

Inflammatory auxo-action in the stem cell division theory of cancer

Yi Luo^{1,2}, jian-hui Xiao^{Corresp. 1,2,3}

¹ Institute of Medicinal Biotechnology, Affiliated Hospital of Zunyi Medical University, Zunyi, China

² Zunyi Municipal Key Laboratory of Medicinal Biotechnology & Guizhou Provincial Research Center for Translational Medicine, Affiliated Hospital of Zunyi Medical University, Zunyi, China

³ Department of Gynaecology and Obstetrics, Affiliated Hospital of Zunyi Medical University, Zunyi, China

Corresponding Author: jian-hui Xiao
Email address: jianhuixiao@126.com

Acute inflammation is a beneficial response to the changes caused by pathogens or injuries that can eliminate the source of damage and restore homeostasis in damaged tissues. However, chronic inflammation causes malignant transformation and carcinogenic effects of cells through continuous exposure to pro-inflammatory cytokines and activation of inflammatory signaling pathways. According to the theory of stem cell division, the essential properties of stem cells, including long life span and self-renewal, make them vulnerable to accumulating genetic changes that can lead to cancer. Inflammation drives quiescent stem cells to enter the cell cycle and perform tissue repair functions. However, as cancer likely originates from DNA mutations that accumulate over time via normal stem cell division, inflammation may promote cancer development, even before the stem cells become cancerous. Numerous studies have reported that the mechanisms of inflammation in cancer formation and metastasis are diverse and complex; however, few studies have reviewed how inflammation affects cancer formation from the stem cell source. Based on the stem cell division theory of cancer, this review summarizes how inflammation affects normal stem cells, cancer stem cells, and cancer cells. We conclude that chronic inflammation leads to persistent stem cells activation, which can accumulate DNA damage and ultimately promote cancer. Additionally, inflammation not only facilitates the progression of stem cells into cancer cells, but also plays a positive role in cancer metastasis.

1 **Inflammatory auxo-action in the stem cell division theory of cancer**

2 Yi Luo^{1,2}, Jian-Hui Xiao^{1,2,3*}

3

4

5

6 ¹ Institute of Medicinal Biotechnology, Affiliated Hospital of Zunyi Medical University 149
7 Dalian Road, HuiChuan District, Zunyi 563003, China

8 ² Zunyi Municipal Key Laboratory of Medicinal Biotechnology & Guizhou Provincial Research
9 Center for Translational Medicine, Affiliated Hospital of Zunyi Medical University, 149 Dalian
10 Road, HuiChuan District, Zunyi 563003, China

11 ³ Department of Gynaecology and Obstetrics, Affiliated Hospital of Zunyi Medical University,
12 149 Dalian Road, Huichuan District, Zunyi 563003, China

13

14

15

16

17

18 *Correspondence to: Jian-Hui Xiao, Guizhou Provincial Research Center for Translational
19 Medicine, Affiliated Hospital of Zunyi Medical University, 149 Dalian Road, HuiChuan District,
20 Zunyi, 563003, People's Republic of China, Email jhxiao@zmu.edu.cn

21

22

23

24

25

26

27

28 **Abstract**

29 Acute inflammation is a beneficial response to the changes caused by pathogens or injuries
30 that can eliminate the source of damage and restore homeostasis in damaged tissues. However,
31 chronic inflammation causes malignant transformation and carcinogenic effects of cells through
32 continuous exposure to pro-inflammatory cytokines and activation of inflammatory signaling
33 pathways. According to the theory of stem cell division, the essential properties of stem cells,
34 including long life span and self-renewal, make them vulnerable to accumulating genetic
35 changes that can lead to cancer. Inflammation drives quiescent stem cells to enter the cell cycle
36 and perform tissue repair functions. However, as cancer likely originates from DNA mutations
37 that accumulate over time via normal stem cell division, inflammation may promote cancer
38 development, even before the stem cells become cancerous. Numerous studies have reported that
39 the mechanisms of inflammation in cancer formation and metastasis are diverse and complex;
40 however, few studies have reviewed how inflammation affects cancer formation from the stem
41 cell source. Based on the stem cell division theory of cancer, this review summarizes how
42 inflammation affects normal stem cells, cancer stem cells, and cancer cells. We conclude that
43 chronic inflammation leads to persistent stem cells activation, which can accumulate DNA
44 damage and ultimately promote cancer. Additionally, inflammation not only facilitates the
45 progression of stem cells into cancer cells, but also plays a positive role in cancer metastasis.

46 **Key words:** stem cell division theory; quiescent stem cells; cancer stem cells; carcinogenesis;
47 cancer metastasis; inflammation;

48

49

50

51

52 **Introduction**

53 Cancer is a major reason for decreasing life expectancy. (1) Cancer-related mortality
54 remains one of the leading causes of death globally, accounting for 13% of all human deaths
55 despite the availability of a range of clinical treatment strategies. (2) The term “cancer” is
56 fearsome to ordinary people because it is rooted in misunderstandings or misconceptions. For a
57 more accurate understanding of cancer, our forefathers (as early as ancient Greece) actively
58 explored its origin and proposed humoral theories, which stated that the imbalance of humors,
59 such as black bile, was responsible for various diseases. (3) With the development of modern
60 science and technology, scientists have presented many different theories for the origin of cancer,
61 including field theory, chemical carcinogenesis, infection, chromosomal abnormalities,
62 mutations, epigenetic changes, non-healing wounds, and immune surveillance theories, over the
63 past century. (4-6) The “monoclonal theory,” proposed by Hanahan and Weinberg in 2000
64 remains the mainstream and widely accepted theory of tumor development mechanisms. (7) It
65 states that tumors originate from a single “rebellious” cell, which eventually grows into the
66 whole tumor. Findings based on this theory have exerted positive effects on cancer prevention
67 and therapy by reducing incidence and mortality. However, it does not provide a complete
68 framework for understanding the cellular origin and pathogenesis of many cancers, which is
69 necessary for providing reliable roadmaps for cancer prevention, diagnosis, prognosis, and
70 treatment. (8) Cancer is known to be ultimately caused by uncontrolled cell growth and
71 proliferation. This feature is similar to the self-renewal ability of stem cells, which is their
72 exclusive potential to generate an unlimited number of cells. Although the current view is that
73 chemotherapy kills most cells in cancer tissues, cancer stem cells (CSCs) are believed to be left
74 behind, which may cause recurrence. (9) Furthermore, cancer cells possessing unique stem cell-
75 like properties have been widely identified in different human cancers over the last two decades.
76 (10, 11) Since the beginning of the new century, the CSC theory has been the focus of an
77 upsurge in research. (12) Nevertheless, the origin of these cancer cells remains controversial.
78 Currently, numerous data point to resident adult stem cells (ASCs) or primitive progenitor cells
79 as the origin of cancer cells, and studies have emphasized their role not only as propagators of

80 repair after tissue damage but also as cancer initiators. (13) Other studies have suggested that the
81 accumulation of mutations from stem cell division is the main trigger of cancer, and there is a
82 strong correlation between cancer risk and stem cell division. (14, 15) In 2015, based on
83 fundamental concepts in cell biology and related evidence, Spanish scientist Lopez-Lazaro
84 proposed the stem cell division theory of cancer. (16, 17)

85 Stem cells in developing embryos can proliferate indefinitely or remain undifferentiated.
86 When an organism needs to repair or remodel a certain tissue, the stem cell genome is activated
87 in a designated manner to differentiate into somatic cells with specialized functions and identities.
88 (18) ASCs with similar functions remain in individuals after embryonic development (Figure 1).
89 It is generally accepted that ASCs naturally exist in a quiescent state, with a small cell size and
90 no cell division. (19) Quiescent stem cells can be “awakened” into the cell cycle in response to
91 local damage signals or other regeneration needs, (20) and mutations leading to cancer tend to
92 occur only in actively dividing cells. (21) Nevertheless, cell division is a major source of DNA
93 alterations in normal stem cells and can lead to the accumulation of various cancer-promoting
94 errors. Recent cancer statistics show a dramatic increase in cancer incidence with age, suggesting
95 that the formation of most cancers requires multistep accumulation of deoxyribonucleic acid
96 (DNA) changes over years or decades (https://seer.cancer.gov/csr/1975_2018/). Accordingly, the
97 researchers also found that stem cells from newborn mice were less likely to become cancerous
98 than those from adult mice. (21) In the past, DNA damage was considered to result only in
99 genomic instability. However, recent evidence has shown that DNA damage can trigger
100 inflammation by activating the cGAS-STING axis and activating NF- κ B through ATM or ATR
101 proteins. (22) Furthermore, DNA damage induces the expression of type I interferon (IFN) and
102 other inflammatory cytokines. (23-25) Thus, the DNA damage generated during stem cell
103 division acts on itself to stimulate abnormal division by activating inflammatory pathways or
104 inducing the production of inflammatory cytokines. Alternatively, inflammation induced by
105 tissue damage can “wake up” quiescent stem cells and cause them to divide, greatly increasing
106 the risk of cancer. (26, 27) Overall, a circulating network of inflammatory signals produced by

107 multiple pathways regulates the process of stem cell division after tissue damage (Figure 2).
108 Combined with the stem cell division theory of cancer, (16, 17) we can conclude that tissue
109 damage creates favorable conditions for the formation of cancer cells, and inflammation
110 inadvertently promotes cancer. (28) In addition, while many factors can increase cancer risk,
111 including smoking, alcohol, high body mass index, pathogens, and radiation, the abnormal
112 activation of inflammatory signals is a common underlying factor associated with all these risk
113 factors. (29) Therefore, investigating the effects of inflammation on the behavior of normal stem
114 cells, CSCs, and cancer cells based on the stem cell division theory of cancer is critical to
115 understanding cancer generation from multiple perspectives. The information we have gathered
116 and reviewed here will be of interest to researchers working in developmental biology and cancer
117 development, as well as those working in the fields of molecular biology, cell biology, and stem
118 cells.

119 **Survey methodology**

120 The PubMed (<https://pubmed.ncbi.nlm.nih.gov/>) and Google Scholar
121 (<https://scholar.google.com/>) repositories were used to search the following terms: stem cells,
122 cancer stem cells, progenitor cells, mesenchymal stem cells, cancer cells, cancer, inflammation,
123 cytokines, cell division, malignant phenotype, tumorigenicity, cancer metastasis, stemness,
124 pluripotency, self-renewal, and differentiation potential.

125

126 **1 Stem cell division theory of cancer**

127 Cancer remains one of the major diseases that threaten human survival and is difficult to
128 cure clinically. (2) Despite significant progress in understanding the signals that drive cancer
129 growth and how to target these signals, effective disease control remains a key scientific and
130 medical challenge. Three well-known reasons behind the difficulty in conquering cancer include
131 tumor immune escape (difficulty of the host immune system in recognizing cancer cells), cancer
132 metastasis, and tumor resistance. Although cancer cells have different biochemical compositions,
133 antigenic structures, and biological behaviors than normal cells, (30, 31) the immune system

134 struggles to identify cancer cells accurately. Tracing the origin of cancer cells is essential for
135 finding strategies to overcome this phenomenon. Cancer cells are mutated from "self" cells,
136 which endows them with good camouflage properties. Cancer metastasis is the other main reason
137 for the failure of clinical treatment of tumors and death of most patients. Although clinical
138 manifestations of cancer suggest that metastasis is a late event, many recent studies have
139 demonstrated that cancer cells begin to metastasize at an early stage, even before diagnosis. (32-
140 34) Researchers analyzed samples from more than 100 patients with breast, colorectal, and lung
141 cancer and found that the genomic drivers required for invasion and metastasis were present in
142 primary tumors. (33) Breast cancer cells with certain molecular changes can spread to other
143 organs even before the primary tumor forms. These cells remain quiescent for long periods
144 before being awakened to form aggressive and lethal metastatic breast cancer cells. (34) This
145 new model of early metastasis challenges our understanding of how cancer spreads and forms
146 metastases. Therefore, the novel biological mechanisms of early cancer spread must be explored
147 to target metastatic cancer cells. A key and well-recognized factor that promotes cancer
148 metastasis is the formation of a microenvironment at a specific site (35) that is favorable for
149 cancer metastasis, including tumor-secreted factors, mobilization of suppressive immune cells,
150 and inflammatory polarization of stromal components at this site. (36) During the multistep
151 process of cancer metastasis, primary cancer cells acquire cellular and phenotypic plasticity to
152 survive and grow in diverse microenvironments. (37) Therefore, understanding the novel
153 biological mechanisms of cancer metastasis, cancer cell origin, and tumor microenvironment is
154 essential.

