



Regulation of LncRNAs and microRNAs in neuronal development and disease

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ABSTRACT

Non-coding RNAs (ncRNAs) are RNAs that do not encode proteins but play important roles in regulating cellular processes. Multiple studies over the past decade have demonstrated the role of microRNAs (miRNAs) in cancer, in which some miRNAs can act as biomarkers or provide therapy target. Accumulating evidence also points to the importance of long non-coding RNAs (lncRNAs) in regulating miRNA-mRNA networks. An increasing number of ncRNAs have been shown to be involved in the regulation of cellular processes, and dysregulation of ncRNAs often heralds disease. As the population ages, the incidence of neurodegenerative diseases is increasing, placing enormous pressure on global health systems. Given the excellent performance of ncRNAs in early cancer screening and treatment, here we attempted to aggregate and analyze the regulatory functions of ncRNAs in neuronal development and disease. In this review, we summarize current knowledge on ncRNA taxonomy, biogenesis, and function, and discuss current research progress on ncRNAs in relation to neuronal development, differentiation, and neurodegenerative diseases.

Subjects Biochemistry, Molecular Biology, Neuroscience

Keywords NcRNAs, LncRNAs, MiRNAs, Neuronal, Neurodegeneration

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INTRODUCTION

In 2019, about 50 million people had dementia due to neurodegenerative diseases, and this number is predicted to increase to 1.25 million by 2060 (*World Alzheimer Report, 2019*). Non-coding RNAs (ncRNAs) are involved in various biological processes, including cell proliferation, differentiation, apoptosis, metabolism, stem cell self-renewal, survival and cell integrity maintenance, synaptic formation, and DNA damage responses (*Hombach & Kretz, 2016*). Interestingly, ncRNAs are particularly abundant in the central nervous system, and alterations in their expression pattern have been linked to neuronal differentiation and function. They may lead to brain aging and neurodegenerative diseases (*Mehta, Dempsey & Vemuganti, 2020*). Given the lack of effective treatments for neurodegenerative diseases and the burden on global health systems, early detection, and treatment are required.

Only 1.5% of the human genome encodes protein, and the remaining genes are called “dark matter”, which are widely transcribed to generate a massive amount of ncRNAs (*Hombach & Kretz, 2016*). With more and more types of ncRNA being discovered and

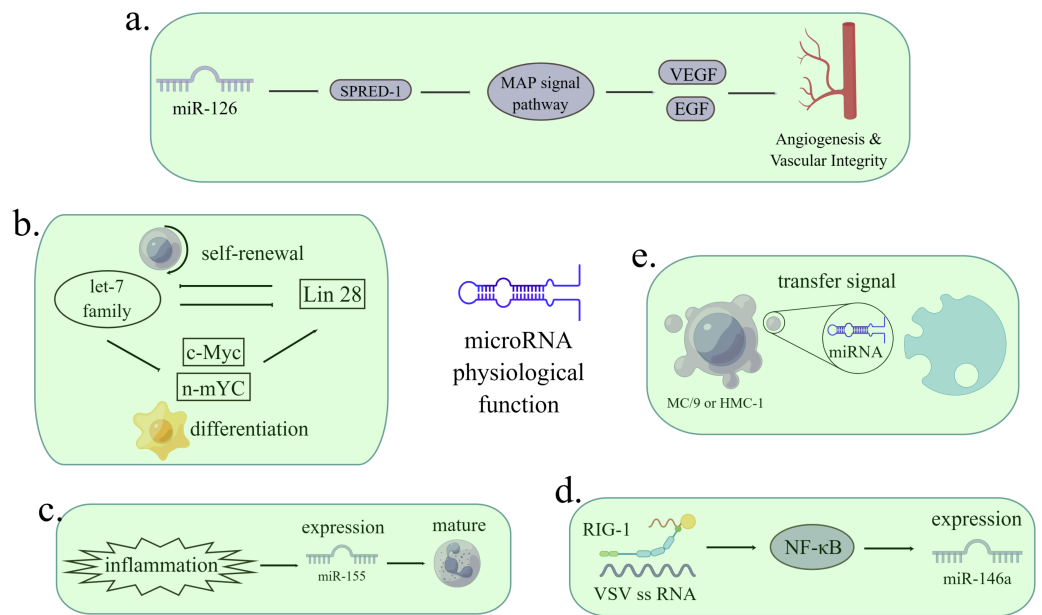


Figure 1 The functions of miRNAs in regulating pathological processes. (A) MiR-126 inhibits the expression of Spred-1, a negative regulator of Ras/MAP kinase signal transduction, thereby enhancing VEGF and FGF and promoting angiogenesis. The loss of miR-126 function reduces the response of MAP kinase to VEGF and FGF, while the function of miR-126 enhances the angiogenesis signal. Spred-1: intracellular inhibitor of angiogenesis signal; VEGF: vascular endothelial growth factor; FGF: fibroblast growth factor. (B) Let-7 family and Lin28 play antagonistic roles in stable self-renewal and differentiation of cells. Let-7 promotes embryonic stem cell differentiation by inhibiting Lin28 and c-Myc. When let-7 is inhibited, cells continue to self-renewal. (C) Under the stimulation of inflammation, the expression of miR-155 is enhanced, thus promoting immature granulocyte numbers in vivo. (D) When infected with a virus, miRNA can either viral target functions and defend against RNA and DNA viruses or can be used by viruses to control cells. After VSV infection, the expression of miR-146a increased in a RIG-1 dependent manner. RIG-1 protein interacts with VSV RNA through its helicase domain, resulting in transcription of pri-miR-1446a in nuclear through NF- κ B, leading to an increase in the number of miR-146a. The reciprocity of miRNAs and viruses can be used as a therapeutic target. VSV: vesicular stomatitis virus. (E) MC/9 and HMC-1 can deliver exosomes containing miRNA to other cells, thus transmitting signals. (Figure made with Figdraw).

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annotated, many ncRNAs have been proved to control the expression of protein-coding genes and are related to cell cycle, proliferation, differentiation, immune response and apoptosis (Friedman *et al.*, 2009). The research on miRNA was relatively earlier and clearer among the various ncRNAs. MicroRNA is widely involved in the physiological and pathological processes of cells. MicroRNA is related to angiogenesis, cell differentiation, inflammatory reaction, virus infection and signal transmission (Valadi *et al.*, 2007; Wang *et al.*, 2008a; O'Connell *et al.*, 2009; Melton, Judson & Blelloch, 2010; Bruscella *et al.*, 2017). Figure 1 MiRNAs regulate physiological and pathological processes such as cancer, gastrointestinal diseases, cardiac diseases, diabetes, and liver diseases (Chen *et al.*, 2019b; Huang, Zhang & Chen, 2022). The targeting oncogenes of tumor suppressor let-7 include MYC, KRAS and HMGA2, but it act as a tumor promoter when limiting immune cells in tumor microenvironment (Balzeau *et al.*, 2017; Pobezinsky & Wells, 2018).

With the discovery of lncRNA, more and more researchers focus on exploiting the functions of lncRNAs and the relationship between miRNAs and lncRNAs (Xu et al., 2020). The functions of lncRNA can be divided into five aspects: (i) Location in genomic imprinting; (ii) chromatin modification; (iii) regulation of cell cycle and apoptosis; (iv) regulation of transcription and mRNA decay; (v) regulation of protein translation. Genomic imprinting is an epigenetic phenomenon in which genes are expressed monoallelically based on parent origin (Bridges, Daulagala & Kourtidis, 2021). It has been found that multiple lncRNAs are at the imprinted genomic loci, and the loss of imprinting causes abnormal gene expression, resulting in disease (Zhu et al., 2013; Cheong et al., 2015). HOTAIR, Kcnq1ot1 and Air can recruit chromatin remodeling complexes to silence genes or regulate epigenetics (Saxena & Carninci, 2011). The accumulation of Gas5 in growth-arrested cells inhibits glucocorticoid response genes and makes cells sensitive to apoptosis (Mourtada-Maarabouni et al., 2009). LncRNA trans-activates STAU1-mediated mRNA decay or targets the sense mRNA transcripts like siRNA (Gong & Maquat, 2011). AS-UCHL1 (ubiquitin carboxy-terminal hydrolase L1) significantly increased the synthesis of UCHL1 protein (Ogawa, Sun & Lee, 2008). LncRNAs were also involved in diseases such as cancer, nervous system disorders, and other diseases (Chen et al., 2017; Chen et al., 2019a).

Because both lncRNA and miRNA are closely related to diseases, more and more research is devoted to developing their potential as biomarkers of diseases. However, experiments often require a lot of time and money, which can be solved through computer models. Although computational models have become an essential method for screening the most promising miRNA-disease pairs, their accuracy and universality still need to be improved (Chen et al., 2018; Chen et al., 2019b). Therefore, computer models can form a reciprocal relationship with experiments. Namely, on the one hand, computer models can guide the most valuable research directions, and on the other hand, experimental results can help optimize computer models (Chen et al., 2019b). The lncRNA disease association (LDA) model is similar to the miRNA disease association (MDA) model. Some LDA models are based on classical models, and some implement random forests and feature selection to reduce the interference of noise and redundant information between these data resources (Yao et al., 2020; Cui et al., 2020; Wang et al., 2021). In addition, computer models can also be used to identify new small molecules targeting miRNAs. At present, there are three methods to predict miRNA-associated small molecules: (i) miRNA structure-based models; (ii) models based on gene expression profiles; (iii) known models based on the association of small miRNAs. More effective prediction models will significantly benefit the screening of compound libraries and the discovery of new miRNA-based small-molecule drug candidates (Chen et al., 2018).

However, the research on ncRNA function and developing computer models related to ncRNA diseases are focused on cancer. Evidence shows that ncRNAs are closely associated with neurons' development, differentiation, and dysfunction. The human central nervous system has roughly equal numbers of neurons and glial cells, and almost all 86 billion neurons are located in the brain (Silbereis et al., 2016). The connectome, one of the most critical components of neural networks and circuits, consists of various neuronal cells and their specific synaptic connections (Van den Heuvel, Bullmore & Sporns, 2016). The

human brain weighs 2.5% of the body but still consumes 18% of its oxygen at rest. Humans evolved to accommodate high levels of neuronal activity, including changes in diet and energy allocation, due to the high metabolic cost of the connectome (*Khaitovich et al., 2008*). Some miRNAs contribute to the development of neurons and maintain the survival of mature neurons (*Yoo et al., 2009*). During neuronal differentiation, miR-124 reduces the level of PTBP1, to increase the expression of correctly spliced PTBP2 (*Makeyev et al., 2007*). When cells differentiate into neurons, miR-124 eliminate the biological effects of REST by inhibiting SCP1 (*Shi & Jin, 2009*). LncRNA is involved in many nervous system processes, including neuronal identity establishment and maintenance, stress response deployment, plasticity, and brain development (*Qureshi & Mehler, 2013*). For example, lncRNA Sox2OT is dynamically regulated in the CNS (*Amaral et al., 2009*). REST suppresses the expression of the nervous system-specific transcriptional gene human accelerated region 1 (HAR1), and HAR1 expression changes may be linked to Huntington's disease phenotype (*Johnson et al., 2010a*). The external environment and internal genetic risk factors can lead to neuronal damage and further neuronal degeneration, and when this damage accumulates beyond an individual's "balanced load," neurodegenerative diseases result (*Armstrong, 2020*).

