Genome-wide identification and characterization of *NHL* gene family in response to alkaline stress, ABA and MEJA treatments in wild soybean (*Glycine soja*) (#75299)

First revision

Guidance from your Editor

Please submit by 3 Nov 2022 for the benefit of the authors .



Structure and Criteria

Please read the 'Structure and Criteria' page for general guidance.



Raw data check

Review the raw data.



Image check

Check that figures and images have not been inappropriately manipulated.

Privacy reminder: If uploading an annotated PDF, remove identifiable information to remain anonymous.

Files

Download and review all files from the <u>materials page</u>.

- 1 Tracked changes manuscript(s)
- 1 Rebuttal letter(s)
- 9 Figure file(s)
- 3 Table file(s)
- 8 Raw data file(s)

i

Structure and Criteria



Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

- 1. BASIC REPORTING
- 2. EXPERIMENTAL DESIGN
- 3. VALIDITY OF THE FINDINGS
- 4. General comments
- 5. Confidential notes to the editor
- You can also annotate this PDF and upload it as part of your review

When ready submit online.

Editorial Criteria

Use these criteria points to structure your review. The full detailed editorial criteria is on your guidance page.

BASIC REPORTING

- Clear, unambiguous, professional English language used throughout.
- Intro & background to show context.
 Literature well referenced & relevant.
- Structure conforms to <u>PeerJ standards</u>, discipline norm, or improved for clarity.
- Figures are relevant, high quality, well labelled & described.
- Raw data supplied (see <u>PeerJ policy</u>).

EXPERIMENTAL DESIGN

- Original primary research within Scope of the journal.
- Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
- Rigorous investigation performed to a high technical & ethical standard.
- Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

- Impact and novelty not assessed.

 Meaningful replication encouraged where rationale & benefit to literature is clearly stated.
- All underlying data have been provided; they are robust, statistically sound, & controlled.



Conclusions are well stated, linked to original research question & limited to supporting results.



Standout reviewing tips



The best reviewers use these techniques

Τ	p

Support criticisms with evidence from the text or from other sources

Give specific suggestions on how to improve the manuscript

Comment on language and grammar issues

Organize by importance of the issues, and number your points

Please provide constructive criticism, and avoid personal opinions

Comment on strengths (as well as weaknesses) of the manuscript

Example

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Your introduction needs more detail. I suggest that you improve the description at lines 57-86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult. I suggest you have a colleague who is proficient in English and familiar with the subject matter review your manuscript, or contact a professional editing service.

- 1. Your most important issue
- 2. The next most important item
- 3. ...
- 4. The least important points

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.



Genome-wide identification and characterization of *NHL* gene family in response to alkaline stress, ABA and MEJA treatments in wild soybean (*Glycine soja*)

Xu Zhang Equal first author, 1 , Yongguo Xue Equal first author, 2 , Haihang Wang 1 , Zaib_un Nisa 3 , Xiaoxia Jin 1 , Lijie Yu 1 , Xinlei Liu 2 , Yang Yu Corresp., 4 , Chao Chen Corresp. 1

Corresponding Authors: Yang Yu, Chao Chen

Email address: yuyang_syu@163.com, chchao@hrbnu.edu.cn

Background: *NDR1/HIN1-like* (*NHL*) family genes are known to be involved in pathogen induced plant responses to biotic stress. Even though the *NHL* family genes have been identified and characterized in plant defense responses in some plants, the roles of these genes associated with the plant abiotic stress tolerance in wild soybean is not fully established yet, especially in response to alkaline stress.

Methods: We identified the potential *NHL* family genes by using Hidden Markov model and wild soybean genome. The maximum-likelihood phylogenetic tree and conserved motifs were generated by using MEME online server and MEGA 7.0 software, respectively. Furthermore, the syntenic analysis was generated with Circos-0.69. Then we used PLANT CARE online software to predict and analyze the regulatory *cis*-acting elements in promoter regions. Hierarchical clustering trees was generated using TM4: MeV4.9 software. Additionally, the expression levels of *NHL* family genes under alkaline stress, ABA and MEJA treatment were identified by gRT-PCR.

Results: In this study, we identified 59 potential *NHL* family genes in wild soybean. We identified that wild soybean *NHL* family genes could be mainly classified into five groups as well as exist with conserved motifs. Syntenic analysis of *NHL* family genes revealed genes location on 18 chromosomes and presence of 65 pairs of duplication genes. Moreover, *NHL* family genes consisted of a variety of putative hormone-related and abiotic stress responsive elements, where numbers of methyl jasmonate (MeJA) and abscisic acid (ABA) responsive elements were significantly larger than other elements. We confirmed the regulatory roles of *NHL* family genes in response to alkaline stress, ABA and MEJA treatment. In conclusion, we identified and provided valuable information on the wild soybean *NHL* family genes, and established a foundation to further explore the potential roles of *NHL* family genes in crosstalk with MeJA or ABA signal transduction mechanisms under alkaline stress.

¹ Harbin Normal University, Harbin, heilongjiang, China

² Heilongjiang Provincial Academy of Agricultural Sciences, Harbin, heilongjiang, China

University of Lahore, Lahore, Pakistan

Shenyang University, Shenyang, China



1	Genome-wide identification and characterization of
2	NHL gene family in response to alkaline stress, ABA
3	and MEJA treatments in wild soybean (Glycine soja)
4	
5	Xu Zhang ^{1, a} , Yongguo Xue ^{2, a} , Haihang Wang ¹ , Zaib_un Nisa ³ , Xiaoxia Jin ¹ , Lijie Yu ¹ ,
6	Xinlei Liu², Yang Yu⁴*, Chao Chen¹,*
7	
8	¹ School of Life Science and Technology, Harbin Normal University, Harbin, P.R. China
9	² Institute of Soybean Research, Heilongjiang Provincial Academy of Agricultural
10	Sciences, Harbin, P.R. China
11	³ Institute of Molecular Biology and Biotechnology, University of Lahore, Defence road
12	campus, Lahore, Pakistan
13	⁴ College of Life Science and Engineering, Shenyang University, Shenyang, Harbin, P.R.
14	China
15	
16	Corresponding Author:
17	Chao Chen; Yang Yu
18	Harbin Normal University, 1 South Normal University Road, Songbei Limin Economic
19	Development Zone, Harbin 150025, P.R. China
20	Email address: chchao@hrbnu.edu.cn
21	a: These authors contributed equally to this work.
22	
23	
23	
24	
25	
20	
26	
27	Abstract



28	Background: NDR1/HIN1-like (NHL) family genes are known to be involved in pathogen
29	induced plant responses to biotic stress. Even though the NHL family genes have been
30	identified and characterized in plant defense responses in some plants, the roles of these
31	genes associated with the plant abiotic stress tolerance in wild soybean is not fully
32	established yet, especially in response to alkaline stress.
33	Methods: We identified the potential NHL family genes by using Hidden Markov model
34	and wild soybean genome. The maximum-likelihood phylogenetic tree and conserved
35	motifs were generated by using MEME online server and MEGA 7.0 software,
36	respectivel - rurthermore, the syntenic analysis was generated with Circos-0.69. Then
37	we used PLANT CARE online software to predict and analyze the regulatory cis-acting
38	elements in promoter regions. Hierarchical clustering trees was generated using TM4:
39	MeV4.9 software. Additionally, the expression levels of \emph{NHL} family genes under alkaline
10	stress, ABA and MEJA treatment were identified by qRT-PCR.
11	Results: In this study, we identified 59 potential NHL family genes in wild soybean. We
12	identified that wild soybean NHL family genes could be mainly classified into five groups
13	as well as exist with conserved motifs. Syntenic analysis of NHL family genes revealed
14	genes location on 18 chromosomes and presence of 65 pairs of duplication genes.
15	Moreover, NHL family genes consisted of a variety of putative hormone-related and
16	abiotic stress responsive elements, where numbers of methyl jasmonate (MeJA) and
17	abscisic acid (ABA) responsive elements were significantly larger than other elements.
18	We confirmed the regulatory roles of NHL family genes in response to alkaline stress,
19	ABA and MEJA treatment. In conclusion, we identified and provided valuable information
50	on the wild soybean NHL family genes, and established a foundation to further explore
51	the potential roles of NHL family genes in crosstalk with MeJA or ABA signal transduction
- 2	machanisms under alkaline stress

55

56

Introduction



70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

57 NHL (NDR1/HIN1-like) family genes are previously reported to be involved in plant defense response against pathogens, such as *Phytophthora infestans*, *Botrytis cinerea*, 58 Pseudomonas syringae (Chen et al. 2018; Chong et al. 2008; Varet et al. 2002). Previous 59 studies revealed that there are at least 29 NHL family members that show homology to 60 NDR1 and HIN1 gene in Arabidopsis based on the non-redundant Genbank database 61 and then increased this number to 45 upon the completed genome sequencing of 62 Arabidopsis, suggesting a potential role for Arabidopsis NDR1/HIN1-like family genes in 63 64 plant-pathogen interactions. The 45 NHL family genes were divided into four groups and shared three conserved sequence motifs—Dormann et al. 2000; Zheng et al. 2004). 65 Analysis of amino acid sequence reveals that a lot of NHL family proteins contain a 66 specific conserved late embryogenesis abundant (LEA) domain and putative 67 68 transmembrane domain (Dormann et al. 2000; Liu et al. 2020b).

