

# Effects of different plant parts of invasive *Solidago* species on the germination and growth of native grassland plant species

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Allelopathy is an important factor influencing whether an invasive plant species can become successfully established in a new range through disrupting the germination and growth of native plant species. Goldenrods (*Solidago* species) are one of the most widespread invasive taxa in Central Europe of North American origin. Owing to their high environmental impact and wide distribution range, invasive *Solidago* species should be controlled in Europe, and the areas invaded by them should be restored. Numerous studies have reported the allelopathic effects of *Solidago gigantea* and *Solidago canadensis*, but the results are inconsistent regarding differences in the allelopathic effects of particular plant parts and in the sensitivity to *Solidago* allelopathic effects among native species as well as between the two invasive species themselves. In this study, we aimed to analyse the effect of water extracts from *S. canadensis* and *S. gigantea* parts (roots, rhizomes, stems, leaves, and inflorescences) on the germination and initial growth of seedlings of 13 grassland species that typically grow in Central Europe. The tested grassland species differed in susceptibility to *Solidago* allelopathy, with the most resistant species being *Daucus carota*, *Leucanhemum vulgare*, *Lolium perenne* and *Trifolium pratense*. The inhibitory effect of 10% water extracts from leaves and flowers was stronger than those from rhizomes, roots, and stems without leaves, regardless of the *Solidago* species. Our study results imply that reducing the allelopathic effect of *Solidago* during habitat restoration requires removal of the aboveground parts, including fallen leaves. The allelopathic effects of roots and rhizomes seem to be of secondary importance.

1 **Effects of different plant parts of invasive *Solidago* species on the germination and growth**  
2 **of native grassland plant species**

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28 **Abstract**

29 Allelopathy is an important factor influencing whether an invasive plant species can become  
30 successfully established in a new range through disrupting the germination and growth of native  
31 plant species. Goldenrods (*Solidago* species) are one of the most widespread invasive taxa in  
32 Central Europe of North American origin. Owing to their high environmental impact and wide  
33 distribution range, invasive *Solidago* species should be controlled in Europe, and the areas  
34 invaded by them should be restored. Numerous studies have reported the allelopathic effects of  
35 *Solidago gigantea* and *Solidago canadensis*, but the results are inconsistent regarding differences  
36 in the allelopathic effects of particular plant parts and in the sensitivity to *Solidago* allelopathic  
37 effects among native species as well as between the two invasive species themselves. In this  
38 study, we aimed to analyse the effect of water extracts from *S. canadensis* and *S. gigantea* parts  
39 (roots, rhizomes, stems, leaves, and inflorescences) on the germination and initial growth of  
40 seedlings of 13 grassland species that typically grow in Central Europe. The tested grassland  
41 species differed in susceptibility to *Solidago* allelopathy, with the most resistant species being  
42 *Daucus carota*, *Leucanhemum vulgare*, *Lolium perenne* and *Trifolium pratense*. The inhibitory  
43 effect of 10% water extracts from leaves and flowers was stronger than those from rhizomes,  
44 roots, and stems without leaves, regardless of the *Solidago* species. Our study results imply that  
45 reducing the allelopathic effect of *Solidago* during habitat restoration requires removal of the  
46 aboveground parts, including fallen leaves. The allelopathic effects of roots and rhizomes seem  
47 to be of secondary importance.

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49 **Key words:** allelopathy, grassland restoration, inhibition, goldenrods, land reclamation, plant  
50 invasion, semi-natural grasslands

## 51 Introduction

52 Allelopathy involves the production of secondary metabolite biochemical substances by  
53 one plant that stimulate or inhibit the germination, growth, and development of adjoining or  
54 neighbouring organisms (Rice, 1984; Cheema, Farooq & Khaliq, 2013; Bachheti et al., 2020; Li  
55 et al., 2021). The allelochemicals are present in various plant tissues, primarily inside the cells  
56 composing the various plant parts, such as leaves, stems, pollen, flowers, fruits, and roots  
57 (Begum et al., 2019; Macías, Mejías & Molinillo, 2019; Bachheti et al., 2020). Allelochemicals  
58 can be released to the environment through leaching from leaves and other aboveground plant  
59 parts, volatilization, root exudation, and litter decomposition (Uddin & Robinson, 2017; Wang et  
60 al., 2021).

61 Allelopathy plays a significant role in both natural and agricultural ecosystems by  
62 influencing seed germination and the growth of seedlings (Chon, Kim & Lee, 2003; Mushtaq &  
63 Siddiqui, 2018). The inhibition of plant growth caused by allelopathy differs based on the plant  
64 tissue (e.g., leaves, stems, roots) from which the allelopathic compounds are released (Begum et  
65 al., 2019; Kato-Noguchi & Kato, 2022). Most research shows that leaf extracts have stronger  
66 effects than those from flowers, stems, and roots (Turk & Tawaha, 2003; Siddiqui, Bhardwaj &  
67 Meghvanshi, 2009; Sodaeizadeh et al., 2009; Meiners, 2014; Debnath, Debnath & Paul, 2016;  
68 Mushtaq & Siddiqui, 2018; Mangao et al., 2020), however, it is not absolute and the effect of  
69 belowground parts can sometimes be stronger (Li & Jin, 2010; Zivanai, Ronald & Nester, 2019).  
70 The sensitivity to allelochemicals differs considerably among plant species (Debnath, Debnath &  
71 Paul, 2016; Sekutowski et al., 2019; Mangao et al., 2020) and even among genotypes within a  
72 species (Meiners, 2014; Appiah, Amoatey & Fujii, 2015). Most allelopathy studies have focused  
73 on the interactions between weeds and crops and have described the negative impacts of weeds  
74 on crops (Turk & Tawaha, 2003; Mangao et al., 2020). Allelopathy studies have also focused on  
75 aspects of eco-friendly agriculture such as the synthesis of agrochemicals to control pests and  
76 diseases, especially in weed management as an alternative to synthetic herbicides (Chon, Kim &  
77 Lee, 2003; Macías, Mejías & Molinillo, 2019; Bachheti et al., 2020; Mangao et al., 2020; Li et  
78 al. 2021; Motmainna et al., 2021, Ullah, Khan & Khan, 2021).

