- 1 Morpho-biochemical characterization and molecular marker marker based genetic diversity
- 2 study in pearl millet [Pennisetum glaucum (L.) R. Br.]
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14 Abstract

- 15 Pearl millet is a key food for millions living the semi-arid and arid regions. The poor's diet
- 16 contains more pearl millet than other grains. The genetic diversity existing in the pearl millet
- 17 germplasm can be used to improve the micronutrient content and grain yield. Effective and
- 18 organized exploitation of diversity at morphological and DNA levels is the strategic strategy to
- 19 for any crop improvement program. In this study, the genetic diversity of 48 pearl millet
- 20 genotypes wasere evaluated for eight morphological traits and eleven biochemical characters.
- 21 and genetic diversity was performed. Genotypes were characterized using twelve SSR and six
- 22 SRAP markers. Significant The significant mean difference among between morphological and
- 23 biochemical-was traits wereas detected in ANOVA. The productive tillers per plant varied from
- 2.65 to 7.60 with a mean of 4.80. The-G grain yield of genotypes varied more than 3x from 15.85
- 25 g (ICMR 07222) to 56.75 g (Nandi 75) with an average of 29.54 g per plant. During the
- experiment, higher protein, iron, and zinc content was wereas recorded in ICMR 12555 (20.6
- onpermitting in protein, nong une mite general mas meteral recorded in Territ 12000 (2010)
- 27 %), –ICMR 08666 (77.38 ppm), and IC 139900 (55.48 ppm), respectively. Substantial
- 29 256.00 ppm (ICMR 12888). The top eight nutrient-dense genotypes flowered in 34–74 days and

variability was observed for grain calcium as it ranged from 100.00 ppm (ICMR 10222) to

- 250.00 ppm (refine 12000). The top eight number dense genotypes nowered in 54 74 days and
- had 5.71–9.39 g 1000 grain weight. Genotype ICMR 08666 was dense superior for Fe, Zn, K,
- and P. The inter-genotype similarity coefficient at the genetic level, generated using DNA

Açıklamalı [Ref1]: Instead of "study in" the word "of" would be better

Açıklamalı [**Ref2**]: Sentence is too long, can be divided in to two sentences.

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32 markers, ranged from 0.616 to 0.877 with a mean of 0.743. A_C_combination of phenotypic

33 morpho-biochemical traits- and genotypic DNA markers-based diversity carried out in this

34 research will help to differentiate the genotypes and diverse genotypes can be used in the

breeding programs to improve the mineral content in pearl millet. under study in a better way.

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Key-words: Genetic Deliversity, Germplasm, Grain minerals, Pearl millet, Variability, Yield

38 1. Introduction

- 39 Pearl millet [Pennisetum glaucum(L.) R. Br.] is a small-seeded C4 plant of the Poaceae family.
- 40 The 1.7 GB genome of this crop is accommodated by 2n=2x=14 chromosomes. Compared to
- other cereals like wheat and rice, it can withstand effectively-under in drought, nutrient-depleted
- 42 soil, and high temperature hot environmental conditions of the hot hostile deserts of Indian and
- 43 African. This hardy nature makes pearl millet a resilient to harsher climatic conditions
- consequently it is cultivated in marginal environments of arid and semi-arid tropical regions of
- 45 India and sSouth-east Asia, sub-Saharan Africa, and much of southern and eastern Africa
- 46 (Ramya et al., 2018). Pearl millet is a key food for millions living the semi-arid and arid regions.
- 47 The grain is mainly consumed as a-human food while biological yield is used as livestock feed.
- 48 The pPearl millet is a primary food for mankind living in dryland agriculture regions.
- 49 Pearl millet accounts for circa 50% of the total global millet production. It is grown on >28
 - million ha, mainly in Africa and India. India is the world's largest producer of millets, harvesting
- 51 11 million tonnes annually, or around 36% of global production. In 2020, India harvested 8.61
- 52 million tonnes of pearl millet grains from <u>a 6.93</u> million ha area with 1,243 kg/ha of productivity
- (Directorate of Millets Development, 2020) (AICPMIP, 2020).
- 54 In any breeding strategy, variation continues to be the key to success. Pearl millet shows
- 55 abundant phenotypic variability for most of the quantitative traits like flowering time, ear head
- 56 length, grain characteristics, tolerance to various (a)biotic stresses as well as nutritional quality
- 57 (Bhattacharjee et al., 2007). Effective and logical utilization of this diversity is the vital to any
- breeding program (Allard et al.,1960). Exploiting this genetic diversity in the pearl millet
- 59 population will-may allow the improvement of micronutrients density in grain and grain yield.

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Micronutrient insufficiency has emerged as a global problem, particularly for those living in underdeveloped nations and consuming carbohydrate—rich cereal—based diets. This deficiency can be managed with pearl millet, a nutritious cereal. Among all coarse cereals, pearl millet grains are dense in minerals like iron (Fe) and zinc (Zn) concentration and essential amino acids. In comparison to wheat (11.8 g/100 g), the protein in this coarse cereal ranges between 9 - 21%, which is higher than sorghum (10.4%), rice (6.8%), and maize (4.7%) (Kaur et al., 2014). The grains of pearl millet areare gluten-free and have a low glycemic index due to their grains have high fibreer content—low glycemic—index. The provitamin-A enriched grains are also a_richer source of fat (5-7 g/100 g) but are scarce in vitamins B and C (Gopalan et al., 2003). Pearl millet grain is encased in a tough fibrous seed that contains a variable amount of inhibitory factors like phytic acid and polyphenols (Arora— et al., 2003). But these factors can be reduced through various approaches like soaking, fermentation, blanching, and roasting (Kaushik and Grewal, 2017) up to a certain extented only. Moreover, the presence of anti-nutrients factors like saponins, tannins, and phytic acid which can reduce nutrient utilization or food uptake hinders the biofortification in millet.