During the last three decades, many NHL family members were isolated and identified to play important roles in triggering plants defense resistance. The HIN1 gene (harpininduced gene), which could be rapidly activated and elicit HR (hypersensitive response) phenomenon when plants exposed to bacterial pathogens (e.g. Pseudomonas syringae pv. syringae), was first isolated in tobacco (Gopalan et al. 1996). The NDR1 gene (nonrace-specific disease resistance gene), which shows sequence similarity with tobacco HIN1, was first identified to play distinct roles in response to both bacterial and fungal pathogen resistance in Arabidopsis (Century et al. 1995; Takahashi et al. 2004). Overexpression of NHL3 gene could enhanced the plant resistance to *Pseudomonas* syringae pv. tomato DC3000, which was a membrane-localized protein in Arabidopsis (Varet et al. 2003). In soybean (Glycine max), two homologs of Arabidopsi NDR1 gene named GmNDR1a and GmNDR1b were identified (Selote et al. 2014). This study showed that the NDR1 protein could interact with RIN4 to play roles in resistance to *Pseudomonas* syringae. The function role of GmNDR1b, also named Gm-NDR1-1, was further determined that played important roles in impairing root pathogenic nematode *Heterodera* glycines and Meloidogyne incognita (McNeece et al. 2017). The StPOTHR1, a NHL family member in potato, could enhanced plants resistance to Phytophthora infestans through effecting the MAP kinase signaling process by interacting with NbMKK5L (Chen et al. 2018). Overexpression of the pepper CaNHL4 enhanced the expression of salicylic acid



(SA)- related and jasmonic acid (JA)-related genes, increased ROS production, and inhibited the infection of the pathogens (Liu et al. 2020a). The interaction of ToxA with NHL10 protein could induce cell death under plant pathogen stress in wheat (Dagvadorj et al. 2022)

Saline-alkaline soils are known to have high content of sodium, bicarbonates and high pH, which consequently causes growth retardation and ultimately leads to death of plants growing in such soils. The total of 434 million ha of global land is affected by alkaline soils (Jin et al. 2006; Wang et al. 2008). In comparison with neutral salts stress, alkaline stress exerts more harmful effects on plant growth (Yang et al. 2008). Alkaline stress can inhibit photosynthesis, N and sugar metabolism, as well as limits the absorption of ions, such as H₂PO₄-, Cl⁻, Al³⁺ and Fe²⁺ (Vondrackova et al. 2015).

It has been documented that *NHL* family genes also play distinct roles in plant abiotic stress resistance. In pepper (*Capsicum annuum* L.), fifteen *NHL* genes were identified in a genome-wide analysis and the responses of these genes were characterized under different abiotic stresses (Liu et al. 2020b). A stress-inducing *NHL* member, *BnNHL18A*, was isolated from *Brassica napus*, which displayed roles in response to different treatments including NaCl, H₂O₂, as well as ethephon and SA (Lee et al. 2006). In Arabidopsis, overexpression of NHL6 increased the sensitivity to salt, osmotic and ABA treatment, and NHL6 could affect the seed germination and early seedling development under these stresses-induced ABA signaling (Bao et al. 2016). Furthermore, studies have shown that some saline or alkaline stress induced transcription factors co-expresses with NHL network to regulate the salt or alkali response(Liu et al. 2020c). The soybean NHLs have been identified to play important roles in regulating seed germination under chilling stress and with ABA treatment (Wang et al. 2022). However, little is known about the wild soybean NHL family genes in response to environment stresses, especially under alkaline stress.

Soybean has been adopted as important crops in the world, particularly for protein and oil production (Yu et al. 2018). Wild soybean, as the ancestor of cultivated soybean, showed a better adaptation to various abiotic stress, such as salt, drought and alkaline. Therefore, the wild soybean has been suggested as a valuable sources to improve the



119

120

121

122

123

124

agronomic traits of soybean(Wen et al. 2009). In previous studies, we have identified a highly adaptable saline-alkali soil tolerant wild soybean (*Glycine soja*) line (G07256). It can survive well in the saline-alkali soil (Ge et al. 2010). By using transcriptome data, we identified some candidate genes in response to alkaline stress. In this study, the NHL family genes in wild soyabe genome were identified and their expression was investigated under the influence of growth hormones in alkaline stress which may enhance the stress responses.

125

126

127

136

137

Materials and Methods

Identification of NHL family genes in wild soybean genome

isoelectric point values of NHL family proteins (Artimo et al. 2012).

To identify all potential genes encoding NHL family genes in wild soybean genome, a 128 Hidden Markov model was first established by using the Arabidopsis and soybean amino 129 acid sequences of NHL family genes as gueries (Gopavajhula et al. 2013; Wang et al. 130 2022). The HMM profile (build 2.3.2) was further used to search in wild soybean genome 131 database to get similar sequences (Finn et al. 2011). Then, the potential genes were 132 identified after removing the overlapping genes and incomplete domains genes through 133 Pfam and SMART database (Finn et al. 2016). The online software ExPASy 134 (https://web.expasy.org/compute pi/) was used to predict the molecular weight and 135

Bioinformatics analysis of NHL family genes

The conserved motifs of all the potential NHL family genes were identified by MEME 138 online server (http://meme-suite.org/) (Bailey et al. 2009). The maximum-likelihood 139 phylogenetic tree was constructed by software MEGA 7.0 (Kumar et al. 2008) software. 140 Then, the TBtools software was used to combine the conserved motifs and phylogenetic 141 tree. The syntenic analysis was generated with Circos-0.69 (http://circos.ca/) (Krzywinski 142 et al. 2009). To analyze the potential regulatory cis-acting elements in the promoters of 143 NHL genes, 3000 bp upstream sequences of the above-mentioned genes were extracted 144 based on the genome database(Sun et al. 2014). Then we used PLANT CARE online 145 software (http://bioinformatics.psb.ugent.be/webtools/plantcare/html/) to predict and 146



164

173

174

analyze the regulatory cis-acting elements in promoter regions (Lescot et al. 2002). To examine the expression profiles of *NHL* family genes under alkaline stress, wild soybean transcriptome data was downloaded (DuanMu et al. 2015) and hierarchical clustering trees were generated using TM4: MeV4.9 software (Saeed et al. 2006).

