

# Skeletal variation in bird domestication: limb proportions and sternum in chicken, with comparisons to mallard ducks and Muscovy ducks

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**Background.** Domestication, including selective breeding, can lead to morphological changes of biomechanical relevance. In birds, limbs proportion and sternum characteristics are of great importance and have been studied in the past for its relation with flight, terrestrial locomotion and animal welfare. In this work we studied the effects of domestication and breed formation in limb proportions in chicken, mallard ducks and Muscovy ducks.

**Methods.** Here we analysed the scaling of limb proportions. First, we quantified the proportional length of three long bones of forelimb (humerus, radius and carpometacarpus) and hind limb (femur, tibiotarsus and tarsometatarsus) of red jungle fowl and its domestic counterpart, chickens (*Gallus gallus*), and of analogous comparisons for mallard duck (*Anas platyrhynchos*) and Muscovy duck (*Cairina moschata*). We compared sternum characteristics in red jungle fowl and 9 chicken breeds.

**Results.** We found a lack of significant change in the proportions of long bones of chicken and Muscovy duck due to domestication, but significant differences in the case of mallard ducks. Variation of evolvability, allometric scaling, and heterochrony may serve to describe some of the patterns of change we report. Flight capacity loss in mallard ducks resulting from domestication may have a relation with the difference in proportions. The lack of variation in proportions that could distinguish domestic from wild forms of chicken and Muscovy ducks may reflect less intensive selection for flight capacity. In chicken, some of the differences identified in the traits discussed are breed-dependent. The study of the sternum revealed a higher frequency of notched carina sterni in the red junglefowl compared to domestic chicken. The condition of crooked keel, however, is not unique to chicken. Despite some similar morphological changes identified across species, this study highlights the lack of universal patterns in domestication and breed formation.

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## Abstract

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## Introduction

Through domestication, much morphological diversity has been generated (Darwin, 1868; Herre & Röhrs, 1990). Domesticated animals exhibit phenotypic changes compared to their wild counterparts (Clutton-Brock, 1999). This is the case in domestic birds, as in recent works on the skulls of pigeons and chickens (Young, et al., 2017; Stange, et al., 2018) and the integument in chickens (Núñez-León, et al., 2019). In contrast, postcranial anatomy, including limb bones, has rarely been dealt with analytically and globally in domesticated species (see Wayne 1986 for dogs; van Grouw 2018 for an overview for many species), despite having been widely studied in bird evolutionary biology (Middleton & Gatesy, 2000; Dyke & Nudds, 2009; Nudds, et al., 2013). Mentioned in Darwin's 1868 'Variation in Animals and Plants Under Domestication', changes in limb length (or lack thereof) in domestic birds can be observed. Especially notable is the reduction of wing length of the mallard duck after domestication (Darwin, 1868). Limb proportions can be of importance in biomechanical processes such as flight ability. Middleton and Gatesy (2000) highlighted the humerus relative proportion in the forelimb, where flightless birds have relatively longer humeri while flight manoeuvrability is being linked to shorter humeri. The selective pressure for flight is thus expected to influence the relative proportion of the different bones in the forelimb (Mason, 1984). Chickens are considered short-

distance non-migrant flyers, however their flight capability is breed-dependent (Schippers, et al., 2013). Although certain breeds are known to have completely lost their flight capacity (e.g. Ukokei and Polish breeds) this is largely attributed to the unfitness of their plumage for such task (Ekarius, 2007; Smith, 2018). Other breeds are short-distance flyers, including the wild type red junglefowl (RJF), but differences in flight capacity have been reported depending on the weight of the breed: the heavier, the less prone to sustained flight (Darwin, 1868; Schippers, et al., 2013).

It is generally agreed that domestic mallard ducks have lost their flight capacity when compared to their wild counterpart, which in contrast to chickens, are long distance migratory birds (Accordi & Barcellos, 2006). This change has been attributed to a gain in weight and reduction in the proportional wing length (Darwin, 1868; Cnotka, 2006). Because domestic Muscovy ducks can fly (Anon., 2016; Anon., 2017), their feathers are trimmed or clipped to discourage flight in breeding farm (Smith, 2007). Unlike the case of differences in wing length between domestic and wild mallards, finding osteological differences between domestic and wild Muscovy ducks has proven more difficult (Angulo, 1998; Stahl, et al., 2006).

Another skeletal element of biomechanical importance in birds is the sternum, the site of attachment of the pectoralis muscle, important for producing mechanical work during downstroke and pronating the wing, and the supracoracoideus muscle, which elevates and supinates the wing during upstroke (Biewener, 2011). This element varies in shape and proportions and is a source of intensive study in poultry research concerned with welfare due to widespread reports of damage ranging between 30 and 100% of commercial hens (Fleming, et al., 2004; Harlander-Matauschek, et al., 2015; Toscano, et al., 2020). Keel damage may include crookedness or bending, as well as fractures (Casey-Trott, et al., 2015). The problem appears somewhat historical as it was referred to by Darwin (1868, p. 282) as ‘generally so much deformed that it is scarcely possible to compare its form strictly in the several breeds’. The crookedness of the keel has been largely studied in the context of its causes, its relation with animal welfare, and its effect on productivity (Blount, 1933; Warren, 1937; Waters, 1949; Fleming, et al., 2004) and the market value of the domestic chicken (Hyre, 1995). Keel crookedness has been reported to have environmental and genetic origins, appearing mostly when chickens are given the opportunity to perch (Blount, 1933; Warren, 1937; Waters, 1949). However, whether crookedness of the keel is associated with domestication in its initial, less

intense phase (Vigne, 2011), or if it appeared only with breed formation and improvement has not been determined. Deformations of the keel can be divided into two types: fractures, defined as sharp bends, fragmented sections or shearing of the keel; and deviations, defined as abnormally shaped bone containing section(s) that vary from an ideally perfect two-dimensional straight plane that has not resulted from fracture (Casey-Trott, et al., 2015). Here we use the term ‘crookedness’ to denote exclusively the deviations, not fractures, of the keel, as defined in Casey-Trott et al. (2015).

In this study, we analysed the limb proportions of three domestic species: chicken (*Gallus gallus*), mallard duck (*Anas platyrhynchos*) and Muscovy duck (*Cairina moschata*) in order to determine whether there has been changes due to domestication and breed formation. We examined this matter in the context of evolvability, allometry, heterochrony, and selective pressures (artificial selection) that affect flight capacity. In the case of chickens, we examined these changes also among a diversity of breeds. Further, we investigated variation in the sternum by coding discrete features that characterize its variation between fowl and chicken in order to study differences among breeds. Specifically, we examined the crookedness of the keel, extensively studied in the past but never comparing breeds (Blount, 1933; Warren, 1937; Waters, 1949; Hyre, 1995; Fleming, et al., 2004; Kittelsen, et al., 2020), plus three other features: presence of a notch on the ventral margin of the carina sterni, sponginess or smoothness of the foramen pneumaticum and pars cardiaca and shape of the caudal end of the trabecula mediana.

## Materials & Methods

**Measurements.** We took measurements of the length of the humerus, radius and carpometacarpus of the forelimb and length of the femur, tibiotarsus and tarsometatarsus and of the hind limb. For each bone, we measured the length as the maximum length of the bone parallel to the measuring device (using digital calipers to the nearest tenth of a mm), coinciding with the anatomical structures shown in *Fig.1*.

We considered the proportion of a bone to be the proportion of that bone with respect to the total length of the limb. The total length of the forelimb was taken as the sum of the lengths of the carpometacarpus, radius and humerus, while the total length of the hind limb was taken as the sum of the lengths of the tarsometatarsus, tibiotarsus and femur (*Fig.2*).

**Specimens.** We compared proportions between the wild and domestic forms of different bird species: *Gallus gallus*, *Anas platyrhynchos* and *Cairina moschata* (Table 1). We chose these three species for their historical importance in bird domestication and wide availability in osteological collections where the labelling as wild and domestic forms is reliable. We only considered adult skeletally mature specimens for this work, with individuals considered adults when the epiphyses of all long bones were externally completely fused to the diaphyses. As the wild forebearer of the chicken we used the RJF (Al-Nasser, et al., 2007; Lawal, et al., 2020) although there has been introgression of grey junglefowl. (Eriksson, et al., 2008; Lawal, et al., 2020). For the mallard duck and the Muscovy duck, we have used their wild counterparts available in the visited museum osteological collections.

