



# Effects of melatonin on growth and antioxidant capacity of naked oat (*Avena nuda* L) seedlings under lead stress

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## ABSTRACT

Melatonin (MT) plays an important role in plant response to abiotic stress. In recent years, lead (Pb) pollution has seriously affected the living environment of plants. In this study, we applied two different concentrations of MT to naked oat seedlings under Pb stress to explore the effect of MT on naked oat seedlings under Pb pollution. The results showed that Pb stress seriously inhibited the growth and development of naked oat seedlings, which was alleviated by MT. MT could increase the soluble protein content and decrease the proline content of naked oat seedlings to maintain the osmotic balance of naked oat seedlings. The application of MT could accelerate the removal of reactive oxygen species (ROS) and improve the activities of superoxide dismutase (SOD), peroxidase (POD) and catalase (CAT), so as to maintain the redox balance in naked oat seedlings. Exogenous melatonin could significantly increase the chlorophyll content of naked oat seedlings under Pb treatment, so as to improve the photosynthesis efficiency of naked oat seedlings. MT could also remarkably up regulate the expression of the genes of *LOX*, *POX* and *Asmap1*, and affect the expression of transcription factors *NAC* and *WRKY1*. It might regulate the expression of downstream genes through MAPKs pathways and TFs to improve the Pb tolerance of naked oat seedlings. These results proved that MT could significantly promote the growth and development of naked oats seedlings under Pb stress, which is expected to be applied in agricultural production practice.

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## INTRODUCTION

After the third Industrial Revolution, science and technology made great progress; however, environmental problems also followed. The increase of motor vehicle exhaust emissions, the abuse of sewage irrigation, pesticides and chemical fertilizers, as well as the rapid development of modern mining industry have seriously polluted the soil, water and atmosphere (*Pellinen, Cherkashina & Gustaytis, 2021; Zhao et al., 2015*). Heavy metal contamination is potentially toxic to humans and animals, among which the heavy metal pollution problems caused by heavy metals such as lead (Pb) and mercury or their compounds, rank first for the biological threat (*Rahman & Singh, 2019*). Metal Pb is

distributed in the atmosphere, water and soil, which leads to the excessive production and accumulation of reactive oxygen species (ROS) in plants through plant growth. Excessive ROS not only destroy plant proteins, carbohydrates, lipids and DNA, but also cause plant oxidative stress (Gill & Tuteja, 2010), leading to intracellular dynamic imbalance, as well as affecting and harming plant growth (Pérez-Chaca et al., 2014). At present, most plants suffer from serious Pb pollution (Pace et al., 2020). Pb could significantly reduce the leaf area and root length of *Zygophyllum fabago* (López-Orenes et al., 2018). Moreover, studies revealed that Pb has negative effects on the biochemical and physiological processes that regulate grain quality, and that high Pb concentrations can destroy several key metabolic processes in plants, such as organelle integrity, membrane stability, mineral metabolism, oxygen release and enzyme activities (Aslam et al., 2020). How to improve the yield and quality of crops under Pb stress and ensure food security has always been one of the popular topics in research.

Since the discovery of melatonin (MT) in plants in 1995 (Hattori et al., 1995), its function in plants has been revealed rapidly. More and more studies have found that MT plays an active role in plant stress resistance (Sun et al., 2021). MT can improve plant growth and development under drought, low temperature and salt stress conditions (Li et al., 2019; Wang, Zhang & Ding, 2020; Xia et al., 2020; Zhou et al., 2020). Under heavy metal stress, MT, as an effective antioxidant (Reiter et al., 2016), can reduce the phytotoxicity of nickel by improving the efficiency of photosynthesis, secondary metabolism and the tolerance of oxidative stress of tomato seedlings (Jahan et al., 2020). MT can also reduce the toxicity of copper by improving copper fixation and scavenging ROS in cucumber (Cao et al., 2019). MT can respond to Pb stress through reducing the absorption of Pb by *Carthamus tinctorius* and regulating the antioxidant system of *Carthamus tinctorius* (Namdjoyan et al., 2020). It can reduce cadmium accumulation and reconstruct microRNA-mediated redox homeostasis to enhance plant tolerance to cadmium stress as well (Lewitt & Hulting, 2017). In addition, MT shows more other positive activities in plants, such as regulating seed germination, root growth and development, leaf senescence, circadian rhythm, postharvest fruit ripening, and plant response to adverse environmental conditions (Sun et al., 2021). It has increasingly become one of the star hormones in plant stress resistance regulation.

Naked oats are plants of the family *Gramineae* and oats, with the scientific name “naked grain type oats” or “naked oats”. Naked oats are native to China and recognized as one of the food crops with the highest nutritional value in the world (Zhao et al., 2020), whose protein content is the highest among cereal crops (Zhang, 2006). As a high-cold, drought-tolerant and high-quality food crop, it has lower requirements for topography (Khan et al., 2019). However, it is more vulnerable to soil pollution. It has been found that when naked oat seeds were in the heavy metal environment, the plant height, root length, and chlorophyll content were reduced (Wang, 2014). Like other crops, the growth and development of naked oats is hindered by heavy metals in the soil. But at present, the research on the metal toxicity to naked oats and the response mechanism of plants to alleviate the toxicity is still relatively insufficient. There are no data on metal toxicity mitigation and protection depending on MT in the growth of naked oats. In this paper, we have more specifically elucidated the damage caused by Pb to naked oat seedlings. For

the first time, we applied different concentrations of MT and investigated the effect of different concentrations of exogenous melatonin on Pb-stressed naked oat seedlings, and demonstrated that MT can promote the growth of naked oat seedlings under Pb stress. This study shows that melatonin can be used in agricultural production.

## MATERIALS & METHODS

### Plant materials and seed germination conditions

The naked oat variety “Jinyan 2” (*Avena nuda* L.) was provided by the Shaanxi Key Laboratory of Biotechnology and stored in a refrigerator at 4 °C. Healthy and full seeds were selected, disinfected for 30s with 75% alcohol, rinsed with distilled water three times, disinfected with 0.1% mercury solution for 7 min, finally rinsed with distilled water six times, moved into a water culture box containing Hoagland nutrient solution (pH  $6.5 \pm 0.1$ ). Hoagland nutrient solution was replaced every three days, and the water culture box was placed in the culture room at 25 °C with 75% relative humidity, and a 14L/10D photoperiod.

### Experimental group design

When the naked oats seedlings grew to the 7th day, the control group was continued to be irrigated with Hoagland nutrient solution, and the treatment group was irrigated with different concentrations of Pb solution (0, 25, 50, 75, 100 mg L<sup>-1</sup>). According to the statistical observation, the Pb treatment condition was set as 75 mg L<sup>-1</sup> in this work. With reference to the previous experiment, the concentration of MT was selected as 50 and 100 μM (*Gao et al., 2018*). The experiment was divided into six treatment groups: control, Pb, MT<sub>50</sub>, MT<sub>100</sub>, MT<sub>50</sub>+Pb and MT<sub>100</sub>+Pb. Naked oat seedlings were cultured for 7 days and then treated with 0 and 75 mg L<sup>-1</sup> Pb(NO<sub>3</sub>)<sub>2</sub>, respectively, the hydroponic solution was renewed every three days. The seedlings grown in Pb-free nutrient solution was used as the control. MT<sub>50</sub> and MT<sub>100</sub> were treated with MT solution of 50 and 100 μM respectively on the basis of the control group. MT<sub>50</sub> +Pb and MT<sub>100</sub>+Pb were root-irrigated with MT solution of 50 and 100 μM on the basis of the Pb treatment group. All six treatment groups were sampled seven days after the start of stress. three groups of parallel repeats were set up for sampling, testing and measurement, respectively.

### Calculation of plant height, fresh weight and dry weight

After seven days of treatment in different treatment groups, naked oat seedlings were randomly selected from each hydroponic culture box, and the plant height, plant fresh weight and plant dry weight were measured. The plant height is the linear distance from the base of the radicle to the top of the leaf. After the seedlings of naked oats were taken out, they were simply washed with tap water, then rinsed with distilled water three times, gently dried with paper towels, and the fresh plants were quickly weighed with electronic scales. The seedlings of naked oats were dried at 105 °C for 30 min, then dried at 80 °C for 24 h, and the dry weight was measured by electronic balance.

