

Irradiation with carbon ion beams affects soybean nutritional quality in early generations (#73152)

1

First submission

Guidance from your Editor

Please submit by **29 May 2022** for the benefit of the authors (and your \$200 publishing discount) .



Structure and Criteria

Please read the 'Structure and Criteria' page for general guidance.



Raw data check

Review the raw data.



Image check

Check that figures and images have not been inappropriately manipulated.

Privacy reminder: If uploading an annotated PDF, remove identifiable information to remain anonymous.

Files

Download and review all files from the [materials page](#).

5 Figure file(s)

1 Raw data file(s)



Structure and Criteria

Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

1. **BASIC REPORTING**
2. **EXPERIMENTAL DESIGN**
3. **VALIDITY OF THE FINDINGS**
4. General comments
5. Confidential notes to the editor

 You can also annotate this PDF and upload it as part of your review

When ready [submit online](#).

Editorial Criteria

Use these criteria points to structure your review. The full detailed editorial criteria is on your [guidance page](#).

BASIC REPORTING

-  Clear, unambiguous, professional English language used throughout.
-  Intro & background to show context. Literature well referenced & relevant.
-  Structure conforms to [Peerj standards](#), discipline norm, or improved for clarity.
-  Figures are relevant, high quality, well labelled & described.
-  Raw data supplied (see [Peerj policy](#)).

EXPERIMENTAL DESIGN

-  Original primary research within [Scope of the journal](#).
-  Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
-  Rigorous investigation performed to a high technical & ethical standard.
-  Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

-  Impact and novelty not assessed. *Meaningful* replication encouraged where rationale & benefit to literature is clearly stated.
-  All underlying data have been provided; they are robust, statistically sound, & controlled.
-  Conclusions are well stated, linked to original research question & limited to supporting results.



The best reviewers use these techniques

Tip

Example

Support criticisms with evidence from the text or from other sources

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Give specific suggestions on how to improve the manuscript

Your introduction needs more detail. I suggest that you improve the description at lines 57- 86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

Comment on language and grammar issues

The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult. I suggest you have a colleague who is proficient in English and familiar with the subject matter review your manuscript, or contact a professional editing service.

Organize by importance of the issues, and number your points

1. Your most important issue
2. The next most important item
3. ...
4. The least important points

Please provide constructive criticism, and avoid personal opinions

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

Comment on strengths (as well as weaknesses) of the manuscript

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

Irradiation with carbon ion beams affects soybean nutritional quality in early generations

Changkai Liu¹, Xue Wang^{1,2}, Yansheng Li¹, Heng Chen^{1,2}, Qiuying Zhang^{Corresp., 1, 3}, Xiaobing Liu¹

¹ Key Laboratory of Mollisols Agroecology, Northeast Institute of Geography and Agroecology, CAS, Harbin, China

² University of Chinese Academy of Sciences, Beijing, China

³ Innovation Academy for Seed Design, CAS, Harbin, China

Corresponding Author: Qiuying Zhang

Email address: zhangqiuying1024@outlook.com

With the increased demand for healthy life, seed nutritional quality of soybean has become as important as yield trait. This study aimed to develop soybean lines with beneficially altered seed compositions by using an early carbon ion beam mutant population generated by different irradiation doses (100, 120 and 140 Gy). Eleven quality traits, including protein, oil, sucrose, soluble sugar, Fe, Mn, Zn, Cu, daidzein, glycitin, and genistin concentrations were analyzed to evaluate the variation of seed nutritional quality of the M₂ and M₃ generations. Protein and oil concentrations changed by 38.5-42.9 % and 18.8-23.8 % in M₂ and M₃ generations, respectively, while soluble sugar and sucrose concentrations changed by 48.1-123.4 mg/g and 22.7-74.7 mg/g, with significant effects by 140 Gy across two generations. In two generations, the responses of Fe, Mn, Zn, and Cu to irradiation doses were different, but the inhibitory effect on Mn concentration was enhanced as the irradiation dose was raised. In general, irradiation raised isoflavone concentrations, but 140 Gy had an inhibitory effect on isoflavone concentrations in the M3 generation. Other indicators, with the exception of sucrose and soluble sugar, were still separated from the M2 generation to the M3 generation. Some nutritional quality traits of soybean can be screened in early generations using ion carbon beam irradiation. A lower irradiation dose is preferable when breeding targets are higher isoflavones and Mn concentrations. A higher irradiation dose can be tried if the breeding targets are higher protein, oil, sucrose, soluble sugar, Fe, Zn, and Cu.

Irradiation with carbon ion beams affects soybean nutritional quality in early generations

Changkai Liu¹, Xue Wang^{1,2}, Yansheng Li¹, Heng Chen^{1,2}, Qiuying Zhang^{1,3*}, Xiaobing Liu¹

1. Key Laboratory of Mollisols Agroecology, Northeast Institute of Geography and Agroecology, CAS, Harbin 150081, China

2. University of Chinese Academy of Sciences, Beijing 100049, China

3. Innovation Academy for Seed Design, CAS, Harbin, 150081, China

Corresponding Author:

Qingying Zhang

Haping Road 138, Harbin, Heilongjiang, 150081, China; Tel: 86-451-86601320; Fax: +86-451-86603736

Email address: zhangqiuying@iga.ac.cn

ABSTRACT

With the increased demand for healthy life, seed nutritional quality of soybean has become as important as yield trait. This study aimed to develop soybean lines with beneficially altered seed compositions by using an early carbon ion beam mutant population generated by different irradiation doses (100, 120 and 140 Gy). Eleven quality traits, including protein, oil, sucrose, soluble sugar, Fe, Mn, Zn, Cu, daidzein, glycitin, and genistin concentrations were analyzed to evaluate the variation of seed nutritional quality of the M₂ and M₃ generations. Protein and oil concentrations changed by 38.5-42.9 % and 18.8-23.8 % in M₂ and M₃ generations, respectively, while soluble sugar and sucrose concentrations changed by 48.1-123.4 mg/g and 22.7-74.7 mg/g, with significant effects by 140 Gy across two generations. In two generations, the responses of Fe, Mn, Zn, and Cu to irradiation doses were different, but the inhibitory effect on Mn concentration was enhanced as the irradiation dose was raised. In general, irradiation raised isoflavone concentrations, but 140 Gy had an inhibitory effect on isoflavone concentrations in the M₃ generation. Other indicators, with the exception of sucrose and soluble sugar, were still separated from the M₂ generation to the M₃ generation. Some nutritional quality traits of soybean can be screened in early generations using ion carbon beam irradiation. A lower irradiation dose is preferable when breeding targets are higher isoflavones and Mn concentrations. A higher irradiation dose can be tried if the breeding targets are higher protein, oil, sucrose, soluble sugar, Fe, Zn, and Cu.

