RZ536-TEL3, RM270-RM17, RM260-RM7102, RM17-RM260, RM452, RM215	<i>Das et al. 2020; Islam et al. 2020; Thangadurai et al. 2020; Swamy et al. 2018</i>
Manganese (Mn)	<i>qMn1-1, qMn2-1, qMn3-1, qMn10-1, qMn2.1, qMn2.1, qMn7.1, qMn1.1, qMn1.2, qMn3.1, qMn3.2, qMn4.1</i>	RM243-RM312, RM6367, RM227-R1925, RM214, RMID2009186, RMID2009463, RM7592793	<i>Das et al. 2020; Mahender et al. 2016; Swamy et al. 2018</i>
Calcium	<i>qCa1-1, qCa4-1, qCa5-1, qCa9-1, qCa10-1, qCa11-1, qCa12-1, qCa1.1, qCa1.1, qCa2.1, qCa2.1, qCa3.1, qCa3.2,</i>	RM403585, RMID1013855, RMID2009186, RM2131264, RM2499734, RM2733626, RM598, RM5626-RM16, RM200-RM227,	<i>Das et al. 2020; Mahender et al. 2016; Swamy et al. 2018</i>

Magnesium (Mg)	<i>qMg1-1, qMg3-1, qMg5-1, qMg9-1, qMg12-1, qMg3.1, qMg3.2, qMg5.1, qMg8.1, qMg9.1, qMg1.1, qMg7.1, qMg8.1, qMg11.1</i>	RM2499734, RM3460782, RM5522491, RM8892951, RM9886119, OSR 21, RM467, RM332,	<i>Das et al. 2020; Mahender et al. 2016; Swamy et al. 2018</i>
Phosphorus (P)	<i>qP1-1, qP3-1, qP8-1, qP9-1, qP12-1, qP1.1, qP2.1, qP2.2, qP5.1, qP6.1, qP11.1, qP11.2</i>	RM,119519, RMID2009186, RM2181296, RM5430212, RMID6009257, RM3411, RM495, RM212, RM70-RM172, RM201,	<i>Das et al. 202; Mahender et al. 2016; Swamy et al. 2018</i>
Potassium (K)	<i>qK1-1 qK.1, qK4-1, qK8-1, qK9-1, qK2.1, qK4.1, qK4.2, qK5.1, qK9.1, qK3.1, qK3.2, qK3.3, qK4.1, qK5.1,</i>	RM2094246, RM4285667, RM4668476, RM5430212, RM9858839, RM3572, RM5501,	<i>Das et al. 2020; Mahender et al. 2016; Swamy et al. 2018</i>
Boron (B)	<i>qB2.1, qB3.1, qB4.1, qB4.2, qB10.1</i>	RMid2009186, RM2645329, RM4314701,	<i>Das et al. 2020; Swamy et al. 2018</i>
Cobalt (Co)	<i>qCo1.1, qCo3.1, qCo4.1, qCo12.1, qCo7.1, qCo10.1</i>	RM827062, RM2785595, RM4572241, RM12958034	<i>Das et al. 2020; Swamy et al. 2018</i>
Copper (Cu)	<i>qCu3.1, qCu4.1, qCu4.2, qCu1.1, qCu1.2, qCu2.1, qCu6.1, qCu8.1</i>	RM3330180, RM4314701, RM4761773	<i>Das et al. 2020; Swamy et al. 2018</i>
Molybdenum (Mo)	<i>qMo1.1, qMo1.2, qMo1.3, qMo2.1, qMo11.1, qMo12.1, qMo12.1, qMo12.1</i>	RM854218, RMID1014853, RM1191519, RM1725183, RMID11006537, RM13030749, RM13044018	<i>Das et al. 2020; Swamy et al. 2018</i>
Sodium (Na)	<i>qNa1.1, qNa1.2, qNa7.1, qNa7.2, qNa10.1, qNa3.1, qNa11.1, qNa11.2</i>	RM267954, RM784044, RMID7003294, RM7962882, RM10635878	<i>Das et al. 2020; Swamy et al. 2018</i>

Phytic acid (PA)	<i>qPA.12</i>	RM247-RM179	<i>Thangadurai et al. 2020</i>
Grain protein content (GPC)	<i>qPC1, qPC2, qPC3, qPC6.1, qPC6.2, qPC8, qPC12.1, qPC1.1, qPC11.1, and qPC11.2, qPC-3, qPC-4, qPC-5, qPC-6 and qPC-10, qPr1 and qPr7, qPro-8, qPro-9 and qPro-10, qGPC1.1, qSGPC2.1 and qSGPC7.1, QTL.pro.1</i>	RM493-RM562, 1008-RM575, RM472-RM104, RM5619-RM1211, RM12532-RM555, RM251-RM282, RM190-RZ516, RM190-RZ516, RM270-C751, R1245-RM234, RM445-RM418, RM184-RM3229B, RM24934-RM25128, 1027-RM287, RM287-RM26755, RM5	<i>Thangadurai et al. 2020; Islam et al. 2020; Mahender et al. 2016</i>
Amino acid content (AAC)	<i>qAa1, qAa7, qAA.8, qAA.4, qAA.3, qAA.2, qAA.1, qAa9, qAA.10</i>	RM493-RM562, RM472-RM104, RM324-RM301, RM322-RM521, RM520-RM468, RM348-RM131, RM125-RM214, RM137-RM556, RM447-RM458, RM328-RM107, RM467-RM271	<i>Thangadurai et al. 2020; Mahender et al. 2016</i>
Amylose (amy)	<i>QTL.amy.6, QTL.amy.7, QTL.amy.8, QTL.amy.11</i>	RM190, RM125, RM284, RM144	<i>Islam et al. 2020</i>