### Determination of chlorophyll content and lead content

The leaf samples frozen in liquid nitrogen from  $-80\text{ }^{\circ}\text{C}$  (fresh weight 0.2 g) were fully ground in liquid nitrogen with 95% anhydrous ethanol, then the volume was fixed to 20 mL,  $4\text{ }^{\circ}\text{C}$  and placed in the dark for 24 h. The values of OD<sub>649</sub> and OD<sub>665</sub> of supernatants were determined after samples were centrifuged for 10 min. The formula for calculating chlorophyll is as follows:  $C = C_a + C_b$ , in which  $C_a = 13.95A_{665} - 6.88A_{649} - 7.32A_{665}$ ,  $C_b$  is  $24.96A_{649} - 7.32A_{665}$ . Chlorophyll content = chlorophyll concentration  $\times$  extraction liquid volume  $\times$  dilution multiple / sample fresh weight (Porra, Thompson & Kriedemann, 1989). The content of Pb in leaves of naked oat seedlings was determined by graphite furnace atomic absorption spectrophotometer (model Z2700), following the method of Shi, Li & Pan (2009). The sample was dried in an air-forced oven at  $60\text{ }^{\circ}\text{C}$  for 48 h. Dry plant material was ground in a stainless-steel blender to pass through a 0.4-mm sieve. Leaf samples were subjected to mixed acid digestion ( $\text{HNO}_3\text{-HClO}_4\text{-HF}$ ). A certified reference material of sediment IRMM-804 with  $(0.42 \pm 0.07)\text{ mg kg}^{-1}$  Pb was purchased from the National Centre for Certificate Reference Materials, China, and was used with all patch of digestions. Dried plant samples were digested in a 4:1 (V V<sup>-1</sup>) mixture of  $\text{HNO}_3\text{-HClO}_4$ . Ten milliliter of mixed acids was added to 0.75 g of plant sample in a high-walled beaker and allowed to stand for 12 h at  $25\text{ }^{\circ}\text{C}$ . The samples were then heated in a sand bath at  $170\text{ }^{\circ}\text{C}$  until clear. After cooling, the solution was diluted to 25 mL with deionized water. The Pb content of the digested solutions was determined by graphite furnace AAS. A reagent blank was incorporated within each batch of analytical samples.

### Determination of antioxidant enzyme activity, soluble protein and proline content

The fresh weight of the leaf sample is 0.2 g. PBS buffer (pH7.4) was added to the sample, which was ground to homogenate in an ice bath, and then centrifuged at  $8,000 \times g$  for 10 min to collect the supernatant. The supernatant was used for the determination of superoxide dismutase (SOD; EC1.15.1.1), peroxidase (POD; EC1.11.1.7) and catalase (CAT; EC1.11.1.6). The activities of SOD, POD and CAT were recorded at 550 nm, 420 nm and 405 nm, respectively, according to Elavarthi & Martin's method (2010).

The content of soluble proteins was detected by kits (Nanjing Institute of Bioengineering, Nanjing, China). 0.1 g of fresh leaves was homogenized with 900  $\mu\text{L}$  buffer and the homogenate was centrifuged for 10 min ( $3000\text{ g}$ ); 50  $\mu\text{L}$  supernatant was added to three mL Coomassie brilliant blue solution. The mixture was incubated at  $25\text{ }^{\circ}\text{C}$  for 30 min and the absorbance of soluble protein content was recorded at 595 nm.

Proline content in seedlings of naked oat was measured according to the ninhydrin method described by Bates, Waldren & Teare (1973). The leaf sample was 0.2 g. Proline was extracted with 3% sulphosalicylic acid and then filtered. A portion of the filtrate was supplemented by the addition of one mL of ninhydrin and glacial acetic acid reagent. The reaction mixture was boiled ( $95\text{ }^{\circ}\text{C}$ ) for 1 h. Then the mixture was placed on ice to stop the reaction, the absorbance of the sample was measured at 520 nm.

### Determination of malondialdehyde and ROS accumulation

Malondialdehyde (MDA) was measured according to the procedure described by [Tan et al. \(2019\)](#). The leaf sample (0.5 g fresh weight) was ground with five mL of 5% trichloroacetic acid and centrifuged for 10 min (8000 g, 4 °C). one mL of supernatant was mixed with one mL of 0.67% thiobarbituric acid (TBA). The mixture was then boiled in water for 30 min, the absorbance at 532 nm and 600 nm was measured respectively to calculate MDA content.

To measure H<sub>2</sub>O<sub>2</sub>, the methods described by [Nawaz et al. \(2018\)](#) was followed. Fresh leaves (100 mg) were ground in a mortar with 900 µL buffer, H<sub>2</sub>O<sub>2</sub> content was recorded at a wavelength of 405 nm. The kit method (Nanjing Jiancheng Institute of Biological Engineering, Nanjing, China) was applied to detect O<sub>2</sub><sup>•-</sup>, Fresh leaves (100 mg) were centrifuged at 4000 g and 25 °C for 10 min. 50 µL of the supernatant was mixed with the four mL reagent solution. The mixture was bathed at 37 °C for 40 min and then two mL reagent solution was added. O<sub>2</sub><sup>•-</sup> content was recorded at a wavelength of 550 nm.

According to the method of DAB staining ([Orozco-Cardenas & Ryan, 1999](#)), fresh leaves were soaked in DAB solution (1 mg mL<sup>-1</sup>), the pH was adjusted to 3.8, vacuum was permeated for 30 min, shaker dark treatment was placed for 8 h, washed twice with 95% anhydrous ethanol, then was boiled in a water bath (20 min), and stored in 50% glycerin after cooling, stored at 4 °C. At least three leaves were selected for each treatment, and photographed with a stereomicroscope.

### Extraction of RNA and analysis of related gene expression

Sequence search and primer design were carried out according to previous work ([Gao et al., 2018](#)). The list of primers is shown in [Table S1](#). The total RNA of naked oats was extracted by TRIzol method. First-strand cDNA was synthesized using the PrimeScript<sup>TM</sup> RT reagent kit with the gDNA Eraser (Takara, Shiga, Japan). qRT-PCR was performed on a Bio-Rad CFX96 Real-Time PCR System (Bio-Rad, Hercules, CA, USA) using FastStart Essential DNA Green Master (Tiangen, Beijing, China). The procedure is as follows: 95 °C for 10 min, 1 cycle, 95 °C for 10 s, 60 °C for 30 s, 40 cycles. Finally, a melting curve was drawn to reconfirm the specificity of the primers by heating the product from 60 °C to 95 °C. The internal reference gene was Actin (KP257585.1) ([Gao et al., 2018](#); [Genty, Briantais & Baker, 1989](#)). Three independent biological replications were performed for each experiment. The relative gene expression levels were calculated according to the 2<sup>-ΔΔCt</sup> method and presented as fold changes.

### Data analysis

All the measurements reported in this study are the means of three replicates. Vertical bars represent ± S.D. SPSS26.0 software was used for data statistics univariate analysis and minimum significant difference (LSD) test to compare the average values. We use prism 8.00 to organize pictures.