79 Allelopathy also plays a significant role in the successful establishment and survival of  
80 invasive plant species in the ecosystems of new ranges owing to potential for interfering with the

81 seed germination, seedling growth, development, and establishment of native plant species  
82 (Uddin & Robinson, 2017; Torawane & Mokot, 2020). The allelopathic effect of some invasive  
83 species can be so strong that introduced has been so-called ‘novel weapon hypothesis’ as an  
84 explanation for invasiveness (Callaway & Ridenour, 2004). The novel weapons hypothesis  
85 explains that the invasion efficiency of an exotic plant species can involve novel biochemical  
86 weapons that act as very strong allelopathic agents against resident vegetation. These agents give  
87 the invaders an advantage that arises from differences in the coevolutionary histories of plant  
88 communities. Allelopathic substances can be relatively ineffective against natural neighbours in  
89 the native range of an invader because they are adapted to its presence; however, newly  
90 encountered plants in invaded communities lack that adaptation. The exploitation by invaders of  
91 the susceptibility of resident species to the allelopathic effect due to evolutionary inadequacy is  
92 known as the advantage against resident species hypothesis (Callaway & Ridenour, 2004; Awty-  
93 Carroll et al., 2020). The novel weapons hypothesis explains successful invasions due to  
94 allelopathy in cases such as *Centaurea diffusa*, *Centaurea maculosa*, *Mikania micrantha*, and  
95 *Alliaria petiolata* in America and China (Callaway & Ridenour, 2004; Chen et al., 2017) as well  
96 as *Solidago canadensis* in China and Europe (Chen et al., 2017; Wei et al., 2020a; Wei et al.,  
97 2020b, Wei et al., 2020c) and *Solidago gigantea* in Europe (Pal et al., 2015).

98 In this study, we focussed on two invasive species, *Solidago gigantea* Aiton (Giant  
99 goldenrod) and *Solidago canadensis* L. (Canadian goldenrod), which are the most widespread  
100 invasive species of North American origin in Central Europe (Meyer, 2022; Popay & Parker  
101 2022). These *Solidago* species have strong negative environmental impacts due to competition  
102 for soil nutrients, water, and space, as well as inhibition of native plants through allelopathy  
103 (Ledger et al., 2015). Because of clonal growth, *Solidago* species form dense stands and decrease  
104 the biodiversity of plants (Pal et al., 2015; Hejda, Pyšek & Jarošík, 2009), arthropods (Moroń et  
105 al., 2009; Lenda et al., 2021), ants (Kajzer-Bonk, Szpiłyk & Woyciechowski, 2016), and birds  
106 (Skórka, Lenda & Tryjanowski, 2010). The alien *Solidago* are able to invade grasslands,  
107 especially recently abandoned once (Moroń et al., 2009; Fenesi et al., 2015; Czarniecka-Wiera et  
108 al., 2020; Szymura, Świerszcz & Szymura, 2022), strongly influencing their plant species  
109 richness and pollinators abundance (Moroń et al., 2009; Fenesi et al., 2015).

110 Ample evidence exists regarding allelopathic impact of *Solidago* species on crops and  
111 forage grass species. For example, *S. canadensis* extracts can inhibit seed germination and  
112 growth performance parameters of *Lactuca sativa* (Wang, Wu & Jiang, 2019; Wei et al., 2020a;  
113 Wei et al., 2020b), *Trifolium pratense* (Zandi et al., 2020), *Pterocypselia laciniata* (Wang et al.,  
114 2017), and *Festuca rubra* and *Festuca pratensis* (Karpavičiene, Daniloviene & Vykertaite,  
115 2019). In addition, *S. gigantea* decreases the germination and growth performance parameters of  
116 *Avena sativa*, *Brassica napus* subsp. *oleifera*, and *Helianthus annuus* (Novak et al., 2018).  
117 Moreover, root extracts from *Solidago* species show inhibitory activity against microorganisms  
118 (Móricz et al., 2020; Móricz et al., 2021). Finally, the allelopathy of *S. canadensis* can reduce the  
119 biodiversity of species-rich plant communities and thus increase the susceptibility of a  
120 community to further invasion (Ledger et al., 2015; Adomako et al., 2019).

