



Effect of ethyl methanesulfonate mediated mutation for enhancing morpho-physio-biochemical and yield contributing traits of fragrant rice

Areeqa Shamshad¹, Muhammad Rashid¹, Ljupcho Jankuloski², Kamran Ashraf^{3,4}, Khawar Sultan⁵, Saud Alamri⁶, Manzer H. Siddiqui⁶, Tehzeem Munir⁵ and Qamar uz Zaman⁵

¹Nuclear Institute for Agriculture and Biology College (NIAB-C), PIEAS, Islamabad, Pakistan

²International Atomic Energy Agency, Joint FAO/IAEA Centre, Plant Breeding and Genetics Section, Vienna, Austria

³Department of Bioengineering and Biotechnology, School of Biotechnology, Kunming University of Science and Technology, Shanghai, China

⁴Department of Food Sciences, Government College University Faisalabad, Sahiwal Campus, Faisalabad, Pakistan

⁵Department of Environmental Sciences, The University of Lahore, Lahore, Pakistan

⁶Department of Botany and Microbiology, College of Science, King Saud University, Riyadh, Saudi Arabia

ABSTRACT

Background. Chemical mutagenesis has been successfully used for increasing genetic diversity in crop plants. More than 800 novel mutant types of rice (*Oryza sativa* L.) have been developed through the successful application of numerous mutagenic agents. Among a wide variety of chemical mutagens, ethyl-methane-sulfonate (EMS) is the alkylating agent that is most commonly employed in crop plants because it frequently induces nucleotide substitutions as detected in numerous genomes.

Methods. In this study, seeds of the widely consumed Basmati rice variety (Super Basmati, *Oryza sativa* L.) were treated with EMS at concentrations of 0.25%, 0.50%, 0.75%, 1.0%, and 1.25% to broaden its narrow genetic base.

Results. Sensitivity to a chemical mutagen such as ethyl methanesulfonate (EMS) was determined in the M1 generation. Results in M1 generation revealed that as the levels of applied EMS increased, there was a significant reduction in the germination percent, root length, shoot length, plant height, productive tillers, panicle length, sterile spikelet, total spikelet, and fertility percent as compared to the control under field conditions. All the aforementioned parameters decreased but there was an increase in EMS mutagens in an approximately linear fashion. Furthermore, there was no germination at 1.25% of EMS treatment for seed germination. A 50% germination was recorded between 0.50% and 0.75% EMS treatments. After germination, the subsequent parameters, viz. root length and shoot length had LD_{50} between 0.50% and 0.75% EMS dose levels. Significant variation was noticed in the photosynthetic and water related attributes of fragrant rice. The linear increase in the enzymatic attributes was noticed by the EMS mediated treatments. After the establishment of the plants in the M1 generation in the field, it was observed that LD_{50} for fertility percentage was at EMS 1.0% level, for the rice variety.

Submitted 24 February 2023

Accepted 10 July 2023

Published 26 September 2023

Corresponding author

Qamar uz Zaman,
qamar.zaman1@envs.uol.edu.pk

Academic editor

Bernardo Franco

Additional Information and
Declarations can be found on
page 17

DOI 10.7717/peerj.15821

© Copyright
2023 Shamshad et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

Conclusion. Hence, it is concluded that for creating genetic variability in the rice variety (Super Basmati), EMS doses from 0.5% to 0.75% are the most efficient, and effective.

Subjects Agricultural Science, Biotechnology, Plant Science

Keywords Lethal dose, Ethyl methanesulfonate, Rice, Mutation, Germination, Productivity

INTRODUCTION

Nearly half of the world population relies mostly on the rice crop as a primary staple food. An agro-ecological landscape and the associated biodiversity and customer quality choices have also been major contributors to the development of new rice varieties which is leading to more genetic diversification and several varietal groupings (*Loko et al., 2021*). To ensure food security, it is essential to increase production by utilizing effective techniques for the efficient enhancement of yield (*Zaghum, Ali & Teng, 2022*). The aromatic local cuisine rice (*i.e.*, pulao or biryani) from the Indian subcontinent known as “Basmati” is made up of one such varietal group and is quite expensive both domestically and abroad. The extra-long, narrow grain, pleasant aroma, and fluffy soft textured Basmati rice variety its origins in the Himalayan foothills which provide the most suitable environment for its growth and its distinguishing features (*Hameed et al., 2019; Malabadi, Kolkar & Chalannavar, 2022*). Besides other breeding tools, irradiation (*i.e.*, fast neutrons, γ -rays, & X-rays) and chemical mutagens (*i.e.*, DEB, EMS, & sodium azide) have been frequently used to produce a broad range of functional mutations in rice (*Gulfishan et al., 2023*). Phenotypic characterization of Super Basmati using ethyl methanesulfonate (EMS) was carried out by earlier researchers (*Hameed et al., 2019*). For functional genomics and breeding studies, a large mutant population of coarse variety Katy was developed using EMS (*Jia et al., 2019*). In earlier studies, the upland rice variety Nagina 22, and the Japonica variety Shengdao 808 had also been used for developing mutants exhibiting tolerance to drought and salinity and natural variation studies (*Shang, Chun & Li, 2021; Zargar et al., 2022*).

Mutagens cause point mutations, making them suitable for creating missense and nonsense mutations that would result in functional mutations (*Shelar et al., 2021*). Ionizing radiation also causes chromosomal rearrangements and deletions (*Le Roux, 2019; Singh, Khar & Verma, 2021*). Mustard gas, methyl-methanesulfonate (MMS), EMS, and nitrosoguanidine are all alkylating agents that have diverse effects on DNA (*Ramesh et al., 2019*). According to *Talebi, Talebi & Shahrokhifar (2012)*, EMS produces mutations by alkylating guanine bases, which results in (mis)matches with thymine rather than cytosine and triggers transitions from G/C to A/T. EMS can also lead to A/T to G/C conversions through mismatches of 3-ethyladenine or G/C to T/A transversions by 7-ethylguanine hydrolysis (*Serrat et al., 2014*). The EMS causes point mutations in the rice genome and is one of the most commonly used mutagens in plants due to its potency and ease of application (*Upadhyaya et al., 2007; Husain, Bano & Khan, 2020*).

Since EMS induces an abundance of non-lethal point mutations (genome-wide), a slight mutant population (roughly ten thousand) is abundant to saturate the genome with

mutations (*Hernández-Muñoz et al., 2019*). The point mutation rate is four mutations per Mb in Arabidopsis (*Kazama et al., 2017*). A significant benefit of using a mutagen like EMS in forward genetic screens depends upon its efficacy in a range of organism types (*Taheri et al., 2017*). In different species, chemical mutagenesis induces a different rate of nucleotide substitutions. In Arabidopsis and maize mutational density was observed per gene (*Talebi, Talebi & Shahrokhifar, 2012*). EMS was also used in other crops like sugarcane where the calli were mutagenized with 0.5% EMS and exposed to 2% (w/v) PEG-6000 for induction of the osmotic stress (*Gadakh et al., 2021*). The mutant, dmc, was obtained from EMS treatment in wheat variety Guomai 301 (*Li et al., 2021*). New approaches have been undertaken in recent years to produce EMS-induced rice varieties at research institutes (*Kumawat et al., 2022*). The LD_{50} dose is first calculated and then used to determine the best dose for inducing mutations in CR1009 and CR1009 sub1 rice (*Khannetah et al., 2021*). Thai highland rice (cv. Dawk Pa-yawm and Dawk Kha 50) was subjected to induced mutagenesis using EMS to create genetic variability (*Awais, Nualsri & Soonsuwon, 2019; Unan et al., 2022*). By leaving out this stage, the mutagen dose can result in either a high or low mutation frequency (*Barr & Fearn, 2016; Espina et al., 2018; Galal & Thabet, 2018*).

Basmati rice has a narrow genetic base and broadening its genetic base using non-Basmati rice material may affect its quality attributes. The transgenic approaches have GMO issues. Hence, changing genes in the living cell is not an easy job. The improvement of the CRISPR-Cas9 system in plants is carried out by gene editing which is more specific to gene removal or removing sequences. In CRISPR Cas9 the mutations are also random but often the intended changes are very precise. The removal of sequences/genes may have negative effects on disease resistance, drought, and salinity tolerance. The objective of CRISPR Cas9 is also to develop resistant plants against biotic and abiotic stresses. In the CRISPR editing system there is the

Possibility of altering off-target genes. However, the individuals resulting from the edit can be scrutinized to ensure that there are no additional changes. This can consume time and significant financial resources. It is possible that, in this sense, the use of EMS may be a better option.

In CRISPR Cas9, the bacterial system is used to protect from viruses and replace the mutant/lethal gene with the healthy copy. This can be done by adding the other DNA that carries the desired sequence in cultured cells. In the mutation breeding approach, mutants developed mostly due to deletions in DNA sequences, and no transgenic approach is involved. Like CRISPR Cas9, the cultured cells (Callus) are screened on hygromycin media to conform to the transgenic. Basmati rice being an exportable commodity, the hygromycin resistance is an issue in Basmati rice patenting. The induced mutation has the option to alter one or the other desirable genes without compromising other traits.

Besides, among the conventional breeding and transgenic approaches, induced mutation is the easiest approach that can be used for creating genetic variability without compromising the quality attributes. Inducing genetic diversity in rice by ionizing radiation has been proven to be successful. Understanding the relative biological effectiveness and efficiency of different mutagens is helpful in mutation breeding before the start of any sound breeding program. Many scientists have undertaken several experimental investigations in

this regard to identify the most efficient mutagenic method for the induction of desirable features in rice (Jankowicz-Cieslak, Mba & Till, 2017; Rashid et al., 2009).