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To improve the nutritional quality and diminish the anti-nutritional factors of pearl millet through any breeding approach, the knowledge about the variability for mineral content, antinutritional factors, and their relation with yield is a prerequisite. Accumulation of both microand anti-nutrients in seeds is a complex mechanism containing numerous genes and is affected by the environment (Anuradha et al., 2017). Due to the confounding effect of the environment, similar genotypes would have different phenotypes due to environmental variabilitysimilar phenotypes may have different genomic constituents. Therefore, it is hard to equate morphobiochemical and genetic variability. In contrast, molecular markers reflect the authentic genetic variability and relationships among accessions than phenotypic markers (Glaszmannet al. 2010). In pearl millet, microsatellite, SNP and RFLP markers have been applied to create linkage maps followed by quantitative trait loci mapping and germplasm characterization (Kumar et al., 2016). The density and genome coverage can be improved by the combination of various markers. In pearl millet, no report is available on the deployment of sequence-related amplified polymorphism (SRAP) markers for genetic diversity assessment though SRAP has been used for linkage map development by Pedraza-Garcia et al. (2010). Therefore, in the current experiment, both SRAP and microsatellite markers have been used to expose the genetic diversity. SRAP

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markers are dominant markers that target genomic coding sequences and have been employed for genetic diversity assessment (Li and Quiros, 2001). With the purpose to contribute to upsurging the nutritional quality and the food safety of pearl millet as well as widen understanding in this area, the current study was designed to analyze the natural variability for grain mineral and anti-nutrients content in grain along with molecular diversity in pearl millet germplasm.

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2. Materials and methods

With two replications, tThe field trial was done in a_randomized complete block design (RCBD) with two replications. The inter- and intra-row distance was 60 and 15 cm, respectively. The recommendations <u>for_crop</u> management practices were followed for uniform plant growth <u>and</u> the <u>a</u> healthy crop stand. The seeds were sown on February 2021. A total of 48 pearl millet genotypes were used for the study (Supp Table 1).

2.1. Morphological characters

At the panicle emergence stage, The panicles were covered with glassine bags to prevent cross-pollination by outside pollen and to collect self-seeds. For phenotyping of grain—based traits, physiologically mature panicles were collected, dried under sunlight, and then manually threshed in bulk.

The experimental material was evaluated for eight morphological traits viz., days to 50% flowering, plant height, panicle diameter (PD, mm), panicle length (PL, cm), number of productive tillers, grain yield, days to maturity, and 1000 grain weight. Except for days to 50% flowering and days to maturity, which were recorded on a plot basis during the study, data on the above traits were collected from randomly tagged five competitive plants in each genotype in both replications. PL-PD was measured with Vernier calipers: on panicles for which PL was recorded.

2.2. Biochemical characters

Before biochemical analysis, grains were cleaned followed by hot air oven drying (80°C for 24 h). Dried grains (10 g) were powdered manually. For mineral analysis, 0.5 g flour was processed in 11 mL of nitric acid (69%) and 1 mL of H₂O₂. The digestions were carried out in HVT50 vessels using rotor 12HVT50 in Multiwave GO/ Multiwave GO plus (Anton Paar GmbH, Austria). For microwave digestion, the initial temperature was kept at 180°C with a ramp of 20 minutes and a hold period of 12 minutes. While the second round of digestion was performed with a ramp time of 10 mins on at 70°C temperature and a holding time of 5 minsutes. Inductively Coupled Plasma Optical Emission Spectrometry (ICP-OES) (model 7000DV of make: Perkin Elmer, made in USA with wintab32 software ver. 5.1) was used to determine the mineral content (Fe, Zn, Ca, Cu, and Mn) in grain after diluting the digested mixture to a volume of 50 ml using distilled water. The flow rate in a peristaltic peristaitic pump was 1.5ml / min. From the acid extract, potassium content was quantified using a flame photometer (Jackson, 1973), while the vanadate-molybdate method of Jackson (1973) was used to estimate the phosphorus. Total phenols was were estimated using the Folin-Ciocalteau reagent as Malik and Singh (1980) and reading was measured at 730 nm using a spectrophotometer. Soxhlet extraction was performed to estimate the crude oil content, and semi_micro-Kjeldahl was employed to determine the crude protein content.

135 2.3. Molecular marker study

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- Genomic DNA was extracted from tender leaves as per Mace et al (2003). Genotyping was done
- 137 using simple sequence repeat (SSR) and sequence-related amplified polymorphism (SRAP)
- markers. For microsatellite marker profiling, markers from the PMES series (Zala et al., 2017)
- were amplified in SensoQuest Thermocycler (Germany). The SSR-PCR reaction conditions were
- as follows: 94°C (initial denaturation) for 5 mins., followed by 35 cycles of 94°C for 45 sec,
- 141 X°C (primer specific) for 45 sec, 72°C for 45sec, and 72°C for 7 mins (final extension).
- 142 The SRAP amplification were was as follows: 94 °C (initial denaturation) for 5 min., followed
- by 5 cycles of 94 °C for 30 s, 35 °C for 45 s, and 72 °C for 90 s, followed by 35 cycles of 94 °C
- 144 for 30 s, X°C (primer specific) for 45 s, 72 °C for 60 s and 72 °C for 10 min (final
- extension).— An-agarose gel (3%) was used to resolve PCR products.