121 The content and type of phenolic compounds differ in plant tissues, including leaves,  
122 flowers, stem, and roots, and the characteristics can also be variable between *S. canadensis* and  
123 *S. gigantea* (Marksa et al., 2020; Kato-Noguchi & Kato, 2022; Zhu et al., 2022). Further, even in  
124 the same species, the concentration of allelochemicals can differ based on location (e.g., native  
125 vs. invasive range). For example, in the case of *S. canadensis*, samples from China had higher  
126 allelochemical contents (total phenolics, total flavones, and total saponins) and stronger  
127 allelopathic effects than samples from North America (Yuan et al., 2013).

128 Because of the high environmental impact, wide distribution range, and locally high  
129 abundance of invasive *Solidago* species, they should be controlled in Europe (Sheppard, Shaw &  
130 Sforza, 2006; Fenesi et al., 2015; Tokarska-Guzik et al., 2015). Furthermore, the habitats invaded  
131 by them should be restored (Nagy et al., 2020; Szymura, Świerszcz & Szymura, 2022). A  
132 reasonable direction of post-invaded ground restoration is species-rich grasslands (Szymura,  
133 Świerszcz & Szymura, 2022). Therefore, the selection of grassland species that are resistant to  
134 the allelopathic effects of *Solidago* species seems to be important for effective restoration of  
135 *Solidago*-invaded lands. The aim of this study was to evaluate the allelopathic effect of *S.*  
136 *canadensis* and *S. gigantea* plant parts (leaves, stems, inflorescences, roots, and rhizomes) on  
137 native grassland species in Central Europe. We hypothesised that (1) the native grassland species  
138 differ in terms of seed germination and seedling growth under the allelopathic effects of invasive  
139 *Solidago* species, (2) the impacts on germination and growth of native species vary depending on

140 the *Solidago* plant parts used to create water-based extracts, and (3) *S. canadensis* and *S.*  
141 *gigantea* differ in terms of their allelopathic influence on native grassland species. Additionally,  
142 we also considered the impact on native species from possible interactions between the three  
143 factors (grassland species, extracts of different parts of *Solidago* plants, and *Solidago* species).

## 144 **Material and methods**

### 145 **Studied species**

146 *Solidago canadensis* and *S. gigantea*, members of the Asteraceae family, are clonal  
147 perennial herbs that can form rhizomes. The inflorescences are fasciculate and thyrsoid, the  
148 capitula are small and numerous, and the florets are yellow (McNeil, 1976). Alien *Solidago*  
149 species occur in soils with a wide range of fertility and moisture levels, creating single-species  
150 stands or co-occurring with each other (Weber, 2001; Weber and Jacobs, 2005). The large-range  
151 dispersal is realised by numerous, wind-dispersed seeds, whereas short-range dispersal involves  
152 vegetative growth through rhizomes (Weber, 2001). The species are able to create large stands in  
153 abandoned fields and meadows, riparian habitats, forest edges, and unmowed road verges  
154 (Weber, 2001; Weber and Jacobs, 2005; Fenesi et al., 2015).

155 *Solidago* species contain bioactive compounds such as cytotoxic compounds, phenolic  
156 compounds, and flavonoids (Wandjou et al., 2020; Shelepova et al., 2020; Kato-Noguchi &  
157 Kato, 2022). Twenty-three phenolic compounds of different phenolic origin were identified in  
158 the leaves and inflorescences of *Solidago* species, and *S. gigantea* was found to have higher  
159 amounts of the compounds than *S. canadensis* (Marksa et al., 2020). Essential oils from *S.*  
160 *canadensis*, composed mainly of mono- and sesquiterpene hydrocarbons, also have phytotoxic  
161 potential (Synowiec et al., 2017). The highest concentration of phenolic compounds in *S.*  
162 *canadensis* was found in leaves during the blooming stage and in roots during the early growing  
163 stage (Baležentienė, 2015).

164

### 165 **Plant material**

166 For each *Solidago* species in the current study, we investigated the impact of different  
167 plant parts, using aqueous solutions of dried and ground plant material (Fig. S1). The roots,  
168 rhizomes, stems, and leaves were collected in July 2020 and the inflorescences in September

169 2020 in Wrocław, Poland (51°05'57.3"N, 17°04'39.0"E; 51°09'43.7"N, 17°06'54.0"E; and  
170 51°09'40.9"N, 17°06'40.7"E). Thirteen grassland species native to Europe were used in the study  
171 (Table 1). These species are typical and widespread in semi-natural grasslands in Central Europe,  
172 are important for pollinators, and grow in similar environmental conditions as *Solidago* species.  
173 We used species from different plant families, concentrating on species from *Poaceae* and  
174 *Fabaceae* families that are the most common in grasslands. The seeds were obtained from  
175 Rieger-Hofmann® GmbH company in a ready-to-use form that did not require additional  
176 treatments (e.g., freezing) before sowing.

### 177 **Allelopathic bioassay**

178         Extracts from the different plant parts of the two *Solidago* species were prepared as 10%  
179 aqueous solutions. The concentration was based on previous allelopathic experiments that  
180 identified a concentration representative of a high degree of invasion (Butcko & Jensen, 2002;  
181 Ravlić, Baličević, & Peharda, 2015; Novak et al., 2018; Sekutowski et al., 2019). To create the  
182 solutions, powdered material (10 g) from each part of the plants was mixed with distilled water  
183 (100 ml). The mixtures were set aside in the dark for 24 h at room temperature (20-25 °C) and  
184 then filtered, using filter paper to remove plant residues from the solutions (Fig. S2).