Given the abundant functions of lncRNA and miRNA related to neuronal development and disease, few similar reviews on this topic have simultaneously discussed the relationship between lncRNA and miRNA and neuronal development, differentiation, and disease. We summarized the significance of miRNA and lncRNA in neurodevelopment and disease and their potential role in the future. We want to draw more attention to the potential roles of lncRNA and miRNA in neurons. We believe lncRNA and miRNA can be used as biomarkers and therapeutic targets for some neurological diseases. At the same time, the network basis of lncRNA-miRNA-mRNA can further expand the relevant research direction. However, because there are few research results on ncRNA as a therapeutic target for neurodegenerative diseases or we have yet to be able to retrieve relevant literature, the content of this part needs to be improved, which is the limitation of this review.

SURVEY METHODOLOGY

After identifying the topic, we searched NCBI for multiple review articles with the keyword "neurons," most of which came from authoritative journals in neurology. In the process of comprehensively reading these review articles, we focused on documenting and listing the outline framework of the articles. We searched the experimental articles according to the content of the framework. When reading experimental articles, we categorize them according to the topics and methods of the study. In the case of controversy in the process of collecting and organizing, we selected articles with a higher impact factor or listed controversial cases. After that, we collected and sorted out the relevant contents and completed the literature review of this article. The parts with similar content are arranged in tables, and pictures supplement the problematic parts to describe them in words.

NON-CODING RNA AND NEURONAL DIFFERENTIATION

Usually, neuron cells can choose to differentiate or proliferate, which means that when a neuronal cell differentiates, its ability to proliferate is inhibited (Xie, Sen & Li, 2010). The Notch signaling pathway involves differentiation and neuronal differentiation (Bian et al., 2021). Hes1 is a classic target of the Notch signaling pathway, and high expression of Hes1 inhibits Ascl1 and maintains neural stem cell quiescence. Inactivation of Hes1 and related genes results in the premature termination of neurogenesis, and the healthy activity of neural stem cells depends on Hes1 oscillations (Sueda et al., 2019). LncND inhibits neuronal differentiation by sponging miR-143-3p, suppressing Notch protein expression (Rani et al., 2016). NBAT-1 inhibits neuroblastoma cell proliferation and promotes neuronal differentiation (Pandey et al., 2014). Without SIRT6, H19 inhibits neurogenesis through the p53/Notch1 pathway (Zhang et al., 2018a) (Fig. 2).

In addition to controlling neuronal differentiation through the Notch signaling pathway, lncRNAs and microRNAs can also affect other mRNAs, thereby controlling neuronal differentiation. PTBP1 (polypyrimidine tract-binding protein 1) interacts with Pkny, and the knockdown of Pkny or PTBP1 enhances neurogenesis (Ramos et al., 2015). LncKdm2b cis-activates Kdm2b and promotes cortical neuronal differentiation (Li et al., 2020). LncRNA-1604 orchestrates neural differentiation by competing with the core transcription factors ZEB1 and ZEB2 for miR-200c (Walgrave et al., 2021). In the Dicer1-deletion mouse model, miRNAs are verified to be involved in the survival of differentiated neurons, the maintenance of differentiated neuronal cells and brain homeostasis, and the generation of neurons during embryonic cortex generation (Lau & Hudson, 2010). MiR-124 promotes neuronal differentiation by targeting DACT1 or Neat1 through the Wnt/ β -catenin signaling pathway (Jiao et al., 2018; Cui et al., 2019).

NON-CODING RNAs REGULATE NEURONAL FUNCTION

The effect of ncRNAs on synapse plasticity

Synaptic scale-related protein dysfunction is associated with neurological diseases, and abnormal ncRNA expression is usually associated with protein dysfunction (Fernandes & Carvalho, 2016). MiRNA biogenesis is affected by cellular homeostasis, and miRNAs have been reported to be associated with synaptic disorders (Zheng et al., 2021). MicroRNAs associated with synaptic plasticity or synaptic disorders are listed in Table 1. LncRNA can control cell differentiation by recruiting transcription mechanisms. KCNA2-AS blocks the recovery of neuronal plasticity after peripheral nerve injury (Briggs et al., 2015). ADEPTR deletion can inhibit activity-dependent synaptic transmission changes and dendritic spine structural plasticity (Grinman et al., 2021).

Neuronal degeneration and ncRNAs

Neuronal death caused by progressive neuronal structure or function loss is considered neurodegeneration. Aging, oxidative damage and inflammation are all important factors causing neurodegeneration. Neuronal degeneration caused by environmental toxicants accelerates with aging (Goedert, Eisenberg & Crowther, 2017). miR-29 is

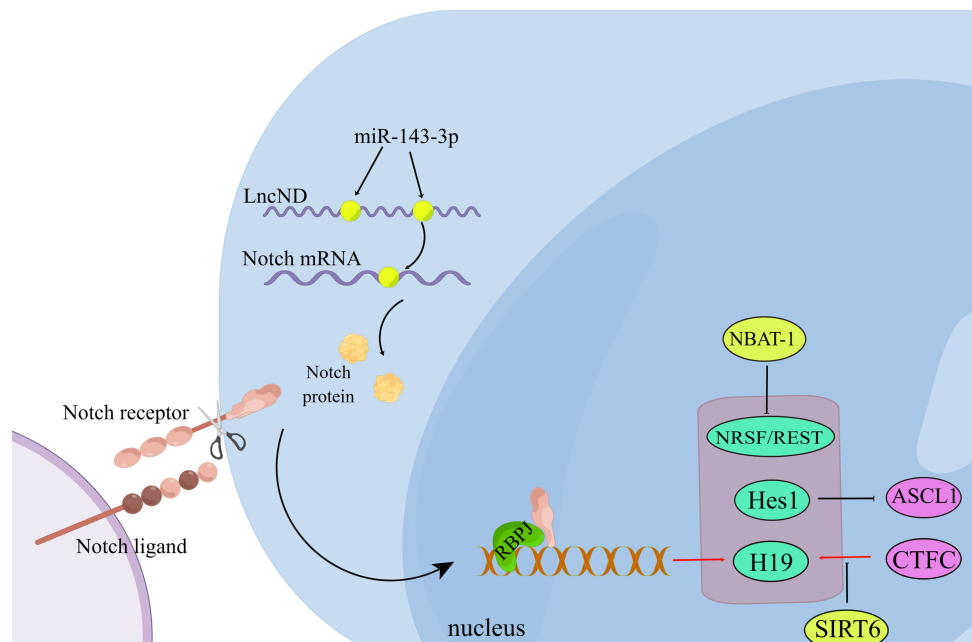


Figure 2 Non-coding RNA regulates neuronal differentiation through the Notch signaling pathway.

During the interaction between the Notch ligand and the Notch receptor, the intracellular domain of Notch is cleaved and then transferred to the nucleus, where it interacts with the transcription complex containing RBPJ, resulting in the expression of various Notch target genes. LncND chelates miR-143-3p and releases the expression of *NOTCH* mRNA, thus increasing the production of NOTCH protein required for the maintenance of neural progenitor cells. (Figure made with Figdraw).

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adaptively upregulated with aging, and downregulation of the miR-29 family promotes neurodegenerative diseases (Ripa et al., 2017; Jauhari, Singh & Yadav, 2018). Because the brain has high oxidative metabolic activity but low antioxidant capacity, the brain is highly vulnerable to oxidative stress damage (Salim, 2017). H19 targets miR-139 to protect H9c2 cells from hypoxia-induced damage (Gong et al., 2017). Inflammation is a significant contributor to cerebral infarction dysfunction. IL-1 stimulates TNF α and IL-1 β production by releasing arachidonic acid, which leads to inflammatory aggregation and brain damage (Zhang et al., 2018b). MiR-223 deficiency significantly improved clinical symptoms of CNS inflammation, demyelination, and EAE and increased resting microglia numbers and brain microglial autophagy (Li et al., 2019).

Neuronal function and ncRNAs

Much work has demonstrated the relationship between ncRNAs and neuronal function. Dicer deletion reduced the number of mature miRNAs, enhancing learning and memory (Konopka et al., 2010). MiR-132 and miR-134 combination might increase the expression of proteins such as Brain-Derived Neurotrophic Factor (BDNF) and Cyclic AMP response element-binding protein (CREB), thereby increasing the formation and maturation of dendritic spines (Im & Kenny, 2012). BS-DRL1 interacts with HMGB1 in neurons and regulates responses to DNA damage and genome stability (Maldonado-Lasuncion et al.,

Table 1 MicroRNAs associated with synaptic plasticity or synaptic disorders.

MicroRNA	Target mRNA	Functions	Refs
miR-34a	Unknown	Increased miR-34a gene expression may lead to dysfunction of synaptic plasticity, energy metabolism, and resting-state network activity.	<i>Sarkar et al. (2016)</i>
miR-92a	GluA1(Gria1)	Regulate expression of synaptic GluA1-containing AMPA receptors during homeostatic scaling.	<i>Letellier et al. (2014)</i>
miR-124	GluA2	Express homeostatic synaptic plasticity.	<i>Hou et al. (2015)</i>
miR-129-5p	Atp2b4 and Dcx	Downregulate Rbfox1 expression and inhibit Atp2b4 and Dcx.	<i>Rajman et al. (2017)</i>
miR-132	MMP-9	Regulate structural plasticity of dendritic spines through MMP-9.	<i>Jasińska et al. (2016)</i>
miR-135a-5p	Rock2/ Add1	Loss of miR-135a-5p results in elevated levels of Rock2 and phosphorylation of Ser726 on Add1, resulting in synaptic dysfunction and memory impairment.	<i>Zheng et al. (2021)</i>
miR-186-5p	GluA2	Increased synaptic expression of GluA2-lacking AMPA receptors, and block synaptic scaling.	<i>Silva et al. (2019)</i>
miR-455-3p	Unknown	High levels of miR-455-3p enhances mitochondrial biogenesis, mitochondrial function, and synaptic activity.	<i>Kumar et al. (2021)</i>
miR-484	Unknown	Predicted targets of miR-484 were enriched in brain proteins involved in the regulation of synaptic transmission and synaptic plasticity.	<i>Wingo et al. (2020)</i>
miR-485	SV2	Negatively regulate dendritic spine density, PSD-95 clustering, and surface expression of GluR2.	<i>Cohen et al. (2011)</i>

Notes.