Keywords: irradiation, soybean, protein, oil, isoflavones, carbohydrates, trace elements

1. Introduction

Soybean is a unique legume crop because of its diverse nutritional value including protein, oil, isoflavones, trace element, and metabolizable energy (Schmutz et al., 2010; Liu et al., 2019). The seed protein concentration of soybean is about 40%, which is about 2-4 times higher than that of corn, rice and wheat. Soybean seed oil concentration is about 20%, contains 8 kinds of amino acids necessary for human, especially lazy acid and tryptophan which cannot be synthesized by human body (Patil et al., 2017). The percentage of carbohydrates in soybean seed is about 20-30%, but the functions are diverse owing to its complex composition (Cober et al., 2000). The ratio of sucrose, starch and dietary fiber in soybean seed influence its nutritional value greatly (Karr-Lilienthal et al., 2005). Soybean seeds are also rich in isoflavones and trace elements, which are essential for human diet (Wu et al., 2020).

Plant germplasms are the most important sources in crop breeding. Mutation breeding compared to cross breeding has an irreplaceable position in breaking the bottleneck of existing germplasm resources and thus creating more beneficial resources. Higher mutation rate and wider mutation spectrum of soybean seed nutritional quality have been reported by physical mutagenesis and chemical mutagenesis (Hajduch et al., 2000; Espina et al., 2018). Carbon ion beams (CIBs), a new mutation mutagenic resource with higher linear energy transfer (LET), higher mutation rates, and wider mutation spectra under lighter damage, has been increasingly applied to the mutation breeding in various plants including soybean (Arase et al. 2011). Therefore, studying the effects of carbon ion beam irradiation on soybean seed quality traits could have greater potential in accelerating the breeding of high-quality soybean varieties.

There was evidence that combined γ -radiation and EMS improved the concentration of soybean oil with higher levels of oleic and lower levels of linolenic acid (Patil et al., 2007). It has also been reported that irradiation of grain cereals and leguminous crops seeds leads to increased protein content and higher carbohydrate and vitamin levels (Jan et al., 2012). Mikuriya et al. (2017) demonstrated that the populations mutagenized by carbon ion beam irradiation exhibited lower isoflavone concentration in soybean seeds, which was closely related to the reduction of leaf chlorophyll concentration. The increased concentration of genistin, genistein, daidzin and glycitein, but decreased concentration of glycitein and daidzein in soybean seedlings were found by laser irradiation (Jin et al., 2011). The increased irradiation dose increased the concentration of Fe, Cu and Zn in soybean seedlings by mutation breeding (Alikamanoglu et al., 2011). Therefore, carbon ion beam irradiation could have greater potential in accelerating the breeding of high-quality soybean varieties. However, the general information of CIBs on soybean population is not available, especially on seed trace elements and isoflavones.

The present study investigated the seed protein, oil, isoflavones, carbohydrates and trace elements in M₂ and M₃ generation generated by carbon ion beam mutagen treatment with the aims in developing soybean lines with beneficially altered seed composition.

2. Materials & Methods

2.1. Plant material and experiment design

Populations of plants harboring mutants of soybean cultivar ‘Dongsheng28’ were previously obtained through carbon ion beam irradiation (CIBR) (Wang et al., 2021). The irradiation was directly targeted at the hilum of each seed with 960 MeV carbon ion beam, the dose for mutagenesis was 0 Gy (control), 100 Gy, 120 Gy and 140 Gy separately.

The treated seeds were planted in the field at the Agronomy Farm of the Northeast Institute of Geography and Agroecology, Chinese Academy of Science (45°73’N, 126°61’E). In the M₁ generation, M₀ seeds were planted according to the irradiation dose in 2018, the seeds of all survival plants were harvested separately. In the M₂ generation (2019), the seeds of individual plants harvested from the last generation were sown in a single line. In total, 47 lines in the treatment of 100 Gy group, 26 lines in the treatment of 120 Gy group and 18 lines in the treatment of 140 Gy group. For each single line, five seeds were harvested from every individual plant to form a block. Half of the block were for the determination of seed nutritional quality and the other half were used as seed for the next generation. In the M₃ generation (2020), 100 seeds were sown for every single line, the harvest method was the same as the M₂ generation.

2.2. Chemical analysis of plant samples

Crude protein

The crude protein concentration was determined using the method of combustion nitrogen analysis by Elementar-Vario (Elementar Analysensysteme GmbH E-III, Germany) (Li et al., 2012). A conversion factor of 6.25 was used to convert total nitrogen to crude protein concentration (Saldivar et al., 2011).

Crude oil

Total oil concentration in seeds was determined by the Soxhlet extraction method. Approximately 0.5 g of dried soybean seed sample with a piece of weighed filter paper was wrapped up, then put in a Soxhlet apparatus in a 60 °C water bath. Adding 200 mL ethyl ether to the Soxhlet apparatus for extracting oil. After a 48-h extraction, the defatted sample was placed in an oven at 45 °C about 12 h, the oil concentration was calculated according to Li et al. (2014).

Soluble total sugar and sucrose

The determination of soluble sugar and sucrose was based on the method of Tu *et al.* (2017). About 0.5 g sample was extracted by 4 mL 80 % ethanol, 80 °C water bath for 30 min. Then the mixture was centrifuged at 4500 r/min⁻¹ for 3 min, the supernatant was removed to a new 15 mL -centrifuge tube. Adding 3 mL 80 % ethanol to the precipitate and repeated above operation twice. Volumes up the supernatant to 10 mL finally. Added 4 mL anthrone (1000 mL 80% H₂SO₄ + 2.5 g anthrone) to 1 mL supernatant, bath in 90 °C water for 10 minutes, measured at 620 nm (Xinshiji T6, Beijing). The determination method of sucrose was basically the same as soluble sugar, but before adding anthrone, it was necessary to add 25 µL 12 mol/L of NaOH and water bath at

100 100 °C for 10 min in order to remove the monosaccharides.

101 *Trace element*

102 Trace element was determined with the modified method of Xue et al. (2006). About 0.5 g sample was weighed
103 and placed in a crucible. After carbonization, it was placed in a muffle furnace at 510 °C for 2 h. After cooling,
104 add 1 mL hydrochloric acid and 1 mL deionized water to dissolve the ash. Volumed up to 25 mL after filtering.
105 The analysis of the trace element was performed by AAS (TAS-990, Beijing).