185         For the experiment, Petri dishes (78.54 cm<sup>2</sup>) with two layers of filter paper were sterilized  
186 for 3 h at 120 °C before use. For each grassland species, a sample of 50 seeds was sterilized with  
187 1% NaClO for 15 min, rinsed three times in distilled water, and placed on filter paper to remove  
188 excess water. Each sample was then sown onto a Petri dishes and soaked with 8 ml of an extract  
189 based on a *Solidago* plant part or with distilled water in the case of the control treatment. Finally,  
190 the dishes were closed and placed in a growth chambers (Versatile Environmental Test  
191 Chambers SANYO, Model - MLR-352H and FRIOCELL, Model – FC 404 EVO) at 20 °C/10 °C  
192 temperature (day/night), with 150  $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$  photosynthetic photon flux density and relative  
193 humidity of approximately 70% for 21 days. During the growth period, the Petri dishes were  
194 watered with distilled water every 3 days (Fig. S3). Four control treatments for each grassland  
195 species and four replications of a particular combination of grassland species (13), *Solidago*  
196 species (2), and *Solidago* plant part (5) were prepared (572 Petri dishes in total). For technical  
197 reasons, the trials for each species were conducted separately. The experiment was conducted at

198 the Institute of Agroecology and Plant Production, Wrocław University of Environmental and  
199 Life Sciences, Poland from October 2020 to September 2021.

## 200 **Measurements**

201 After 21 days, the experiment was terminated, the number of germinated seeds were calculated.  
202 Ten random seedlings were selected for each trial, with fewer seedling selected if germination  
203 was poor. The hypocotyl (stem) and root lengths (cm) of the seedlings were measured, using a  
204 linear scale and a binocular microscopy. Afterward, the total fresh mass of 10 seedlings from  
205 each trial was weighed in grams.

## 206 **Data analysis**

207 The germination percentage of seeds (GP) was determined for each Petri dish separately by using  
208 the following formula (1) (International Seed Testing Association, 1985):

$$209 \quad GP \% = \frac{\text{Number of seeds germinated}}{\text{Total number of seeds plated}} \times 100$$

210 The effect of inhibition (EI) on germination, shoot and root length, and seedling weight were  
211 calculated in comparison with the control, according to the following formula (2) (Hsu & Chou,  
212 1992):

$$213 \quad EI = \frac{C - T}{C} \times 100$$

214 where C is the measurement associated with the control for a particular species and T is the  
215 measurement associated with a particular treatment. Positive values of EI indicated inhibition,  
216 values around zero revealed a lack of effect, and negative values showed that treatment had a  
217 positive effect on germination or growth.

218 In statistical analysis, to reduce the pseudo-replication problem (Morrison & Morris,  
219 2000) we considered a Petri dish to be the smallest, independent sample unit; therefore, results of  
220 individual measurements were averaged per dish. Additionally, strong germination inhibition in  
221 many cases precluded performing measurements other than an assessment of seed germination  
222 because of a lack of seedlings. To focus on the main output of the evaluation, we set the values  
223 of inhibition in such cases to 100% for statistical analyses. Correlations between measured traits

224 were checked using Spearman's rank correlations coefficient using Past software (Hammer,  
225 Harper & Ryan, 2001). Differences in average values between groups were analysed within a  
226 generalised linear model framework with glm2 (Marschner, 2011) package in the R environment  
227 (R Core Team, 2022), with assumed normal distribution and identity link. The significance of  
228 differences between treatments, including the interactions, were tested using analysis of deviance  
229 with F test using the stats package (R Core Team, 2022). For the post hoc tests, emmeans (Lenth,  
230 2022) and multcomp (Hothorn, Bretz & Westfall, 2008) packages were used with applied  
231 Bonferroni corrections.

232 To clarify the effect of the experiment on a particular plant species, we also performed  
233 additional analyses using the leaves and flowers of both *Solidago* species together. These  
234 analyses were prompted by observed significant differences between the leaves and flowers and  
235 the remaining plant parts; the differences were the same for both *Solidago* species (see Results).  
236 To better visualise the differences between particular species, we applied discriminant analysis,  
237 also considering only the effect of flowers and leaves of both *Solidago* species in Past software  
238 (Hammer, Harper & Ryan, 2001).

## 239 Results

240 In general, we observed that extracts had a negative effect on seed germination, which  
241 was expressed as positive values of inhibition effect (Fig. 1). There were significant correlations  
242 between measured traits, showing that inhibition of germination usually also reduces the biomass  
243 and the shoot and root lengths of the studied species (Table 2, Fig. 3).