This research aims at determining an optimal EMS dose (-50% lethal dose) in comparison to the standard (control) in order to generate variability, keeping typical characteristics of Super Basmati rice. Various EMS concentrations were applied to Super Basmati rice (Fragrant Rice) seeds and systematically assessed the survival and lethal doses during germination, seedling lethality, morpho-physio-biochemical, and yield attributes. Based on these observations, we may determine the optimal dose for EMS mutagenesis in the Super Basmati rice cultivar improvement against different ecological extremes conditions.

MATERIALS & METHODS

Plant material

In this research work a total of 400 seeds of the rice cultivar Super Basmati (*Oryza sativa* L. spp.) were selected by collecting from the Plant Breeding and Genetics Division of the Nuclear Institute for Agriculture and Biology (NIAB), Faisalabad, Pakistan.

EMS mutagenesis

Super Basmati seeds were soaked in ultrapure water (-100 mL) up to a height of 5 cm above the seeds and stored at room temperature overnight for 20 h. The 50 mL of EMS with (v/v) concentrations (0.25%, 0.50%, 0.75%, 1.0%, and 1.25%) were added after decanting the ultrapure water (resistivity of 18.2 M Ω . cm at 25 °C). The seeds were then transferred and rinsed with 100 mL ultrapure water (five times and 4 min each) and 200 mL ultrapure water (four times, and 15 min each), and after that treated seeds were incubated at room temperature for 12 h. The processed seeds were then washed in continuously running tap water for about 4 h before being placed in Petri dishes for further analysis (Talebi, Talebi & Shahrokhifar, 2012) (Fig. 1).

Photosynthetic attributes

Photosynthetic rate (A), transpiration rate (E), and stomatal conductance (g_s) were measured on completely lengthened uppermost leaves with a portable photosynthesis system (Infra-Red Gas Analyzer) at a light saturating intensity between 9:00 am to 12:00 noon on a full-sun day. Samples (~5 g) of leaves from each treatment were collected on test tubes containing acetone (85% each sample) were macerated in the same tube. Samples were kept in the dark for 24 h to allow the extraction of photosynthetic pigments. Tubes were centrifuged for 10 min at 4,000 \times at 4 °C to remove cellular debris. Supernatants were measured in a spectrophotometer (Halo DB-20/ DB-20S, UK) at 470, 647, and 664.5 nm to measure contents of chlorophyll (Lichtenthaler, 1987).

Water related attributes

For the determination of water related attributes, three penultimate leaves of each treatment were harvested at the tillering stage. A pressure chamber was used to determine the leaf water potential. After that leaf samples were frozen and thawed sap was extracted and the osmotic potential was determined using digital Osmoter (Wescor, Logan, UT, USA) (Farooq, Wahid & Lee, 2009).

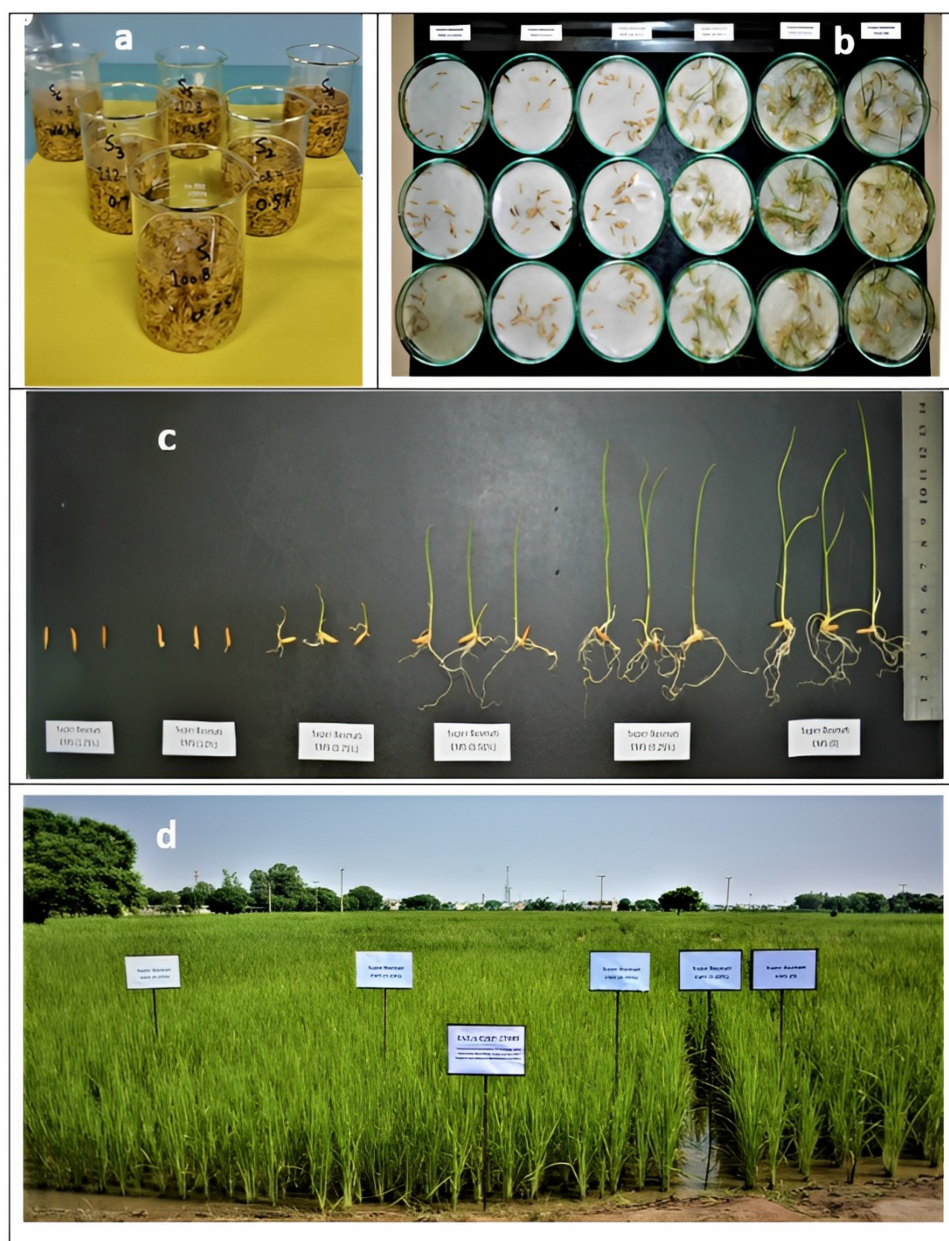


Figure 1 Chemical induced mutagenesis in Super Basmati: (A) soaking paddy seeds of Super Basmati in 4 ultrapure water for 20 h, (B) sowing of EMS treated seeds of Super Basmati in Petri plates, (C) effect of 5 EMS on seedling, (D) sowing of EMS treated seed in the field.

Full-size DOI: 10.7717/peerj.15821/fig-1

Activities of enzymatic antioxidants

An aliquot of fresh green leaf sample amount was homogenized with five mL of 50 mM Tris-HCl buffer (pH 8.0) for CAT and 50 mM KH_2PO_4 buffer (pH 7.0) for POX and APX determination. The homogenate was centrifuged at 5000 rpm for 20 min and the supernatant was then used as enzyme extract. The CAT (EC: 1.11.1.6) activity was assayed as

described by *Islam et al. (2009)*. The POX (EC: 1.11.1.7) and APX (EC: 1.11.1.11) activities were assayed as described by *Zeng, Liu & Xu (2011)*.

Lethal dose study in EMS mutagenesis

In addition to the control, a total of about 40 seeds were planted on filter paper that had been dipped in 5 mL of ultrapure water in Petri plates following the EMS-induced treatments. Petri plates were then placed at 25 °C for 7 days in an incubator. The number of seeds that were grown under controlled conditions was counted and observed after seven days of germination. The germinated seeds from each applied EMS concentration, as compared to the control, were shifted to plastic pots and later in the rice field. In the greenhouse, the seedlings were irrigated with distilled water. After two weeks, the shoot and root lengths were measured using the sandwich blotter technique (*Ariraman et al., 2014*). After sown on the nursery bed, the emergence was recorded for each dose after germination. Parameters such as the height of the plant, panicle length, productive tillers, total spikelet, sterile spikelet, and fertility percent were measured at the physical maturity of rice plants.

Statistical analysis

For lethal dose determination, the rice variety was treated with five levels of EMS concentrations and then sown in triplicate in a randomized block design. The least significant difference (LSD) test with p -values less than 0.05 was employed to analyze the average variance for all investigated parameters between treated and control plants. The statistical evaluation was carried out using Statistix 8.13 software. Principle component analysis (PCA) was carried out using Minitab-19.

RESULTS

Effect of EMS-induced mutagenesis on germination

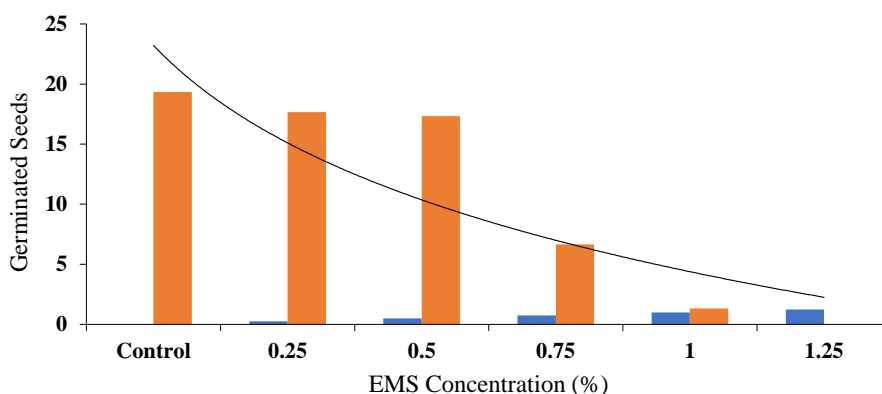
The experimental data indicated that different doses of EMS caused variations in the germination of aromatic rice. The seed germination attributes of the control along with the treated seeds of Basmati rice are shown in [Table 1](#). The major differences in the values of seed germination after the EMS treatment at varying doses of 0.25%, 0.50%, 0.75%, 1.00%, and 1.25% were highly significant at the 5% concentration. In all cases, there is an inhibitory effect, but it occurs to different degrees depending on the dose level. In the case of seed germination, data indicate that EMS had a retarding growth effect, or even inhibit it, depending on the dose applied, as compared to the control ([Fig. 2](#)). Under the conditions of EMS treatments of 0.25% and 0.50% levels, seeds showed the highest germination percentages, 91.4% & 89.6%, respectively, among all other EMS treatments. Low seed germination percentages were recorded in the higher doses at 0.75% and 1% levels to be 34.4% and 6.9%, respectively, while 1.25% treatment did not register any germination.