Figure 1

Figure 1

The importance of pigmented rice in India with respect to the (a) varieties cultivated in different parts; (b) elemental content in different coloured-rice and (c) different nutraceutical benefits of pigmented rice.

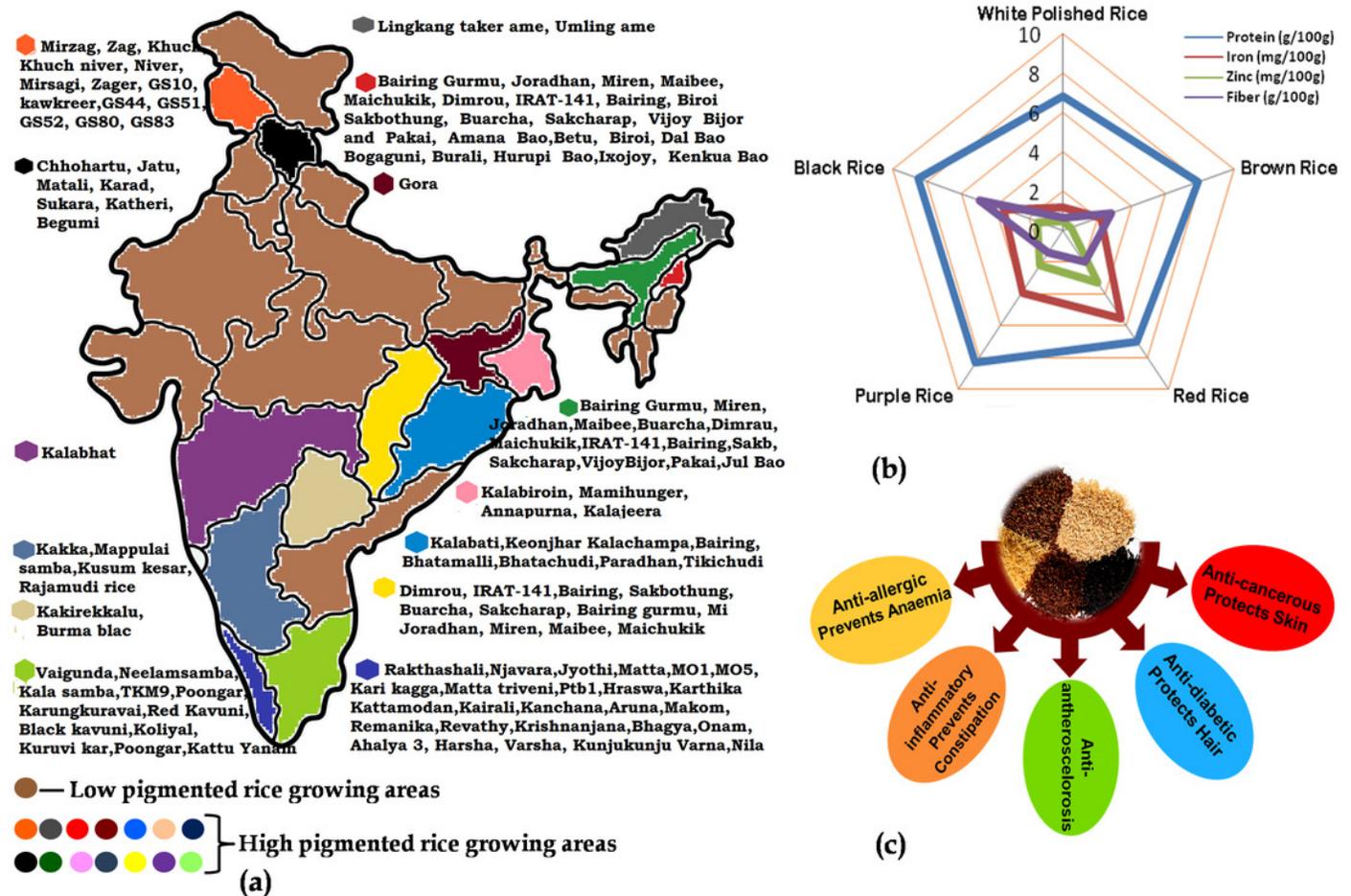


Figure 2

Figure 2

Genes Regulating Starch Biosynthesis in the endosperm through various transcriptional factors where, different abbreviations indicates amylose content (AC), eating & cooking quality (ECQ), nutritional quality (NQ) and appearance quality(AQ), granule-bound starch synthase (GBSS), soluble starch synthase (SS) and starch branching enzymes (SBE).