244 We observed that all experimental treatments and their interactions significantly affected  
245 all germination and growth characteristics. The results of statistical comparisons for the entire  
246 experiment are shown in Table 3. The detailed results, with *post hoc* comparisons, are presented  
247 in the Appendix (Tables S1-S4). The results consistently show that leaves and flowers cause  
248 stronger inhibition compared with roots, rhizomes, and stems (Fig. 1 and Fig. S4-S7), regardless  
249 of *Solidago* species. Considering only the effects of leaf and flower extracts, we observed  
250 significant differences between examined species in germination (Fig. 2) as well as other  
251 measured traits (Fig. 3). The species *Trifolium pratense*, *Daucus carota*, *Lolium perenne*, and  
252 *Leucanhemum vulgare* were more resistant to the allelopathic effects of *Solidago*, while  
253 *Campanula patula*, *Lychnis flos-cuculi*, *Lotus corniculatus*, *Trifolium repens*, *Festuca*

254 *arundinacea*, *Festuca pratensis*, *Festuca rubra*, *Phleum pratense*, and *Poa pratensis* were more  
255 sensitive (Fig. 2 and 3).

256 In spite of the observed germination inhibition, extracts from root, rhizome, and stem for  
257 some tested species slightly enhanced the growth of shoots (*Trifolium repens*, *Lychnis flos-*  
258 *cuculi*, *Phleum pratense*, *Trifolium pratense*, *Lolium perenne*, *Leucanhemum vulgare*, *Daucus*  
259 *carota*, *Festuca arundinacea*, and *Festuca pratensis*) and roots (*Daucus carota*, *Lotus*  
260 *corniculatus*, and *Trifolium pratense*) as well as the biomass (*Phleum pratense*, *Festuca*  
261 *arundinacea*, *Trifolium repens*, and *Trifolium pratense*) (Fig. S4-S7).

262

## 263 Discussion

264 Numerous previous studies report the effect of water extracts from *S. gigantea* and *S.*  
265 *canadensis* on the germination and growth of other plants. Most of the studies focussed on *S.*  
266 *canadensis* (e.g. Wang, Wu & Jiang, 2019; Wei et al., 2020a; Wei et al., 2020c; Zandi et al.,  
267 2020), while a lower number focussed on *S. gigantea* (e.g. Pal et al., 2015; Baličević, Ravlić &  
268 Živković, 2015; Sekutowski et al., 2019). The results are inconsistent because target species,  
269 extract concentrations, and tissues from which the extracts were produced varied between  
270 studies.

271 Our results revealed that the tested grassland species differ in susceptibility to *Solidago*  
272 allelopathy, which confirms our first hypothesis. The most resistant species in our experiment  
273 were *Daucus carota*, *Leucanhemum vulgare*, *Lolium perenne*, and *Trifolium pratense*. Chen, Mei  
274 and Tang (2005) and Megenhardt (2015) observed that grasses were more sensitive than forbs  
275 and legumes to the allelopathic impact of *S. canadensis*. However, in our experiment we did not  
276 observe such a pattern, and between *Trifolium repens* and *T. pratense*, legumes belonging to the  
277 same genus, *T. pratense* was more resistant than *T. repens*. Species detected here as being more  
278 resistant to the allelopathic effects of *Solidago* are considered useful in restoring semi-natural  
279 grasslands (da Silva, Overbeck & Soares, 2017; Thiébaud, Tarayre & Rodríguez-Pérez, 2019;  
280 Zandi et al., 2020). The obtained results suggest that the above-mentioned species should be  
281 prioritised when grasslands are restored on sites invaded by *Solidago*.

282 Different parts of *Solidago* were assumed to differ in their allelopathic effect (Marksa et  
283 al., 2020; Kato-Noguchi & Kato, 2022; Zhu et al., 2022). Allelopathic studies have most often  
284 used aboveground parts such as leaves, stems, and flowers, with leaves being most commonly  
285 found to have an effect (Wang, Wu & Jiang, 2019; Wei et al., 2020a; Wei et al., 2020c; Zandi et  
286 al., 2020; Kato-Noguchi & Kato, 2022; Zhu et al., 2022). Direct comparisons revealed that  
287 aboveground parts of *S. canadensis* had a significant allelopathic effect, but the effect of  
288 belowground parts was not significant (Yu et al., 2022). It was also observed that extracts from  
289 *S. canadensis* rhizome stimulated the germination of *Raphanus sativus* seeds and lengthened  
290 their shoots (Anžlovar & Anžlovar, 2012). In contrast, other studies showed that *S. canadensis*  
291 rhizome extract inhibited seed germination and root growth of several native Chinese plant  
292 species (Chen, Mei & Tang, 2005) and *Zoysia japonica* (Sun et al., 2022). In the case of *S.*  
293 *gigantea*, water extracts of its rhizomes and roots increases the dry biomass of *Echinochloa crus-*  
294 *galli* and *Amaranthus retroflexus* (Sekutowski et al., 2019), while leaf and stem extracts reduced  
295 the growth of *E. crus-galli*. However, Pal et al. (2015) showed that root extracts reduced the  
296 shoot growth of plant species native to Europe. The observed differences could be due to  
297 different reactions of specific target species as well as different concentrations of the extracts.  
298 When the extract was at a low concentration (1%), it could even increase the growth of lettuce  
299 (Wang, Wu & Jiang, 2019; Wei et al., 2020b). In addition, Ye et al. showed that a water extract  
300 of *S. canadensis* shoots up to a concentration of 12.5% could increase the growth of *Zea mays*  
301 (Ye, Meng, & Wu, 2019). However, it should be noted that *Z. mays* seeds are exceptionally  
302 large, and the allelopathic effect could be weakened due to good isolation of the embryo. In our  
303 study, 10% extracts of leaves and flowers consistently showed stronger effects compared with  
304 extracts of rhizomes, roots, and stems without leaves, regardless of the *Solidago* species used  
305 (Fig. 1). Additional noise in data could be also related to differences in the allelopathic effect of  
306 leaves and stems. If stems with leaves are used for preparing solutions (e.g. Baličević, Ravlić &  
307 Živković, 2015; Sekutowski et al., 2019; Ye, Meng, & Wu, 2019), then the uncontrolled  
308 proportion of leaf versus stem biomass could change the results obtained. There is great  
309 variability in the chemical composition of potentially allelopathic substances between particular  
310 parts, seasons and geographical locations in the case of *Solidago* species (See Marksa et al.,  
311 2020; Kato-Noguchi & Kato, 2022; Zhu et al., 2022). Therefore, it is impossible to attribute the