MMP-9, matrix metalloproteinase-9; Add1, adducin 1; PSD-95, postsynaptic density95

2019). Furthermore, lncRNAs were found to serve a functional role in gender differences in depression susceptibility. LINC00473 reduced the amplitude and frequency of sEPSCs only in mPFC pyramidal neurons in female mice and the mPFC and several other forebrain regions in depressed females (*Gururajan, 2020; Issler et al., 2020*).

NON-CODING RNAS IN NEURODEGENERATION

Major neurodegenerative diseases include Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), frontotemporal lobar dementia (FTLD), and amyotrophic lateral sclerosis (ALS). Discrete populations of neurons are lost or damaged in nearly every neurodegenerative disease (*Malhi et al., 2014*). Here, we discussed the relationship between ncRNAs and several common neurodegenerative diseases; other neurodegenerative diseases are listed in [Table 2](#).

Non-coding RNAs and Alzheimer's Disease

Alzheimer's disease (AD) is the prevalent CNS degenerative disease, which can lead to mood disturbances, cognitive decline and even death (*Kumar, Singh & Ekavali, 2015*). The most striking pathological feature of AD is the "accumulation" of amyloid beta (A β) peptides and intracellular neurofibrillary tangles (NFTs) (*Iqbal et al., 2005*). Research shows that ncRNA is associated with increased risk of AD. BACE1 (β -site amyloid precursor protein cleaving enzyme 1) catalyzes APP cleavage to generate β -amyloid peptides. BACE1-AS and

Table 2 Illustrative list of ncRNAs that are disrupted in neuronal disorders.

Type	Disease	Involved ncRNAs	Refs
	AD	BACE1-AS, GDNFOS, 17A, NAT-Rad18, BC200, Sox2OT, NDM29, 51A, 1810014B01Rik	Mus, Hof & Tiedge (2007), Parenti et al. (2007), Airavaara et al. (2011), Arisi et al. (2011), Massone et al. (2011), Massone et al. (2012), Ciarlo et al. (2013) and Liu et al. (2014)
	ALS/FTLD	C9ORF72, NEAT1-2	Nishimoto et al. (2013) and Zu et al. (2013)
	AS	UBE3A-AS	Johnstone et al. (2006)
	Autism	ST7OT (anti-sense to ST7)	Vincent et al. (2002)
	BWS	LIT1 (anti-sense KvLQT1), Peg8, H19	Horike et al. (2000), Okutsu et al. (2000) and Sparago et al. (2004)
	GABA neuropathies	Evf-2 (anti-sense Dlx6)	Feng et al. (2006)
	Fragile X syndrome	BC1	Zalfa et al. (2005)
	HD	HDD-AS, HAR1, TUG1, NEAT1, DGCR5, MEG3	Johnson et al. (2009), Johnson et al. (2010b), Chung et al. (2011) and Johnson (2012)
lncRNA	Long-term memory disorders	Anti-NOS (anti-sense nNOS)	Korneev et al. (2005)
	Neuronal hyperexcitability	EVF2	Bond et al. (2009)
	PD	Uchl1-AS, PINK1-AS (naPINK1), Sox2OT, BC200, 1810014B01Rik	Scheele et al. (2007), Carriero et al. (2015) and Luo et al. (2015)
	PWS	UBE3A-AS, IPW, ZNF127-AS (anti-sense ZNF127)	Wevrick & Francke (1997), Jong et al. (1999) and Chamberlain & Brannan (2001)
	Schizophrenia	GOMAFU, DISC2 (anti-sense DISC1), PSZA11q14 (anti-sense DLG2)	Polesskaya et al. (2003), Millar et al. (2004) and Barry et al. (2014)
	Spinocerebellar ataxia 8	SCA8 (ATXN8OS)	Daughters et al. (2009)
miRNA	AD	miR-29, miR-146, let-7, miR-9, miR-124, miR-138, miR-181, miR-125, miR-485, miR-107, miR-200, miR-34	Scheele et al. (2007), Lu et al. (2008), Nowak & Michlewski (2013) and Yang et al. (2017)
	ASD	miR-30	Nowak & Michlewski (2013)
	Down's syndrome	let-7, miR-125, mir-155, miR-802	Kuhn et al. (2010) and Nowak & Michlewski (2013)
	Fragile X syndrome	miR-124, miR-132, miR-125	Nowak & Michlewski (2013)
	HD	miR-9, miR-124, miR-132	Nowak & Michlewski (2013)
	PD	miR-7, miR-184, let-7, miR-133, miR-34	Gehrke et al. (2010) and Nowak & Michlewski (2013)
	Rett's syndrome	miR-146a, miR-146b, miR-29, miR-382, miR-132,	Urduinguio et al. (2010), Wu et al. (2010) and Nowak & Michlewski (2013)
	Spinal motor neuron disease	miR-9	Haramati et al. (2010)
	Schizophrenia	miR-134, miR-181, miR-219, miR-198	Nowak & Michlewski (2013)
	Spinocerebellar ataxia type 1	miR-19, miR-101, miR-100	Haramati et al. (2010)
circRNA	AD	ciRS-7	Lukiw (2013)
T-UCR	Idiopathic neurodevelopmental disease	T-UCRs uc.195, uc.392, uc.46 and uc.222	Lukiw (2013)
snoRNA	PWS	snoRNA cluster at 15q11–q13 imprinted locus	Kishore & Stamm (2006), Horsthemke & Wagstaff (2008) and Sahoo et al. (2008)

Notes.

There is not necessarily a clear delineation between classes of non-coding RNA (ncRNA).

AD, Alzheimer's disease; ALS/FTD, Amyotrophic lateral sclerosis and Frontotemporal dementia; AS, Angelman syndrome; BWS, Beckwith–Wiedemann Syndrome; HD, Huntington's disease; PD, Parkinson's disease; PWS, Prader–Willi Syndrome; ASD, Autism spectrum disorders.

miR-485-5p may share the same binding site with the sixth exon of the BACE1 mRNA transcript, preventing miRNA-induced repression of BACE1 mRNA (Roberts, Morris & Wood, 2014). MiR-9, miR-125b, and miR-128 are expressed in the Alzheimer's disease brain over the average adult abundance (Lukiw, 2007), whereas miR-107 is reduced in early AD and may regulate BACE1 to promote disease progression (Wang et al., 2008b). The MiR-29 family is significantly reduced in AD patients, accompanied by abnormally high levels of BACE1 protein (Shioya et al., 2010). The neuronal sortilin-related receptor gene (SORL1) interacts with APP in the endosome or trans-Golgi network, affecting trafficking and proteolytic processing, thereby increasing the risk of AD (Schipper et al., 2007). 51A reduces the synthesis of SORL1 variant A by driving the SPRL1 splicing shift, thereby impairing APP processing and increasing A β production (Qiu-Lan et al., 2009).

Non-coding RNAs and Parkinson's Disease

Parkinson's disease (PD) primarily affects the motor system of the CNS. With the deterioration of PD, there will also be autonomic dysfunction and other non-motor symptoms (Reich & Savitt, 2019). Hereditary PD family cases are mainly caused by mutations in the genes SNCA, PARKIN, UCHL1, PINK1, DJ-1, and LRKK2 (Xiromerisiou et al., 2010). AS-Uchl1 induces Uchl1 expression by promoting Nurr1 translation. Uchl1 overexpression may be beneficial for treating neurodegenerative diseases (Carriero et al., 2015). The knockdown of NEAT1 significantly inhibits autophagy in PD, thereby attenuating dopaminergic neuron damage (Yan et al., 2018). Alpha-synuclein (SNCA) is detrimental to dopamine neurons, both miR-7 and miR-153 effects on SNCA expression additively (Mouradian, 2012). Downregulation of miR-34b/c occurred in several brain regions in PD patients, which underlies early mitochondrial dysfunction in PD (Wang et al., 2008c). Screening for abnormal expression of ncRNAs in PD patients and models is beneficial to the finding of novel biomarkers or therapeutic targets, but further studies on the pathogenesis of PD are still needed.

Non-coding RNAs and Huntington's disease

Huntington's disease (HD) is a fatal dominant neurodegenerative disorder induced by repeat expansions of cytosine-adenine-guanine trinucleotides in the Huntington gene. Its symptoms include chorea, mental problems and dementia (Zuccato et al., 2003). Overexpression of NEAT1 contributes neuroprotection against neuronal damage in HD through a cell survival pathway under stress conditions (Choudhry et al., 2015). Human accelerated region 1 (HAR1) is specifically expressed in the nervous system, and the level of HAR1 in the striatum of HD patients is significantly lower than that of untreated patients (Sunwoo et al., 2017). miR-9, miR-9*, miR-29b, and miR-124 are down-regulated in HD, while miR-330 is up-regulated in HD (Packer et al., 2008). miR-29a is up-regulated in HD but down-regulated in mouse cortex (Johnson et al., 2008). This difference may be due to the different methods used to analyze miRNA expression and continuing problems with RNA quality and integrity in the postmortem human brain (Johnson & Buckley, 2009).

Non-coding RNAs and Other Neurodegeneration Disease

In a mouse model of Amyotrophic Lateral Sclerosis (ALS), an antisense oligonucleotide (ASO) inhibitor of miR-129-5p significantly increases survival and improves the neuromuscular phenotype of treated mice (Lu et al., 2021). MiR-155 may be a candidate for co-silencing of miR-129 as its function is mainly involved in CNS inflammation by regulating microglia (Koval et al., 2013). A co-role of miR-183/96/182 has been demonstrated in the pathogenesis of ALS/FTD-related aging and cognitive function (Jawaid et al., 2019). MiR-146a-5p may play a vital role in regulating neurogenesis in the pathological process of depression. The DG is a critical region for neurogenesis in the adult brain, and microglia-derived exosomes transport miR-146a-5p to the DG region to regulate neuronal function.

EARLY DIAGNOSIS AND TREATMENT BASED ON MIRNA

Research shows that early diagnosis can reduce the risk of Alzheimer's disease by one-third (Norton et al., 2014). However, the existing diagnostic methods are invasive or costly (Dolgin, 2018). In addition, due to individual differences, the accuracy of these methods is low (McKhann et al., 2011). It is reported that the miRNA profile in brain tissue and blood of patients with neurodegenerative diseases has changed, so miRNA has excellent potential as a biomarker (Lee et al., 2021). The cost of new drug discovery is very high, and the period is extended (Mohs & Greig, 2017). It is an ideal solution to change the expression of specific miRNAs by inhibitors or endogenous substances or to use miRNA-targeted drug delivery (Hu et al., 2018; Ouyang et al., 2022). When the expression of miRNAs is changed in cells, it may affect drug sensitivity and regulate drug resistance to standard cancer therapy, thus having a more substantial therapeutic effect (Adams et al., 2017).