Plant material, growth condition and stress treatment

The wild soybean cultivar DN50 was grown in 1/4 Hoagland nutrient solutions in the 152 growth chamber with 22-28 °c room temperature, 70-80% relative humidity and 8 h 153 dark/16 h light. The healthy and plump seeds were rinsed with 75% ethanol for 1 min, 154 and then washed with sterile water before germination (Qiao et al. 2020). After two days, 155 the germinated seedlings were transferred in to 1/4 strength Hoagland nutrient solutions 156 to be cultured and the nutrient solutions were changed every two days. Twelve days later, 157 the young seedlings were treated with alkaline (NaHCO₃) stress or exogenous hormones 158 159 (ABA and MeJA), respectively. For alkaline stress, the 12-days old seedlings were transferred into 1/4 Hoagland solution with 50 mM NaHCO₃. For exogenous hormones 160 treatment, the 12-days old seedlings were transferred into 1/4 Hoagland solution with 50 161 μM ABA or 1/4 Hoagland solution with 50 μM MeJA. The roots were harvested and stored 162 in liquid nitrogen at 0 h, 1 h and 3 h after treatment for RNA extraction. 163

Transcript expression analysis by qRT-PCR

Total RNA was extracted with the OminiPlant RNA isolation kit (Kangwei), and the cDNAs 165 were synthesized using the First Stand cDNA Synthesis kit (Toyobo) for qRT-PCR. The 166 167 gRT-PCR was then performed with UtraSYBR Mixture (Baioleibo) and ABI 7500 sequencer. The primers of wild soybean NHL genes and GsGADPH were listed in 168 supplementary Table S1 which are used in our study. Here the GsGADPH gene was used 169 as an internal control in wild soybean (Huis et al. 2010). The gRT-PCR data was 170 calculated with three independent biological replicates using 2-ΔΔCT method and Student's 171 *t*-test. 172

Results

Identification of NHL genes in wild soybean



To identify the *NHL* family genes in wild soybean, the amino acid sequences of the *NHL* family genes from Arabidopsis and soybean were queried against the wild soybean genome via BLAST from NCBI. A total of 208 NHL candidate sequences were obtained based on the Hidden Markov model. All candidate sequences were then subjected to Pfam and SMART database to remove the redundant sequences or incomplete domain sequences. As a result, 59 genes were obtained as potential *NHL* family genes in wild soybean genome.

As shown in Table 1, 59 predicted wild soybean *NHL* genes were named based on the location within the reference genome from *GsNHL1* to *GsNHL59*. Then the chemical properties of these proteins were determined including the protein sequence lengths, molecular weights (MW), and theoretical isoelectric points (pl). The protein sequence length was ranged from 149 (*GsNHL33*) to 348 (*GsNHL17*) amino acids residues. The MW varied from 16.37778 (*GsNHL33*) to 40.00232 (*GsNHL17*) kDa and the pl values ranged from 7.82 (*GsNHL31*) to 10.24 (*GsNHL43*).

Phylogenetic and conserved motifs analysis of wild soybean NHL genes

To confirm the evolutionary relationships of *NHL* family genes in wild soybean, a maximum-likelihood phylogenetic tree was constructed with the full-length protein sequences from soybean, Arabidopsis and wild soybean. The result showed that the genes could be divided into six groups and the other 7 ungroup genes (supplementary Fig. S1).

Based on conserved motif sequences, the 59 wild soybean *NHL* family genes could be further classified into five groups (group 1a, group 1b, group 2, group 3a and group 3b) and the other 11 ungroup genes (Fig. 1a). MEME motif analysis also revealed that wild soybean *NHL* family proteins shared ten conserved motif sequences (Fig. 1b, supplementary Fig. S2). Most of wild soybean *NHL* family proteins contain conserved motif 1 and motif 2. Interestingly, we found that some wild soybean *NHL* family proteins in the same group shared a similar motif composition. For example, motif 5 is mainly present within group 1b, group 2 and group 3b genes, while most genes of group 1a and group 1b contain motif 7 and 9. The motif 3 only located in group 1a genes. The similar motif arrangement among the proteins of wild soybean *NHL* family suggested that the



205 protein architecture was conserved within subgroups, which indicated that the proteins in 206 the same group may have similar function in plant development and resistance responses 207 under stress conditions. However, functions of these conserved motifs are still need to be 208 further explored.

Chromosomal locations and syntenic analysis

The analysis of gene duplication events could drive the potential evolution mechanisms of the wild soybean *NHL* family genes. In this study, 59 wild soybean *NHL* family genes were found randomly distributed among 18 chromosomes, with the exception of 8 and 17 (Fig. 2). Gene duplication plays significant roles in plant organismal evolution and functional diversity (Bowers et al. 2003). Further, a total of 65 pairs of *NHL* syntenic paralogs were identified in wild soybean genome. These results indicated that the wild soybean *NHL* family have been exhibited a high gene family expansion.

Identification of cis-acting elements of NHL gene promoters in wild soybean

To explore the potential roles of wild soybean *NHL* family genes in response to abiotic stress, the promoter sequences 3 kb upstream regions of the ATG were predicted using information within the PlantCARE online tool. The results showed that the wild soybean *NHL* family genes displayed a variety of putative hormone-related and abiotic stress responsive elements (Fig. 3, supplementary Table S2). For example, the plant hormone-related responsive elements include Methyl jasmonate (MeJA), abscisic acid (ABA, ABRE), gibberellin (GA), salicylic acid (SA) and Auxin responsive elements. Interestingly, we found that the numbers of MeJA and ABA responsive elements were significantly larger than the other plant hormone responsive elements, indicating the potential roles of *NHL* family genes in MeJA and ABA signaling pathways. We also identified some response elements including MBS, LTR and TC-rich, which respond to drought, low temperatures and general stress, respectively. Collectively, these results strongly suggested that the roles of wild soybean *NHL* family genes are likely associated with plant abiotic stresses and hormone stimuli.

232 Expression analysis of NHL genes in response to alkaline treatment in wild

soybean



235

236

237

238

239

240

241

242

243

244

245

246

247

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

To assess the potential roles of NHL family genes participate in the defense responses towards alkaline stress, we generated a heat map of NHL family genes based on the wild soybean transcriptome data under alkaline stress. The results showed that 24 genes were differently induced under alkaline stress. Among them, 18 of NHL family genes were significantly up-regulated, while six genes showed down-regulation patterns (Fig. 4). To further confirm the expression of NHL family genes in response to alkaline treatment, we selected 12 of the up-regulated genes to detect their expression patterns under 50 mM NaHCO₃ stress by using qRT-PCR analysis. As shown in Fig. 5, the expression patterns of 11 up-regulated genes were roughly consistent with the transcriptome data under alkaline stress, except that GsNHL29 had contrary results. In addition, the expression of GsNHL9, GsNHL44, GsNHL45 and GsNHL47 showed higher expression levels at 3 h point than the other genes (Fig. 5e, i-k). In conclusion, the qRT-PCR analysis confirmed the results that *GsNHL* family genes possibly participate in responses to alkaline stress.

Effects of different phytohormone treatments including ABA and MeJA on the expression of NHL genes 248

The plant hormones play regulatory roles in plant responses to various stresses. In this study, we found that the numbers of MeJA and ABA responsive elements in wild soybean NHL family genes were significantly larger than the other plant hormone responsive elements. Here, to explore if the NHL family genes could participate in ABA and MeJA signaling pathways, we analyzed the transcript expression levels of the 12 NHL family genes mentioned above under ABA and MeJA treatments by using qRT-PCR analysis. As shown in Fig. 6, nine genes were up-regulated under MeJA treatment and GsNHL29 was down-regulated. In addition, GsNHL6 and GsNHL11 had contrary expression pattern at 1 h and 3 h (Fig. 6b, f). Under ABA treatment, only GsNHL44 and GsNHL51 were upregulated and seven genes were down regulated (Fig. 7). GsNHL4 had contrary expression pattern at 1 h and 3 h (Fig. 7a). GsNHL6 and GsNHL45 showed no significant expression changes (Fig. 7b, g). Collectively, these results indicated that NHL family genes participate in ABA and MeJA signaling pathways and play different roles in response to ABA or MeJA signaling pathway.

Discussion



Previously, studies have identified that *NHL* family genes are involved in plant development and pathogens attack resistance (Bao et al. 2016; Chen et al. 2021). Many *NHL* family genes have been identified in plant species, such as tomato, pepper and soybean (Dormann et al. 2000; Liu et al. 2020b; Wang et al. 2022). However, the wild soybean *NHL* family genes have not been identified, especially the roles of *NHL* family genes in regulating alkaline stress. Hence, this research was based on bioinformatics analysis about wild soybean *NHL* family genes in order to understand their structure and location, and mainly potential roles were investigated in response to plant hormones and alkaline stress treatments.

In this study, 59 wild soybean *NHL* family genes were identified in accordance with the soybean and Arabidopsis *NHL* related genes (Table 1). We found that NHL family proteins varied markedly in protein sequence length and molecular weight, indicating the divergent evolution in wild soybean *NHL* family genes. However, the high pl value showed NHL families are alkaline proteins.