## RESULTS

### MT improves the growth of naked oat seedlings under lead stress

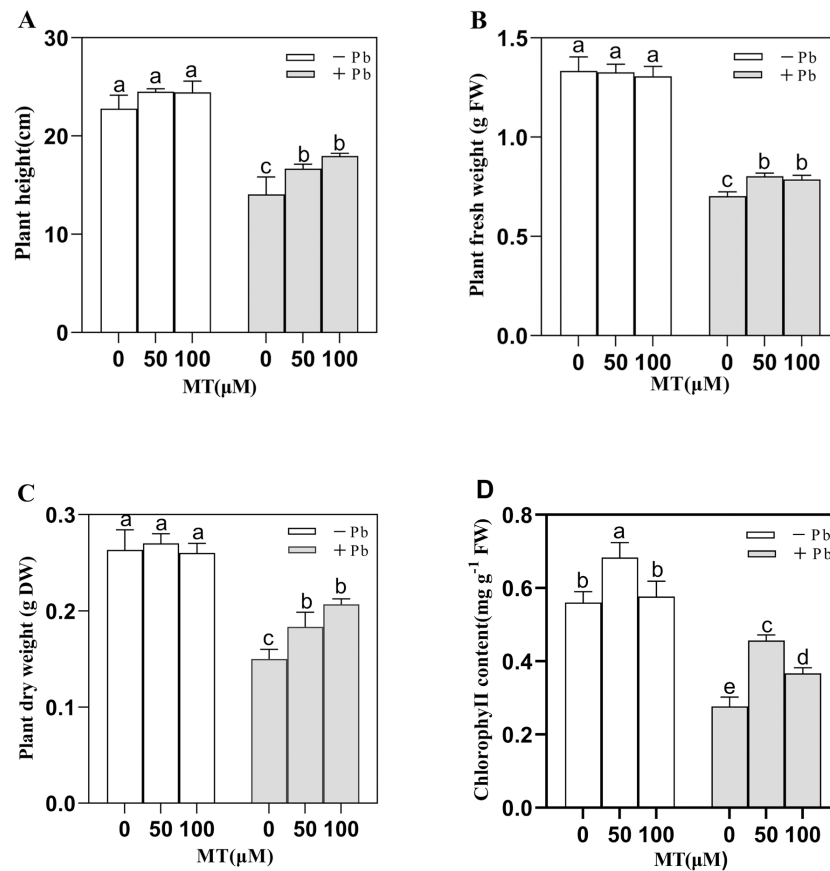
In the simulated experimental group (Control, Pb, MT<sub>50</sub>, MT<sub>100</sub>, MT<sub>50</sub> +Pb, MT<sub>100</sub> +Pb), on the 7th day (Fig. 1), the Pb solution decreased the plant height, fresh weight, dry weight and total chlorophyll content of naked oat seedlings. Under Pb stress, MT treatment improved seedlings growth with the plant height of seedlings increasing by 27.8% in MT<sub>100</sub> +Pb group, and 18.6% in MT<sub>50</sub> +Pb group compared with Pb group (Fig. 1A). The fresh weight of seedlings decreased by 47.2% after Pb treatment, which was significantly alleviated by the application of MT (Fig. 1B). Compared with the the Pb group, the dry weight of MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb groups increased by 20% and 33.3%, respectively (Fig. 1C). The chlorophyll content of the seedlings treated with 50 and 100  $\mu$ M MT increased by 21.4% and 1.7% respectively compared with the control group, and the effect of 50  $\mu$ M MT was more significant. Pb stress decreased the total chlorophyll content, but after MT treatment, the total chlorophyll content of naked oat seedlings increased considerably, with leaves also turning to green. Furthermore, the toxic symptoms were alleviated, with the total chlorophyll content increasing by 66.7% and 33.3% respectively compared with the Pb group. It can be observed that Pb inhibited the growth of naked oat seedlings, while the application of exogenous MT improved the tolerance of naked oat seedlings to Pb.

### Exogenous application of MT enhances the activity of antioxidant enzymes in naked oat seedling cells

SOD, POD and CAT are important protective enzymes in plants, and they are all important indicators of plant resistance to stress. Pb stress changed the antioxidant enzyme activity of naked oat seedlings (Fig. 2), in which SOD, POD, CAT activity increased. After 50 and 100  $\mu$ M MT pretreatment, SOD activity was significantly increased by 84.0% and 76.4%, respectively. Interestingly, the efficiency of MT to increase SOD activity decreased under Pb stress, but after MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb treatment, it increased 31.8% and 28.1% respectively compared with Pb group. Compared with the Pb stress group, the POD activity of the group treated with MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb increased by 50.7% and 62.0%, while the activity of CAT increased by 49.5% and 36.0%. The results showed that exogenous MT could enhance the activity of antioxidant enzymes in naked oats cells under Pb stress, so as to strengthen the antioxidant system of plants and reduce the toxicity of Pb to naked oats.

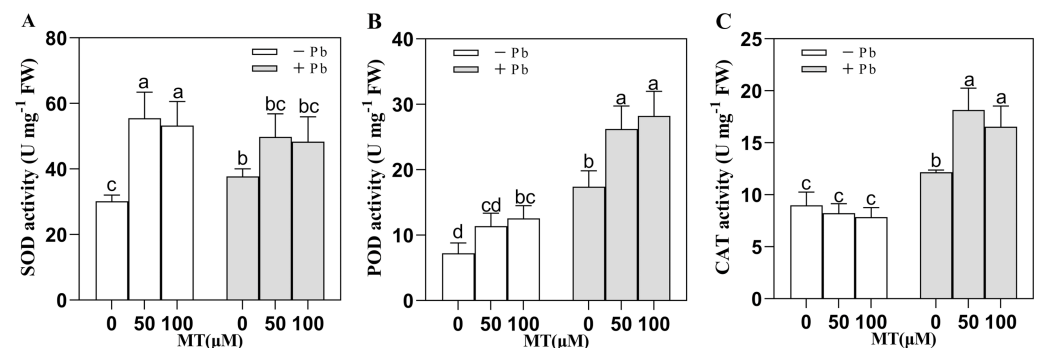
### Changes of osmotic regulation substances in naked oat seedlings

After naked oat seedlings cells sensed the external Pb stress, plant soluble proteins and proline increased or decreased accordingly to maintain the redox balance and alleviate the Pb poisoning of plant cells (Fig. 3). MT<sub>50</sub> and MT<sub>100</sub> treatment did not cause significant changes in intracellular soluble protein content in the control group, and Pb stress decreased the soluble protein content, but after 50  $\mu$ M or 100  $\mu$ M MT pretreatment, the soluble protein content increased. Compared with the Pb group, the soluble protein content of MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb group increased by 99.3% and 96.9%, respectively (Fig. 3A). The content of proline increased by 31.5% and 24.2% in MT<sub>50</sub> and MT<sub>100</sub> treatments, respectively compared with the control group (Fig. 3B). Pb stress significantly increased



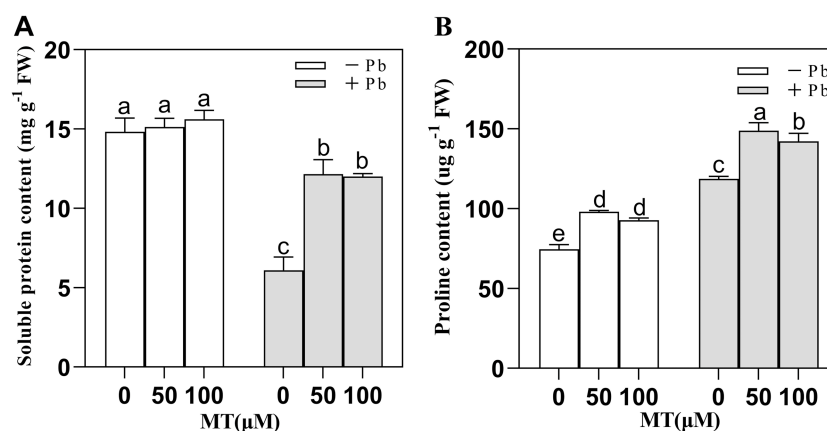
**Figure 1** Effects of different concentrations of MT on the growth of naked oat seedlings under lead stress. (A) Plant height; (B) plant fresh weight; (C) plant dry weight; (D) chlorophyll content. – Pb means lead-free and it is not a stress, + Pb means 75 mg L<sup>-1</sup> lead stress. Data are shown as means ± SD (*n* = 3). Different letters indicate a significant difference between control and treatment (*P* < 0.05).

Full-size [DOI: 10.7717/peerj.13978/fig-1](https://doi.org/10.7717/peerj.13978/fig-1)



**Figure 2** Effect of MT treatment on enzyme activity of naked oat seedlings under lead stress. (A) SOD; (B) POD; (C) CAT. – Pb means lead-free and it is not a stress, + Pb means 75 mg L<sup>-1</sup> lead stress. Data are shown as means ± SD (*n* = 3). Different letters indicate a significant difference between control and treatment (*P* < 0.05).