155 Although the target cells of the transforming mutation are unknown for most cancers, recent
156 evidence suggests that cancer is a stem cell-based disease, and embryonic, adult, or pluripotent
157 stem cells are the origin cells. (38) Research shows that the number of stem cell divisions in
158 tissues is positively correlated with cancer incidence, and 65% of cancers can be explained by
159 this number. (39) Therefore, Miguel Lopez-Lazaro proposed a theory of stem cell division in
160 cancer, which states that 1) tumors originate from normal stem cells; 2) the main determinant of

161 cancer development is the damage undergone by stem cells during division; 3) the accumulation
162 of sufficient damage in stem cells leads to the production of tumor stem cells, which are
163 responsible for tumor formation; and 4) metastasis occurs when stem cells (uncontrolled,
164 precancerous, or cancerous) or their malignant progeny leave their natural tissue (not necessarily
165 the primary tumor) and form tumors elsewhere. (9, 10)

166 The stem cell division theory of cancer is controversial; however, the idea that cancer
167 originates from normal stem cells has a solid biological basis. Several observational studies have
168 shown that age is the most important risk factor for cancer. (40, 41) Stem cells possess genome
169 passed on from fertilized eggs, and remain in our body until death; therefore, they are the only
170 cells that acquire and accumulate DNA changes throughout our lives. (42) Self-renewal, which is
171 the most important property of stem cells, is strikingly similar to that of cancer cells. Many
172 canonical signaling pathways associated with cancer, such as Notch, sonic hedgehog, and Wnt
173 signaling, also regulate normal stem cell development. Emerging cancer cells likely take
174 advantage of the self-renewal cell division machinery normally expressed in stem cells.
175 Understanding the signaling network of normal stem cells and cancer cell development will help
176 to trace the initial cancer cells and identify targets for anticancer therapy. Stem cell functions,
177 including splitting rate and migration ability, are established within the niche. (43-45) The stem
178 cell niche is a dynamic and special microenvironment that activates certain intracellular signaling
179 pathways by producing factors that directly act on stem cells, ultimately determining the fate of
180 cells, such as division, differentiation, or apoptosis. Additionally, tissue stem cells, which are
181 usually in a quiescent state, activate self-renewal and differentiation programs to maintain tissue
182 homeostasis and repair wounds. (46) Tissue damage accompanies the generation of sterile and
183 pathogenic inflammation. (47) Thus, inflammatory signals regulate stem cell function in many
184 ways and undoubtedly affect the accumulation of stem cell mutations.

185

186 **2 Is the inflammatory response a friend or foe to cancer?**

187 The typical triggers of inflammation (infection and tissue damage) are marked by leukocyte

188 infiltration, a process by which the innate immune system helps repair damaged tissue by
189 activating immune and non-immune cells against pathogens. (48) Currently, the medical
190 community has made it clear that inflammation is closely related to cancer occurrence,
191 development, and efficacy of anticancer treatment and acts as a “double-edged sword”.
192 Inflammation can be classified into acute and chronic inflammation. Acute inflammation is the
193 natural defense of the body against damaged cells, viruses, and other harmful stimuli; it sets in
194 quickly and helps the body to heal itself. During this process, the immune cells and chemicals
195 involved can kill pathogens, promote tissue repair, prevent tumor growth, and activate the
196 immune system through stimuli and inflammatory factors, thereby acting as tumor suppressors.
197 (49-51) For example, induced acute inflammation is used to treat bladder cancer, (52) and high
198 concentrations of tumor necrosis factor (TNF) can induce an antitumor response in a mouse
199 model of sarcoma. (53) Acute inflammation is short-lived and quickly resolved. If inflammation
200 cannot be resolved in time, it results in chronic inflammation, which promotes the development
201 of malignant tumors through continuous exposure to pro-inflammatory factors and activation of
202 signaling pathways, such as NF- κ B and signal transducer and activator of transcription (STAT) 3.
203 When cells become malignant, chronic inflammation signals play a critical role in promoting
204 cancer cell proliferation, evading immune surveillance, and promoting angiogenesis to support
205 the growth and spread of cancer. For example, low and sustained levels of TNF- α can induce
206 tumor phenotypes, and interleukin (IL) 1 β has been identified as a key molecule contributing to
207 colorectal cancer (CRC) development. (54, 55) IL-6, a pro-inflammatory cytokine with typical
208 pro-cancer effects, plays a key role in promoting proliferation and inhibiting apoptosis by
209 binding to its receptor IL-6Ra and co-receptor glycoprotein130 (gp130) to activate transcription
210 factors, including STAT1 and STAT3. (56)

211 Inflammation is an adaptive response triggered by infection and tissue damage and is
212 differentiated by pathogen-associated molecular patterns (PAMPs) and damage-associated
213 molecular patterns (DAMPs). PAMPs, such as membrane-associated lipids and
214 lipopolysaccharides, are exogenous components specific to invading microorganisms. In contrast,

215 inflammation without pathogens and their products is called sterile inflammation and is triggered
216 by endogenous danger signs. DAMPs are released upon tissue injury (57) and initial
217 inflammation and leukocyte recruitment are prerequisites for effective tissue repair. (58)
218 Researchers have now found that both PAMPs and DAMPs are recognized by pattern
219 recognition receptors (PRRs) to initiate immune responses. These receptors include Toll-like
220 receptors (TLRs), RIG-i-like receptors (RLRs), C-type lectin receptors (CLRs), and Nod-like
221 receptors (NLRs), which induce cytokine and IFN production by activating associated
222 inflammatory pathways. (59) After identifying PAMP and DAMP, TLRs bind to adaptor proteins
223 containing Toll-IL-1-resistance (TIR) domains, such as myeloid differentiation primary response
224 gene 88 (MYD88) or Toll-receptor-associated activator of interferon (TRIF). MyD88 and TRIF
225 recruit and activate mitogen-activated protein kinases (MAPKs) and I κ B kinases (IKK) and
226 ultimately induce the expression of inflammatory cytokines by activating transcription factor
227 activating protein 1 (AP-1) and NF- κ B, respectively. (60-61) Thus, the production of PAMPs
228 and DAMPs is necessary for tissue repair and regeneration; however, under persistent
229 inflammation, they can also lead to the development of cancer. (61) For example, pro-
230 inflammatory microenvironments mediated by the TLR2-MyD88-NF- κ B pathway support
231 epithelial ovarian CSCs-driven repair and self-renewal. (62)

232

233 **3 Influence of inflammation on non-malignant stem cell behavior**

234 ASCs can undergo extensive cell division while retaining the capacity to generate stem cells
235 and differentiate into specialized cells. This enables ASCs to replenish damaged tissues and help
236 maintain healthy tissue in the body throughout an organism's lifespan. (63) ASCs can remain
237 quiescent without dividing for extended periods until they are activated. (64) Accordingly, ASCs
238 are suspected to be the origin of cancer cells because of their long lifespan and propensity for
239 extensive cell division. (65)

240 ***3.1 Inflammatory response: the key to awakening dormant stem cells***

241 Historically, ASCs have been considered to exist in either a quiescent state, in which the

242 cell is not actively cycling, or an activated state, in which the cell has entered the cell cycle. (19,
243 66) In the quiescent state, stem cells are in the reversible G0 phase and can re-enter the cell cycle
244 in response to normal physiological stimuli. This is different from the G0 phase for terminally
245 differentiated cells or senescent cells, which have left cell cycle and cannot re-enter. (19) The
246 non-proliferative quiescent state of ASCs can protect the cell's DNA from mutations acquired
247 during continuous cell division, (67) making them less likely to acquire cancer.

248 Many factors can trigger ASCs to exit the quiescent state. ASCs activation can be induced
249 during tissue damage through mechanisms such as cell–cell contact and cell–extracellular matrix
250 interaction. (68) In addition, the mechanism by which inflammation awakens quiescent stem
251 cells to promote the regeneration of damaged tissues has been widely reported. (69) Reactive
252 oxygen species (ROS) generated during tissue damage and infection are important cues linking
253 inflammation to intestinal stem cells (ISCs) proliferation through activating Jun N-terminal
254 kinases (JNKs) and inhibiting nuclear factor erythroid2-related factor 2 (Nrf2) signal. (70, 71)
255 Lipopolysaccharide (LPS) -induced transient mild inflammation stimulates the selective
256 activation and proliferation of stationary stem cell populations, such as Clara-like cells (CLCs).
257 (72) Moreover, IFN- γ treatment increased the expression of bone marrow stromal cell antigen 2
258 (BST2), a cell surface protein essential for INF γ -dependent hematopoietic stem cell (HSC)
259 activation, thereby disrupting HSC quiescence and promoting excessive terminal differentiation.
260 (73) Additionally, IL-6 and TNF directly act on epithelial cells by binding to the IL-6 receptor,
261 gp130 heterotetramer, and TNF receptor 1 (TNFR1), thereby promoting the regeneration of
262 damaged intestinal mucosa. (74, 75) IL-22 acts on epithelial cells and fibroblasts to stimulate
263 proliferation, inhibit cell death, and delay terminal differentiation. The binding of IL-22 to its
264 receptor leads to the activation of Janus kinases (JAK)-STAT3, MAPK, extracellular signal-
265 regulated kinase (ERK), and JNK. (76) Therefore, inflammation is an important mediator that
266 mobilizes quiescent stem cells to regenerate after tissue damage.

267 ***3.2 Inflammation mobilizes tissue stem cells: regeneration and tumorigenicity coexist***

268 After tissue damage, inflammatory cells and cytokines are the main components of the

269 tissue stem cell niche. Tissue stem cells, which are activated from a quiescent state owing to
270 niche changes, repair damaged tissues through proliferation and differentiation and exhibit strong
271 regenerative capabilities. (77) For example, multiple subsets of innate lymphoid cells can play
272 context-specific roles in stem cell regeneration and differentiation. (78, 79) However, mutations
273 can occur with cell division. Although stem cells can self-renew for extended periods, but this
274 also poses an inherent challenge. Being the longest-lived cells in an organism, they are at an
275 increased risk of acquiring genomic damage, which undoubtedly increases the likelihood of
276 accumulation of mutations in the stem cell genome. Gradual accumulation of genomic mutations
277 throughout life may lead to carcinogenesis. Therefore, under the influence of inflammation, the
278 regeneration of tissue stem cells coexists with tumorigenic ability. Although mutations and
279 cancer incidence in some tissues depend primarily on exposure to external mutagens, (80)
280 intrinsic factors, such as the number of cell divisions in the tissue, appear to dominate other
281 cancer types. (19) For example, a high-fat diet can drive a surge in intestinal stem cells and
282 produce a range of other cells that behave very similarly to stem cells by multiplying indefinitely
283 and differentiating into other types of cells. Stem and stem-like cells are more likely to cause
284 intestinal tumors. (81, 82) Wnt/ β -catenin signaling plays a key role in intestinal epithelial
285 homeostasis by promoting stem cell renewal. If Wnt signaling is overstimulated, stem cells
286 divide uncontrollably. Mice with mutations in the Wnt signaling pathway develop polyps that
287 eventually develop into colon cancer. (83)

288 ***3.3 Influence of inflammatory response on adult stem cell function***

289 The function of ASCs has been established in the stem cell niche, which regulates stem cell
290 survival and function by producing factors that directly act on stem cells. (43) Inflammatory
291 mediators, which are important niche components after tissue injury, influence the behavior of
292 stem cells in various ways. Numerous studies have reported that the inflammatory
293 microenvironment affects the biological characteristics and behavior of stem cells, both in vitro
294 and in vivo (Table 1). For example, IL-1-induced mesenchymal stem cells (MSCs) show
295 increased granulocyte colony-stimulating factor (G-CSF) expression through the type 1 IL-1

296 receptor, which results in decreased secretion of LPS-activated microglia inflammatory
297 mediators, priming MSCs for in vitro transformation to an anti-inflammatory and pro-nutritional
298 phenotype. (84) Moreover, a combination of four pro-inflammatory cytokines (IL-1 α , IL-13,
299 TNF- α , and IFN- γ) secreted by T cells can promote the continuous expansion of muscle stem
300 cells (MuSCs) in vitro for over 20 generations. (85) IL-22 promotes ISC-mediated epithelial
301 regeneration by inducing STAT3 phosphorylation. (78) The inflammatory milieu mediated by
302 IL-1 and TNF- α preconditioning initially stimulates the regeneration of gingival mesenchymal
303 stem/progenitor cells (G-MSCs); this positive effect disappears as inflammation persists,
304 followed by a short-term stimulatory effect on osteogenesis. (86) After exposing cultured MuSCs
305 to the inflammatory mediator prostaglandin E2 (PGE2) for one day, the number of cells
306 increased six-fold compared to the control group. (87) Thus, these studies confirmed that
307 inflammatory factors activate downstream stem cell pathways, enhancing their self-renewal and
308 in vivo efficacy. In addition, when MSCs are exposed to an inflammatory environment, they
309 release more functional growth factors, exosomes, and chemokines, thereby regulating the
310 inflammatory microenvironment and promoting tissue regeneration. (88) In vitro studies have
311 confirmed that some inflammatory factors have profound effects on stem cell behavior,
312 especially proliferation and immune phenotypes. Considering the stem cell division theory of
313 cancer and the coexistence of stem cell proliferation and tumorigenicity, it is uncertain whether
314 the inflammatory microenvironment, especially the influence of inflammatory factors on stem
315 cell behavior, is beneficial or harmful to cancer formation. However, it is certain that stem cells
316 retain their biological properties, such as self-renewal and tissue repair, after coming into contact
317 with inflammatory factors. Partial inflammatory factors can maintain or even improve stem cell
318 function; that is, inflammatory factor treatment indirectly maintains the integrity of the stem cell
319 genome.