Previous studies revealed that NHL protein family could be classified into six groups by investigating the relationship of soybean, Arabidopsis and rice(Wang et al. 2022). This result was consistent with our findings that wild soybean *NHL* family genes be divided into six groups (supplementary Fig. 1). On the basis of conserved motif analysis clustered, we found most of wild soybean *NHL* family genes could be classified into five groups, which was also consistent with the results of phylogenetic tree analysis of *NHL* family genes in soybean (Wang et al. 2022). In addition, each group almost shared a similar motif composition, which indicated that the groups may have similar roles in plant development progress (Fig. 1).

Further, the wild soybean *NHL* family genes from the same group were mostly located in different chromosomes and located near the edges of the chromosomes, suggesting a strategy to exert their functions in the whole wild soybean genome. The pairs of NHL syntenic paralogs also indicated that the wild soybean *NHL* family have been exhibited a high gene family expansion, which might play significant roles in gene functional diversity (Fig. 2).



294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

In addition, we found that conserved motif 1, 3, 4, 5 and 10 belong to the LEA-2 domain (Fig. 1b, Supplementary Fig. S2). This result also consistent with previous study (Liu et al. 2020b). Furthermore, we found that LEA-2 domain belongs to the LEA_2 subgroup which are widely known as a late embryogenesis abundant proteins and play significant roles under abiotic stress responses (Jin et al. 2019). For example, the rice LEA proteins showed accumulation during the salinity-triggered growth, while degradation in LEA proteins was observed during plant recovery from salt stress (Chourey et al. 2003). The tea plant LEA genes were significantly induced under stress conditions, such as drought, ABA, low and high temperature (Jin et al. 2019). Overexpression of *IpLEA* could show high tolerance to salt and drought stress in *Ipomoea pescaprae* by mediating water homeostasis and as a reactive oxygen species scavenger (Zheng et al. 2019). Thus, this evidence indicated the potential roles of wild soybean *NHL* family genes in response to environmental stresses.

The cis-acting regulatory elements play important roles as molecular switches to control various biological processes, including hormonal and various stress responses (Sun et al. 2021). The *cis*-acting regulatory elements analysis showed that the promoter regions of wild soybean NHL family genes contain a variety of putative hormone-related and abiotic stress responsive elements (Fig. 3). Previous studies have been shown that NHL family genes participate in the plant hormone-mediated pathways. For example, overexpression of AtNHL1 and AtNHL8 in soybean could enhance plants resistance to Heterodera glycines by mediating the jasmonic acid and ethylene pathways, confirming the roles of these genes in plant defense response (Maldonado et al. 2014). NHL6 participates in the abiotic stresses-induced ABA signaling at seed germination and early seedling stages in Arabidopsis (Bao et al. 2016). BnNHL18A could be significantly induced by NaCl, ethephon, methyl jasmonate or salicylic acid treatment in Brassica napus (Lee et al. 2006). Also, LTR is a cis-element responsive to low-temperature stress (Brown et al. 2001). TC-rich cis-element has been identified to be involved in stress mediated plant defense responses (Sazegari et al. 2015). In conclusion, these evidence strongly suggested that NHL family genes may be involved in stress resistance and plant hormones responses in wild soybean.



324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

Alkaline stress is one of the most harmful abiotic stresses, which leading to a series of regulatory mechanisms in plants, such as ion balance, osmotic adjustment, pH regulation, and ROS scavenging mechanisms. Previously, we identified a highly adaptable saline-alkali soil tolerant wild soybean line which can survive well in the saline-alkali soil. Then, we explored the differentially expressed genes of wild soybean seedlings treated with 50 mM NaHCO₃ by RNA sequencing (DuanMu et al. 2015). In this study, we mainly intend to explore the potential roles of wild soybean *NHL* family genes in response to alkaline stress. According to the transcriptome data, a total of 24 genes were significantly induced under alkaline stress (Fig. 4), and qRT-PCR confirmed the results that wild soybean *NHL* family genes may play positive role in response to alkaline stress (Fig. 5).

During abiotic stress responsive processes, plant hormones such as MeJA, ABA, SA, GA and Auxin also play important roles and have cross talks in signal transduction pathways (Ku et al. 2018). In our previous study, we also identified that plant hormones have crosstalk with plant alkaline stress resistance response. For example, the wild soybean gene ERF71 could regulate endogenous auxin accumulation when plants treated with alkaline solution (Yu et al. 2017). The TIFY10 gene could act as a regulator in response to alkaline stress and jasmonate signaling in wild soybean (Zhu et al. 2011). On the other hand, we found that wild soybean *NHL* family genes comprised of a variety of putative hormone-related responsive elements, and the numbers of MeJA and ABA responsive elements were significantly larger than others. Thus, we concluded that the wild soybean NHL family genes have crosstalk with MeJA or ABA signal transduction under alkaline stress. gRT-PCR analysis showed that nine genes were up-regulated under MeJA treatment, and these genes were all up-regulated under alkaline stress (Fig. 6). This finding is consistent with the previous studies that wild soybean TIFY10a overexpression lines enhanced the alkaline stress resistance and also increased the jasmonate content of the transgenic alfalfa (Zhu et al. 2014). However, in comparison with MeJA treatment, gRT-PCR analysis showed a different expression, in which only two wild soybean NHL family genes were up-regulated and seven genes were downregulated under ABA treatment (Fig. 7). For example, GsNHL7, GsNHL8, GsNHL9, GsNHL12 and GsNHL7 were up-regulated under alkaline stress and MeJA treatment, while were down-regulated under ABA treatment. In addition, our previous studies



355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

identified that overexpression of wild soybean *NAC019* or *SKP21* in *Arabidopsis* could contribute to alkaline stress tolerance, but reduced ABA sensitivity(Cao et al. 2017; Liu et al. 2015). In conclusion, these results speculated that wild soybean *NHL* family genes have crosstalk with MeJA or ABA signal transduction under alkaline stress, and some genes may display different roles in ABA or MeJA signal transduction in response to alkaline stress.

Conclusions

In conclusion, in this study, we identified 59 potential *NHL* family genes in wild soybean. We identified the phylogenetic relationship, conserved domains, gene duplication events and *cis*-acting elements in promoter regions. We also confirmed that wild soybean *NHL* family genes may play important regulatory roles in response to alkaline stress, ABA and MEJA treatment. Taken together, our results established a foundation for characterization of wild soybean *NHL* family genes in response to alkaline stress, ABA and MEJA treatment. However, the function analysis of up-regulated genes under ABA, MEJA or alkaline stress, such as *GsNHL4*, *GsNHL44* and *GsNHL51*, is of great significance in the future. On the other hand, more jobs are need to do for exploring the potential roles of NHL family genes, especially the roles in crosstalk with MeJA or ABA signal transduction pathways under alkaline stress in wild soybean.