Despite great potential, the biological application of miRNA still has some insuperable limitations. First, miRNA is prone to degrade *in vivo* due to circulating RNase or cell endocytosis chamber, and its half-life is very short (Ramachandran & Chen, 2008). Second, the acceptable delivery methods for human beings are limited. These methods have low penetration efficiency of the blood-brain barrier (BBB) or are unable to target delivery, or the delivery dose is limited (Lee et al., 2019; Ul Islam et al., 2020). Third, although transfection reagents or nanoparticles can solve the problem of penetrating the blood-brain barrier, the toxicity of small therapeutic oligonucleotides is still a problem (Ramachandran & Chen, 2008; Ruberti, Barbato & Cogoni, 2012). Last, the relationship between miRNAs and target genes is not always a 1:1 match, and some imitations of miRNAs may lead to off-target effects (Loganatharaj & Randall, 2017).

Despite these limitations, a miRNA is still a powerful tool for the early detection and treatment of neurodegenerative diseases. Thus, we suggest the following points to exploit the potential of miRNA fully. (i) Conduct statistical analysis with the help of bioinformatics to accurately guide the research direction. (ii) Combining with endogenous competitive RNA (ceRNA) mechanism, further improve the linkage map between miRNA and neurodegenerative diseases to seek more accurate targets. (iii) Further modify delivery materials (such as nanoparticles) and oligonucleotides to improve penetration and reduce toxicity.

CONCLUDING REMARKS

Here, we introduced ncRNAs and the nervous system separately and discussed the roles of ncRNAs in neuronal differentiation, function, and diseases. Although the mechanisms by which ncRNAs affect neuronal function and dysfunction are not fully understood, this review summarizes the current relevant research results. Because the differentiation of the nervous system happens mainly in a specific period, ncRNAs often delay or advance differentiation by acting on specific signaling pathways, thereby affecting the differentiation process. The current findings suggest that neurodegenerative diseases are often the result of disturbances at the protein level. LncRNAs and microRNAs often compete and alter protein stability, thereby changing the ratio of protein concentrations. Due to individual differences and the complexity of the nervous system, there are sometimes two independent studies with opposing results. In the end, our exploration of neuronal remains the tip of the iceberg. It cannot be excluded that with further research on ncRNAs and neurons in the future, more targets will be unlocked for therapeutic purposes.

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Cheng Xuan performed the experiments, prepared figures and/or tables, and approved the final draft.
- Enyu Yang performed the experiments, prepared figures and/or tables, and approved the final draft.
- Shuo Zhao performed the experiments, prepared figures and/or tables, and approved the final draft.

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- Yaping Zhang conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Zhenggang Jiang conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Xianfeng Ding performed the experiments, authored or reviewed drafts of the article, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

This is a literature review and does not have raw data.

REFERENCES

- Adams BD, Parsons C, Walker L, Zhang WC, Slack FJ. 2017. Targeting noncoding RNAs in disease. *The Journal of Clinical Investigation* 127:761–771 DOI 10.1172/JCI84424.
- Airavaara M, Pletnikova O, Doyle ME, Zhang YE, Troncoso JC, Liu QR. 2011. Identification of novel GDNF isoforms and cis-antisense GDNFOS gene and their regulation in human middle temporal gyrus of Alzheimer disease. *Journal of Biological Chemistry* 286:45093–45102 DOI 10.1074/jbc.M111.310250.
- Amaral PP, Neyt C, Wilkins SJ, Askarian-Amiri ME, Sunkin SM, Perkins AC, Mattick JS. 2009. Complex architecture and regulated expression of the Sox2ot locus during vertebrate development. *RNA* 15:2013–2027 DOI 10.1261/RNA.1705309.
- Arisi I, D’Onofrio M, Brandi R, Felsani A, Capsoni S, Drovandi G, Felici G, Weitschek E, Bertolazzi P, Cattaneo A. 2011. Gene expression biomarkers in the brain of a mouse model for Alzheimer’s disease: mining of microarray data by logic classification and feature selection. *Journal of Alzheimer’s Disease* 24:721–738 DOI 10.3233/JAD-2011-101881.
- Armstrong R. 2020. What causes neurodegenerative disease? *Folia Neuropathologica* 58:93–112 DOI 10.5114/FN.2020.96707.
- Balzeau J, Menezes MR, Cao S, Hagan JP. 2017. The LIN28/let-7 pathway in cancer. *Frontiers in Genetics* 8:31 DOI 10.3389/FGENE.2017.00031.
- Barry G, Briggs JA, Vanichkina DP, Poth EM, Beveridge NJ, Ratnu VS, Nayler SP, Nones K, Hu J, Bredy TW, Nakagawa S, Rigo F, Taft RJ, Cairns MJ, Blackshaw S, Wolvetang EJ, Mattick JS. 2014. The long non-coding RNA Gomafu is acutely regulated in response to neuronal activation and involved in schizophrenia-associated alternative splicing. *Molecular Psychiatry* 19:486–494 DOI 10.1038/mp.2013.45.
- Bian W, Tang M, Jiang H, Xu W, Hao W, Sui Y, Hou Y, Nie L, Zhang H, Wang C, Li N, Wang J, Qin J, Wu L, Ma X, Chen J, Wang W, Li X. 2021. Low-density-lipoprotein-receptor-related protein 1 mediates Notch pathway activation. *Developmental Cell* 56:2902–2919 DOI 10.1016/J.DEVCEL.2021.09.015.

- Bond AM, Vangompel MJW, Sametsky EA, Clark MF, Savage JC, Disterhoft JF, Kohtz JD. 2009.** Balanced gene regulation by an embryonic brain ncRNA is critical for adult hippocampal GABA circuitry. *Nature Neuroscience* **12**:1020–1027 DOI [10.1038/nn.2371](https://doi.org/10.1038/nn.2371).
- Bridges MC, Daulagala AC, Kourtidis A. 2021.** LNCcation: lncRNA localization and function. *The Journal of Cell Biology* **220**(2):e202009045 DOI [10.1083/JCB.202009045](https://doi.org/10.1083/JCB.202009045).
- Briggs JA, Wolvetang EJ, Mattick JS, Rinn JL, Barry G. 2015.** Mechanisms of long non-coding RNAs in mammalian nervous system development, plasticity, disease, and evolution. *Neuron* **88**:861–877 DOI [10.1016/j.neuron.2015.09.045](https://doi.org/10.1016/j.neuron.2015.09.045).
- Bruscella P, Bottini S, Baudesson C, Pawlowsky JM, Feray C, Trabucchi M. 2017.** Viruses and miRNAs: more friends than foes. *Frontiers in Microbiology* **8**:824 DOI [10.3389/fmicb.2017.00824](https://doi.org/10.3389/fmicb.2017.00824).
- Carrieri C, Forrest ARR, Santoro C, Persichetti F, Carninci P, Zucchelli S, Gustincich S. 2015.** Expression analysis of the long non-coding RNA antisense to Uchl1 (AS Uchl1) during dopaminergic cells' differentiation in vitro and in neurochemical models of Parkinson's disease. *Frontiers in Cellular Neuroscience* **9**:114 DOI [10.3389/fncel.2015.00114](https://doi.org/10.3389/fncel.2015.00114).
- Chamberlain SJ, Brannan CI. 2001.** The Prader-Willi syndrome imprinting center activates the paternally expressed murine Ube3a antisense transcript but represses paternal Ube3a. *Genomics* **73**:316–322 DOI [10.1006/geno.2001.6543](https://doi.org/10.1006/geno.2001.6543).
- Chen X, Guan NN, Sun YZ, Li JQ, Qu J. 2018.** MicroRNA-small molecule association identification: from experimental results to computational models. *Briefings in Bioinformatics* **21**:47–61 DOI [10.1093/BIB/BBY098](https://doi.org/10.1093/BIB/BBY098).
- Chen X, Sun YZ, Guan NN, Qu J, Huang ZA, Zhu ZX, Li JQ. 2019a.** Computational models for lncRNA function prediction and functional similarity calculation. *Briefings in Functional Genomics* **18**:58–82 DOI [10.1093/BFGP/ELY031](https://doi.org/10.1093/BFGP/ELY031).
- Chen X, Xie D, Zhao Q, You ZH. 2019b.** MicroRNAs and complex diseases: from experimental results to computational models. *Briefings in Bioinformatics* **20**:515–539 DOI [10.1093/bib/bbx130](https://doi.org/10.1093/bib/bbx130).
- Chen X, Yan CC, Zhang X, You ZH. 2017.** Long non-coding RNAs and complex diseases: from experimental results to computational models. *Briefings in Bioinformatics* **18**:558–576 DOI [10.1093/BIB/BBW060](https://doi.org/10.1093/BIB/BBW060).
- Cheong CY, Chng K, Ng S, Chew SB, Chan L, Ferguson-Smith AC. 2015.** Germline and somatic imprinting in the nonhuman primate highlights species differences in oocyte methylation. *Genome Research* **25**:611–623 DOI [10.1101/GR.183301.114](https://doi.org/10.1101/GR.183301.114).
- Choudhry H, Albukhari A, Morotti M, Haider S, Moralli D, Smythies J, Schödel J, Green CM, Camps C, Buffa F, Ratcliffe P, Ragoussis J, Harris AL, Mole DR. 2015.** Tumor hypoxia induces nuclear paraspeckle formation through HIF-2 α dependent transcriptional activation of NEAT1 leading to cancer cell survival. *Oncogene* **34**:4482–4490 DOI [10.1038/onc.2014.378](https://doi.org/10.1038/onc.2014.378).
- Chung DW, Rudnicki DD, Yu L, Margolis RL. 2011.** A natural antisense transcript at the Huntington's disease repeat locus regulates HTT expression. *Human Molecular Genetics* **20**:3467–3477 DOI [10.1093/hmg/ddr263](https://doi.org/10.1093/hmg/ddr263).