2.4. Statistical analysis

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The mean value of traits was figured out, and analysis of variance (ANOVA) was performed in accordance with Panse and Sukhatme (1978) in Microsoft Excel 2013. A P phenotypic trait-based dissimilarity matrix was constructed using Manhattan coefficients with Numerical Taxonomy and Multivariate Analysis System (NTSYSpc 2.0;—(Rohlf 1998). The amplified products of SSR and SRAP markers were scored in 1 (presence) and 0 (absence) fashion. Polymorphism information content (PIC), Multiplex ratio (MR), effective multiplex ratio (EMR) marker index (MI), and resolving power (Rp) value were estimated following Sharma et al. (2016) in Microsoft Excel 2013. In NTSYSpc 2.0, The SIMQUAL program used the Jaccard's similarity (J) coefficient to compute the genetic similarity between genotypes, SAHN clustering method was used to construct the unweighted pair group method with arithmetic mean (UPGMA) dendrogram (Sneath and Sokal, 1973).

3. Results and discussion

3.1. ANOVA

Genetic variability, a quantitative measure of genetic dissimilarities, is the total of genetic diversity within a population. The genetic differences among genotypes are the result of natural selection over a period of time. Greater variability present in the base breeding germplasm ensures good opportunities to produce desired genetic architecture of a plant. The ANOVA resulted that genotypic variations were very significant at a 5% level of probability for all the traits, showing an ample genetic diversity among the genotypes under study. (Table 1). This also suggested that there is sufficient scope to select superior breeding material which can be exploited in pearl millet breeding programs.

168 <u>3.2.3.1.</u> Character variance analysis

169 <u>3.2.1.3.1.1.</u> Morphological parameters

170 Early flowering is a desirable trait for pearl millet as it is a crop of (semi)-arid and arid regions.

Earliness becomes an important trait in areas where scanty and erratic rains aggravate the

moisture stress condition during the growth stage of the crop and leads to-a post-flowering

moisture stress (Yadav et al., 2011). In the-current study, though, the population mean for days

to 50% flowering was 54.69 days but the days to 50% flowering ranged from 34 (IC 370523) to

175 77 days (ICMR 07999). Earlier literature also recorded similar values for days to 50% flowering

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176 for example 49.06 days by Govindaraj et al. (2011), 53.10 days by Sonali et al. (2019), and 55.61 days by Pallavi et al. (2020). PH is an important trait that governs tradeoffs between competition 177 178 and resource distribution, which is decisive for productivity (He et al., 2021). Semi-dwarf 179 genotypes are better than their tall counterparts because of their reduced lodging vulnerability 180 and better response to nitrogen Azhaguvel et al. (2003). In the present experiment, PH ranged from 110.10 cm (ICMR 06555) to 205.35 cm (IC 332715) with an average of 149.42 cm. The 181 182 results indicated that most of the studied genotypes are semi-dwarf in nature and with better 183 management dwarfism supports the grain yield. 184 Panicle size (length and diameter) are two important traits which that have direct positive 185 correlations with grain yield in pearl millet (Vengadessan et al., 2013). Hence, the improvement 186 of sink-size relation traits is a key objective for pearl millet improvement programs. PL in the 187 present study ranged from 15.55 cm (ICMR 11888) to 38.05 cm (IC-332716) with an average of 188 24.03 while the diameter ranged from 1.03 cm (ICMR 10999) to 2.15 cm (ICMR 09333) with an 189 average of 1.53 cm. Abubakar et al. (2019) observed a similar range and mean in pearl millet (2.26 cm) for panicle diameter. Similarly, results for PL are comparable with previous reports 190 (Sharma et al., 2018, Rani et al., 2019). The number of productive tillers per plant varied from 191 2.65 (ICMR 08222, ICMR 11999) to 7.60 (IC 370523) with an average of 4.80 (Table 2). 192 193 According to Sile et al. (2004), non-tillering millet genotypes produced bold seeds than the 194 genotype that produced tillers. Similarly, Maman et al. (2004) also reported that, a reduction in productive tillers from 10 to 3 or 5 improved seed yields by 15-30%. Yadav et al (2021) reported 195 196 that private—sector hybrids are generally having less number of effective tillers/plants. But still, farmers in drought-prone areas prefer high tillering hybrids because tillering is a strategy of 197 198 adaptation to intermittent drought spells (Yadav et al., 2016). 199 In cereal breeding, yield, a complex trait, is one of the supreme traits which is influenced by 200 several associated traits. This makes grain yield a complicated trait. The G grain yield of 201 genotypes varied more than 3x from 15.85 g (ICMR 07222) to 56.75 g (Nandi 75) with an 202 average of 29.54 g per plant. Large variability was also observed for 1000 grain weight (TGW) 203 which is determined by the form, size, and density of the grain and these are directly related to 204 total grain yield. TGW ranged from 4.93 g (ICMR 06888) to 10.45 g (ICMR 06555) with an average of 7.14 g. A diversity assessment of 21,594 pearl millet genotypes from 50 nations 205

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206 revealed huge variability for 1000 seed weight the TGW (1.5 to 21.3 g) (Upadhyaya et al., 2007). Three-fold variability for TGW (6-16 g) was earlier recorded by Pujar et al (2018). 207

