Enhanced biomass and thermotolerance of *Arabidopsis* by *SiERECTA* in *Setaria italica* L.

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Foxtail millet is commonly used as food and forage grass. ERECTA (ER) is a receptor-like kinase that can improve plant biomass and stress resistance. Sorghum *SbER10_X1* gene was used as a probe to obtain *ER* family genes on the *Setaria italica* genomes (*SiERs*), and detect the functional characteristics of SiERs family. Herein, the structural features, expression patterns, and thermotolerance of SiERs function were identified by bioinformatics analysis, real-time PCR and transgenosis estimation. Results showed that *SiERs* had four members: two members were located on chromosome 1 with a total of six copies (*SiER1_X1*, *SiER1_X2*, *SiER1_X3*, *SiER1_X4*, *SiER1_X5*, and *SiER1_X6*), and two were on chromosome 4, namely, *SiER4* (*SiER4_X1* and *SiER4_X2*) and *SiERL1.* Among them, *SiER1_X4* and *SiER4_X1* were expressed the highest in aboveground organs of foxtail millet, and actively responded to treatments with abscisic acid, brassinolide, gibberellin, and indole acetic acid. After overexpression of *SiER1_X4* and *SiER4_X1* in *Arabidopsis*, the plant height and biomass of transgenic *Arabidopsis* significantly increased. Under high-temperature treatment, comparing to wild type, transgenic seedlings survived better after restoration of well-water culture. Tansgenic lines showed higher SOD and POD activities, and the expression of *AtHSF1* and *AtBl1* gene significantly increased. These results indicated that SiER1_X4 and SiER4_X1 played important regulatory roles in plant growth and thermotolerance. The two genes provided potential breeding targets or biotechnological methods to improve the biomass of forage grass and thermotolerance of field crops.

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ABSTRACT

Foxtail millet is commonly used as food and forage grass. ERECTA (ER) is a receptor-like kinase that can improve plant biomass and stress resistance. Sorghum *SbER10_X1* gene was used as a probe to obtain *ER* family genes on the *Setaria italica* genomes (*SiERs*), and detect the functional characteristics of SiERs family. Herein, the structural features, expression patterns, and thermotolerance of SiERs function were identified by bioinformatics analysis, real-time PCR and transgenosis estimation. Results showed that *SiERs* had four members: two members were located on chromosome 1 with a total of six copies (*SiER1_X1*, *SiER1_X2*, *SiER1_X3*, *SiER1_X4*, *SiER1_X5*, and *SiER1_X6*), and two were on chromosome 4, namely, *SiER4* (*SiER4_X1* and *SiER4_X2*) and *SiERL1*. Among them, *SiER1_X4* and *SiER4_X1* were expressed the highest in aboveground organs of foxtail millet, and actively responded to treatments with abscisic acid, brassinolide, gibberellin, and indole acetic acid. After overexpression of *SiER1_X4* and *SiER4_X1* in *Arabidopsis*, the plant height and biomass of transgenic *Arabidopsis* significantly increased. Under high-temperature treatment, comparing to wild type, transgenic seedlings survived better after restoration of well-water culture. Tansgenic lines showed higher SOD and POD activities, and the expression of *AtHSF1* and *AtBl1* gene significantly increased. These results indicated that SiER1_X4 and SiER4_X1 played important regulatory roles in plant growth and thermotolerance. The two genes provided potential breeding targets or biotechnological methods to improve the biomass of forage grass and thermotolerance of field crops.

Subjects Bioinformatics, Functional genomics, Plant physiology

Keywords *SiERECTA* family, Expression characteristics, Thermotolerance, Biomass, Foxtail millet

INTRODUCTION

Foxtail millet is an annual C_4 crop that can be used as food and forage grass (*Singh et al., 2021*). In arid and semi-arid regions, foxtail millet shows strong tolerance to various abiotic stresses (drought, salinity, high temperature, etc.) (*Aidoo et al., 2016*). However, the natural factors relied by field crops is increasingly complex. The human demand for food and energy is intensified by the global population growth and per capita income increase. To cope with this severe challenge, crop breeding varieties need to be improved by traditional breeding, functional gene screening, genome editing, and other technologies. The foxtail millet genome contains many excellent genes related to drought resistance, barren tolerance, high yield, and high light efficiency (*Zhang et al., 2012*). The rational utilization of foxtail millet functional genes is an important strategy to ensure food security, solve energy crisis, and promote development of animal husbandry.

ERECTA (ER) belongs to receptor-like kinases (RLKs) involved in the regulation of plant photosynthesis and transpiration efficiency, thereby increasing biomass and plant resistance (*Masle et al., 2005*; *van Zanten et al., 2009*). *SbER2-1* was isolated from the drought-tolerant model plant sorghum, overexpression of *SbER2-1* in maize conferred the increased drought tolerance, expecially in regard to improved water-use efficiency (*Li et al., 2019*). When *Arabidopsis AtER* gene was overexpressed in tomato and rice, the biomass of transgenic lines was increased and heat tolerance was enhanced (*Shen et al., 2015*). Further studies had shown that the fusion gene of chitin elicitor receptor kinase 1 and *ER* (*CERK1n-ER*) can induce the production of chitooligosaccharides and improve the heat tolerance of *Arabidopsis* (*Chen et al., 2020*). In poplar, overexpression of *PdER* gene in *Arabidopsis* resulted in reduced stomatal density, thereby influencing transpiration, water-use efficiency and drought tolerance (*Li et al., 2021*). Interference of MAPK cascade reaction through the interaction of *ER* with *BAK1* gene increased the resistance of *Arabidopsis* to the necrotrophic fungus *Plectosphaerella cucumerina BMM* (*PcBMM*) (*Jorda et al., 2016; Mei et al., 2021*). These results showed that the ER family had broad application prospects in regulating plant development and stress resistance.

In the current research, the characteristics of *SiER* family members (*SiER*s) in the foxtail millet genome were analyzed. The *cis*-regulatory elements of *SiERs* promoters and the amino acid motif structure were predicted, and the evolutionary relationship of *SiERs* family in monocotyledonous and dicotyledonous plants was clarified. Moreover, *SiER1_X4*, and *SiER4_X1* genes were isolated, and the expression levels in five main foxtail millet varieties, tissue expression specificity, and hormone-induced expression patterns of the two genes were identified. Finally, the biomass and thermotolerance of transgenic *Arabidopsis,* overexpresing *SiER1_X4* and *SiER4_X1* genes, respectively, were evaluated. The findings provided the functional genes for improvement of potential production and stress resistance in gramineous crops.

MATERIALS AND METHODS

Phylogenetic analysis of SiERs family in *Setaria italica*

Two *SiER* gene tags from foxtail millet (Seita.4G086700.1 and Seita.1G338900.1) were obtained with the sorghum *SbER10_X1* gene (XM_002437978.2) as a reference sequence after BLAST in the Phytozome v12.1 database. Four families of *SiER*s members were obtained by searching NCBI database with the two *SiER* tags to predict the complete CDS and chromosome-position information. The exon distribution (GSDS 2.0), *cis*-regulatory elements of promoters (Plant CARE), subcellular localization characteristics (Plant-mPLoc) and motif structure (MEME) of *SiERs* family were predicted. Moreover, The conserved functional domains (PROSITE and SMART databases), amino acid size, molecular weight, and isoelectric point (ProtParam) of SiERs proteins were analyzed. Supplementary Material Annex 1 (Table S1) listed all databases and their URLs available at the journal's website.

Based on the functional domains of SiERs, the amino acid sequences of the published ER family in monocot and dicot plants with similarity above 80% were downloaded from NCBI database (Annex 2, *SiER1_X4* gene was listed in Annex 4), to produce SiERs phylogenetic tree by MEGA5.0 software with a threshold of 1000 replications for bootstrap, according to the neighbor-joining method (Tamura *et al*. 2013).

Genes isolation and subcellular localization of SiER1_X4 and SiER4_X1

Due to the abundant transcription of *SiERs* in the pedicel tissue of the Dungu variety at heading stage, total RNA from pedice was extracted with RNAprep Pure Kit (Tiangen, DP432, China), and cDNA was synthesized with a PrimeScript First-Strand cDNA Synthesis Kit (Takara, 6110A, Japan). Taking the pedicel cDNA as material, specific primers (*SiER1_X4-F2*/*SiER1_X4-R2* and *SiER4_X1-F3* /*SiER4_X1-R3* in Annex 3) were designed to separate *SiER1_X4* and *SiER4_X1* fragment, respectively. The PCR reaction system (50 μL) was as follows: 25 μL of 2× PCR buffer, 10 μL of dNTP (2 mM), 1.5 μL of Primer-F(10 mM), 1.5 μL of Primer-R (10 mM), 1 μL of KOD FX (1.0 U/mL, KFX-101, Toyobo, China), 5 μL of cDNA as template, 6 μL of ddH_2O. The PCR procedure was as follows: 94 ℃ for 2 min, 40 cycles (98 ℃ for 10 s, 65℃ for renaturation in both *SiER1_X4* and *SiER4_X1* gene, lasting for 30s, 68℃ for 4 min for extension), and 68℃ for 10 min.

The code fragment of *SiER1_X4* and *SiER4_X1* (without the stop codon) was separated through *SiER1_X4-gfpF1*/*SiER1_X4-gfpR1* and *SiER4_X1-gfpF1* /*SiER4_X1-gfpR1* primers (Annex 3). The same PCR procedure and reaction system as above were used, except for the 62 ℃ and 61 ℃ for renaturation in *SiER1_X4-gfp* and *SiER4_X1-gfp* gene, respectively. The fusion-protein was generated as below: PCR products of *SiER1_X4* and *SiER4_X1* were differentially integrated into the N terminal of green fluorescent protein vector (pJIT16318-GFP), which included CaMV35S promoter. pJIT16318-SiER1_X4 and pJIT16318-SiER4_X1 were transferred into wheat mesophyll protoplasts (isolation from 10-day-old wheat seedlings) via the PEG4000-mediated method (Cui *et al*. 2019). The transformed cells were incubated at 22℃ in darkness for 18-20 h, and then observed and photographed under a confocal laser scanning microscope (LSM700; CarlZeiss, Germany).