Effect of EMS mutagenesis on seedling growth attributes

The length of the roots and shoots revealed that EMS-induced mutagenesis had a substantial effect on their growth as indicated by the size. According to the measurements and

Table 1 Mean value of germination attribute of fragrant rice by following EMS mutagenesis.

Treatment	Germination	
	Observed	% Control
Control	19.33	100
0.25%	17.67	91.4
0.50%	17.33	89.6
0.75%	6.66	34.4
1.00%	1.33	6.9
1.25%	0	0
LSD%	0.94	
C.V%	5.07	

**Figure 2** Effect of different concentrations of EMS mutagenesis on seed germination of fragrant rice.

Full-size  DOI: [10.7717/peerj.15821/fig-2](https://doi.org/10.7717/peerj.15821/fig-2)

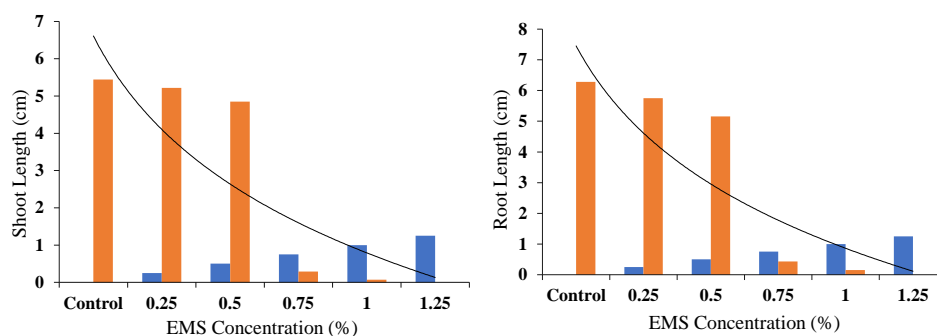
observations, shoot length decreased in proportion to the amount of EMS applied (Table 2 & Fig. 3). It was evident that when the concentration of EMS was increased, the root length decreased as compared to the control. The EMS 0.25% level exhibited the highest shoot length of 5.22 cm as compared to the control (5.44 cm). In other treatments of EMS viz. 0.5%, 1.0%, and 1.25%, a decreasing trend in shoot length of 4.85, 0.29, and 0.07 cm, respectively, were recorded as compared to the control. Among the EMS treatments, 0.25% level exhibited a maximum root length (5.75 cm) than the other treatments of 0.5% EMS (5.16 cm), 0.75% EMS (0.43 cm), and 1% EMS (0.15 cm). At a 1.0% concentration of EMS, the shortest root length was recorded in the experiment. A decreasing trend in the length of shoot and root was observed with the increase in the dose of EMS. When Basmati rice was treated with EMS concentrations greater than 1%, no seed germination was observed for the genotype under consideration.

Effect of EMS mutagenesis on photosynthetic attributes

Various concentrations of EMS treatments significantly affected the photosynthetic attributes of fragrant rice. It was observed that by increasing the concentration of EMS treatments a linear increase was noticed up to 0.75% of EMS treatment. At 1%

Table 2 Average value of root and shoot length attributes of fragrant rice by EMS-induced mutagenesis.

Treatment	Root length (cm)		Shoot length (cm)	
	Observed	% Control	Observed	% Control
Control	6.28	100	5.44	100
0.25%	5.75	91.56	5.22	95.96
0.50%	5.16	82.17	4.85	89.15
0.75%	0.43	6.85	0.29	5.33
1.00%	0.15	2.39	0.07	1.29
1.25%	0	0	0	0
LSD%	0.16		0.53	
C.V%	3.01		11.34	

**Figure 3** Effect of different concentrations of EMS mutagenesis on root and shoot growth of fragrant rice.

Full-size DOI: [10.7717/peerj.15821/fig-3](https://doi.org/10.7717/peerj.15821/fig-3)

Table 3 Effect of different concentrations of EMS mutagenesis on photosynthetic attributes of fragrant rice.

Treatments	Chlorophyll Contents (mg L ⁻¹)	Photosynthetic Rate (μmol CO ₂ m ⁻² s ⁻¹)	Transpiration Rate (mmol H ₂ O m ⁻² s ⁻¹)	Stomatal Conductance (mmol H ₂ O m ⁻² s ⁻¹)
Control	4.87 B	10.84 D	4.83 E	0.08 D
0.25%	5.03 B	14.97 BC	6.22 D	0.06 CD
0.50%	5.30 A	17.57 B	9.04 B	0.35 AB
0.75%	5.40 A	13.96 A	10.18 A	0.41 A
1.00%	5.26 A	12.33 CD	8.33 C	0.26 BC
1.25%	0.00 C	0.00 E	0.00 F	0.00 D
HSD ($p \leq 0.05$)	0.17	2.57	0.68	0.15

Notes.

Within each column, mean data followed by the same letters are not statistically different ($p \leq 0.05$ HSD test).

concentration, there was a decrease in all the photosynthetic attributes that was statistically at par with the 0.25% EMS treatment. The maximum of all the photosynthetic attributes was observed at 1.00% EMS treatment (Table 3).

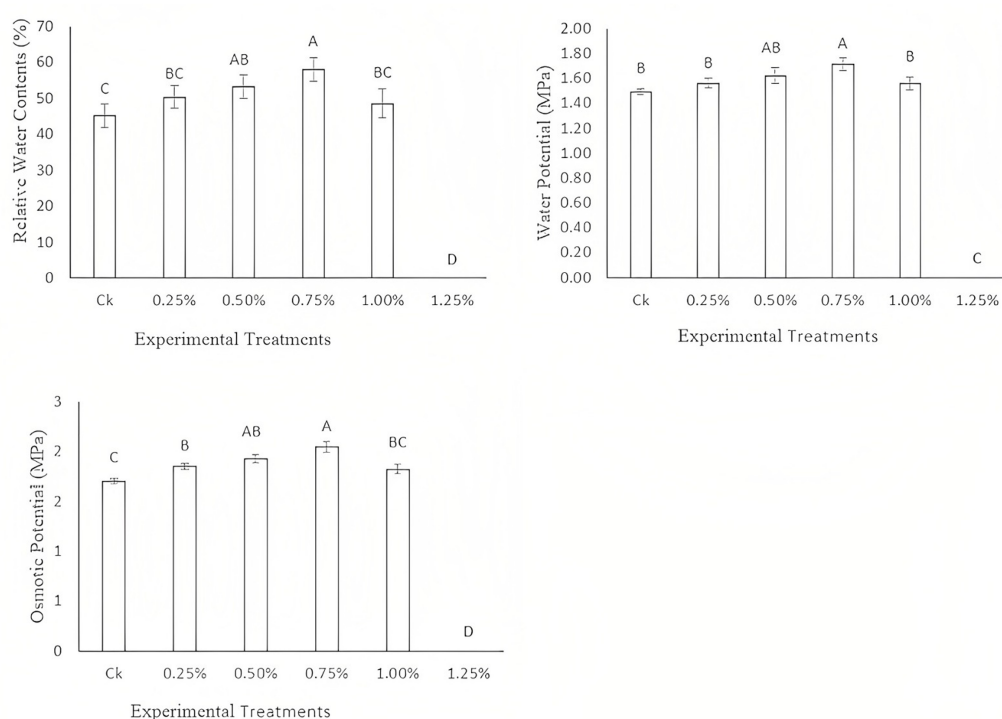


Figure 4 Effect of different concentrations of EMS mutagenesis on water related attributes of fragrant rice.

Full-size DOI: [10.7717/peerj.15821/fig-4](https://doi.org/10.7717/peerj.15821/fig-4)

Effect of EMS mutagenesis on water related attributes

It is observed from the experimental data of the current study that there exists a linear increase in the water related attributes after the EMS mutagenesis. Maximum relative water contents, water potential, and osmotic potential were noticed at 0.75% EMS treatment in the fragrant rice. However, at 1.00% there is a decrease in the water related attributes that was statistically at par with the concentration of 0.25%. A minimum of all the water related attributes was noticed under control conditions (Fig. 4).

Effect of EMS mutagenesis on enzymatic antioxidants attributes

Significant variation was observed in the activities of enzymatic antioxidants by the EMS treatments. A linear increase was observed in all of the activities by the EMS treatments. A minimum activity was noticed at control while the maximum activity was observed at 1.00% EMS treatment. At 1.25% concentration, there was no value because of no germination (Table 4).

Effect of EMS mutagenesis on phenological and yield contributing attributes

The maximum plant height (Table 5 & Fig. 5) after attaining maturity level was measured to be 104.6 cm and 105.0 cm in 0.25% EMS treatment and control, respectively. The minimum plant height (94.67 cm) was measured in 1% dose of EMS. In the present study,

Table 4 Effect of different concentrations of EMS mutagenesis on enzymatic antioxidants of fragrant rice.

