

Is there a hybridization barrier between *Gentiana lutea* color morphs?

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In *Gentiana lutea* two varieties are described, *G. lutea* var. *aurantiaca* with orange corolla colors and *G. lutea* var. *lutea* with yellow corolla colors. Both varieties co-occur in NW Spain. However, it is not known whether a hybridization barrier exists between these *G. lutea* color varieties. If that is the case, this barrier might reflect that *G. lutea* varieties are the product of a secondary contact or that they are currently under an ongoing speciation process. We know that abiotic environmental factors are not related to flower color variation among *G. lutea* populations in NW Spain. But, pollinators cause selection on flower color in this species, if *G. lutea* is dependent on pollinators, they might be currently driving or reinforcing the existence of these two morphs. Thus, we aim to test the compatibility between flower color varieties in *G. lutea* and its dependence on pollen vectors. Within a *G. lutea* population we analyzed differences in reproductive success (number, weight, viability and germinability of seeds) depending on fertilization treatments (autogamy and xenogamy within variety and among varieties). Our results confirmed that reproductive success is higher within color varieties than among varieties, due to seed viability reduction on hybrids from different varieties, and that *G. lutea* reproductive success is strongly dependent on pollinators. Regardless of the allopatric or sympatric origin of *G. lutea* flower color, we conclude that a partial hybridization barrier exists between *G. lutea* varieties, which are at least partially driven by the dependence of *G. lutea* on pollinators.

1 Is there a hybridization barrier between *Gentiana lutea* color morphs?

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11 Abstract

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lutea reproductive success is strongly dependent on pollinators. Regardless of the allopatric or sympatric origin of *G. lutea* flower color, we conclude that a partial hybridization barrier exists between *G. lutea* varieties, which are at least partially driven by the dependence of *G. lutea* on pollinators.

Keywords Pollen vectors, *Gentiana lutea*, flower color morphs, self-incompatibility, hybridization barrier, outcrossing success.

Introduction

Pollen vectors can drive plant evolution and diversification throughout their selection on floral traits (Darwin, 1859; Darwin, 1862; Thompson, 1994; Barrett & Harder, 1996; Charlesworth, 2006). In particular, animal pollinators exert selective pressures on plant traits through natural selection (Stebbins, 1970) and many flowering plants rely on those pollen vectors to reproduce. Species widely differ in their dependence on pollen vectors for seed production, from complete autogamy to xenogamy, requiring cross-pollination for successful reproduction (Darwin, 1877; Axelrod, 1960). Most angiosperms need vectors for pollen transference between plants, which are mainly insects but can also be other animals and to a lesser extent wind or water (Harder & Barrett, 1996; Ackerman, 2000; Ollerton, Winfree & Tarrant, 2011). Different floral strategies were developed to attract these animals (Ghazoul, 2006), which may affect plant fitness to a large degree (Waser, 1983; Conner & Rush, 1996). This plant-pollinator relationship promotes evolution of species involved in such an interaction. Thus, the degree of dependence on animal pollinators might affect the strength of selection and therefore the likelihood of species diversification.

Flower color variation in polymorphic species may originate from different selective pressures exerted by pollinators, which favor isolation between different color morphs and

cause sympatric diversification (Gegear & Burns, 2007). Thus, pollinator behavior may favor one of the color morphs, with individuals visiting one color morph more frequently than a different flower color morph (Hopkins & Rausher, 2012). Additionally, many plant species show floral divergences due to geographic isolation suffered by climatic changes during Quaternary period (Gómez & Lunt, 2007; Vargas et al., 2009; Martín-Bravo et al., 2010; Fernández-Mazuecos & Vargas, 2011; Blanco-Pastor, Vargas & Pfeil, 2012; Blanco-Pastor & Vargas, 2013; Fernández-Mazuecos et al., 2013). Thus, a secondary contact between flower color varieties that remained geographical isolated after a long time, could explain reproductive incompatibility between floral color morphs. Despite the origin of the phenotypic floral variation, diversification between flower color varieties may be reinforced by pollinator preferences (Campbell, Alarcon & Wu, 2003).

Gentiana lutea L. shows flower color variation from orange to yellow both, within and among populations (Sobral et al., 2015) at the western extreme of the distribution range. Two different varieties are described for *Gentiana lutea* L. depending on flower color: *Gentiana lutea* L. var. *aurantiaca* (M. Laínz) showing orange corolla colors and *Gentiana lutea* L. var. *lutea* showing yellow corollas (Laínz, 1982; Renobales, 2003). *Gentiana lutea*'s flower color is a trait with a genetic basis (Zhu et al., 2002; Zhu et al., 2003) and there are genetic differences among populations (González-López et al., 2014). We know that flower color variation among populations in this species across NW Spain, is not related to abiotic environmental factors such as elevation, temperature, radiation and rainfall (Veiga et al., 'in press'). Additionally, we know that pollinators exert selective pressures on *G. lutea* flower color (Veiga et al., 2015) and play a role in flower color differentiation among *G. lutea* populations (Sobral et al., 2015). But, it is not completely understood the degree of dependence of *G. lutea* on pollen vectors to successfully reproduce. Rossi (2012) described *G. lutea* as a partial self-compatible species. However, others cite *G. lutea* as a self-incompatible

species, which may need animal pollinators to reproduce (Hegi, 1927; Kéry, Matthies & Spillmann, 2000; Kozuharova & Anchev, 2006; González-López et al., 2014).