Thermotolerance identification of transgenic *Arabidopsis*

SiER1_X4 and *SiER4_X1* segments (without the stop codon for fusion-protein development) were separated by primers of *SiER1_X4-1302F1*/*SiER1_X4-1302R1* and *SiER4_X1-1302F1*/ *SiER4_X1-1302R1*, respectively (Annex 3). The same PCR procedure and reaction system were used as above, except for 64 ℃ and 62 ℃ for renaturation in *SiER1_X4-1302* and *SiER4_X1-1302* gene, respectively. The PCR products of *SiER1_X4* and *SiER4_X1* were inserted into pCAMBIA1302 vector (CaMV35S promoter) to obtain the fusion vectors of pCAMBIA1302-SbER1_X4 and pCAMBIA1302-SbER4_X1, respectively. According to the steps of *Agrobacterium tumefaciens*-mediated transformation system (Bradley *et al*. 1997), the targeted fusion vectors were transformed into *Arabidopsis* (Columbia ecotype). The offspring seeds were screened under antibiotics to obtain homozygous transgenic *SiER1_X4* and *SiER4_X1* lines. The test steps were described as Chen et al. (*Chen et al., 2020*).

The stable transgenic lines overexpressing target genes were selected to cultivate on MS medium for 3 days (without antibiotics), and then move into a light incubator to grow for 7 days. Seedlings of the similar size were transplanted into flower pots (6.8×6.8 cm) with nine plants in each pot and ten pots per transgenic line. After 10 days of continuous growth in a greenhouse (26℃ growth with an 8 h/16 h dark/light, photon flux density of 525μmol·s^{-1} m^{-2}), five pots per transgenic line were treated in a light incubator at 42°C for 48 h and 60 h, and the five remains were well cultivated at 26℃ for later biomass investigation (Control).

After high-temperature treatment for 60 h, the leaves of partially transgenic and wild-type (WT) lines were collected, some samples were used to determine SOD and POD activity, referring to the procedures described by *Zheng et al.* (2020), the rest was quickly frozen in liquid nitrogen, and stored at -80°C for qRT-PCR. All the remaining treated lines were transferred to the greenhouse, well cultivation was resumed for 11 days, to observe the recovery growth of *Arabidopsis* plants, the number of plants with green leaves was counted to assess the survival rate of transgenic and WT lines after high-temperature treatment. Four individual plants from each line were served as biological replicates.

Plant material and hormone-induction treatment

Five foxtail millet germplasm resources (Dabaigu, Dungu, Jingu21, Yugu1, and Kuanjiu) were pre-germinated for 4 days. Seedlings with the similar germination were transplanted to flower pots (35 × 35 cm) with forty plants in each pot, and the flower pots were placed in a light incubator for growth (humidity 60%; temperature 23 °C/20 ℃ day/night; 16 h/8 h light/dark; light intensity 525 μmol·s^{-1}·m^{-2}). After 6 days, mixture of the stems and leaves from a single plant for each variety was collected. After culturing the remaining plants for 15 days, seedlings were taken out with roots, rinsed off the soil, and placed on filter paper to dry instantaneously, then cultured in hormone solution and deionized water (control). The concentration of hormone solution was as follows: abscisic acid (ABA) 100 μM, brassinolides (BRs) 0.75 μM, gibberellin (GA_3) 30 mM and indole

acetic acid (IAA) 10 μM (*Zheng et al., 2016*). Samples (mixture of stems and leaves) were separately collected for qRT-PCR. The treatment periods were 0, 1, 2, 4, 6, 12, 24, 48, and 60 h.

In May 2021, the Dungu variety was planted in the experimental field, embryo and coleoptile were collected at the germ stage. Roots, stems, flag leaves, flag leaf sheaths, pedicels, and inflorescence samples were collected at the flowering stage. Seeds were collected at the maturity stage. All samples were quickly frozen in liquid nitrogen after collection and stored at -80 °C for later detection of *SiERs* expression patterns in diverse organs. Three individual plants were selected as biological replicates for each sample collection.

Total RNA extraction and qRT-PCR analysis

The procedures of total RNA extraction and cDNA synthesization were as above. Nine cDNA sequences of *SiERs* family were aligned to design specific primers for *SiER1_X4* and *SiER4_X1* qRT-PCR expression. The high-temperature related gene *AtHSFA1a* and superoxide suppressor gene *AtBl1* were used to detect the molecular-response mechanism of *SiER1_X4* and *SiER4_X1* transgenic *Arabidopsis* after high-temperature stress (*Yoshida et al., 2011*; *Ishikawa et al., 2013*). The primers of *SiER1_X4* (*SiER1_X4–qRTF2*/*SiER1_X4–qRTR2*), *SiER4_X1* (*SiER4_X1–qRTF1*/ *SiER4_X1–qRTR1*), *AtHSFA1a* (*AtHSFA1a-qRTF2*/*AtHSFA1a-qRTR2*), and *AtBl1*(*AtBI1-qRTF1*/*AtBI1-qRTR1*), as well as the reference genes (*SiActin-qRTF1*/*SiActin-qRTR1* and *AtActin-qRTF5*/ *AtActin-qRTR5*), are listed in Annex 3. The target-gene-expression level was detected by qRT-PCR analysis with the ABI Prism 7500 system (Applied Biosystems, USA). Three technical replicates and three biological replicates were conducted for all experiments, and the $2^{-\Delta\Delta Ct}$ method was used for quantification (*Liu et al., 2013*).

Data processing and statistical analysis

qRT-PCR data was analyzed in accordance with the procedure of Zheng and Hu (2016). Error analysis was conducted with SPSS Statistics Software version 18.0 (SPSS18.0 IBM, USA) based on the biological replicates of three individual plants. The related indicators of agronomic traits were also statistically analyzed using SPSS18.0 software. The data of all graphs was represented as the mean ±standard error. The graphics were analyzed and produced with OriginPro 2018C SR1 and Excel 2010 software.

RESULTS

Characteristics and phylogenetic relationship of *SiERs* family of foxtail millet

Four genes were found in the *SiERs* family of foxtail millet. Among them, *SiERL4* (gene ID: LOC101753243) and *SiER4* (gene ID: LOC10175555 8097) were distributed on chromosome 4, and *SiER1* was located on chromosome 1 with two genes (gene ID: LOC101780996 and gene ID: LOC117840131) (Table 1). Further analysis (Fig. 1) showed that 1 copy and 26 exons were found in *SiERL4* sequences (XM_004964364.4), and 2 copies and 27 exons in *SiER4* sequences. In exon 25, 6 amino acids were less encoded in *SiER4_X2*

(XM_004964885.3) than in *SiER4_X1* (XM_004964884.4). Three copies were found in the LOC101780996 gene of *SiER1,* exons 1 and 2 were lacking in *SiER1_X3* (XM_014804622.2), 22 exons were found in the other two copies, 5 amino acids were lacking in exon 20 of *SiER1_X2* (XM_014804623.2), and valine was lacking in exon 21 of *SiER1_X1* (XM_014804625.2). Three copies were found in the LOC117840131 gene of *SiER1*, each of which contained 27 exons, compared with *SiER1_X5* (XM_034720593.1), mutations were found in exon 9 and 25 of *SiER1_X4* (Annex 4), and one amino acid was lost in exon 26 of *SiER1_X6* (XM_034720600.1). The amino acid structure prediction indicated that SiER4 family was larger, and the LOC101780996 of SiER1 was smaller. The nine copies of four genes in the SiERs family were all predicted to be transmembrane proteins, a typical feature of ER family protein, whereas total 15 LRR tandem regions in SiERL4 protein, 13 LRR regions in SiER4, 9 LRR regions in LOC101780996 (SiER1), and 14 LRR regions in LOC117840131 (SiER1) (Annex 5).

In the published ER family, cluster analysis showed four categories (Fig. 2): Category I and Category II contained the monocotyledonous plants, the six copies of SiER1 family and rice ER protein constituted the first category, in which SiER1_X1 and SiER1_X6, and SiER1_X4 and SiER1_X5 were closely related. Category II was composed of two copies of SiER4 family, as well as ERs of sorghum, maize, goatgrass, wheat, barley, and brachypodium. SiER4 family was closely related to sorghum and maize. Category III was composed of ER family of dicotyledon as soybean and grape. Category IV was constituted by SiERL4 and *Arabidopsis* AtER and AtERL. This finding showed that in the evolution of ER families of different species, ERL was another branching direction, the phylogenetic relationship of SbER1 family was close to modern aquatic plants, whereas that of SbER4 family was closer to field xerophytic plants.

***SiERs* gene structure and its cis-regulatory elements**

The *cis*-regulatory elements of *SiERs* family promoters were primarily involved in regulating three types of plant functional responses as follows (Table 2): (a) cell development process, including seed development, endosperm formation, meristem and mesophyll cell differentiation, cell-development cycle changes, etc.; (b) hormone-response mechanisms, including regulation pathways mediated by salicylic acid, methyl jasmonate, abscisic acid, gibberellin and auxin; (c) biological metabolic reactions, including light response, drought and low temperature induction, adversity defense, anaerobic induction, circadian rhythm regulation, etc. This finding suggests that the SiERs family could participate in the regulation of plant growth and development, and may increase plant resistance to external stress.

SiERs family belonged to a typical receptor-like kinase (Annex 5), including the N-terminal signal-peptide region, the leucine tandem region (LRRs), the transmembrane region, and the C-terminal serine/threonine kinase domain. ER families of different species greatly differed in amino acid residues in the N-terminal signal-peptide region and transmembrane region (Annex 6). The 15 motif-conserved structures in the SiERs

family can be divided into two categories (Fig. 3): The first category included SiER1_X1, SiER1_X2, and SiER1_X3, whereas the remaining six copies were classified into the second category. In the first category, motif 14 and 13, encoding the N-terminal signal-peptide region and the 1-3 LRR tandem domains, respectively, were lacking. Motif 8, encoding No.4 and 5 of the LRR region, was additionally lacking in SiER1_X3. In the second category, except for SiERL4 that lacked motif 15 and 12 (encoding 13-14 LRR structures and transmembrane region, respectively), the other SiER proteins were all equipped with 15 completely conserved motifs structure. This finding showed that no significant difference existed in the motif distribution of SiER family members, except for some amino acid change during the SiERs evolution, which speculated that the function of SiERs could be conserved in the foxtail millet.

The gene-structure characteristics of different *SiERs* copies revealed the following (Fig. 4): *SiERs* exons differed in the length, exon 25, 26, and 27 near the 3′-UTR region were larger, which encoded the threonine/serine kinase region of ER proteins. Exons near the 5′-UTR region had different cascade numbers, which mainly encoded the leucine tandem region of ER proteins. From these characteristics, it was speculated that SiERs proteins had similar regulatory functions, which received upstream signal and transmitted them into the cell, to induce downstream genes effects by phosphorylation. In the LOC101780996 genes, *SiER1_X3* lacked the first two exons, and the distribution of other exons was similar. The first intron of *SiER4* family (LOC101758097) was larger, resulting in the largest sequence of *SiER4* family. *SiERL4* (LOC101753243) had 26 exons and was divided into a separate branch. It was reported that ER family often constituted 27 exons, and *ERL* only belonged to *ERECTA-LIKE1* family (*Masle et al., 2005*; *Pillitteri et al., 2012*). In this study, both of *SiER1_X4* and *SiER4_X1* had 27 exons, showed typical gene-structure of *SiERs* family, and were selected to be isolated for their functional characteristics.