208 3.2.2.3.1.2. Biochemical parameters

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Compared ing to to other main cereal crops, pearl millet yields more nutritious grains that are a 210 great source of with protein, calcium, phosphorus, iron, and zinc (Devos et al., 2006). Currently, 211 the commercially grown varieties/hybrids of pearl millet produce grains with an average Fe and 212 Zn content of 42 and 32 ppm (parts per million), respectively (Rai et al. 2016). However, a much 213 wider variability for these micro_nutrients has been reported in germplasm collections (Rai et al. 214 2014). Iron is an essential element for blood production and for the growth and development of 215 the body. Zinc is essential for the development of a strong immune system. The values of iron 216 content in the current study ranged from 31.58 (ICMR 07777) to 77.38 (ICMR 08666) with an 217 average of 49.69. Zinc content ranged from 29.34 (ICMR 08111) to 55.48 (IC 139903) with an 218 average of 39.36 (Table 2). A similar mean value was ere—observed by Velu et al. (2007; Fe: 45.50), Anuradha et al. (2017; Fe: 57.65, Zn: 46.61), Anuradha et al. (2018; Fe: 55.73, Zn: 42.75), Sonali et al. (2019; Fe: 53.57, Zn: 40.39) and Yadav et al. (2020; Fe: 50.60, Zn: 38.60).

In the human body, fats and carbohydrates metabolism, absorption of Ca, and the control of blood sugar are all impacted by manganese. It is also essential for standard brain/nerve functioning and bone mineral density. The values of manganese content ranged from 7.20 ppm (ICMR 07222) to 17.63 ppm (ICMR 08444) with an average of 14.04 ppm. The outcome is in congruence with Anuradha et al. (2017), Kumar et al. (2020) and Govindaraj et al. (2020a). Similarly, a low value of Mn (8 ppm) was recorded by Oshodi et al. (1999).

Calcium is very important for the contraction of muscle, the development of strong bones and teeth, blood clotting, the transmission of nerve impulses, and in the regulation g-heart beats in cells. This is claimed that a high intake of cereal grains increases the chances of calcium deficiency. But this is not true with pearl millet as substantial variability was observed for grain calcium as it ranged from 100.00 ppm (ICMR 10222) to 256.00 ppm (ICMR 12888) with an average of 199.31 ppm. Higher variability for Ca (85-249 ppm) was also recorded by Govindaraj et al. (2020b) in pearl millet core collection. In the current study, 50% of the genotypes had high calcium (>200 ppm).

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metalloenzymes playing role in metabolism Fe and cellular respiration. In the current study, the 236 237 grain Cu ranged from 4.92 ppm (IC 332703) to 22.59 ppm (GHB 558) with an average of 9.86 ppm. The range of grain Cu in various studies are is different like 4.14-15.35 ppm in Anuradha 238 et al. (2018), 4-7 ppm in Govindaraj et al. (2020), 3.19-4.76 ppm in Warrier et al (2020). 239 240 The transport of water, nutrients, and carbohydrates within plant cells is linked with potassium. It is a crucial mineral for the activation of several enzymes that control the synthesis of protein, 241 242 starch, and adenosine triphosphate (ATP) in plants. The potassium ranged from 1800 ppm 243 (ICMR 07222) to 6000 ppm (ICMR 10999) with an average of 4700 ppm. Large variability for potassium was also recorded in 122 commercial pearl millet cultivars (3675-5375ppm; 244 245 Govindaraj et al. 2020a) and core collection (3667-5133 ppm; Govindaraj et al. 2020b). 246 The body needs phosphorus to produce protein for the development, upkeep, and repairing of 247 cells and tissues. Additionally, it participates in the production of ATP. The values of 248 phosphorus content ranged from 2200 (IC 139900) to 3600 ppm (ICMR 08666) with an average of 3112 ppm. ICMR 06555 was statistically at par with IC 139900. 249 250 Pearl millet is also a promising source of protein. Studies indicated that protein in pearl millet is 251 circa 11.8 %, which is better than rice (8.6 %), and maize (9.2 %) and comparable with sorghum 252 (10.7 %). Moreover, pearl millet grain is enriched with glutamate which is a precursor of γ -253 aminobutyric acid (GABA) (Tomar et al., 2021). In the current study the protein content in 254 ranged from 8.26% (IC 332716) to 20.06% (ICMR 12555) with an average of 13.73%. ICMR 255 07444 (9.89%) was statistically at par with IC 332716. The study of Pujar et al. (2020) reported 256 grain protein content variation between 6 - 18%, with a mean of 11%. The augmentation of pearl millet in daily food can reduce the risk of protein malnutrition in an economical way. Moreover, 257

protein extracted from pearl millet can be exploited to design protein-enriched functional foods.

The lipid content ranged from 2.72% (ICMR 06999) to 6.95% (ICMR 08444) with an average of

4.68%. A comparable range and mean were observed by Arulselvi et al. (2007; 5.12%), Abdalla

et al. (2007; 2.70-7.10%) and Tomar et al. (2021; 5.24-9.99). The lipid content in pearl millet

was 1.5 times higher sorghum and corn. Though, the high lipids have been documented as

possible causes for the rancidity of millet flour. However, the shelf life of flour can be increased

Copper is essential for the synthesis of elastin and collagen. It is a key cofactor of many

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by hydrothermal treatment, irradiation, cooling storage, or a combination of more than one technology. (Goyal and Chugh, 2017)