Treatments	APX activity ($\mu\text{mol}/\text{min}/\text{g FW}$)	POX activity ($\mu\text{mol}/\text{min}/\text{g FW}$)	CAT activity ($\mu\text{mol}/\text{min}/\text{g FW}$)
Control	5.33 E	9.10 E	2.04 C
0.25%	8.65 D	13.18 D	2.86 C
0.50%	11.30 C	16.30 C	3.70 B
0.75%	14.30 B	21.23 B	4.11 B
1.00%	16.93 A	27.97 A	4.93 A
1.25%	0.00 F	0.00 F	0.00 D
HSD ($p \leq 0.05$)	0.99	1.54	0.82

Notes.

Within each column, mean data followed by the same letters are not statistically different ($p \leq 0.05$ HSD test).

Table 5 Mean value of phonological and yield attribute of fragrant rice by following EMS mutagenesis.

Treatment	Plant height (cm)		Productive tillers		Panicle length (cm)	
	Observed	% Control	Observed	% Control	Observed	% Control
Control	104.97	100	3.90	100	26.9	100
0.25%	104.57	99.62	3.67	94.10	26.9	100
0.50%	104	99.08	3.60	92.31	25.63	95.28
0.75%	103	98.12	3.50	89.74	25.3	94.05
1.00%	94.67	90.19	3.17	81.28	24.67	91.71
1.25%	0	0	0.00	0	0	0
LSD%	4.93		0.52		0.90	
C.V%	3.25		9.87		2.35	

EMS treatment at the highest concentration (1.25%) has shown an inhibitory effect as compared to the control. Productive tillers, panicle length, and total spikelet (Table 5) showed a maximum length of 3.67, 26.9, and 111.9 cm at 0.25% EMS treatments whereas control ranged as 3.90, 26.9, and 112.9 cm of productive tillers, panicle length, and total spikelet, respectively. However, at a 1.5% EMS level, an inhibitory effect for productive tillers, panicle length, and total spikelet was recorded as compared to the control. It was observed that the proportion of sterile spikelets increased as the applied EMS concentration increased. When Super Basmati was treated with a concentration of 0.25%, the highest value in sterile spikelets (85.2) was observed. Figure 3C indicates that as the applied EMS concentration was increased, fertility decreased. In the control group, the highest fertility rate (13.12%) was observed. When the Super Basmati cultivar was subjected to a concentration of 0.25%, the highest fertility rate (11.48%) was observed. At 1% EMS concentration, the lowest sterile spikelets, and fertility were registered. An EMS treatment above 1% concentration, showed a total inhibition of sterile spikelet, and fertility (Table 6).

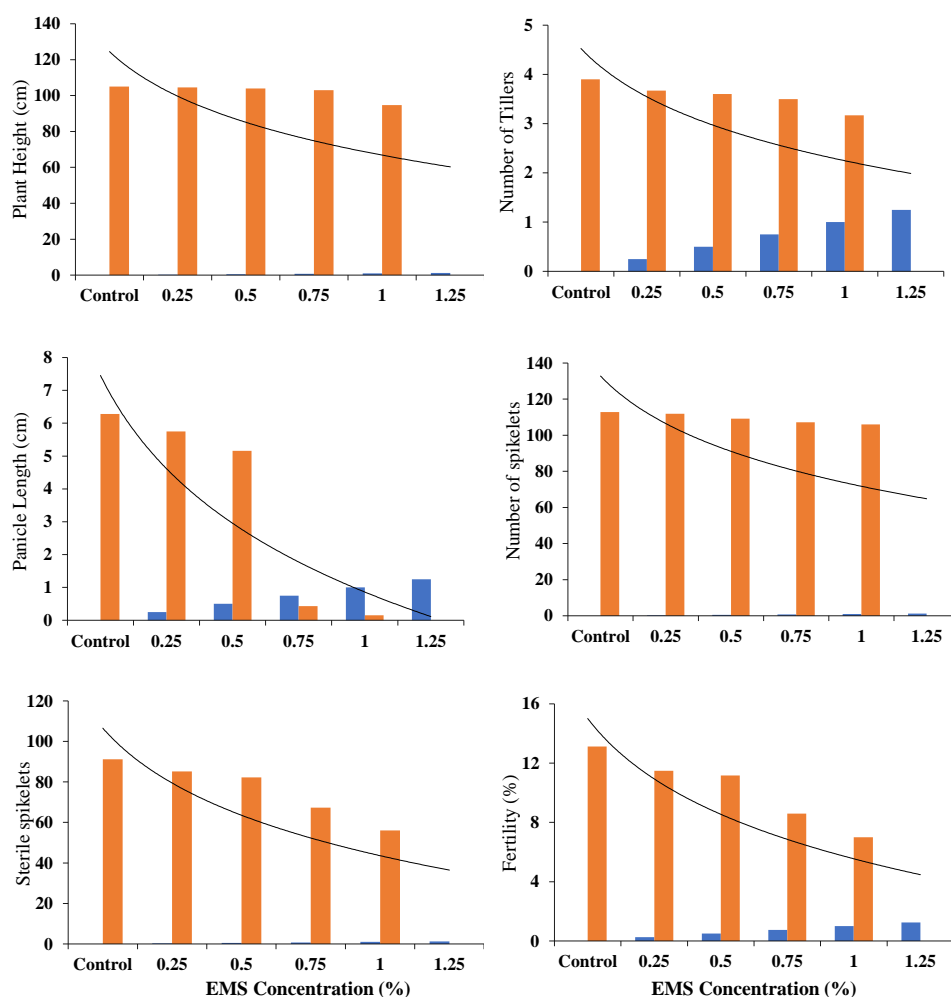


Figure 5 Effect of different concentrations of EMS mutagenesis on phonological and yield attributes of fragrant rice.

[Full-size !\[\]\(666e09182d4cd268646ea700ea60dcdf_img.jpg\) DOI: 10.7717/peerj.15821/fig-5](https://doi.org/10.7717/peerj.15821/fig-5)

Lethal dose effects

An EMS treatment at various concentration levels was evaluated on the aromatic rice variety Super Basmati to determine the LD_{50} based on the germination rate, growth, and yield attributes of rice (Fig. 6). The results obtained in this investigation indicated that the germination and all other measured attributes decreased when the EMS dose was increased. The LD_{50} values for seed germination (0.069%), root length (0.6%), shoot length (0.625%), plant height (1.125%), productive tillers (1.125%), panicle length (1.125%), total spikelet (1.126%), sterile spikelet (1.06%), and productivity (1.05%) for Super Basmati rice variety were carefully determined (Fig. 7).

DISCUSSION

EMS mutagenesis resulted in a major reduction in germination under the prevailing field conditions. As the EMS concentration increased, there was a substantial decrease in seed

Table 6 Mean value of yield attribute of fragrant rice by following EMS mutagenesis.

Treatment	Total spikelet		Sterile spikelet		Fertility (%)	
	Observed	% Control	Observed	% Control	Observed	% Control
Control	112.9	100	91.13	100	13.12	100
0.25%	111.9	99.11	85.2	93.49	11.48	87.5
0.50%	109.2	96.72	82.27	90.28	11.17	85.14
0.75%	107.2	94.95	67.33	73.88	8.6	65.55
1.00%	106	93.89	56	61.45	7	53.35
1.25%	0	0	0	0	0	0
LSD%	42.23		5.26		0.99	
C.V%	27.72		4.65		6.50	

germination. The EMS has shown to be one of the most potent chemical mutagens as an alkylating agent. According to previous research work, it has been documented that polyploids are more tolerant than diploids (*Chopra, 2005*). The Basmati rice used in the study is also diploid. The percentage reduction in seed germination might have been caused by the influence and impact of mutagens on the meristematic tissues of the seed. The decrease in seed germination at higher dose levels of the mutagens may be attributed to the disturbances at the cellular level with implications at the physiological level.

Earlier research work has shown that in okra (*Abelmoschus esculentus*), germination percentage generally decreased with increasing dose concentrations of gamma rays and the EMS levels (*Gupta et al., 2016*). Reduced germination percentage with increasing doses of gamma radiation has also been reported in pinus (*Ariraman et al., 2014*), rye (*Khah & Verma, 2015*), and chickpea plants (*Shah et al., 2008*). A gradual reduction in germination percentage was also observed with an increase in the concentration of mutagen, reaching more than 50% lethality at 0.5% EMS level in two genotypes of tobacco plants (*Dhakshanamoorthy, Selvaraj & Chidambaram, 2010*). In this study, seeds of the Super Basmati rice variety were treated with chemical mutagen EMS viz., 0.25%, 0.5%, 0.75%, 1%, and 1.25% concentrations. In the laboratory germination test, it was observed that an increase in the level of EMS had an overall adverse effect. Similar results have been reported in a previous study of *capsicum annum* (*Hasan et al., 2022*) that seeds treated with 1.5% of EMS dose in M1 generations had the lowest germination percentage (–84%) among all treatments. The germination percentage was found to be profoundly inhibited by EMS treatment in two varieties (Co1 and Co2) of soybean plants (*Karthika & Subba, 2006*). The mutagenic reaction is more or less linear with the dosage quantity. Plant survival to maturity is dependent on the type and extent of chromosomal damage according to a previous study on radiation mutation (*Naaz et al., 2022*). Germination inability, plant growth, and survival can be reduced as the occurrence of chromosomal damage increases with growing radiation dose (*Sood et al., 2016*). Furthermore, genes close to the centromere are more sensitive to mutagenic treatment than genes further apart. Significant change in the chlorophyll contents were noticed by the EMS treatment group and increased in the frequency of chlorophyll mutants depends on the dose or concentration of the mutagen