Expression patterns of *SiERs* in different foxtail millet varieties and diverse organs

Among the five common foxtail millet varieties in China, *SiER1_X4* and *SiER4_X1* showed the highest expression levels in Dungu, whereas *SiER1_X4* showed the lowest expression level in Yugu 1, as well as the lowest expression level of *SiER4_X1* in Dabaigu (Fig. 5). Compared with *SiER1_X4*, *SiER4_X1* showed a higher expression level in the five foxtail millet varieties. This finding showed that *SiERs* had different transcription levels in different foxtail millet varieties and *SiER4_X1* may have a stronger regulatory function on the development of foxtail millet. Dungu was selected as an important material for subsequent gene-expression analysis.

In different organs of Dungu, *SiER1_X4* and *SiER4_X1* genes were highly expressed in aboveground organs but rarely expressed in underground root (Fig. 6). Taking root organ as a reference, the expression level of the two genes in the pedicel were both the highest, reaching 70 and 61 times of that in root, respectively. The expression level in panicle ranked the second (only 36 and 31 times, respectively). The expression levels

in leaves and kernels were similar, both of which were at a low level. Thus, the functional roles of *SiERs* probably differed in regulating the development of different organs of foxtail millet, and the transcription levels of *SiER4_X1* gene in different organs were significantly higher than that of *SiER1_X4*.

Expression patterns of *SiER1_X4* and *SiER4_X1* under hormone induction and subcellular localization analysis

Upon treatments with the hormones ABA, BRs, GA_3, and IAA, *SiER1_X4* and *SiER4_X1* established stable expression levels in the respective control samples, whereas the extremely and significantly increased expression level was observed in the treated samples ($P<0.01$). With prolonged hormone-treatment time, the expression levels of the two genes showed a response pattern of initial increase and then decrease (Fig. 7). After treatment with ABA, the expression levels of the two genes rapidly increased. At 2 h, the expression reached the highest level, those of *SiER1_X4* and *SiER4_X1* were 7.1 and 8.6 times of the respective control, respectively. After treatment with BRs for 2 h, the expression levels of *SiER1_X4* and *SiER4_X1* gene gradually increased, the expression was the highest at 6 h. Upon treatment with GA_3, the expression levels of the two genes rapidly increased after 2 h, and the expression was the highest at 4 h, which was 15.9 and 7.0 times of the control, respectively, then the expression level rapidly decreased. After auxin (IAA) treatment, the expression of *SiER1_X4* slowly increased, whereas the expression of *SiER4_X1* rapidly increased. At 6 h and 12 h, the expression of the two genes reached their highest levels, respectively. Thus, compared with IAA treatment, the transcription level of *SiER4_X1* gene was higher under the other three treatments. This finding showed that SiERs could actively respond to hormone induction and might participate in the regulation of millet development and stress-resistance related physiological processes.

The ORF fragments of *SiER1_X4* and *SiER4_X1* were 2973 bp and 2991 bp, respectively (Annex 7). The subcellular localization analysis showed that the fluorescence signals of the two fusion proteins were located on the cell membrane and chloroplast of wheat mesophyll protoplasts, whereas the control pJIT16318-GFP was distributed on the cell membrane, cytoplasm and nucleus (Fig. 8). This result indicated that SiER1_X4 and SiER4_X1 primarily acted on cell membranes and chloroplasts, which was consistent with the above-mentioned prediction of SiERs as transmembrane proteins.

Overexpression of SiERs in Arabidopsis thaliana increased the biomass

SiER1_X4 and *SiER4_X1* were transferred into *Arabidopsis*, and the T_4 generation plants were investigated (Fig. 9). In the transgenic lines *OxSiER1_X4#3* and *OxSiER4_X1#13*, the expression levels of *SiER1_X4* and *SiER4_X1* were 66 and 9 times those of control lines (WT), respectively. Compared with WT, the plant height of the two transgenic lines significantly increased ($P<0.01$), the main stem diameter and the biomass per plant were significantly higher than that of WT lines ($P<0.05$), indicating that overexpression of *SiER1_X4* and *SiER4_X1* gene could enhance the biomass of *Arabidopsis*, which had significant implications for improving

the biomass of forage crops, such as sorghum and foxtail millet. Among them, the total number of siliques per plant of *SiER1_X4* transgenic line was significantly more than that of WT ($P<0.05$), as well as only slightly more for *SiER4_X1* line. Meanwhile, plant height, total number of siliques per plant, and biomass per plant of *SiER1_X4* transgenic *Arabidopsis* were higher than those of *SiER4_X1* transgenic lines.

Thermotolerance of *Arabidopsis thaliana* overexpressing *SiERs* genes

After treating *Arabidopsis* overexpressing *SiER1_X4* and *SiER4_X1* genes under high-temperature (42℃), the plant leaves withered and some plants showed local necrosis. After well cultivation recovering for 11 days, only a few plants of WT lines showed vital signs, and the others all died, whereas the survival rate of transgenic *Arabidopsis* was extremely and significantly higher than that of WT ($P< 0.01$), especially the *SiER4_X1* transgenic plants, which showed a stronger ability to restore growth (Fig. 10a and 10b). Further determination of the antioxidant-enzyme activity of *Arabidopsis* showed that the SOD activity of *SiER1_X4* and *SiER4_X1* lines before and after high-temperature treatment was significantly higher than that of WT plants, as well as the POD activity of both transgenic lines ($P< 0.01$) (Fig. 10c). Under high-temperature stress, the SOD and POD activities of *SiER4_X1* lines were slightly higher than those of *SiER1_X4* lines.

Detection of high-temperature regulation gene (*AtHSF1*) and superoxide suppressor gene (*AtBl1*) found that the expression level of *AtHSF1* in the transgenic lines was extremely and significantly higher than that of WT line ($P<0.01$) (Fig. 10d). Particularly, after high-temperature induction, the *AtHSF1* expression level of transgenic lines significantly increased. Before high-temperature treatment, the expression level of *AtBl1* did not significantly differ between the transgenic lines and WT. After high-temperature treatment, the expression level of *AtBl1* increased and reached a significant difference in the *SiER4_X1* lines ($P<0.05$). Before and after high-temperature treatment, the *AtHSF1* expression level of WT line did not change significantly, whereas the expression level of *AtBl1* significantly increased ($P<0.05$). This finding showed that overexpression of *SiER1_X4* and *SiER4_X1* genes can improve the high-temperature tolerance of *Arabidopsis*, which may be due to the influence of heat-related gene expression in the regulatory pathway and induce the variable activity of related antioxidant enzymes. Moreover, *SiER4_X1* showed a better regulatory function than *SiER1_X4*.

DISCUSSION

The characteristics of gene families have become an important means to analyze their function. The accuracy and reliability of analysis on the evolutionary features depend on genome-sequencing information. This study found that the foxtail millet genome contained four *SiER* family members (*SiERs*), two genes were distributed on the first chromosome, with a total of 6 copies, and two genes were distributed on the fourth chromosome, with three copies. In rice, wheat, sorghum, cotton, tobacco crops, the ER family also had two members, and each member had different spliceosomes, resulting in an uneven distribution of the number of introns and

exons in the genome (*Liu et al., 2019*). In foxtail millet, the spliceosomes in different copies of *SiERs* had obviously different forms, indicating that the relationship of *SiERs* famlily was more complicated in the evolutionary process. In eukaryotes, the gain or loss of introns is one of the evolutionary mechanisms of a gene family (*Roy et al., 2007*), the difference in the number of introns affected the target-gene expression level. The introns of *AtER* genes in *Arabidopsis* were absent, leading to the reduced target protein by 500–900 times (*Karve et al., 2011*). With decreased LRR tandem amount in the extracellular region of soybean GmER (decreased exons), shading treatment had increased the hypocotyl length, leaf area, and petiole length of *Arabidopsis* (*Du et al., 2018*). We speculated that different spliceosomes of *SiERs* resulted in great difference in regulatory functions.

Before the emergence of monocotyledonous and dicotyledonous plants, the ER family evolved into two large families, namely, ER and ERL. Later, with the occurrence of gene-replication events, multiple copies of ER and ERL families gradually formed (*Liu et al. 2019*). In the present study, the ER family can be clearly divided into four categories: aquatic monocot, terrestrial monocot, dicot, and *Arabidopsis* ER and ERL families. Among them, six copies on the first chromosome were closely related to aquatic monocots (rice), and two copies on the fourth chromosome were closely related to terrestrial monocots. Further analysis of amino acid sequences of SiERs in other species showed that different ER families greatly differed in amino acid residues in the N-terminal signal-peptide recognition and transmembrane regions. ER family was a transmembrane protein that can sense external stimuli, activate the expression of intracellular signal factors, and regulate the physiological response of cells (*Shpak et al., 2004*). The most important function of ER was phosphorylation. the amino acid position difference in the transmembrane region influenced the phosphorylation event, and the N-terminal extension region was one of the components of overall kinase folding and was critical to the kinase activity (*Kosentka et al., 2017*).

The ER family was reported to involve in light-induced undergrowth (*van Zanten et al., 2010*), improve drought resistance of maize (*Li et al., 2019*), participate in the regulation of non-host resistance of rice blast disease and coordinately regulate the resistance of *Arabidopsis* to the quantitative traits of *Verticillium* wilt together with ABA and methyl jasmonate (*Häffner et al., 2014*). Moreover, it inhibited cell division and promote cell elongation (*Qu et al., 2017*). In the current research, *SiERs* promoters contained the core elements related to abscisic acid, low temperature, drought, methyl jasmonate, anaerobic induction, and light response, speculating that SiERs played an important role in plant resistance and photosynthesis. However, no study has been reported about the mechanism of low-temperature and anaerobic-induced response. *van Zanten et al.* (*2009*) also reported that ER affected the photoelectron-transfer capacity and carboxylation rate of ribulose diphosphate carboxylase (Rubisco), thereby increasing the photosynthetic capacity of *Arabidopsis*. SiERs had two common *cis*-acting elements, G-Box and TCCC-motif, which were involved in the light-response process. Moreover, SiER1_X4 and SiER4_X1 were both located on chloroplasts, implying that SiERs were involved in

the photosynthetic process. These results indicated great application potential for improving foxtail millet photosynthesis and plant biomass.