In *Gallus gallus*, we measured 29 RJFs and 103 domestic chickens for the forelimb and 28 RJFs and 99 chickens for the hind limb (Table 2). We excluded the modern meat lines (broilers) due to their short life span, resulting in an insufficient osteological maturation to be considered valid as adult specimens with the criterion used in this work, since the epiphyses of their long bones are not completely fused to the diaphyses (Zuidhof, et al., 2014; Bennett, et al., 2018). In *Anas platyrhynchos*, 56 wild and 32 domestic specimens were measured. Of the 32 domestic mallards, seven were from known breeds (four Aylesbury, two Black Labrador or Buenos Aires Drake and one Khaki Campbell). Given the low number of labelled breeds, we only made the distinction between wild and domestic mallards. In *Cairina moschata*, 39 wild and 17 domestic specimens were measured.

The specimens used for this study are housed in the Paläontologisches Institut und Museum, Universität Zürich (Switzerland), Naturmuseum Senckenberg (Germany), Staatssammlung für Anthropologie und Paläoanatomie München (Germany), Zoologisches Institut/Populationsgenetik (former Institut für Haustierkunde), Christian-Albrechts-Universität zu Kiel (Germany), Natural History Museum at Tring, Bird Collection, General and Darwin collection (United Kingdom).

**Analyses.** We performed independent two-tailed student's t-tests to check for differences in the limb bones proportions in their respective limb between wild and domestic forms for all species. Further, we performed independent two-tailed student's t-tests to check for differences in limb length, as a proxy for body size, between domestic and wild forms of the three species in this study (Fig.3). In the case of *Gallus gallus*, we also tested if there are significant differences in

the proportions among breeds (*Table 3*). For this, we used only domestic breeds with sample size greater than four, namely Araucana, bantam, Chabo, cochin, Ukokkei, ‘Italiener’, Kömpfer, Shamo and Polish as well as the RJF. Due to violation of assumptions of normality and homoscedasticity (*Table S1* and *Table S2* in supplementary materials), we performed non-parametric Kruskal-Wallis tests.

In order to determine the explanatory power of body size on long bone proportions in *Gallus gallus*, *Anas platyrhynchos*, and *Cairina moschata*, we performed linear regressions using hind limb and forelimb total length as a proxy of body size, respectively. We contrasted the log-transformed values of each bone with the log-transformed values of the total length of the correspondent limb. Total limb length is the best approximation available, since the age of the used specimen might be variable, even though all were skeletally mature, and using the mean breed weight or size could be more problematic. To determine if the regression slopes were equal to 1 (null hypothesis of isometry), we determined if the 95% confidence intervals of the slopes included 1.0. We assumed positive or negative allometry to best explain a given scaling relationship if the confidence interval of the regression did not include 1, with a slope of  $> 1.0$  or  $< 1.0$ , respectively. Similar as for the adult sample described here, we extracted data about limb bone dimensions of chicken embryos and juveniles from the literature and determined allometric growth patterns as described for the adult sample. In order to illustrate the distribution of proportions of the hind limb bones in the morphospace of *Gallus gallus* during pre-hatching development, data were taken from Thomas, (2014) and Faux and Field (2017) and we performed the same analyses as for the regression slopes of the three species previously mentioned. The results are displayed in *Fig. 4* and *Table 4*.

In the following summary of feature variations, we account for five characters of the sternum of *Gallus gallus* across nine different domestic breeds and the RJF as their wild counterpart. Sternum characters were coded in 63 *Gallus gallus* specimens from which 12 were RJF and the rest belonged to domestic breeds.

#### **Description of character states.**

- Carina sterni is defined as ‘straight’ when the direction of growth is maintained in a straight line, as opposed to ‘crooked’, in which the growth is tortuous with one or multiple pronounced torsions (*Fig. 5a*).

- Carina sterni is defined as ‘notched’ when the direction of the lower margin of the carina in lateral view changes direction, creating a notch. It is defined as ‘non notched’ if this does not happen (*Fig.5b*).
- Pars cardiaca (PC) and Foramen pneumaticum (FP) are described as ‘spongy’ or ‘smooth’ depending on the type of external appearance, either porous or flat respectively (*Fig.5c*).
- The caudal end of the trabecula mediana of the sternum is differentiated between ‘fanned’ (end wider than the rest of the trabecula in dorsal view) and ‘straight’ (end as wide as the rest of the trabecula in dorsal view; *Fig.5d*).

## Results

The two-tailed, independent t-test for the proportion of long bones of the forelimbs and hind limbs performed comparing wild versus domestic in three species: *Gallus gallus*, *Anas platyrhynchos* and *Cairina moschata* show that significant differences in limb proportions only occur in *Anas platyrhynchos* (*Table2*). Domestic mallard ducks, compared to their wild counterpart, have proportionally longer humeri and shorter radii and carpometacarpi in the case of the wing, and proportionally longer tarsometatarsi and shorter femurs in the case of the hind limb (*Fig.6*).

The Kruskal-Wallis test comparing limb proportions among chicken breeds revealed significant differences in the selected breeds (*Table 3*). The detailed pairwise comparison is shown in *Fig.7* and the supplementary materials for breeds with sample size equal or greater than five (*Table S3*).

A brief comparison of the total forelimb length, showing the substantial size variation, in different breeds of *Gallus gallus* is provided in *Table 5*. The lengths of the long bones are strongly and significantly correlated with the total size of the corresponding limb as a proxy for body size in chicken, mallard ducks, and Muscovy ducks (*Fig.8, Table 6*). Pairwise comparisons of total limb length between wild and domestic forms in each species, as a proxy for body size, show that the domestic varieties are larger than the wild ones in *Gallus gallus* and *Anas platyrhynchos* but no significant differences were observed in *Cairina moschata* (*Fig.3*). Our observations on the characters of the sternum are summed up in *Table 7*.

# Discussion

Given the previous studies on differences between wild and domestic animals and the importance in locomotion of forelimb and hind limb proportions and sternum characteristics, we aimed to determine whether differences exist between wild and domestic forms of three bird species and to shed light on the possible underlying mechanisms.

We observed no evidence of significant differences between wild and domestic forms of *Gallus gallus* in the proportional length of the long bones of the forelimbs (humerus, radius and carpometacarpus) and hind limbs (femur, tibiotarsus and tarsometatarsus; *Table 1*). However, when the RJF and breeds of domestic chicken were analysed as individual groups, significant differences were detected (*Fig.7, Table S3*). One possible interpretation is for diversification to have taken multiple directions in the case of the limb proportions in domestic birds, but selection has not provoked a unique directionality, maintaining the same mean proportions between domestic and wild forms in all bones. The most notable case of breed-specific specialisation is the Chabo, also known as Japanese bantam. Chabo is a true bantam breed (i.e. it does not present a large variant) with characteristically short legs (Roberts, 2008). The short leg trait is associated with a dominant allele of gene *Cp*, which is lethal in homozygosis (Shibuya, et al., 1972; Shibuya & Kuroda, 1973) and is selected as a distinct characteristic of the breed (Van Grouw, 2018). The *Cp* gene accelerates differentiation of chondrocytes accompanied by suppression of their proliferation, leading to stunted limb long bone growth (Shibuya, et al., 1972). Similar phenotypes are also known in domestic dogs (e.g. Dachshund, Corgi) and cats, and represent cases of specialisation in breed formation (Rimbault & Ostrander, 2012).

Our results further show that proportions of the bones in *Anas platyrhynchos* vary significantly when comparing wild and domestic forms (*Table 1; Fig.6*). Unlike wild and domestic *Gallus gallus*, domestic forms of the mallard duck thus show a set of proportions in the long bones that is significantly different from the wild form. Since body size differences were marked between the wild and domestic forms of the three investigated species, with the domestic forms being on average larger than their wild relatives in *Gallus gallus* and *Anas platyrhynchos* (*Fig.3*), we compared the log-transformed length of the long bones with the log-transformed total length of the limb as a proxy for body size to investigate the influence of allometric scaling on limb size proportions (*Fig.8, Table 6*). Limb proportions were found to scale with size in an inconsistent way in the different species (*Table 6*). The observed difference of limb proportions in wild and