Full-size [DOI: 10.7717/peerj.13978/fig-2](https://doi.org/10.7717/peerj.13978/fig-2)



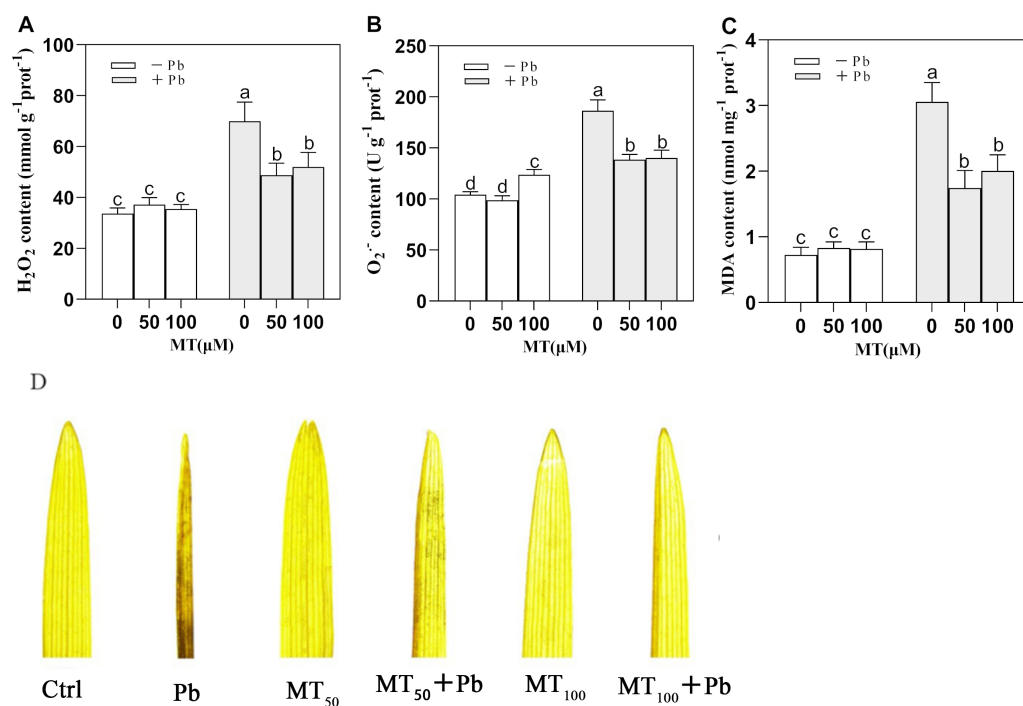
**Figure 3** Effects of different concentrations of melatonin on osmotic regulatory substances in naked oat seedlings under lead stress. (A) soluble protein content; (B) proline content. – Pb means lead-free and it is not a stress, + Pb means 75 mg L<sup>-1</sup> lead stress. Data are shown as means ± SD (*n* = 3). Different letters indicate a significant difference between control and treatment (*P* < 0.05).

Full-size DOI: 10.7717/peerj.13978/fig-3

proline content, and the effect of MT treatment was more significant. In MT<sub>50</sub> +Pb treatment group it increased by 25.4% while for MT<sub>100</sub> +Pb group the number was 19.8%. Therefore, MT might eliminate the increase of ROS in naked oat seedlings caused by Pb stress, stabilize the molecular structure of soluble proteins and enhance the tolerance of naked oats to heavy metal Pb stress. At the same time, the ROS of naked oat seedlings were elevated under Pb stress, and MT application could effectively alleviate the accumulation of ROS in naked oat seedlings.

Pb stress caused a significant increase in the contents of H<sub>2</sub>O<sub>2</sub> and O<sub>2</sub><sup>•-</sup> in young leaves of naked oats (Fig. 4). MT could significantly alleviate this phenomenon. Compared with the Pb group, the contents of H<sub>2</sub>O<sub>2</sub> in MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb groups decreased by 30.3% and 25.7%, respectively, and the content of O<sub>2</sub><sup>•-</sup> decreased by 25.7% and 24.7%, respectively. In the experiment, the content of MDA increased significantly under Pb stress, but MT treatment could reduce the content of MDA, and the content of MDA decreased by 42.9% and 34.4% in MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb groups, respectively (Fig. 4C). DAB histochemical staining of naked oat seedling leaves (Fig. 4D) showed that Pb stress induced oxidative stress of naked oats, and a large amount of H<sub>2</sub>O<sub>2</sub> was accumulated in seedling leaves, which affected the leaf morphology. After Pb poisoning, the seedling leaves became smaller and narrower, and the DAB staining was aggravated. After MT treatment, the symptoms of the leaves treated with MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb were relieved, and the accumulation of H<sub>2</sub>O<sub>2</sub> was reduced. The above results showed that after the Pb ion entered into naked oat seedlings, oxidative stress produced O<sub>2</sub><sup>•-</sup>, H<sub>2</sub>O<sub>2</sub> and other reactive oxygen species, which could be reduced by external application of MT. It is speculated that MT might be used as an antioxidant to help plants remove excessive ROS, lessen the degree of membrane lipid peroxidation in naked oat leaves, improve their tolerance under Pb stress, and improve the growth of naked oat seedlings.





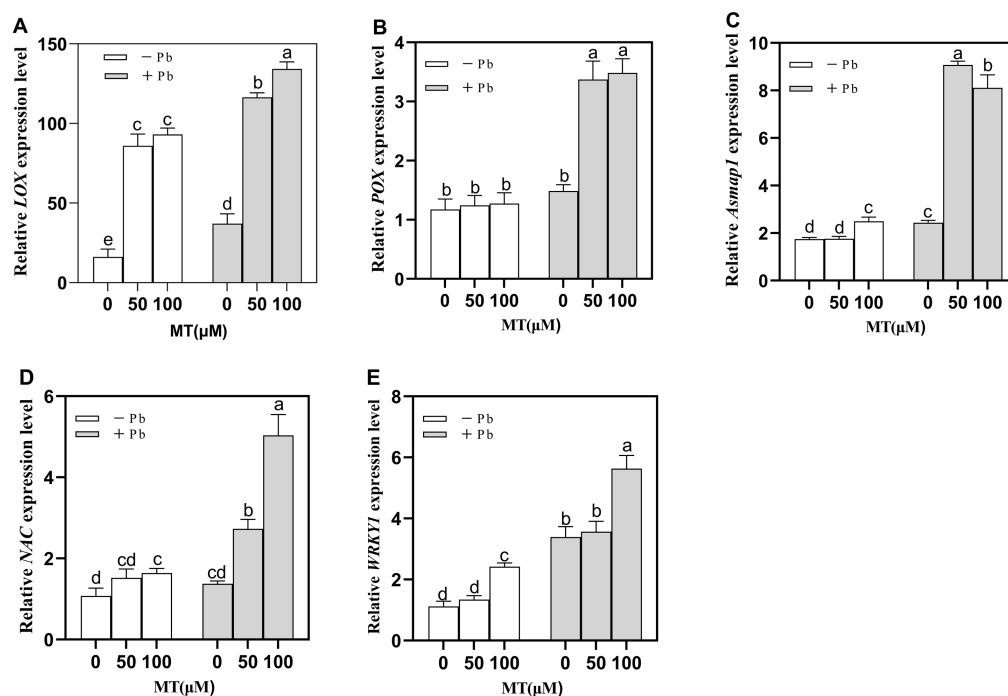
**Figure 4** Effects of different concentrations of MT on the changes of intracellular reactive oxygen species in naked oat seedlings under lead stress. (A) H<sub>2</sub>O<sub>2</sub> content; (B) O<sub>2</sub><sup>•-</sup> content; (C) MDA content; (D) DAB staining histochemical analysis. - Pb means lead-free and it is not a stress, + Pb means 75 mg L<sup>-1</sup> lead stress. Data are shown as means ± SD (*n* = 3). Different letters indicate a significant difference between control and treatment (*P* < 0.05).