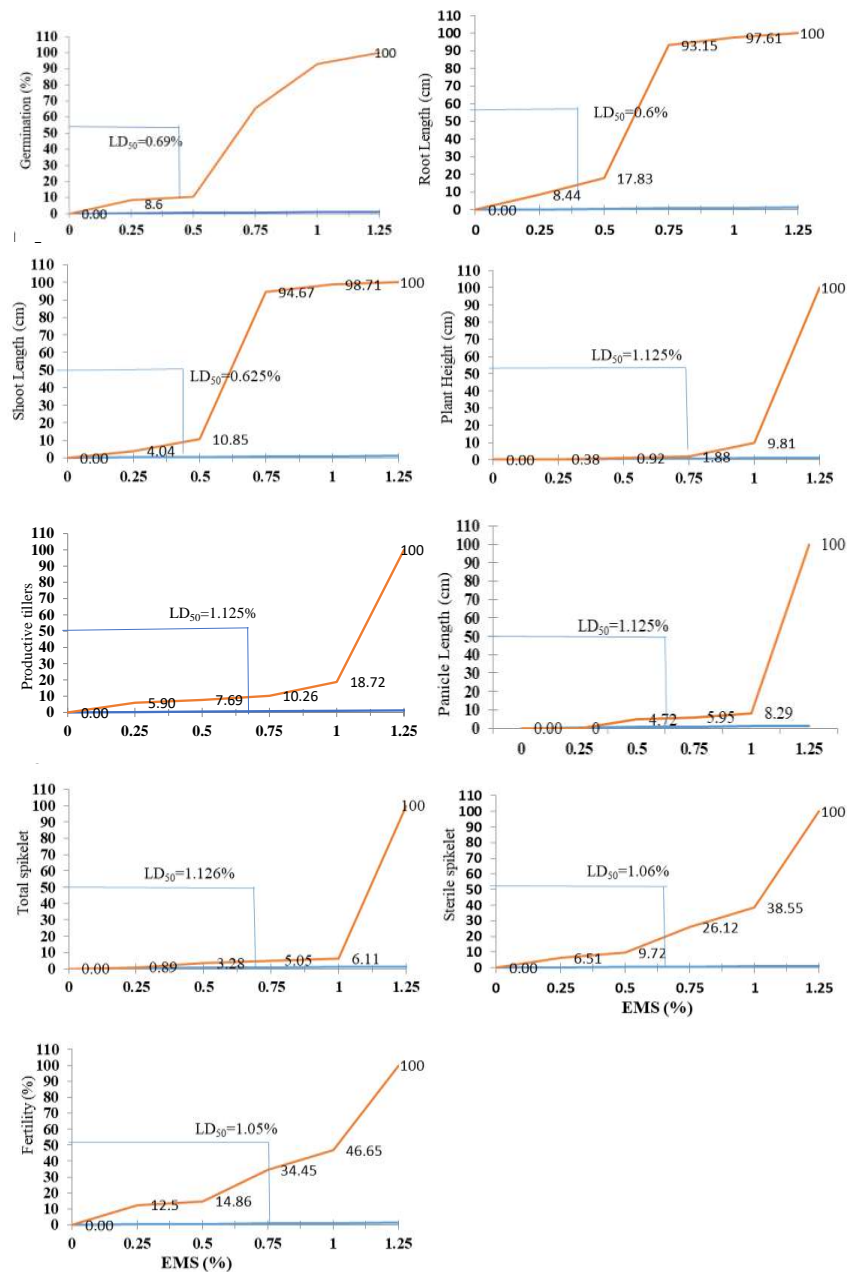


Figure 6 Effect of chemical mutagen (EMS) LD₅₀ value on germination, growth, and yield attributes of Super Basmati.

Full-size DOI: [10.7717/peerj.15821/fig-6](https://doi.org/10.7717/peerj.15821/fig-6)

used (Bado et al., 2015). The activation of RNA or protein synthesis may be responsible for the stimulating effect of physical mutation on germination. It can happen after the seeds have been processed during the early stages of germination (Zhang et al., 2021). Shoot length is most commonly used as an index to classify and report the biological effects of different physical and chemical mutagens in M1 (Boyane, 2015). Shoot length and the dosage of physical or chemical mutagens have been shown to have a linear relationship.

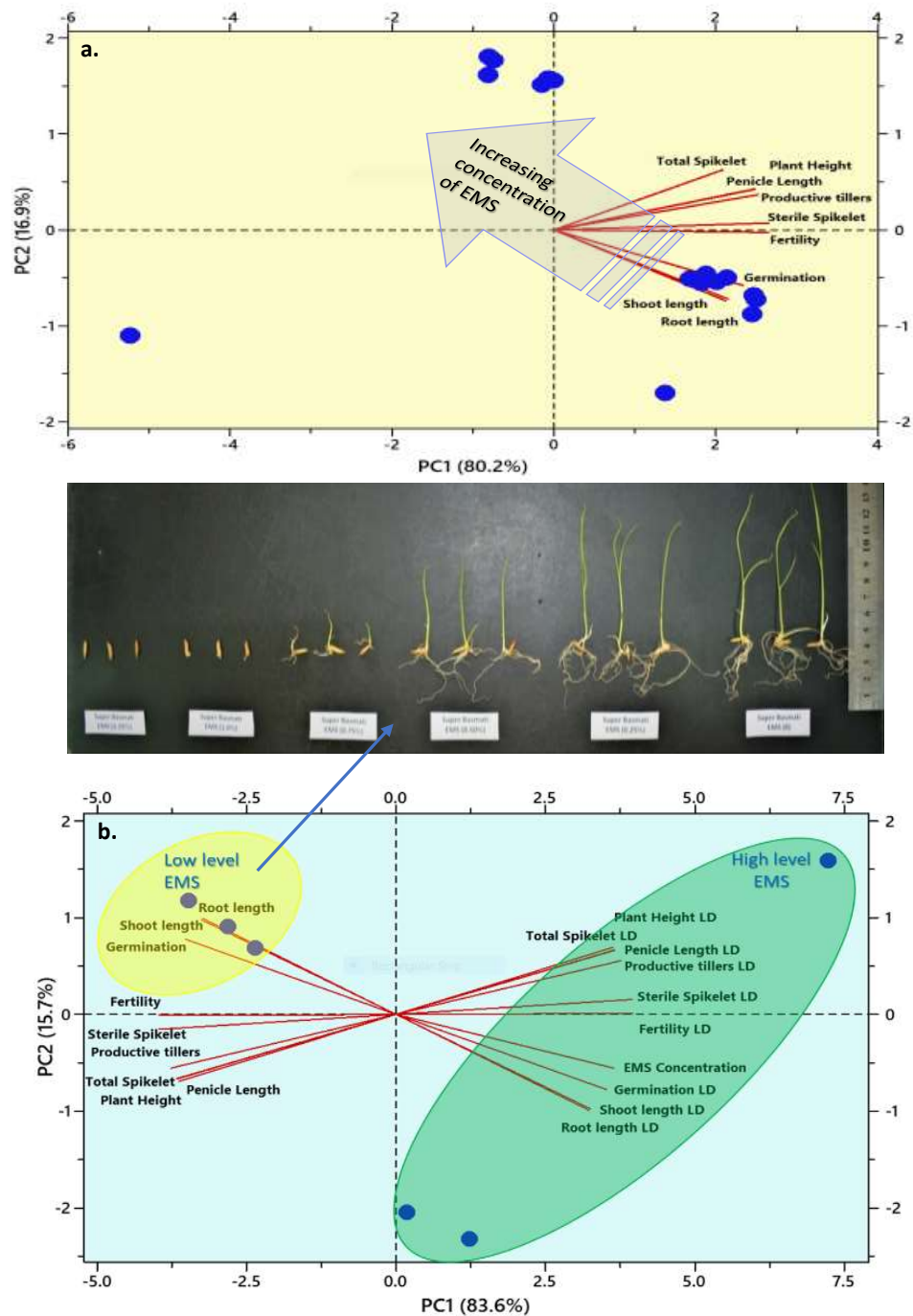


Figure 7 Biplots from Principal components analysis of the measured parameter of the Super Basmati rice. Lines indicate determined parameters and circles show the chemical mutagens levels. Image point to changes in the lengths of roots and shoots. The principal components analysis revealed a systematic trend in EMS treated plants. Two groups formed due to mutual association with the determined variables. An arrow indicates that as the concentration of EMS increases, germination, shoot, and root lengths decrease. An outlier (plot a) shown on the negative PC1 and PC2 quadrant represent the highest EMS level at which germination is inhibited. A cluster of measured parameters (plot b) groups traits at low and high levels of mutagen. A low level of EMS dose results in higher fertility and growth parameters such as root length.

Full-size DOI: [10.7717/peerj.15821/fig-7](https://doi.org/10.7717/peerj.15821/fig-7)

Measured data of this study showed that increases in the EMS concentrations caused decreases in shoot length. Our findings also revealed that when the rice variety of Super Basmati was treated with EMS, the shoot length decreased significantly as compared to the control. The concentration of applied EMS had an important impact on the root length of Super Basmati rice. Every subsequent increase in the EMS concentration resulted in a reduction in root length. Enhancement or inhibition of germination, shoot length, and other biological responses are commonly observed in low or high dose treated plants ([Deoli & Hasenstein, 2018](#)).

According to [Ramchander, Pillai & Ushakumari \(2014\)](#), a low dose of irradiation induces growth stimulation by either modifying the hormonal signaling network in plant cells or growing the cells' anti-oxidative ability. Plants can easily withstand everyday stress factors such as light intensity and temperature variations in the environment. The cell cycle arrests at the G2/M phase during the somatic cell division and various types of damages in the entire genome have been associated with the high dose treatments ([Ahmad et al., 2022](#); [Jan, Parween & Siddiqi, 2012](#)). Variability was assessed in this analysis by the mean values of the shoot and root lengths, both of which decreased as the concentration of EMS increased. When the radiation amount is sufficient to reduce root length percentage, the root lengths do not exceed a few millimeters in size as reported in a physical mutation analysis ([Chaudhuri, 2002](#)). After irradiation, seeds are unable to germinate due to metabolic disorders ([Sood et al., 2016](#)).