domestic mallards (*Table 1; Fig. 6*) could thus be interpreted as the result of allometric scaling due to particularly pronounced size differences in the wild vs. the domestic sample (*Fig. 3*). Specifically, in the case of the mallard's forelimb, the humerus scales positively allometrically, the radius isometrically, and the carpometacarpus negatively allometrically, leading to proportionally longer humeri and shorter radii in larger specimens (*Fig. 2, Fig. 8, Table 6*). Although no ontogenetic study is available for mallard ducks so far as a comparison, this same tendency for allometric scaling of the limb bones can be extracted from the data available in the literature regarding embryological growth of *Gallus gallus* (Faux & Field, 2017) and ontogenetic development after hatching (Thomas, et al., 2016; Jackson & Diamond, 1996) (*Fig. 4A and Fig. 4B*). As the body size of the embryo and hatched birds increases, their limb proportions change in a similar tendency (except tibiotarsus and tarsometatarsus in post-hatching growth, *Fig. 4, Table 4*) as we observe in differently sized adult chickens (*Fig. 8a,b, Table 6*). This can also be observed for wing growth in *Larus californicus*, which shows a progressive decrease in the humerus length relative to the radius and carpometacarpus (Middleton & Gatesy, 2000). This suggests that the differences in proportions of the limbs among adult chickens might be at least partially related to allometric scaling during development and growth. Differences in limb proportions among breeds could thus be related to different sizes of breeds. As differences in limb proportions between the wild and the domestic groups are not apparent in chicken and Muscovy ducks (*Fig. 6, Table 1*), although allometric scaling of limb bones was observed (*Fig. 8, Table 6*), a lack of pronounced size differences (*Fig. 3*) between wild and domestic could explain the lack of proportional differences. The literature shows implications of different allometries in limbs. An apparent inverse correlation of the proportion of the humerus in the wing with aerial maneuverability in birds has been proposed in the past (Middleton & Gatesy, 2000). Moreover, flightlessness is reportedly linked to a proportionally long humerus compared to the rest of the long bones of the wing in theropods (Middleton & Gatesy, 2000) and steamer ducks (*Tachyeres*) (Livezey & Humphrey, 1986). Experiments on the domestication of the wild mallard resulted in bigger body sizes and rapid loss (after three generations) of the ability to fly (Darwin, 1868). Mason (1984) proposed that this loss of flight capacity might be due to the loss of positive selection for flying ability: under the care of humans, mallards are usually granted access to a reliable source of food at a specific

location, thus flight becomes less essential for survival. An alternative explanation could be that the selection of the species for specific traits could have pleiotropic effects that lead to a reduction of flight capacity. These hypotheses remain to be tested in future work. The loss of the capacity to fly may be related to the proportion of the limbs, which upon less selection due to lack of necessity to fly may have become significantly different from their wild counterpart (*Table S2*). Other skeletal, muscular and perhaps neurological changes may also be related to the loss of flight capacity. In the case of the hind limb, we observed in domestic forms of the mallard duck a relative shortening of the femur and elongation of the tarsometatarsus. Gatesy and Middleton (1997) reported an apparent gradient of cursoriality increasing with tarsometatarsal proportion among theropod species with similar Tibiotarsus/Femur ratios. The pattern we observe in the domestic mallard duck, together with a decrease in flight adaptations (above), is consistent with a more terrestrial, although not necessarily highly cursorial, lifestyle of the domesticated vs. the wild form.

Changes in limb proportions are not observed in *Cairina moschata*. Its proportions remain indistinguishable from the wild form, as opposed to *Anas platyrhynchos* (*Table 1*). Changes in flight habits might not have been similar between the wild and domestic forms of mallard ducks and Muscovy ducks. While some populations of wild mallard ducks are migratory (Bellerose & Crompton, 1970), domestic mallards rarely fly as much and many breeds lack flight capacity entirely (Smith, et al., 1989). Muscovy ducks, on the other hand, although non-migratory birds (Accordi & Barcellos, 2006), have been reported to fly frequently (Anon., 2016; Anon., 2017). This suggesting that their flight habits might not have changed so markedly as to produce a different morphology. Alternatively, and related to the allometric scaling relationships considered above, it is noteworthy that in contrast to the other two species, limb bones of the forelimb were found to consistently scale isometrically in the Muscovy duck, suggesting restricted evolvability concerning forelimb proportions in this species. An analogous conclusion was reached when postulating lower evolvability in cats versus dogs when reporting largely isometric versus allometric in the skulls, respectively (Sánchez-Villagra, et al., 2017).

The variations on the sternum of *Gallus gallus* (*Table 7*) show that the RJF generally possesses a notched carina sterni, a smooth surface of the pars cardiaca and foramen pneumaticum and a fanned caudal end of the trabecula mediana. The RJF develops a crooked keel in half of our observed specimens (*Table 7*). The crookedness of the keel has been shown to be hereditary and

influenced by early roosting conditions (Blount, 1933; Warren, 1937; Waters, 1949). Our observations reveal that this is not a feature unique to domestic varieties, as has been previously reported (Kittelsen, et al., 2020).

Most domestic variants exhibit straight carina sterni, with the notable exception of the Chabo and Ukokkei breeds. One possible explanation might be that the shortness of the legs in the Chabo and the inability to fly of the Ukokkei are responsible for a more direct contact with higher pressure on the keel with the perching structure of their environments, as it has been shown to be a determinant factor for its appearance (Blount, 1933; Warren, 1937; Waters, 1949; Pickel, et al., 2011; Casey-Trott, et al., 2015). Behaviour, especially flying and perching, could be an important factor distinguishing breeds but could not be assessed in this study.

The notch in the carina sterni seems to be a rare condition in domestic breeds, while it is the most common condition in the wild form, contrary to the other features observed with a more general distribution. The sponginess of the pars cardiaca and foramen pneumaticum is a rare condition in most breeds except for the case of the Araucana breed. Most breeds show a fanned end of the sternum as is the case of the wild form. In summary, these results reveal that the crookedness of the keel is not unique to domestic forms and, while the prevalence of the notch in the carina sterni can be distinguishing between the wild and domestic forms, the variation of the different characters are not segregated among the breeds.

## Conclusions

This study shows how the allometric growth of *Gallus gallus* can help us understand the variation in limb proportions of differently sized breeds. Furthermore, we show that significant differences in the proportions of limb bones of wild and domestic *Anas platyrhynchos* exist, whereas no significant differences between wild and domestic forms of *Gallus gallus* and *Cairina moschata* were found. We offer possible explanations for these differences such as an extension of the allometric growth of domestic Mallard ducks, loss of positive selection for flying ability, and/or pleiotropic effects or evolvability constraints, which could inspire future research. Furthermore, we observed that the condition of crooked keel is not unique to domestic forms of *Gallus gallus* and a higher number of cases of notched carina sterni can be found in RJF. These findings add new observations to the study of keel characteristics that can be added to the body of knowledge of domestication and poultry welfare.

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# **Table 1**(on next page)

Results of two-tailed, independent t-test for the lengths of long bones of the forelimbs and hind limbs of domestic and wild forms of *Gallus gallus*, *Anas platyrhynchos* and *Cairina moschata*.

\*: the number of degrees of freedom is approximated by the Welch-Satterthwaite formula.

**Bold**: null hypothesis of equality of means rejected. Df = degrees of freedom.

Species	Bone	Df	T	p-value
<i>Gallus gallus</i>	Humerus	70.893*	-0.746	0.4584
	Radius	140	1.674	0.0963
	Carpometacarpus	140	-1.603	0.1112
	Femur	133	1.477	0.1420
	Tibiotarsus	135	-1.111	0.2685
	Tarsometatarsus	135	-0.982	0.3279
<i>Anas platyrhynchos</i>	Humerus	86	5.212	<b>&lt; 0.0001</b>
	Radius	45.049*	-2.520	<b>0.0154</b>
	Carpometacarpus	43.519*	-3.166	<b>0.0028</b>
	Femur	86	-1.989	<b>0.0498</b>
	Tibiotarsus	45.171*	-1.001	0.3222
	Tarsometatarsus	86	3.042	<b>0.0031</b>
<i>Cairina mostacha</i>	Humerus	54	-0.199	0.8430
	Radius	54	1.390	0.1701
	Carpometacarpus	54	-0.899	0.3728
	Femur	54	0.730	0.4684
	Tibiotarsus	54	-0.408	0.6849
	Tarsometatarsus	54	-0.271	0.7873

# **Table 2**(on next page)

Breeds of chicken (*Gallus gallus*) and number of specimens measured in this study.