- Ciarlo E, Massone S, Penna I, Nizzari M, Gigoni A, Dieci G, Russo C, Florio T, Cancedda R, Pagano A. 2013.** An intronic ncRNA-dependent regulation of SORL1 expression affecting A β formation is upregulated in post-mortem Alzheimer's disease brain samples. *Disease Models & Mechanisms* **6**:424–433 DOI [10.1242/DMM.009761](https://doi.org/10.1242/DMM.009761).
- Cohen JE, Lee PR, Chen S, Li W, Fields RD. 2011.** MicroRNA regulation of homeostatic synaptic plasticity. *Proceedings of the National Academy of Sciences of the United States of America* **108**:11650–11655 DOI [10.1073/pnas.1017576108](https://doi.org/10.1073/pnas.1017576108).
- Cui Y, Yin Y, Xiao Z, Zhao Y, Chen B, Yang B, Xu B, Song H, Zou Y, Ma X, Dai J. 2019.** LncRNA Neat1 mediates miR-124-induced activation of Wnt/ β -catenin signaling in spinal cord neural progenitor cells. *Stem Cell Research & Therapy* **10**(1):400 DOI [10.1186/S13287-019-1487-3](https://doi.org/10.1186/S13287-019-1487-3).
- Cui Z, Liu JX, Gao YL, Zhu R, Yuan SS. 2020.** LncRNA-disease associations prediction using bipartite local model with nearest profile-based association inferring. *IEEE Journal of Biomedical and Health Informatics* **24**:1519–1527 DOI [10.1109/JBHI.2019.2937827](https://doi.org/10.1109/JBHI.2019.2937827).
- Daughters RS, Tuttle DL, Gao W, Ikeda Y, Moseley ML, Ebner TJ, Swanson MS, Ranum LPW. 2009.** RNA gain-of-function in spinocerebellar ataxia type 8. *PLOS Genetics* **5**:e1000600 DOI [10.1371/journal.pgen.1000600](https://doi.org/10.1371/journal.pgen.1000600).
- Dolgin E. 2018.** Alzheimer's disease is getting easier to spot. *Nature* **559**:S10–S12 DOI [10.1038/D41586-018-05721-W](https://doi.org/10.1038/D41586-018-05721-W).
- Feng J, Bi C, Clark BS, Mady R, Shah P, Kohtz JD. 2006.** The Evf-2 noncoding RNA is transcribed from the Dlx-5/6 ultraconserved region and functions as a Dlx-2 transcriptional coactivator. *Genes and Development* **20**:1470–1484 DOI [10.1101/gad.1416106](https://doi.org/10.1101/gad.1416106).
- Fernandes D, Carvalho AL. 2016.** Mechanisms of homeostatic plasticity in the excitatory synapse. *Journal of Neurochemistry* **139**:973–996 DOI [10.1111/JNC.13687](https://doi.org/10.1111/JNC.13687).
- Friedman RC, Farh KKH, Burge CB, Bartel DP. 2009.** Most mammalian mRNAs are conserved targets of microRNAs. *Genome Research* **19**:92–105 DOI [10.1101/GR.082701.108](https://doi.org/10.1101/GR.082701.108).
- Gehrke S, Imai Y, Sokol N, Lu B. 2010.** Pathogenic LRRK2 negatively regulates microRNA-mediated translational repression. *Nature* **466**:637–641 DOI [10.1038/nature09191](https://doi.org/10.1038/nature09191).
- Goedert M, Eisenberg DS, Crowther RA. 2017.** Propagation of Tau aggregates and neurodegeneration. *Annual Review of Neuroscience* **40**:189–210 DOI [10.1146/ANNUREV-NEURO-072116-031153](https://doi.org/10.1146/ANNUREV-NEURO-072116-031153).
- Gong C, Maquat LE. 2011.** lncRNAs transactivate STAU1-mediated mRNA decay by duplexing with 3' UTRs via Alu elements. *Nature* **470**:284–290 DOI [10.1038/NATURE09701](https://doi.org/10.1038/NATURE09701).
- Gong LC, Xu HM, Guo GL, Zhang T, Shi JW, Chang H. 2017.** Long non-coding RNA H19 protects H9c2 cells against hypoxia-induced injury by targeting MicroRNA-139. *Cellular Physiology and Biochemistry* **44**:857–869 DOI [10.1159/000485354](https://doi.org/10.1159/000485354).
- Grinman E, Nakahata Y, Avchalumov Y, Espadas I, Swarnkar S, Yasuda R, Puthanveetil SV. 2021.** Activity-regulated synaptic targeting of lncRNA ADEPTR mediates

- structural plasticity by localizing Sptn1 and AnkB in dendrites. *Science Advances* 7(16):eabf0605 DOI 10.1126/sciadv.abf0605.
- Gururajan A.** 2020. Sex differences in susceptibility to depression: a role for LncRNAs. *Neuron* 106:871–872 DOI 10.1016/j.neuron.2020.05.016.
- Haramati S, Chapnik E, Sztainberg Y, Eilam R, Zwang R, Gershoni N, McGlenn E, Heiser PW, Wills AM, Wirguin I, Rubin LL, Misawa H, Tabin CJ, Brown R, Chen A, Hornstein E.** 2010. miRNA malfunction causes spinal motor neuron disease. *Proceedings of the National Academy of Sciences of the United States of America* 107:13111–13116 DOI 10.1073/pnas.1006151107.
- Hombach S, Kretz M.** 2016. Non-coding RNAs: classification, biology and functioning. *Advances in Experimental Medicine and Biology* 937:3–17 DOI 10.1007/978-3-319-42059-2_1.
- Horike SI, Mitsuya K, Meguro M, Kotobuki N, Kashiwagi A, Notsu T, Schulz TC, Shirayoshi Y, Oshimura M.** 2000. Targeted disruption of the human LIT1 locus defines a putative imprinting control element playing an essential role in Beckwith-Wiedemann syndrome. *Human Molecular Genetics* 9:2075–2083 DOI 10.1093/hmg/9.14.2075.
- Horsthemke B, Wagstaff J.** 2008. Mechanisms of imprinting of the Prader-Willi/Angelman region. *American Journal of Medical Genetics, Part A* 146:2041–2052 DOI 10.1002/ajmg.a.32364.
- Hou Q, Ruan H, Gilbert J, Wang G, Ma Q, Yao WD, Man HY.** 2015. MicroRNA miR124 is required for the expression of homeostatic synaptic plasticity. *Nature Communications* 6:10045 DOI 10.1038/NCOMMS10045.
- Hu WZ, Tan CL, He YJ, Zhang GQ, Xu Y, Tang JH.** 2018. Functional miRNAs in breast cancer drug resistance. *OncoTargets and Therapy* 11:1529–1541 DOI 10.2147/OTT.S152462.
- Huang L, Zhang L, Chen X.** 2022. Updated review of advances in microRNAs and complex diseases: experimental results, databases, webservers and data fusion. *Briefings in Bioinformatics* 23(6):bbac397 DOI 10.1093/BIB/BBAC397.
- Im HI, Kenny PJ.** 2012. MicroRNAs in neuronal function and dysfunction. *Trends in Neurosciences* 35:325–334 DOI 10.1016/j.tins.2012.01.004.
- Iqbal K, Alonso AC, Chen S, Chohan MO, El-akkad E, Gong C, Khatoon S, Li B, Liu F, Rahman A, Tanimukai H, Grundke-iqbal I.** 2005. Tau pathology in Alzheimer disease and other tauopathies. *Biochimica et Biophysica Acta—Molecular Basis of Disease* 1739:198–210 DOI 10.1016/j.bbadis.2004.09.008.
- Issler O, Van der Zee YY, Ramakrishnan A, Wang J, Tan C, Loh YHE, Purushothaman I, Walker DM, Lorsch ZS, Hamilton PJ, Peña CJ, Flaherty E, Hartley BJ, Torres-Berrío A, Parise EM, Kronman H, Duffy JE, Estill MS, Calipari ES, Labonté B, Neve RL, Tamminga CA, Brennand KJ, Dong Y, Shen L, Nestler EJ.** 2020. Sex-specific role for the long non-coding RNA LINC00473 in depression. *Neuron* 106:912–926 DOI 10.1016/j.neuron.2020.03.023.

- Jasińska M, Miłek J, Cymerman IA, Łęski S, Kaczmarek L, Dziembowska M. 2016. miR-132 regulates dendritic spine structure by direct targeting of matrix metalloproteinase 9 mRNA. *Molecular Neurobiology* 53:4701–4712 DOI 10.1007/S12035-015-9383-Z.
- Jauhari A, Singh T, Yadav S. 2018. Expression of miR-145 and its target proteins are regulated by miR-29b in differentiated neurons. *Molecular Neurobiology* 55:8978–8990 DOI 10.1007/S12035-018-1009-9.
- Jawaid A, Woldemichael BT, Kremer EA, Laferriere F, Gaur N, Afroz T, Polymenidou M, Mansuy IM. 2019. Memory decline and its reversal in aging and neurodegeneration involve miR-183/96/182 biogenesis. *Molecular Neurobiology* 56:3451–3462 DOI 10.1007/S12035-018-1314-3.
- Jiao S, Liu Y, Yao Y, Teng J. 2018. miR-124 promotes proliferation and neural differentiation of neural stem cells through targeting DACT1 and activating Wnt/ β -catenin pathways. *Molecular and Cellular Biochemistry* 449:305–314 DOI 10.1007/S11010-018-3367-Z.
- Johnson R. 2012. Long non-coding RNAs in Huntington's disease neurodegeneration. *Neurobiology of Disease* 46:245–254 DOI 10.1016/j.nbd.2011.12.006.
- Johnson R, Buckley NJ. 2009. Gene dysregulation in Huntington's disease: REST, microRNAs and beyond. *Neuromolecular Medicine* 11:183–199 DOI 10.1007/s12017-009-8063-4.
- Johnson R, Richter N, Jauch R, Gaughwin PM, Zuccato C, Cattaneo E, Stanton LW. 2010a. Human accelerated region 1 noncoding RNA is repressed by REST in Huntington's disease. *Physiological Genomics* 41:269–274 DOI 10.1152/PHYSIOLGENOMICS.00019.2010.
- Johnson R, Richter N, Jauch R, Gaughwin PM, Zuccato C, Cattaneo E, Stanton LW. 2010b. Human accelerated region 1 noncoding RNA is repressed by REST in Huntington's disease. *Physiological Genomics* 41:269–274 DOI 10.1152/PHYSIOLGENOMICS.00019.2010/SUPPL_FILE/FIGS1.PDF.
- Johnson R, Teh CHL, Jia H, Vanisri RR, Pandey T, Lu ZH, Buckley NJ, Stanton LW, Lipovich L. 2009. Regulation of neural macroRNAs by the transcriptional repressor REST. *Rna* 15:85–96 DOI 10.1261/rna.1127009.
- Johnson R, Zuccato C, Belyaev ND, Guest DJ, Cattaneo E, Buckley NJ. 2008. A microRNA-based gene dysregulation pathway in Huntington's disease. *Neurobiology of Disease* 29:438–445 DOI 10.1016/j.nbd.2007.11.001.
- Johnstone KA, DuBose AJ, Futtner CR, Elmore MD, Brannan CI, Resnick JL. 2006. A human imprinting centre demonstrates conserved acquisition but diverged maintenance of imprinting in a mouse model for Angelman syndrome imprinting defects. *Human Molecular Genetics* 15:393–404 DOI 10.1093/hmg/ddi456.
- Jong MTC, Gray TA, Ji Y, Glenn CC, Saitoh S, Driscoll DJ, Nicholls RD. 1999. A novel imprinted gene, encoding a RING zinc-finger protein, and overlapping antisense transcript in the Prader-Willi syndrome critical region. *Human Molecular Genetics* 8:783–793 DOI 10.1093/hmg/8.5.783.
- Khaitovich P, Lockstone HE, Wayland MT, Tsang TM, Jayatilaka SD, Guo AJ, Zhou J, Somel M, Harris LW, Holmes E, Pääbo S, Bahn S. 2008. Metabolic