Full-size DOI: 10.7717/peerj.13978/fig-4

## Exogenous MT application increases the expression of genes related to oat seedlings

The expression of Lipoxygenase (*LOX*) and peroxygenase (*POX*) of naked oats changed significantly after exogenous application of MT (Figs. 5A and 5B). A total of 50 and 100 μM MT significantly increased the expression of *LOX* by 431.2% and 475.5%. Under Pb stress, the expression of *LOX* in MT<sub>50</sub>+Pb and MT<sub>100</sub>+Pb groups was also significantly increased by 214.3% and 262.3% (Fig. 5A). *POX* gene expression also increased significantly after Pb stress, and exogenous MT pretreatment increased 127.7% and 135.1% respectively (Fig. 5B). The results showed that MT treatment changed the expression level of lipid peroxidase in naked oat cells under Pb stress, which may protect plants from oxidative stress by up-regulating *LOX* and *POX*.

*Asmap1* gene is a MAPK (mitogen activated protein kinase) protein in naked oats. After MT pretreatment, there was no significant change in MAPK cascade response in the Control group, after Pb treatment, *Asmap1* gene expression increased significantly by 273.3% and 233.7% respectively (Fig. 5C) in MT<sub>50</sub>+Pb and MT<sub>100</sub>+Pb groups treated with MT, indicating that MT<sub>50</sub> can induce a stronger MAPK cascade response. It is suggested that MT can induce gene upregulation in MAPK cascade reaction and enhance the tolerance of naked oat seedlings to Pb stress.



**Figure 5** Effects of different concentrations of MT on the expression of *LOX* (A), *POX* (B), *Asmap1* (C), *NAC* (D), *WRKY1* (E) in leaves of naked oat seedlings under lead stress. – Pb means lead-free and it is not a stress, + Pb means 75 mg L<sup>-1</sup> lead stress. Data are shown as means ± SD (n = 3). Different letters indicate a significant difference between control and treatment (P < 0.05).

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Compared to the Pb group (Fig. 5D), the gene (*NAC*) expression of MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb groups increased significantly by 99.1% and 267.2% respectively. Compared to the Pb group, the gene (*WRKY1*) expression of MT<sub>50</sub> +Pb and MT<sub>100</sub> +Pb groups increased by 5.0% and 66.1%. It is speculated that exogenous MT may affect the expression of related TFs in naked oat seedlings in some way, and further enhance the tolerance of naked oat seedlings to Pb stress.

## DISCUSSION

MT can reduce the oxidative stress damage caused by heavy metals in some plants (Sarafi *et al.*, 2017). The differences of its concentration and treatment methods, stress, as well as environments have distinctive effects on the same plant. The Pb tolerance of plants is the result of synergistic function in plants, and one of the indicators is plant biomass (Liu, Li & Shen, 2007). Under 75 mg L<sup>-1</sup> Pb stress, the plant height, fresh weight and dry weight of naked oats seedlings were reduced, the growth slowed down, and serious wilting appeared. After exogenous MT pretreatment, the wilting growth state of naked oats seedlings was effectively alleviated and gradually returned to normal, this is similar to Hasan *et al.* (2015).

Photosynthesis maintains plant growth and development, and the change of chlorophyll content in chloroplast can be used as a marker to measure photosynthetic efficiency (Lee *et al.*, 2003; Xiong, Zhao & Li, 2006). In our experiment, Pb stress led to the decrease

of chlorophyll content which could be increased by MT treatment in varying degrees. Chlorophyll content is closely related to the dry weight of the plant (Hu et al., 2018), the increase of chlorophyll content may also increase the dry weight of naked oat seedlings. So we speculated that MT could improve the photosynthetic efficiency and help the seedlings to restore growth.

Singh et al. (2010) found that under the stress of heavy metal Pb, a large amount of ROS in indica rice cells could accumulate and destroy the balance in the oxidation system and aggravate membrane lipid peroxidation. The activities of SOD, POD and CAT in naked oats seedlings induced by Pb stress did not change significantly, while exogenous MT increased the activities of the three. The increase of activity was related to the concentration of MT, in which POD and CAT changed significantly, while SOD activity was not sensitive to Pb. Overall, MT application enhanced the activity of antioxidant enzymes, which is similar to the findings of Xia et al. (2020). At the same time, metal Pb caused a large accumulation of ROS and a significant increase of  $\text{H}_2\text{O}_2$  and  $\text{O}_2^{\bullet-}$  in naked oat cells, and MT could effectively alleviate this phenomenon, lessen the oxidative damage of young leaves (Fig. 5), and help naked oat seedlings to resist the toxicity of metal Pb.

Heavy metals can damage intracellular soluble proteins (Yadav, 2010).  $75 \text{ mg L}^{-1}$  Pb stress significantly decreased the content of soluble proteins in naked oat seedlings while MT treatment increased it. It is speculated that MT as an active oxygen scavenger maintains intracellular homeostasis and alleviates metal-induced protein damage, which is consistent with the discovery by Namdjoyan et al. (2020). Free proline in plant osmotic regulators increases under stress, which helps to alleviate the toxic effects caused by abiotic stress and protect plant cells from the harm of free radicals (Soussana et al., 2004). The proline content of young leaves of naked oats treated with Pb was higher than that of the control group, which may be because the infiltration of metal Pb ions induced the change of water potential in the seedlings and destroyed the equilibrium state. MT under Pb stress can increase the content of proline, and proline can regulate cell osmotic pressure (Lehmann et al., 2010). It may regulate cell osmotic pressure balance and enhance the tolerance of naked oats to Pb by clearing excess ROS in cells. The content of MDA, the product of membrane lipid peroxidation, indirectly reflects the degree of damage to the membrane system (Atalay, Gegotek & Skrzydlewska, 2021). This study showed that the application of melatonin significantly reduced MDA levels, further demonstrating that melatonin can maintain osmotic balance in naked oat seedlings. In conclusion, the exogenous application of melatonin maintained the osmotic balance of naked oat seedlings, and we speculate that the increase in fresh weight of naked oat seedlings may be related to this.

Usually,  $\text{H}_2\text{O}_2$ , as a signal molecule with low concentration, participates in a variety of physiological and metabolic regulation in plants (Nazir, Fariduddin & Khan, 2020) and regulates the expression of transcription factors in bacteria, animals and lower eukaryotes (Marinho et al., 2014). Exogenous application of melatonin can significantly reduce the  $\text{H}_2\text{O}_2$  content in naked oat seedlings. We speculate that MT may promote the expression of stress resistance genes by reducing the  $\text{H}_2\text{O}_2$  concentration. MAPK cascade reaction is a common cascade reaction in abiotic stress (Wang, He & Yang, 2012). *Asmap1* is an important gene in MAPK. We found that *Asmap1* was significantly up-regulated after

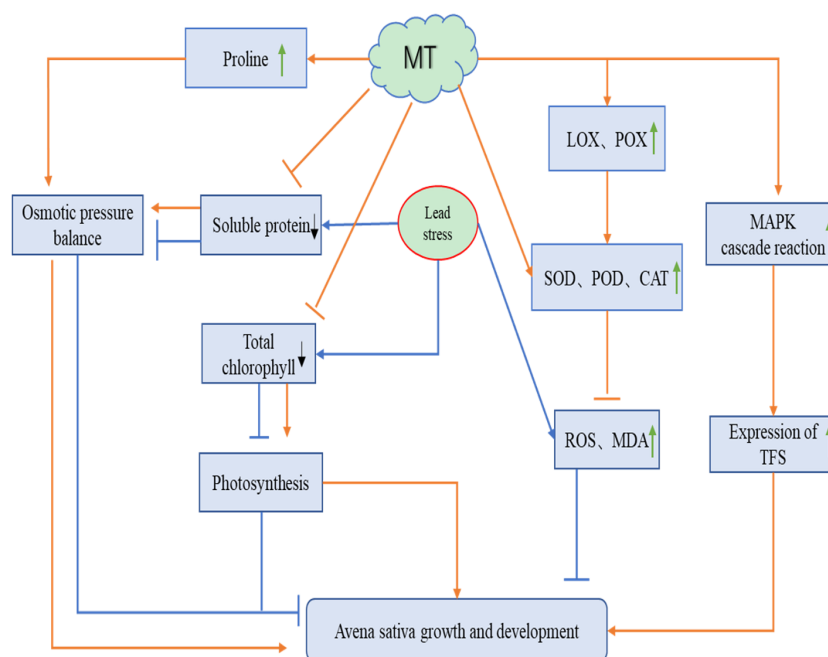
melatonin was applied. Therefore, MT may enhance plant Pb tolerance by affecting MAPK pathway. Transcription factor (TFs) regulates the expression of coding genes, especially the expression of early stress response genes and a series of downstream target genes, which may enhance plant stress resistance (Hoang et al., 2017). Genes such as NAC and WRKY are involved in a variety of abiotic stress responses, and the overexpression of NAC and WRKY genes can regulate the senescence process of *Arabidopsis thaliana* leaves (Guo, Cai & Gan, 2004). In this study, exogenous MT significantly increased the expression of NAC and WRKY1. MT may also promote the growth of naked oat seedlings under Pb stress by affecting the expression of TFs. In general, MT may affect the MAPK cascade and the expression of TFs, thus improving the Pb tolerance of naked oat seedlings.