320 In vivo studies have confirmed that the tissue repair function of bone marrow MSCs in
321 various inflammatory diseases depends on inflammation regulation and production of various
322 growth factors. (89, 90) Thus, MSCs localized in damaged tissues can alleviate inflammation and

323 promote tissue stem/progenitor and other resident cells to regenerate normal functioning cells
324 and improve the tissue microenvironment by acting synergistically. For example, in vivo ISC
325 turnover is heavily regulated by the crypt niche. Macrophages have been identified as a key
326 component of intestinal crypts, and their production of PGE2 promotes Wnt/ β -catenin signaling
327 by binding to the ISC prostaglandin E receptors (EP, also know as Ptger) 1 and EP4, which in
328 turn promotes their self-renewal. (91) Furthermore, when mice were injected with PGE2
329 intramuscularly, muscle regeneration was significantly promoted and muscle strength increased.
330 Conversely, when MuSC responses to PGE2 were inhibited, EP4 expression was reduced, or
331 nonsteroidal anti-inflammatory drugs were administered to suppress PGE2 production, strength
332 recovery was hampered. (87) After intestinal injury, innate lymphocytes produce a large amount
333 of IL-22, which directly targets ISCs, enhances the growth of small intestinal organoids, and
334 promotes ISC proliferation and expansion. (78) The interaction between ISCs and T cells in the
335 local microenvironment has been elucidated, suggesting that T helper (Th) 1, Th2, and Th17
336 cells and their cytokines IFN- γ , IL-13, IL-17, and other pro-inflammatory signals can promote
337 the differentiation of Lgr5+ISCs, while regulatory T cells (Treg) cells and their cytokine IL-10
338 promote ISC self-renewal. (92) Skin-resident Tregs express high levels of the Notch ligand
339 family member Jagged 1 (Jag1). Jag1 expression on Treg promotes hair follicle regeneration by
340 enhancing hair follicle stem cell proliferation and differentiation. (93) Additionally, impaired
341 communication between aging keratinocytes and immune cells causes difficulty in wound
342 healing in aging skin. (94) In summary, in vivo studies have confirmed the importance of
343 inflammatory signaling in regulating tissue regeneration by promoting the proliferation and
344 differentiation of tissue-resident stem cells. Both in vitro and in vivo studies have confirmed that
345 the inflammatory environment profoundly affects the biological functions and behavior of stem
346 cells, including enhanced self-renewal and tissue repair. Based on the stem cell division theory of
347 cancer, cancer cells are derived from normal stem cells; therefore, the inflammatory
348 microenvironment is a pivotal factor affecting cancer cell formation from the source. However,
349 under different conditions, the microenvironments of stem cells are complex, and the influence

350 of the inflammatory microenvironment on the function of stem cells and their development into
351 cancer requires further exploration.

352

353 **4 Influence of inflammation on cancer stem cell behavior**

354 CSCs, a small subgroup of malignant tumor cells, have enhanced self-renewal, metastasis,
355 and spread as well as treatment resistance, and play a key role in the occurrence, recurrence and
356 metastasis of tumors. (95) Currently, it is considered that there are many similarities between
357 CSCs and ASCs, especially in their self-renewal, signal pathways and some stemness
358 transcription factors. Many signaling pathways that contribute to the survival, proliferation, self-
359 renewal, and differentiation characteristics of ASCs are abnormally activated or inhibited in
360 CSCs. (96, 97) For example, Wnt signaling is one of the key cascades regulating stem cell
361 development and survival; however, its increased activity induces stem cell signatures in
362 colorectal cancer (CRC) cells. (98, 99) The PI3K/Akt/mTOR pathway is also important for stem
363 cell survival, proliferation, and migration, and its continued activation in CSCs induces
364 tumorigenesis, cancer metastasis, and drug resistance. (100) Recently, a mainstream study
365 showed that CSCs originate from ASCs or progenitor cells, (86) and that many oncogenic factors,
366 including inflammation, induce the formation of CSCs by activating the pathways necessary for
367 them. (101)

368 ***4.1 Inflammation drives the generation of cancer stem cells***

369 In some tumors, CSCs are considered to be a mutated version of non-malignant
370 stem/progenitor cells and a precursor to differentiated cancer cells that can induce tumor
371 formation and differentiation into various cell types within the tumor. Chronic inflammation, that
372 is, secreted factors from stromal and immune cells, triggers signaling changes in stem/progenitor
373 cells that abnormally activate stem cell-related molecular pathways and promote the
374 transformation of non-malignant cells into CSCs, ultimately inducing tumor formation. The IL-
375 6/JAK/STAT3 signaling axis plays a central role in regulating the generation of CSC in human
376 breast cancer cell lines. Compared with other tumor cell types, IL-6/JAK2/STAT3 pathway is

377 preferentially active in CD44⁺CD24⁻ breast cancer cells, and JAK2 inhibition reduces their
378 number and blocks the growth of xenograft. (102) Furthermore, IL-6 regulates the expression of
379 CSC-related Oct-4 genes in non-CSC through the IL-6/JAK1/STAT3 signal transduction
380 pathway. (103) TNF- α induced inflammation promotes tumorigenesis and cancer progression
381 and is a key signal for the generation of CSCs. For example, TNF- α triggers chromosomal
382 instability in hepatocellular progenitors by modulating ubiquitin D and checkpoint kinase 2 and
383 enhances the self-renewal of hepatocellular progenitors through the TNFR1/Src/STAT3 pathway,
384 which collaboratively promotes the transformation of hepatocellular progenitors into
385 hepatocellular carcinoma stem cells. (104) Additionally, TNF- α induces malignant
386 transformation of ISCs by activating NF- κ B and Wnt/ β -catenin pathways. (105) Overall, the
387 cellular and molecular mechanisms of inflammatory induction of CSC generation have been
388 gradually elucidated, providing a new molecular classification for the individualized treatment of
389 cancer.

390 *4.2 Inflammation maintains the malignant phenotype of cancer stem cells*

391 The complex interaction between CSCs and their microenvironment can further regulate the
392 growth of CSCs. Inflammatory cytokines, such as ILs, TNF, and chemokines, maintain the
393 stemness state of CSCs in a variety of ways (Table 2). IL-6-mediated signal transduction has
394 been extensively studied. For example, the transcription factor CCAA T/enhancer binding
395 protein delta (C/EBP- δ) amplifies IL-6 and hypoxia-inducible factor 1 (HIF-1) signaling by
396 directly targeting the IL-6 receptor (IL6RA), whose deletion or depletion reduces the expression
397 of stem cell factors and stem cell markers, the formation of blobs, and self-renewal. (106) The
398 expression of circATP5B in glioma stem cells (GSC) was significantly up-regulated and
399 promoted the proliferation of GSC. Mechanically, circATP5B acts as a miR-185-5p sponge to
400 up-regulate the expression of homeobox protein Hox-B5 (HOXB5). HOXB5 is overexpressed in
401 gliomas and transcriptionally regulates IL6 expression. (107) IL-6 also promotes the cell
402 stemness and invasiveness of glioblastoma by inhibiting the expression of miR-142-3p. (108)
403 Alternatively, TNF- α -mediated signaling pathways have also been highlighted. Long-term

404 exposure to TNF- α can increase the proportion of CSCs in oral squamous cell carcinoma,
405 thereby enhancing its pellet-forming ability, stem cell transcription factor expression, and
406 tumorigenicity. (109) CircKPNB1 overexpression can promote the proliferation, invasion, and
407 stem cell viability of GSC. Mechanistically, circKPNB1 regulates protein stability and nuclear
408 translocation in SPI1. SPI1 promotes the malignant phenotype of GSC through TNF- α mediated
409 NF- κ B signaling. (110) Other inflammatory factors, such as PGE2, chemokine (C-C motif)
410 ligand (CCL) 16, and IL-17, (111-113) can promote the malignant phenotype of CSCs through
411 relevant signaling pathways. Overall, the evidence suggests that inflammatory cytokines are
412 involved in the mechanism of CSCs stemness, and targeting inflammatory cytokines may be a
413 useful adjunct to cancer therapy.

414

415 **5 Influence of inflammation on the spread of cancer**

416 The importance of intracellular communication between cancer and immune cells in the
417 tumor microenvironment has long been recognized. (114) When stimulated, host immune cells
418 secrete cytokines and other tiny inflammatory proteins to fight cancer; however, these cytokines
419 sometimes activate cancer cells and lead to specific mutations and epigenetic changes. (115) In
420 most cases, chronic inflammation is required for the development of cancer. (116, 117) Cases of
421 chronic inflammation resulting from tissue damage leading to cancer have lost their clinical
422 novelty. For example, the tobacco carcinogen nicotine-derived nitrosamine ketone significantly
423 promotes lung cancer by increasing the expression of CCL20. (118) Obesity also induces chronic
424 inflammation (activation of IL-6 and TNF- α) and promotes hepatocellular carcinoma. (119)
425 Helicobacter pylori infection is associated with gastric cancer and gastric mucosa-associated
426 lymphoid tissue lymphoma. (120) Human papillomavirus infection leads to lesions of the
427 cervical squamous epithelium and progression to cervical cancer. (121) Although an immune
428 response to pathogens is a natural defense, all these pathogens can cause long-term infections,
429 which can induce chronic inflammation and, ultimately, cancer cell formation. These examples
430 demonstrate that almost all forms of cancer, regardless of carcinogenic factors, are associated

431 with immune activation and chronic inflammation. Therefore, it is important to explore the
432 influence of the inflammatory microenvironment on the behavior of cancer cells.