312 allelopathic effect observed here to particular chemical substances and differences in their  
313 concentrations between plant parts.

314         The results also seem to confirm the general hypothesis that allelopathic effects differ  
315 between the examined *Solidago* species (Table 2); however, we also observed strong effects  
316 associated with interactions, which impeded making straightforward conclusions. Previous  
317 studies reported inhibition of seed germination and seedling growth in numerous plant species  
318 caused by allelopathic effects of both *S. canadensis* (Wang, Wu & Jiang, 2019; Wei et al.,  
319 2020a; Wei et al., 2020c; Zandi et al., 2020) and *S. gigantea* (Pal et al., 2015; Baličević, Ravlić  
320 & Živković, 2015); however, Marksa et al. (2020) found that the leaves of *S. gigantea* contained  
321 more active antioxidant compounds than leaves of *S. canadensis*, which suggested that the first  
322 species had a stronger allelopathic potential. We assumed that comparing the differences in  
323 allelopathic effects of extracts from various parts of *Solidago* and the differences in the  
324 susceptibility of target species, the differences between the two invasive species would have  
325 minor practical implications. In practice, the results of a pot experiment showed that *S.*  
326 *canadensis* and *S. gigantea* have similar competitive abilities (Szymura & Szymura, 2016).

## 327 **Conclusions and practical implications**

328         The results of our investigation yield some practical implications. First, to reduce the  
329 allelopathic effect of *Solidago* during habitat restoration, the aboveground parts should be  
330 removed, including fallen leaves, since the leaves have a strong allelopathic effect. The effects of  
331 roots and rhizomes seem to be of secondary importance, and the results of other experiments  
332 have shown that restoration is possible without extraction of the belowground parts (Szymura,  
333 Świerszcz & Szymura, 2022). Second, given the expected allelopathic effects of *Solidago*,  
334 relatively resistant species such as *Daucus carota*, *Leucanhemum vulgare*, *Lolium perenne*, and  
335 *Trifolium pratense* should be favoured for site restoration. Third, the difference in allelopathic  
336 effects of leaves versus stems suggest that these two plant parts should be considered separately,  
337 and not mixed, in allelopathic trials. Our results suggest that hard-to-control differences in  
338 fractions of leaves and stem biomass in a plant material used to produce extracts may  
339 significantly influence experimental outcomes.

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**Table 1** (on next page)

Common grassland species used for the experiment.

1 **Table 1.** Common grassland species used for the experiment.

No	Species	Abbreviation	Family
1	<i>Daucus carota</i> L.	DC	Apiaceae
2	<i>Leucanthemum vulgare</i> Lam.	LV	Asteraceae
3	<i>Campanula patula</i> L.	CP	Campanulaceae
4	<i>Lychnis flos-cuculi</i> L.	LF	Caryophyllaceae
5	<i>Lotus corniculatus</i> L.	LC	Fabaceae
6	<i>Trifolium pratense</i> L.	TP	Fabaceae
7	<i>Trifolium repens</i> L.	TR	Fabaceae
8	<i>Festuca arundinacea</i> L.	FA	Poaceae
9	<i>Festuca pratensis</i> Huds.	FP	Poaceae
10	<i>Festuca rubra</i> L.	FR	Poaceae
11	<i>Lolium perenne</i> L.	LP	Poaceae
12	<i>Phleum pratense</i> L.	PhP	Poaceae
13	<i>Poa pratensis</i> L.	PoP	Poaceae

2

**Table 2** (on next page)

Results of Spearman's rank correlations (lower part) and associated p values (upper part) among analysed traits.<sup>a</sup>

<sup>a</sup>Germ\_inhib = seeds germination inhibition; Shoot\_inhib = seedlings shoot length inhibition; Root\_inhib = seedlings root length inhibition; Weight\_inhib = seedlings weight inhibition.

- 1 **Table 2.** Results of Spearman's rank correlations (lower part) and associated p values (upper part)  
 2 among analysed traits.<sup>a</sup>

	<b>Germ_inhib</b>	<b>Weigh_inhib</b>	<b>Root_inhib</b>	<b>Shoot_inhib</b>
<b>Germ_inhib</b>		<0.001	<0.001	<0.001
<b>Weigh_inhib</b>	0.77		<0.001	<0.001
<b>Root_inhib</b>	0.75	0.83		<0.001
<b>Shoot_inhib</b>	0.60	0.69	0.56	

3 <sup>a</sup>Germ\_inhib = seeds germination inhibition; Shoot\_inhib = seedlings shoot length inhibition;

4 Root\_inhib = seedlings root length inhibition; Weight\_inhib = seedlings weight inhibition.

5

**Table 3**(on next page)

Results of statistical comparisons (F and p) for seed germination (Germ\_inhib), inhibition of seedling shoot (Shoot\_inhib), root (Root\_inhib), and weight (Weight\_inhib), and their interactions ( $p < 0.05$ ).<sup>a</sup>

<sup>a</sup>df = degree of freedom.