LOX and POX genes in plants are involved in a variety of signal pathways (Santino et al., 2013), helping the body resist adverse external factors. Under salt stress, the activity of lipoxygenase in the cells of citrus seedlings increased (Ben-Hayyim et al., 2001), and the 9-LOX gene expression increased three to six times under osmotic stress (Fedina et al., 2004). Similarly, naked oat seedlings in this study showed slightly elevated expression levels of LOX and POX under Pb stress. LOX may help activate antioxidant enzyme system (Cho et al., 2012). The expression levels of LOX and POX were significantly increased in naked oat seedlings after exogenous application of different concentrations of MT, and we speculate that exogenous MT may enhance the resistance of oat seedlings by regulating the expression levels of LOX and POX. This is similar to the results of the study by Gao et al. (2018).

This study provides the first evidence that MT may help to alleviate the damage suffered by naked oat seedlings under Pb stress. In some areas, crops are experiencing heavy metal contamination (Onakpa, Njan & Kalu, 2018), especially in acidic soils, which tend to accumulate more heavy metals. It is urgent to reduce the negative impact of heavy metals in soils on crops. On the one hand, MT can alleviate heavy metal toxicity in plants, while at the same time improve soil enzyme activity and soil quality by altering the composition of soil bacterial and fungal communities, thereby promoting plant growth (Acuña Castroviejo et al., 2014; Li et al., 2018; Madigan et al., 2019; Moustafa-Farag et al., 2020). On the other hand, MT may also alleviate abiotic stresses to which plants are subjected in other ways (Chang et al., 2021), such as by affecting circadian rhythms. Soil Pb concentrations in some vegetable gardens were much higher than those in this experiment (Amorim et al., 2021), and the therapeutic effect of MT remained stable in the short term (Yan et al., 2021). In summary, MT has the potential to be applied in future agricultural practices as one of the substances to improve the growing environment of crops such as naked oats.

## CONCLUSIONS

A total of 75 mg L<sup>-1</sup> Pb stress severely affected the growth of naked oat seedlings, while different concentrations of MT all alleviated the damage caused by Pb stress to oat. We speculated that MT may act on naked oat seedlings under Pb stress in the following ways (Fig. 6): exogenous MT can increase chlorophyll content and enhance photosynthesis in oat seedlings. It can also promote changes in osmoregulatory substances such as



**Figure 6** Potential role of MT on the growth and development of naked oat seedlings under lead stress. The orange line indicates the pathway of action for melatonin (MT) and the blue line indicates the pathway of action for lead stress. Green arrows indicate an increase, black arrows indicate a decrease.

Full-size [DOI: 10.7717/peerj.13978/fig-6](https://doi.org/10.7717/peerj.13978/fig-6)

proline to restore the growth and development of Pb-stressed naked oat seedlings. Pb stress significantly increased the level of reactive oxygen species in oat seedlings, while exogenous MT was able to significantly reduce the level of reactive oxygen species in oat seedlings by increasing the activity of antioxidant enzymes. Exogenous MT up-regulated the expression of genes related to the antioxidant system, activated the MAPK cascade reaction and promoted the expression of TFs genes, thus improving the resistance of naked oat seedlings to Pb. The relationship between the mechanism of the function of MT and related signaling pathways in naked oat seedlings under heavy metal Pb stress needs to be further explored.

## ADDITIONAL INFORMATION AND DECLARATIONS

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### Competing Interests

The authors declare there are no competing interests.

### Author Contributions

- Kai Wang performed the experiments, prepared figures and/or tables, and approved the final draft.
- Jinjin He performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Ningbo Zhao analyzed the data, prepared figures and/or tables, and approved the final draft.
- Yajing Zhao analyzed the data, prepared figures and/or tables, and approved the final draft.
- Fangbing Qi analyzed the data, prepared figures and/or tables, and approved the final draft.
- Fenggui Fan conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Yingjuan Wang conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.

### Data Availability

The following information was supplied regarding data availability:

The raw measurements are available in the [Supplementary File](#).

### Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.13978#supplemental-information>.

## REFERENCES

- Amorim HCS, Hurtarte LCC, Vergütz L, Silva IR, Costa ODV, Pacheco AA, Fontes MP. 2021.** Lead speciation and availability affected by plants in a contaminated soil. *Chemosphere* **285**:131468 DOI [10.1016/j.chemosphere.2021.131468](https://doi.org/10.1016/j.chemosphere.2021.131468).
- Aslam M, Aslam A, Sheraz M, Ali B, Ulhassan Z, Najeeb U, Zhou W, Gill RA. 2020.** Lead toxicity in cereals: mechanistic insight into toxicity, mode of action, and management. *Frontiers in Plant Science* **11**:587785 DOI [10.3389/fpls.2020.587785](https://doi.org/10.3389/fpls.2020.587785).
- Atalay S, Gęgotek A, Skrzydlewska E. 2021.** Protective effects of cannabidiol on the membrane proteome of UVB-irradiated keratinocytes. *Antioxidants* **10**:402 DOI [10.3390/antiox10030402](https://doi.org/10.3390/antiox10030402).
- Bates LS, Waldren RP, Teare I. 1973.** Rapid determination of free proline for water-stress studies. *Plant and Soil* **39**:205–207 DOI [10.1007/BF00018060](https://doi.org/10.1007/BF00018060).