433 ***5.1 Inflammatory microenvironment reprograms cancer cell fate***

434 Cytokines and growth factors produced during chronic inflammation may have
435 multifunctional effects on tumor formation and growth, both directly on tumor cells and
436 indirectly by promoting favorable conditions in the microenvironment. DNA damage is a bridge
437 between chronic inflammation and cancer. (122) During inflammation, reactive oxygen and
438 nitrogen species are created to fight pathogens and stimulate tissue repair and regeneration;
439 however, they also damage DNA, which in turn promotes mutations that lead to genomic
440 instability that can initiate and promote cancer development. (123-125) The ability of
441 inflammation to reprogram cancer cell fate is linked to cancer development. Persistent
442 inflammation promotes cancer development by activating the proliferation, survival, and
443 metastasis of cancer cells. (126, 127) For example, CCL2-CCR2 signaling promotes cancer
444 progression by supporting cancer cell proliferation and survival, inducing cancer cell migration
445 and invasion, and stimulating inflammation and angiogenesis. (128) IL-6 directly affects tumor
446 growth by enhancing the proliferation and survival of malignant cells in multiple myeloma, non-
447 Hodgkin's lymphoma, and hepatocellular carcinoma. (129, 130) It has been found that TNF- α
448 and IL-1 β can activate the hypoxia signaling pathway in human liver cancer cells, regulate tumor
449 cells, and directly affect tumor growth. (131) Chronic inflammation caused by smoking can
450 cause neutrophils to exhibit suicidal anti-infective behavior, which, in addition to awakening
451 cancer cells that are dormant for years or even decades in patients with cancer, causes tumor
452 recurrence. (132) Conversely, anticancer compounds inhibit cancer initiation and progression by
453 downregulating pro-inflammatory cytokine levels. (133-135)

454 ***5.2 Inflammatory microenvironment promotes cancer cell metastasis***

455 Epithelial-to-mesenchymal transition (EMT) is an embryonic process that loosens cell-cell
456 adhesion complexes and confers enhanced migratory and invasive properties to cells, and it is
457 exploited by cancer cells during metastasis. Cancer cells that undergo EMT are more aggressive

458 and exhibit enhanced invasiveness, stem cell-like characteristics, and anti-apoptotic capabilities.
459 Inflammation is a potent inducer of EMT in tumors; conversely, EMT can also stimulate cancer
460 cells to produce pro-inflammatory factors. (136) Researchers have comprehensively summarized
461 the progression of inflammation and EMT in cancer cells. (136) Briefly, the EMT/inflammatory
462 axis promotes the aggressiveness of primary tumors, and EMT and inflammatory markers are
463 associated with poor prognosis in multiple cancer patient cohorts. In addition to acting as a direct
464 inducer of EMT in cancer cells, inflammation can act as an intermediate mediator of cancer
465 development. For example, microenvironment dysregulation due to microbial interactions
466 promotes tumor development. Microbes do not directly influence tumor behavior and require
467 mediators to promote tumor development. The current view is that microorganisms regulate
468 tumor immunity through their derived metabolites, toxins, antigens, and other substances; they
469 regulate tumor cell metabolism and reshape the tumor microenvironment to promote tumor
470 occurrence and progression. Inflammation acts as an intermediary in this process. (137) In
471 addition, the regulatory balance between tumors and immunity is a cyclic process. For example,
472 one study found that tumor cells secreted granulocyte-macrophage colony-stimulating factor to
473 stimulate macrophages, which were activated and secreted CCL18, promoting tumor cell EMT
474 and eventually leading to lung metastasis of breast cancer cells. (138) Altogether, the main cause
475 of cancer death is not the primary tumor itself but the depletion of distant organs and tissue
476 metastases. Although the mechanism underlying this process is unclear, inflammation-mediated
477 EMT plays an important role.

478 In general, malignant tumors are difficult to treat because, in addition to the ability of
479 cancer cells to proliferate, invade, and metastasize indefinitely, they can evade immune
480 surveillance. A series of immunotherapeutic approaches based on immune evasion mechanisms
481 have been developed and clinically applied in the past few decades. Unlike traditional
482 chemoradiotherapy, immunotherapy mainly uses immune cells inside and outside the tumor
483 microenvironment to identify and attack cancer cells (139), which theoretically makes
484 immunotherapy more specific with few side effects. Nonetheless, downregulation of major

485 histocompatibility complex class I antigen presentation, which frequently occurs in solid cancers,
486 limits the effectiveness of these therapies. Research has confirmed that cells that appear to be in
487 the quiescent phase are resistant to immune system attacks. (140) The ability of long-lived stem
488 cells to evade immune surveillance may be due to their mostly non-proliferating quiescent state,
489 which may be an important feature of stem cells that develop into cancer.

490

491 **6 Conclusions**

492 The inactive division of CSCs is key to cancer metastasis, recurrence and drug resistance.
493 Although compounds and methods have been found to specifically inhibit or eliminate CSCs,
494 (141-143) no systematic studies have been conducted to achieve the eradication of CSCs in vivo.
495 While the origin of CSCs can vary by tissue evidence suggests that ASCs can transform into
496 CSCs under continuous abnormal activation of pathways related to cell proliferation and survival.
497 This abnormal activation is typically triggered by the changes in stem cell niche, and
498 inflammation is one of the key factors that can mediate such changes. ASCs are usually present
499 in a quiescent state and function to maintain tissue homeostasis. During tissue injury, ASCs exit
500 the quiescent state, driven by complex signals including inflammation, and enter the cell cycle to
501 repair the damaged tissue through division and differentiation. However, prolonged periods of
502 ASCs division increase the risk of cancer. (Figure 3) This partly explains why ISC, skin stem
503 cells, and stomach epithelial cells renew more quickly than other cells in the body, and they are
504 at a high risk of developing cancer. (38, 144, 145) Conversely, non-regenerating cells such as
505 nerve cells and cardiomyocytes have a significantly low risk of developing cancer at their tissue
506 sites.

507 Although the concept of inflammation-inducing cancers has gained acceptance over time,
508 its underlying mechanism has not been clearly explained. Based on the stem cell division theory
509 of cancer, this review highlights the impact of inflammation on normal stem cells, CSCs, and
510 cancer cells. It also speculates on the potential of inflammation to promote the transformation of
511 ASCs into CSCs and to facilitate cancer metastasis. Ultimately, a comprehensive understanding

512 of how inflammation influences the behavior of stem cells and cancer cells could revolutionize
513 our comprehension of numerous diseases, paving the way for the development of novel
514 therapeutic interventions. For example, chemotherapy kills only proliferating cancer cells, but
515 the survival of quiescent cancer cells, or cancer stem cells (non-dividing), is crucial for cancer to
516 return. The tumor microenvironment, especially changes in the composition and number of
517 inflammatory cells or cytokines, can activate cancer cells or CSCs to divide. So dare to imagine
518 the appropriate combination of inflammation and chemotherapy, the treatment of cancer can play
519 a better effect?

520 **Abbreviations**

521 AP-1, activating protein 1; ASCs, adult stem cells; ATM, ataxia telangiectasia-mutated gene;
522 ATR, ATM and rad3-related gene; BST2, bone marrow stromal cell antigen 2; CCL, chemokine
523 (C-C motif) ligand; C/EBP- δ , CCAA T/enhancer binding protein delta; cGAS, cyclic GMP-AMP
524 synthase; CLCs, clara-like cells; CLRs, C-type lectin receptors; CRC, colorectal cancer; CSCs,
525 cancer stem cells; DAMPs, damage-associated molecular patterns; EMT, epithelial-to-
526 mesenchymal transition; EP, prostaglandin E receptor; G-MSCs, gingival mesenchymal
527 stem/progenitor cells; gp130, glycoprotein 130; GSC, glioma stem cells; HIF-1 α , hypoxia-
528 inducible factor 1 alpha; HOXB5, homeobox protein Hox-B5; HSC, hematopoietic stem cell;
529 IFN, interferon; IL, Interleukin; IKK, I κ B kinases; ISCs, intestinal stem cells; Jag1, Jagged 1;
530 JAK, janus kinases; JNKs, jun N-terminal kinases; LPS, lipopolysaccharides; MAPK, mitogen-
531 activated protein kinase; MSCs, mesenchymal stem cells; MuSCs, muscle stem cells; MYD88,
532 myeloid differentiation primary response gene 88; NF- κ B, nuclear factor-k-gene binding; NLRs,
533 Nod-like receptors; Nrf2, Nuclear factor erythroid2-related factor 2; PAMPs, pathogen-
534 associated molecular patterns; PGE2, prostaglandin E2; PRRs, pattern recognition receptors;
535 RLRs, RIG-i-like receptors; ROS, reactive oxygen species; STAT, signal transducer and
536 activator of transcription; STING, stimulator of interferon genes; Th, T helper; TIR, Toll-IL-1-
537 resistance; TLRs, Toll-like receptors; TNF, tumor necrosis factor; Treg, regulatory T cells; TRIF,
538 toll-receptor-associated activator of interferon.

539 **Ethics approval and consent to participate**

540 Not applicable.

541 **References**

- 542 1. Bray F, Laversanne M, Weiderpass E, Soerjomataram I. The ever-increasing importance of cancer as a
543 leading cause of premature death worldwide. *Cancer*. 2021; 127 (16): 3029-30.
- 544 2. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018:
545 GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J*
546 *Clin*. 2018; 68 (6): 394-424.
- 547 3. Karpozilos A, Pavlidis N. The treatment of cancer in Greek antiquity. *Eur J Cancer*. 2004 ;40 (14): 2033-40.
- 548 4. Liu J. The dualistic origin of human tumors. *Semin Cancer Biol*. 2018; 53:1-16.
- 549 5. Allegra A, Alonci A, Penna G, Innao V, Gerace D, Rotondo F, Musolino C. The cancer stem cell hypothesis:
550 a guide to potential molecular targets. *Cancer Invest*. 2014; 32 (9): 470-95.
- 551 6. Sell S. On the stem cell origin of cancer. *Am J Pathol*. 2010; 176 (6): 2584-494.
- 552 7. Hanahan D WR. The hallmarks of cancer. *Cell*. 2000; 100 (1): 57-70.
- 553 8. Pandya R, Grace San Diego K, Shabbir T, Modi AP, Wang J, Dhahbi J, Barsky SH. The cell of cancer
554 origin provides the most reliable roadmap to its diagnosis, prognosis (biology) and therapy. *Med Hypotheses*.
555 2021; 157: 110704.
- 556 9. Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. *Nat Rev Cancer*. 2005; 5 (4): 275-84.
- 557 10. Zhou HM, Zhang JG, Zhang X, Li Q. Targeting cancer stem cells for reversing therapy resistance:
558 mechanism, signaling, and prospective agents. *Signal Transduct Target Ther*. 2021; 6 (1): 62.
- 559 11. Paul R, Dorsey JF, Fan Y. Cell plasticity, senescence, and quiescence in cancer stem cells: biological and
560 therapeutic implications. *Pharmacol Ther*. 2022; 231: 107985.
- 561 12. Hung KF, Yang T, Kao SY. Cancer stem cell theory: Are we moving past the mist? *J Chin Med Assoc*.
562 2019; 82 (11): 814-8.
- 563 13. White AC, Lowry WE. Refining the role for adult stem cells as cancer cells of origin. *Trends Cell Biol*.
564 2015; 25 (1): 11-20.
- 565 14. Tomasetti C, Vogelstein B. Cancer etiology. Variation in cancer risk among tissues can be explained by the

- 566 number of stem cell divisions. *Science*. 2015; 347 (6217): 78-81.
- 567 15. Tomasetti C, Li L, Vogelstein B. Stem cell divisions, somatic mutations, cancer etiology, and cancer
568 prevention. *Science*. 2017; 355 (6331).
- 569 16. López-Lázaro M. Stem cell division theory of cancer. *Cell Cycle*. 2015; 14 (16): 2547-8.
- 570 17. López-Lázaro M. The stem cell division theory of cancer. *Crit Rev Oncol Hematol*. 2018; 123: 95-113.
- 571 18. Miller FD, Kaplan DR. Mobilizing endogenous stem cells for repair and regeneration: are we there yet?
572 *Cell Stem Cell*. 2012; 10 (6): 650-2.
- 573 19. Cheung TH, Rando TA. Molecular regulation of stem cell quiescence. *Nat Rev Mol Cell Biol*. 2013; 14 (6):
574 329-40.
- 575 20. Machado L, Geara P, Camps J, Dos Santos M, Teixeira-Clerc F, Van Herck J, Varet H, Legendre R,
576 Pawlotsky JM, Sampaolesi M, Voet T, Maire P, Relaix F, Mourikis P. Tissue damage induces a conserved
577 stress response that initiates quiescent muscle stem cell activation. *Cell Stem Cell*. 2021; 28 (6): 1125-1135.e7.
- 578 21. Zhu L, Finkelstein D, Gao C, Shi L, Wang Y, López-Terrada D, Wang K, Utley S, Pounds S, Neale G,
579 Ellison D, Onar-Thomas A, Gilbertson RJ. Multi-organ mapping of cancer risk. *Cell*. 2016; 166 (5): 1132-
580 1146.e7.
- 581 22. Zhao Y, Simon M, Seluanov A, Gorbunova V. DNA damage and repair in age-related inflammation. *Nat*
582 *Rev Immunol*. 2023; 23 (2): 75-89.
- 583 23. Brzostek-Racine S, Gordon C, Van Scoy S, Reich NC. The DNA damage response induces IFN. *J*
584 *Immunol*. 2011; 187 (10):5336-5345.
- 585 24. Kondo T, Kobayashi J, Saitoh T, Maruyama K, Ishii KJ, Barber GN, Komatsu K, Akira S, Kawai T. DNA
586 damage sensor MRE11 recognizes cytosolic double-stranded DNA and induces type I interferon by regulating
587 STING trafficking. *Proc Natl Acad Sci U S A*. 2013; 110 (8): 2969-74.
- 588 25. Härtlova A, Erttmann SF, Raffi FA, Schmalz AM, Resch U, Anugula S, Lienenklaus S, Nilsson LM,
589 Kröger A, Nilsson JA, Ek T, Weiss S, Gekara NO. DNA damage primes the type I interferon system via the
590 cytosolic DNA sensor STING to promote anti-microbial innate immunity. *Immunity*. 2015; 42 (2): 332-343.
- 591 26. Nichane M, Javed A, Sivakamasundari V, Ganesan M, Ang LT, Kraus P, Lufkin T, Loh KM, Lim B.
592 Isolation and 3D expansion of multipotent Sox9⁺ mouse lung progenitors. *Nat Methods*. 2017; 14 (12): 1205-