1 **Table 3.** Results of statistical comparisons (F and p) for seed germination (Germ\_inhib), inhibition  
 2 of seedling shoot (Shoot\_inhib), root (Root\_inhib), and weight (Weight\_inhib), and their  
 3 interactions ( $p < 0.05$ ).<sup>a</sup>

	df	Germ_inhib		Shoot_inhib		Root_inhib		Weight_inhib	
		F	p	F	p	F	p	F	p
<i>Solidago</i>	1	6.3	0.012	13.3	<0.001	4.0	0.045	14.1	<0.001
<b>Part</b>	4	458.9	<0.001	148.8	<0.001	501.7	<0.001	474.4	<0.001
<b>Species</b>	12	149.5	<0.001	67.8	<0.001	98.3	<0.001	63.7	<0.001
<i>Solidago</i> × <b>Part</b>	4	12.5	<0.001	6.1	<0.001	15.5	<0.001	10.9	<0.001
<i>Solidago</i> × <b>Species</b>	12	4.6	<0.001	13.3	<0.001	3.2	<0.001	2.9	0.001
<b>Part</b> × <b>Species</b>	48	12.2	<0.001	6.8	<0.001	13.9	<0.001	8.4	<0.001
<i>Solidago</i> × <b>Part</b> × <b>Species</b>	48	3.4	<0.001	3.9	<0.001	3.7	<0.001	2.6	<0.001

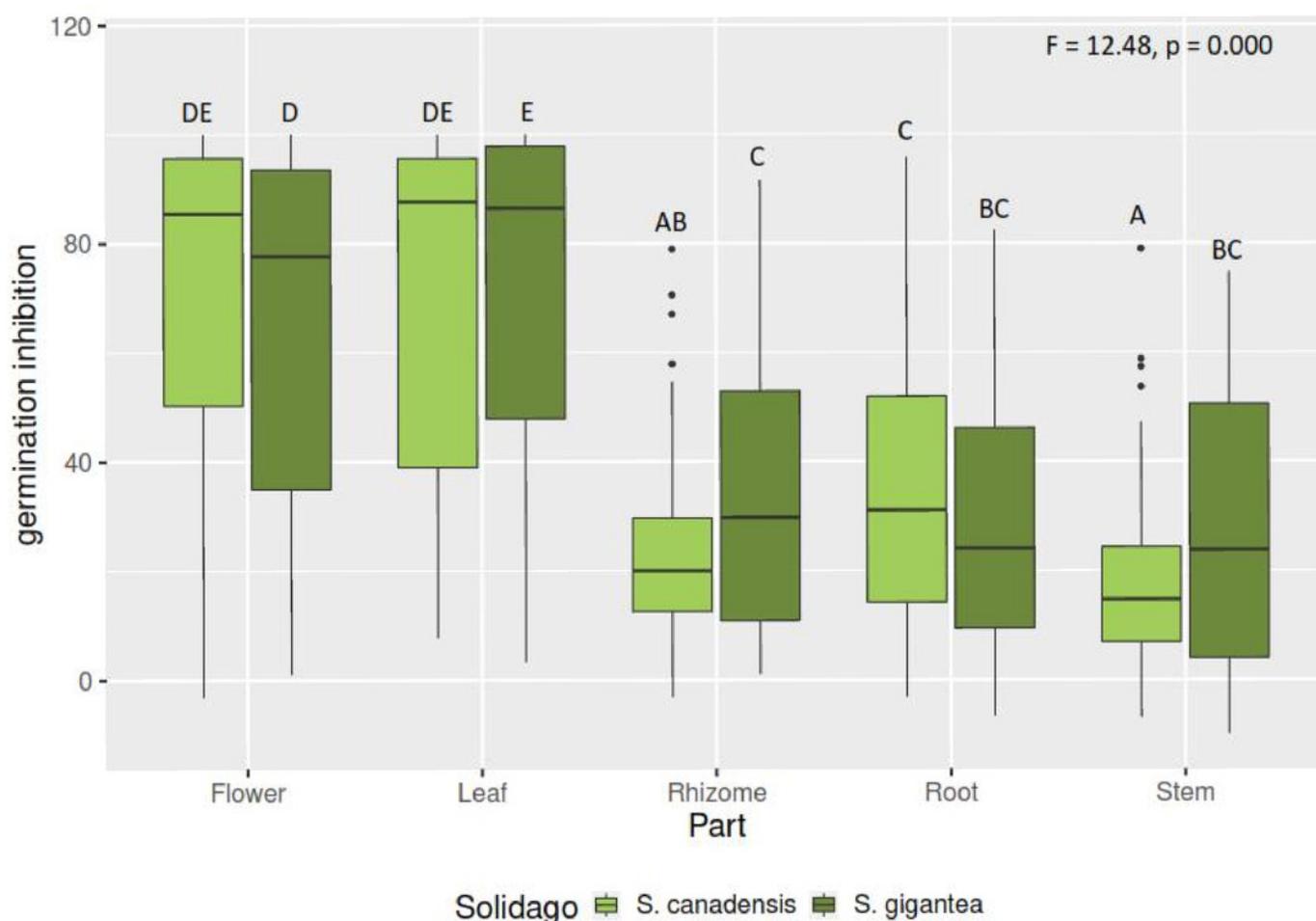
4 <sup>a</sup>df = degree of freedom.