Breed	N <sub>Forelimb</sub>	N <sub>Hind limb</sub>	Breed	N <sub>Forelimb</sub>	N <sub>Hind limb</sub>
RJF	29	28	Pennsylvania Naked Neck	2	<u>2</u>
‘African cock’	1	<u>1</u>	Rhode Island	1	<u>0</u>
Appenzeller Barthuhn	3	<u>3</u>	Rumpless fowl	1	<u>1</u>
Appenzeller Spitzhaubenhuhn	5	<u>5</u>	Seidenhuhn	5	<u>5</u>
Araucana	13	13	Shamo	8	8
Bantam	7	8	Silver pencil hamburghen	1	0
Brahmahuhn	1	1	Polish	5	5
Burmese Bantam	1	1	Spanish cock	2	2
Chabo	16	16	Sultan	2	1
Cochin	6	<u>6</u>	Tail-Less Bantam	1	<u>1</u>
Dorking cock	1	<u>1</u>	Ukokkei	10	<u>10</u>
Game	1	<u>1</u>	Welsumer	3	<u>2</u>
Gold spangle Polish	1	<u>1</u>	White crested rumpless Turkish fowl	1	<u>1</u>
Golden pencil hamburgh	1	<u>1</u>	White dorkinghen	1	<u>1</u>
Italiener	2	<u>2</u>	White Leghorn	4	<u>4</u>
Kulm	1	<u>1</u>	Zwergwelsumer	2	<u>2</u>
Light sussex	3	<u>2</u>			

# **Table 3**(on next page)

Results of Kruskal-Wallis tests for comparison among breeds of *Gallus gallus*.

Kruskal-Wallis rank sum test				
Limb	Bone	Chi-squared	df	p-value
Forelimb	Humerus	47	7	6.84E-08
	Radius	37	7	5.58E-06
	Carpometacarpus	16	7	0.02805
Hind limb	Femur	37	7	4.64E-06
	Tibiotarsus	23	7	0.001848
	Tarsometatarsus	33	7	2.63E-05

1

# **Table 4**(on next page)

Results of linear regressions of long bone lengths vs. total limb length of embryological development (data taken from Faux and Field, 2017) and ontogeny (data taken from Thomas, 2014).

$a$  = regression slope with corresponding 95% confidence interval (ci); scaling pattern = interpretation of isometry (ci including 1,  $a=1$ ), negative allometry (ci not including 1,  $a<1$ ), or positive allometry (ci not including 1,  $a>1$ );  $r^2$  = coefficient of determination;  $F$  =  $F$  statistic with associated degrees of freedom (df);  $p$  =  $p$ -value.

Study	Bone	a (ci)	Scaling pattern	r <sup>2</sup>	F (df)	p
<u>Faux and Field (2017)</u>	<u>Femur</u>	<u>0.963</u> ( <u>0.928</u> , <u>0.998</u> )	<u>Negative allometry</u>	<u>0.96</u> <u>28</u>	<u>3.242*10<sup>3</sup></u> ( <u>1, 23</u> )	<u>&lt;2.2*</u> <u>10<sup>-16</sup></u>
	<u>Tibiotarsus</u>	<u>1.005</u> ( <u>0.958</u> , <u>1.051</u> )	<u>Isometry</u>	<u>0.98</u> <u>86</u>	<u>1.993*10<sup>3</sup></u> ( <u>1, 23</u> )	<u>&lt;2.2*</u> <u>10<sup>-16</sup></u>
	<u>Tarsometatarsus</u>	<u>1.033</u> ( <u>1.001</u> , <u>1.065</u> )	<u>Positive allometry</u>	<u>0.99</u> <u>48</u>	<u>4.403*10<sup>3</sup></u> ( <u>1, 23</u> )	<u>&lt;2.2*</u> <u>10<sup>-16</sup></u>
<u>Thomas (2014)</u>	<u>Femur</u>	<u>0.930</u> ( <u>0.909</u> , <u>0.952</u> )	<u>Negative allometry</u>	<u>0.99</u> <u>43</u>	<u>7.549*10<sup>3</sup></u> ( <u>1, 43</u> )	<u>&lt;2.2*</u> <u>10<sup>-16</sup></u>
	<u>Tibiotarsus</u>	<u>1.040</u> ( <u>1.027</u> , <u>1.053</u> )	<u>Positive allometry</u>	<u>0.99</u> <u>83</u>	<u>2.5*10<sup>4</sup></u> ( <u>1,43</u> )	<u>&lt;2.2*</u> <u>10<sup>-16</sup></u>
	<u>Tarsometatarsus</u>	<u>1.016</u> ( <u>0.994</u> , <u>1.038</u> )	<u>Isometry</u>	<u>0.99</u> <u>51</u>	<u>8.788*10<sup>3</sup></u> ( <u>1,43</u> )	<u>&lt;2.2*</u> <u>10<sup>-16</sup></u>

**Table 5**(on next page)

Mean length of the forelimb and hind limb of the different chicken breeds in mm.

Measurements belonging to RJF are shown in bold.

	Bantam	Chabo	Red junglefowl	Ukokkei	Polish	White Leghorn	Appenzeller Spitzhaubenhuhn	Araucana	Shamo
Forelimb length (mm)	115,47	123,5719	158,8323	175,508	179,2575	192,5175	197,032	202,0969	231,7588
Hind limb length (mm)	175,915	172,9388	241,4441	263,119	269,7775	297,655	307,306	312,8046	365,585

1

# Table 6 (on next page)

Results of linear regressions of long bone lengths vs. total limb length in *Gallus gallus*, *Anas platyrhynchos* and *Cairina moschata*.

a = regression slope with corresponding 95% confidence interval (ci); scaling pattern = interpretation of isometry (ci including 1,  $a=1$ ), negative allometry (ci not including 1,  $a<1$ ), or positive allometry (ci not including 1,  $a>1$ );  $r^2$  = coefficient of determination; F = F statistic with associated degrees of freedom (df); p = p-value.

Species	Bone	a (ci)	Scaling pattern	r <sup>2</sup>	F (df)	p
<i>Gallus gallus</i>	Humerus	0.937 (0.926, 0.948)	Negative allometry	0.9954	3.002*10 <sup>4</sup> (1, 140)	<2.2*10 <sup>-16</sup>
	Radius	1.069 (1.054, 1.085)	Positive allometry	0.9923	1.809*10 <sup>4</sup> (1, 140)	<2.2*10 <sup>-16</sup>
	Carpometacarpus	1.004 (0.981, 1.027)	Isometry	0.9813	7.342*10 <sup>3</sup> (1, 140)	<2.2*10 <sup>-16</sup>
	Femur	0.896 (0.881, 0.911)	Negative allometry	0.9903	1.38*10 <sup>4</sup> (1, 135)	<2.2*10 <sup>-16</sup>
	Tibiotarsus	1.005 (0.998, 1.013)	Isometry	0.998	6.822*10 <sup>4</sup> (1, 135)	<2.2*10 <sup>-16</sup>
	Tarsometatarsus	1.105 (1.088, 1.122)	Positive allometry	0.9918	1.636*10 <sup>4</sup> (1, 135)	<2.2*10 <sup>-16</sup>
<i>Anas platyrhynchos</i>	Humerus	1.038 (1.016, 1.061)	Positive allometry	0.9898	8.336*10 <sup>3</sup> (1, 86)	<2.2*10 <sup>-16</sup>
	Radius	0.985 (0.965, 1.006)	Isometry	0.9907	9.137*10 <sup>3</sup> (1, 86)	<2.2*10 <sup>-16</sup>
	Carpometacarpus	0.956 (0.926, 0.986)	Negative allometry	0.9791	9.903*10 <sup>3</sup> (1, 86)	<2.2*10 <sup>-16</sup>
	Femur	0.977 (0.941, 0.996)	Negative allometry	0.9844	5.423*10 <sup>3</sup> (1, 86)	<2.2*10 <sup>-16</sup>
	Tibiotarsus	0.99 (0.973, 1.008)	Isometry	0.9933	1.268*10 <sup>4</sup> (1, 86)	<2.2*10 <sup>-16</sup>
	Tarsometatarsus	1.054 (1.023, 1.085)	Positive allometry	0.9819	4.677*10 <sup>3</sup> (1, 86)	<2.2*10 <sup>-16</sup>
<i>Cairina moschata</i>	Humerus	0.992 (0.971, 1.013)	Isometry	0.9941	9.336*10 <sup>3</sup> (1, 54)	<2.2*10 <sup>-16</sup>
	Radius	0.987 (0.954, 1.021)	Isometry	0.9849	3.513*10 <sup>3</sup> (1, 54)	<2.2*10 <sup>-16</sup>
	Carpometacarpus	1.026 (0.968, 1.083)	Isometry	0.9595	1.28*10 <sup>3</sup> (1, 54)	<2.2*10 <sup>-16</sup>
	Femur	0.988 (0.954, 1.022)	Isometry	0.9846	3.455*10 <sup>3</sup> (1, 54)	<2.2*10 <sup>-16</sup>
	Tibiotarsus	0.97 (0.95, 0.99)	Negative allometry	0.9944	9.641*10 <sup>3</sup> (1, 54)	<2.2*10 <sup>-16</sup>
	Tarsometatarsus	1.068 (1.029, 1.108)	Positive allometry	0.9819	2.933*10 <sup>3</sup> (1, 54)	<2.2*10 <sup>-16</sup>

# **Table 7** (on next page)

Table of characters and character states by breed.

