

analysis

2023-03-29

- Importing data
- Building MEMs and testing for spatial autocorrelation in species composition
- making pRDA model

```
library(readr)
library(vegan)
library(tidyverse)
library(usdm)
library(adespatial)
library(adegraphics)
```

Importing data

```
data <- read_csv("peerj-79676-Naas_T-data.csv",
                 col_types = cols(`Wetland type` =
                                   col_factor(levels = c("Temp vle",
                                   "Large dam", "Small dam", "River edge",
                                   "Fynbos pool")), temporary = col_factor(levels = c("temp",
                                   "perm")), Fish = col_factor(levels = c("no-fish",
                                   "fish"))))

data #wetland type, hydroperiod, and fish treated as factors
```

```
## # A tibble: 50 × 29
##   Site Site ...1 Date ...2 Latit...3 Longi...4 Wetla...5 tempo...6 catch...7 area perim...8
##   <chr> <chr> <dbl> <dbl> <dbl> <fct> <fct> <chr> <dbl> <dbl>
## 1 Site_01 Zilver... 170803 -34.3 19.2 Temp v... temp G40L 6169 445
## 2 Site_02 Zilver... 170617 -34.4 19.4 Temp v... temp G40L 6450 324
## 3 Site_03 Zilver... 170612 -34.4 19.4 Large ... perm G40L 2785 668
## 4 Site_04 Anton ... 170801 -34.4 19.4 Small ... temp G40L 296 65
## 5 Site_05 Steenk... 170801 -34.4 19.4 Temp v... temp G40L 684 151
## 6 Site_06 Arum L... 170617 -34.4 19.5 Small ... perm G40L 60 30
## 7 Site_07 Jan Ma... 170801 -34.4 19.5 Large ... perm G40L 2296 204
## 8 Site_08 Jan Ma... 170801 -34.4 19.5 Small ... perm G40L 1305 133
## 9 Site_09 Chris ... 170905 -34.4 19.5 Large ... perm G40L 2767 197
## 10 Site_10 Chris ... 