Overexpression of *SiER*s could promote *Arabidopsis* biomass accumulation, which was primarily due to the increase of stem thickness and plant height of transgenic plants, whereas the amount of fruit holder was uncertain. This finding was similar to previous results (*Xing et al., 2011*; *Masle et al., 2005*). Under high-temperature treatment, *Arabidopsis* overexpressing *SiERs* had strong survival ability, the SOD activity of transgenic lines significantly increased. Increased SOD activity could eliminate the damage to cells inflicted by reactive oxygen species produced by plants under high-temperature stress (*De et al. 2012*). *SiERs* maybe involved in the regulation of SOD synthesis activity at high temperature and alleviate the damage to cells inflicted by O^{2-} and H_2O_2 during adversity. Moreover, the expression levels of high-temperature regulation gene (*AtHSF1*) and superoxide suppressor gene (*AtBl1*) confirmed the above statement, and the specific mechanism requires further study. Overexpression of *Arabidopsis ER* genes in tomatoes and rice could significantly improve the heat tolerance and biomass of tomatoes and rice (*Shen et al. 2015*). In *Arabidopsis*, transforming CERK1n-ERc complex factors showed that under high-temperature stress, the H_2O_2 and related electrolyte content in transgenic *Arabidopsis* were less, and the ability to withstand high temperatures was significantly increased (*Chen et al. 2020*). This finding was similar to our current results, thereby providing an important basis for the next step to reveal the molecular mechanism of high-temperature tolerance of crops.

CONCLUSIONS

This study analyzed the characteristics of *SiER* family members (*SiER*s) in the foxtail millet. We found that the foxtail-millet genome contained four *SiERs* member, two genes were distributed on the first chromosome, with a total of 6 copies, and two genes were distributed on the fourth chromosome, with three copies. Among them, *SiER1_X4* and *SiER4_X1* actively responded to the induced reaction of ABA, BRs, GA_3, and IAA, with a higher expression level in aboveground organs of foxtail millet. Comparing to wild type, the transgenic lines of *Arabidopsis* overexpressing the two genes enhanced the plant height and biomass accumulation, and showed the higher SOD and POD activities under high temperature, reflecting an increased thermotolerance in *Arabidopsis* plants. These results provided potential breeding targets or biotechnological methods to improve the forage crop production under harsh environments.

ADDITIONAL INFORMATION AND DECLARATIONS

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Competing Interests

The authors declare that they have no competing interests.

Author Contributions

- Jia Cheng Zheng conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Xiao Yi Huang conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Jie Qin Li conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Chao Wu Zeng conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Qing Yuan He conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Wan Zhao conceived and designed the experiments, read and commented on the manuscript, authored or reviewed drafts of the paper, and approved the final draft.
- Hai Zhou Chen conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Qiu Wen Zhan conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Zhao Shi Xu conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability Statement

The data that support this study are available at https://www.icloud.com/iclouddrive/.

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506 DOI 10.1071/CP20434.
507

Figure legends

Figure 1 Nucleotide sequence characteristics of *SiER* family genes. Asteris represented the mutation locus of amino acids.

Figure 2 Phylogenetic tree of ER family proteins in monocots and dicots. Each category was represented by the same symbol with the same color. Numbers beside the branches represented bootstrap values based on 1000 replications. Plant species and NCBI accession numbers of proteins in phylogenetic tree were listed in Annex 2.

Figure 3 Motif analysis of SiERs amino acid sequence

Figure 4 Intron–exon structure of *SiERs* family in monocots and dicots. Each gene was represented by the same symbol with the same color. Numbers beside the branches represented the bootstrap values based on 1000 replications.

Figure 5 Expression profiles of *SiER1_X4* and *SiER4_X1* gene in five varieties of foxtail millet (n=9). The capital letters represented the greatly significant difference of the same gene expression among different foxtail millet varieties ($P < 0.01$). The primers of *SiER1_X4* gene (*SiER1_X4 －qRTF2/SiER1_X4 －qRTR2*), *SiER4_X1 gene* (*SiER4_X1 －qRTF1/ SiER4_X1 －qRTR1*) and reference gene (*SiActin-qRTF1/SiActin-qRTR1*) were listed in Annex 3.

Figure 6 Expression profiles of *SiER1_X4* and *SiER4_X1* genes during foxtail millet growth stages (n=9). Foxtail millet variety Dungu cDNA was used to detect expression patterns of the two genes. The capital letters represented the greatly significant difference of the same gene expression among different foxtail millet growth stages ($P < 0.01$). The primers of *SiER1_X4* gene (*SiER1_X4 －qRTF2/SiER1_X4 －qRTR2*), *SiER4_X1 gene* (*SiER4_X1 －qRTF1/ SiER4_X1 －qRTR1*) and reference gene (*SiActin-qRTF1/SiActin-qRTR1*) were listed in Annex 3.

Figure 7 Expression patterns of *SiER1_X4* and *SiER4_X1* genes after hormone induction. Foxtail millet variety Dungu cDNA was used to detect expression patterns of the two genes (n=9). (a) abscisic acid (ABA) treatment (100 μM); (b) brassinolide (BR) treatment (0.75 μM); (c) gibberellin (GA_3) treatment (30 mM); and (d) auxin (IAA) treatment (10 μM).

Figure 8 Subcellular localization of SiER1_X4 and SiER4_X1 fusion proteins in wheat mesophyll protoplasts. SbER1_X4-GFP, SbER4_X1-GFP, and pJIT16318-GFP (control) were transiently expressed in wheat mesophyll protoplasts, respectively. Images were captured using a confocal microscope (scale bar = 40 μm).

Figure 9 Biomass-related traits of transgenic *Arabidopsis***.** WT was the wild *Arabidopsis* lines, *OxSiER4_X1#13* and *OxSiER1_X4#3* were *Arabidopsis* lines transfected from *SiER4_X1* and *SiER1_X4* genes, respectively. (a) *Arabidopsis* plants grown for 30 days; (b) plant stalk of *Arabidopsis* grown for 30 days; (c)

549 the fragment isolated from *SiER1_X4* and *SiER4_X1* genes (Annex 7); (d) detection of overexpression level of
550 transgenic *Arabidopsis* (n=9); (e) plant height of transgenic *Arabidopsis* (n=6); (f) main stem diameter of
551 transgenic *Arabidopsis* (n=7); (g) total number of siliques per plant of transgenic *Arabidopsis* (n=6); and (h)
552 biomass per plant of transgenic *Arabidopsis* (n=9). Asterisk represented a significant difference (*$P < 0.05$;
553 **$P < 0.01$), the same was as below.
554
555 **Figure 10** Detection of thermotolerance of transgenic *Arabidopsis.* WT was the wild type of *Arabidopsis* lines,
556 *OxSiER4_X1#13* and *OxSiER1_X4#3* were *Arabidopsis* lines transfected from *SiER4_X1* and *SiER1_X4* genes,
557 respectively. HN and HS represented well-culture and high-temperature stress plants, respectively. (a) restored
558 culturing for 11 d after high-temperature stress of transgenic *Arabidopsis*; (b) survival rate of transgenic
559 *Arabidopsis* after high-temperature stress (n=5); (c) SOD and POD activity of transgenic *Arabidopsis* (n=4);
560 and (d) expression identification of *AtHSF1* and *AtBl1* gene in transgenic *Arabidopsis* (n=9). Uppercase and
561 lowercase letters represented a significant difference at 0.01 and 0.05 level, respectively.
562
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565

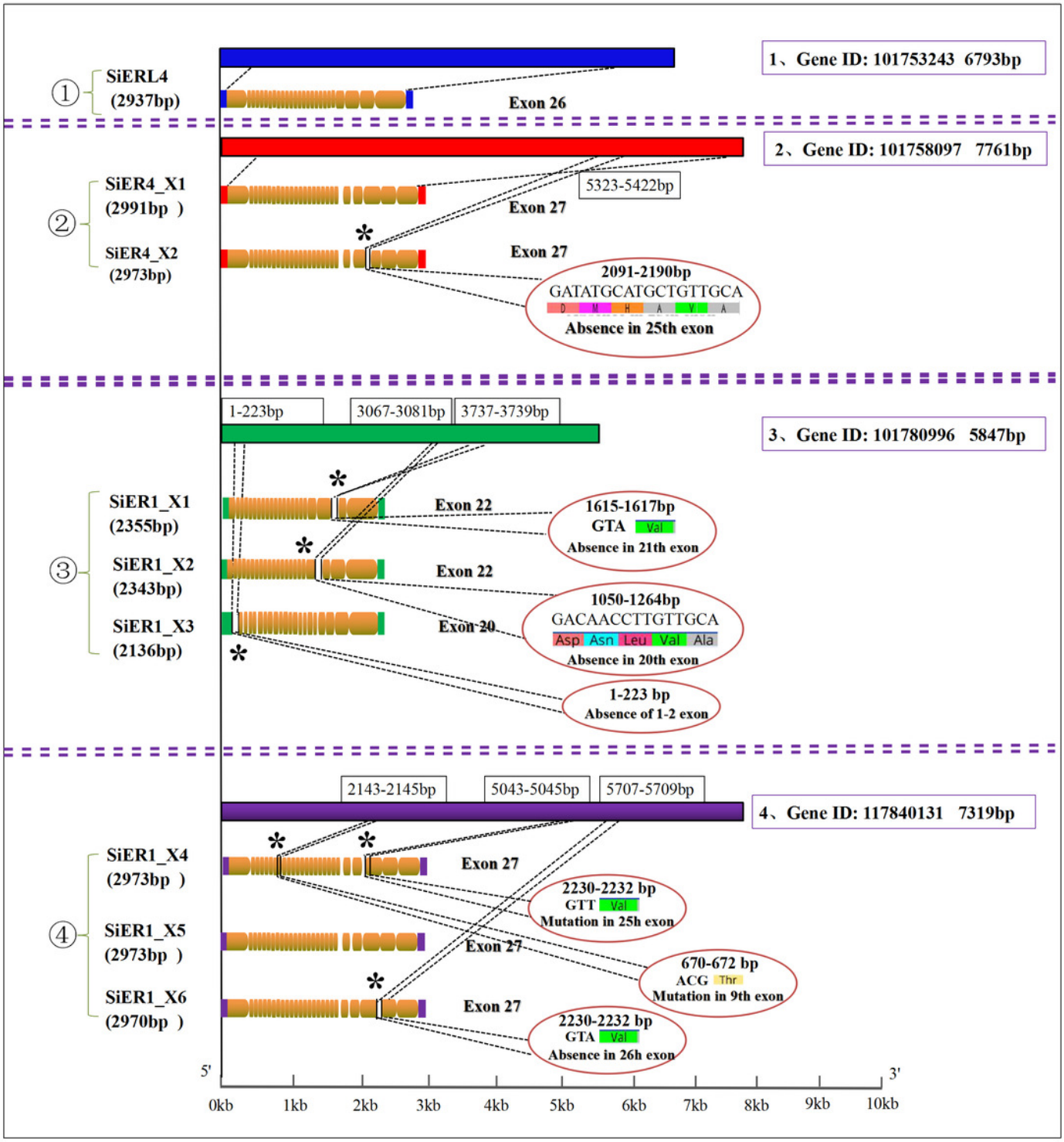
566 **Annex legends**

567

568 **Table S1** The URLs list of biological database
569
570 **Table S2** NCBI accession numbers of proteins in phylogenetic tree
571
572 **Table S3** The primers sequence related to PCR amplication
573
574
575 **Figure S1** The coding sequence of *SiER1_X4* gene
576
577 **Figure S2** The conserved structure domain of SiER family members
578
579 **Figure S3** Alignment of ERECTA family in N-terminal and transmembrane domains
580
581 **Figure S4** Isolation of *SiER1_X4* and *SiER4_X1* genes fragments.
582