Characters and character states: carina sterni (straight or crooked), carina sterni (notched or non-notched), pars cardiaca (PC) and foramen pneumaticum (FP) (spongy or smooth), trabecula mediana: (fanned or straight). Numbers in the table indicate the number of specimens per character state.

1

	<u>Carina sterni</u>		<u>Carina sterni</u>		<u>PC and FP</u>		<u>Trabecula mediana</u>	
<u>Breed</u>	<u>Crooked</u>	<u>Straight</u>	<u>Notched</u>	<u>Non-notched</u>	<u>Spongy</u>	<u>Smooth</u>	<u>Fanned</u>	<u>Straight</u>
<u>Araucana</u>	<u>1</u>	<u>6</u>	<u>1</u>	<u>6</u>	<u>4</u>	<u>3</u>	<u>3</u>	<u>2</u>
<u>Bantam</u>	<u>0</u>	<u>7</u>	<u>2</u>	<u>5</u>	<u>2</u>	<u>5</u>	<u>7</u>	<u>0</u>
<u>Chabo</u>	<u>8</u>	<u>8</u>	<u>3</u>	<u>13</u>	<u>6</u>	<u>10</u>	<u>11</u>	<u>5</u>
<u>Cochin</u>	<u>1</u>	<u>3</u>	<u>0</u>	<u>4</u>	<u>1</u>	<u>3</u>	<u>1</u>	<u>3</u>
<u>Ukokkei</u>	<u>3</u>	<u>0</u>	<u>1</u>	<u>2</u>	<u>1</u>	<u>2</u>	<u>2</u>	<u>1</u>
<u>Italiener</u>	<u>2</u>	<u>1</u>	<u>0</u>	<u>3</u>	<u>1</u>	<u>2</u>	<u>1</u>	<u>2</u>
<u>Kömpfer</u>	<u>0</u>	<u>1</u>	<u>0</u>	<u>1</u>	<u>0</u>	<u>1</u>	<u>1</u>	<u>0</u>
<u>RJF</u>	<u>6</u>	<u>6</u>	<u>9</u>	<u>3</u>	<u>0</u>	<u>12</u>	<u>10</u>	<u>2</u>
<u>Shamo</u>	<u>2</u>	<u>4</u>	<u>2</u>	<u>4</u>	<u>2</u>	<u>4</u>	<u>2</u>	<u>3</u>
<u>Polish</u>	<u>3</u>	<u>1</u>	<u>1</u>	<u>3</u>	<u>3</u>	<u>1</u>	<u>2</u>	<u>2</u>
<u>TOTAL</u>	<u>26</u>	<u>37</u>	<u>19</u>	<u>44</u>	<u>20</u>	<u>43</u>	<u>40</u>	<u>20</u>

# Figure 1

Anatomical structures used to define measurements parallel to the longitudinal axis of the limb bones (indicated with red dots).

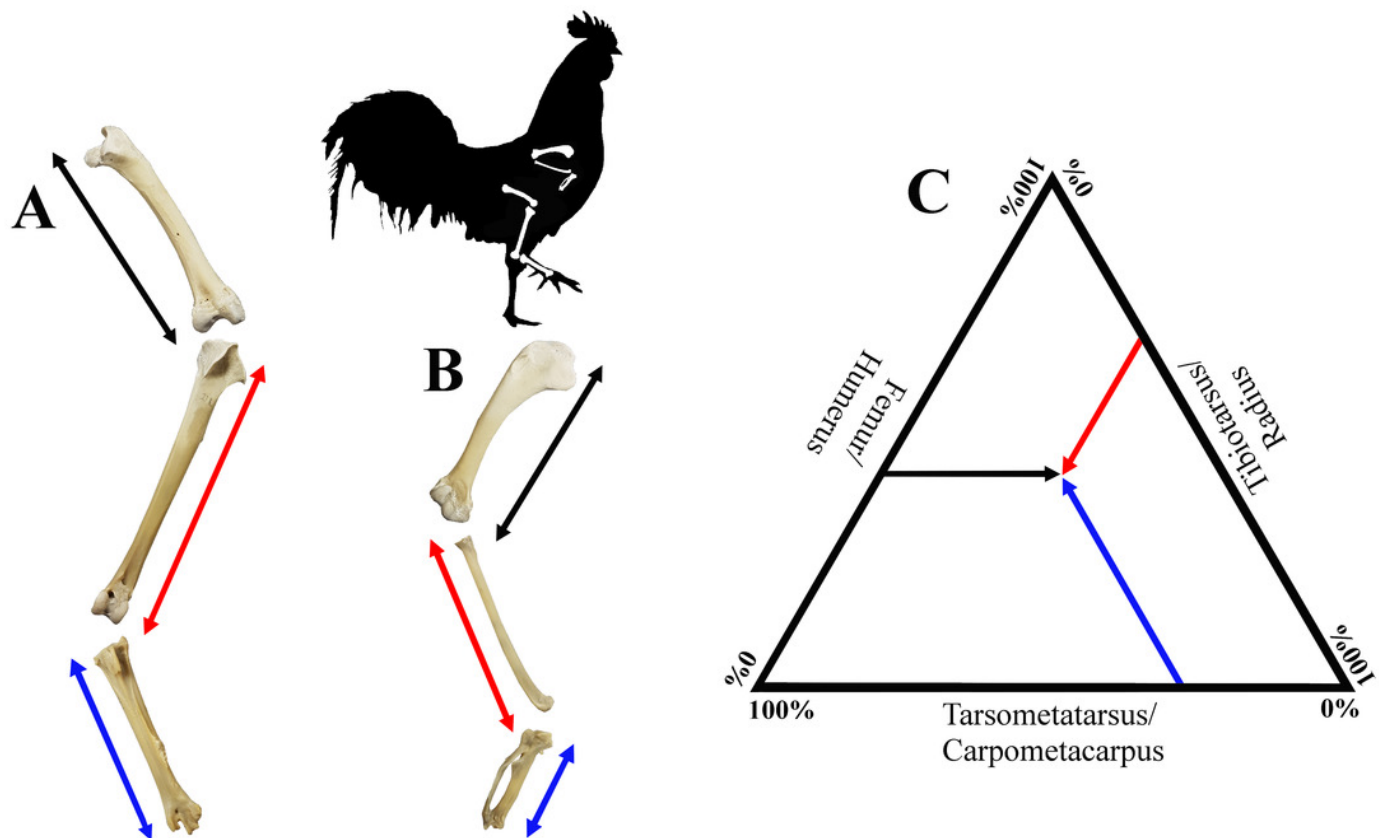
Humerus (A): caput humeri (B) and cotylus ventralis (C); Radius (D): cotyla humeralis (E) and facies articularis radiocarpalis (F); Carpometacarpus (G): trochlea carpalis (H) and facies articularis digitalis minor (I); Femur (J): trochanter femoris (K) and condylus lateralis (L); Tibiotarsus (M): crista cnemialis cranialis (N) and epicondylus lateralis (O); Tarsometatarsus (P): crista medialis hypotarsi (Q) and trochlea metatarsi III (R) Shown bones belonging to domestic chicken (*Gallus gallus*, Araucana)



# Figure 2

Forelimb and hind limb proportions of the three bones in each limb represented graphically in a ternary plot.