170905 -34.4 19.5 Small ... perm G40L 1027 123
## # ... with 40 more rows, 19 more variables: pH <dbl>, Conductivity <dbl>,
## # `Lm Bass` <dbl>, `Tilapia M` <dbl>, Fish <fct>, Terapin <chr>, Crab <chr>,
## # Frog <dbl>, Xl <dbl>, Af <dbl>, Sg <dbl>, Td <dbl>, Ca <dbl>, Sw <dbl>,
## # Av <dbl>, Sp <dbl>, Sc <dbl>, Sb <dbl>, Hh <dbl>, and abbreviated variable
## # names 1`Site name`, 2`Date visited`, 3Latitude, 4Longitude,
## # 5`Wetland type`, 6temporary, 7catchment, 8perimeter
```

```
coords <- data[,4:5]

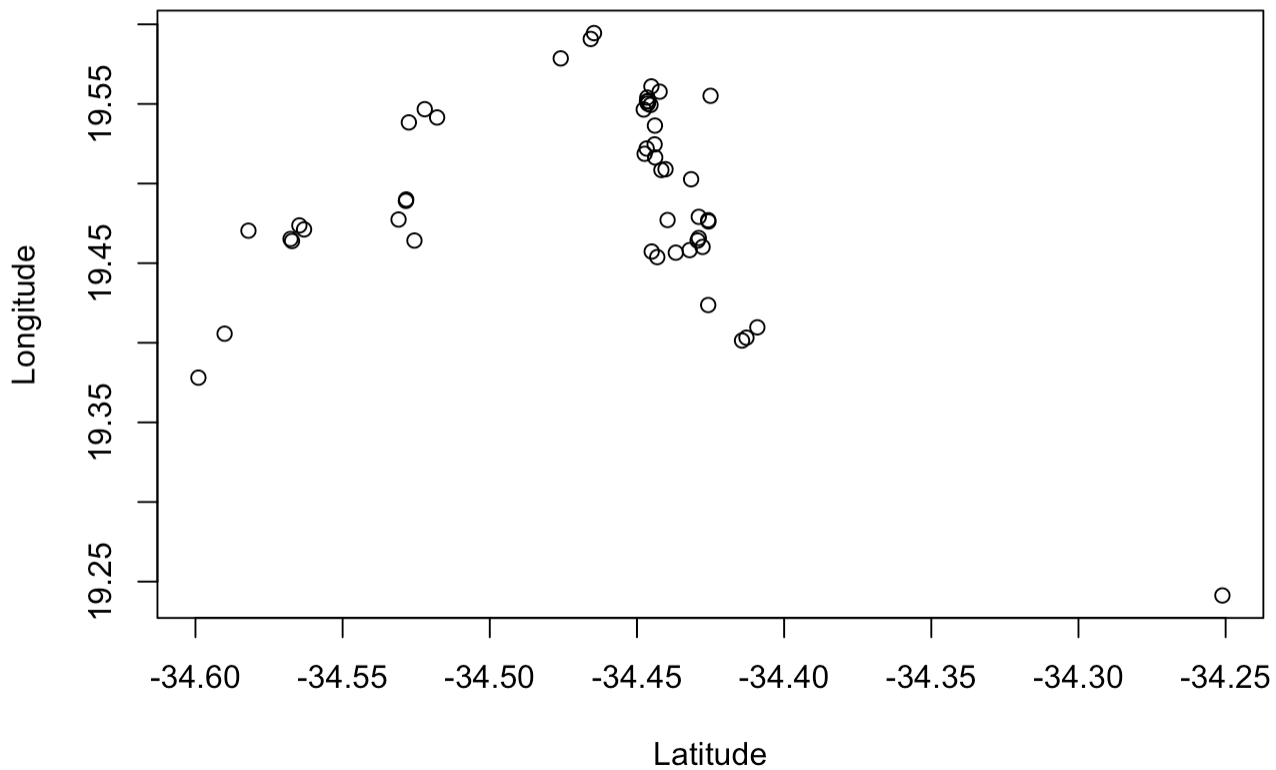
sp_compos <- data %>%
  dplyr::select(Xl:Hh)

env <- data %>%
  dplyr::select(c(`Wetland type`, temporary, area, perimeter, pH, Conductivity, Fish))
env
```

```
## # A tibble: 50 × 7
##   `Wetland type` temporary area perimeter pH Conductivity Fish
##   <fct> <fct> <dbl> <dbl> <dbl> <dbl> <fct>
## 1 Temp vlei temp 6169 445 7.5 95 no-fish
## 2 Temp vlei temp 6450 324 7.5 300 no-fish
## 3 Large dam perm 2785 668 7.3 210 no-fish
## 4 Small dam temp 296 65 8.4 1450 no-fish
## 5 Temp vlei temp 684 151 7.5 400 no-fish
## 6 Small dam perm 60 30 7.4 94 no-fish
## 7 Large dam perm 2296 204 7.3 1200 fish
## 8 Small dam perm 1305 133 7.4 700 no-fish
## 9 Large dam perm 2767 197 7.3 700 no-fish
## 10 Small dam perm 1027 123 7.7 1000 no-fish
## # ... with 40 more rows
```

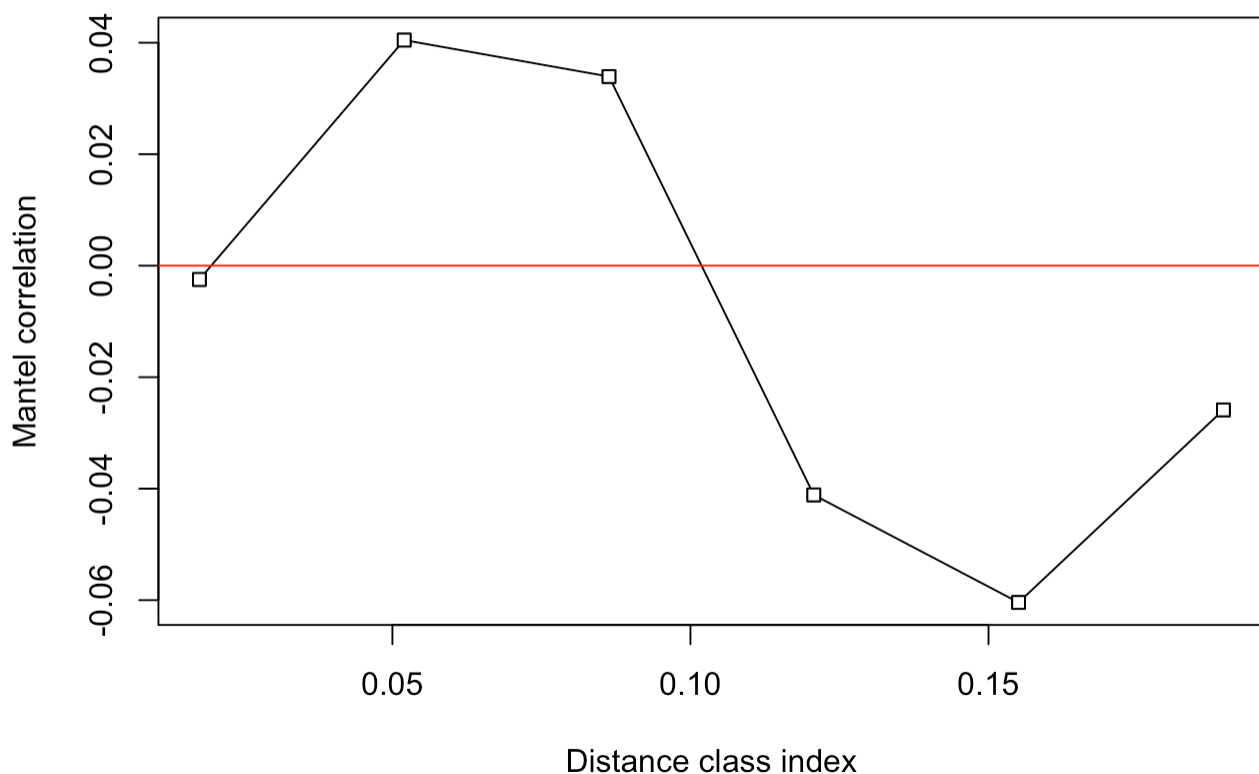
Building MEMs and testing for spatial autocorrelation in species composition

```
dec <- decostand(sp_compos, "hell")#transforming species composition  
plot(data[,4:5])
```



```
##  
## Mantel Correlogram Analysis  
##  
## Call:  
##  
## mantel.correlog(D.eco = vegdist(sp_compos), XY = coords)  
##  
##      class.index      n.dist  Mantel.cor Pr(Mantel) Pr(corrected)  
## D.cl.1      0.0176250 378.0000000  -0.0024746      0.518      