5

## Figure 1

Inhibition of grassland species germination caused by leaf, flower, root, rhizome, and stem extracts of *Solidago* species, and results of tests (F and p).

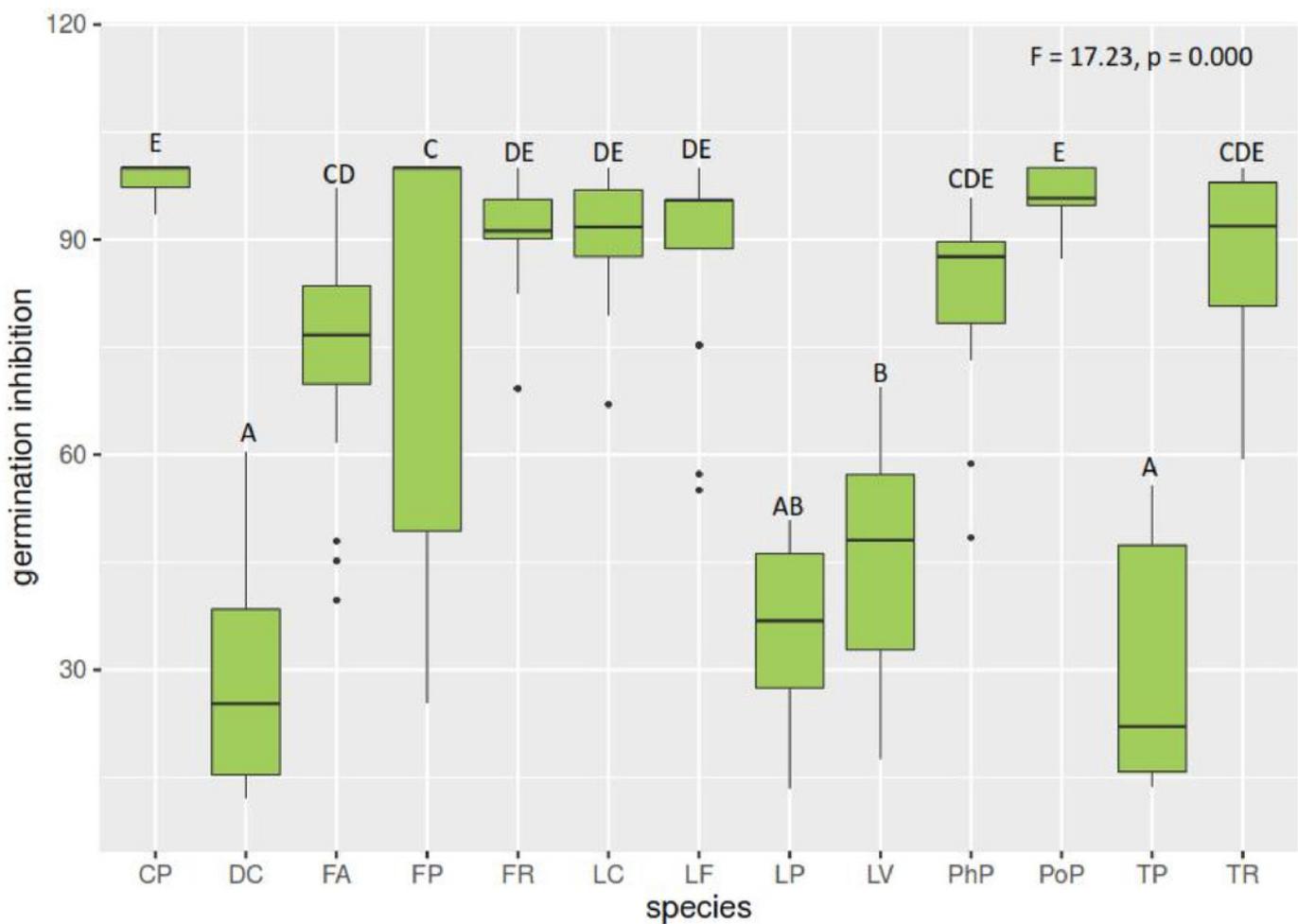
The different letters above boxes indicate significant differences detected by post hoc comparisons.



## Figure 2

Inhibition of grassland species germination caused by *Solidago* allelopathy, and results of tests (F and p).

Inhibition of grassland species germination caused by *Solidago* allelopathy, and results of tests (F and p). The different letters above boxes indicate significant differences detected by post hoc comparisons. Species name abbreviations are presented in Table 1. The graph shows the results for combined leaf and flower extracts only.



## Figure 3

Discriminant analysis for *Solidago* allelopathy inhibition of seed germination, seedling root length, and shoot length and weight of tested grassland species.

Species name abbreviations are presented in Table 1. The graph shows the results for merged leaf and flower extracts only.