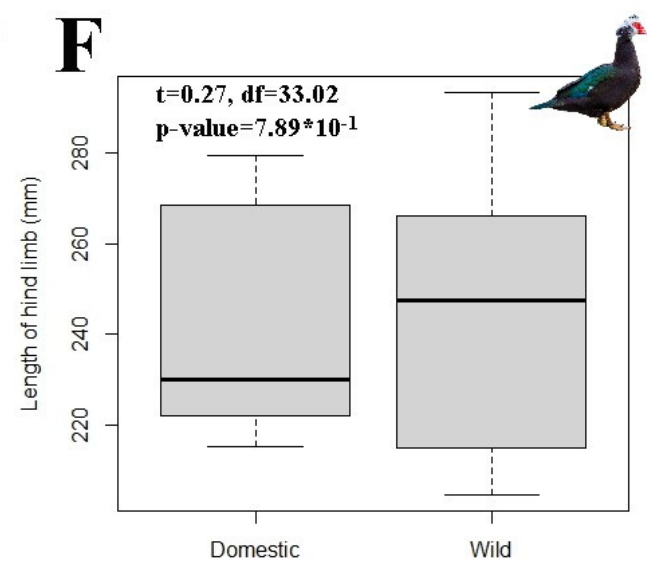
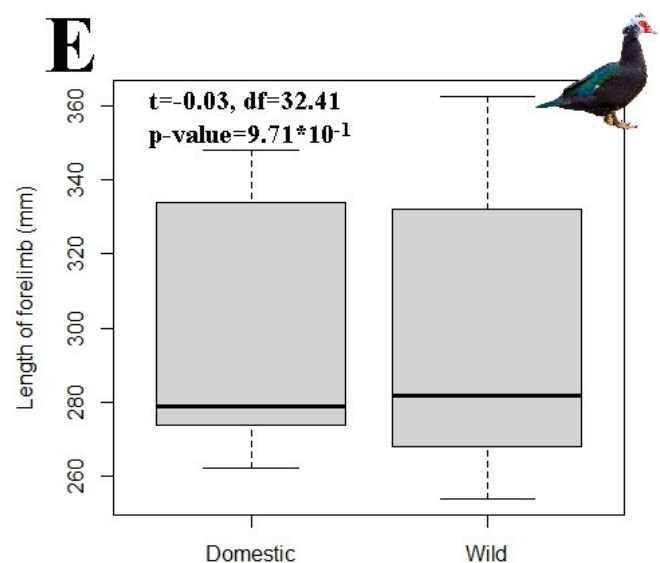
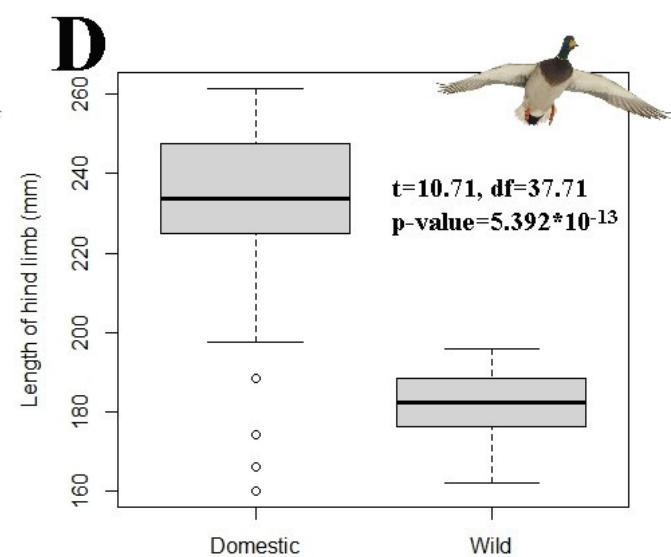
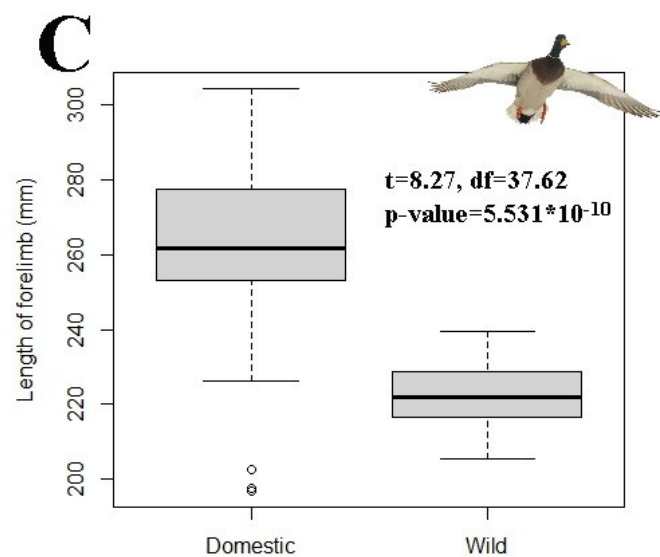
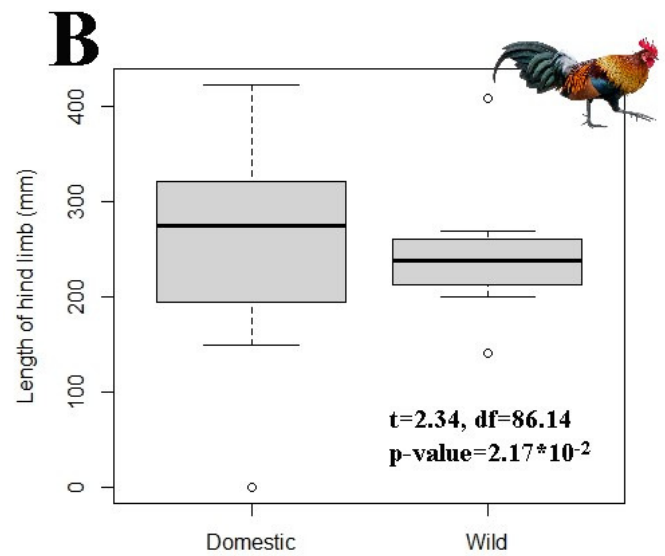
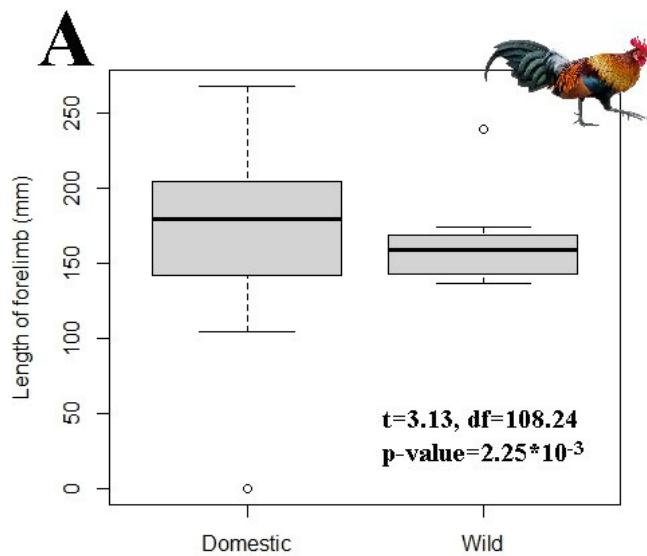
For the hind limb (A), the considered bones are femur (black), tibiotarsus (red) and tarsometatarsus (blue); for the forelimb (B), the considered bones are humerus (black), radius (red) and carpometacarpus (blue); a ternary plot is used to show the proportions for each limb (C).



# Figure 3

Boxplots comparing limb length between wild and domestic forms.

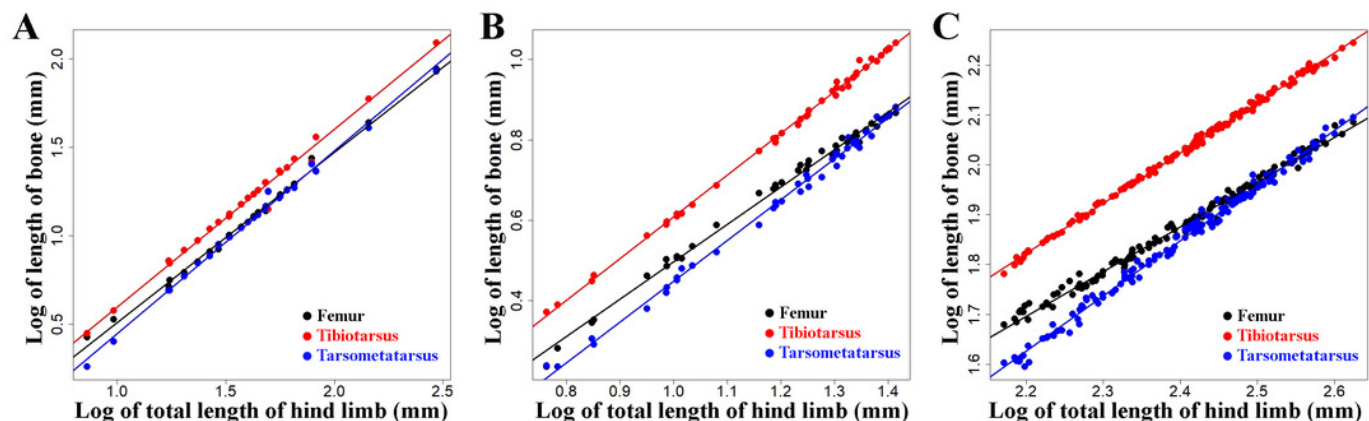
(A and B) For chicken; (C and D) for Mallard ducks and (E and F) for Muscovy ducks. T test results are included and show t statistic (t) with associated degrees of freedom (df); p = p-value.