Abbreviations

NHL NDR1/HIN1-like

qRT-PCR quantitative real-time PCR

MW molecular weight

pl isoelectric point

ABA abscisic acid

MeJA methyl jasmonate

SA salicylie acid

GA gibberellin



MBS drought

LTR low temperature responsive

TC-rich defense and stress responsive

373

374

375

380 381

382

383

384

385

386

393

394

395

400

401

402

403

404

405

References

- Artimo P, Jonnalagedda M, Arnold K, Baratin D, Csardi G, de Castro E, Duvaud S, Flegel V, Fortier A,
 Gasteiger E, Grosdidier A, Hernandez C, Ioannidis V, Kuznetsov D, Liechti R, Moretti S, Mostaguir
 K, Redaschi N, Rossier G, Xenarios I, and Stockinger H. 2012. ExPASy: SIB bioinformatics resource
 portal. *Nucleic Acids Research* 40:W597-W603. 10.1093/nar/gks400
 - Bailey TL, Boden M, Buske FA, Frith M, Grant CE, Clementi L, Ren J, Li WW, and Noble WS. 2009. MEME SUITE: tools for motif discovery and searching. *Nucleic Acids Research* 37:W202-W208. 10.1093/nar/gkp335
 - Bao Y, Song W-M, Pan J, Jiang C-M, Srivastava R, Li B, Zhu L-Y, Su H-Y, Gao X-S, Liu H, Yu X, Yang L, Cheng X-H, and Zhang H-X. 2016. Overexpression of the NDR1/HIN1-Like Gene NHL6 Modifies Seed Germination in Response to Abscisic Acid and Abiotic Stresses in Arabidopsis. *Plos One* 11. 10.1371/journal.pone.0148572
- Bowers JE, Chapman BA, Rong JK, and Paterson AH. 2003. Unravelling angiosperm genome evolution by phylogenetic analysis of chromosomal duplication events. *Nature* 422:433-438. 10.1038/nature01521
- 390 Brown APC, Dunn MA, Goddard NJ, and Hughes MA. 2001. Identification of a novel low-temperature-391 response element in the promoter of the barley (Hordeum vulgare L) gene blt101.1. *Planta* 392 213:770-780. 10.1007/s004250100549
 - Cao L, Yu Y, Ding X, Zhu D, Yang F, Liu B, Sun X, Duan X, Yin K, and Zhu Y. 2017. The Glycine soja NAC transcription factor GsNAC019 mediates the regulation of plant alkaline tolerance and ABA sensitivity. *Plant Molecular Biology* 95:253-268. 10.1007/s11103-017-0643-3
- Century KS, Holub EB, and Staskawicz BJ. 1995. NDR1, A LOCUS OF ARABIDOPSIS-THALIANA THAT IS
 REQUIRED FOR DISEASE RESISTANCE TO BOTH A BACTERIAL AND A FUNGAL PATHOGEN.

 Proceedings of the National Academy of Sciences of the United States of America 92:6597-6601.

 10.1073/pnas.92.14.6597
 - Chen L, He F, Long R, Zhang F, Li M, Wang Z, Kang J, and Yang Q. 2021. A global alfalfa diversity panel reveals genomic selection signatures in Chinese varieties and genomic associations with root development. *Journal of Integrative Plant Biology* 63:1937-1951. 10.1111/jipb.13172
 - Chen Q, Tian Z, Jiang R, Zheng X, Xie C, and Liu J. 2018. StPOTHR1, a NDR1/HIN1-like gene in Solanum tuberosum, enhances resistance against Phytophthora infestans. *Biochemical and Biophysical Research Communications* 496:1155-1161. 10.1016/j.bbrc.2018.01.162
- 406 Chong J, Le Henanff G, Bertsch C, and Walter B. 2008. Identification, expression analysis and characterization of defense and signaling genes in Vitis vinifera. *Plant Physiology and Biochemistry* 46:469-481. 10.1016/j.plaphy.2007.09.010



423

424

425

426

427

428

429

430

431 432

433

434

435

436

437

438

439

440

441

442

443

444

- 409 Chourey K, Ramani S, and Apte SK. 2003. Accumulation of LEA proteins in salt (NaCl) stressed young 410 seedlings of rice (Oryza sativa L.) cultivar Bura Rata and their degradation during recovery from 411 salinity stress. *Journal of Plant Physiology* 160:1165-1174. 10.1078/0176-1617-00909
- Dagvadorj B, Outram MA, Williams SJ, and Solomon PS. 2022. The necrotrophic effector ToxA from Parastagonospora nodorum interacts with wheat NHL proteins to facilitate Tsn1-mediated necrosis. *Plant Journal* 110:407-418. 10.1111/tpj.15677
- Dormann P, Gopalan S, He SY, and Benning C. 2000. A gene family in Arabidopsis thaliana with sequence similarity to NDR1 and HIN1. *Plant Physiology and Biochemistry* 38:789-796. 10.1016/s0981-9428(00)01186-4
- DuanMu H, Wang Y, Bai X, Cheng S, Deyholos MK, Wong GK-S, Li D, Zhu D, Li R, Yu Y, Cao L, Chen C, and Zhu Y. 2015. Wild soybean roots depend on specific transcription factors and oxidation reduction related genesin response to alkaline stress. *Functional & Integrative Genomics* 15:651-660. 10.1007/s10142-015-0439-y
 - Finn RD, Clements J, and Eddy SR. 2011. HMMER web server: interactive sequence similarity searching. *Nucleic Acids Research* 39:W29-W37. 10.1093/nar/gkr367
 - Finn RD, Coggill P, Eberhardt RY, Eddy SR, Mistry J, Mitchell AL, Potter SC, Punta M, Qureshi M, Sangrador-Vegas A, Salazar GA, Tate J, and Bateman A. 2016. The Pfam protein families database: towards a more sustainable future. *Nucleic Acids Research* 44:D279-D285. 10.1093/nar/gkv1344
 - Ge Y, Li Y, Zhu Y-M, Bai X, Lv D-K, Guo D, Ji W, and Cai H. 2010. Global transcriptome profiling of wild soybean (Glycine soja) roots under NaHCO3 treatment. *Bmc Plant Biology* 10. 10.1186/1471-2229-10-153
 - Gopalan S, Bauer DW, Alfano JR, Loniello AO, He SY, and Collmer A. 1996. Expression of the Pseudomonas syringae avirulence protein AvrB in plant cells alleviates its dependence on the hypersensitive response and pathogenicity (Hrp) secretion system in eliciting genotype-specific hypersensitive cell death. *Plant Cell* 8:1095-1105.
 - Gopavajhula VR, Chaitanya KV, Khan PAA, Shaik JP, Reddy PN, and Alanazi M. 2013. Modeling and analysis of soybean (Glycine max. L) Cu/Zn, Mn and Fe superoxide dismutases. *Genetics and Molecular Biology* 36:225-236. 10.1590/s1415-47572013005000023
 - Huis R, Hawkins S, and Neutelings G. 2010. Selection of reference genes for quantitative gene expression normalization in flax (Linum usitatissimum L.). *Bmc Plant Biology* 10. 10.1186/1471-2229-10-71
 - Jin H, Plaha P, Park JY, Hong CP, Lee IS, Yang ZH, Jiang GB, Kwak SS, Liu SK, Lee JS, Kim YA, and Lim YP. 2006. Comparative EST profiles of leaf and root of Leymus chinensis, a xerophilous grass adapted to high pH sodic soil. *Plant Science* 170:1081-1086. 10.1016/j.plantsci.2006.01.002
 - Jin X, Cao D, Wang Z, Ma L, Tian K, Liu Y, Gong Z, Zhu X, Jiang C, and Li Y. 2019. Genome-wide identification and expression analyses of the LEA protein gene family in tea plant reveal their involvement in seed development and abiotic stress responses. *Scientific Reports* 9. 10.1038/s41598-019-50645-8
- Krzywinski M, Schein J, Birol I, Connors J, Gascoyne R, Horsman D, Jones SJ, and Marra MA. 2009. Circos:
 An information aesthetic for comparative genomics. *Genome Research* 19:1639-1645.
 10.1101/gr.092759.109
- Ku Y-S, Sintaha M, Cheung M-Y, and Lam H-M. 2018. Plant Hormone Signaling Crosstalks between Biotic and Abiotic Stress Responses. *International Journal of Molecular Sciences* 19. 10.3390/ijms19103206
- Kumar S, Nei M, Dudley J, and Tamura K. 2008. MEGA: A biologist-centric software for evolutionary analysis of DNA and protein sequences. *Briefings in Bioinformatics* 9:299-306. 10.1093/bib/bbn017



469

470

471

475 476

477

478

479

480

481 482

483

484

485

486 487

488

489

490

491

492

493 494

495

496 497

- Lee SB, Ham BK, Park JM, Kim YJ, and Paek KH. 2006. BnNHL18A shows a localization change by stressinducing chemical treatments. *Biochemical and Biophysical Research Communications* 339:399-457 406. 10.1016/j.bbrc.2005.10.210
- Lescot M, Dehais P, Thijs G, Marchal K, Moreau Y, Van de Peer Y, Rouze P, and Rombauts S. 2002.