When rice plants are exposed to lower dosages of mutagens, they exhibit defensive responses that involve structural changes in the photosynthetic machinery. Increasing the concentration of EMS enhanced the concentration of photosynthetic attributes in a linear pattern, however, at 1.25% EMS concentration plant growth and development was affected due to poor rate of germination. Maximum chlorophyll content was seen when *capsicum annuum* was treated with 0.1% EMS for 3 h, according to [Ahmad & Asif \(2023\)](#). [Saba & Mirza \(2002\)](#) reported a similar finding by discovering that tomato plants treated with 0.5% EMS for three hours had the maximum chlorophyll content along with other photosynthetic attributes. Enhancement in the water related attributes might be due to the better growth and stay green character of rice plants ([Chaudhari, Verma & Chaudhary, 2015](#)). However, lower and extremely higher doses could cause a significant change in the DNA of the rice plants, as has been reported for mutagenesis studies in rice ([Abid et al., 2018](#); [Viana et al., 2019](#)). Except for control and 1.25% EMS concentration, all treatments exhibited a notable improvement in water content, but the 1.00% exhibited a decline in the water related attributes. According to [Elyadini et al. \(2021\)](#), one of the two adaption mechanisms maintaining a high level of tissue elasticity or lowering osmotic pressure can contribute to the maintenance of a relatively high value of the relative water content under EMS treatment leading to a positive effect on the rice plants for improving its overall productivity.

The enzymatic antioxidants are essential enzymes in plant cells that remove H₂O₂ from various organelles like the cytosol and chloroplast to prevent oxidative damage ([Zahra et al., 2021](#)). High APX activity was observed in this study and various researchers have also observed an increase in APX activity during the increased concentrations of EMS

treatments (*Hamid et al., 2015*). Therefore, the production of reactive oxygen species in cells that lack water content causes cell damage, which eventually culminates in cell death (*Bali & Sidhu, 2019*). The balance in the enzymatic antioxidants, whose activity was raised at moderate levels of EMS treatments, is one of the antioxidant systems that regulate oxidative stress through a variety of adaptive ways (*Palace et al., 1998*). Under higher levels of EMS, enzymatic antioxidant activity was found to be elevated in wheat, as it was in many other crop species (*Devi, Kaur & Gupta, 2012*). Cell walls, vacuoles, extracellular spaces, and cytosol, all contain APX. This enzyme, which is known as a stress indicator, has a broad range of phenolic substrate selectivity and is attracted to H₂O₂ than catalase. It can use H₂O₂ to produce phenoxy chemicals, which ultimately polymerize lignin, a component of the cell wall (*Štolfa et al., 2015*).

The EMS-treated seeds may develop a mutant basmati rice variety. This is possibly due to the pleiotropic impact of mutated genes or mutations on various genome loci (*Muqaddasi & Arif, 2012*). Several morphological mutations in legume plants have also been identified (*Goyal et al., 2019*) and few of mutations have been shown to affect multiple attributes. A combination of the elevated amount of dose and the period of treatment resulted in higher seedling death and lower yield in the plant characteristics in the EMS-treated seeds. Similar results were recorded in an experiment of EMS-treated fenugreek seeds, where no callus cultures were developed when treated with EMS levels above 1% (*Basu, Acharya & Thomas, 2008*). In this analysis, LD₅₀ values for yield contributing traits included the plant height (1.125%), productive tillers (1.125%), panicle length (1.125%), complete spikelet (1.126%), sterile spikelet (1.06%), and fertility (1.05%) which were found in seeds treated with 0, 0.25, 0.5, 0.75, 1, and 1.25 percent EMS, resulting in an inverse association between all of these yielding traits (*Kozgar, Goyal & Khan, 2011*). The efficacy of the current study decreased as the concentration of EMS increased. This observation was also confirmed by the findings in black gram (*Usharani & Kumar, 2015*), chickpea (*Singh et al., 2015*) and cowpea plants (*Nair & Mehta, 2014*).

The variation in LD₅₀ for the Super Basmati rice variety at the different EMS (%) concentrations has been observed in mutation studies, and it is thought to be mainly due to the biological material, scale, maturity, hardness, and moisture content at the time of exposure of breeding material (*Thakur, Paul & Kumar, 2020*). There is sufficient evidence that the radiation-induced sterility of M1 panicles is passed on to subsequent generations (*Jyothilekshmi, 2012*). Physiological damage induces a significant portion of sterility, which is not passed on to the next generation. It is found in this research work that with the increasing doses of mutagen treatments, induced panicle sterility. These findings are consistent with those of previous researchers (*El-Degwy, 2013; Siddiqui & Singh, 2010*) who found that gamma-ray treatment caused rice plants to become highly sterile. In determining the yield potential of these mutants, it will be vital to analyze the heritability in a multi-location yield trial that incorporates suitable experimental design to assess whether these mutations will perform consistently across different environments.

CONCLUSIONS

Physicochemical mutagenesis has been employed to produce genetic variability in crops including rice plants. The EMS induced mutagenesis is a promising exploratory tool to search for novel players for improving agronomic and yield contributing traits. Germination, seedling growth, and yield attributes were significantly influenced by variations in EMS concentration treatments. There was no germination observed upon the application of a 1.25% concentration of EMS treatment for seed germination and 50% germination was recorded between EMS 0.50% and EMS 0.75% treatments. After the cultivation of rice plants of the M1 generation in the field, it was observed that LD_{50} for fertility occurred at EMS 1.0% for the investigated rice variety. The EMS treatment demonstrated a negative biological influence such as reduced germination and abnormal seedling development of Basmati rice plants. It is, therefore, concluded that for creating genetic variability in the rice variety of Super Basmati, the EMS doses from 0.5% to 0.75% are more useful and effective for improving the overall performance of fragrant rice. Furthermore, mutants with yield related value-added traits will be available for the scientific community for advanced level research and will also serve as a public genetic resource for development and breeding programs.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by the Researchers Supporting Project number (RSP2023R194), King Saud University, Riyadh, Saudi Arabia. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:
King Saud University, Riyadh, Saudi Arabia: RSP2023R194.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Areeqa Shamshad conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Muhammad Rashid conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Ljupcho Jankuloski analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Kamran Ashraf analyzed the data, prepared figures and/or tables, and approved the final draft.
- Khawar Sultan analyzed the data, prepared figures and/or tables, and approved the final draft.

- Saud Alamri conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Manzer H. Siddiqui conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Tehzeem Munir analyzed the data, prepared figures and/or tables, and approved the final draft.
- Qamar uz Zaman conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The raw measurements are available in the [Supplemental Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.15821#supplemental-information>.

REFERENCES

- Abid M, Ali S, Qi LK, Zahoor R, Tian Z, Jiang D, Snider JL, Dai T. 2018.** Physiological and biochemical changes during drought and recovery periods at tillering and jointing stages in wheat (*Triticum aestivum* L.). *Scientific Reports* **8**:4615 DOI [10.1038/s41598-018-21441-7](https://doi.org/10.1038/s41598-018-21441-7).
- Ahmad I, Ahmad I, Muhammad Z, Ullah B. 2022.** Response of sorghum vulgare L. cultivars to gamma irradiation, a preliminary approach. *Journal of Applied Research in Plant Sciences* **3**:215–223 DOI [10.38211/joarps.2022.3.1.26](https://doi.org/10.38211/joarps.2022.3.1.26).
- Ahmad A, Asif A. 2023.** Mutagenesis in medicinal plants. In: *Omics studies of medicinal plants*. United Kingdom: CRC Press.
- Ariraman M, Gnanamurthy S, Dhanavel D, Bharathi T, Murugan S. 2014.** Mutagenic effect on seed germination, seedling growth and seedling survival of Pigeon pea (*Cajanus cajan* (L.) Millsp). *International Letters of Natural Sciences* **21**:41–49 DOI [10.56431/p-g8xcn6](https://doi.org/10.56431/p-g8xcn6).
- Awais A, Nualsri C, Soonsuwon W. 2019.** Induced mutagenesis for creating variability in Thailand's upland rice (cv. Dawk Pa-yawm and Dawk Kha 50) using ethyl methane sulphonate (EMS). *Sarhad Journal of Agriculture* **35**:293–301.
- Bado S, Forster BP, Nielen S, Ali AM, Lagoda PJ, Till BJ, Laimer M. 2015.** Plant mutation breeding: current progress and future assessment. *Plant Breeding Reviews* **39**:23–88.
- Bali AS, Sidhu GPS. 2019.** Abiotic stress-induced oxidative stress in wheat. In: Hasanuz-zaman M, Nahar K, Hossain M, eds. *Wheat production in changing environments*. Singapore: Springer.
- Barr J, Fearn R. 2016.** Genetic instability of RNA viruses. In: *Genome stability*. Academic Press, 21–35 DOI [10.1016/B978-0-12-803309-8.00002-1](https://doi.org/10.1016/B978-0-12-803309-8.00002-1).