0.518  
## D.cl.2      0.0519805 422.0000000   0.0404736      0.119      0.238  
## D.cl.3      0.0863360 608.0000000   0.0339321      0.192      0.384  
## D.cl.4      0.1206916 428.0000000  -0.0411226      0.133      0.476  
## D.cl.5      0.1550471 344.0000000  -0.0604176      0.109      0.545  
## D.cl.6      0.1894026 124.0000000  -0.0258708      0.340      0.680  
## D.cl.7      0.2237582  48.0000000          NA          NA          NA  
## D.cl.8      0.2581137   8.0000000          NA          NA          NA  
## D.cl.9      0.2924692 22.0000000          NA          NA          NA  
## D.cl.10     0.3268247 14.0000000          NA          NA          NA  
## D.cl.11     0.3611803 32.0000000          NA          NA          NA  
## D.cl.12     0.3955358 22.0000000          NA          NA          NA
```

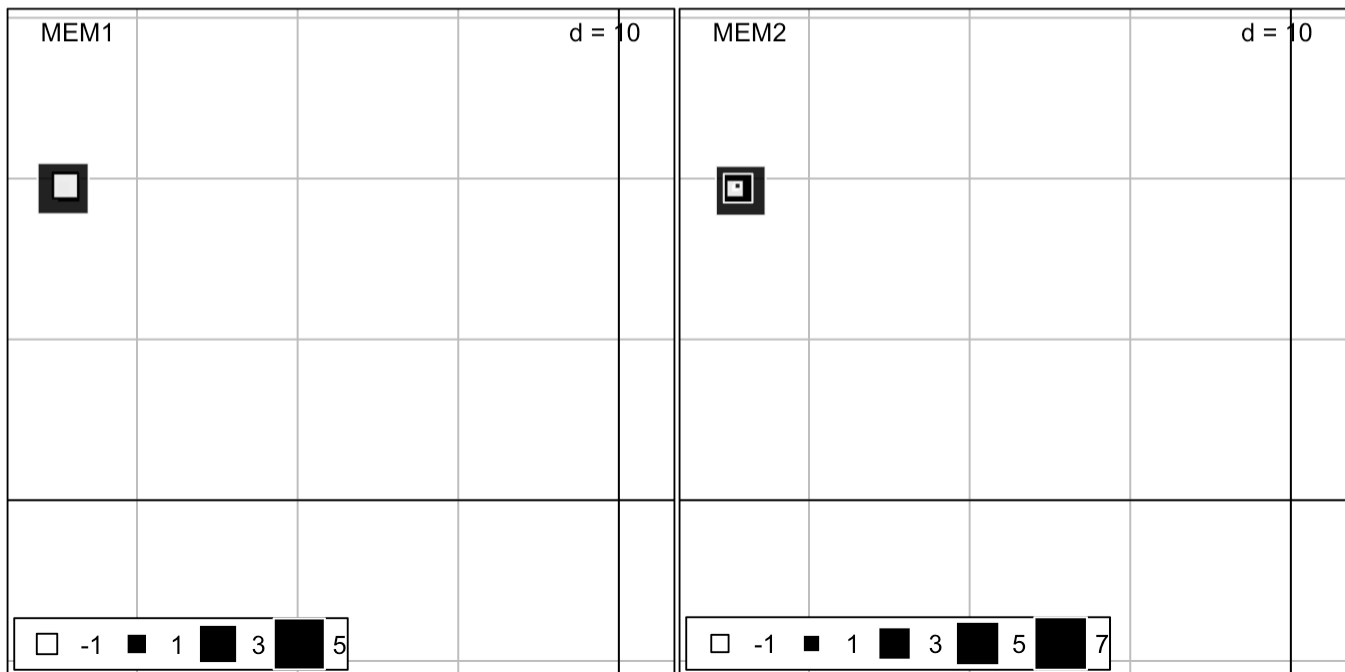
```
plot(mantel.correlog(vegdist(sp_compos), XY = coords))
```



```
mems <- dbmem(data[,4:5], MEM.autocor = "positive")#building MEMs and only taking eig
envectors with positive Moran's I
s.value(coords, mems[,1:2], include.origin = FALSE)
```

```
## Warning: Unused parameters: include
```

```
## Warning: Unused parameters: include
```



```
MEMs <- cbind(MEM1=mems$MEM1, MEM2=mems$MEM2)
```

making pRDA model

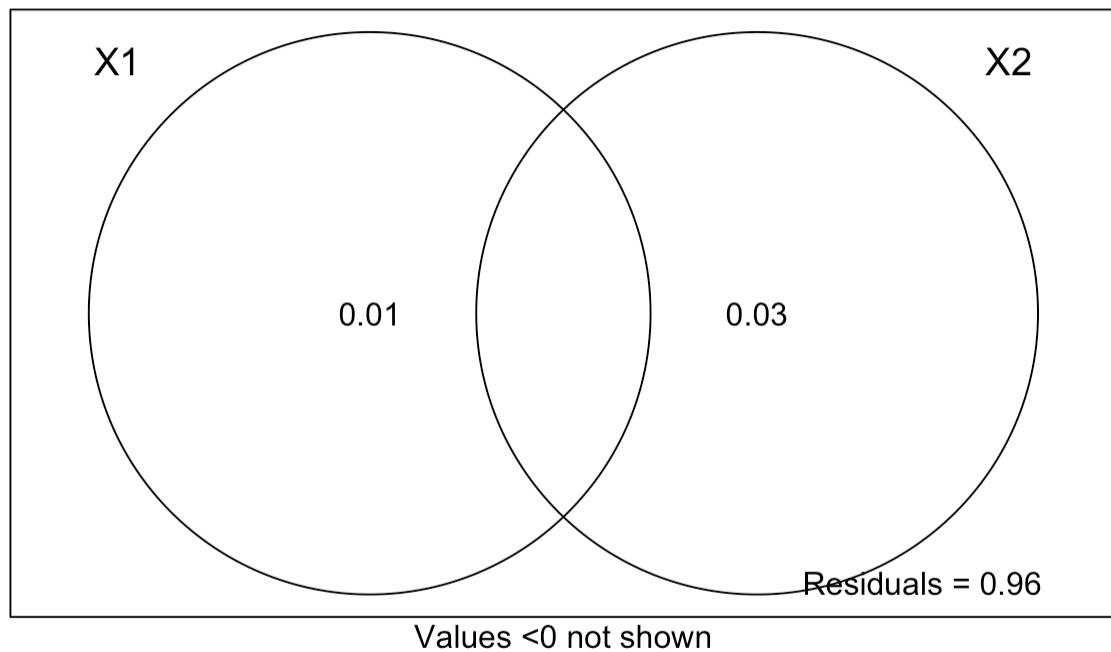
```
gm <- rda(dec~`Wetland type`+ temporary+ area+ perimeter+ pH+ Conductivity+Fish + Co
ndition(MEMs), data=env)
gm
```

```
## Call: rda(formula = dec ~ `Wetland type` + temporary + area + perimeter
## + pH + Conductivity + Fish + Condition(MEMs), data = env)
##
##              Inertia Proportion Rank
## Total          0.63751      1.00000
## Conditional    0.01718      0.02695    2
## Constrained    