Figure 1

Figure 1 Nucleotide-sequence characteristics of *SiERs* family genes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①
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(2937bp)
Exon 26
2、Gene ID: 101758097 7761bp
②
SiER4_X1
(2991bp)
SiER4_X2
(2973bp)
5323-5422bp
Exon 27
Exon 27
2091-2190bp
GATATGCATGCTGTTGCA
D M H A V A
Absence in 25th exon
1-223bp
3067-3081bp
3737-3739bp
3、Gene ID: 101780996 5847bp
③
SiER1_X1
(2355bp)
SiER1_X2
(2343bp)
SiER1_X3
(2136bp)
Exon 22
Exon 22
Exon 20
1615-1617bp
GTA Val
Absence in 21th exon
1050-1264bp
GACAACCTTGTTGCA
Asp Asn Leu Val Ala
Absence in 20th exon
1-223 bp
Absence of 1-2 exon
2143-2145bp
5043-5045bp
5707-5709bp
4、Gene ID: 117840131 7319bp
④
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(2973bp)
SiER1_X5
(2973bp)
SiER1_X6
(2970bp)
Exon 27
Exon 27
Exon 27
2230-2232 bp
GTT Val
Mutation in 25h exon
670-672 bp
ACG Thr
Mutation in 9th exon
2230-2232 bp
GTA Val
Absence in 26h exon
5'
3'
0kb
1kb
2kb
3kb
4kb
5kb
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8kb
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10kb

Figure 2

Figure 2 Phylogenetic tree of ER family proteins in monocots and dicots.

Each category is represented by the same symbol with the same color. Numbers beside the branches represent bootstrap values based on 1000 replications. Plant species and NCBI accession numbers of proteins in phylogenetic tree are listed in Annex 2.

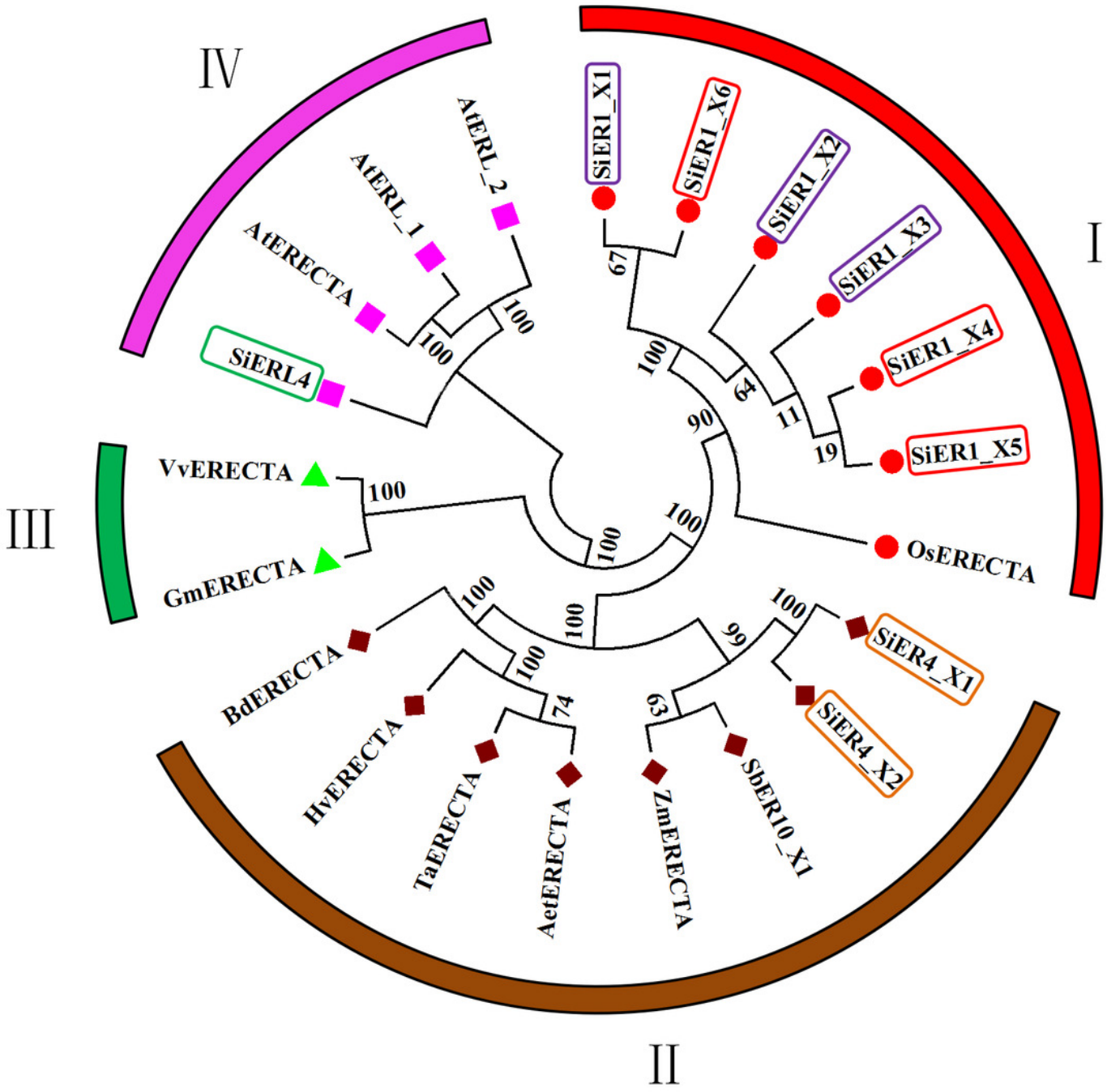
IV
I
III
II
AtERL_2
AtERL_1
AtERECTA
SiERL4
SiER1_X1
SiER1_X6
SiER1_X2
SiER1_X3
SiER1_X4
SiER1_X5
OsERECTA
VvERECTA
GmERECTA
BdERECTA
HvERECTA
TaERECTA
AetERECTA
ZmERECTA
SbER10_X1
SiER4_X1
SiER4_X2
100
100
67
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Figure 3

Figure 3 Motif analysis of SiERs amino acid sequence

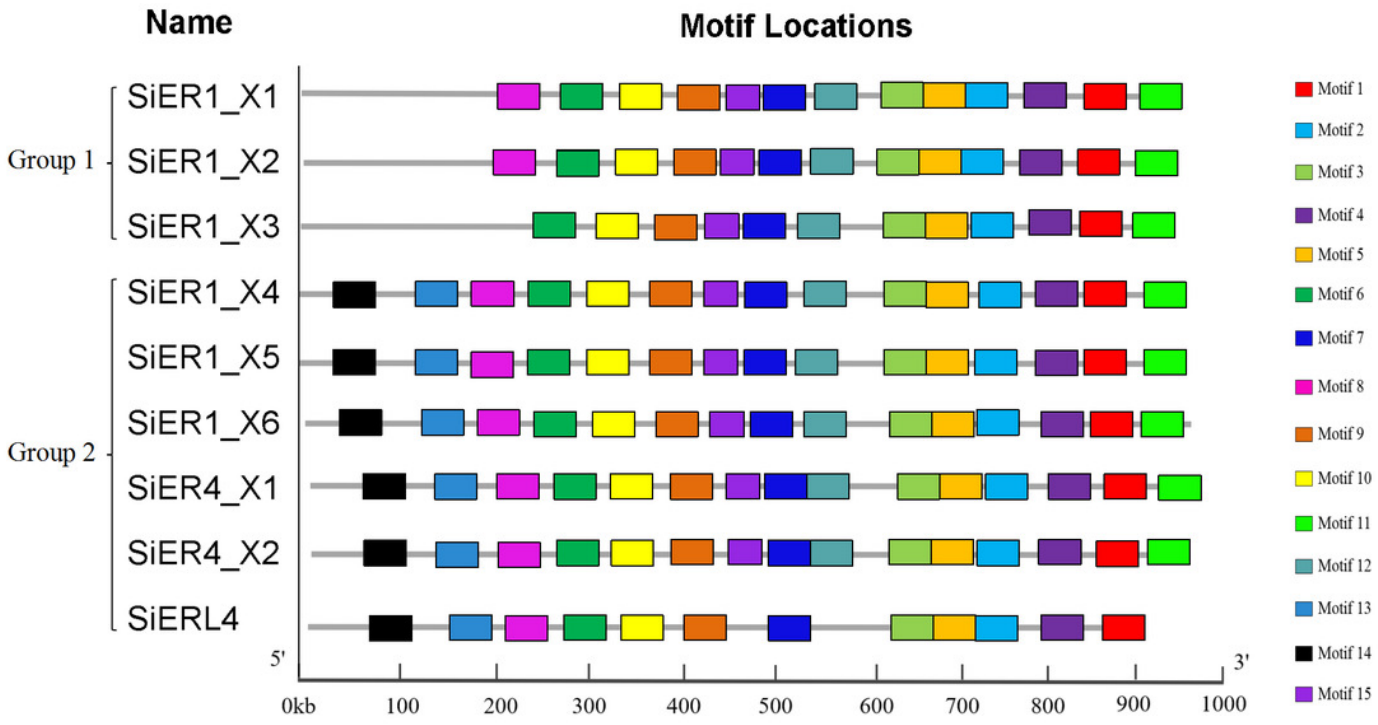

Figure 4

Figure 4 Intron–exon structures of *SiERs* family in monocots and dicots.

Each gene is represented by the same symbol with the same color. Numbers beside the branches represented the bootstrap values based on 1000 replications.