- changes in schizophrenia and human brain evolution. *Genome Biology* **9**(8):R124 DOI [10.1186/GB-2008-9-8-R124](https://doi.org/10.1186/GB-2008-9-8-R124).
- Kishore S, Stamm S. 2006.** The snoRNA HBII-52 regulates alternative splicing of the serotonin receptor 2C. *Science* **311**:230–232 DOI [10.1126/science.1118265](https://doi.org/10.1126/science.1118265).
- Konopka W, Kiryk A, Novak M, Herwerth M, Parkitna JR, Wawrzyniak M, Kowarsch A, Michaluk P, Dzwonek J, Arnsperger T, Wilczynski G, Merken-schlager M, Theis FJ, Köhr G, Kaczmarek L, Schütz G. 2010.** MicroRNA loss enhances learning and memory in mice. *Journal of Neuroscience* **30**:14835–14842 DOI [10.1523/JNEUROSCI.3030-10.2010](https://doi.org/10.1523/JNEUROSCI.3030-10.2010).
- Korneev SA, Straub V, Kemenes I, Korneeva EI, Ott SR, Benjamin PR, O’Shea M. 2005.** Timed and targeted differential regulation of nitric oxide synthase (NOS) and anti-NOS genes by reward conditioning leading to long-term memory formation. *Journal of Neuroscience* **25**:1188–1192 DOI [10.1523/JNEUROSCI.4671-04.2005](https://doi.org/10.1523/JNEUROSCI.4671-04.2005).
- Koval ED, Shaner C, Zhang P, Du Maine X, Fischer K, Tay J, Nelson Chau B, Wu GF, Miller TM. 2013.** Method for widespread microRNA-155 inhibition pro-longs survival in ALS-model mice. *Human Molecular Genetics* **22**:4127–4135 DOI [10.1093/HMG/DDT261](https://doi.org/10.1093/HMG/DDT261).
- Kuhn DE, Nuovo GJ, Terry AV, Martin MM, Malana GE, Sansom SE, Pleister AP, Beck WD, Head E, Feldman DS, Elton TS. 2010.** Chromosome 21-derived microRNAs provide an etiological basis for aberrant protein expression in hu-man down syndrome brains. *Journal of Biological Chemistry* **285**:1529–1543 DOI [10.1074/jbc.M109.033407](https://doi.org/10.1074/jbc.M109.033407).
- Kumar A, Singh A, Ekavali . 2015.** Pharmacological reports review article a review on Alzheimer’ s disease pathophysiology and its management: an update. *Pharmacologi-cal Reports* **67**(2):195–203 DOI [10.1016/j.pharep.2014.09.004](https://doi.org/10.1016/j.pharep.2014.09.004).
- Kumar S, Morton H, Sawant N, Orlov E, Bunquin LE, Pradeepkiran JA, Alvir R, Reddy PH. 2021.** MicroRNA-455-3p improves synaptic, cognitive functions and extends lifespan: relevance to Alzheimer’s disease. *Redox Biology* **48**:102182 DOI [10.1016/J.REDOX.2021.102182](https://doi.org/10.1016/J.REDOX.2021.102182).
- Lau P, Hudson LD. 2010.** MicroRNAs in neural cell differentiation. *Brain Research* **1338**:14–19 DOI [10.1016/J.BRAINRES.2010.04.002](https://doi.org/10.1016/J.BRAINRES.2010.04.002).
- Lee CY, Ryu IS, Ryu JH, Cho HJ. 2021.** miRNAs as therapeutic tools in Alzheimer’s disease. *International Journal of Molecular Sciences* **22**(23):13012 DOI [10.3390/IJMS222313012](https://doi.org/10.3390/IJMS222313012).
- Lee SWL, Paoletti C, Campisi M, Osaki T, Adriani G, Kamm RD, Mattu C, Chiono V. 2019.** MicroRNA delivery through nanoparticles. *Journal of Controlled Release* **313**:80–95 DOI [10.1016/J.JCONREL.2019.10.007](https://doi.org/10.1016/J.JCONREL.2019.10.007).
- Letellier M, Elramah S, Mondin M, Soula A, Penn A, Choquet D, Landry M, Thoumine O, Favereaux A. 2014.** miR-92a regulates expression of synaptic GluA1-containing AMPA receptors during homeostatic scaling. *Nature Neuroscience* **17**:1040–1042 DOI [10.1038/NN.3762](https://doi.org/10.1038/NN.3762).
- Li W, Shen W, Zhang B, Tian K, Li Y, Mu L, Luo Z, Zhong X, Wu X, Liu Y, Zhou Y. 2020.** Long non-coding RNA LncKdm2b regulates cortical neuronal differentiation by cis-activating Kdm2b. *Protein and Cell* **11**:161–186 DOI [10.1007/s13238-019-0650-z](https://doi.org/10.1007/s13238-019-0650-z).

- Li Y, Zhou D, Ren Y, Zhang Z, Guo X, Ma MK, Xue Z, Lv J, Liu H, Xi Q, Jia L, Zhang L, Liu Y, Zhang Q, Yan J, Da Y, Gao F, Yue J, Yao Z, Zhang R. 2019. Mir223 restrains autophagy and promotes CNS inflammation by targeting ATG16L1. *Autophagy* 15:478–492 DOI 10.1080/15548627.2018.1522467.
- Liu TE, Huang Y, Chen J, Chi H, Yu Z, Wang J, Chen C. 2014. Attenuated ability of BACE1 to cleave the amyloid precursor protein via silencing long noncoding RNA BACE1—AS expression. *Molecular Medicine Reports* 10:1275–1281 DOI 10.3892/mmr.2014.2351.
- Loganatharaj R, Randall TA. 2017. The limitations of existing approaches in improving MicroRNA target prediction accuracy. *Methods in Molecular Biology* 1617:133–158 DOI 10.1007/978-1-4939-7046-9_10.
- Lu YL, Liu Y, McCoy MJ, Yoo AS. 2021. MiR-124 synergism with ELAVL3 enhances target gene expression to promote neuronal maturity. *Proceedings of the National Academy of Sciences of the United States of America* 118(22):e2015454118 DOI 10.1073/pnas.2015454118.
- Lu Y, Okubo T, Rawlins E, Hogan BLM. 2008. Epithelial progenitor cells of the embryonic lung and the role of microRNAs in their proliferation. *Proceedings of the American Thoracic Society* 5:300–304 DOI 10.1513/pats.200710-162DR.
- Lukiw WJ. 2007. Micro-RNA speciation in fetal, adult and Alzheimer's disease hippocampus. *Neuroreport* 18:297–300 DOI 10.1097/WNR.0B013E3280148E8B.
- Lukiw WJ. 2013. Circular RNA (circRNA) in Alzheimer's disease (AD). *Frontiers in Genetics* 4:307 DOI 10.3389/fgene.2013.00307.
- Luo Y, Hoffer A, Hoffer B, Qi X. 2015. Mitochondria: a therapeutic target for Parkinson's disease? *International Journal of Molecular Sciences* 16:20704–20730 DOI 10.3390/ijms160920704.
- Makeyev EV, Zhang J, Carrasco MA, Maniatis T. 2007. The MicroRNA miR-124 promotes neuronal differentiation by triggering brain-specific alternative pre-mRNA splicing. *Molecular Cell* 27:435–448 DOI 10.1016/J.MOLCEL.2007.07.015.
- Maldonado-Lasuncion I, Atienza M, Sanchez-Espinosa MP, Cantero JL. 2019. Aging-related changes in cognition and cortical integrity are associated with serum expression of candidate MicroRNAs for Alzheimer disease. *Cerebral Cortex* 29:4426–4437 DOI 10.1093/CERCOR/BHY323.
- Malhi GS, Coulston CM, Fritz K, Lampe L, Bargh DM, Ablett M, Lyndon B, Sapsford R, Theodoros M, Woolfall D, Van der Zyppe A, Hopwood M, Mitchell AJ. 2014. Unlocking the diagnosis of depression in primary care: which key symptoms are GPs using to determine diagnosis and severity? *The Australian and New Zealand Journal of Psychiatry* 48:542–547 DOI 10.1177/0004867413513342.
- Massone S, Ciarlo E, Vella S, Nizzari M, Florio T, Russo C, Cancedda R, Pagano A. 2012. Biochimica et Biophysica Acta NDM29, a RNA polymerase III-dependent non coding RNA, promotes amyloidogenic processing of APP and amyloid β secretion. *BBA—Molecular Cell Research* 1823:1170–1177 DOI 10.1016/j.bbamcr.2012.05.001.
- Massone S, Vassallo I, Fiorino G, Castelnuovo M, Barbieri F, Borghi R, Tabaton M, Robello M, Gatta E, Russo C, Florio T, Dieci G, Cancedda R, Pagano A. 2011.

- Neurobiology of disease 17A, a novel non-coding RNA, regulates GABA B alternative splicing and signaling in response to inflammatory stimuli and in Alzheimer disease. *Neurobiology of Disease* 41:308–317 DOI 10.1016/j.nbd.2010.09.019.
- McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR, Kawas CH, Klunk WE, Koroshetz WJ, Manly JJ, Mayeux R, Mohs RC, Morris JC, Rossor MN, Scheltens P, Carrillo MC, Thies B, Weintraub S, Phelps CH. 2011.** The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's & Dementia* 7:263–269 DOI 10.1016/J.JALZ.2011.03.005.
- Mehta SL, Dempsey RJ, Vemuganti R. 2020.** Role of circular RNAs in brain development and CNS diseases. *Progress in Neurobiology* 186:101746 DOI 10.1016/J.PNEUROBIO.2020.101746.
- Melton C, Judson RL, Blelloch R. 2010.** Opposing microRNA families regulate self-renewal in mouse embryonic stem cells. *Nature* 463:621–626 DOI 10.1038/NATURE08725.
- Millar JK, James K, Brandon NJ, Thomson PA. 2004.** DISC1 and DISC2: discovering and dissecting molecular mechanisms underlying psychiatric illness. *Annals of Medicine* 36:367–378 DOI 10.1080/07853890410033603.
- Mohs RC, Greig NH. 2017.** Drug discovery and development: role of basic biological research. *Alzheimer's & Dementia* 3:651–657 DOI 10.1016/J.TRCL.2017.10.005.
- Mouradian MM. 2012.** MicroRNAs in Parkinson's disease. *Neurobiology of Disease* 46:279–284 DOI 10.1016/J.NBD.2011.12.046.
- Mourtada-Maarabouni M, Pickard MR, Hedge VL, Farzaneh F, Williams GT. 2009.** GAS5, a non-protein-coding RNA, controls apoptosis and is downregulated in breast cancer. *Oncogene* 28:195–208 DOI 10.1038/onc.2008.373.
- Mus E, Hof PR, Tiedge H. 2007.** Dendritic BC200 RNA in aging and in Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America* 104:10679–10684 DOI 10.1073/pnas.0701532104.
- Nishimoto Y, Nakagawa S, Hirose T, Okano HJ, Takao M, Shibata S, Suyama S, Kuwako KI, Imai T, Murayama S, Suzuki N, Okano H. 2013.** The long non-coding RNA nuclear-enriched abundant transcript 1-2 induces paraspeckle formation in the motor neuron during the early phase of amyotrophic lateral sclerosis. *Molecular Brain* 6:31 DOI 10.1186/1756-6606-6-31.
- Norton S, Matthews FE, Barnes DE, Yaffe K, Brayne C. 2014.** Potential for primary prevention of Alzheimer's disease: an analysis of population-based data. *The Lancet. Neurology* 13:788–794 DOI 10.1016/S1474-4422(14)70136-X.
- Nowak JS, Michlewski G. 2013.** MiRNAs in development and pathogenesis of the nervous system. *Biochemical Society Transactions* 41:815–820 DOI 10.1042/BST20130044.
- O'Connell RM, Chaudhuri AA, Rao DS, Baltimore D. 2009.** Inositol phosphatase SHIP1 is a primary target of miR-155. *Proceedings of the National Academy of Sciences of the United States of America* 106:7113–7118 DOI 10.1073/PNAS.0902636106.
- Ogawa Y, Sun BK, Lee JT. 2008.** Intersection of the RNA interference and X-inactivation pathways. *Science* 320:1336–1341 DOI 10.1126/SCIENCE.1157676.

