

Whole-genome sequence and genesis of an avian influenza virus H5N1 isolated from a healthy chicken in a live bird market in Indonesia: Accumulation of mammalian adaptation markers in avian hosts

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Background. Influenza A viruses are a major pathogen that causes significant clinical and economic harm to many animals. In Indonesia, the highly pathogenic avian influenza (HPAI) H5N1 virus has been endemic in poultry since 2003 and has caused sporadic deadly infections in humans. The genetic bases that determine host range have not yet been fully elucidated. We analyzed whole-genome sequence of a recent H5 isolate to reveal the evolution toward its mammalian adaptation. **Methods.** We determined the whole-genome sequence of A/chicken/East Java/Av1955/2022 (hereafter, “Av1955”) from a healthy chicken in April 2022 and conducted phylogenetic and mutational analysis. **Results.** Phylogenetic analysis revealed that Av1955 belonged to the H5N1 clade 2.3.2.1c (Eurasian lineage). The six gene segments (PB1, PB2, HA, NP, NA, and NS) out of the eight segments derived from viruses of H5N1 Eurasian lineage, one (PB2) from the H3N6 subtype and the remaining one (M) from H5N1 clade 2.1.3.2b (Indonesian lineage). The donor of the PB2 segment was a reassortant among three viruses of H5N1 Eurasian and Indonesian lineages and the H3N6 subtype. The HA amino acid sequence contained multiple basic amino acids at the cleavage site. Mutation analysis revealed that Av1955 possessed the maximal number of mammalian adaptation marker mutations. **Conclusions.** Av1955 was a virus of H5N1 Eurasian lineage. The HA protein contains an HPAI H5N1-type cleavage site sequence, while the virus was isolated from a healthy chicken suggesting its low pathogenicity nature. The virus has increased mammalian adaptation markers by mutation and intra- and inter-subtype reassortment, gathering gene segments possessing the most abundant maker mutations among previously circulating viruses. The increasing

mammalian adaptation mutation in avian hosts suggests that they might be adaptive to infection in mammalian and avian hosts. It highlights the importance of genomic surveillance and adequate control measures for H5N1 infection in live poultry markets.

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28 Abstract**29 Background.**

30

31 Influenza A viruses are a major pathogen that causes significant clinical and economic harm to
32 many animals. In Indonesia, the highly pathogenic avian influenza (HPAI) H5N1 virus has been
33 endemic in poultry since 2003 and has caused sporadic deadly infections in humans. The genetic
34 bases that determine host range have not yet been fully elucidated. We analyzed whole-genome
35 sequence of a recent H5 isolate to reveal the evolution toward its mammalian adaptation.

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37 Methods.

38

39 We determined the whole-genome sequence of A/chicken/East Java/Av1955/2022 (hereafter,
40 “Av1955”) from a healthy chicken in April 2022 and conducted phylogenetic and mutational
41 analysis.

42

43 Results.

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45 Phylogenetic analysis revealed that Av1955 belonged to the H5N1 clade 2.3.2.1c (Eurasian
46 lineage). The six gene segments (PB1, PB2, HA, NP, NA, and NS) out of the eight segments
47 derived from viruses of H5N1 Eurasian lineage, one (PB2) from the H3N6 subtype and the
48 remaining one (M) from H5N1 clade 2.1.3.2b (Indonesian lineage). The donor of the PB2
49 segment was a reassortant among three viruses of H5N1 Eurasian and Indonesian lineages and
50 the H3N6 subtype. The HA amino acid sequence contained multiple basic amino acids at the
51 cleavage site. Mutation analysis revealed that Av1955 possessed the maximal number of
52 mammalian adaptation marker mutations.

53

54 Conclusions.

55

56 Av1955 was a virus of H5N1 Eurasian lineage. The HA protein contains an HPAI H5N1-type
57 cleavage site sequence, while the virus was isolated from a healthy chicken suggesting its low
58 pathogenicity nature. The virus has increased mammalian adaptation markers by mutation and
59 intra- and inter-subtype reassortment, gathering gene segments possessing the most abundant
60 marker mutations among previously circulating viruses. The increasing mammalian adaptation
61 mutation in avian hosts suggests that they might be adaptive to infection in mammalian and
62 avian hosts. It highlights the importance of genomic surveillance and adequate control measures
63 for H5N1 infection in live poultry markets.

64

65 Keywords:

66 Highly pathogenic avian influenza H5N1, Clade 2.3.2.1c Eurasian lineage, Low pathogenic
67 avian influenza H3N6, Whole-genome sequence, Human health, Inter-subtype reassortment,
68 Intra-subtype reassortment, Mammalian adaptation marker mutation

69

70 Introduction

71 Influenza A viruses are a major pathogen that causes significant clinical and economic harm to
72 various species, including poultry, pigs, horses, marine mammals, and humans (Peiris et al.,
73 2007; Webster et al., 1992). The surface antigenicity of the virus particles divides them into 18
74 hemagglutinin (HA) (H1 to H18) and 11 neuraminidase (NA) (N1 to N11) subtypes (Tong et al.,
75 2013). Highly pathogenic avian influenza (HPAI) H5N1 viruses are capable of sporadic human
76 infection. They have the potential to cause severe sickness with a high case fatality rate among
77 people who have been hospitalized and proven to have the virus. To date, there have been 861
78 confirmed disease cases in people, and 455 have led to death (WHO, 2021). Indonesia reported
79 200 confirmed human cases and 168 fatalities. The 168 fatalities account for over one-third of all
80 deaths among all affected countries. However, Indonesia had only two cases in 2014 and 2015,
81 zero in 2016, and one in 2017, the last.