 PlantCARE, a database of plant cis-acting regulatory elements and a portal to tools for in silico analysis of promoter sequences. *Nucleic Acids Research* 30:325-327. 10.1093/nar/30.1.325
- 461 Liu A, Yu Y, Duan X, Sun X, Duanmu H, and Zhu Y. 2015. GsSKP21, a Glycine soja S-phase kinase-associated
 462 protein, mediates the regulation of plant alkaline tolerance and ABA sensitivity. *Plant Molecular* 463 *Biology* 87:111-124. 10.1007/s11103-014-0264-z
- Liu C, Peang H, Li X, Liu C, Lv X, Wei X, Zou A, Zhang J, Fan G, Ma G, Ma L, and Sun X. 2020a. Genome-wide analysis of NDR1/HIN1-like genes in pepper (Capsicum annuum L.) and functional characterization of CaNHL4 under biotic and abiotic stresses. *Horticulture research* 7:93-93. 10.1038/s41438-020-0318-0
 - Liu C, Peng H, Li X, Liu C, Lv X, Wei X, Zou A, Zhang J, Fan G, Ma G, Ma L, and Sun X. 2020b. Genome-wide analysis of NDR1/HIN1-like genes in pepper (Capsicum annuum L.) and functional characterization of CaNHL4 under biotic and abiotic stresses. *Horticulture research* 7:93-93. 10.1038/s41438-020-0318-0
- Liu J, Shen F, Xiao Y, Fang H, Qiu C, Li W, Wu T, Xu X, Wang Y, Zhang X, and Han Z. 2020c. Genomicsassisted prediction of salt and alkali tolerances and functional marker development in apple rootstocks. *Bmc Genomics* 21. 10.1186/s12864-020-06961-9
 - Maldonado A, Youssef R, McDonald M, Brewer E, Beard H, and Matthews B. 2014. Modification of the expression of two NPR1 suppressors, SNC1 and SNI1, in soybean confers partial resistance to the soybean cyst nematode, Heterodera glycines. *Functional Plant Biology* 41:714-726. 10.1071/fp13323
 - McNeece BT, Pant SR, Sharma K, Niruala P, Lawrence GW, and Klink VP. 2017. A Glycine max homolog of NON-RACE SPECIFIC DISEASE RESISTANCE 1 (NDR1) alters defense gene expression while functioning during a resistance response to different root pathogens in different genetic backgrounds. *Plant Physiology and Biochemistry* 114:60-71. 10.1016/j.plaphy.2017.02.022
 - Qiao Y, Lu W, Wang R, Nisa Z, Yu Y, Jin X, Yu L, and Chen C. 2020. Identification and Expression Analysis of Strigolactone Biosynthetic and Signaling Genes in Response to Salt and Alkaline Stresses in Soybean (Glycine max). *DNA and Cell Biology* 39:1850-1861. 10.1089/dna.2020.5637
 - Saeed AI, Hagabati NK, Braisted JC, Liang W, Sharov V, Howe EA, Li J, Thiagarajan M, White JA, and Quackenbush J. 2006. TM4 microarray software suite. In: Kimmel A, and Oluver B, eds. *DNA Microarrays, Part B: Databases and Statistics*, 134-+.
 - Sazegari S, Niazi A, and Ahmadi FS. 2015. A study on the regulatory network with promoter analysis for Arabidopsis DREB-genes. *Bioinformation* 11:101-106. 10.6026/97320630011101
 - Selote D, Shine MB, Robin GP, and Kachroo A. 2014. Soybean NDR1-like proteins bind pathogen effectors and regulate resistance signaling. *New Phytologist* 202:485-498. 10.1111/nph.12654
 - Sun C, Zhang C, Wang X, Zhao X, Chen F, Zhang W, Hu M, Fu S, Yi B, and Zhang J. 2021. Genome-Wide Identification and Characterization of the IGT Gene Family in Allotetraploid Rapeseed (Brassica napus L.). *DNA and Cell Biology* 40:441-456. 10.1089/dna.2020.6227
 - Sun X, Luo X, Sun M, Chen C, Ding X, Wang X, Yang S, Yu Q, Jia B, Ji W, Cai H, and Zhu Y. 2014. A Glycine Soja 14-3-3 Protein GsGF14o Participates in Stomatal and Root Hair Development and Drought Tolerance in Arabidopsis thaliana. *Plant and Cell Physiology* 55:99-118. 10.1093/pcp/pct161
- Takahashi Y, Berberich T, Yamashita K, Uehara Y, Miyazaki A, and Kusano T. 2004. Identification of tobacco
 HIN1 and two closely related genes as spermine-responsive genes and their differential
 expression during the Tobacco mosaic virus-induced hypersensitive response and during leaf- and
 flower-senescence. *Plant Molecular Biology* 54:613-622. 10.1023/b:Plan.0000038276.95539.39

517

518

528

529

530

531

- Varet A, Hause B, Hause G, Scheel D, and Lee J. 2003. The Arabidopsis NHL3 gene encodes a plasma membrane protein and its overexpression correlates with increased resistance to Pseudomonas syringae pv. tomato DC3000. *Plant Physiology* 132:2023-2033. 10.1104/pp.103.020438
- Varet A, Parker J, Tornero P, Nass N, Nurnberger T, Dangl JL, Scheel D, and Lee J. 2002. NHL25 and NHL3,
 two NDR1/HIN1-like genes in Arabidopsis thaliana with potential role(s) in plant defense.
 Molecular Plant-Microbe Interactions 15:608-616. 10.1094/mpmi.2002.15.6.608
- Vondrackova S, Szakova J, Drabek O, Tejnecky V, Hejcman M, Muellerova V, and Tlustos P. 2015.
 Aluminium Uptake and Translocation in Al Hyperaccumulator Rumex obtusifolius Is Affected by
 Low-Molecular-Weight Organic Acids Content and Soil pH. *Plos One* 10.
 10.1371/journal.pone.0123351
- Wang J, Wu R, Shangguan T, Chen G, Zheng Y, Tao X, Li S, Wang Y, and Xu S. 2022. NDR1/HIN1-like genes
 may regulate Glycine max seed germination under chilling stress through the ABA pathway. *Plant Growth Regulation*. 10.1007/s10725-022-00894-x
 - Wang Y, Ma H, Liu G, Xu C, Zhang D, and Ban Q. 2008. Analysis of Gene Expression Profile of Limonium bicolor under NaHCO3 Stress Using cDNA Microarray. *Plant Molecular Biology Reporter* 26:241-254. 10.1007/s11105-008-0037-4
- Wen Z, Ding Y, Zhao T, and Gai J. 2009. Genetic diversity and peculiarity of annual wild soybean (G. soja Sieb. et Zucc.) from various eco-regions in China. *Theoretical and Applied Genetics* 119:371-381. 10.1007/s00122-009-1045-y
- Yang CW, Wang P, Li CY, Shi DC, and Wang DL. 2008. Comparison of effects of salt and alkali stresses on the growth and photosynthesis of wheat. *Photosynthetica* 46:107-114. 10.1007/s11099-008-0018-8
- 525 Yu M, Liu Z, Jiang S, Xu N, Chen Q, Qi Z, and Lv W. 2018. QTL mapping and candidate gene mining for 526 soybean seed weight per plant. *Biotechnology & Biotechnological Equipment* 32:908-914. 527 10.1080/13102818.2018.1438851
 - Yu Y, Duan X, Ding X, Chen C, Zhu D, Yin K, Cao L, Song X, Zhu P, Li Q, Nisa ZU, Yu J, Du J, Song Y, Li H, Liu B, and Zhu Y. 2017. A novel AP2/ERF family transcription factor from Glycine soja, GsERF71, is a DNA binding protein that positively regulates alkaline stress tolerance in Arabidopsis. *Plant Molecular Biology* 94:509-530. 10.1007/s11103-017-0623-7
- Zheng J, Su H, Lin R, Zhang H, Xia K, Jian S, and Zhang M. 2019. Isolation and characterization of an atypical
 LEA gene (IpLEA) from Ipomoea pes-caprae conferring salt/drought and oxidative stress tolerance.
 Scientific Reports 9. 10.1038/s41598-019-50813-w
- Zheng MS, Takahashi H, Miyazaki A, Hamamoto H, Shah J, Yamaguchi I, and Kusano T. 2004. Up-regulation
 of Arabidopsis thaliana NHL10 in the hypersensitive response to Cucumber mosaic virus infection
 and in senescing leaves is controlled by signalling pathways that differ in salicylate involvement.
 Planta 218:740-750. 10.1007/s00425-003-1169-2
- Zhu D, Bai X, Chen C, Chen Q, Cai H, Li Y, Ji W, Zhai H, Lv D, Luo X, and Zhu Y. 2011. GsTIFY10, a novel
 positive regulator of plant tolerance to bicarbonate stress and a repressor of jasmonate signaling.
 Plant Molecular Biology 77:285-297. 10.1007/s11103-011-9810-0
- Zhu D, Li R, Liu X, Sun M, Wu J, Zhang N, and Zhu Y. 2014. The Positive Regulatory Roles of the TIFY10
 Proteins in Plant Responses to Alkaline Stress. *Plos One* 9. 10.1371/journal.pone.0111984



Table 1(on next page)

Protein information of NHL family genes in wild soybean.