The metal—chelating ability of phytic acid makes it is—an antinutritional phytochemical as it declines the bio-availability of ions like Mn, Ca, Mg, Fe, and Zn (Marathe et al., 2018). In the current study, the phytate ranged from 201.5 mg/100g (ICMR 08111) to 542.50 mg/100g (GHB 558) with an average of 282.39. Abdalla et al. (2007) also recorded—A a similar range from 354-795 mg/100g of phytate was detected by Abdalla et al. (2007where; 354-795). The study of by Gabaza et al. (2018) reported that phytate in pearl millet grains ranges between 580 mg/100g to 1380 mg/100g which is similar to sorghum and maize. The range of phytate in the current study is supported with—by the result of Pushparaj and Urooj (2014) in Indian cultivars where it was between 0.26 - 0.99 g/100 g. The result suggested that phytic acid content in pearl millet grain is significantly lower than in rice (0.68-1.03%; Liu, 2005), oat (0.5–1.2%; Peterson, 2001), soybean soybean (1.0–2.22%; Lolas et al. 1976) and wheat (0.2 - 2.9%; Gupta et al. 2015). Hence, regular

Polyphenols have many health benefits as having antioxidant activity. Moreover, phytic acid is considered to be beneficial in dropping cholesterol and reduces reduceing cancer risk. The values of total phenolic acid ranged with from 75.16mg/100g (ICMR 12555) to 44.41mg/100g (Nandi 75) with an average of 60.26mg/100g. Higher phenol content in grain makes pearl millet a good food to maintain the redox potential of cells and to quench the ROS species. Phenolic may be particularly important in the treatment of postprandial hyperglycemia since it has been documented that it reduces intestinal—glucosidase and pancreatic—amylase (Shobana et al. 2009).

consumption will possibly not hamper the bioavailability of minerals.

285 3.3.3.2. Nutrient-dense genotypes

Genotypes dense in multiple nutrients can directly be released as <u>a variety</u> after evaluating on its their yield performance over the locations for multiple years. Such genotypes can be exploited in <u>a hybridization</u> program. In the current study, the top eight nutrient-dense genotypes flowered in 34–74 days and had 5.71–9.39 g TGW (Table 3). Top genotypes had Fe content of 61.07-77.38 (ICMR 08666) ppm, Zn content of 45.11-55.48 (IC 139900) ppm, Mn content of 16.2-17.63 (ICMR 08444) ppm, Ca content of 230.5-256 (ICMR 12888) ppm, Cu content of 14.32-22.59 (GHB 558) ppm, Kcontent of52.3-60.15(ICMR 10999) ppm, and P content of 33.28-36.72 (ICMR 08666). IC 139900 was superior for both Fe (71.22 ppm) and Zn (55.48 ppm). Genotype

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ICMR 08666 was dense for Fe, Zn, K_a and P. Out of eight high-Fe genotypes, only two genotypes had > 75 ppm. Thus current experiment also identified the best genotypes that had a higher content of multiple nutrients. Earlier, Govindraj et al (2020) also identified genotypes having a high content of multiple nutrients.

298 3.4.3.3. Phenotypic diversity analysis

Phenotypic diversity is important for pearl millet breeding. The interactions between the genome and all of its growing micro- and mega-environments lead to the phenotype of the plant (Fasoula et al., 2020). The mean value of each trait was used to generate the Manhattan dissimilarity coefficient and dendrogram (Sokal and Michener, 1958) using NTSYS-pc 2.02 program me (Rohlf 1998). The genotypes were divided into nine major clusters based on the Manhattan dissimilarity coefficient. The average dissimilarity value among genotypes was calculated to be 0.16, demonstrating modest phenotypic variability (Table 4). The dissimilarity between genotypes ranged from 0.08 (IC 139899 and ICMR 07888) to 0.27 (Nandi 75 and ICMR 07222) showing the maximum and minimum similarity for the respective pair of genotypes.

Cluster I comprise seven genotypes, characterized by high values of DFF, days to maturity, lipid, potassium and Low values of NPT. Cluster II consists of twenty five 25 genotypes. Cluster III contains four genotypes, namely ICMR 10222, Nandi 75, ICMR 12111—and ICMR 10888. This cluster is characterized by more GY, days to maturity—and 1000 grain weight TGW. Cluster IV has four genotypes, namely ICMR 08999, ICMR 11888, ICMR 139900 and ICMR 07777. This cluster is characterized by more lipid content, manganese content, and days to maturity. Cluster V has three genotypes (IC 332716, GHB 558, ICMR 07444) which is are characterized by low values of head diameter—and lipid content. Cluster VI consists of two genotypes (ICMR 08111, and ICMR 12333). This cluster is characterized by more number of productive tillers per plant with low content of zinc. Cluster VII has only one genotype (ICMR 12555) which has high values for characters like days to maturity, protein content, manganese content and 1000 grain weight TGW (Table 4).

320 <u>3.5.3.4.</u> Molecular <u>marker--based diversity</u>study

During recent days molecular markers have been commonly using for assessing genetic diversity. The combined use of two or more markers (both dominant and codominant) for genetic

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Açıklamalı [Ref23]: Material and methods not here.

Açıklamalı [Ref24]: Not capitalized.