82
83 The HPAI H5N1 subtype was first discovered in Indonesia in 2003 (Li et al., 2004),
84 spreading to numerous regions and killing over 16 million chickens by the end of 2007 (Lam et
85 al., 2008; Putri et al., 2019). The sequence of the hemagglutinin gene classified it into clade 2.1.
86 This clade subsequently branched into clades 2.1.1 to 2.1.3 in Indonesia (WHO/OIE/FAO/H5N1,
87 2008). Clade 2.1.3 further branched into clades 2.1.3.1 to 2.1.3.3; then clade 2.1.3.2 into clades
88 2.1.3.2a and b (WHO/OIE/FAO/H5N1, 2014). Clade 2.1.1 viruses were predominantly isolated
89 from HPAI-infected chickens during outbreaks between 2003 and 2005. Clades 2.1.2 and 2.1.3
90 were isolated from birds between 2003 and 2005 and humans in 2005. Clade 2.1.3.2, a branch of
91 clade 2.1.3, became prevalent in poultry and humans in 2007. Clade 2.1.3.2b, a descendant of
92 clade 2.1.3.2, became prevalent from 2010 to 2012.

93
94 An incursion of a new HPAI H5N1 clade, clade 2.3.2.1c, from the mainland of Southeast
95 Asia occurred in Indonesia in 2012 (Dharmayanti et al., 2014). The clade became prevalent as
96 early as 2013. Shimizu et al. (2016) reported the isolations of three avian influenza viruses in
97 East Java, Indonesia: H5N1 clade 2.3.2.1c (Av154) from an HPAI outbreak in a turkey farm in
98 2013, H5N1 clade 2.1.3.2b (Av240) from an ill chicken at a live poultry market in 2014, and
99 H3N6 (Av39) from a mildly ill duck at a live poultry market in 2013. Clade 2.3.2.1c was
100 designated as Eurasian lineage, while clade 2.1.3.2b was designated as Indonesian lineage based
101 on the place of emergence.

102
103 Clade 2.3.2.1c caused only one fatal infection in humans in Indonesia, the last case in 2017.
104 The genetic bases that determine host range have not yet been fully elucidated. Influenza virus
105 evolves by the mutation of the genes and by the reassortment of the gene segments. Mertens et
106 al. (2013) identified 149 phenotypic marker mutations related to host tropism or increased
107 pathogenicity in mammals from previously reported literature. Recently Rehman et al. (2022)
108 isolated an avian influenza H5N1 virus, A/chicken/East Java/Av1955/2022 (hereafter,
109 "Av1955"), from a healthy chicken at a live poultry market in East Java, Indonesia. In this study,
110 we determined the whole-genome sequence to identify the clade/lineage and some characteristic
111 features of the amino acid sequences of the viral proteins. We compared the phenotypic marker
112 mutations between Av1955 and its ancestral viruses to reveal the evolution toward its
113 mammalian adaptation.

114

115 **Materials and Methods**

116 **Ethical approval**

117 The Animal Care and Use Committee at the Faculty of Veterinary Medicine, Universitas
118 Airlangga, Surabaya, Indonesia gave their approval for every step of this study (Approval no.
119 1.KE.028.03.2021).

120

121 **Virus**

122

123 Saifur et al. (2022) isolated an avian influenza A H5 virus, Av1955 (A/chicken/East
124 Java/Av1955/2022), from a tracheal swab sample collected from a healthy chicken at a live
125 poultry market in East Java, Indonesia, on March 5, 2022. For this research, they provided a one-
126 mL aliquot of an allantoic fluid from an infected egg. The harvest was positive for the
127 hemagglutination test with chicken red blood cells. It was also positive for 1-step TaqMan real-
128 time RT-PCR tests targeting the influenza A virus M gene and H5 HA gene using sets of primers
129 and probes as previously described (Shimizu et al., 2016) (data, not shown). All procedures were
130 performed in the BSL3 laboratory of the Institute of Tropical Disease, Airlangga University.

131

132 **Whole-genome sequencing**

133

134 For genome analysis by next-generation sequencing, total RNA was isolated from the allantoic
135 harvest using a QIAamp viral Minikit (Qiagen, Tokyo, Japan). Linear polyacrylamide was used
136 as a transporter rather than tRNA. A TruSeq RNA sample preparation kit version 2 was utilized
137 to compile an RNA library (Illumina, Japan). The library was loaded in the flow cell of the 300-
138 cycle MiSeq reagent kit version 2 (Illumina, USA). A MiSeq system (Illumina) sequenced the
139 barcode-labeled multiplex library, which had two runs of 150 bp each. The system created the
140 FASTQ files of sequence reads removing the primer and adaptor sequences. For analysis, the
141 files were imported into CLC Genomics Workbench version 8.1 (CLC bio, Japan). The reads
142 were mapped to the genomes of 27 reference viruses of influenza type A virus, including all
143 subtypes of HA (H1 to H18) and NA (N1 to N11). Tentative complete 8-segment genome
144 sequences were constructed from the assembled consensus with the best coverage and common
145 sequences of type A influenza viruses at the 5' end (12 nucleotides [nt]) and 3' end (13 nt) of the
146 genome segments. The reads were mapped again to the tentative complete genome sequence to
147 assemble Av1955 consensus sequences. The assembled sequences covered 97.4 to 99.5% of the
148 complete genome-segments mostly lacking the 5' and 3' consensus sequences. The sequences of
149 the 8 genome-segments were submitted to the GISAID database with isolate ID:
150 EPI_ISL_13690275.

151

152 **Genetic analysis**

153

154 BLAST analysis in the GISAID EpiFlu database (<https://gisaid.org>) on 10 August 2022
155 identified viruses with the highest identity to the nucleotide sequence of one or more gene
156 segments of Av1955.

157

158 The genetic information processing software Genetyx v14 (Genetyx Co., Tokyo, Japan)
159 generated the phylogenetic trees using the RAxML with 100 bootstrap replicates. The trees are
160 rooted in the A/South Carolina/1/1918(H1N1) (HA: GenBank AF117241) or A/Brevig
161 Mission/1/1918(H1N1) (NA: AF250356, PB2: DQ208309, PB1: DQ208310, PA: DQ208311,
162 NP: AY744935, M: AY130766, NS: AF333238).