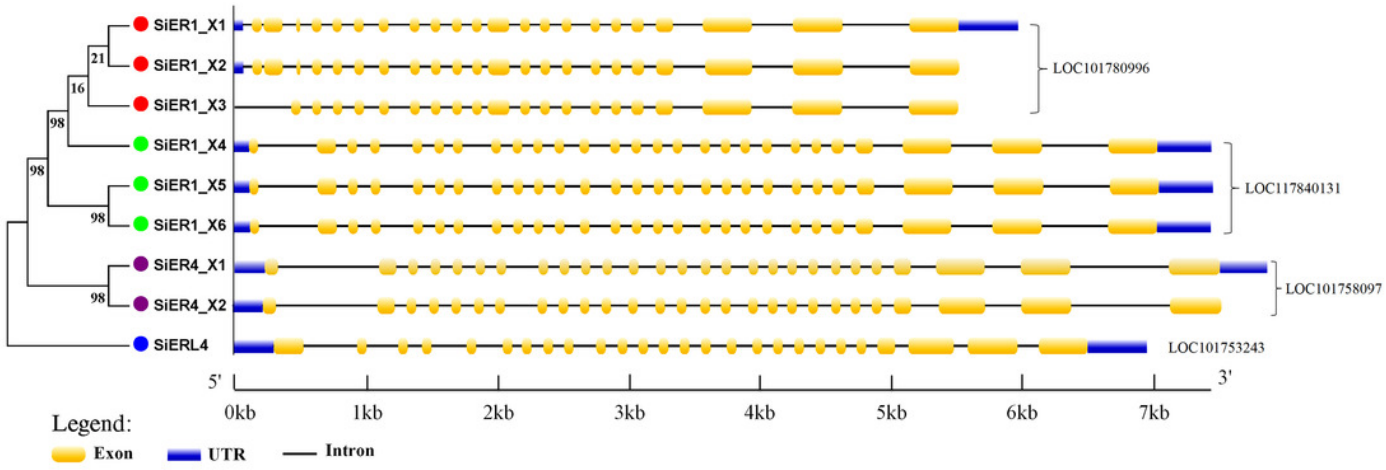

Figure 5

Figure 5 Expression profiles of *SiER1_X4* and *SiER4_X1* gene in five varieties of foxtail millet (n=9).

The primers of *SiER1_X4* gene (*SiER1_X4–qRTF2/SiER1_X4–qRTR2*), *SiER4_X1 gene* (*SiER4_X1–qRTF1/ SiER4_X1–qRTR1*) and reference gene (*SiActin-qRTF1/SiActin-qRTR1*) were listed in Annex 3.

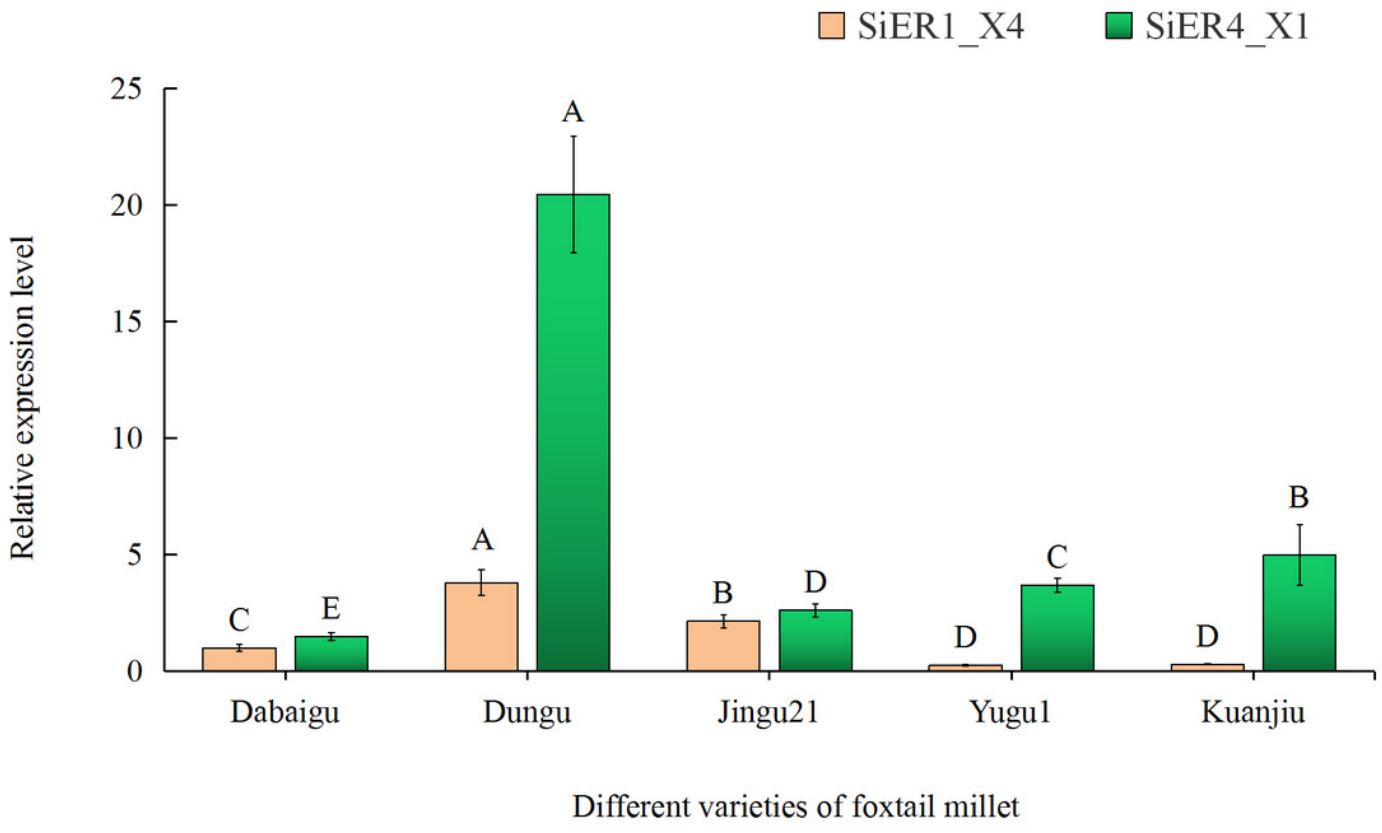

Figure 6

Figure 6 Expression profiles of *SiER1_X4* and *SiER4_X1* genes during foxtail-millet growth stages (n=9).

Foxtail millet variety Dungu cDNA was used to detect expression patterns of the two genes. The primers of *SiER1_X4* gene (*SiER1_X4–qRTF2/SiER1_X4–qRTR2*), *SiER4_X1 gene* (*SiER4_X1–qRTF1/ SiER4_X1–qRTR1*) and reference gene (*SiActin-qRTF1/SiActin-qRTR1*) were listed in Annex 3.

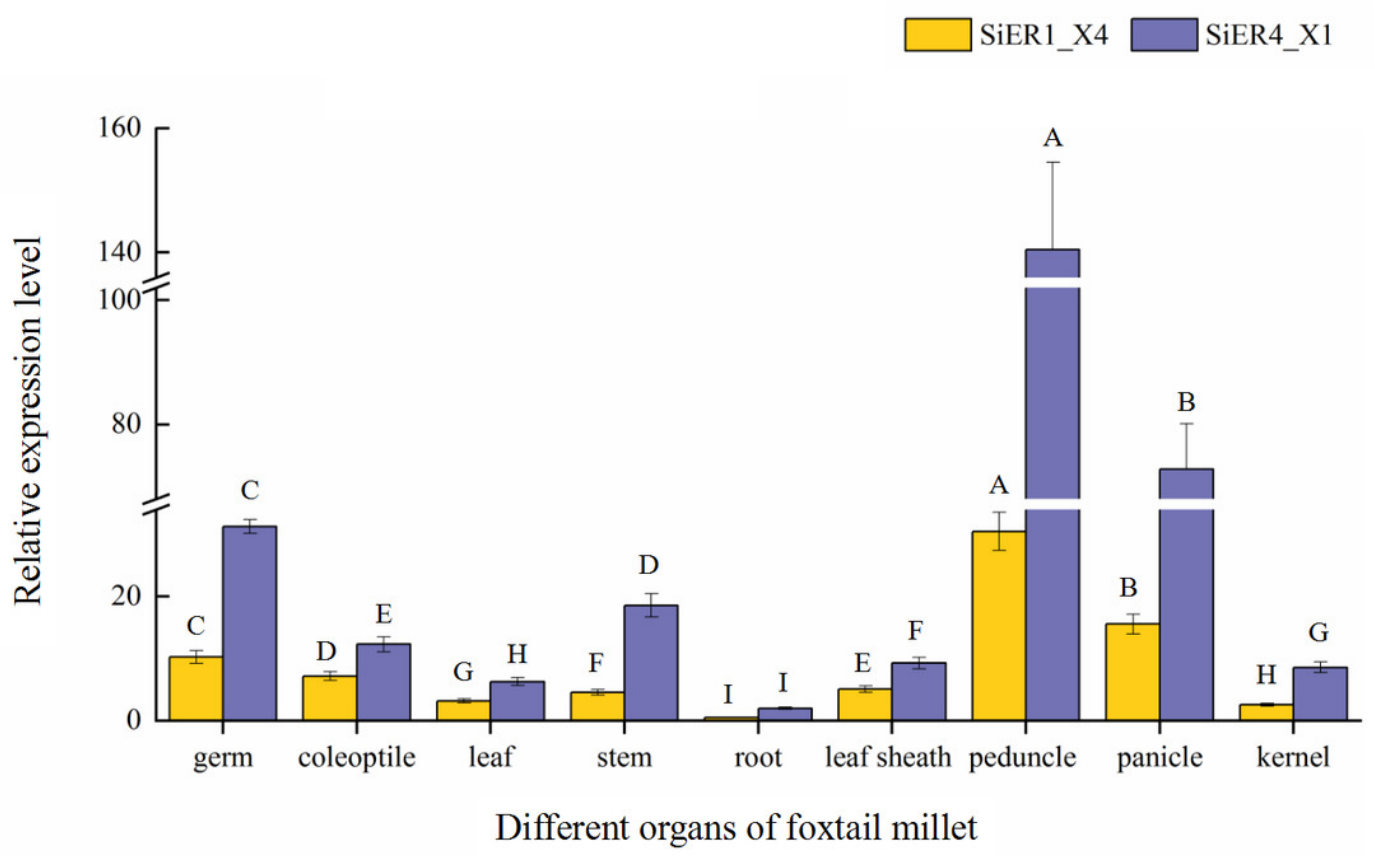

Figure 7

Figure 7 Expression patterns of *SiER1_X4* and *SiER4_X1* genes after hormone induction.

Foxtail millet variety Dungu cDNA was used to detect expression patterns of the two genes (n=9). (a) abscisic acid (ABA) treatment (100 μM); (b) brassinolide (BR) treatment (0.75 μM); (c) gibberellin (GA_3) treatment (30 mM); and (d) auxin (IAA) treatment (10 μM).