106 *Isoflavones*

107 Isoflavones determination was according to the method described by Hoeck et al. (2000) with high-performance
108 liquid chromatography (HPLC). Weighing about 0.5 g sample into a 15 mL centrifuge tube, and 9 mL 80 %
109 methanol was added. Then ultrasonically extracted at 60 °C for 30 min, centrifuged at 5000 rpm for 5 min, the
110 supernatant was collected into a 25 mL volumetric flask. Adding 6 mL 90 % methanol to precipitation twice.
111 Finally diluted the supernatant to 25 mL with 10 % methanol. The filtrate through 0.45 µm filter membrane was
112 used for the determination of isoflavones. The conditions of HPLC were: RPC18 stainless steel chromatography
113 column; the mobile phase was 0.1% acetic acid and 0.1% acetic acid acetonitrile; the flow rate was 1.0 mL/min;
114 the column temperature was 40 °C and the wavelength was 260 nm.

115 **2.3. Statistical Analysis**

116 Excel 2016 and SPSS 25.0 were used for the analysis of statistical data. The frequency distribution
117 histograms were created by SPSS 25.0. The figures of the effects of different irradiation doses on the quality
118 traits were created with Graphpad Prism 8. The line was the average under the different irradiation dose.
119 Pearson method was used to analyse the correlation of different nutrition quality traits.

120 **3. Result**

121 **3.1. Effects of CIBR on soybean protein and oil concentration**

122 As a result of mutagenic treatments, a wide range of variability for protein concentration and oil concentration
123 was found in the M₂ and M₃ generation (**Fig 1**).

124 In the M₂ generation, the range of protein and oil concentration of all three CIBR doses was 38.5-42.9% and
125 18.8-23.8% respectively. The coefficient variation of protein ranged 2.0-2.2 % in M₂ generation and ranged 5.1-
126 6.9 % in M₃ generations. In the M₃ generation, seed protein generally increased compared with control. To oil,
127 compared with 100 Gy, higher irradiation doses (120 Gy and 140 Gy) exhibited positive effects on seed oil
128 concentration.

129 In the M₃ generation, 140 Gy treatment exhibited positive effects on both protein and oil concentration, and their
130 concentrations of most lines were higher than control in this group.

131 **3.2. Effects of CIBR on seed soluble sugar and sucrose concentration**

Compared to protein and oil concentration, the variation of the soluble sugar and sucrose concentration was more diverse. Besides the variations were relatively consistent in the M₂ and M₃ generations.

In terms of the concentration of soluble sugar, the range in the M₂ generation and M₃ generation was 78.0-123.4 mg/g and 48.1-98.8 mg/g respectively. Compared with the control treatment, mutagenic treatments generally increased the concentration of soluble sugar, especially at 140 Gy. The range of sucrose concentration was 39.6-74.7 mg/g and 22.7-71.8 mg/g in the M₂ generation and M₃ generation respectively. Overall, the distribution of sucrose was basically the same as soluble sugar, higher doses induced a more significant positive effect on sucrose concentration (**Fig. 2**).

3.3 Effects of CIBR on seed trace element concentrations

The effects of carbon ion beam irradiation on Fe, Mn, Zn and Cu concentrations are showed in **Fig 3**.

In the M₂ generation, Zn concentration ranged from 24.9-35.8 µg/g for all **CIBR** doses. The maximum concentration of Zn under **CIBR** treatment was **28.7 %** higher than control (27.8±0.53 µg/g). In the M₃ generation, the Zn concentration ranged from 31.5 to 42.6 µg/g, and **89.0 %** lines were concentrated at 36.0-42.0 µg/g. In the M₂ generation, irradiation treatments showed obvious positive effect on Zn concentration, especially at 100 Gy and 140 Gy doses with the highest coefficient variation of 7.25 % at 100 Gy. In M₃ generation however, higher irradiation dose (120 Gy and 140 Gy) increased Zn concentration.

Irradiation treatment generally decreased Fe concentration in the M₂ generation. The range of Fe concentration was 63.7-89.5 µg/g. Compared to control (80.66±1.35 µg/g), **only 12.1% lines showed higher Fe concentration**. In the group of 140 Gy, the Fe concentration of **55.6% lines** was higher than control. In the M₃ generation, the range of Fe was 31.9-69.8 µg/g, and over 70.3% lines were concentrated at 40.0-55.0 µg/g. On average across the different irradiation dose groups, the Fe concentration in over 50% lines was higher than control, and the concentration over 90% lines at 140 Gy was higher than control.

To Mn element, the concentration range in the M₂ and M₃ generations was 17.1 - 23.3 µg/g and 13.8-32.5 µg/g, respectively. No significant differences were found among different irradiation doses in the M₂ generation. However, in the M₃ generation, the 100 Gy and 120 Gy treatment generally increased Mn concentration. In the group of highest irradiation dose (140 Gy), Mn concentration in most lines was lower than control.

The ranges of Cu concentrations in M₂ and M₃ generations were 9.1-15.0 µg/g and 5.6-15.6 µg/g, respectively. In the M₂ generation, irradiation treatment generally increased Cu concentration, while the effects were different in the M₃ generation with most significant effect at 120 Gy.

3.4. Effects of CIBR on seed isoflavones concentration

The concentration of different isoflavones all showed diverse variation in the M₂ generation and M₃ generation (**Fig 4**).

To daidzin, in the M₂ generation, the range was 265-851 µg/g. Compared to control (421±16µg/g), irradiation treatment generally increased the daidzin concentration. Higher **CV** was found in 100 Gy and 120 Gy treatment than 140 Gy, while treatment of 140 Gy had a more significant positive effect on daidzin concentration, and

167 daidzin concentration in this group was higher than control. In the M₃ generation, the range was 215 -735 µg/g.
168 Since the range in control was 468±18 µg/g, the positive effect was decreased. In the group of 120 Gy, about
169 half lines showed higher daidzin concentrations, while the daidzin concentration of most lines in the group of
170 140 Gy was lower than control.

171 To glycitin, the range was similar in the two generations, which were 30.3-98.6 µg/g and 36.5-111.8 µg/g
172 respectively. The distribution was different though there were no significant differences among different
173 irradiation doses. The lines having glycitin concentration higher or lower than the average of control was all
174 about 50%.

175 Genistin had higher CV compared with the other two isoflavones, reaching 514-1821 µg/g in M₂ generation and
176 104-1147 µg/g in M₃ generation. The maximum genistin concentration was 169 % and 53 % higher than control
177 in M₂ and M₃ generation. The treatment of 100 Gy and 120 Gy consistently showed a discrete distribution in the
178 two generations. However, 140 Gy exhibited a positive effect on genistin concentration in the M₂ generation but
179 a negative effect in the M₃ generation.

180 **3.5. Correlation analysis of different nutritional quality indexes in M₂ and M₃ generations**

181 In order to compare the correlation among different nutritional quality indexes, the correlation of 11 quality
182 indexes was analyzed in this study, as showed in **Table 1** and **Table 2**.