- 593 12.
- 594 27. Qi K, Li N, Zhang Z, Melino G. Tissue regeneration: The crosstalk between mesenchymal stem cells and
595 immune response. *Cell Immunol.* 2018; 326: 86-93.
- 596 28. Nolan E, Bridgeman VL, Ombrato L, Karoutas A, Rabas N, Sewnath CAN, Vasquez M, Rodrigues FS,
597 Horswell S, Faull P, Carter R, Malanchi I. Radiation exposure elicits a neutrophil-driven response in healthy
598 lung tissue that enhances metastatic colonization. *Nat Cancer.* 2022; 3 (2): 173-187.
- 599 29. GBD 2019 Cancer Risk Factors Collaborators. The global burden of cancer attributable to risk factors,
600 2010-19: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet.* 2022; 400 (10352): 563-
601 591.
- 602 30. Liebelt BD, Finocchiaro G, Heimberger AB. Principles of immunotherapy. *Handb Clin Neurol.* 2016; 134:
603 163-81.
- 604 31. Pardoll D. Cancer and the Immune System: Basic Concepts and Targets for Intervention. *Semin Oncol.*
605 2015; 42 (4): 523-38.
- 606 32. Hosseini H, Obradović MMS, Hoffmann M, Harper KL, Sosa MS, Werner-Klein M, Nanduri LK, Werno
607 C, Ehrl C, Maneck M, Patwary N, Haunschild G, Gužvić M, Reimelt C, Grauvogl M, Eichner N, Weber F,
608 Hartkopf AD, Taran FA, Brucker SY, Fehm T, Rack B, Buchholz S, Spang R, Meister G, Aguirre-Ghiso JA,
609 Klein CA. Early dissemination seeds metastasis in breast cancer. *Nature.* 2016; 540 (7634): 552-8.
- 610 33. Hu Z, Li Z, Ma Z, Curtis C. Multi-cancer analysis of clonality and the timing of systemic spread in paired
611 primary tumors and metastases. *Nat Genet.* 2020; 52 (7): 701-8.
- 612 34. Harper KL, Sosa MS, Entenberg D, Hosseini H, Cheung JF, Nobre R, Avivar-Valderas A, Nagi C, Girnius
613 N, Davis RJ, Farias EF, Condeelis J, Klein CA, Aguirre-Ghiso JA. Mechanism of early dissemination and
614 metastasis in Her2+ mammary cancer. *Nature.* 2016; 540 (7634): 558-92.
- 615 35. Bado IL, Zhang W, Hu J, Xu Z, Wang H, Sarkar P, Li L, Wan YW, Liu J, Wu W, Lo HC, Kim IS, Singh S,
616 Janghorban M, Muscarella AM, Goldstein A, Singh P, Jeong HH, Liu C, Schiff R, Huang S, Ellis MJ, Gaber
617 MW, Gugala Z, Liu Z, Zhang XH. The bone microenvironment increases phenotypic plasticity of ER+ breast
618 cancer cells. *Dev Cell.* 2021; 56 (8): 1100-17.e9.
- 619 36. Liu Y, Cao X. Characteristics and significance of the pre-metastatic Niche. *Cancer Cell.* 2016; 30 (5): 668-

- 620 81.
- 621 37. Zhang W, Bado IL, Hu J, Wan YW, Wu L, Wang H, Gao Y, Jeong HH, Xu Z, Hao X, Lege BM, Al-Ouran
622 R, Li L, Li J, Yu L, Singh S, Lo HC, Niu M, Liu J, Jiang W, Li Y, Wong STC, Cheng C, Liu Z, Zhang XH.
623 The bone microenvironment invigorates metastatic seeds for further dissemination. *Cell*. 2021; 184 (9): 2471-
624 86.e20.
- 625 38. Sánchez-Danés A, Hannezo E, Larsimont JC, Liagre M, Youssef KK, Simons BD, Blanpain C. Defining
626 the clonal dynamics leading to mouse skin tumour initiation. *Nature*. 2016; 536 (7616): 298-303.
- 627 39. Quesenberry PJ, Goldberg LR. Stem cell divisions and cancer. *Leukemia*. 2015; 29 (10): 1959.
- 628 40. Zinger A, Cho WC, Ben-Yehuda A. Cancer and aging - the inflammatory connection. *Aging Dis*. 2017; 8
629 (5): 611-27.
- 630 41. Hsu T. Educational initiatives in geriatric oncology - Who, why, and how? *J Geriatr Oncol*. 2016; 7 (5):
631 390-6.
- 632 42. IL W. Stem cells: units of development, units of regeneration, and units in evolution. *Cell*. 2000; 100 (1):
633 157-68.
- 634 43. Ge Y, Miao Y, Gur-Cohen S, Gomez N, Yang H, Nikolova M, Polak L, Hu Y, Verma A, Elemento O,
635 Krueger JG, Fuchs E. The aging skin microenvironment dictates stem cell behavior. *Proc Natl Acad Sci U S A*.
636 2020; 117 (10): 5339-50.
- 637 44. Duckworth CA. Identifying key regulators of the intestinal stem cell niche. *Biochem Soc Trans*. 2021; 49
638 (5): 2163-76.
- 639 45. Li Y, Guo W. Neural stem cell niche and adult neurogenesis. *Neuroscientist*. 2021; 27 (3): 235-45.
- 640 46. Hsu YC, Fuchs E. A family business: stem cell progeny join the niche to regulate homeostasis. *Nat Rev*
641 *Mol Cell Biol*. 2012; 13 (2): 103-14.
- 642 47. Zindel J, Kubes P. DAMPs, PAMPs, and LAMPs in immunity and sterile inflammation. *Annu Rev Pathol*.
643 2020; 15: 493-518.
- 644 48. Sun SC. The non-canonical NF- κ B pathway in immunity and inflammation. *Nat Rev Immunol*. 2017; 17
645 (9): 545-58.
- 646 49. Castaño Z, San Juan BP, Spiegel A, Pant A, DeCristo MJ, Laszewski T, Ubellacker JM, Janssen SR,

- 647 Dongre A, Reinhardt F, Henderson A, Del Rio AG, Gifford AM, Herbert ZT, Hutchinson JN, Weinberg RA,
648 Chaffer CL, McAllister SS. IL-1 β inflammatory response driven by primary breast cancer prevents metastasis-
649 initiating cell colonization. *Nat Cell Biol.* 2018; 20 (9): 1084-97.
- 650 50. Rossi DJ, Jamieson CH, Weissman IL. Stems cells and the pathways to aging and cancer. *Cell.* 2008; 132
651 (4): 681-96.
- 652 51. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer
653 suppression and promotion. *Science.* 2011; 331 (6024): 1565-70.
- 654 52. Askeland EJ, Newton MR, O'Donnell MA, Luo Y. Bladder cancer immunotherapy: BCG and beyond. *Adv*
655 *Urol.* 2012; 2012: 181987.
- 656 53. Havell EA, Fiers W, North RJ. The antitumor function of tumor necrosis factor (TNF), I. Therapeutic
657 action of TNF against an established murine sarcoma is indirect, immunologically dependent, and limited by
658 severe toxicity. *J Exp Med.* 1988; 167 (3): 1067-85.
- 659 54. Balkwill F. TNF-alpha in promotion and progression of cancer. *Cancer Metastasis Rev.* 2006; 25 (3): 409-
660 16.
- 661 55. Muthusami S, Ramachandran IK, Babu KN, Krishnamoorthy S, Guruswamy A, Queimado L, Chaudhuri G,
662 Ramachandran I. Role of inflammation in the development of colorectal cancer. *Endocr Metab Immune Disord*
663 *Drug Targets.* 2021; 21 (1): 77-90.
- 664 56. Hirano T. IL-6 in inflammation, autoimmunity and cancer. *Int Immunol.* 2021; 33 (3): 127-148.
- 665 57. Medzhitov R. Origin and physiological roles of inflammation. *Nature.* 2008; 454 (7203): 428-35.
- 666 58. McDonald B, Pittman K, Menezes GB, Hirota SA, Slaba I, Waterhouse CC, Beck PL, Muruve DA, Kubes
667 P. Intravascular danger signals guide neutrophils to sites of sterile inflammation. *Science.* 2010; 330 (6002):
668 362-366.
- 669 59. Wicherska-Pawłowska K, Wróbel T, Rybka J. Toll-like receptors (TLRs), NOD-like receptors (NLRs), and
670 RIG-I-like receptors (RLRs) in innate immunity. TLRs, NLRs, and RLRs ligands as immunotherapeutic agents
671 for hematopoietic diseases. *Int J Mol Sci.* 2021; 22 (24): 13397.
- 672 60. O'Neill LA, Golenbock D, Bowie AG. The history of Toll-like receptors - redefining innate immunity. *Nat*
673 *Rev Immunol.* 2013; 13 (6): 453-60.

- 674 61. Gong T, Liu L, Jiang W, Zhou R. DAMP-sensing receptors in sterile inflammation and inflammatory
675 diseases. *Nat Rev Immunol*. 2020; 20 (2): 95-112.
- 676 62. Chefetz I, Alvero AB, Holmberg JC, Lebowitz N, Craveiro V, Yang-Hartwich Y, Yin G, Squillace L,
677 Gurrea Soteras M, Aldo P, Mor G. TLR2 enhances ovarian cancer stem cell self-renewal and promotes tumor
678 repair and recurrence. *Cell Cycle*. 2013; 12 (3): 511-521.
- 679 63. Goodell MA, Nguyen H, Shroyer N. Somatic stem cell heterogeneity: diversity in the blood, skin and
680 intestinal stem cell compartments. *Nat Rev Mol Cell Biol*. 2015; 16 (5): 299-309.
- 681 64. Sueda R, Kageyama R. Regulation of active and quiescent somatic stem cells by Notch signaling. *Dev*
682 *Growth Differ*. 2020; 62 (1): 59-66.
- 683 65. Rossi F, Noren H, Jove R, Beljanski V, Grinnemo KH. Differences and similarities between cancer and
684 somatic stem cells: therapeutic implications. *Stem Cell Res Ther*. 2020; 11 (1): 489.
- 685 66. Rossi L, Lin KK, Boles NC, Yang L, King KY, Jeong M, Mayle A, Goodell MA. Less is more: unveiling
686 the functional core of hematopoietic stem cells through knockout mice. *Cell Stem Cell*. 2012; 11 (3): 302-317.
- 687 67. Walter D, Lier A, Geiselhart A, Thalheimer FB, Huntscha S, Sobotta MC, Moehrle B, Brocks D, Bayindir
688 I, Kaschutnig P, Muedder K, Klein C, Jauch A, Schroeder T, Geiger H, Dick TP, Holland-Letz T, Schmezer P,
689 Lane SW, Rieger MA, Essers MA, Williams DA, Trumpp A, Milsom MD. Exit from dormancy provokes
690 DNA-damage-induced attrition in haematopoietic stem cells. *Nature*. 2015; 520 (7548): 549-552.
- 691 68. Cho IJ, Lui PP, Obajdin J, Riccio F, Stroukov W, Willis TL, Spagnoli F, Watt FM. Mechanisms, hallmarks,
692 and implications of stem cell quiescence. *Stem Cell Reports*. 2019; 12 (6): 1190-1200.
- 693 69. Schuettpelz LG, Link DC. Regulation of hematopoietic stem cell activity by inflammation. *Front Immunol*.
694 2013; 4: 204.
- 695 70. Hochmuth CE, Biteau B, Bohmann D, Jasper H. Redox regulation by Keap1 and Nrf2 controls intestinal
696 stem cell proliferation in *Drosophila*. *Cell Stem Cell*. 2011; 8 (2): 188-199.
- 697 71. Biteau B, Hochmuth CE, Jasper H. JNK activity in somatic stem cells causes loss of tissue homeostasis in
698 the aging *Drosophila* gut. *Cell Stem Cell*. 2008; 3 (4): 442-455.
- 699 72. Verckist L, Pintelon I, Timmermans JP, Brouns I, Adriaensen D. Selective activation and proliferation of a
700 quiescent stem cell population in the neuroepithelial body microenvironment. *Respir Res*. 2018; 19 (1): 207.