0.18152      0.28474   10
## Unconstrained  0.43880      0.68830   11
## Inertia is variance
##
## Eigenvalues for constrained axes:
##   RDA1   RDA2   RDA3   RDA4   RDA5   RDA6   RDA7   RDA8   RDA9   RDA10
## 0.09671 0.03645 0.02549 0.01227 0.00469 0.00315 0.00166 0.00084 0.00020 0.00005
##
## Eigenvalues for unconstrained axes:
##   PC1   PC2   PC3   PC4   PC5   PC6   PC7   PC8   PC9   PC10
## 0.16202 0.09384 0.07409 0.04716 0.02274 0.01410 0.01213 0.00533 0.00341 0.00269
##   PC11
## 0.00129
```

```
varpart(sp_compos, env, MEMs)
```

```
##
## Partition of variance in RDA
##
## Call: varpart(Y = sp_compos, X = env, MEMs)
##
## Explanatory tables:
## X1:  env
## X2:  MEMs
##
## No. of explanatory tables: 2
## Total variation (SS): 86178202
##           Variance: 1758739
## No. of observations: 50
##
## Partition table:
##           Df R.squared Adj.R.squared Testable
## [a+c] = X1    10   0.21141      0.00921    TRUE
## [b+c] = X2     2   0.07047      0.03092    TRUE
## [a+b+c] = X1+X2 12   0.27619      0.04144    TRUE
## Individual fractions
## [a] = X1|X2    10           0.01052    TRUE
## [b] = X2|X1     2           0.03224    TRUE
## [c]             0          -0.00132   FALSE
## [d] = Residuals           0.95856   FALSE
## ---
## Use function 'rda' to test significance of fractions of interest
```

```
plot(varpart(sp_compos, env, MEMs))
```



```
anova.cca(gm)
```

```
## Permutation test for rda under reduced model
## Permutation: free
## Number of permutations: 999
##
## Model: rda(formula = dec ~ `Wetland type` + temporary + area + perimeter + pH + Co
nductivity + Fish + Condition(MEMs), data = env)