183 In the M₂ generation, protein concentration was negatively correlated with oil and soluble sugar, but positively
184 correlated with Zn and Cu concentration. While oil concentration was negatively correlated with Zn and Cu
185 concentration. Sucrose concentration was positively correlated with Fe and Cu concentration. Zn concentration
186 was negatively correlated Fe and glycitin concentration, but positively correlated with Cu concentration.
187 Daidzein, glycitin and genistin concentrations were significantly correlated.

188 In the M₃ generation, protein concentration was positively correlated with Zn concentration, but significantly
189 negatively correlated with Fe concentration. Oil concentration was significantly positively correlated with the
190 concentrations of sucrose and soluble sugar, but negatively correlated with Cu concentration. Sucrose and
191 soluble sugar concentrations were positively correlated. Sucrose concentration was negatively correlated with
192 Mn concentration, while Fe and Mn concentrations were both positively correlated with that of daidzin and
193 genistin. Consistent with M₂ generation, daidzein, glycitin and genistin concentrations were significantly
194 correlated.

195 Classifying the lines in M₃ generation by clustering 11 quality indicators (**Fig 5**), four categories: A, B, C, and
196 D could be classified as shown in Table 3. Category A includes 16 lines with relatively higher concentration of
197 Zn, Cu, Fe and sucrose; Category B includes 21 lines with relatively higher oil, sucrose and soluble sugar
198 concentration; Category C includes 27 lines with relatively higher protein, Mn and isoflavone concentration;
199 Category D contains 27 lines with higher concentration of soluble sugar, Fe, Mn, Cu and isoflavones.

200 **4. Discussion**

Seed protein and oil concentration are important quality indicators of soybean. Studies have shown that mutation breeding is an effective way to break the significant negative correlation between protein and oil, which can create germplasm resources with high protein and oil concentration (Chaudhary et al., 2015). The change range of the protein and oil concentration is about 5%, similar results in fast neutron irradiation methods are also revealed (Bolon et al., 2011). Though the negative correlation between protein and oil concentration were still found from the M_2 generation to the M_3 generation, higher irradiation doses induced the high-oil and high-protein mutants with the possibility of stable inheritance, thus the irradiation treatment may break the negative correlation between protein and oil. This was consistent with the results of Patil et al. (2007). Therefore, if the screening population is large enough, higher irradiation dose has the potential in creating lines with both high-oil and high-protein mutants in breeding program.

Hayashi and Aoki (1985) demonstrated that the irradiation treatment promoted the accumulation of sucrose in potatoes. EMS treatment can increase seed soluble sugar concentration up to two times compared to control in soybean (Espina et al., 2018). Consistent with these results, in present study, a wider range changes in sucrose and soluble sugar concentrations from the offspring population by carbon ion beam irradiation were also found. The distribution and variation were similar in the M_2 and M_3 generation, and irradiation treatments generally increased the concentration of soluble sugar and sucrose. Furthermore, the higher irradiation dose had more positive effect on their concentrations. In this regard, the trait of soluble sugar and sucrose may be more easily inherited stably in the early generations.

The concentration of trace elements is closely related to the varieties, planting conditions, soil and other environmental factors (Wang et al., 2000). Correlation analysis in present study showed that trace elements were closely associated with protein, oil, carbohydrate and isoflavone ingredients. Therefore, the position of trace elements in soybean seed nutrition quality traits is irreplaceable. The different distribution of trace elements in the M_2 generation and M_3 generation demonstrated that these traits were not stabilized in early generations. Although irradiation treatment had positive effect on the concentration of trace elements in this study, the responses of Fe, Mn, Zn, Cu to irradiation dose were different. For example, lower irradiation dose exhibited more significant positive effect on Mn than other trace elements, while 120 Gy was a better choice to increase the concentration of Cu element.

Much progress has been achieved in breeding high-isoflavone germplasm resources of soybean through the selection of existing resources and crossing combinations in recent years (Wu et al., 2020). The present study showed that higher radiation doses of carbon ion beam reduced the isoflavone concentration in the M_3 generation, which was different from other nutrition quality indicators. Therefore, in order to get more high-isoflavone mutants, the radiation dose should be appropriately reduced. There was evidence that isoflavone concentration is negatively correlated with protein concentration, but positively correlated with 100-seed weight, and small seed and brown coat seed showed higher isoflavone concentration (Seguin et al., 2004; Sakai et al., 2005; Lee et al., 2008). In this study, isoflavone concentration was closely related with trace element concentration, but the correlations were not consistent in the M_2 and M_3 generations, which indicates that the traits have not been stably inherited in these generations. Moreover, many studies demonstrated that isoflavone concentration is greatly affected by environmental factors (Sivesind et al., 2005). Therefore, the application of radiation mutation breeding to screen specific high-isoflavone germplasm still requires more systematic investigation.

It is difficult to screen mutants with all excellent nutrition quality indicators in breeding program, nevertheless it is feasible to focus some indicators. Our cluster analysis of the M₃ generation lines provides the reference for the screening of mutants in breeding. As the key generations of mutation breeding, the M₂ and M₃ generations can reflect the mutagenic effects of some traits, however, many nutrition quality traits are still separated in these generations. Therefore, further research is needed to determine in which generation these traits can stably be inherited.

Acknowledgments

This work was partially funded by the Major Program of National Science and Technology of China, grant number **2021YFD1201103-03** and the Strategic Priority Research Program of the Chinese Academy of Sciences, grant number **XDA24030403-3**.

References

- Alikamanoglu S, Yaycili O, Sen A. 2011. Effect of gamma radiation on growth factors, biochemical parameters, and accumulation of trace elements in soybean plants (*Glycine max* L. Merrill). *Biol Trace Elem Res* 141: 283-293.
- Arase S, Yoshihiro H, Jun A, Megumi K, Tetsuya Y, Keisuke K, Narumi I, Tanaka A, Kanazawa A. 2011. Optimization of ion-beam irradiation for mutagenesis in soybean: Effects on plant growth and production of visibly altered mutants. *Plant Biotechnol-Nar* 28(3): 323-329.
- Bolon YT, Stec AO, Michno JM, Roessler J, Bhaskar PB, Ries L, Dobbels AA, Campbell BW, Young NP, Anderson JE. 2014. Genome resilience and prevalence of segmental duplications following fast neutron irradiation of soybean. *Genetics* 198(3): 967-81.
- Chaudhary J, Patil GB, Sonah H, Deshmukh RK, Vuong TD, Valliyodan B, Nguyen HT. 2015. Expanding omics resources for improvement of soybean seed composition traits. *Front Plant Sci* 6(31): 1021.
- Cober ER, Voldeng HD. 2000. Developing high-protein, high-yield soybean populations and lines. *Crop Sci* 40(1): 39-42.
- Espina MJ, Sabbir A, Angelina B, Ekundayo A, Zeinab Y, Prakash A, Vince P, Ali T. 2018. Development and phenotypic screening of an Ethyl Methane Sulfonate mutant population in soybean. *Front Plant Sci* 9: 394.
- Hajduch M, Debre F, Bohmova B, Pretova A. 2000. Effect of sodium azide and gamma-irradiation on the seed protein composition of soybean. *Biologia* 55(1): 115-120.
- Hayashi T, Aoki S 1985) Effect of irradiation on the carbohydrate metabolism responsible for sucrose accumulation in potatoes. *J Agr Food Chem* 33(1): 14-17.
- Hoeck JA, Fehr WR, Murphy PA, Welke GA. 2000. Influence of genotype and environment on isoflavone