- 701 73. Florez MA, Matatall KA, Jeong Y, Ortinou L, Shafer PW, Lynch AM, Jaksik R, Kimmel M, Park D, King
702 KY. Interferon gamma mediates hematopoietic stem cell activation and niche relocalization through BST2.
703 *Cell Rep.* 2020; 33 (12): 108530.
- 704 74. Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, Scheller J, Rose-John S,
705 Cheroutre H, Eckmann L, Karin M. IL-6 and Stat3 are required for survival of intestinal epithelial cells and
706 development of colitis-associated cancer. *Cancer Cell.* 2009; 15 (2): 103-13.
- 707 75. Böhm F, Köhler UA, Speicher T, Werner S. Regulation of liver regeneration by growth factors and
708 cytokines. *EMBO Mol Med.* 2010; 2 (8): 294-305.
- 709 76. Nikoopour E, Bellemore SM, Singh B. IL-22, cell regeneration and autoimmunity. *Cytokine.* 2015; 74 (1):
710 35-42.
- 711 77. Naik S, Larsen SB, Cowley CJ, Fuchs E. Two to tango: dialog between immunity and stem cells in health
712 and disease. *Cell.* 2018; 175 (4): 908-20.
- 713 78. Lindemans CA, Calafiore M, Mertelsmann AM, O'Connor MH, Dudakov JA, Jenq RR, Velardi E, Young
714 LF, Smith OM, Lawrence G, Ivanov JA, Fu YY, Takashima S, Hua G, Martin ML, O'Rourke KP, Lo YH,
715 Mokry M, Romera-Hernandez M, Cupedo T, Dow L, Nieuwenhuis EE, Shroyer NF, Liu C, Kolesnick R, van
716 den Brink MRM, Hanash AM. Interleukin-22 promotes intestinal stem cell mediated epithelial regeneration.
717 *Nature.* 2015; 528 (7583).
- 718 79. von Moltke J, Ji M, Liang HE, Locksley RM. Tuft-cell-derived IL-25 regulates an intestinal ILC2-
719 epithelial response circuit. *Nature.* 2016; 529 (7585): 221-225.
- 720 80. Yoshida K, Gowers KHC, Lee-Six H, Chandrasekharan DP, Coorens T, Maughan EF, Beal K, Menzies A,
721 Millar FR, Anderson E, Clarke SE, Pennycuik A, Thakrar RM, Butler CR, Kakiuchi N, Hirano T, Hynds RE,
722 Stratton MR, Martincorena I, Janes SM, Campbell PJ. Tobacco smoking and somatic mutations in human
723 bronchial epithelium. *Nature.* 2020; 578 (7794): 266-272.
- 724 81. Beyaz S, Mana MD, Roper J, Kedrin D, Saadatpour A, Hong SJ, Bauer-Rowe KE, Xifaras ME, Akkad A,
725 Arias E, Pinello L, Katz Y, Shinagare S, Abu-Remaileh M, Mihaylova MM, Lamming DW, Dogum R, Guo G,
726 Bell GW, Selig M, Nielsen GP, Gupta N, Ferrone CR, Deshpande V, Yuan GC, Orkin SH, Sabatini DM,
727 Yilmaz ÖH. High-fat diet enhances stemness and tumorigenicity of intestinal progenitors. *Nature.* 2016; 531

- 728 (7592): 53-8.
- 729 82. Beyaz S, Chung C, Mou H, Bauer-Rowe KE, Xifaras ME, Ergin I, Dohnalova L, Biton M, Shekhar K,
730 Eskiocak O, Papciak K, Ozler K, Almeqdadi M, Yueh B, Fein M, Annamalai D, Valle-Encinas E, Erdemir A,
731 Dogum K, Shah V, Alici-Garipcan A, Meyer HV, Özata DM, Elinav E, Kucukural A, Kumar P, McAleer JP,
732 Fox JG, Thaiss CA, Regev A, Roper J, Orkin SH, Yilmaz ÖH. Dietary suppression of MHC class II expression
733 in intestinal epithelial cells enhances intestinal tumorigenesis. *Cell Stem Cell*. 2021; 28 (11): 1922-35.e5.
- 734 83. Degirmenci B, Valenta T, Dimitrieva S, Hausmann G, Basler K. GLI1-expressing mesenchymal cells form
735 the essential Wnt-secreting niche for colon stem cells. *Nature*. 2018; 558 (7710): 449-53.
- 736 84. Redondo-Castro E, Cunningham C, Miller J, Martuscelli L, Aoulad-Ali S, Rothwell NJ, Kielty CM, Allan
737 SM, Pinteaux E. Interleukin-1 primes human mesenchymal stem cells towards an anti-inflammatory and pro-
738 trophic phenotype in vitro. *Stem Cell Res Ther*. 2017; 8 (1): 79.
- 739 85. Fu X, Xiao J, Wei Y, Li S, Liu Y, Yin J, Sun K, Sun H, Wang H, Zhang Z, Zhang BT, Sheng C, Wang H,
740 Hu P. Combination of inflammation-related cytokines promotes long-term muscle stem cell expansion. *Cell*
741 *Res*. 2015; 25 (6): 655-673.
- 742 86. Zhang F, Si M, Wang H, Mekhemar MK, Dörfer CE, Fawzy El-Sayed KM. IL-1/TNF- α inflammatory and
743 anti-inflammatory synchronization affects gingival stem/progenitor cells' regenerative attributes. *Stem Cells Int*.
744 2017; 2017: 1349481.
- 745 87. Ho ATV, Palla AR, Blake MR, Yucel ND, Wang YX, Magnusson KEG, Holbrook CA, Kraft PE, Delp SL,
746 Blau HM. Prostaglandin E2 is essential for efficacious skeletal muscle stem-cell function, augmenting
747 regeneration and strength. *Proc Natl Acad Sci U S A*. 2017; 114 (26): 6675-6684.
- 748 88. Liu Y, Zhang Z, Wang B, Dong Y, Zhao C, Zhao Y, Zhang L, Liu X, Guo J, Chen Y, Zhou J, Yang T,
749 Wang Y, Liu H, Wang S. Inflammation-stimulated MSC-derived small extracellular vesicle miR-27b-3p
750 regulates macrophages by targeting CSF-1 to promote temporomandibular joint condylar regeneration. *Small*.
751 2022; 11: e2107354.
- 752 89. Shi Y, Hu G, Su J, Li W, Chen Q, Shou P, Xu C, Chen X, Huang Y, Zhu Z, Huang X, Han X, Xie N, Ren
753 G. Mesenchymal stem cells: a new strategy for immunosuppression and tissue repair. *Cell Res*. 2010; 20 (5):
754 510-518.

- 755 90. Wang Y, Chen X, Cao W, Shi Y. Plasticity of mesenchymal stem cells in immunomodulation: pathological
756 and therapeutic implications. *Nat Immunol.* 2014; 15 (11): 1009-1016.
- 757 91. Zhu P, Lu T, Wu J, Fan D, Liu B, Zhu X, Guo H, Du Y, Liu F, Tian Y, Fan Z. Gut microbiota drives
758 macrophage-dependent self-renewal of intestinal stem cells via niche enteric serotonergic neurons. *Cell Res.*
759 2022; 32 (6): 555-569.
- 760 92. Biton M, Haber AL, Rogel N, Burgin G, Beyaz S, Schnell A, Ashenberg O, Su CW, Smillie C, Shekhar K,
761 Chen Z, Wu C, Ordovas-Montanes J, Alvarez D, Herbst RH, Zhang M, Tirosh I, Dionne D, Nguyen LT,
762 Xifaras ME, Shalek AK, von Andrian UH, Graham DB, Rozenblatt-Rosen O, Shi HN, Kuchroo V, Yilmaz OH,
763 Regev A, Xavier RJ. T helper cell cytokines modulate intestinal stem cell renewal and differentiation. *Cell.*
764 2018; 175 (5): 1307-20.
- 765 93. Ali N, Zirak B, Rodriguez RS, Pauli ML, Truong HA, Lai K, Ahn R, Corbin K, Lowe MM, Scharschmidt
766 TC, Taravati K, Tan MR, Ricardo-Gonzalez RR, Nosbaum A, Bertolini M, Liao W, Nestle FO, Paus R,
767 Cotsarelis G, Abbas AK, Rosenblum MD. Regulatory T cells in skin facilitate epithelial stem cell
768 differentiation. *Cell.* 2017; 169 (6): 1119-1129.
- 769 94. Keyes BE, Liu S, Asare A, Naik S, Levorse J, Polak L, Lu CP, Nikolova M, Pasolli HA, Fuchs E. Impaired
770 epidermal to dendritic T cell signaling slows wound repair in aged skin. *Cell.* 2016; 167 (5): 1323-1338.
- 771 95. Huang T, Song X, Xu D, Tiek D, Goenka A, Wu B, Sastry N, Hu B, Cheng SY. Stem cell programs in
772 cancer initiation, progression, and therapy resistance. *Theranostics.* 2020; 10 (19): 8721-8743.
- 773 96. Clara JA, Monge C, Yang Y, Takebe N. Targeting signalling pathways and the immune microenvironment
774 of cancer stem cells - a clinical update. *Nat Rev Clin Oncol.* 2020; 17 (4): 204-232.
- 775 97. Yang L, Shi P, Zhao G, Xu J, Peng W, Zhang J, Zhang G, Wang X, Dong Z, Chen F, Cui H. Targeting
776 cancer stem cell pathways for cancer therapy. *Signal Transduct Target Ther.* 2020; 5 (1): 8.
- 777 98. Zhan T, Rindtorff N, Boutros M. Wnt signaling in cancer. *Oncogene.* 2017; 36 (11): 1461-1473.
- 778 99. Essex A, Pineda J, Acharya G, Xin H, Evans J; Reproducibility Project: Cancer Biology. Replication Study:
779 Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *Elife.* 2019; 8: e45426.
- 780 100. Karami Fath M, Ebrahimi M, Nourbakhsh E, Zia Hazara A, Mirzaei A, Shafieyari S, Salehi A,
781 Hoseinzadeh M, Payandeh Z, Barati G. PI3K/Akt/mTOR signaling pathway in cancer stem cells. *Pathol Res*

- 782 *Pract.* 2022; 237: 154010.
- 783 101. Gasmi I, Machou C, Rodrigues A, Brouillet A, Nguyen TC, Rousseau B, Guillot A, Rodriguez C,
784 Demontant V, Ait-Ahmed Y, Calderaro J, Luciani A, Pawlotsky JM, Lafdil F. Interleukin-17 programs liver
785 progenitor cell transformation into cancer stem cells through miR-122 downregulation with increased risk of
786 primary liver cancer initiation. *Int J Biol Sci.* 2022; 18 (5): 1944-1960.
- 787 102. Marotta LL, Almendro V, Marusyk A, Shipitsin M, Schemme J, Walker SR, Bloushtain-Qimron N, Kim
788 JJ, Choudhury SA, Maruyama R, Wu Z, Gönen M, Mulvey LA, Bessarabova MO, Huh SJ, Silver SJ, Kim SY,
789 Park SY, Lee HE, Anderson KS, Richardson AL, Nikolskaya T, Nikolsky Y, Liu XS, Root DE, Hahn WC,
790 Frank DA, Polyak K. The JAK2/STAT3 signaling pathway is required for growth of CD44⁺CD24⁻ stem cell-
791 like breast cancer cells in human tumors. *J Clin Invest.* 2011; 121 (7): 2723-2735.
- 792 103. Kim SY, Kang JW, Song X, Kim BK, Yoo YD, Kwon YT, Lee YJ. Role of the IL-6-JAK1-STAT3-Oct-4
793 pathway in the conversion of non-stem cancer cells into cancer stem-like cells. *Cell Signal.* 2013; 25 (4): 961-
794 969.
- 795 104. Li XF, Chen C, Xiang DM, Qu L, Sun W, Lu XY, Zhou TF, Chen SZ, Ning BF, Cheng Z, Xia MY, Shen
796 WF, Yang W, Wen W, Lee TKW, Cong WM, Wang HY, Ding J. Chronic inflammation-elicited liver
797 progenitor cell conversion to liver cancer stem cell with clinical significance. *Hepatology.* 2017; 66 (6): 1934-
798 1951.
- 799 105. Zhao X, Ma L, Dai L, Zuo D, Li X, Zhu H, Xu F. TNF- α promotes the malignant transformation of
800 intestinal stem cells through the NF- κ B and Wnt/ β -catenin signaling pathways. *Oncol Rep.* 2020; 44 (2): 577-
801 588.
- 802 106. Balamurugan K, Mendoza-Villanueva D, Sharan S, Summers GH, Dobrolecki LE, Lewis MT, Sterneck E.
803 C/EBP δ links IL-6 and HIF-1 signaling to promote breast cancer stem cell-associated phenotypes. *Oncogene.*
804 2019; 38 (20): 3765-3780.
- 805 107. Zhao J, Jiang Y, Zhang H, Zhou J, Chen L, Li H, Xu J, Zhang G, Jing Z. The SRSF1/circATP5B/miR-
806 185-5p/HOXB5 feedback loop regulates the proliferation of glioma stem cells via the IL6-mediated
807 JAK2/STAT3 signaling pathway. *J Exp Clin Cancer Res.* 2021; 40 (1): 134.
- 808 108. Chiou GY, Chien CS, Wang ML, Chen MT, Yang YP, Yu YL, Chien Y, Chang YC, Shen CC, Chio CC,