163

164 The amino acid sequences of the viral proteins were decoded from the genome nucleotide
165 sequences and analyzed for nonsynonymous mutations. The putative host adaptation mutations
166 were analyzed according to the evaluation of phenotypic markers described by Mertens et al.
167 (2013).

168

169 **Isolate ID (EPI_ISL) in the GISAID database of viruses used as a reference**

170 A/duck/East Java/Av39/2013 (H3N6) (hereafter, Av39): EPI_ISL_307026, A/chicken/East
171 Java/Av240/2014 (H5N1 Indonesian lineage) (HA clade 2.1.3.2b) (hereafter, Av240):
172 EPI_ISL_307019, A/turkey/East Java/Av154/2013 (H5N1 Eurasian lineage) (HA clade 2.3.2.1c)
173 (hereafter, Av154): EPI_ISL_307002, A/eagle/Jakarta Timur/20616-206-III/2016 (H5N1)
174 (hereafter, Eg2061): EPI_ISL_266799, A/chicken/East Java/Av1210/2017 (H5N1) (hereafter,
175 Av1210): EPI_ISL_401823, A/chicken/East Java/Spg119/2018 (H5N1) (hereafter, Spg119):
176 EPI_ISL_365528, and A/chicken/East Java/Av1534/2019 (H5N1) (hereafter, Av1534):
177 EPI_ISL_401824

178

179 **Results**

180 **HA and NA subtypes of Av1955**

181 The BLAST analysis revealed that the HA sequence of Av1955 was most close to Spg119 of
182 H5N1 (the sharing identity, 97.7%) and the NA to Av1210 of H5N1 (97.3%) among the viruses
183 in the GISAD database (Table 1). The HA and NA segments were most close to Av154 of H5N1
184 Eurasian lineage (HA clade 2.3.2.1c) (95.1% and 96.2%, respectively) among the three isolates
185 (Av39, Av240, and Av154) during 2013-2014 in East Java (Table 1). In the phylogenies, the HA
186 and NA segments are on the same branch with Av154 (Figure 1). These results clearly indicate
187 that Av1955 is a virus of the H5N1 Eurasian lineage (HA clade 2.3.2.1c).

188

189 **Genesis of Av1955**

190 The BLAST analysis revealed that the genome sequence of Av1955 was most close to Spg119
191 for the PB2 segment (the sharing identity, 98.0%), HA (97.7%), NP (98.3%), and NS (98.5%); to
192 Av1534 for the PB1 (97.9%) and PA (98.1%); to Av1210 for the NA (97.3%); and to Eg2061 for
193 the M (98.9%) among the viruses in the GISAD database (Table 1). These results indicated that
194 Av1955 acquired the PB2, HA, NP, and NS segments from an Spg119-like virus; the PB1 and
195 PA from an Av1534-like virus; the NA from an Av1210-like virus; and the M from an Eg2061-
196 like virus. These four putative donors of the segments are H5N1 Eurasian lineage since all HA

197 and NA closely cluster with Av154 of H5N1 Eurasian lineage in the phylogenies of HA and NA
198 (Figure 1).

199
200 Av1955 was most close to Av39 for the PB2 (93.2%); to Av154 for the PB1 (95.9%), PA
201 (96.0%), HA (95.2%), NP (97.0%), NA (96.2%), and NS (97.2%); and Av240 for the M (98.1%)
202 among the three isolates in East Java during 2013-2014. In the phylogenies, the PB2 of Av1955
203 closely clusters with Spg119 and Av39; all of the PB1, PA, HA, NP, and NA cluster with
204 Eg2061, Av1210, Spg119, Av1534, and Av154; the M with Eg2061, Spg119, and Av154; and
205 the NS with Av1210, Spg119, Av1534, and Av154.

206
207 The PB2, M, and NS of Eg2061 closely cluster with Av240, while the other six segments with
208 Av154, indicating that Eg2061 is a reassortant between Av240 and Av154; Eg2061 acquired
209 the PB2, M and NS segments from an Av240-like virus, and the other six segments from an
210 Av154-like virus.

211
212 The PB2 and M of Spg119 closely cluster with Av39 and Av240, respectively, while the other
213 six segments with Av154, indicating that Spg119 is a triple reassortant between Av39, Av240,
214 and Av154; Spg119 acquired the PB2 segment from an Av39-like virus, the M segment from an
215 Av240-like virus, and the other six segments from an Av154-like virus.

216
217 From the described results, we proposed a model of the genesis of Av1955 by multi-steps of
218 inter- and intra-subtype reassortment, as illustrated in Figure 2.

219

220 **Characteristic amino acid sequences of Av1955**

221 Table 2 summarizes the findings of an analysis of Av1955 amino acid sequences of the receptor
222 binding, cleavage, and glycosylation sites in HA, deletions in NA and NS1, truncation of PB1-
223 F2, and amantadine and rimantadine resistant mutations in M2. At the receptor binding region of
224 the HA protein, the amino acid sequences were E-186, G-221, Q-222, and G-224 (H5
225 numbering), which are consistent with the avian type specifically binding 2,3-linked sialic acid
226 receptor. There were no mutations in this region among the seven reference viruses. The amino
227 acid sequence at the cleavage site of the HA protein was PQRE-RRRKR for Av1955, Av154,
228 and the four donor strains and was PQRESRRKKR for Av240; all of these possessed five
229 consecutive basic amino acid residues as typical highly pathogenic avian influenza A viruses.
230 Av39 had PEKQT---R at the site, only one basic amino acid residue R, as specific low
231 pathogenic viruses. A glycosylation site, NST, was present at 154-156 in Av240, while the
232 particular sequence was absent in seven other viruses, including Av1955 as a result of
233 substitution(s) to GST for Av39; DNA for Av154; NNA for Eg2061, Av1210, Spg119, and
234 Av1534; and CNA for Av1955.