overcome the limitation of morphological, biochemical and protein based markers which are 324 325 affected by genotype x environmental interaction. The calculation of genetic diversity helps in the characterization of germplasm apart from crop improvement. 326 327 Forty-eight genotypes of pearl millet were analysed using SSRs and SRAP markers 328 (Table 5). During the experiment, a total of 50 SSR markers were screened for amplification. Out 329 of 50 SSR markers, 32 (60%) primers showed amplification. Of these 30 SSR markers, 12 330 (37.5%) markers were found, of 30 SSR primers certified polymorphic. These 12 polymorphic SSRs markers generated 65 allelesamplicons. The molecular weight of the amplicon with band 331 332 sizes ranging ranged from 85bp (PMES 190) to 292bp (PMES 171). In previous reports with PMES-series SSR markers, Earlier, Zala et al. (2017) recorded amplicon size from 101 to 285 bp 333 334 was reported by Zala et al. (2017). The number of polymorphic bands/amplicons per SSR marker ranged from 2 to 13, with a mean of 6.71. Zala et al. (2017) reported 2.20 alleles. All SSR 335 336 amplicons were found polymorphic—The percentage polymorphism is 100%. The PIC value, the 337 informativeness of a primer, for each marker was computed for the estimation of marker allelic 338 variation based on considering the allele frequencies in studied genotypes. PIC designates the 339 informativeness of a primer. The mean PIC of SSR markers was 0.28 on average though it and varied from 0.132 (PMES 171) to 0.499 (PMES 173). This range was comparable with Zala et 340 al. (2017) where PIC was between ; 0.188 to 0.375). 341 The Rp was estimated considering the proportion of genotypes containing the amplicon. The 342 343 primer that might best differentiate the cultivar can easily be identified by the value of the Rp 344 and PIC. In the current study, Rp varied from 1.625 (PMES 162) to 2.375 (PMES 168), with an average of 1.98. Mean Rp was between 0.154 (PMES 171) to 1.000 (PMES 173). The PI ranged 345 346 from 0.998 (PMES 173) to 1.963 (PMES 168), though the mean PI value was 1.420. MI is considered to be an inclusive measure of the efficiency to detect polymorphism. The SSR MI 347 was 19.092. 348 349 In the case of the SRAP markers, of 25 SRAP, 6 (24%) were polymorphic. The polymorphic SRAPs amplified 119 amplicons. The product size for SRAPs ranged from 94 (Em2+Me2) to 350 351 1357 bp (Em6+Me3). The polymorphic bands ranged from 10 (Em6+Me3) to 34 (Em2+Me2),

with a mean of 21.67. Liu et al. (2008) observed a polymorphic band detected with each ranging

diversity study has been better than the individual marker system. Molecular markers help

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- 353 from 6 to 17, with an average of 11.76. Bhatt et al. (2017) had a band size from 120 to 500 bp in
- 354 cumin. This suggested that in different crops SRAP amplicon size will be highly variable. PIC
- oscillated from 0.224 (Em5+Me4) 0.324 (Em6+Me2), with an average of 0.26. Bhatt et al.
- 356 (2017) reported a PIC value (0.34).
- The mean PI of SRAP markers was 5.69, through the maximum PI was for Em6+Me2 (8.108)
- 358 and the lowest value from for Em6+Me3(2.335). Rp ranged from 4.625 (Em5+Me4) to 13.250
- 359 (Em6+Me2), with an average value of 8.27. Mean Rp was between 0.272 (Em5+Me4) 0.542
- 360 (Em6+Me3) with an average of 0.40. Liu et al. (2008) stated higher RP values ranged from 2.229
- 361 to 8.457 with an average of 4.927. Fraction The fraction of polymorphism, MR, EMR and MI for
- 362 SRAPs are 1.00, 10.00, 10.00, and 15.65, respectively (Table 6).
- 363 3.6.3.5. *Inter-Genotype Genetic Relationship*
- 364 Forty-eight pearl millet genotypes were divided into seven major groups by the dendrogram
- 365 created using pooled data from SSR and SRAP markers based on Jaccard's similarity matrix.
- 366 Cluster I, II, III, IV, V, VI and VII had with 1, 1, 11, 8, 3, 22, and 2 genotypes, respectively
- 367 (Figure 1). In the study of Nehra et al. (2017), with SSR markers, 49 accessions were clustered
- 368 into eight core clusters. Kumar et al. (2020) alienated 18 lines into three clusters in pearl millet
- 369 using 74 SSRs. In the current study, the inter-accession genetic coefficient of similarity ranged
- 370 from 0.616 to 0.877 while the average similarity was 0.743. ICMR 098888 and GHB 905 has a
- genetic distance (0.384) indicative that both genotypes are having moderate-genomic difference
- 372 level and can be crossed to create a bi-parental mapping population. The minimum genetic
- distance (0.123) was between IC 139899 and IC 332727, demonstrating that these accessions
- have more similarity in microsatellite locus. Moreover, on the based of on diversity results,
- breeders can select diverse genotypes for combining ability and heterosis analysis for traits
- 376 studied in the current study.

4. Conclusion

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- 378 The genetic diversity for morphological and grain biochemical traits, an outcome of natural
 - selection with the cross-pollination nature of pearl millet, was revealed by analysis of variance.
- 380 Variability for grain micronutrient content was found greater with a wide range in the
- population. Genotypes namely ICMR 08666 and IC 139903 were superior for iron and zinc

Açıklamalı [Ref25]: Genomic or genetic?

383	and can further be utilized for genetic biofortification. In the present study, phenotypic diversity
384	analysis grouped all genotypes into nine different clusters. Among all clusters, three clusters
385	were share had only single genotype with better phenotypic value for most of the grain
386	biochemical parameters. But phenotype is a total outcome of the genotype and its interaction
387	with the environment. Genetic markers are found effective in this study, they help to identify
388	ICMR 098888 and GHB 905 as diverse genotypes for making a bi-parental mapping population.
389	5. Conflict of Interest
390	All authors declared that the research was conducted in the absence of any commercial or
391	financial relationship that could be construed as a potential conflict of interest.
392	6. Authors' Contribution
393	SK, MPP, AAS & VK: Conceptualization & Designing the experiment, DFG, RP & DK:
394	performed the experiments, RP & SK: analyzing the data, VK: Contribution of experimental
395	material, DFG, SK & RP: Prepared the figures and tables, SK & AAS: Critically revised the
396	manuscript for important content,
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l 403	References
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content, respectively. Genotype ICMR 08666 was also found promising for Zn, K and P content