- 809 Lu KH, Ma HI, Chen KH, Liu DM, Miller SA, Chen YW, Huang PI, Shih YH, Hung MC, Chiou SH.
810 Epigenetic regulation of the miR142-3p/interleukin-6 circuit in glioblastoma. *Mol Cell*. 2013; 52 (5): 693-706.
811 109. Lee SH, Hong HS, Liu ZX, Kim RH, Kang MK, Park NH, Shin KH. TNF α enhances cancer stem cell-
812 like phenotype via Notch-Hes1 activation in oral squamous cell carcinoma cells. *Biochem Biophys Res*
813 *Commun*. 2012; 424 (1): 58-64.
- 814 110. Jiang Y, Zhao J, Liu Y, Hu J, Gao L, Wang H, Cui D. CircKPNB1 mediates a positive feedback loop and
815 promotes the malignant phenotypes of GSCs via TNF- α /NF- κ B signaling. *Cell Death Dis*. 2022; 13 (8): 697.
- 816 111. Wang D, Fu L, Sun H, Guo L, DuBois RN. Prostaglandin E2 promotes colorectal cancer stem cell
817 expansion and metastasis in mice. *Gastroenterology*. 2015; 149 (7): 1884-1895.e4.
- 818 112. Shen W, Zhang X, Tang J, Zhang Z, Du R, Luo D, Liu X, Xia Y, Li Y, Wang S, Yan S, Yang W, Xiang
819 R, Luo N, Luo Y, Li J. CCL16 maintains stem cell-like properties in breast cancer by activating
820 CCR2/GSK3 β / β -catenin/OCT4 axis. *Theranostics*. 2021; 11 (5): 2297-2317.
- 821 113. Xiang T, Long H, He L, Han X, Lin K, Liang Z, Zhuo W, Xie R, Zhu B. Interleukin-17 produced by
822 tumor microenvironment promotes self-renewal of CD133+ cancer stem-like cells in ovarian cancer.
823 *Oncogene*. 2015; 34 (2): 165-176.
- 824 114. Stoeltzing O, Meric-Bernstam F, Ellis LM. Intracellular signaling in tumor and endothelial cells: The
825 expected and, yet again, the unexpected. *Cancer Cell*. 2006; 10 (2): 89-91.
- 826 115. Galdiero MR MG, Mantovani A. Cancer inflammation and cytokines. *Cold Spring Harb Perspect Biol*.
827 2018; 10 (8): a028662.
- 828 116. Coussens LM, Werb Z. Inflammation and cancer. *Nature*. 2002; 420 (6917): 860-867.
- 829 117. Anuja K, Roy S, Ghosh C, Gupta P, Bhattacharjee S, Banerjee B. Prolonged inflammatory
830 microenvironment is crucial for pro-neoplastic growth and genome instability: a detailed review. *Inflamm Res*.
831 2017; 66 (2): 119-128.
- 832 118. Wang GZ, Cheng X, Li XC, Liu YQ, Wang XQ, Shi X, Wang ZY, Guo YQ, Wen ZS, Huang YC, Zhou
833 GB. Tobacco smoke induces production of chemokine CCL20 to promote lung cancer. *Cancer Lett*. 2015; 363
834 (1): 60-70.
- 835 119. Park EJ, Lee JH, Yu GY, He G, Ali SR, Holzer RG, Osterreicher CH, Takahashi H, Karin M. Dietary and

- 836 genetic obesity promote liver inflammation and tumorigenesis by enhancing IL-6 and TNF expression. *Cell*.
837 2010; 140 (2): 197-208.
- 838 120. Fischbach W, Malfertheiner P. *Helicobacter Pylori* infection. *Dtsch Arztebl Int*. 2018; 115 (25): 429-36.
- 839 121. Institute for Research on Aging CGARNAECoMABSBCoMBRCoHB. Integrated genomic and
840 molecular characterization of cervical cancer. *Nature*. 2017; 543 (7645): 378-384.
- 841 122. Punt S, Dronkers EA, Welters MJ, Goedemans R, Koljenović S, Bloemena E, Snijders PJ, Gorter A, van
842 der Burg SH, Baatenburg de Jong RJ, Jordanova ES. A beneficial tumor microenvironment in oropharyngeal
843 squamous cell carcinoma is characterized by a high T cell and low IL-17(+) cell frequency. *Cancer Immunol*
844 *Immunother*. 2016; 65 (4): 393-403.
- 845 123. Kay J, Thadhani E, Samson L, Engelward B. Inflammation-induced DNA damage, mutations and cancer.
846 *DNA Repair (Amst)*. 2019; 83: 102673.
- 847 124. Colotta F, Allavena P, Sica A, Garlanda C, Mantovani A. Cancer-related inflammation, the seventh
848 hallmark of cancer: links to genetic instability. *Carcinogenesis*. 2009; 30 (7): 1073-1081.
- 849 125. Pikor L, Thu K, Vucic E, Lam W. The detection and implication of genome instability in cancer. *Cancer*
850 *Metastasis Rev*. 2013; 32 (3-4): 341-352.
- 851 126. Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, Gutkovich-Pyest E, Urieli-Shoval S,
852 Galun E, Ben-Neriah Y. NF-kappaB functions as a tumour promoter in inflammation-associated cancer.
853 *Nature*. 2004; 431 (7007): 461-466.
- 854 127. Kiraly O, Gong G, Olipitz W, Muthupalani S, Engelward BP. Inflammation-induced cell proliferation
855 potentiates DNA damage-induced mutations in vivo. *PLoS Genet*. 2015; 11 (2): e1004901.
- 856 128. Xu M, Wang Y, Xia R, Wei Y, Wei X. Role of the CCL2-CCR2 signalling axis in cancer: Mechanisms
857 and therapeutic targeting. *Cell Prolif*. 2021; 54 (10): e13115.
- 858 129. Aggarwal BB, Shishodia S, Sandur SK, Pandey MK, Sethi G. Inflammation and cancer: how hot is the
859 link? *Biochem Pharmacol*. 2006; 72 (11): 1605-1621.
- 860 130. He G, Karin M. NF- κ B and STAT3 - key players in liver inflammation and cancer. *Cell Res*. 2011; 21 (1):
861 159-168.
- 862 131. Hellwig-Bürgel T, Rutkowski K, Metzen E, Fandrey J, Jelkmann W. Interleukin-1beta and tumor necrosis

- 863 factor-alpha stimulate DNA binding of hypoxia-inducible factor-1. *Blood*. 1999; 94 (5): 1561-1567.
- 864 132. Albregues J, Shields MA, Ng D, Park CG, Ambrico A, Poindexter ME, Upadhyay P, Uyeminami DL,
865 Pommier A, Küttner V, Bružas E, Maiorino L, Bautista C, Carmona EM, Gimotty PA, Fearon DT, Chang K,
866 Lyons SK, Pinkerton KE, Trotman LC, Goldberg MS, Yeh JT, Egeblad M. Neutrophil extracellular traps
867 produced during inflammation awaken dormant cancer cells in mice. *Science*. 2018; 361 (6409): eaao4227.
- 868 133. Keshari AK, Singh AK, Kumar U, Raj V, Rai A, Kumar P, Kumar D, Maity B, Nath S, Prakash A, Saha
869 S. 5H-benzo[h]thiazolo[2,3-b]quinazolines ameliorate NDEA-induced hepatocellular carcinogenesis in rats
870 through IL-6 downregulation along with oxidative and metabolic stress reduction. *Drug Des Devel Ther*. 2017;
871 11: 2981-95.
- 872 134. Barker EC, Kim BG, Yoon JH, Tochtrop GP, Letterio JJ, Choi SH. Potent suppression of both
873 spontaneous and carcinogen-induced colitis-associated colorectal cancer in mice by dietary celastrol
874 supplementation. *Carcinogenesis*. 2018; 39 (1): 36-46.
- 875 135. Chikara S, Mamidi S, Sreedasyam A, Chittam K, Pietrofesa R, Zuppa A, Moorthy G, Dyer N,
876 Christofidou-Solomidou M, Reindl KM. Flaxseed consumption inhibits chemically induced lung
877 tumorigenesis and modulates expression of phase II enzymes and inflammatory cytokines in A/J mice. *Cancer*
878 *Prev Res (Phila)*. 2018; 11 (1): 27-37.
- 879 136. Suarez-Carmona M, Lesage J, Cataldo D, Gilles C. EMT and inflammation: inseparable actors of cancer
880 progression. *Mol Oncol*. 2017; 11 (7): 805-823.
- 881 137. Sepich-Poore GD, Zitvogel L, Straussman R, Hasty J, Wargo JA, Knight R. The microbiome and human
882 cancer. *Science*. 2021; 371 (6536): eabc4552.
- 883 138. Su S, Liu Q, Chen J, Chen J, Chen F, He C, Huang D, Wu W, Lin L, Huang W, Zhang J, Cui X, Zheng F,
884 Li H, Yao H, Su F, Song E. A positive feedback loop between mesenchymal-like cancer cells and
885 macrophages is essential to breast cancer metastasis. *Cancer Cell*. 2014; 25 (5): 605-620.
- 886 139. Yost KE, Ciliberto G, Wells DK, Qi Y, Wang C, Kageyama R, McNamara KL, Granja JM, Sarin KY,
887 Brown RA, Gupta RK, Curtis C, Bucktrout SL, Davis MM, Chang ALS, Chang HY. Clonal replacement of
888 tumor-specific T cells following PD-1 blockade. *Nat Med*. 2019; 25 (8): 1251-1259.
- 889 140. Bruschini S, Ciliberto G, Mancini R. The emerging role of cancer cell plasticity and cell-cycle quiescence

890 in immune escape. *Cell Death Dis.* 2020; 11 (6): 471.

891 141. Shi Y, Lim SK, Liang Q, Iyer SV, Wang HY, Wang Z, Xie X, Sun D, Chen YJ, Tabar V, Gutin P,
892 Williams N, De Brabander JK, Parada LF. Gboxin is an oxidative phosphorylation inhibitor that targets
893 glioblastoma. *Nature.* 2019; 567 (7748): 341-346.

894 142. Sato K, Padgaonkar AA, Baker SJ, Cosenza SC, Rechkoblit O, Subbaiah DRCV, Domingo-Domenech J,
895 Bartkowski A, Port ER, Aggarwal AK, Ramana Reddy MV, Irie HY, Reddy EP. Simultaneous
896 CK2/TNIK/DYRK1 inhibition by 108600 suppresses triple negative breast cancer stem cells and
897 chemotherapy-resistant disease. *Nat Commun.* 2021; 12 (1): 4671.

898 143. Ohta Y, Fujii M, Takahashi S, Takano A, Nanki K, Matano M, Hanyu H, Saito M, Shimokawa M,
899 Nishikori S, Hatano Y, Ishii R, Sawada K, Machinaga A, Ikeda W, Imamura T, Sato T. Cell-matrix interface
900 regulates dormancy in human colon cancer stem cells. *Nature.* 2022; 608 (7924): 784-794.

901 144. Hayakawa Y, Nakagawa H, Rustgi AK, Que J, Wang TC. Stem cells and origins of cancer in the upper
902 gastrointestinal tract. *Cell Stem Cell.* 2021; 28 (8): 1343-1361.