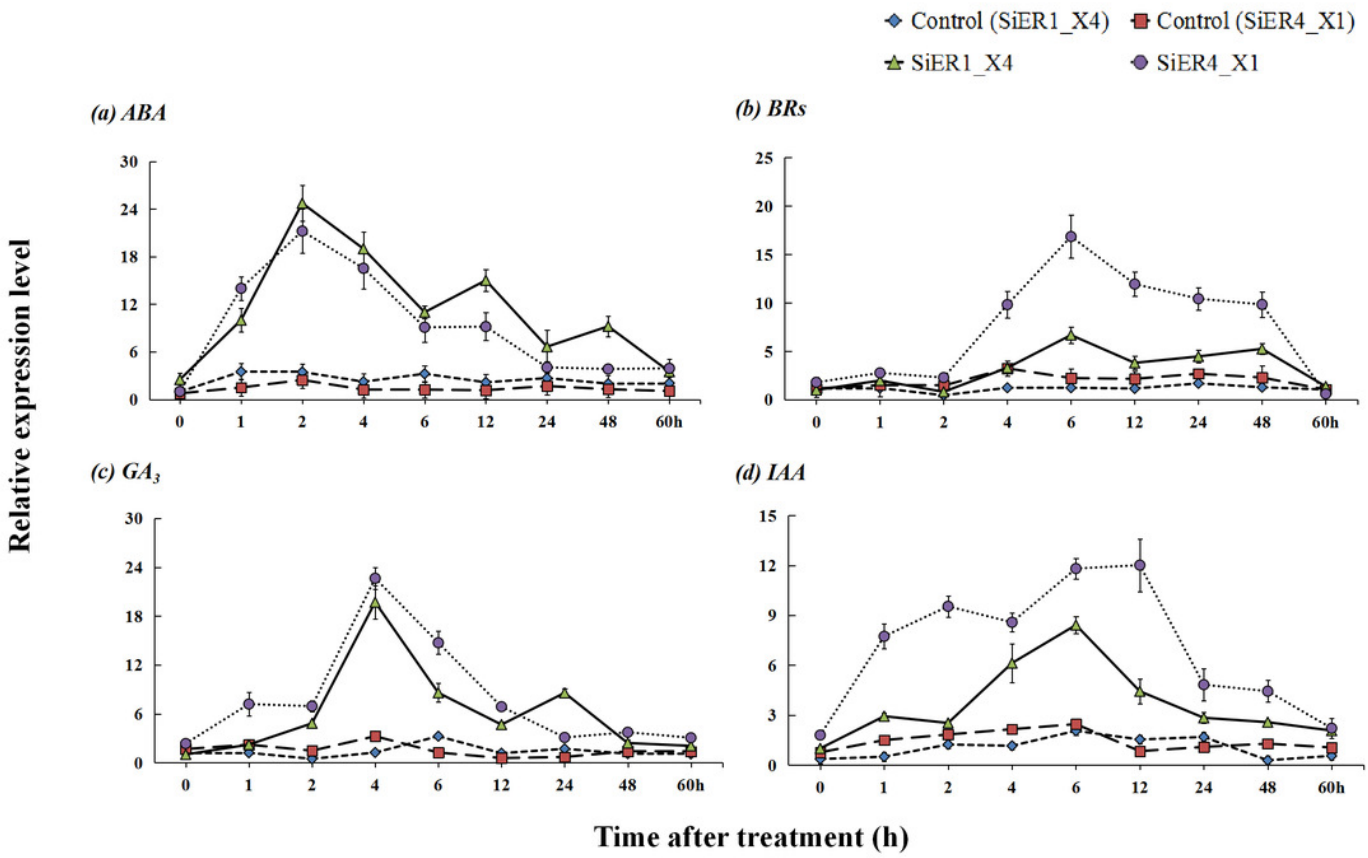

Figure 8

Figure 8 Subcellular localization of SiER1_X4 and SiER4_X1 fusion proteins in wheat mesophyll protoplasts.

SbER1_X4-GFP, SbER4_X1-GFP, and pCAM35-GFP (control) were transiently expressed in wheat mesophyll protoplasts, respectively. Images were captured using a confocal microscope (scale bar = 40 μm).

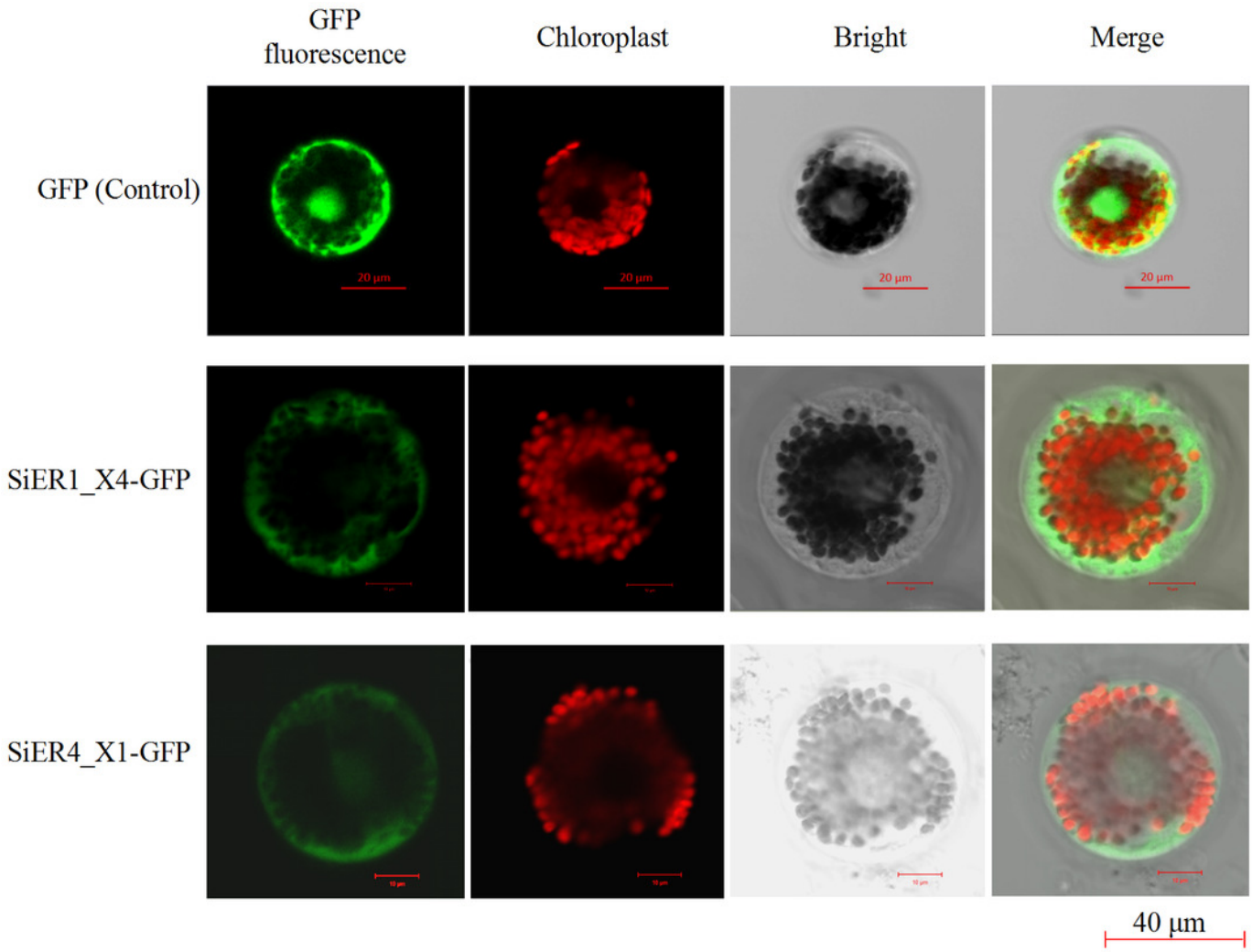

Figure 9

Figure 9 Biomass-related traits of transgenic *Arabidopsis.*

WT was the wild *Arabidopsis* strains, *OxSiER4_X1#13* and *OxSiER1_X4#3* were *Arabidopsis* strains transfected from *SiER4_X1* and *SiER1_X4* genes, respectively. (a) *Arabidopsis* plants grown for 30 days; (b) plant stalk of *Arabidopsis* grown for 30 days; (c) the fragment isolated from *SiER1_X4* and *SiER4_X1* genes (Annex 7); (d) detection of overexpression level of transgenic *Arabidopsis* (n=9); (e) plant height of transgenic *Arabidopsis* (n=15); (f) main stem diameter of transgenic *Arabidopsis* (n=15); (g) total number of siliques per plant of transgenic *Arabidopsis* (n=15); and (h) biomass per plant of transgenic *Arabidopsis* (n=15).