# Figure 4

Bivariate plots reflecting the allometric scaling in hind limb of embryological and adult *Gallus gallus*.

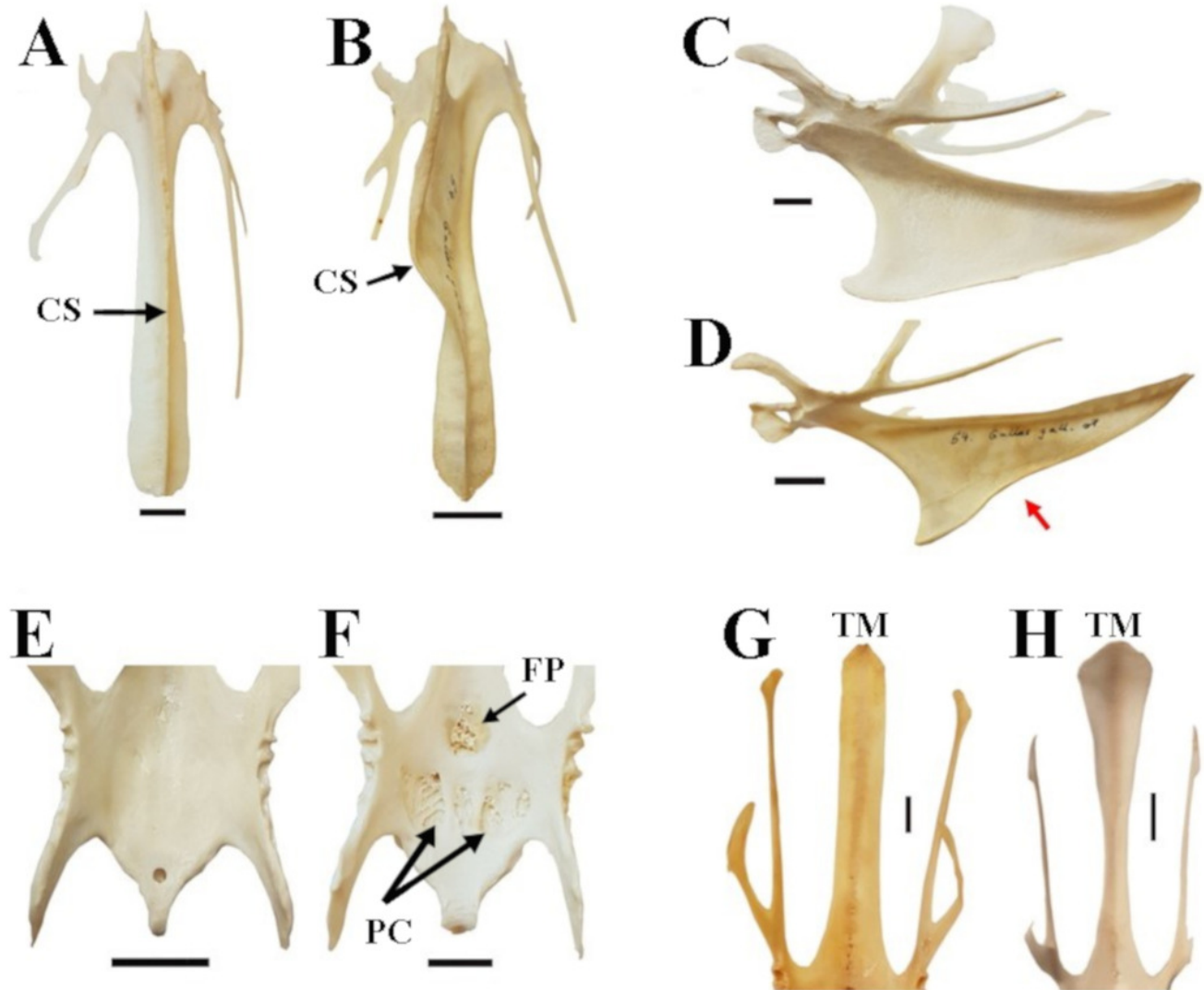
(A) Measurements taken from embryos at day 7 of incubation to day 17 (data taken from Faux and Field, 2017). (B) Measurements taken from juveniles from 1 week after hatching to week 36 (data taken from Thomas, 2014). (C) Variation of proportions of adults of chicken breeds and RJF sampled in this study.



# Figure 5

Characters and character states of the sternum in *Gallus gallus*.

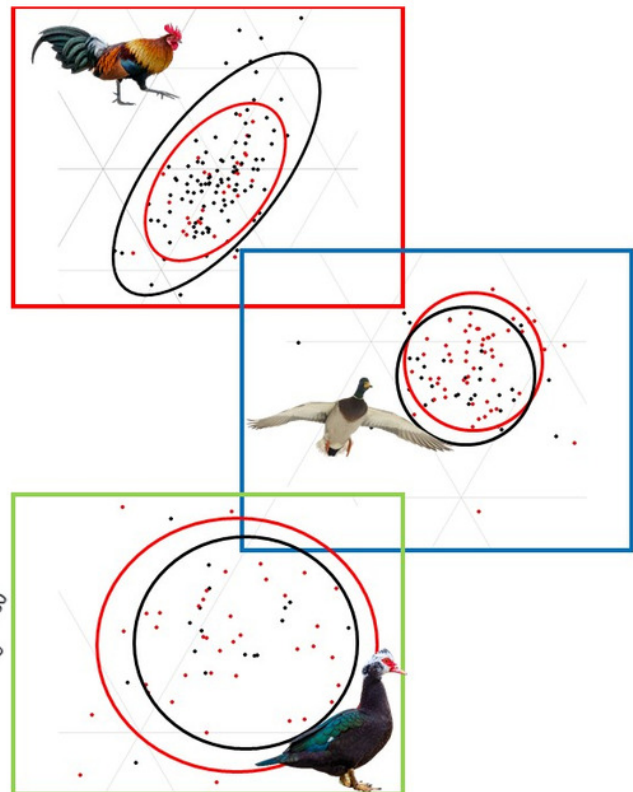
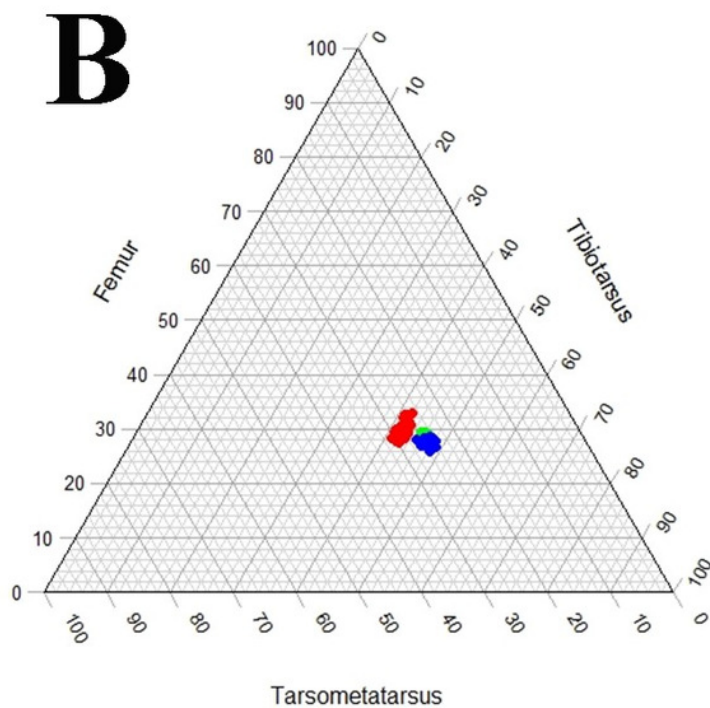
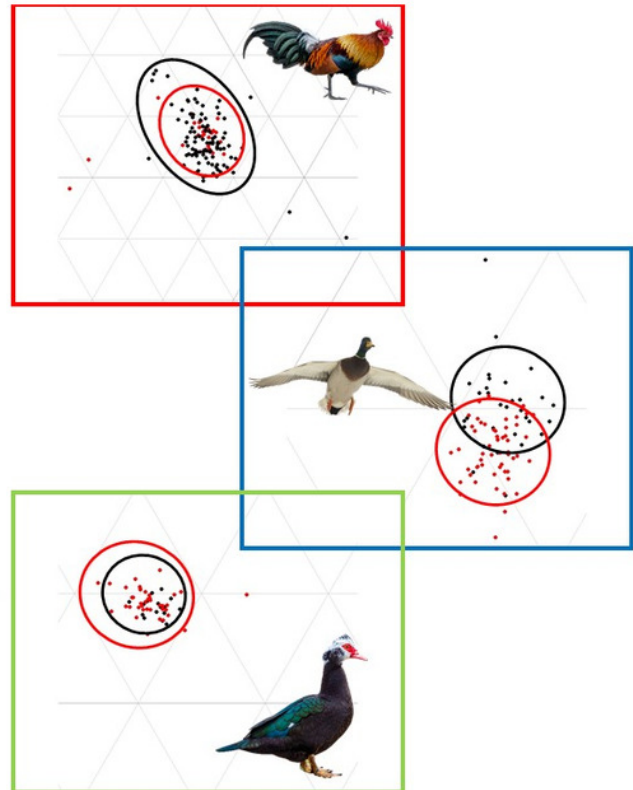
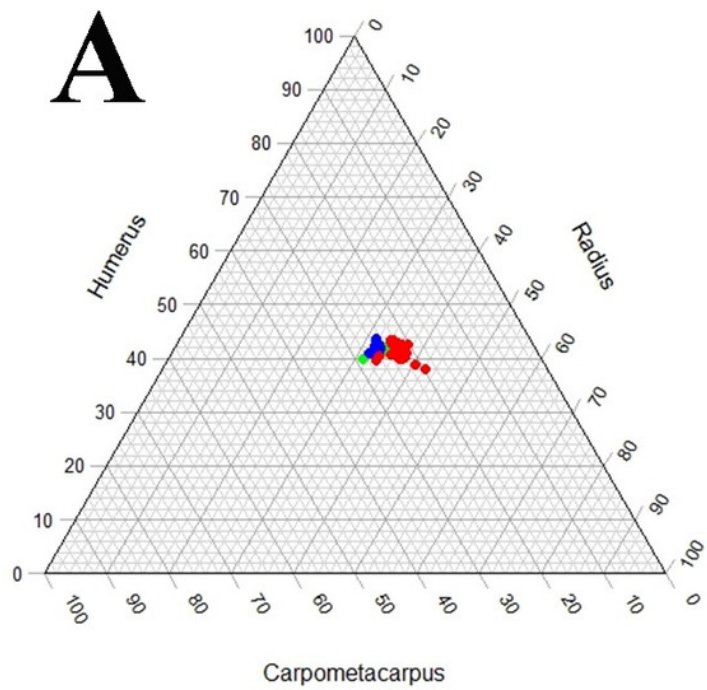
(A) Straight carina sterni (CS) (Araucana SAPM-AV-05259), (B) crooked carina sterni (RJF SAPM-AV-05218), (C) non-notched carina sterni (Araucana SAPM-AV-05274), (D) notched carina sterni (RJF SAPM-AV-05218) where notch is indicated by the red arrow, (E) smooth surface and poorly defined concavities of PC and FP (RJF SAPM-AV-05234), (F) spongy well-defined concavities of PC and FP (Araucana SAPM-AV-05259)), (G) Straight end of the trabecula mediana (TM) in dorsal view (Araucana SAPM-AV-05264), (H) fanned end of the trabecula mediana (TM) in dorsal view (RJF SAPM-AV-05266). Specimens are form the Staatssammlung für Anthropologie und Paläoanatomie München. Scale = 1cm.