903 145. Aliluev A, Tritschler S, Sterr M, Oppenländer L, Hinterdobler J, Greisle T, Irmeler M, Beckers J, Sun N,
904 Walch A, Stemmer K, Kindt A, Krumsiek J, Tschöp MH, Luecken MD, Theis FJ, Lickert H, Böttcher A. Diet-
905 induced alteration of intestinal stem cell function underlies obesity and prediabetes in mice. *Nat Metab.* 2021;
906 3 (9): 1202-1216.

907

Figure 1

Tissue formation and distribution of adult stem cells in the body.

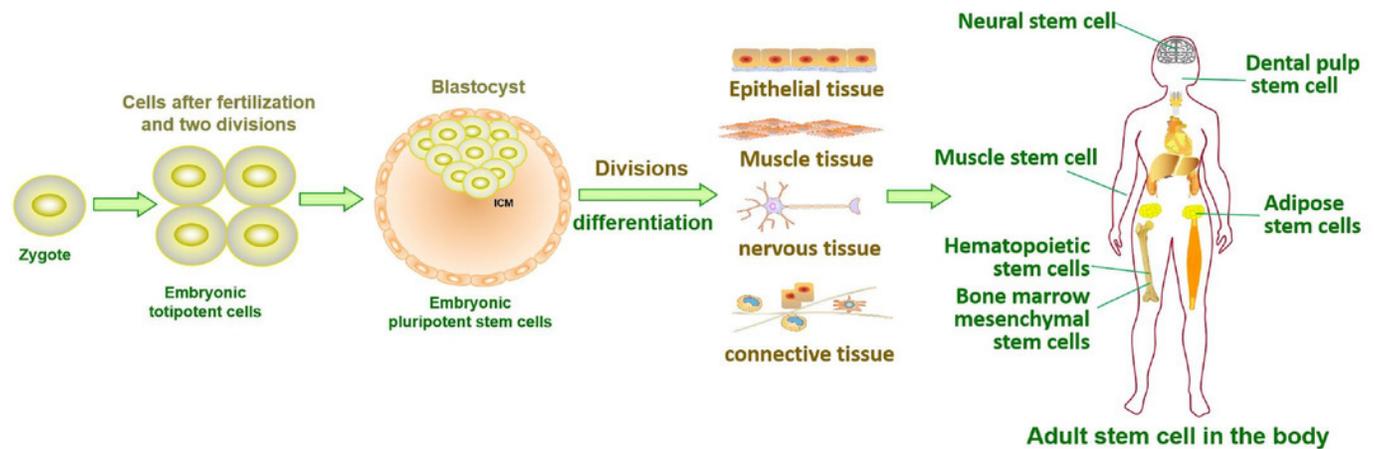


Figure 2

Multi-pathway inflammatory signaling and tissue repair

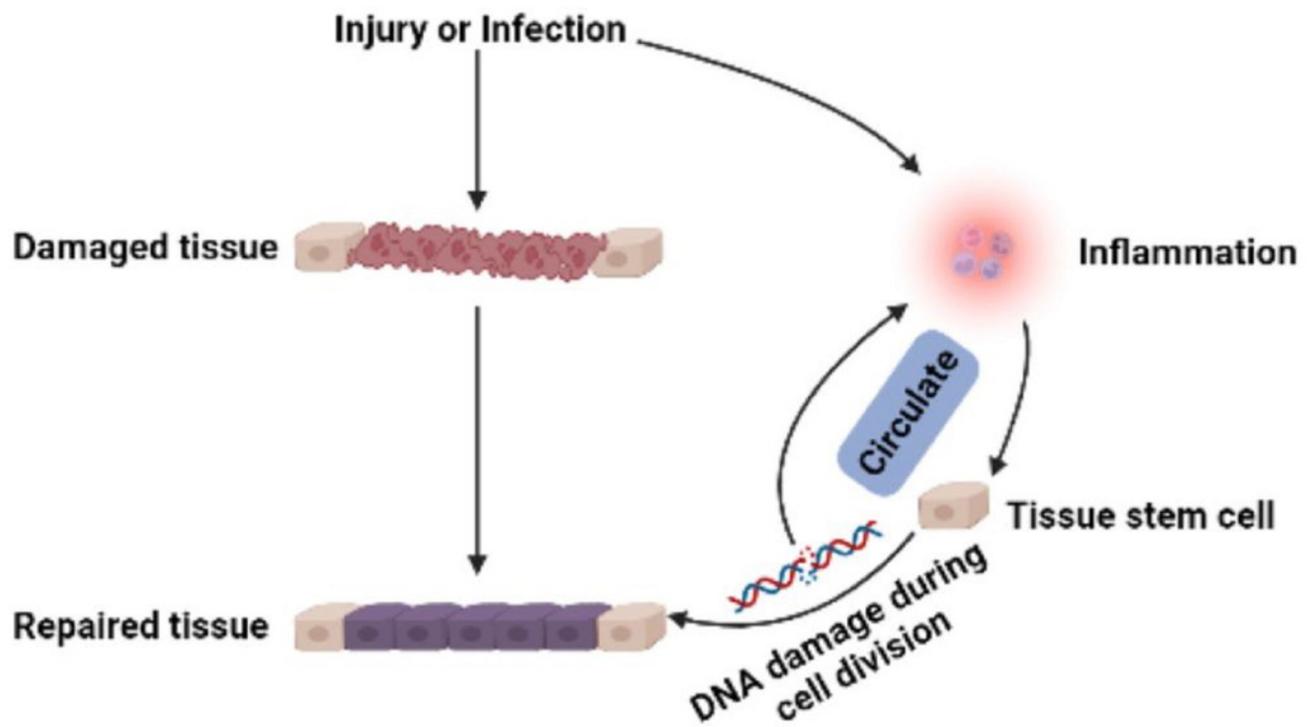


Figure 3

The role of inflammatory response in the stem cell division theory of cancer.

Tissue stem cells are quiescent in the niche. (A) The organism is infected with a pathogen or has endogenous tissue damage. (B) Tissue stem cells produce PAMPs, DAMPs, and ROS upon stimulation by pathogens or endogenous danger signals. (C) PAMPs and DAMPs activate pattern recognition receptors (PRR). The PRR signaling pathway is the promoter of a cascade of reactions that ultimately lead to the migration of immune cells to infection site. The innate immune response is subsequently activated, while the adaptive immune response can be activated directly or indirectly. (D) Recruited immune cells alter the tissue stem cell niche, causing inflammatory cells and factors to become its major components. (E) Inflammatory cells or factors act directly or indirectly on tissue stem cells to activate their regenerative capacity. (F) In response to repeated inflammatory stimulation, tissue stem cells undergo long-term division, resulting in the accumulation of genomic mutations, and eventually mutate into cancer cells. (G) Inflammation also reprograms the fate of cancer cells, or cancer stem cells, and promotes the spread of cancer.

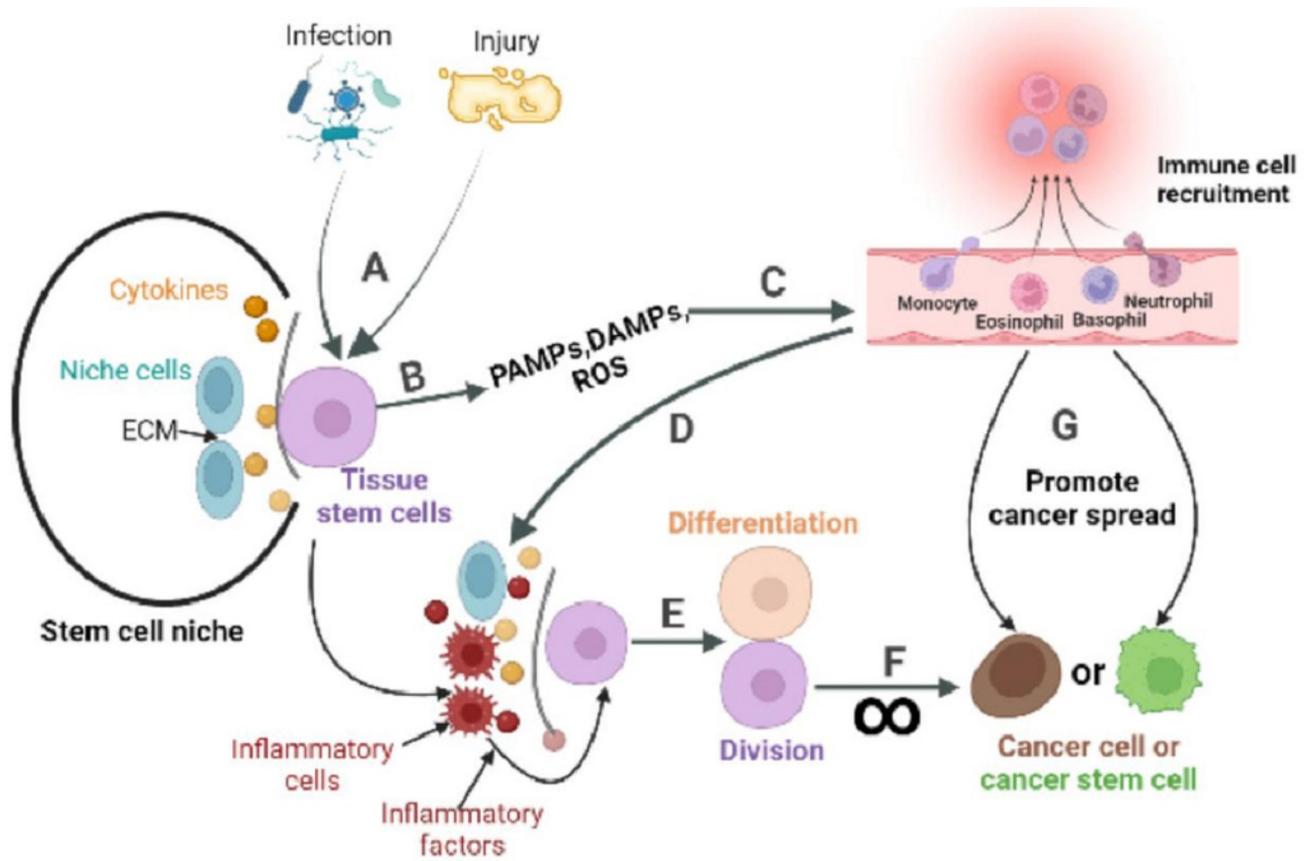


Table 1 (on next page)

Inflammatory soluble factors or immune cells and their effect on stem cell function

Soluble factors or immune cell	Cellular origin	Stem cell type	Effects on stem cell function	References
IL-1	Macrophages	Bone marrow derived MSC	Prime MSCs towards an anti-inflammatory and pro-trophic phenotype in vitro	(84)
IL-1 α /IL-13/TNF- α /IFN- γ	T cell/ macrophages	Muscle stem cells (MuSCs)	Serially expand MuSCs in vitro for over 20 passages	(85)
IL-22	T cell	Intestinal stem cell (ISC)	Augment the growth of mouse and human intestinal organoids, increase ISC proliferation and expansion.	(78)
PGE2	Inflammatory cells	ISC	Promote ISC self-renewal	(91)
PGE2	Inflammatory cells	MuSC	Promote MuSC proliferation	(87)
Tregs	T cell	Hair follicle stem cells (HFSCs)	Augment HFSCs proliferation and differentiation	(93)
IFN- γ / IL-13/ IL-17	T cell	ISC	Promote ISC differentiation	(92)
IL-10	T cell	ISC	Promote ISC self-renewal	(92)
IL-1/TNF- α	Tcell/ macrophages	Gingival mesenchymal stem/progenitor cells (G-MSCs)	Stimulate G-MSCs regeneration and osteogenesis	(86)

Table 2 (on next page)

Inflammatory soluble factors and their related signals promote malignant phenotypes of cancer stem cells

Soluble factors and Inflammation related signal transduction pathways	Cancer stem cell type	Effects on cancer stem cell function	References
IL-6/HIF1	breast cancer stem cell	stemness markers, sphere formation, self-renewal, metastases	(106)
SRSF1/circATP5B/miR-185-5p/HOXB5-IL-6/JAK2/STAT3	glioma stem cells	proliferation	(107)
TNF- α /NF- κ B	Glioma stem cells	cell viabilities, proliferation, invasion, neurospheres formation abilities, stemness markers	(110)
PGE2/NF- κ B	Colorectal Cancer Stem Cell	Expansion and Metastasis	(111)
CCL16/CCR2/GSK3 β / β -catenin/OCT4	breast cancer stem cell	Stemness markers	(112)
IL-17/NF- κ B/p38MAPK	ovarian cancer stem cell	self-renewal	(113)