235

236 There were deletions in NA at 49-68 and NS at 80-84 with Av1955 and all reference viruses
237 except Av39. The Av1955 PB1-F2 was truncated at 25, while that of the reference viruses,
238 except for Av39 and Av240, was truncated at 57. Av39 and Av240 possessed 87 open reading
239 frames for PB1-F2, almost full of 90. The M2 of Av1955 was a resistant type to amantadine and
240 rimantadine.

241

242 Mammalian adaptation markers in the amino acid sequences of Av1955

243 Martens et al. (2013) identified 149 phenotypic marker mutations related to the host tropism or
244 increased pathogenicity in mammals from previously reported literature. They distribute to PB2
245 (34), PB1 (16), PB1-F2 (1), PA (18), HA (38), NP (10), NA (2), M1 (6), M2 (4), NS1 (16), and
246 NS2 (4). We analyzed the presence or absence of these mutations in Av1955 and seven reference
247 viruses. Table 3 summarizes the results showing the substitutions positive for at least one of the
248 eight viruses. There were no marker mutations in NP and NA proteins. Av1955 had 11 markers
249 in PB2, while the donor of Spg119 had 10. The increase indicates that the virus acquired one
250 marker mutation in the PB2 (R318K) by a substitution mutation. A marker increase also
251 occurred with NS1 (7 to 8 by P215T).

252
253 The markers of the donor segments were the same number as the recipient, Av1955, with PB1
254 (8), PA (3), HA (5), M1 (3), M2 (0), and NS2 (1). The markers in the transferred proteins were
255 the maximum among the four donor viruses except for the HA. The HA donor, Spg119, had 5
256 marker mutations in HA, while Av1210 and Av1534 had 6 and 7, respectively. Av1955 acquired
257 39 mammalian adaptation markers in its proteins, an 8% increase from 36 of Spg119 (the donor
258 of 3 genome segments).

259 260 Discussion

261 Sequencing and phylogenetic analysis (Figure 1) identified the HA and NA of Av1955
262 (A/chicken/East Java/Av1955/2022) as an H5N1 subtype of a Eurasian lineage (HA clade
263 2.3.2.1c). However, Av1955 possesses the PB2 and M genome segments derived from viruses of
264 the H3N6 subtype and H5N1 Indonesian lineage, respectively. The phylogenies indicated that
265 the donor of the PB2 segment, Spg119, was a triple-reassortant among H3N6 and H5N1
266 Indonesian and Eurasian lineages. The donor of the M segment, Eg20616, was a double-
267 reassortant between H5N1 Indonesian and Eurasian lineages. The genesis of Av1955 might
268 involve two steps of genome-segment reassortment (Figure 2). It is worth noting that the M
269 genome segment of the Indonesian lineage has remained in the virus of Eurasian lineage after the
270 7-year disappearance of the H5N1 Indonesian lineage; the last isolate was in 2015 in East Java
271 (A Nastri, 2019, unpublished data).

272
273 There were no mammalian adaptation mutations in the receptor binding region of the HA
274 protein among the seven reference viruses and Av1955. Their amino acid sequences were E-186,
275 G-221, Q-222, and G-224 (H5 numbering), consistent with the avian type specifically binding to
276 a 2,3-linked sialic acid receptor on the cell surface of avian hosts. If the mutations occur, viruses
277 are unable to infect avian hosts. The HA protein of Av1955 contains an HPAI H5N1-type
278 cleavage site sequence with multiple basic amino acid residues (PQRERRRKR). In contrast, the
279 virus was isolated from a healthy chicken suggesting its low pathogenicity nature. Av1955 lost a
280 glycosylation site at 154-156 on the HA protein; the loss has been reported to increase
281 pathogenicity in mice (Suguitan et al., 2012). Av1955 has deletions at 49-68 on the NA and 80-
282 84 on the NS1, both of which have been shown to increase virulence in mice (Matsuoka et al.,
283 2009; Zhou et al., 2009; Long et al., 2008). The Av1955 PB1-F2 truncated at 25 of the total
284 length, 90. The deletion of the main body has been related to increased virulence in mammals
285 (Mettier et al., 2021). Av1955 has amino acid residues of A-27 and N-31 of the M2 protein,

286 which are known to render the phenotype of resistance to amantadine and rimantadine (Belshe et
287 al., 1988; Lan et al., 2010; Pinto et al., 1992). The M genome segment originated from the
288 Av240-like virus of the H5N1 Indonesian lineage (HA clade 2.1.3.2b). Av240 had already
289 acquired the resistant mutations, V27A and N31N, while one of the ancestors of Indonesian
290 lineage, A/Indonesia/5/2005 (H5N1) (HA clade 2.1.3.2), had not yet. It was reported that these
291 mutations occurred during 2003-2008 in human and avian infections in Indonesia (Dharmayanti
292 et al., 2010). Since amantadine had been subscribed to influenza patients but never used for
293 poultry in Indonesia, the avian viruses possessing the M segment with resistant mutations might
294 come from human-to-avian transmission.

295

296 Av1955 possesses 39 mammalian adaptation marker mutations out of 149 identified by
297 Martens et al. (2013), while the four viruses of the genome-segment donor have 34-36 and the
298 three ancestral viruses isolated during 2013-2014 have 27-34. The virus has increased the
299 number of the markers by substitution mutations and intra- and inter-subtype reassortment
300 gathering gene segments possessing the most abundant marker mutations among circulating
301 viruses. The increasing mammalian adaptation mutation in avian hosts suggests that they might
302 be adaptive to infection in mammalian and avian hosts. It highlights the importance of genomic
303 surveillance and adequate control measures for H5N1 infection in live poultry markets.