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Table 1. Analysis of variance (ANOVA) of morpho-biochemical traits in pearl millet

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	Source of variation and Mean squares						
Trait	Replication (df = 1) Genotypes (df = 47)		Error $(df = 47)$				
Days to 50% flowering	0.167	322.311*	5.954				
Plant height	66.833	917.408*	62.211				
Head diameter	0.027	0.159*	0.027				
Panicle length	1.927	46.955*	3.880				
Productive tillers per plant	0.027	3.712*	0.217				
Grain yield per plant	11.003	203.764*	30.441				
Days to maturity	2.344	9.292*	3.471				
1000 grain weight	0.000176	2.863*	0.104				
Protein content	2.004	10.363*	0.705				
Lipid content	0.062	3.851*	0.052				
Iron content	14.015	276.360*	16.714				
Zinc content	41.12	79.610*	13.100				
Manganese content	2.154	11.232*	1.576				
Calcium content	661.5	2061.205*	332.691				
Copper content	0.218	43.735*	0.551				
Potassium content	57.722	102.837*	14.517				
Phosphorus content	5.782	12.004*	5.093				
Phytate content	895.482	8396.470*	541.246				
Total phenolic acid	26.471	112.792*	36.582				

^{*}Significant at 5% level of probability

Açıklamalı [Ref26]: Phenological trait

Açıklamalı [Ref27]: Yield trait

Açıklamalı [Ref28]: Yield component

Table 2. Mean performance of morpho-biochemical traits in pearl millet

Trait	Mean	Range	S. Em	CD @ 5%	CV%	Açıklamalı [Ref29]: What is S.Em?
Days to 50% flowering	54.69	34.00 (IC-370523) - 77.00 (ICMR 07999)	1.73	4.91	4.46	Açıklamalı [Ref30]: All abbreviations used in a given table
Plant height (cm)	149.42	110.10 (ICMR 06555) - 205.35 (IC-332715)	5.58	15.87	5.28	should be explained in full below the table.
Head Diameter (cm)	1.53	1.03 (ICMR 10999) - 2.15 (ICMR 09333)	0.12	0.33	10.75	
Panicle length (cm)	24.03	15.55 (ICMR 11888) - 38.05 (IC-332716)	1.39	3.96	8.2	
Productive tillers per plant	4.80	2.65(ICMR 08222, ICMR 11999)-7.60(IC-370523)	0.33	0.94	9.7	
Grain yield per plant	29.54	15.85 (ICMR 07222) - 56.75 (Nandi 75)	3.9	11.1	18.67	
Days to maturity	85.64	80.50 (ICMR 12333) - 89.00(ICMR 08999, ICMR 07777)	5.82	16.57	9.71	
1000 grain weight	7.14	4.93 (ICMR 06888) - 10.45 (ICMR 06555)	0.23	0.65	4.51	
Protein content (%)	13.73	8.26(AICRP-PM-12)-20.06(ICMR 12555)	0.59	1.68	6.12	
Lipid content (%)	4.68	2.72(ICMR 06999)-6.95(ICMR 08444)	0.16	0.46	4.89	
Iron content (ppm)	49.69	31.58 (ICMR 07777)-77.38 (ICMR 08666)	2.89	8.22	8.23	
Zinc content (ppm)	39.36	29.34 (ICMR 08111)-55.48 (IC 139903)	2.65	7.53	9.5	
Manganese content (ppm)	14.04	7.20 (ICMR 07222)-17.63(ICMR 08444)	0.89	2.53	8.94	
Calcium content (ppm)	199.31	100.00 (ICMR 10222)–256.00 (ICMR 12888)	12.9	36.69	9.15	
Copper content (ppm)	9.86	4.92 (AICRP-PM- 6)-22.59(GHB 558)	0.52	1.49	7.52	
Potassium content (ppm)	4798	1800 (ICMR 07222)-6020(ICMR 10999)	2.69	7.67	7.94	
Phosphorus content (ppm)	3112	2258(AICRP-PM-62)-3672(ICMR 08666)	1.6	4.54	7.25	
Phytate content (mg/100g)	282.39	201.5(ICMR 08111)-542.5(GHB 558)	16.45	46.8	8.24	
Total phenolic acid (mg/100g)	60.26	75.16(ICMR 12555)-44.41(Nandi 75)	4.28	12.17	10.04	

Table 3. Top eight genotypes for different nutritional traits with their agronomic performance

Trait	Top 8 genotypes	Range		
		DFF	TGW (g)	
Fe	ICMR 08666, ICMR 11888, IC 332727, IC 139900, ICMR 12555, ICMR 06111, ICMR 08333, ICMR 07888	37.00-72.00	5.91-8.45	
Zn	IC 139903, ICMR 08999, ICMR 06222, ICMR 12999, ICMR 10999, ICMR 08666, IC 332715, IC 332703	42.00-70.50	5.94-9.39	
Mn	ICMR 08444, ICMR 12555, ICMR 09888, ICMR 08333, ICMR 06666, ICMR 12666, ICMR 09222, IC 332703	45.00-74.50	5.91-8.01	
Ca	ICMR 12888, ICMR 12777, ICMR 07444, ICMR 09333, ICMR 11777, IC 332703, ICMR 08999, ICMR 08333	35.50-72.50	5.91-8.36	
Cu	GHB 558, IC 332715, IC 332716, ICMR 06666, ICMR 07222, ICMR 07444, ICMR 07777, ICMR 10999	42.00-73.50	5.71-8.02	
K	GHB 558, GHB 732, GHB 905, ICMR 08666, ICMR 10999, ICMR 12555, ICMR 12666, ICMR 12777	43.50-74.00	6.16-8.99	
P	ICMR 08666, ICMR 06888, ICMR 06111, IC 370523, IC 332727, IC 139899, Nandi 75, ICMR 06666	34.00-73.50	4.93-9.34	