- Basu SK, Acharya SN, Thomas JE. 2008.** Genetic improvement of fenugreek (*Trigonella foenum-graecum* L.) through EMS induced mutation breeding for higher seed yield under western Canada prairie conditions. *Euphytica* **160**:249–258
[DOI 10.1007/s10681-007-9545-9](https://doi.org/10.1007/s10681-007-9545-9).
- Boyane AB. 2015.** The effect of ethylmethanesulfonate (EMS) on morphological characteristics and seed quality development of Vernonia (*Centropalus pauciflorus* var. *ethiopica* Willd.). Masters Degree, University of Kwazulu Natal.
- Chaudhari AK, Verma S, Chaudhary B. 2015.** Ethyl methanesulphonate and sodium azide effects on seedling growth and chlorophyll mutations in psoralea corylifolia IC 111228. *Journal of Crop Improvement* **29**:602–618
[DOI 10.1080/15427528.2015.1070391](https://doi.org/10.1080/15427528.2015.1070391).
- Chaudhuri SK. 2002.** A simple and reliable method to detect gamma irradiated lentil (*Lens culinaris* Medik.) seeds by germination efficiency and seedling growth test. *Radiation Physics and Chemistry* **64**:131–136 [DOI 10.1016/S0969-806X\(01\)00467-4](https://doi.org/10.1016/S0969-806X(01)00467-4).
- Chopra V. 2005.** Mutagenesis: investigating the process and processing the outcome for crop improvement. *Current Science* **89**:353–359.
- Deoli NT, Hasenstein KH. 2018.** Irradiation effects of MeV protons on dry and hydrated Brassica rapa seeds. *Life Sciences in Space Research* **19**:24–30
[DOI 10.1016/j.lssr.2018.08.004](https://doi.org/10.1016/j.lssr.2018.08.004).
- Devi R, Kaur N, Gupta AK. 2012.** Potential of antioxidant enzymes in depicting drought tolerance of wheat (*Triticum aestivum* L.). *Indian Journal of Biochemistry and Biophysics* **49**(4):257–265.
- Dhakshanamoorthy D, Selvaraj R, Chidambaram A. 2010.** Physical and chemical mutagenesis in *Jatropha curcas* L. to induce variability in seed germination, growth and yield traits. *Romanian Journal of Biology—Plant Biology* **55**:113–125.
- El-Degwy IS. 2013.** Mutation induced genetic variability in rice (*Oryza sativa* L.). *International Journal of Agriculture and Crop Sciences* **5**:2789–2794.
- Elyadini M, Gaaadaoui A, ElHajjaji S, Labjar N, Labhili M, Gaboune F, Azeqour M. 2021.** Induced mutagenesis for improving water stress tolerance in durum wheat (*Triticum turgidum* L. subsp. durum). In: *E3S Web of Conferences*. EDP Sciences, 00107.
- Espina MJ, Ahmed CS, Bernardini A, Adeleke E, Yadegari Z, Arelli P, Pantalone V, Taheri A. 2018.** Development and phenotypic screening of an ethyl methane sulfonate mutant population in soybean. *Frontiers in Plant Science* **9**:394
[DOI 10.3389/fpls.2018.00394](https://doi.org/10.3389/fpls.2018.00394).
- Farooq M, Wahid A, Lee D-J. 2009.** Exogenously applied polyamines increase drought tolerance of rice by improving leaf water status, photosynthesis and membrane properties. *Acta Physiologiae Plantarum* **31**:937–945 [DOI 10.1007/s11738-009-0307-2](https://doi.org/10.1007/s11738-009-0307-2).
- Gadakh S, Patel D, Narwade A, Singh D. 2021.** Screening of EMS induced drought tolerant sugarcane (*Saccharum* spp. Complex) mutants employing physiological, molecular and biochemical approaches. *Indian Journal of Genetics and Plant Breeding* **81**:81–87.

- Galal O, Thabet A. 2018.** Cytological and molecular effects of silver nanoparticles (AgNPs) on *Vicia faba* M1 Plants. *Journal of Agricultural Chemistry and Biotechnology* **9**:269–275 DOI [10.21608/jacb.2018.36317](https://doi.org/10.21608/jacb.2018.36317).
- Goyal S, Wani MR, Laskar RA, Raina A, Amin R, Khan S. 2019.** Induction of morphological mutations and mutant phenotyping in black gram [*Vigna mungo* (L.) Hepper] using gamma rays and EMS. *Vegetos* **32**:464–472 DOI [10.1007/s42535-019-00057-w](https://doi.org/10.1007/s42535-019-00057-w).
- Gulfishan M, Bhat TA, Mir RA, Jahan A, Shabir F, Khan SA. 2023.** Revolutionizing plant biology. In: *Biotechnologies and genetics in plant mutation breeding. Plant mutagenesis: terms and applications*, United Kingdom: CRC Press.
- Gupta N, Sood S, Singh Y, Sood D. 2016.** Determination of lethal dose for gamma rays and ethyl methane sulphonate induced mutagenesis in okra (*Abelmoschus esculentus* (L.) Moench.). *SABRAO Journal of Breeding & Genetics* **48**:344–351.
- Hameed K, Khan MS, Sadaqat HA, Awan FS. 2019.** Phenotypic characterization of super basmati ethyl methane sulfonate (EMS) induced mutants. *Pakistan Journal of Agricultural Sciences* **56**(2):377–384.
- Hamid R, Kamili AN, Mahmooduzzafar O, Gücel S, Öztürk M, Ahmad P. 2015.** Analysis of physiobiochemical attributes, some key antioxidants and esculin content through HPLC in *in vitro* grown *Cichorium intybus* L. treated with ethylmethane sulfonate. *Plant Growth Regulation* **76**:233–241 DOI [10.1007/s10725-014-9992-y](https://doi.org/10.1007/s10725-014-9992-y).
- Hasan N, Choudhary S, Jahan M, Sharma N, Naaz N. 2022.** Mutagenic potential of cadmium nitrate [Cd (NO₃)₂] and ethyl-methane sulphonate [EMS] in quantitative and cyto-physiological characters of *Capsicum annum* L. cultivars. *Ecological Genetics and Genomics* **22**:100110 DOI [10.1016/j.egg.2021.100110](https://doi.org/10.1016/j.egg.2021.100110).
- Hernández-Muñoz S, Pedraza-Santos ME, López PA, Gómez-Sanabria JM, Morales-García JL. 2019.** Mutagenesis in the improvement of ornamental plants. *Revista Chapingo Serie horticultura* **25**:151–167 DOI [10.5154/r.rchsh.2018.12.022](https://doi.org/10.5154/r.rchsh.2018.12.022).
- Husain S, Bano F, Khan S. 2020.** *Studies on Cytomorphological Response of Vicia faba L. to Maleic hydrazide and Hydrazine hydrate*. Maude Avenue, Sunnyvale, CA: LAP LAMBERT Academic Publishing.
- Islam M, Parveen F, Hossain K, Khatun S, Karim MR, Kim G, Absar N, Haque MS. 2009.** Purification and biochemical characterization of lipase from the dorsal part of *Cirrhinus reba*. *Thai Journal of Agricultural Science* **42**:71–80.
- Jan S, Parween T, Siddiqi T. 2012.** Effect of gamma radiation on morphological, biochemical, and physiological aspects of plants and plant products. *Environmental Reviews* **20**:17–39 DOI [10.1139/a11-021](https://doi.org/10.1139/a11-021).
- Jankowicz-Cieslak J, Mba C, Till BJ. 2017.** Mutagenesis for crop breeding and functional genomics. In: Jankowicz-Cieslak J, Tai T, Kumlehn J, Till B, eds. *Biotechnologies for plant mutation breeding: protocols*. Singapore: Springer, Cham.
- Jia Y, Wang Z, Jia MH, Rutger JN, Moldenhauer KA. 2019.** Development and characterization of a large mutant population of a rice variety katy for functional genomics studies and breeding. *Crop Breeding, Genetics and Genomics* **1**:e190014.

- Jyothilekshmi S. 2012.** Induction of lodging resistance in upland rice (*Oryza sativa* L.) through mutagenesis. Doctoral dissertation, Department of Plant Breeding and Genetics, College of Horticulture, Vellanikkara.
- Karthika I, Subba B. 2006.** Effect of gamma rays and EMS on two varieties of soybean. *Asian Journal of Biological Sciences* 5:721–724.
- Kazama Y, Ishii K, Hirano T, Wakana T, Yamada M, Ohbu S, Abe T. 2017.** Different mutational function of low-and high-linear energy transfer heavy-ion irradiation demonstrated by whole-genome resequencing of Arabidopsis mutants. *The Plant Journal* 92:1020–1030 DOI 10.1111/tbj.13738.
- Khah MA, Verma RC. 2015.** Assessment of the effects of gamma radiations on various morphological and agronomic traits of common wheat (*Triticum aestivum* L.) var. WH-147. *European Journal of Experimental Biology* 5:6–11.
- Khannetah K, Pushpam R, Ganesan K, Kumar K, Chandrashekar C, Pillai MA. 2021.** Appraising LD50 dosage for physical mutagen (Gamma rays) in CR1009 and CR1009 sub1 rice varieties. *Journal of Pharmacognosy and Phytochemistry* 10:2715–2719.
- Kozgar MI, Goyal S, Khan S. 2011.** EMS induced mutational variability in *Vigna radiata* and *Vigna mungo*. *Research Journal of Botany* 6:31 DOI 10.3923/rjb.2011.31.37.
- Kumawat S, Raturi G, Dhiman P, Sudhakarn S, Rajora N, Thakral V, Yadav H, Padalkar G, Sharma Y, Rachappanavar V. 2022.** Opportunity and challenges for whole-genome resequencing-based genotyping in plants. In: *Genotyping by sequencing for crop improvement*. John Wiley & Sons Ltd.
- Le Roux W. 2019.** *Induced mutagenesis through gamma irradiation of embryogenic callus and selection for drought tolerance in sugarcane*. Stellenbosch: Stellenbosch University.
- Li J, Jiang Y, Zhang J, Ni Y, Jiao Z, Li H, Wang T, Zhang P, Guo W, Li L. 2021.** Key auxin response factor (ARF) genes constraining wheat tillering of mutant dmc. *PeerJ* 9:e12221 DOI 10.7717/peerj.12221.
- Lichtenthaler HK. 1987.** Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. In: *Methods in enzymology*. Elsevier, 350–382.
- Loko YLE, Ewedje E-E, Orobiyi A, Djedatin G, Toffa J, Gbemavo CD, Tchakpa C, Gavodo D, Sedah P, Sabot F. 2021.** On-farm management of rice diversity, varietal preference criteria, and farmers' perceptions of the African (*Oryza glaberrima* Steud.) versus Asian rice (*Oryza sativa* L.) in the Republic of Benin (West Africa): implications for breeding and conservation. *Economic Botany* 75:1–29 DOI 10.1007/s12231-021-09515-6.
- Malabadi RB, Kolkar K, Chalannavar R. 2022.** White, and brown rice-nutritional value and health benefits: arsenic toxicity in rice plants. *International Journal of Innovation Scientific Research and Review* 4:3065–3082.
- Muqaddasi Q, Arif M. 2012.** Ethyle methane sulphonate (EMS) induced mutagenic attempts to create genetic variability in Basmati rice. *Journal of Plant Breeding and Crop Science* 4:101–105.