304

305 **Conclusions.**

306 Av1955 was a virus of the H5N1 Eurasian lineage (HA clade 2.3.2.1c). The virus has increased
307 mammalian adaptation markers by substitution mutation and intra- and inter-subtype
308 reassortment, gathering gene segments possessing the most abundant marker mutations among
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313

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319

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450
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Figure 1

Phylogenetic analysis of the eight gene segments of Av1955

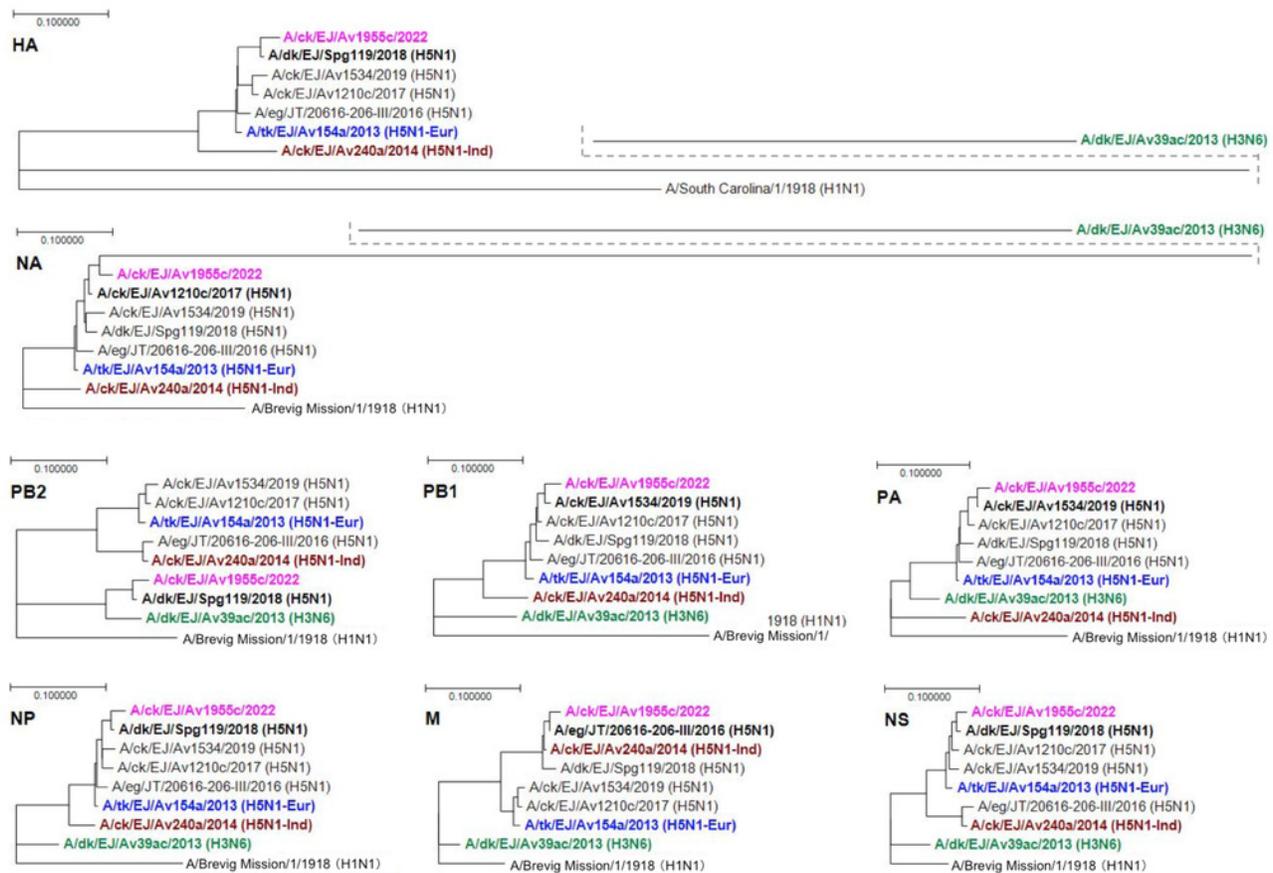


Figure 1:
Phylogenetic analysis of the eight gene segments of Av1955.

The genetic information processing software Genetyx v14 (Genetyx Co., Tokyo, Japan) generated the phylogenetic trees using the RAxML with 100 bootstrap replicates. The tree is rooted in the A/Brevig Mission /1/1918 or A/South Carolina/1/1918 (H1N1). Four viruses with the highest identity to the nucleotide sequence of one or more gene segments of A/ck/EJ/Av1955/2022 (bold and pink color) were included in the phylogenies; these were A/eg/JT/2061-206-III/2016 (H5N1), A/ck/EJ/Av1210c/2017 (H5N1), A/ck/EJ/Spg119/2018 (H5N1) and A/ck/EJ/Av1534/2019 (H5N1). Three viruses isolated from poultry in East Java in 2013-2014 were also included; A/dk/EJ/Av39ac/2013 (H3N6) (bold and green), A/ck/EJ/Av240/2014 (H5N1-Ind) (bold and brown), and A/tk/EJ/Av154/2013 (H5N1-Eur). (bold and blue). Bold and black highlighted the virus at the nearest position to A/ck/EJ/Av1955c/2022 in each of the eight genome segments. “ck”: chicken, “eg”: eagle, “dk”: duck, “tk”: turkey, “EJ”: East Java, “JT”: Jakarta Timur, “-Ind”: Indonesian lineage (HA clade 2.1.3.2b), and “-Eur”: Eurasian lineage (HA clade 2.3.2.1c). Isolate ID (EPI_ISL) in the GISAID database of the viruses was noted in Materials and Methods.