Table 1 Protein information of NHL family genes in wild soybean.

Number	Gene ID	Gene Name	Chr	Amino acid residues	MW (kDa)	pl
1	GsNHL1	GlysoPl483463.01G116400	1	230	25.81956	8.95
2	GsNHL2	GlysoPl483463.01G198800	1	190	21.52359	10.15
3	GsNHL3	GlysoPl483463.02G162500	2	208	23.91979	9.79
4	GsNHL4	GlysoPl483463.02G162700	2	245	27.62919	9.26
5	GsNHL5	GlysoPl483463.02G199000	2	256	28.76769	10.12
6	GsNHL6	GlysoPl483463.02G230000	2	274	30.45435	9.86
7	GsNHL7	GlysoPI483463.03G161800	3	222	24.93808	9.58
8	GsNHL8	GlysoPl483463.03G162100	3	208	24.13909	9.46
9	GsNHL9	GlysoPl483463.03G162300	3	230	26.77082	9.14
10	GsNHL10	GlysoPl483463.03G162400	3	204	23.49148	9.53
11	GsNHL11	GlysoPl483463.03G162500	3	210	23.77761	9.81
12	GsNHL12	GlysoPI483463.03G213100	3	239	26.31572	9.37
13	GsNHL13	GlysoPl483463.03G218900	3	198	21.55946	9.83
14	GsNHL14	GlysoPl483463.03G162200	3	228	26.24044	9.50
15	GsNHL15	GlysoPl483463.04G093700	4	256	28.40712	9.51
16	GsNHL16	GlysoPl483463.04G185500	4	211	23.22106	9.69
17	GsNHL17	GlysoPl483463.05G146000	5	348	40.00232	10.00
18	GsNHL18	GlysoPI483463.05G185300	5	214	22.94057	9.54
19	GsNHL19	GlysoPl483463.06G095800	6	260	28.63512	9.04
20	GsNHL20	GlysoPl483463.06G124800	6	224	25.31161	8.97
21	GsNHL21	GlysoPI483463.07G008800	7	255	27.95721	9.68
22	GsNHL22	GlysoPI483463.07G045200	7	253	28.11260	9.46
23	GsNHL23	GlysoPl483463.07G092400	7	210	24.10902	9.43
24	GsNHL24	GlysoPl483463.07G092500	7	208	24.15102	9.82
25	GsNHL25	GlysoPl483463.09G134000	9	204	23.68964	9.53
26	GsNHL26	GlysoPI483463.09G151000	9	315	34.59544	9.85

27	GsNHL27	GlysoPl483463.10G070800	10	247	27.67418	9.09
28	GsNHL28	GlysoPl483463.10G070900	10	216	24.78271	8.71
29	GsNHL29	GlysoPl483463.10G071000	10	228	26.25031	9.64
30	GsNHL30	GlysoPl483463.10G071100	10	210	24.00494	10.02
31	GsNHL31	GlysoPl483463.10G071200	10	200	22.61333	7.82
32	GsNHL32	GlysoPl483463.10G071700	10	223	24.65133	9.10
33	GsNHL33	GlysoPl483463.10G105800	10	149	16.37778	9.39
34	GsNHL34	GlysoPl483463.10G214600	10	228	26.04372	9.56
35	GsNHL35	GlysoPl483463.11G015000	11	179	19.59170	9.55
36	GsNHL36	GlysoPl483463.11G145400	11	215	23.56234	9.96
37	GsNHL37	GlysoPl483463.11G186300	11	246	27.22966	8.88
38	GsNHL38	GlysoPl483463.12G084400	12	214	23.57643	9.96
39	GsNHL39	GlysoPl483463.12G153200	12	222	24.646.62	9.34
40	GsNHL40	GlysoPl483463.12G178100	12	218	24.24817	9.81
41	GsNHL41	GlysoPl483463.13G250400	13	233	26.33392	7.93
42	GsNHL42	GlysoPl483463.13G263900	13	272	29.99915	10.01
43	GsNHL43	GlysoPl483463.13G298400	13	255	27.84143	10.24
44	GsNHL44	GlysoPl483463.14G036900	14	274	30.37919	9.65
45	GsNHL45	GlysoPl483463.14G166200	14	259	29.21918	10.15
46	GsNHL46	GlysoPl483463.15G014800	15	310	34.24409	9.95
47	GsNHL47	GlysoPl483463.16G088600	16	208	22.74147	9.23
48	GsNHL48	GlysoPl483463.16G179000	16	193	20.53805	9.30
49	GsNHL49	GlysoPl483463.18G044000	18	245	27.85761	9.97
50	GsNHL50	GlysoPl483463.18G046800	18	238	26.32375	8.06
51	GsNHL51	GlysoPl483463.19G161400	19	215	24.27526	9.74
52	GsNHL52	GlysoPI483463.19G161500	19	228	26.41171	9.34
53	GsNHL53	GlysoPl483463.19G161600	19	228	26.43265	9.34
54	GsNHL54	GlysoPI483463.19G161700	19	281	23.90965	9.81
55	GsNHL55	GlysoPl483463.19G161800	19	282	31.18464	8.22
56	GsNHL56	GlysoPI483463.19G209900	19	245	26.96660	9.77

E - 10				
		0	M	ı
-	C	C	Ш.	,

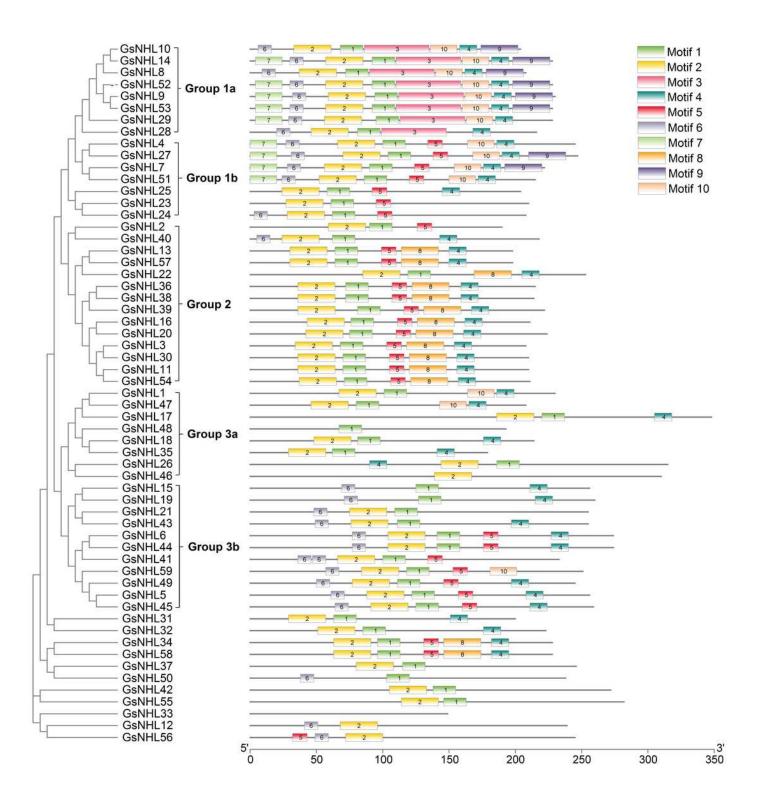
Manuscript to be reviewed

57	GsNHL57	GlysoPl483463.19G216600	19	198	21.53948	9.89
58	GsNHL58	GlysoPl483463.20G108700	20	228	26.04367	9.36
59	GsNHL59	GlysoPI483463.20G179000	20	251	27.97813	9.74

Figure 1. The phylogenetic and conserved domain analysis of NHL family proteins.

a The phylogenetic analysis of wild soybean *NHL* family proteins. The maximum-likelihood (ML) phylogenetic tree was constructed based on 1000 replications for each branch. **b** The motif composition of *NHL* family proteins was identified using MEME online software, and the motif were displayed by boxes of different numbers and colors. The TBtools software was used to combine the conserved motifs and phylogenetic tree.