- 271 contents of soybean. Crop Sci 40(1): 48-51.
- 272 Jan S, Parween T, Siddiqi TO, Mahmooduzzafar. 2012. Effect of gamma radiation on morphological,
273 biochemical, and physiological aspects of plants and plant products. Environ Rev 20: 17-39.
- 274 Jin L, Shen B, Yao B, Shi J, Ju D, Tian J. 2011. Effect of he-ne laser illumination exposition on content of soy
275 isoflavone in its seedling. Journal of Agriculture University 33(3): 264-268. (in Chinese with English
276 abstract)
- 277 Karr-Lilienthal LK, Kadzere CT, Grieshop CM, Fahey Jr GC. 2005. Chemical and nutritional properties of
278 soybean carbohydrates as related to nonruminants: a review. Livest Prod Sci 97(1): 1-12.
- 279 Lee SJ, Kim JJ, Moon HI, Ahn JK, Chun SC, Jung WS, Lee OK, Chung IM. 2008. Analysis of isoflavones and
280 phenolic compounds in korean soybean (*Glycine max* (L.) Merrill) seeds of different seed weights. J Agr
281 Food Chem 56(8): 2751-2758.
- 282 Li YS, Du M, Zhang QY, Wang GH, Jin J, Herbert S, Liu XB. 2014. Planting date influences fresh pod yield
283 and seed chemical compositions of vegetable soybean. Hortscience 49: 1376-1380.
- 284 Li YS, Du M, Zhang QY, Wang GH, Liu XB. 2012. Greater differences exist in seed protein, oil, total soluble
285 sugar and sucrose content of vegetable soybean genotypes [*Glycine max* (L.) Merrill] in Northeast China.
286 Aust J Crop Sci 6: 1681-1686.
- 287 Liu C, Li Y, Tu B, Wang X, Tian B, Zhang Q, Liu XB. 2019. Seed nutritional quality comparison of vegetable
288 soybean genotypes at fresh pod and mature stage. Emir J Food Agr 31(6): 405-414.
- 289 Mikuriya S, Kasai M, Nakashima K, Natasia Hase Y, Yamada T, Abe J, Kanazawa A. 2017. Frequent generation
290 of mutants with coincidental changes in multiple traits via ion-beam irradiation in soybean. Genes Genet
291 Syst 92(3): 16.
- 292 Patil A, Taware SP, MD Oak, Tamhankar SA, Rao VS. 2007. Improvement of oil quality in soybean [*Glycine*
293 *max* (L.) merrill] by mutation breeding. J Am Oil Chem Soc 84: 1117-1124.
- 294 Patil G, Mian R, Vuong T, Pantalone V, Nguyen H. 2017. Molecular mapping and genomics of soybean seed
295 protein: a review and perspective for the future. Theor Appl Genet 130(2): 1975-1991.
- 296 Sakai T, Kikuchi A, Shimada H, Takada Y, Kono Y, Shimada S. 2005. Evaluation of isoflavone contents and
297 compositions of soybean seed and its relation with seeding time. Jpn J Crop Sci 74(2): 156-164.
- 298 Saldivar X, Wang YJ, Chen P, Hou A. 2011. Changes in chemical composition during soybean seed
299 development. Food Chem 124(4): 1369-1375.
- 300 Schmutz J., Cannon S, Schlueter J, Ma J, Mitros T, Nelson W, Hyten DL, Song Q, Thelen JJ, Cheng JJ. 2010.
301 Genome sequence of the palaeopolyploid soybean. Nature 463(7294): 178-83.
- 302 Seguin P, Zheng W, Smith DL, Deng W. 2004. Isoflavone content of soybean cultivars grown in eastern Canada.

303 J Sci Food Agr 84 (11), 1327-1332.

304 Sivesind E, Seguin P. 2005. Effects of the environment, cultivar, maturity, and preservation method on red clover
305 isoflavone concentration. J Agr Food Chem 53(16): 6397-6402.

306 Tu B, Liu C, Tian B, Zhang Q, Liu X, Herbert SJ. 2017. Reduced abscisic acid content is responsible for
307 enhanced sucrose accumulation by potassium nutrition in vegetable soybean seeds. J Plant Res 130: 551-
308 558.

309 Wang HF, Takematsu N, Ambe S. 2000. Effects of soil acidity on the uptake of trace elements in soybean and
310 tomato plants. Appl Radiat Isotopes 52(4): 803-811.

311 Wang X, Liu CK, Tu BJ, Li YS, Zhang QY, Liu XB. 2021. Effects of carbon ion beam irradiation on phenotypic
312 variations and biochemical parameters in early generations of soybean plants. Agriculture-Basel 11(2): 98.

313 Wu D, Li D, Zhao X, Zhan Y, Teng W, Qiu L, Han Y. 2020. Identification of a candidate gene associated with
314 isoflavone content in soybean seeds using genome-wide association and linkage mapping. Plant J 104(4):
315 950-963.

316 Xue GQ, Liu, Q, Han YQ, Wei HG, Dong T. 2006. Determination of thirteen metal elements in the plant
317 foeniculum vulgare Mill. by flame atomic absorption spectrophotometry. Spectrosc Spect Anal 26: 1935-
318 1938. (in Chinese with English abstract)

319

Legend of figures

Figure 1. The distribution of seed protein and oil concentration in M_2 and M_3 generations.

(**A** and **B**. Frequency distribution of protein concentration; **C** and **D**. Frequency distribution of oil concentration; **E** and **F**. Effect of different irradiation doses on protein concentration; **G** and **H**. Effect of different irradiation doses on oil concentration; M_2 generation: **A**, **C**, **E** and **G**; M_3 generation: **B**, **D**, **F** and **H**)

Figure 2. The distribution of seed soluble sugar and sucrose concentration in M_2 and M_3 generations.