Figure 2

The genesis of Av1955 by multiple reassortments

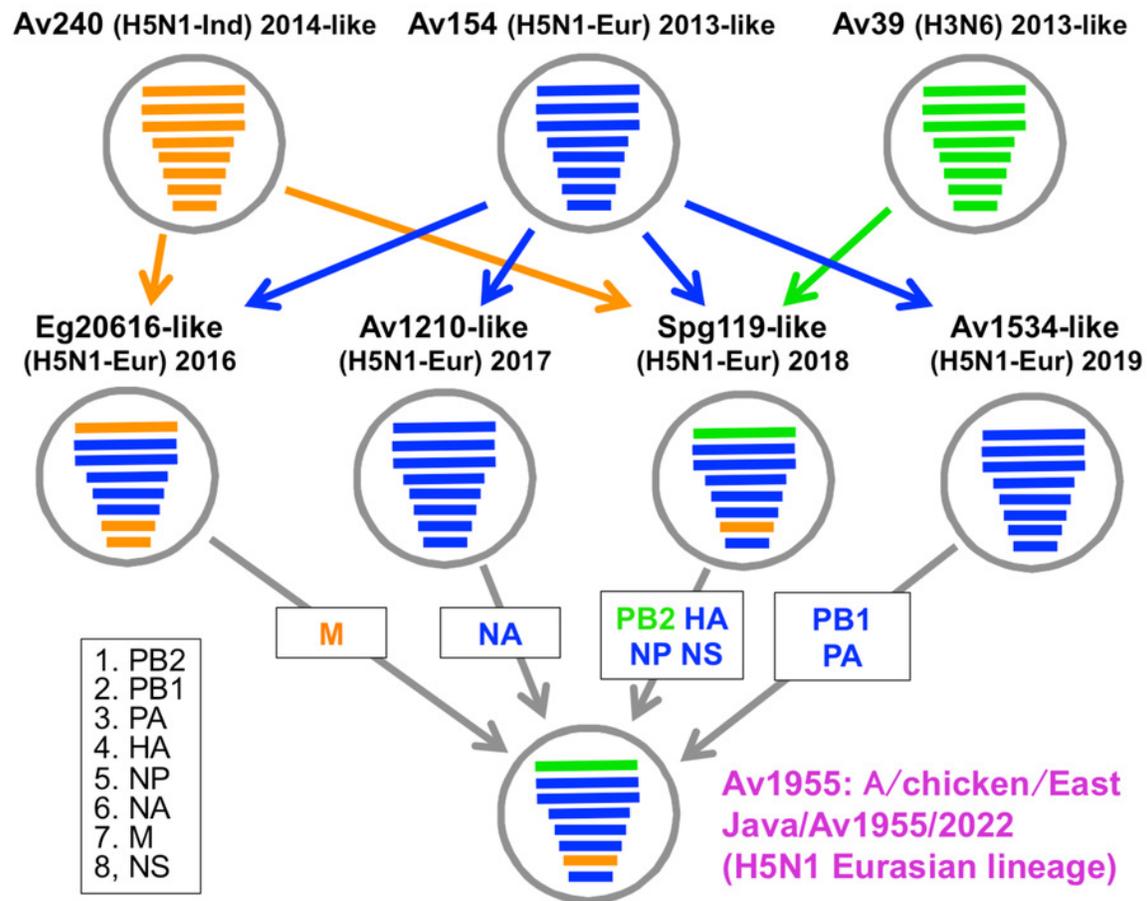


Figure 2:

The genesis of Av1955 by multiple reassortments.

Av39: A/duck/East Java/Av39/2013 (H3N6), **Av240:** A/chicken/East Java/Av240/2014 (H5N1 Indonesian lineage), **Av154:** A/turkey/East Java/Av154/2013 (H5N1 Eurasian lineage), **Eg2061:** A/eagle/Jakarta Timur/20616-206-III/2016 (H5N1 Eurasian lineage), **Av1210:** A/chicken/East Java/Av1210/2017 (H5N1 Eurasian lineage), **Spg119:** A/chicken/East Java/Spg119/2018 (H5N1 Eurasian lineage), and **Av1534:** A/chicken/East Java/Av1534/2019 (H5N1 Eurasian lineage). Isolate ID (EPI_ISL) in the GISAID database of the viruses was noted in Materials and Methods.

Table 1 (on next page)

Viruses with the highest identity to the nucleotide sequence of one or more gene segments of Av1955

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| Genome segment | Identity to the eight genome segments of Av1955, % | | | | | | | |
|----------------|--|-------------------|-------------------|-------------------------------------|--------------------|--------------------|--------------------|--------|
| | Isolates in 2013-2014* | | | Viruses with the highest identity** | | | | Av1955 |
| | Av39 H3N6 | Av240 H5N1-Ind | Av154 H5N1-Eur | Eg2061 H5N1 | Av1210 H5N1 | Spg119 H5N1 | Av1534 H5N1 | |
| PB2 | 93.2 | 85.2 | 85.3 | 85.2 | 85.2 | <u>98.0</u> | 85.0 | 100.0 |
| PB1 | 87.6 | 91.3 | 95.9 | 96.0 | 97.4 | 96.7 | <u>97.9</u> | 100.0 |
| PA | 90.9 | 87.6 | 96.0 | 95.3 | 96.8 | 95.4 | <u>98.2</u> | 100.0 |
| HA | 53.8 | 87.0 | 95.2 | 94.6 | 94.4 | <u>97.7</u> | 93.8 | 100.0 |
| NP | 90.3 | 93.5 | 97.0 | 96.7 | 96.6 | <u>98.3</u> | 96.6 | 100.0 |
| NA | 54.0 | 87.7 | 96.2 | 95.0 | <u>97.3</u> | 96.0 | 95.5 | 100.0 |
| M | 90.4 | 98.1 | 93.8 | <u>98.9</u> | 93.3 | 97.0 | 93.4 | 100.0 |
| NS | 88.8 | 93.0 | 97.2 | 92.4 | 97.9 | <u>98.5</u> | 97.2 | 100.0 |

4 * "Bold" indicates the highest identity to Av1955 among Av39, Av154, and Av240.