##      Df Variance      F Pr(>F)
## Model   10  0.18152 1.5306  0.022 *
## Residual 37  0.43880
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova.cca(gm, by = "term")#only variable that is significant is wetland type
```

```
## Permutation test for rda under reduced model
## Terms added sequentially (first to last)
## Permutation: free
## Number of permutations: 999
##
## Model: rda(formula = dec ~ `Wetland type` + temporary + area + perimeter + pH + Co
nductivity + Fish + Condition(MEMs), data = env)
##           Df Variance      F Pr(>F)
## `Wetland type`  4  0.10443 2.2014  0.008 **
## temporary      1  0.00774 0.6525  0.697
## area           1  0.01668 1.4067  0.236
## perimeter      1  0.01118 0.9429  0.432
## pH             1  0.00693 0.5845  0.727
## Conductivity   1  0.01441 1.2155  0.324
## Fish           1  0.02014 1.6985  0.130
## Residual      37  0.43880
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
anova.cca(gm, by = "axis")
```

```
## Permutation test for rda under reduced model
## Forward tests for axes
## Permutation: free
## Number of permutations: 999
##
## Model: rda(formula = dec ~ `Wetland type` + temporary + area + perimeter + pH + Co
nductivity + Fish + Condition(MEMs), data = env)
##           Df Variance      F Pr(>F)
## RDA1        1  0.09671 8.1544  0.011 *
## RDA2        1  0.03645 3.0739  0.785
## RDA3        1  0.02549 2.1493  0.955
## RDA4        1  0.01227 1.0345  1.000
## RDA5        1  0.00469 0.3959  1.000
## RDA6        1  0.00315 0.2659  1.000
## RDA7        1  0.00166 0.1396  1.000
## RDA8        1  0.00084 0.0711  1.000
## RDA9        1  0.00020 0.0173  1.000
## RDA10       1  0.00005 0.0044  1.000
## Residual    37  0.43880
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
RsquareAdj(gm)#pretty small adjR2
```

```
plot(gm)
```