(**A** and **B**. Frequency distribution of soluble sugar concentration; **C** and **D**. Frequency distribution of sucrose concentration; **E** and **F**. Effect of different irradiation doses on soluble sugar concentration; **G** and **H**. Effect of different irradiation doses on sucrose concentration; M_2 generation: **A**, **C**, **E** and **G**; M_3 generation: **B**, **D**, **F** and **H**)

Figure 3. The distribution of seed trace elements concentration in M_2 and M_3 generations

(**A** and **E**. Frequency distribution of Zn concentration; **B** and **F**. Frequency distribution of Fe concentration; **C** and **G**. Frequency distribution of Mn concentration; **D** and **H**. Frequency distribution of Cu concentration; **I** and **M**. Effect of different irradiation doses on Zn concentration; **J** and **N**. Effect of different irradiation doses on Fe concentration; **K** and **O**. Effect of different irradiation doses on Mn concentration; **L** and **P**. Effect of different irradiation doses on Cu concentration; M_2 generation: **A-D**, **I-L**; M_3 generation: **E-H**, **M-P**)

Figure 4. The distribution of seed isoflavones concentration in M_2 and M_3 generations

(**A** and **D**. Frequency distribution of daidzin concentration; **B** and **E**. Frequency distribution of Glycitin concentration; **C** and **F**. Frequency distribution of Genistin concentration; **G** and **J**. Effect of different irradiation doses on daidzin concentration; **J** and **N**. Effect of different irradiation doses on Glycitin concentration; **K** and **O**. Effect of different irradiation doses on Genistin concentration; M_2 generation: **A-C**, **G-H**; M_3 generation: **D-F**, **J-L**)

Figure 5. Cluster analysis of seed quality traits in M_3 generation lines

(The number in the figure is the line number, 1-47 is the treatment of 100 Gy, 48-73 is the treatment of 120 Gy and 74-91 is the treatment of 140 Gy)

Legend of tables

Table1. Spearman correlation analyses of seed quality traits in the soybean M₂ population

	Protein	Oil	SUG	Sucrose	Zn	Fe	Mn	Cu	Daidzin	Glycitin	Genistin
Protein	1.000										
Oil	-0.260*	1.000									
SUG	-0.208*	0.266*	1.000								
Sucrose	0.127	0.030	-0.144	1.000							
Zn	0.302**	-0.319**	-0.120	0.019	1.000						
Fe	-0.087	0.144	0.153	0.277**	-0.248*	1.000					
Mn	-0.030	0.017	-0.132	-0.076	-0.004	0.075	1.000				
Cu	0.230*	-0.398**	-0.188	0.243*	0.397**	-0.046	-0.201	1.000			
Daidzin	-0.026	0.090	-0.057	0.020	-0.133	-0.061	0.043	0.075	1.000		
Glycitin	-0.114	0.145	-0.061	0.036	-0.241*	-0.080	-0.048	0.057	0.881**	1.000	
Genistin	-0.046	0.181	0.072	0.137	-0.048	0.071	0.120	-0.050	0.715**	0.649**	1.000

Notes: ** and *represent significant difference at $p = 0.01$ and 0.05 level

Table2. Spearman correlation analyses of seed quality traits in the soybean M₃ population

	Protein	Oil	SUG	Sucrose	Zn	Fe	Mn	Cu	Daidzin	Glycitin	Genistin
Protein	1.000										
Oil	0.110	1.000									
SUG	0.172	0.357**	1.000								
Sucrose	-0.097	0.381**	0.572**	1.000							
Zn	0.212*	-0.020	-0.176	0.131	1.000						
Fe	-0.271**	-0.163	-0.074	0.066	0.113	1.000					
Mn	-0.082	-0.061	-0.019	-0.407**	-0.130	0.413**	1.000				
Cu	-0.060	-0.364**	-0.107	-0.071	0.131	-0.058	-0.204	1.000			
Daidzin	-0.101	0.002	0.135	-0.105	-0.281**	0.216*	0.383**	-0.132	1.000		
Glycitin	-0.153	0.035	0.088	-0.062	-0.200	0.117	0.268*	-0.164	0.756**	1.000	
Genistin	-0.084	0.007	0.107	-0.137	-0.339**	0.232*	0.448**	-0.119	0.961**	0.655**	1.000

Notes: ** and *represent significant difference at $p = 0.01$ and 0.05 level

360

361

362

Table 3. The average of quality traits based on the cluster analysis in M₃ generation

Category	Protein	Oil	SUG	Sucrose	Zn	Fe	Mn	Cu	Daidzin	Glycitin	Genistin
A	38.14	23.02	72.93	48.74	40.04	48.42	22.92	11.73	264.80	51.18	402.58
B	38.29	24.06	77.85	49.33	38.52	44.18	23.48	9.55	368.41	65.31	584.89
C	38.75	23.35	73.91	42.29	38.28	44.90	25.72	9.51	480.40	74.57	803.09
D	37.84	23.46	78.15	46.88	37.93	52.26	27.37	10.38	575.16	78.45	1003.17

363

Figure 1

Figure 1. The distribution of seed protein and oil concentration in M_2 and M_3 generations.

(A and B. Frequency distribution of protein concentration; C and D. Frequency distribution of oil concentration; E and F. Effect of different irradiation doses on protein concentration; G and H. Effect of different irradiation doses on oil concentration; M_2 generation: A, C, E and G; M_3 generation: B, D, F and H)