5 ** " Bold and underlined" indicate the highest identity to Av1955 among viruses in the EpiFlu
6 database of GISAID with whole-genome sequence

Table 2 (on next page)

Characteristic amino acid sequences of Av1955: the receptor binding, cleavage, and glycosylation sites in HA, deletion in NA and NS1, truncation of PB1-F2, and amantadine resistant mutation in M2.

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| Amino acid mutation | | | Amino acid sequence* | | | | | | | | Pheno- type** | |
|---------------------|---------------------------------|-----------------------------|-----------------------------|--|---|---|---|---|---|---|------------------|-----|
| Pro- tein | Mutation | Site, H5 numbering | Isolates in 2013-2014 | | | Viruses with the highest identity | | | | | | |
| | | | Av39 | Av240 | Av154 | Eg2061 | Av1210 | Spg119 | Av1534 | Av1955 | | |
| HA | Receptor binding | E186G /D | E | E | E | E | E | E | E | E | E | [1] |
| | | G221D | G | G | G | G | G | G | G | G | G | [2] |
| | | Q222L | Q | Q | Q | Q | Q | Q | Q | Q | Q | [3] |
| | | G224S | G | G | G | G | G | G | G | G | G | [4] |
| | Multiple basic amino acid | 321-330 Cleav- age site | PEKQT ---- R↓G | PQRES <u>RRKK</u> <u>R↓G</u> | PQRE - <u>RRRK</u> <u>R↓G</u> | [5] | |
| | Loss of glycosylati on | Loss of 154- 156 (NXT/S) | <u>Lost</u> (GST) | NST | <u>Lost</u> (DNA) | <u>Lost</u> (NNA) | <u>Lost</u> (NNA) | <u>Lost</u> (NNA) | <u>Lost</u> (NNA) | <u>Lost</u> (CNA) | [6] | |
| NA | Deletion | 49-68 | Not deleted | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | [7] | |
| NS1 | Deletion | 80-84 | TIASV | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | <u>Deleted</u> | [8] | |
| PB1-F2 | Trancation | at 12 | at 87 | at 87 | at 57 | <u>at 25</u> | [9] | |
| M2 | Resistance to amantadine | V27A | V | <u>A</u> | I | <u>A</u> | I | <u>A</u> | V | <u>A</u> | [10] | |
| | | S31/N/G | S | <u>N</u> | S | <u>N</u> | S | <u>N</u> | S | <u>N</u> | [11] | |

3 * "Bold and underlined" indicate the mammalian adaptation markers.

4 ** [1]: Increased virus binding to α 2,6 (Glaser et al., 2005); [2]: Change in receptor binding
5 affinity from avian to human receptors (Rogers et al., 1983); [3]: Change in receptor binding
6 recognition from α 2,3 to α 2,6 (Connor et al., 1994) , Airborne transmissible in mammals
7 (Maines et al., 2011); [4]: Increased virus binding to α 2,6, (Imai et al., 2012), Airborne
8 transmissible in mammals (Maines et al., 2011); [5]: Increased virulence in mice (Klenk &
9 Garten, 1994; Suguitan et al., 2012; Yudhawati et al., 2020); [6]: Increase virus binding to α 2,6
10 and pathogenicity in mice (Wang et al., 2010; Zhang et al., 2015); [7]: Enhanced virulence in
11 mice (Matsuoka et al., 2009; Zhou et al., 2009); [8]: Enhance virulence in mice associated with
12 D92E shift (Long et al., 2008); [9]: Attenuate virulence in mammals (Dharmayantiet al., 2010);
13 [10]: Reduce susceptibility to amantadine (Hay et al., 1985); [11]: Reduce susceptibility to
14 amantadine and rimantadine (Belshe et al.,1988).

Table 3 (on next page)

Mammalian adaptation markers (Mertens et al., 2013) in the amino acid sequences of Av1955

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| Mammalian adapt. marker mutation | | Amino acid at the marker site** | | | | | | | | Pheno- type*** |
|-------------------------------------|-----------|---------------------------------|----------|----------|-----------------------------------|----------|----------|----------|----------|-------------------|
| | | Isolates in 2013-2014 | | | Viruses with the highest identity | | | | Av1955 | |
| Protein | Mutation* | Av39 | Av240 | Av154 | Eg2061 | Av1210 | Spg119 | Av1534 | | Av1955 |
| PB2 | T63I | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | [1] |
| | L89V | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | [1], [2] |
| | G309D | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | [1], [2] |
| | R318K | R | R | R | R | R | R | R | <u>K</u> | [2] |
| | T339K | <u>K</u> | T | T | A | T | <u>K</u> | T | <u>K</u> | [1], [2] |
| | Q368R | <u>R</u> | <u>R</u> | Q | <u>R</u> | Q | <u>R</u> | Q | <u>R</u> | [1], [2] |
| | H447Q | <u>Q</u> | <u>Q</u> | <u>Q</u> | <u>Q</u> | <u>Q</u> | <u>Q</u> | <u>Q</u> | <u>Q</u> | [1], [2] |
| | R477G | <u>G</u> | <u>G</u> | <u>G</u> | <u>G</u> | <u>G</u> | <u>G</u> | <u>G</u> | <u>G</u> | [1], [2] |
| | I495V | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | [1], [2] |
| | K526R | K | <u>R</u> | K | <u>R</u> | K | K | K | K | [2], [3] |
| | V667I | <u>I</u> | V | V | V | V | V | V | V | [4] |
| | A676T | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | [2], [3] |
| | K702R | <u>R</u> | K | K | K | K | <u>R</u> | K | <u>R</u> | [4] |
| Subtotal | | 11 | 9 | 6 | 10 | 7 | 10 | 6 | 11 | |
| PB1 | A3V | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | [1], [2] |
| | L13P | <u>P</u> | <u>P</u> | <u>P</u> | <u>P</u> | <u>P</u> | <u>P</u> | <u>P</u> | <u>P</u> | [1], [2], [5] |
| | R207K | <u>K</u> | R | <u>K</u> | <u>K</u> | <u>K</u> | <u>K</u> | <u>K</u> | <u>K</u> | [2] |
| | K328N | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | [2] |
| | S375N | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | S | <u>N</u> | <u>N</u> | [1], [2], [5] |
| | H436Y | <u>Y</u> | <u>Y</u> | <u>Y</u> | <u>Y</u> | <u>Y</u> | <u>Y</u> | <u>Y</u> | <u>Y</u> | [1], [2] |
| | L473V | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | <u>V</u> | [2] |
| | M677T | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | [1] |
| Subtotal | | 8 | 7 | 8 | 8 | 8 | 7 | 8 | 8 | |
| PA | H266R | <u>R</u> | <u>R</u> | <u>R</u> | <u>R</u> | <u>R</u> | <u>R</u> | <u>R</u> | <u>R</u> | [2], [3] |
| | A404S | A | A | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | [5] |
| | S/A515T | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | [2], [3] |
| | Subtotal | | 2 | 2 | 3 | 3 | 3 | 3 | 3 | |