DFF: days to 50% flowering; TGW: 1000-grain weight

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Table 4. Variability for mean values of 19 quantitative traits in nine groups identified by the Manhattan dissimilarity coefficient

Number of genotypes in each cluster										
Trait	7	25	4	4	3	2	1	1	1	
Days to 50% flowering	72.86	50.24	50.50	44.88	57.67	62.00	53.50	56.50	70.50	
Plant height (cm)	126.56	152.87	174.28	139.36	172.68	139.78	125.45	135.95	151.25	
Head Diameter (cm)	1.66	1.55	1.49	1.59	1.41	1.16	1.65	1.45	1.03	
Panicle length (cm)	23.71	23.68	25.99	23.90	30.53	19.23	22.00	26.00	17.95	
Productive tillers per plant	3.67	4.98	4.36	5.29	4.70	6.70	5.90	4.70	3.50	
Grain yield per plant	24.72	29.63	48.99	22.53	27.23	22.33	47.75	15.85	28.35	
Days to maturity	86.50	85.12	86.13	88.38	87.00	81.50	85.00	86.00	84.00	
1000 grain weight	7.83	7.12	7.79	6.50	7.04	6.32	7.27	5.71	6.16	
Protein content (%)	12.50	14.49	12.72	12.61	9.71	13.69	20.06	15.72	15.71	
Lipid content (%)	5.67	4.36	4.72	5.64	3.25	4.13	2.82	6.92	6.64	
Iron content (ppm)	52.03	50.39	37.14	55.66	46.22	43.33	69.65	48.98	46.16	
Zinc content (ppm)	36.59	40.25	35.21	47.05	36.07	30.54	35.13	44.98	48.79	
Manganese content (ppm)	13.63	14.94	13.65	13.30	14.40	9.66	17.28	7.20	10.39	
Calcium content (ppm)	191.57	208.04	144.75	207.25	215.83	158.75	229.00	204.50	218.50	
Copper content (ppm)	9.40	8.02	7.19	13.85	19.78	9.44	8.18	17.29	19.29	
Potassium content (ppm)	45.70	49.33	46.08	49.31	49.00	45.58	52.30	18.00	60.15	
Phosphorus content (ppm)	31.43	31.81	30.68	27.54	31.98	29.23	27.75	31.52	32.07	
Phytate content (mg/100g)	283.74	272.46	327.68	248.10	398.32	206.55	287.75	307.10	250.95	
Total phenolic acid (mg/100g)	58.59	58.50	58.21	66.53	65.50	59.57	75.16	56.96	73.15	

SSR marker system										
Maker name	Band size (bp)	TB	Polymorphism (%)	PΙ	PIC	RP	Mean RP			
PMES153	145-154	4	100	1.14	0.29	2.00	0.50			
PMES157	135-151	4	100	1.46	0.37	1.96	0.49			
PMES160	134-150	4	100	1.36	0.34	2.13	0.53			
PMES161	155-233	3	100	1.23	0.41	2.04	0.68			
PMES162	132-162	7	100	1.35	0.19	1.63	0.23			
PMES168	222-284	10	100	1.96	0.19	2.38	0.24			
PMES170	154-195	9	100	1.43	0.16	2.00	0.22			
PMES171	151-292	13	100	1.71	0.13	2.00	0.15			
PMES173	216-203	8	100	0.99	0.50	2.00	1.00			
PMES185	176-200	8	100	1.55	0.19	1.83	0.23			
PMES190	85-104	6	100	1.53	0.26	1.92	0.32			
PMES199	213-305	4	100	1.27	0.32	1.83	0.46			
Average		6.17		1.42	0.28	1.98	0.42			
SRAP marker	system									
Em6+Me2	205-1234	25	100	8.11	0.32	13.25	0.53			
Em2+Me2	94-1152	34	100	7.96	0.26	10.20	0.30			
Em1+Me2	223-1125	18	100	5.33	0.30	6.88	0.38			
Em5+Me4	120-1020	17	100	3.81	0.22	4.63	0.27			
Em6+Me3	450-1357	10	100	2.33	0.23	5.42	0.54			
Em2+Me3	313-1065	26	100	6.57	0.25	9.25	0.36			
Average		21.67		5.69	0.26	8.27	0.4			

TB: Total Number of Bands; PI: Primer index; PIC: Polymorphic Information Content; RP: Resolving Power.

Table 6. Comparison between SSR and SRAP marker system

Marker system	Total markers	TB	PB	FP	H_{av}	MR	EMR	MI
SSR	12	74.00	74.00	1.00	3.35	5.69	5.69	19.092
SRAP	6	130.00	130.00	1.00	1.57	10.00	10.00	15.655

TB: Total Number of Bands; PB: Number Polymorphic Bands; PI: Primer index; PIC: PolymorphicInformation Content; RP: Resolving Power; FP: Fractionation of Polymorphism; Hav: Average PIC; MR: Multiplex Ratio; EMR: Effective Multiplex Ratio; MI: Marker Index.

Fig 1. Dendrogram based on the genetic coefficient of similarity among pearl millet genotypes