- Ben-Hayyim G, Gueta-Dahan Y, Avsian-Kretchmer O, Weichert H, Feussner I. 2001.** Preferential induction of a 9-lipoxygenase by salt in salt-tolerant cells of *Citrus sinensis* L. Osbeck. *Planta* **212**:367–375 DOI [10.1007/s004250000397](https://doi.org/10.1007/s004250000397).
- Cao YY, Qi CD, Li S, Wang Z, Wang X, Wang J, Ren S, Li X, Zhang N, Guo YD. 2019.** Melatonin alleviates copper toxicity via improving copper sequestration and ROS scavenging in cucumber. *Plant & Cell Physiology* **60**:562–574 DOI [10.1093/pcp/pcy226](https://doi.org/10.1093/pcp/pcy226).
- Acuña Castroviejo D, Escames G, Venegas C, Díaz-Casado ME, Lima-Cabello E, López LC, Rosales-Corral S, Tan D-X, Reiter RJ. 2014.** Extrapineal melatonin: sources, regulation, and potential functions. *Cellular and Molecular Life Sciences* **71**:2997–3025 DOI [10.1007/s00018-014-1579-2](https://doi.org/10.1007/s00018-014-1579-2).
- Chang T, Zhao Y, He H, Xi Q, Fu J, Zhao Y. 2021.** Exogenous melatonin improves growth in hulless barley seedlings under cold stress by influencing the expression rhythms of circadian clock genes. *PeerJ* **9**:e10740 DOI [10.7717/peerj.10740](https://doi.org/10.7717/peerj.10740).
- Cho K, Kim YC, Woo JC, Rakwal R, Agrawal GK, Yoeun S, Han O. 2012.** Transgenic expression of dual positional maize lipoxygenase-1 leads to the regulation of defense-related signaling molecules and activation of the antioxidative enzyme system in rice. *Plant Science* **185**:238–245 DOI [10.1016/j.plantsci.2011.10.016](https://doi.org/10.1016/j.plantsci.2011.10.016).
- Elavarthi S, Martin B. 2010.** Spectrophotometric assays for antioxidant enzymes in plants. *Plant Stress Tolerance* **639**:273–280 DOI [10.1007/978-1-60761-702-016](https://doi.org/10.1007/978-1-60761-702-016).
- Fedina EO, Karimova FG, Chechetkin IR, Tarchevsky IA, Khripach VA. 2004.** Contribution of lipoxygenase metabolism to the brassinosteroid signaling pathway. *Doklady Biochemistry & Biophysics* **395**:80–83 DOI [10.1023/b:doobi.0000025551.80557.42](https://doi.org/10.1023/b:doobi.0000025551.80557.42).
- Gao W, Zhang Y, Feng Z, Bai Q, He J, Wang Y. 2018.** Effects of melatonin on antioxidant capacity in naked oat seedlings under drought stress. *Molecules* **23**:1580 DOI [10.3390/molecules23071580](https://doi.org/10.3390/molecules23071580).
- Genty B, Briantais J-M, Baker NR. 1989.** The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta (BBA)-General Subjects* **990**:87–92 DOI [10.1016/s0304-4165\(89\)80016-9](https://doi.org/10.1016/s0304-4165(89)80016-9).
- Gill SS, Tuteja N. 2010.** Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry* **48**:909–930 DOI [10.1016/j.plaphy.2010.08.016](https://doi.org/10.1016/j.plaphy.2010.08.016).
- Guo Y, Cai Z, Gan S. 2004.** Transcriptome of Arabidopsis leaf senescence. *Plant Cell & Environment* **27**:521–549 DOI [10.1111/j.1365-3040.2003.01158.x](https://doi.org/10.1111/j.1365-3040.2003.01158.x).
- Hasan MK, Ahammed GJ, Yin L, Shi K, Xia X, Zhou Y, Yu J, Zhou J. 2015.** Melatonin mitigates cadmium phytotoxicity through modulation of phytochelatin biosynthesis, vacuolar sequestration, and antioxidant potential in *Solanum lycopersicum* L. *Frontiers in Plant Science* **6**:601 DOI [10.3389/fpls.2015.00601](https://doi.org/10.3389/fpls.2015.00601).
- Hattori A, Migitaka H, Iigo M, Itoh M, Yamamoto K, Ohtani-Kaneko R, Hara M, Suzuki T, Reiter RJ. 1995.** Identification of melatonin in plants and its effects on plasma melatonin levels and binding to melatonin receptors in vertebrates. *Biochemistry and Molecular Biology International* **35**:627–634.

- Hoang XLT, Nhi DNH, Thu NBA, Thao NP, Tran LP. 2017.** Transcription factors and their roles in signal transduction in plants under abiotic stresses. *Current Genomics* **18**:483–497 DOI [10.2174/1389202918666170227150057](https://doi.org/10.2174/1389202918666170227150057).
- Hu Y, Zhang Y, Yu W, Hänninen H, Song L, Du X, Zhang R, Wu J. 2018.** Novel insights into the influence of seed sarcotesta photosynthesis on accumulation of seed dry matter and oil content in *torreya grandis* cv. *Merrillii*. *Frontiers in Plant Science* **8**:2179 DOI [10.3389/fpls.2017.02179](https://doi.org/10.3389/fpls.2017.02179).
- Jahan MS, Guo S, Baloch AR, Sun J, Shu S, Wang Y, Ahammed GJ, Kabir K, Roy R. 2020.** Melatonin alleviates nickel phytotoxicity by improving photosynthesis, secondary metabolism and oxidative stress tolerance in tomato seedlings. *Ecotoxicology and Environmental Safety* **197**:110593 DOI [10.1016/j.ecoenv.2020.110593](https://doi.org/10.1016/j.ecoenv.2020.110593).
- Khan TA, Nadeem F, Chen L, Wang X, Zeng Z, Hu Y. 2019.** Enhancing naked oat (*Avena nuda* L.) productivity with minimal indirect nitrogen loss and maximum nitrogen use efficiency through integrated use of different nitrogen sources. *PLOS ONE* **14**:e0213808 DOI [10.1371/journal.pone.0213808](https://doi.org/10.1371/journal.pone.0213808).
- Lee S, Moon JS, Ko TS, Petros D, Goldsbrough PB, Korban SS. 2003.** Overexpression of arabidopsis phytochelatin synthase paradoxically leads to hypersensitivity to cadmium stress. *Plant Physiology* **131**:656–663 DOI [10.1104/pp.014118](https://doi.org/10.1104/pp.014118).
- Lehmann S, Funck D, Szabados L, Rentsch D. 2010.** Proline metabolism and transport in plant development. *Amino Acids* **39**:949–962 DOI [10.1007/s00726-010-0525-3](https://doi.org/10.1007/s00726-010-0525-3).
- Lewitt MS, Hulting AL. 2017.** Corrigendum to Professor Kerstin Hall (1929-2017): Pioneer in the field of Growth Hormone and IGF Research [Growth Horm. IGF Res. 34 (2017) 28-30. DOI: 10.1016/j.ghir.2017.05.001. Epub 2017 May 3]. *Growth Hormone & IGF Research* **35**:28–30 DOI [10.1016/j.ghir.2017.06.003](https://doi.org/10.1016/j.ghir.2017.06.003).
- Li C, Zhao Q, Gao T, Wang H, Zhang Z, Liang B, Wei Z, Liu C, Ma F. 2018.** The mitigation effects of exogenous melatonin on replant disease in apple. *Journal of Pineal Research* **65**:e12523 DOI [10.1111/jpi.12523](https://doi.org/10.1111/jpi.12523).
- Li J, Yang Y, Sun K, Chen Y, Chen X, Li X. 2019.** Exogenous melatonin enhances cold, salt and drought stress tolerance by improving antioxidant defense in tea plant (*Camellia sinensis* (L.) O. Kuntze). *Molecules* **24**:1826 DOI [10.3390/molecules24091826](https://doi.org/10.3390/molecules24091826).
- Liu MM, Li JN, Shen YX. 2007.** Effect of Pb<sup>2+</sup> pollution on seed germination and seedling growth of multiflora ryegrass. *Grass Science* **24**:52–54.
- López-Orenes A, Dias MC, Ferrer MÁ, Calderón A, Moutinho-Pereira J, Correia C, Santos C. 2018.** Different mechanisms of the metalliferous *Zygophyllum fabago* shoots and roots to cope with Pb toxicity. *Environmental Science and Pollution Research* **25**:1319–1330 DOI [10.1007/s11356-017-0505-1](https://doi.org/10.1007/s11356-017-0505-1).
- Madigan AP, Egidi E, Bedon F, Franks AE, Plummer KM. 2019.** Bacterial and fungal communities are differentially modified by melatonin in agricultural soils under abiotic stress. *Frontiers in Microbiology* **10**:2616 DOI [10.3389/fmicb.2019.02616](https://doi.org/10.3389/fmicb.2019.02616).
- Marinho HS, Real C, Cyrne L, Soares H, Antunes F. 2014.** Hydrogen peroxide sensing, signaling and regulation of transcription factors. *Redox Biology* **2**:535–562 DOI [10.1016/j.redox.2014.02.006](https://doi.org/10.1016/j.redox.2014.02.006).