- Okutsu T, Kuroiwa Y, Kagitani F, Kai M, Aisaka K, Tsutsumi O, Kaneko Y, Yokomori K, Surani MA, Kohda T, Kaneko-Ishino T, Ishino F. 2000.** Expression and imprinting status of human PEG8/IGF2AS, a paternally expressed antisense transcript from the IGF2 locus, in Wilms' tumors. *Journal of Biochemistry* **127**:475–483 DOI [10.1093/oxfordjournals.jbchem.a022630](https://doi.org/10.1093/oxfordjournals.jbchem.a022630).
- Ouyang Q, Liu K, Zhu Q, Deng H, Le Y, Ouyang W, Yan X, Zhou W, Tong J. 2022.** Brain-penetration and neuron-targeting DNA nanoflowers co-delivering miR-124 and rutin for synergistic therapy of Alzheimer's disease. *Small* **18**(14):e2107534 DOI [10.1002/SMLL.202107534](https://doi.org/10.1002/SMLL.202107534).
- Packer AN, Xing Y, Harper SQ, Jones L, Davidson BL. 2008.** The bifunctional microRNA miR-9/miR-9* regulates REST and CoREST and is downregulated in Huntington's disease. *Journal of Neuroscience* **28**:14341–14346 DOI [10.1523/JNEUROSCI.2390-08.2008](https://doi.org/10.1523/JNEUROSCI.2390-08.2008).
- Pandey GK, Mitra S, Subhash S, Hertwig F, Kanduri M, Mishra K, Fransson S, Ganeshram A, Mondal T, Bandaru S, Malin O, Akyu LM, Abrahamsson J, Pfeifer S, Larsson E, Shi L, Peng Z. 2014.** Article the risk-associated long noncoding RNA NBAT-1 controls neuroblastoma progression by regulating cell proliferation and neuronal differentiation. *Cancer Cell* **26**:722–737 DOI [10.1016/j.ccell.2014.09.014](https://doi.org/10.1016/j.ccell.2014.09.014).
- Parenti R, Paratore S, Torrisi A, Cavallaro S. 2007.** A natural antisense transcript against Rad18, specifically expressed in neurons and upregulated during β -amyloid-induced apoptosis. *European Journal of Neuroscience* **26**:2444–2457 DOI [10.1111/j.1460-9568.2007.05864.x](https://doi.org/10.1111/j.1460-9568.2007.05864.x).
- Pobezinsky LA, Wells AC. 2018.** Let's fight cancer: let-7 is a tool to enhance antitumor immune responses. *Future Oncology* **14**:1141–1145 DOI [10.2217/FON-2018-0037](https://doi.org/10.2217/FON-2018-0037).
- Polesskaya OO, Haroutunian V, Davis KL, Hernandez I, Sokolov BP. 2003.** Novel putative nonprotein-coding RNA gene from 11q14 displays decreased expression in brains of patients with schizophrenia. *Journal of Neuroscience Research* **74**:111–122 DOI [10.1002/jnr.10752](https://doi.org/10.1002/jnr.10752).
- Qiu-Lan M, Galasko DR, Ringman JM, Vinters HV, Edland SD, Pomakian J, Ubeda OJ, Rosario ER, Teter B, Frautschy SA, Cole GM. 2009.** Reduction of SorLA/LR11, a sorting protein limiting beta-amyloid production, in Alzheimer disease cerebrospinal fluid. *Archives of Neurology* **66**:448–457 DOI [10.1001/ARCHNEUROL.2009.22](https://doi.org/10.1001/ARCHNEUROL.2009.22).
- Qureshi IA, Mehler MF. 2013.** Long non-coding RNAs: novel targets for nervous system disease diagnosis and therapy. *Neurotherapeutics* **10**:632–646 DOI [10.1007/S13311-013-0199-0](https://doi.org/10.1007/S13311-013-0199-0).
- Rajman M, Metge F, Fiore R, Khudayberdiev S, Aksoy-Aksel A, Bicker S, Ruedell Reschke C, Raof R, Brennan GP, Delanty N, Farrell MA, O'Brien DF, Bauer S, Norwood B, Veno MT, Krüger M, Braun T, Kjems J, Rosenow F, Henshall DC, Dieterich C, Schratt G. 2017.** A microRNA-129-5p/Rbfox crosstalk coordinates homeostatic downscaling of excitatory synapses. *The EMBO Journal* **36**:1770–1787 DOI [10.15252/EMBJ.201695748](https://doi.org/10.15252/EMBJ.201695748).
- Ramachandran V, Chen X. 2008.** Degradation of microRNAs by a family of exoribonucleases in Arabidopsis. *Science* **321**:1490–1492 DOI [10.1126/SCIENCE.1163728](https://doi.org/10.1126/SCIENCE.1163728).

- Ramos AD, Andersen RE, Liu SJ, Nowakowski TJ, Hong SJ, Gertz CC, Salinas RD, Zarabi H, Kriegstein AR, Lim DA. 2015. The long noncoding RNA Pnky regulates neuronal differentiation of embryonic and postnatal neural stem cells. *Cell Stem Cell* 16:439–447 DOI 10.1016/J.STEM.2015.02.007.
- Rani N, Nowakowski TJ, Zhou H, Godshalk SE, Lisi V, Kriegstein AR, Kosik KS. 2016. A primate lncRNA mediates notch signaling during neuronal development by sequestering miRNA. *Neuron* 90:1174–1188 DOI 10.1016/J.NEURON.2016.05.005.
- Reich SG, Savitt JM. 2019. Parkinson's disease. *The Medical Clinics of North America* 103:337–350 DOI 10.1016/J.MCNA.2018.10.014.
- Ripa R, Dolfi L, Terrigno M, Pandolfini L, Savino A, Arcucci V, Groth M, Terzibasi Tozzini E, Baumgart M, Cellerino A. 2017. MicroRNA miR-29 controls a compensatory response to limit neuronal iron accumulation during adult life and aging. *BMC Biology* 15(1):9 DOI 10.1186/S12915-017-0354-X.
- Roberts TC, Morris KV, Wood MJA. 2014. The role of long non-coding RNAs in neurodevelopment, brain function and neurological disease. *Philosophical Transactions of the Royal Society B: Biological Sciences* 369(1652):20130507 DOI 10.1098/RSTB.2013.0507.
- Ruberti F, Barbato C, Cogoni C. 2012. Targeting microRNAs in neurons: tools and perspectives. *Experimental Neurology* 235:419–426 DOI 10.1016/J.EXPNEUROL.2011.10.031.
- Sahoo T, Del Gaudio D, German JR, Shinawi M, Peters SU, Person RE, Garnica A, Cheung SW, Beaudet AL. 2008. Prader-Willi phenotype caused by paternal deficiency for the HBII-85 C/D box small nucleolar RNA cluster. *Nature Genetics* 40:719–721 DOI 10.1038/ng.158.
- Salim S. 2017. Oxidative stress and the central nervous system. *The Journal of Pharmacology and Experimental Therapeutics* 360:201–205 DOI 10.1124/JPET.116.237503.
- Sarkar S, Jun S, Rellick S, Quintana DD, Cavendish JZ, Simpkins JW. 2016. Expression of microRNA-34a in Alzheimer's disease brain targets genes linked to synaptic plasticity, energy metabolism, and resting state network activity. *Brain Research* 1646:139–151 DOI 10.1016/J.BRAINRES.2016.05.026.
- Saxena A, Carninci P. 2011. Long non-coding RNA modifies chromatin: epigenetic silencing by long non-coding RNAs. *BioEssays* 33:830–839 DOI 10.1002/BIES.201100084.
- Scheele C, Petrovic N, Faghihi MA, Lassmann T, Fredriksson K, Rooyackers O, Wahlestedt C, Good L, Timmons JA. 2007. The human PINK1 locus is regulated in vivo by a non-coding natural antisense RNA during modulation of mitochondrial function. *BMC Genomics* 8:74 DOI 10.1186/1471-2164-8-74.
- Schipper HM, Maes OC, Chertkow HM, Wang E. 2007. MicroRNA expression in Alzheimer blood mononuclear cells. *Gene Regulation and Systems Biology* 1:263–274.
- Shi Y, Jin YX. 2009. MicroRNA in cell differentiation and development. *Science in China. Series C, Life Sciences* 52:205–211 DOI 10.1007/S11427-009-0040-5.
- Shioya M, Obayashi S, Tabunoki H, Arima K, Saito Y, Ishida T, Satoh J. 2010. Aberrant microRNA expression in the brains of neurodegenerative diseases miR-29a decreased