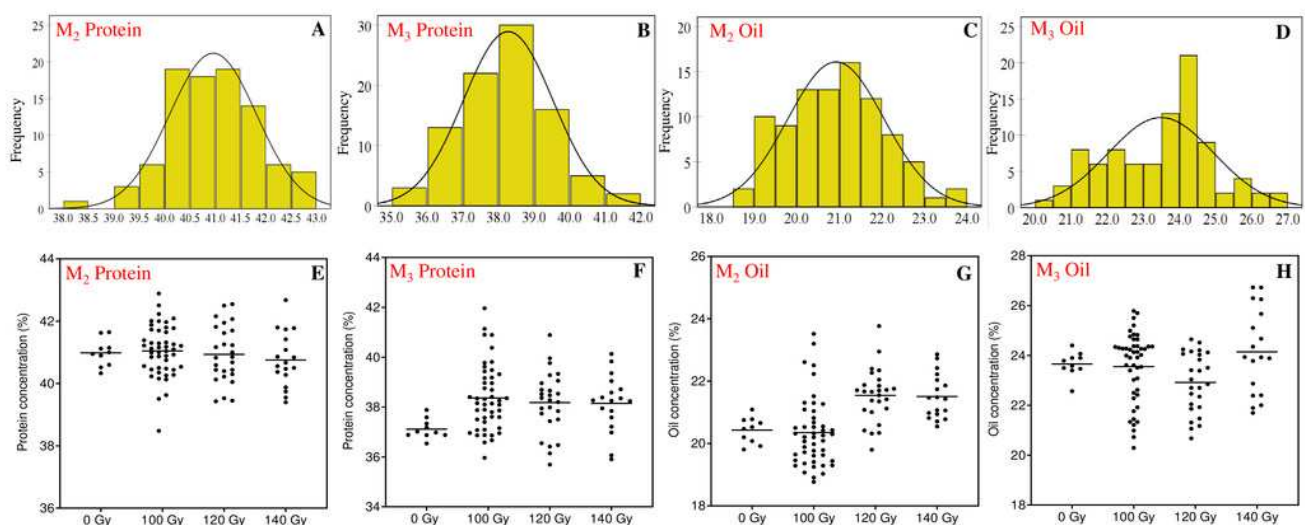


Figure 2

Figure 2. The distribution of seed soluble sugar and sucrose concentration in M_2 and M_3 generations.

(**A** and **B**. Frequency distribution of soluble sugar concentration; **C** and **D**. Frequency distribution of sucrose concentration; **E** and **F**. Effect of different irradiation doses on soluble sugar concentration; **G** and **H**. Effect of different irradiation doses on sucrose concentration; M_2 generation: **A**, **C**, **E** and **G**; M_3 generation: **B**, **D**, **F** and **H**)

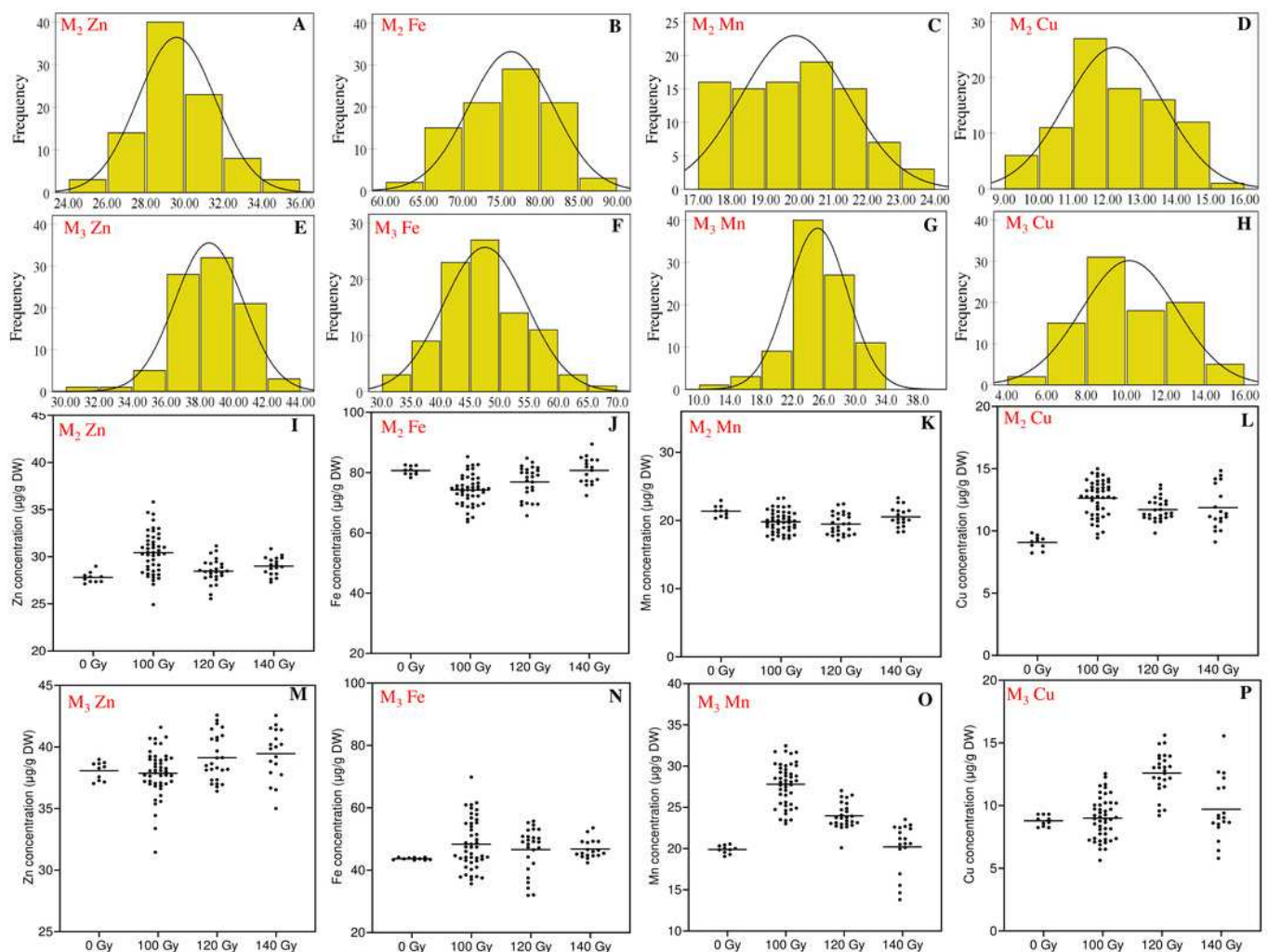


Figure 3

Figure 3. The distribution of seed trace elements concentration in M_2 and M_3 generations

(**A** and **E**. Frequency distribution of Zn concentration; **B** and **F**. Frequency distribution of Fe concentration; **C** and **G**. Frequency distribution of Mn concentration; **D** and **H**. Frequency distribution of Cu concentration; **I** and **M**. Effect of different irradiation doses on Zn concentration; **J** and **N**. Effect of different irradiation doses on Fe concentration; **K** and **O**. Effect of different irradiation doses on Mn concentration; **L** and **P**. Effect of different irradiation doses on Cu concentration; M_2 generation: **A-D, I-L**; M_3 generation: **E-H, M-P**)