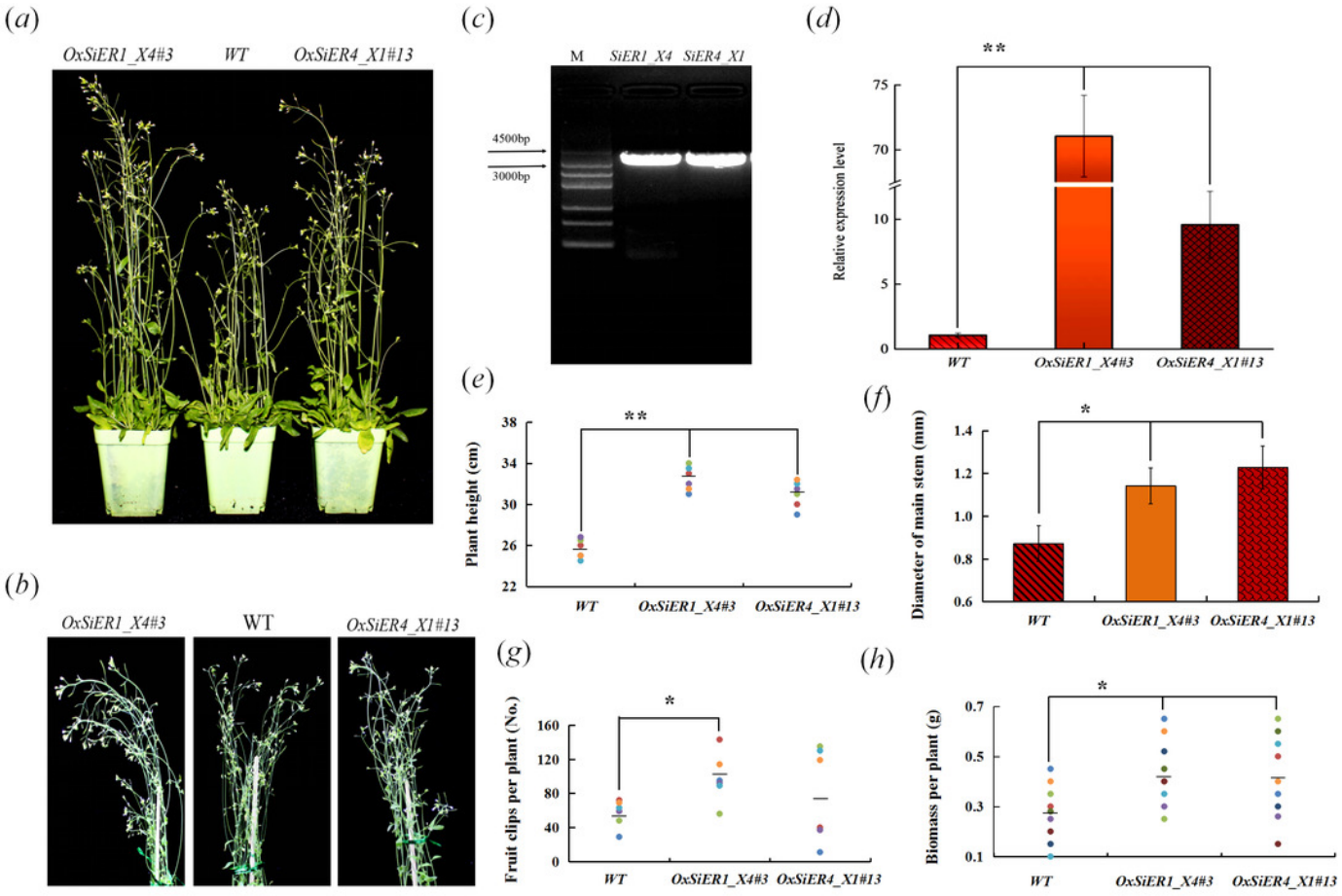

Figure 10

Figure 10 Detection of high-temperature tolerance of transgenic *Arabidopsis.*

WT was the wild type of *Arabidopsis* strains, *OxSiER4_X1#13* and *OxSiER1_X4#3* were *Arabidopsis* strains transfected from *SiER4_X1* and *SiER1_X4* genes, respectively: (a) rewatered culture after high-temperature stress of transgenic *Arabidopsis*; (b) survival rate of transgenic *Arabidopsis* after high-temperature stress (n=40); (c) SOD and POD activity of transgenic *Arabidopsis* (n=4); and (d) expression identification of *AtHSF1* and *AtBl1* gene in transgenic *Arabidopsis* (n=9).

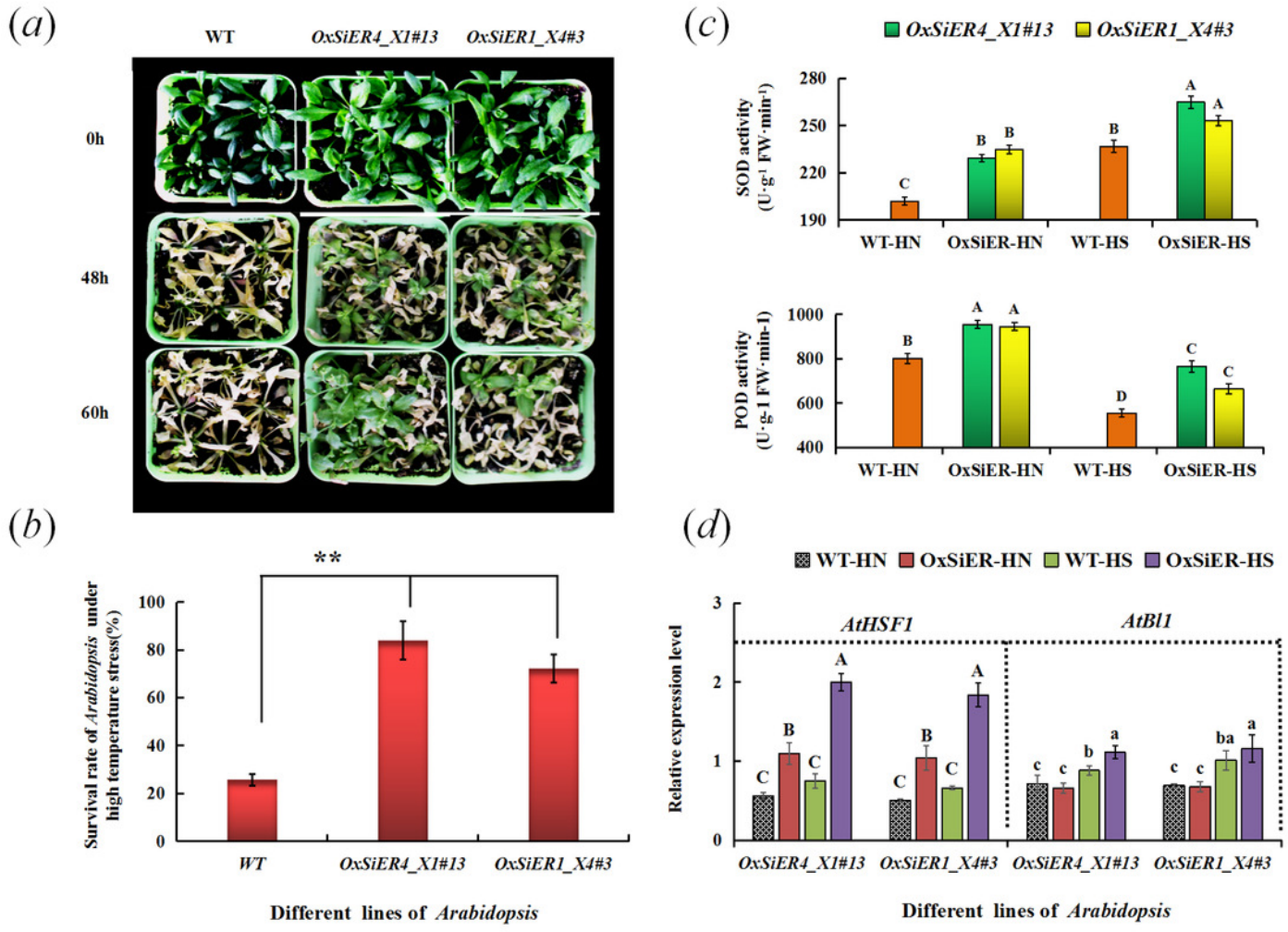

Table 1(on next page)

Table 1 The characteristics of putative *SiER* genes in *Setaria italica* L.

1 **Table 1 The characteristics of putative *SiER* genes in *Setaria italica* L.**

Name	Nucletide				Protein					Location
	Gene ID (NCBI) DNA	Gene length (bp)	Locus (NCBI) mRNA	Number of Exons	Protein accession (NCBI)	pI	Subcellular Location	Length of Protein (AA)	MW (KDa)	
SiER1_X1	LOC101780996	5847	XM_014804625.2	22	XP_014660111.1	5.77	Cell membrane	786	86	Chr. I
SiER1_X2			XM_014804623.2	22	XP_014660109.1	5.83	Cell membrane	782	85	Chr. I
SiER1_X3			XM_014804622.2	20	XP_014660108.1	5.90	Cell membrane	713	78	Chr. I
SiER1_X4	LOC117840131	7319	Annex 4	27	Annex 4 listing	5.45	Cell membrane	991	108	Chr.I
SiER1_X5			XM_034720593.1	27	XP_034576484.1	5.50	Cell membrane	991	108	Chr. I
SiER1_X6			XM_034720600.1	27	XP_034576491.1	5.50	Cell membrane	990	108	Chr. I
SiER4_X1	LOC101758097	7761	XM_004964884.4	27	XP_004964941.1	5.87	Cell membrane.	997	109	Chr.IV
SiER4_X2			XM_004964885.3	27	XP_004964942.1	5.90	Cell membrane.	991	108	Chr. IV
SiERL4	LOC101753243	6793	XM_004964364.4	26	XP_004964421.1	5.55	Cell membrane.	979	106	Chr. IV

2 Note: pI is isoelectric point; MW is the molecular weight of amino acids.

3

4

Table 2(on next page)

Table 2 Functional characteristic of *cis*-acting elements of *SiERs* promoters in *Setaria italica* L.

1 **Table 2 Functional characteristic of *cis*-acting elements of *SiERs* promoters in *Setaria italica* L.**

Code	Funtional elements of SiER promoters				Functional characteristic	Note
	SiER1 (LOC101780996)	SiER1 (LOC117840131)	SiER4 (LOC101758097)	SiERL4 (LOC101753243)		
1	RY-element		RY-element	RY-element	seed-specific regulation	cell development process
2				GCN4_motif	endosperm expression	
3	CAT-box		CAT-box	CAT-box	meristem expression	
4				HD-Zip 1	the palisade mesophyll cells differentiation	
5	MSA-like	MSA-like	MSA-like		involved in cell cycle regulation	
6	TCA-element		TCA-element	TCA-element、	salicylic acid responsiveness	hormone-response mechanisms
7	GARE-motif, TATC-box		GARE-motif, TATC-box	P-box	gibberellin responsive	
8	TGACG-motif, CGTCA-motif	TGACG-motif, CGTCA-motif	CGTCA-motif, TGACG-motif、	CGTCA-motif, TGACG-motif	methyl jasmonate responsiveness	
9	ABRE	ABRE	ABRE	ABRE	abscisic acid responsiveness	
10	TGA-element		TGA-element		auxin responsive	
11				TC-rich repeats	defense and stress responsiveness	biological metabolic reactions
12	Box 4, Sp1, GTGGC-motif, G-Box, TCCC-motif, GATA-motif, TCT-motif, ATCT-motif, GT1-motif	G-Box, Gap-box, GTGGC-motif, GT1-motif, TCCC-motif,	Box4, GT1-motif, G-Box, GTGGC-motif, Sp1, ATCT-motif, GATA-motif, TCCC-motif, TCT-motif,	TCCC-motif, Sp1, Box4, TCT-motif, L-box, G-Box, 3-AF1 binding site	light responsive	
13	MBS	MBS	MBS	MBS	drought inducibility	
14	LTR	LTR	LTR	LTR	low temperature responsiveness	
15	ARE	ARE	ARE, GC-motif	ARE	the anaerobic induction	
16	GC-motif	GC-motif		GC-motif	anoxic specific inducibility	
17	Circadian		Circadian		element involved in circadian control	

2 Note: Functional characteristics of *cis*-acting elements of *SbER* promoters were predicted in the PlantCARE database (http://bioinformatics.psb.ugent.be/webtools/plantcare/html/)