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5 **Table 3 (Continued)**

| Mammalian adapt. | Amino acid at the marker site** | Pheno- |
|------------------|---------------------------------|--------|
|------------------|---------------------------------|--------|

| marker mutation | | Isolates in 2013-2014 | | | Viruses with the highest identity | | | | Av1955 | type*** |
|-----------------|-----------------|-----------------------|----------|----------|-----------------------------------|----------|----------|----------|----------|----------|
| Protein | Mutation* | Av39 | Av240 | Av154 | Eg2061 | Av1210 | Spg119 | Av1534 | | |
| HA | D94N | D | S | <u>N</u> | <u>N</u> | <u>N</u> | <i>N</i> | <u>N</u> | <u>N</u> | [6], [7] |
| | S133A | G | S | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | [6] |
| | Q/H/I138 L/N | G | <u>L</u> | Q | Q | Q | Q | Q | Q | [1] |
| | I151T | <u>T</u> | I | I | I | I | I | I | I | [6] |
| | N154D | G | N | <u>D</u> | N | N | S | N | G | [4] |
| | S155N | S | S | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | <u>N</u> | [2], [6] |
| | T156A | T | T | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | [4], [6] |
| | T188I | T | T | T | T | <u>I</u> | T | <u>I</u> | T | [6] |
| | K189S/R | N | M | <u>R</u> | K | <u>R</u> | K | <u>R</u> | K | [2], [6] |
| | V210I | <u>I</u> | V | V | V | V | V | V | V | [6] |
| | A263T | S | A | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | <u>T</u> | [1] |
| Subtotal | | 2 | 1 | 7 | 5 | 6 | 5 | 7 | 5 | |
| M1 | V151/T | V | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | <u>I</u> | [1] |
| | N30D | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | <u>D</u> | [1] |
| | T215A | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | [1] |
| | Subtotal | 2 | 2 | 3 | 3 | 3 | 3 | 3 | 3 | |
| M2 | L55F | <u>F</u> | L | L | L | L | L | L | L | [4] |
| | Subtotal | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | |
| NS1 | A/P42S | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | <u>S</u> | [1], [8] |
| | D87E | D | D | <u>E</u> | D | <u>E</u> | <u>E</u> | <u>E</u> | <u>E</u> | [1] |
| | T/D92E | <u>E</u> | <u>E</u> | <u>E</u> | <u>E</u> | <u>E</u> | <u>E</u> | <u>E</u> | <u>E</u> | [1] |
| | L98F | <u>F</u> | <u>F</u> | <u>F</u> | <u>F</u> | <u>F</u> | <u>F</u> | <u>F</u> | <u>F</u> | [1] |
| | I101M | <u>M</u> | <u>M</u> | <u>M</u> | <u>M</u> | <u>M</u> | <u>M</u> | <u>M</u> | <u>M</u> | [1] |
| | T127N | <u>N</u> | A | T | A | T | T | T | T | [1] |
| | V149A | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | [1], [8] |
| | N200S | <u>S</u> | G | D | G | <u>S</u> | <u>S</u> | G | <u>S</u> | [9] |
| | P215T | P | P | P | P | P | P | P | <u>T</u> | [5] |
| Subtotal | 7 | 5 | 6 | 5 | 7 | 7 | 6 | 8 | | |
| NS2 | T48A | <u>A</u> | <u>A</u> | T | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | <u>A</u> | [8] |
| | Subtotal | 1 | 1 | 0 | 1 | 1 | 1 | 1 | 1 | |
| Total | | 34 | 27 | 33 | 35 | 35 | 36 | 34 | 39 | |

6 *H5 Numbering

7 ** "Bold and underlined" indicate the mammalian adaptation markers. "Bold, underlined, and

8 italic" indicate the mammalian adaptation markers on the genome segment donor.

9 ***[1]: Pathogenic in mice, Increased virulence in mammals; [2]: Increased polymerase activity,
10 Increased replication in mammals; [3]: Increased virulence in mammals and birds; [4]: Enhanced
11 transmission, Airborne transmissibility in mammals; [5]: Human host marker; Mammalian host
12 marker; [6]: Increased virus binding to $\alpha 2,6$; [7]: Enhanced virus fusion; [8]: Antagonism of IFN
13 induction, Escape of antiviral host response; [9]: Decreased IFN antagonism.