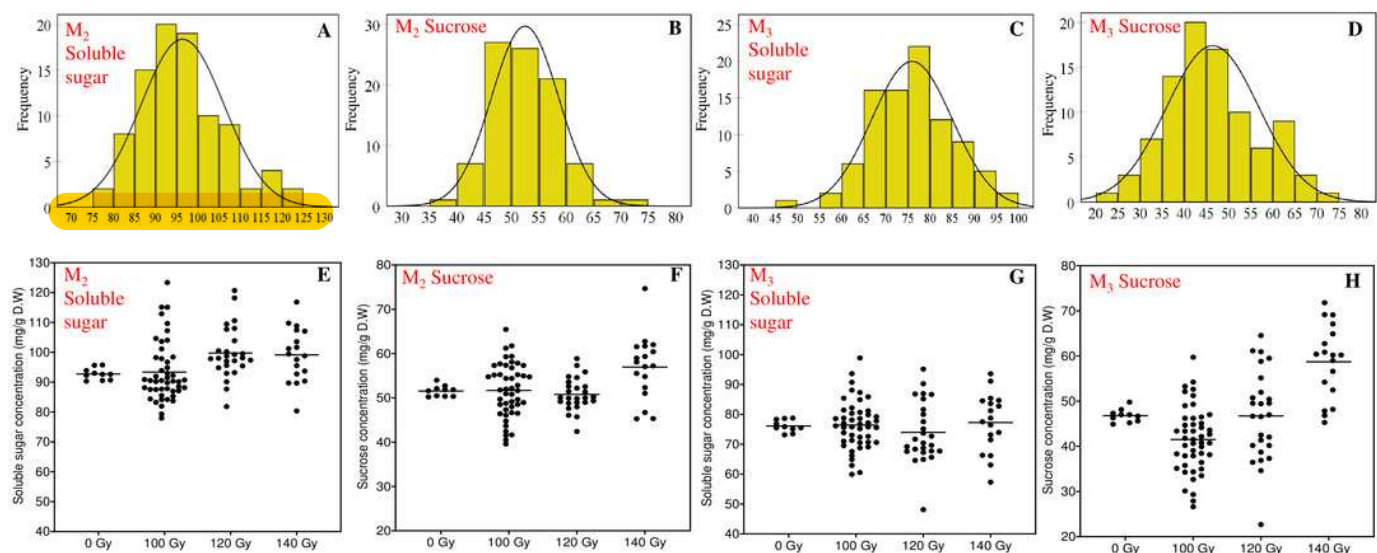


Figure 4

Figure 4. The distribution of seed isoflavones concentration in M_2 and M_3 generations

(**A** and **D**. Frequency distribution of daidzin concentration; **B** and **E**. Frequency distribution of Glycitin concentration; **C** and **F**. Frequency distribution of Genistin concentration; **G** and **J**. Effect of different irradiation doses on daidzin concentration; **J** and **N**. Effect of different irradiation doses on Glycitin concentration; **K** and **O**. Effect of different irradiation doses on Genistin concentration; M_2 generation: **A-C, G-H**; M_3 generation: **D-F, J-L**)

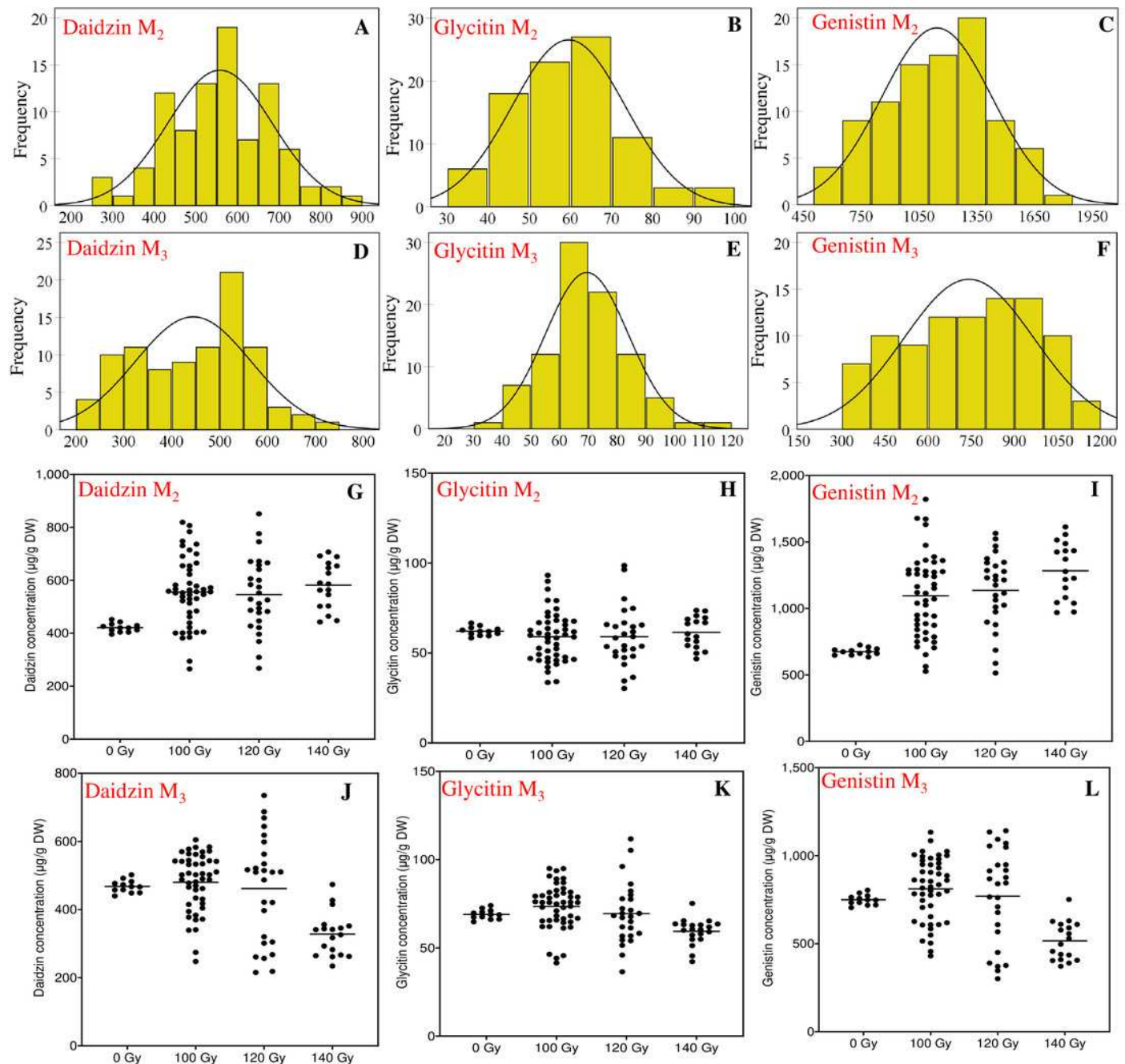


Figure 5

Figure 5. Cluster analysis of seed quality traits in M_3 generation lines

(The number in the figure is the line number, 1-47 is the treatment of 100 Gy, 48-73 is the treatment of 120 Gy and 74-91 is the treatment of 140 Gy)