- in Alzheimer disease brains targets neurone navigator 3. *Neuropathology and Applied Neurobiology* **36**:320–330 DOI 10.1111/j.1365-2990.2010.01076.x.
- Silbereis JC, Pochareddy S, Zhu Y, Li M, Sestan N. 2016.** The cellular and molecular landscapes of the developing human central nervous system. *Neuron* **89**:248–268 DOI 10.1016/j.neuron.2015.12.008.
- Silva MM, Rodrigues B, Fernandes J, Santos SD, Carreto L, Santos MAS, Pinheiro P, Carvalho AL. 2019.** MicroRNA-186-5p controls GluA2 surface expression and synaptic scaling in hippocampal neurons. *Proceedings of the National Academy of Sciences of the United States of America* **116**:5727–5736 DOI 10.1073/PNAS.1900338116.
- Sparago A, Cerrato F, Vernucci M, Ferrero GB, Silengo MC, Riccio A. 2004.** Microdeletions in the human H19 DMR result in loss of IGF2 imprinting and Beckwith-Wiedemann syndrome. *Nature Genetics* **36**:958–960 DOI 10.1038/ng1410.
- Sueda R, Imayoshi I, Harima Y, Kageyama R. 2019.** High Hes1 expression and resultant Ascl1 suppression regulate quiescent vs. active neural stem cells in the adult mouse brain. *Genes & Development* **33**:511–523 DOI 10.1101/GAD.323196.118.
- Sunwoo JS, Lee ST, Im W, Lee M, Byun JI, Jung KH, Park K II, Jung KY, Lee SK, Chu K, Kim M. 2017.** Altered expression of the long noncoding RNA NEAT1 in Huntington’s disease. *Molecular Neurobiology* **54**:1577–1586 DOI 10.1007/s12035-016-9928-9.
- Ul Islam S, Shehzad A, Bilal Ahmed M, Lee YS. 2020.** Intranasal delivery of nanoformulations: a potential way of treatment for neurological disorders. *Molecules* **25**(8):1929 DOI 10.3390/MOLECULES25081929.
- Urduingio RG, Fernandez AF, Lopez-Nieva P, Rossi S, Huertas D, Kulis M, Liu CG, Croce C, Calin GA, Esteller M. 2010.** Disrupted microRNA expression caused by Mecp2 loss in a mouse model of Rett syndrome. *Epigenetics* **5**:656–663 DOI 10.4161/epi.5.7.13055.
- Valadi H, Ekström K, Bossios A, Sjöstrand M, Lee JJ, Lötvall JO. 2007.** Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nature Cell Biology* **9**:654–659 DOI 10.1038/ncb1596.
- Van den Heuvel MP, Bullmore ET, Sporns O. 2016.** Comparative connectomics. *Trends in Cognitive Sciences* **20**:345–361 DOI 10.1016/J.TICS.2016.03.001.
- Vincent JB, Petek E, Thevarkunnel S, Kolozsvari D, Cheung J, Patel M, Scherer SW. 2002.** The RAY1/ST7 tumor-suppressor locus on chromosome 7q31 represents a complex multi-transcript system. *Genomics* **80**:283–294 DOI 10.1006/geno.2002.6835.
- Walgrave H, Balusu S, Snoeck S, Vanden Eynden E, Craessaerts K, Thrupp N, Wolfs L, Horré K, Fourné Y, Ronisz A, Silajdžić E, Penning A, Tosoni G, Callaerts-Vegh Z, D’Hooge R, Thal DR, Zetterberg H, Thuret S, Fiers M, Frigerio CS, De Strooper B, Salta E. 2021.** Restoring miR-132 expression rescues adult hippocampal neurogenesis and memory deficits in Alzheimer’s disease. *Cell Stem Cell* **28**:1805–1821 DOI 10.1016/J.STEM.2021.05.001.
- Wang B, Zhang C, Du X, Zhang J. 2021.** lncRNA-disease association prediction based on latent factor model and projection. *Scientific Reports* **11**(1):19965 DOI 10.1038/S41598-021-99493-5.

- Wang G, Van der Walt JM, Mayhew G, Li YJ, Züchner S, Scott WK, Martin ER, Vance JM. 2008c. Variation in the miRNA-433 binding site of FGF20 confers risk for Parkinson disease by overexpression of α -Synuclein. *American Journal of Human Genetics* 82:283–289 DOI 10.1016/j.ajhg.2007.09.021.
- Wang S, Aurora AB, Johnson BA, Qi X, McAnally J, Hill JA, Richardson JA, Bassel-Duby R, Olson EN. 2008a. The endothelial-specific microRNA miR-126 governs vascular integrity and angiogenesis. *Developmental Cell* 15:261–271 DOI 10.1016/j.DEVCEL.2008.07.002.
- Wang WX, Rajeev BW, Stromberg AJ, Ren N, Tang G, Huang Q, Rigoutsos I, Nelson PT. 2008b. The expression of microRNA miR-107 decreases early in Alzheimer's disease and may accelerate disease progression through regulation of beta-site amyloid precursor protein-cleaving enzyme 1. *The Journal of Neuroscience* 28:1213–1223 DOI 10.1523/JNEUROSCI.5065-07.2008.
- Wevrick R, Francke U. 1997. An imprinted mouse transcript homologous to the human imprinted in Prader-Willi syndrome (IPW) gene. *Human Molecular Genetics* 6:325–332 DOI 10.1093/hmg/6.2.325.
- Wingo TS, Yang J, Fan W, Min Canon S, Gerasimov ES, Lori A, Logsdon B, Yao B, Seyfried NT, Lah JJ, Levey AI, Boyle PA, Schneider JA, De Jager PL, Bennett DA, Wingo AP. 2020. Brain microRNAs associated with late-life depressive symptoms are also associated with cognitive trajectory and dementia. *NPJ Genomic Medicine* 5:6 DOI 10.1038/S41525-019-0113-8.
- World Alzheimer Report. 2019. Alzheimer's Disease International (ADI). Available at <https://www.alzint.org/resource/world-alzheimer-report-2019/> (accessed on 08 July 2022).
- Wu H, Tao J, Chen PJ, Shahab A, Ge W, Hart RP, Ruan X, Ruan Y, Sun YE. 2010. Genome-wide analysis reveals methyl-CpG-binding protein 2-dependent regulation of microRNAs in a mouse model of Rett syndrome. *Proceedings of the National Academy of Sciences of the United States of America* 107:18161–18166 DOI 10.1073/pnas.1005595107.
- Xie HR, Sen HL, Li GY. 2010. SH-SY5Y human neuroblastoma cell line: in vitro cell model of dopaminergic neurons in Parkinson's disease. *Chinese Medical Journal* 123:1086–1092 DOI 10.3760/cma.j.issn.0366-6999.2010.08.021.
- Xiomerisiou G, Dardiotis E, Tsimourtou V, Kounta PM, Paterakis KN, Kapsalaki EZ, Fountas KN, Hadjigeorgiou GM. 2010. Genetic basis of Parkinson disease. *Neurosurgical Focus* 28:1–7 DOI 10.3171/2009.10.FOCUS09220.
- Xu J, Jing WK, Jun JQ, Feng DX. 2020. Roles of miRNA and lncRNA in triple-negative breast cancer. *Journal of Zhejiang University. Science. B* 21:673–689 DOI 10.1631/JZUS.B1900709.
- Yan W, Chen ZY, Chen JQ, Chen HM. 2018. LncRNA NEAT1 promotes autophagy in MPTP-induced Parkinson's disease through stabilizing PINK1 protein. *Biochemical and Biophysical Research Communications* 496:1019–1024 DOI 10.1016/j.bbrc.2017.12.149.

- Yang C, Li Z, Li Y, Xu R, Wang Y, Tian Y, Chen W. 2017. Long non-coding RNA NEAT1 overexpression is associated with poor prognosis in cancer patients: a systematic review and meta-analysis. *Oncotarget* 8:2672–2680 DOI [10.18632/oncotarget.13737](https://doi.org/10.18632/oncotarget.13737).
- Yao D, Zhan X, Zhan X, Kwok CK, Li P, Wang J. 2020. A random forest based computational model for predicting novel lncRNA-disease associations. *BMC Bioinformatics* 21(1):126 DOI [10.1186/S12859-020-3458-1](https://doi.org/10.1186/S12859-020-3458-1).
- Yoo AS, Stahl BT, Chen L, Crabtree GR. 2009. MicroRNA-mediated switching of chromatin-remodelling complexes in neural development. *Nature* 460:642–646 DOI [10.1038/NATURE08139](https://doi.org/10.1038/NATURE08139).
- Zalfa F, Adinolfi S, Napoli I, Kühn-Hölsken E, Urlaub H, Achsel T, Pastore A, Bagni C. 2005. Fragile X mental retardation protein (FMRP) binds specifically to the brain cytoplasmic RNAs BC1/BC200 via a novel RNA-binding motif. *Journal of Biological Chemistry* 280:33403–33410 DOI [10.1074/jbc.M504286200](https://doi.org/10.1074/jbc.M504286200).
- Zhang L, Xia R, Jia J, Wang L, Li K, Li Y, Zhang J. 2018b. Oleanolic acid protects against cognitive decline and neuroinflammation-mediated neurotoxicity by blocking secretory phospholipase A2 IIA-activated calcium signals. *Molecular Immunology* 99:95–103 DOI [10.1016/J.MOLIMM.2018.04.015](https://doi.org/10.1016/J.MOLIMM.2018.04.015).
- Zhang W, Wan H, Feng G, Qu J, Wang J, Jing Y, Ren R, Liu Z, Zhang L, Chen Z, Wang S, Zhao Y, Wang Z, Yuan Y, Zhou Q, Li W, Liu GH, Hu B. 2018a. SIRT6 deficiency results in developmental retardation in cynomolgus monkeys. *Nature* 560:661–665 DOI [10.1038/S41586-018-0437-Z](https://doi.org/10.1038/S41586-018-0437-Z).
- Zheng K, Hu F, Zhou Y, Zhang J, Zheng J, Lai C, Xiong W, Cui K, Hu YZ, Han ZT, Zhang HH, Chen JG, Man HY, Liu D, Lu Y, Zhu LQ. 2021. miR-135a-5p mediates memory and synaptic impairments via the Rock2/Adducin1 signaling pathway in a mouse model of Alzheimer's disease. *Nature Communications* 12(1):1903 DOI [10.1038/S41467-021-22196-Y](https://doi.org/10.1038/S41467-021-22196-Y).
- Zhu JJ, Fu HJ, Wu YG, Zheng XF. 2013. Function of lncRNAs and approaches to lncRNA-protein interactions. *Science China. Life Sciences* 56:876–885 DOI [10.1007/S11427-013-4553-6](https://doi.org/10.1007/S11427-013-4553-6).
- Zu T, Liu Y, Bañez Coronel M, Reid T, Pletnikova O, Lewis J, Miller TM, Harms MB, Falchook AE, Subramony SH, Ostrow LW, Rothstein JD, Troncoso JC, Ranum LPW. 2013. RAN proteins and RNA foci from antisense transcripts in C9ORF72 ALS and frontotemporal dementia. *Proceedings of the National Academy of Sciences of the United States of America* 110:E4968–77 DOI [10.1073/pnas.1315438110](https://doi.org/10.1073/pnas.1315438110).
- Zuccato C, Tartari M, Crotti A, Goffredo D, Valenza M, Conti L, Cataudella T, Leavitt BR, Hayden MR, Timmusk T, Rigamonti D, Cattaneo E. 2003. Huntingtin interacts with REST/NRSF to modulate the transcription of NRSE-controlled neuronal genes. *Nature Genetics* 35:76–83 DOI [10.1038/ng1219](https://doi.org/10.1038/ng1219).