- Moustafa-Farag M, Elkelish A, Dafea M, Khan M, Arnao MB, Abdelhamid MT, El-Ezz AA, Almoneafy A, Mahmoud A, Awad M, Li L, Wang Y, Hasanuzzaman M, Ai S. 2020. Role of melatonin in plant tolerance to soil stressors: salinity, pH and heavy metals. *Molecules* 25:5359 DOI 10.3390/molecules25225359.
- Namdjoyan S, Soorki AA, Elyasi N, Kazemi N, Simaei M. 2020. Melatonin alleviates lead-induced oxidative damage in safflower (*Carthamus tinctorius* L.) seedlings. *Ecotoxicology* 29:108–118 DOI 10.1007/s10646-019-02136-9.
- Nawaz MA, Jiao Y, Chen C, Shireen F, Zheng Z, Imtiaz M, Bie Z, Huang Y. 2018. Melatonin pretreatment improves vanadium stress tolerance of watermelon seedlings by reducing vanadium concentration in the leaves and regulating melatonin biosynthesis and antioxidant-related gene expression. *Journal of Plant Physiology* 220:115–127 DOI 10.1016/j.jplph.2017.11.003.
- Nazir F, Fariduddin Q, Khan TA. 2020. Hydrogen peroxide as a signalling molecule in plants and its crosstalk with other plant growth regulators under heavy metal stress. *Chemosphere* 252:126486 DOI 10.1016/j.chemosphere.2020.126486.
- Onakpa MM, Njan AA, Kalu OC. 2018. A review of heavy metal contamination of food crops in Nigeria. *Annals of Global Health* 84:488–494 DOI 10.29024/aogh.2314.
- Orozco-Cardenas M, Ryan CA. 1999. Hydrogen peroxide is generated systemically in plant leaves by wounding and systemin via the octadecanoid pathway. *Proceedings of the National Academy of Sciences of the United States of America* 96:6553–6557 DOI 10.1073/pnas.96.11.6553.
- Pace R, Liberati D, Sconocchia P, De Angelis P. 2020. Lead transfer into the vegetation layer growing naturally in a Pb-contaminated site. *Environmental Geochemistry and Health* 42:2321–2329 DOI 10.1007/s10653-019-00429-w.
- Pellinen V, Cherkashina T, Gustaytis M. 2021. Assessment of metal pollution and subsequent ecological risk in the coastal zone of the Olkhon Island, Lake Baikal, Russia. *Science of the Total Environment* 786:147441 DOI 10.1016/j.scitotenv.2021.147441.
- Pérez-Chaca MV, Rodríguez-Serrano M, Molina AS, Pedranzani HE, Zirulnik F, Sandalio LM, Romero-Puertas MC. 2014. Cadmium induces two waves of reactive oxygen species in Glycine max (L.) roots. *Plant, Cell & Environment* 37:1672–1687 DOI 10.1111/pce.12280.
- Porra RJ, Thompson WA, Kriedemann PE. 1989. Determination of accurate extinction coefficients and simultaneous equations for assaying chlorophylls a and b extracted with four different solvents: verification of the concentration of chlorophyll standards by atomic absorption spectroscopy. *Biochimica et Biophysica Acta (BBA)-Bioenergetics* 975:384–394 DOI 10.1016/s0005-2728(89)80347-0.
- Rahman Z, Singh VP. 2019. The relative impact of toxic heavy metals (THMs) (arsenic (As), cadmium (Cd), chromium (Cr)(VI), mercury (Hg), and lead (Pb)) on the total environment: an overview. *Environmental Monitoring and Assessment* 191:419 DOI 10.1007/s10661-019-7528-7.

- Reiter RJ, Mayo JC, Tan DX, Sainz RM, Alatorre-Jimenez M, Qin L. 2016. Melatonin as an antioxidant: under promises but over delivers. *Journal of Pineal Research* 61:253–278 DOI 10.1111/jpi.12360.
- Santino A, Taurino M, Domenico SD, Bonsegna S, Poltronieri P, Pastor V, Flors V. 2013. Jasmonate signaling in plant development and defense response to multiple (a)biotic stresses. *Plant Cell Reports* 32:1085–1098 DOI 10.1007/s00299-013-1441-2.
- Sarafi E, Tsouvaltzis P, Chatzissavvidis C, Siomos A, Therios I. 2017. Melatonin and resveratrol reverse the toxic effect of high boron (B) and modulate biochemical parameters in pepper plants (*Capsicum annuum* L.). *Plant Physiology and Biochemistry* 112:173–182 DOI 10.1016/j.plaphy.2016.12.018.
- Shi J, Li L, Pan G. 2009. Variation of grain Cd and Zn concentrations of 110 hybrid rice cultivars grown in a low-Cd paddy soil. *International Journal of Environmental Science and Technology* 21:168–172 DOI 10.1016/S1001-0742(08)62246-9.
- Singh R, Tripathi RD, Dwivedi S, Kumar A, Trivedi PK, Chakrabarty D. 2010. Lead bioaccumulation potential of an aquatic macrophyte *Najas indica* are related to antioxidant system. *Bioresource Technology* 101:3025–3032 DOI 10.1016/j.biortech.2009.12.031.
- Soussana JF, Loiseau P, Vuichard N, Ceschia E, Balesdent J, Chevallier T, Arrouays D. 2004. Carbon cycling and sequestration opportunities in temperate grasslands. *Soil Use and Management* 20:219–230 DOI 10.1079/sum2003234.
- Sun C, Liu L, Wang L, Li B, Jin C, Lin X. 2021. Melatonin: a master regulator of plant development and stress responses. *Journal of Integrative Plant Biology* 63:126–145 DOI 10.1111/jipb.12993.
- Tan XL, Fan ZQ, Kuang JF, Lu WJ, Reiter RJ, Lakshmanan P, Su XG, Zhou J, Chen JY, Shan W. 2019. Melatonin delays leaf senescence of Chinese flowering cabbage by suppressing ABFs-mediated abscisic acid biosynthesis and chlorophyll degradation. *Journal of Pineal Research* 67:e12570 DOI 10.1111/jpi.12570.
- Wang HQ. 2014. Study on the physiological toxicity of three heavy metals, Pb-Zn-Hg, to *Avena sativa*. Shanxi University.
- Wang H, He JJ, Yang XL. 2012. Stress effects of heavy metals on plants and their relationship to signal transduction. *Teaching Biology* 37:7–9.
- Wang M, Zhang S, Ding F. 2020. Melatonin mitigates chilling-induced oxidative stress and photosynthesis inhibition in tomato plants. *Antioxidants* 9:218 DOI 10.3390/antiox9030218.
- Xia H, Ni Z, Hu R, Lin L, Deng H, Wang J, Tang Y, Sun G, Wang X, Li H, Liao M, Lv X, Liang D. 2020. Melatonin alleviates drought stress by a non-enzymatic and enzymatic antioxidative system in kiwifruit seedlings. *International Journal of Molecular Sciences* 21:852 DOI 10.3390/ijms21030852.
- Xiong ZT, Zhao F, Li MJ. 2006. Lead toxicity in *Brassica pekinensis* Rupr.: effect on nitrate assimilation and growth. *Environmental Toxicology* 21:147–153 DOI 10.1002/tox.20167.

- Yadav SK. 2010.** Heavy metals toxicity in plants: An overview on the role of glutathione and phytochelatins in heavy metal stress tolerance of plants. *South African Journal of Botany* **76**:167–179 DOI [10.1016/j.sajb.2009.10.007](https://doi.org/10.1016/j.sajb.2009.10.007).
- Yan F, Wei H, Ding Y, Li W, Liu Z, Chen L, Tang S, Ding C, Jiang Y, Li G. 2021.** Melatonin regulates antioxidant strategy in response to continuous salt stress in rice seedlings. *Plant Physiology and Biochemistry* **165**:239–250 DOI [10.1016/j.plaphy.2021.05.003](https://doi.org/10.1016/j.plaphy.2021.05.003).
- Zhang JP. 2006.** Study on naked oats nutrition components and functions. *Food and Fermentation Industries* **32**:128–130.
- Zhao FJ, Ma Y, Zhu YG, Tang Z, McGrath SP. 2015.** Soil contamination in China: current status and mitigation strategies. *Environmental Science & Technology* **49**:750–759 DOI [10.1021/es5047099](https://doi.org/10.1021/es5047099).
- Zhao N, Yang J, Fang X, Li L, Yan H, Liu D. 2020.** First report of *colletotrichum cereale* causing anthracnose on *avena nuda* in China. *Plant Disease* **105**:1216 DOI [10.1094/pdis-10-20-2109-pdn](https://doi.org/10.1094/pdis-10-20-2109-pdn).
- Zhou R, Wan H, Jiang F, Li X, Yu X, Rosenqvist E, Ottosen CO. 2020.** The Alleviation of photosynthetic damage in tomato under drought and cold stress by high CO<sub>2</sub> and melatonin. *International Journal of Molecular Sciences* **21**:5587 DOI [10.3390/ijms21155587](https://doi.org/10.3390/ijms21155587).