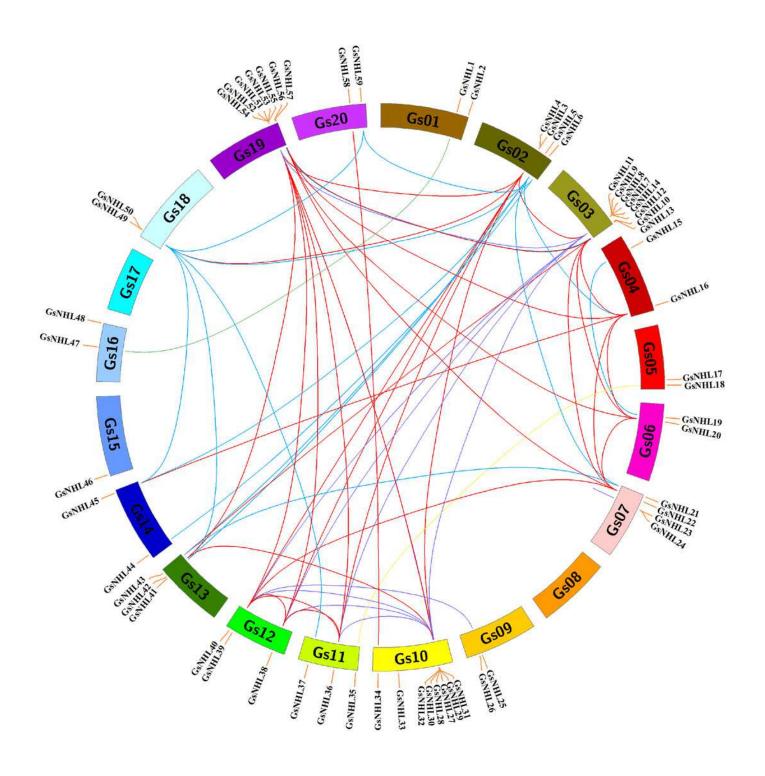




Chromosomal locations and syntenic analysis of NHL family genes in wild soybean.

The Circos-0.69 software was used to generate the chromosomes as a circle. The pair of duplication genes were are identified and connected by different color lines.

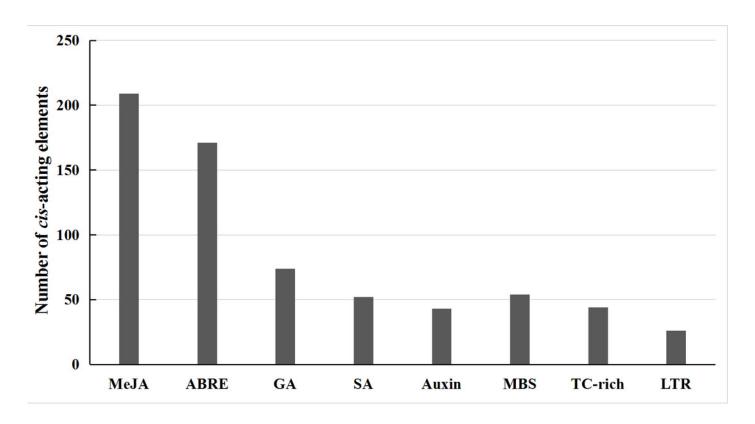






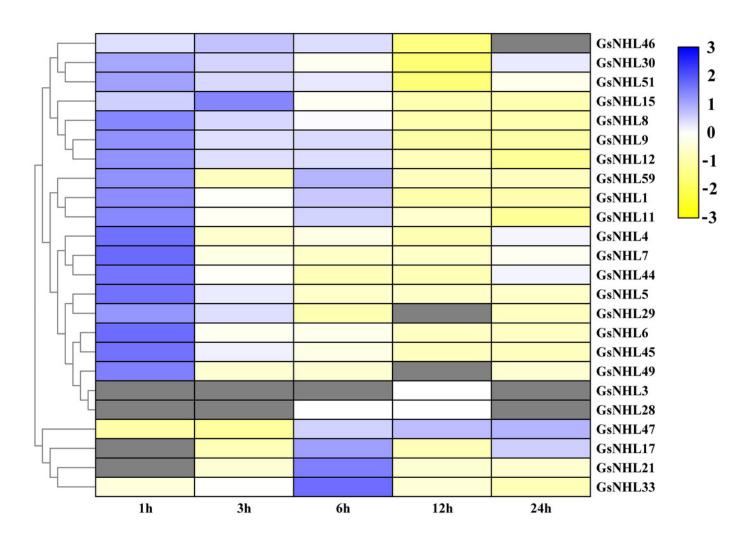
Analysis of *cis*-acting elements of putative *NHL* family gene promoters related to hormones and abiotic stress responses.

The potential regulatory *cis*-acting elements were analyzed in the 3000 bp upstream of translation start site by using the PLANT CARE online software.



Expression patterns of NHL family genes in response to alkaline stress.

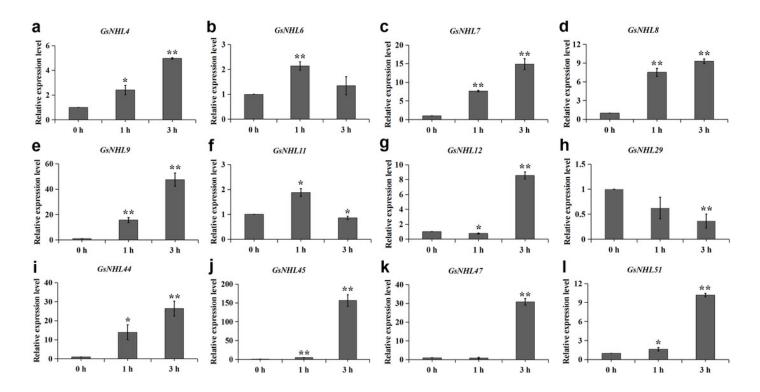
The wild soybean transcriptome data were used to detect the expression pattern of NHL family genes under alkaline stress. The TM4: MeV4.9 software was used to generate the heat map. The blue and yellow colors represent high or low expression levels (|Log2| fold change |Log2| > 2, P < 0.05), respectively.





Expression analysis of wild soybean NHL family genes in response to alkaline stress.

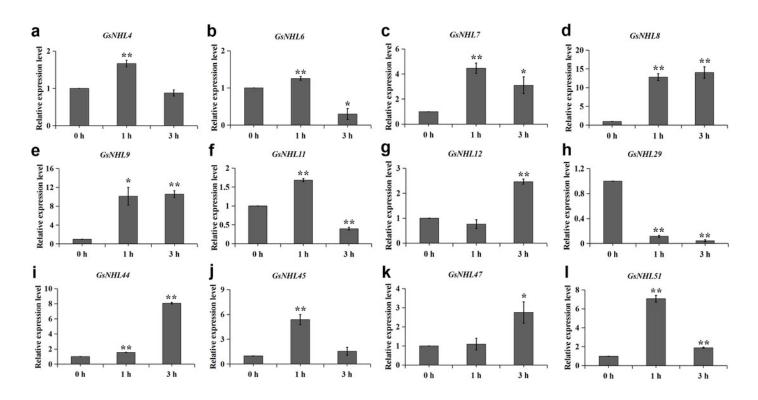
a-I The wild soybean seedlings were treated with 50 mM NaHCO₃ for 0, 1 and 3 h. The qRT-PCR results were analyzed using the $2^{-\Delta\Delta CT}$ method and Student's t-test.





Expression analysis of wild soybean NHL family genes in response to MeJA.

The wild soybean seedlings were treated with 50 μ M MeJA for 0, 1 and 3 h. The qRT-PCR results were analyzed using the 2- $\Delta\Delta$ CT method and Student's t-test.





Expression analysis of wild soybean NHL family genes in response to ABA.

a-I The wild soybean seedlings were treated with 50 μ M ABA for 0, 1 and 3 h. The qRT-PCR results were analyzed using the $2^{-\Delta\Delta CT}$ method and Student's t-test.