# Figure 6

Ternary plots illustrating the proportions in *Gallus gallus* (red), *Anas platyrhynchos* (blue) and *Cairina moschata* (green)

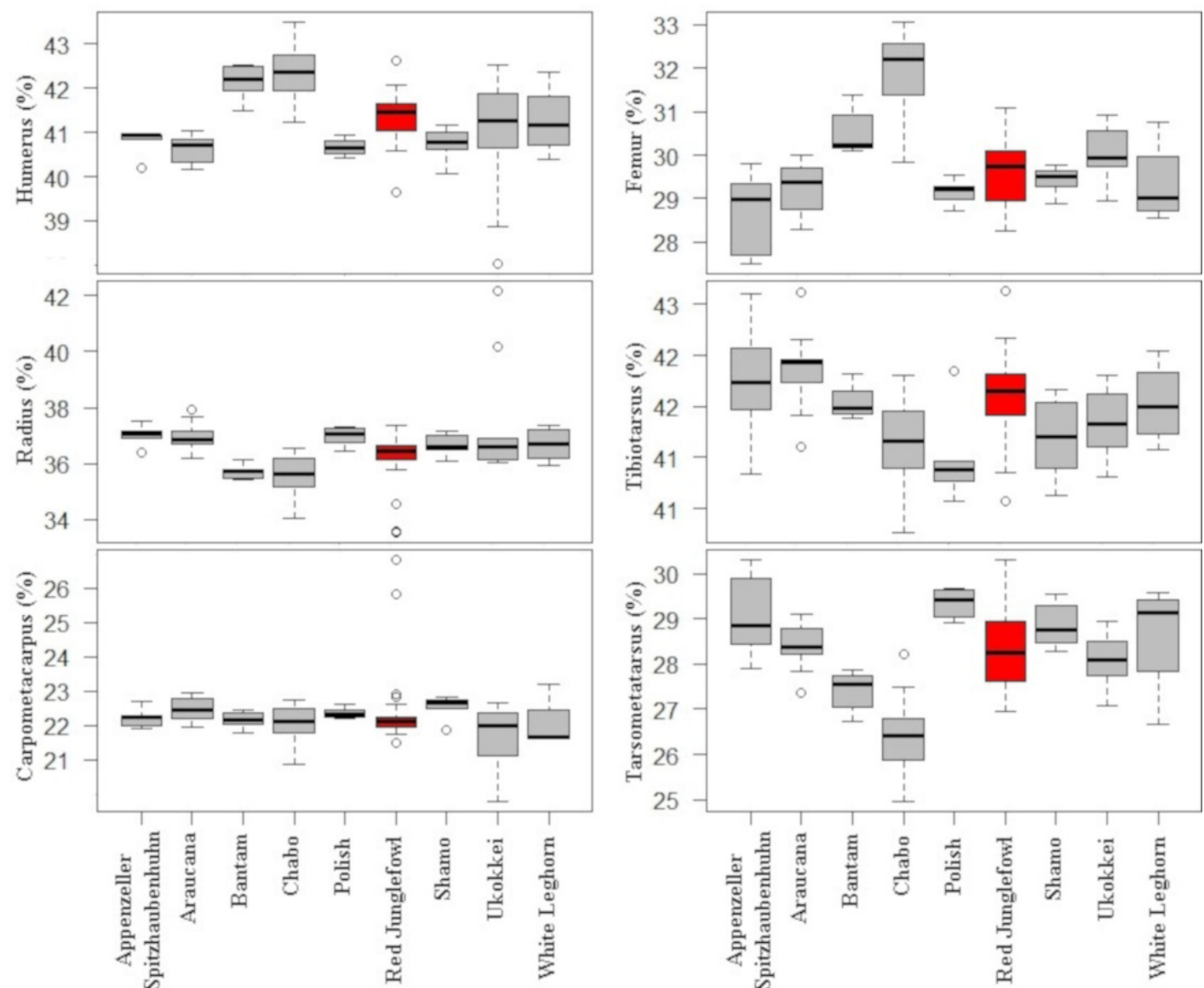
Axes representing the proportions of each bone in the limb as a percentage. (A) for forelimb, (B) for hind limb. Each plot is accompanied with zoomed in areas correspondent to each species differentiating wild forms (red dots) and domesticated forms (black dots). Circles in zoomed areas capture where most points for wild or domestic forms fall. (For statistical comparison of proportions see *Table 1*).



# Figure 7

Comparison of limb bones proportions among breeds of chicken.

Each figure depicts the proportion of one bone in its limb (forelimb bones shown on the left panels, hind limb bones on the right panels). On the vertical axis, the proportion of the bone in the limb. On the horizontal axis, the name of the breed to which the above distribution belongs. RJF is highlighted in red.



# Figure 8

Log-transformed length of individual bone of the forelimb and hind limb vs. log-transformed length of the respective limb as a proxy for body size.

(A and B) for *Gallus gallus*, (C and D) for *Anas platyrhynchos*, (E and F) for *Cairina moschata*.

Domestic breeds are represented with a hollow circle and wild forms are represented with a filled dot. Included are regression equations, regression lines, R squared values, F statistics including degrees of freedom in brackets, and the p-values. Measurements in centimetres.