- Naaz N, Choudhary S, Sharma N, Hasan N, Al Shaye NA, Abd El-Moneim D. 2022.** Frequency and spectrum of M2 mutants and genetic variability in cyto-agronomic characteristics of fenugreek induced by caffeine and sodium azide. *Frontiers in Plant Science* **13**:1030772 DOI [10.3389/fpls.2022.1030772](https://doi.org/10.3389/fpls.2022.1030772).
- Nair R, Mehta A. 2014.** Induced mutagenesis in cowpea [*Vigna unguiculata* (L.) Walp] var. Arka Garima. *Indian Journal of Agricultural Research* **48**:247–257 DOI [10.5958/0976-058X.2014.00658.1](https://doi.org/10.5958/0976-058X.2014.00658.1).
- Palace V, Brown S, Baron C, Fitzsimons J, Woodin B, Stegeman J, Klaverkamp J. 1998.** An evaluation of the relationships among oxidative stress, antioxidant vitamins and early mortality syndrome (EMS) of lake trout (*Salvelinus namaycush*) from Lake Ontario. *Aquatic Toxicology* **43**:195–208 DOI [10.1016/S0166-445X\(97\)00107-0](https://doi.org/10.1016/S0166-445X(97)00107-0).
- Ramchander S, Pillai MA, Ushakumari R. 2014.** Determination of lethal dose and effect of ethyl methane sulphonate in rice varieties. *Trends in Biosciences* **7**:1151–1156.
- Ramesh M, Vanniarajan C, Ravikesavan R, Aiyana KEA, Mahendran P. 2019.** Determination of lethal dose and effect of EMS and gamma ray on germination percentage and seedling parameters in barnyard millet variety Co (Kv) 2. *Electronic Journal of Plant Breeding* **10**:957–962 DOI [10.5958/0975-928X.2019.00123.6](https://doi.org/10.5958/0975-928X.2019.00123.6).
- Rashid M, Ren-hu L, Wei J, Yong-han X, Fu-lin W, Yue-zhi T, Jun-mei W, Cheema AA, Jin-qing C, He G. 2009.** Genomic diversity among Basmati rice (*Oryza sativa* L) mutants obtained through ⁶⁰Co gamma radiations using AFLP markers. *African Journal of Biotechnology* **8**(24):6777–6783 .
- Saba N, Mirza B. 2002.** Ethyl methane sulfonate induced genetic variability in *Lycopersicon esculentum*. *International Journal of Agriculture And Biology* **4**:89–92.
- Serrat X, Esteban R, Guibourt N, Moysset L, Nogués S, Lalanne E. 2014.** EMS mutagenesis in mature seed-derived rice calli as a new method for rapidly obtaining TILLING mutant populations. *Plant Methods* **10**:1–14 DOI [10.1186/1746-4811-10-1](https://doi.org/10.1186/1746-4811-10-1).
- Shah TM, Mirza JI, Haq MA, Atta BM. 2008.** Radio sensitivity of various chickpea genotypes in M1 generation I-Laboratory studies. *Pakistan Journal of Botany* **40**:649–665.
- Shang J, Chun Y, Li X. 2021.** Map-based cloning and natural variation analysis of the PAL3 gene controlling panicle length in rice. *Chinese Bulletin of Botany* **56**:520–532.
- Shelar A, Singh AV, Maharjan RS, Laux P, Luch A, Gemmati D, Tisato V, Singh SP, Santilli MF, Shelar A. 2021.** Sustainable agriculture through multidisciplinary seed nanoprimering: prospects of opportunities and challenges. *Cells* **10**:2428 DOI [10.3390/cells10092428](https://doi.org/10.3390/cells10092428).
- Siddiqui S, Singh S. 2010.** Induced genetic variability for yield and yield traits in Basmati rice. *World Journal of Agricultural Sciences* **6**:331–337.
- Singh AP, Pandey BK, Deveshwar P, Narnoliya L, Parida SK, Giri J. 2015.** JAZ repressors: potential involvement in nutrients deficiency response in rice and chickpea. *Frontiers in Plant Science* **6**:975.
- Singh H, Khar A, Verma P. 2021.** Induced mutagenesis for genetic improvement of Allium genetic resources: a comprehensive review. *Genetic Resources and Crop Evolution* **68**:2669–2690 DOI [10.1007/s10722-021-01210-8](https://doi.org/10.1007/s10722-021-01210-8).

- Sood S, Jambulkar S, Sood A, Gupta N, Kumar R, Singh Y. 2016.** Median lethal dose estimation of gamma rays and ethyl methane sulphonate in bell pepper (*Capsicum annuum* L.). *SABRAO Journal of Breeding and Genetics* **48**:528–535.
- Taheri S, Abdullah TL, Jain SM, Sahebi M, Azizi P. 2017.** Tilling, high-resolution melting (HRM), and next-generation sequencing (NGS) techniques in plant mutation breeding. *Molecular Breeding* **37**:1–23 DOI [10.1007/s11032-016-0586-4](https://doi.org/10.1007/s11032-016-0586-4).
- Talebi AB, Talebi AB, Shahrokhifar B. 2012.** Ethyl methane sulphonate (EMS) induced mutagenesis in Malaysian rice (cv. MR219) for lethal dose determination. *American Journal of Plant Sciences* **3**(12):1661–1665 DOI [10.4236/ajps.2012.312202](https://doi.org/10.4236/ajps.2012.312202).
- Thakur G, Paul S, Kumar A. 2020.** Mutagenic effectiveness and efficiency of ethyl methane sulphonate (EMS) mutagen in linseed (*Linum usitatissimum* L.). *Indian Society of Oilseeds Research* **37**:260–266.
- Unan R, Deligoz I, Al-Khatib K, Mennan H. 2022.** Protocol for ethyl methane-sulphonate (EMS) mutagenesis application in rice. *Open Research Europe* **1**:19 DOI [10.12688/openreseurope.13317.3](https://doi.org/10.12688/openreseurope.13317.3).
- Upadhyaya NM, Bhat RS, Upadhyaya NM, Chaudhury A, Raghavan C, Qiu F, Wang H, Wu J, McNally K, Leung H. 2007.** Chemical-and irradiation-induced mutants and TILLING. In: *Rice functional genomics: challenges, progress and prospects*. New York, NY: Springer.
- Usharani K, Kumar CA. 2015.** Induced polygenic variability using combination treatment of gamma rays and ethyl methane sulphonate in blackgram (*Vigna mungo* (L.) Hepper). *African Journal of Biotechnology* **14**:1702–1709 DOI [10.5897/AJB2015.14438](https://doi.org/10.5897/AJB2015.14438).
- Viana VE, Pegoraro C, Busanello C, Oliveira ACostade. 2019.** Mutagenesis in rice: the basis for breeding a new super plant. *Frontiers in Plant Science* **10**:1326 DOI [10.3389/fpls.2019.01326](https://doi.org/10.3389/fpls.2019.01326).
- Štolfa I, Pfeiffer TŽ, Špoljarić D, Teklić T, Lončarić Z. 2015.** Heavy metal-induced oxidative stress in plants: response of the antioxidative system. In: *Reactive oxygen species and oxidative damage in plants under stress*. New York, NY: Springer, Cham.
- Zaghum MJ, Ali K, Teng S. 2022.** Integrated genetic and omics approaches for the regulation of nutritional activities in rice (*Oryza sativa* L.). *Agriculture* **12**:1757 DOI [10.3390/agriculture12111757](https://doi.org/10.3390/agriculture12111757).
- Zahra S, Hussain M, Zulfiqar S, Ishfaq S, Shaheen T, Akhtar M. 2021.** EMS-based mutants are useful for enhancing drought tolerance in spring wheat. *Cereal research communications* **50**:767–778 DOI [10.1007/s42976-021-00220-7](https://doi.org/10.1007/s42976-021-00220-7).
- Zargar SM, Mir RA, Ebinezer LB, Masi A, Hami A, Manzoor M, Salgotra RK, Sofi NR, Mushtaq R, Rohila JS. 2022.** Physiological and multi-omics approaches for explaining drought stress tolerance and supporting sustainable production of rice. *Frontiers in Plant Science* **12**:3242 DOI [10.3389/fpls.2021.803603](https://doi.org/10.3389/fpls.2021.803603).
- Zeng C-L, Liu L, Xu G-Q. 2011.** The physiological responses of carnation cut flowers to exogenous nitric oxide. *Scientia Horticulturae* **127**:424–430 DOI [10.1016/j.scienta.2010.10.024](https://doi.org/10.1016/j.scienta.2010.10.024).

Zhang K, Zhang Y, Sun J, Meng J, Tao J. 2021. Deterioration of orthodox seeds during ageing: Influencing factors, physiological alterations and the role of reactive oxygen species. *Plant Physiology and Biochemistry* **158**:475–485
[DOI 10.1016/j.plaphy.2020.11.031](https://doi.org/10.1016/j.plaphy.2020.11.031).