The aim of this study is to test if there is some degree of incompatibility between *Gentiana lutea* color varieties due to a partial hybridization barrier, which might be caused or reinforced by selective pressures exerted by pollinators on *G. lutea* flower color. If a hybridization barrier exists between *G. lutea* color morphs we might expect crossings among varieties to have a lower reproductive success (Hauser, Jørgensen & Østergård, 1998; Hauser, Shaw & Østergård, 1998). Additionally, if *G. lutea* color morphs are under a diversification process driven by selective pressures exerted by pollinators (Veiga et al., 2015; Sobral et al., 2015), we would expect *G. lutea* to strongly depend on the pollinating vectors exerting this selection. Thus, *G. lutea* crossings among different individuals would have higher reproductive success than crossings within-individual plants. In order to investigate our hypothesis, we proposed the following questions; (i) Are there any differences in reproductive success (seed number, seed weight, seed viability and seed germinability) between within-color variety crossings and among-color variety crossings? (ii) Are there any differences in reproductive success between autogamic seeds and seeds coming from among individual crossings?

Materials & Methods

Gentiana lutea L. (Gentianaceae) is a herbaceous perennial plant distributed along the Central and Southern European mountains typically growing on livestock grazing grasslands and hillsides from montane to sub-alpine habitats, approximately from 800 to 2.500 m a.s.l. (Hesse, Rees & Müller-Schärer, 2007; Anchisi et al., 2010). This long-lived species presents a rhizome, which develops one (rarely two or three) unbranched stout stem with basal leaves, approximately growing up to 200 cm tall (Renobales, 2012). Flowering occurs in summer

(June-July) when the fertile stems show flowers, grouped in pseudo-whorls, which bloom spirally; and the inflorescence develops in succession from apex to base (Kozuharova, 1994). There are two different varieties that differ in flower color: *Gentiana lutea* L. var. *aurantiaca* (M. Láinz) with orange flowers and *Gentiana lutea* var. *lutea* L. with yellow flowers (Láinz, 1982; Renobales, 2003). *Gentiana lutea* flowers show rotate corollas with lobes (Renobales, 2012), which facilitate the access of pollinators to the nectaries; and are visited by at least 11 families of insects, belonging to four orders (Rossi et al., 2014). The main flower visitors of *G. lutea* plants at the Cantabrian Mountains are bumblebees, followed by cuckoo bumblebees and honeybees (Sobral et al., 2015). Fruits hold many flattened, elliptic and winged seeds disseminated through anemochory (Struwe & Albert, 2002).

We carried out experimental manipulations at one *G. lutea* population in León, Spain (43° 03' N; 6° 04' W; 1600 m a.s.l.) in July 2012. For field experiments, we received field permit from Environmental Territorial Service from León, Territorial Delegation of Government of Spain, Regional Government of Castilla and León (Identifier:12_LE_325_RNA_PuebladeLilio_INV; Reference: 06.01.013.016/ROT/abp; File number: AEN/LE/103/12). Note that in this study, we will distinguish two different flower color classes (orange and yellow). Both varieties do not differ in their UV reflectance but they do differ in their visible reflectance (see Veiga et al., 2015); thus, we use visible reflectance to indentify color varieties. We haphazardly chose five flowers on each of 25 orange-flowered plants and 25 yellow-flowered plants. Flower buds, except the control group, were bagged with tulle before flower opening and until fruit formation in order to avoid contact with pollen vectors.

We created the following treatments: (1) Control group (C), in which we applied no treatment, so natural pollination occurred; (2) Spontaneous autogamy (Sa), in which pollen was not applied manually and only pollen from the same flower may spontaneously arrive to the ovary; (3) Facilitated autogamy (Fa), in which we applied pollen manually from the flower's

own anthers; (4) Facilitated xenogamy within varieties (F_{xw}), in which we emasculated flowers by cutting the stamens before pollen release in order to prevent the entry of flower's own pollen, and applied pollen manually from other plants of the same flower color; (5) Facilitated xenogamy among varieties (F_{xa}), in which we emasculated the flowers and applied pollen manually from plants with different flower color.

Reproductive success can be quantified as the number of fertile descendants produced by an individual throughout its life. It is not feasible to quantify reproductive success in long-lived species in these terms, so seed production is considered a good estimate (see the review of Kingsolver et al., 2001). Other measures of reproductive success are seed viability and seed germinability. We used four measures of reproductive success: number of seeds, weight of seeds (mg), rate of viable seeds (seed viability) and germination rate (seed germinability). Due to manipulations some bags were opened, thus plants with the five treatments were down to 26. 130 ripped fruits were collected before opening, being careful to ensure that the seeds were fully formed. On each fruit we measured seed weight (mg) and counted the number of seeds and the number of ovules not developed; from the sum of non-fertilized ovules and the seeds we obtained the total number of ovules. Total number of ovules did not vary among treatments (data not shown), thus we used the absolute number of seeds produced in each fruit, instead of calculating the seed production relative to the ovules on each fruit.

We also measured seed germinability and seed viability. We haphazardly chose up to 20 seeds in each fruit and distributed them on filter paper in petri plates. Seed number was very low in the autogamy treatments; thus, seeds from spontaneous autogamy (S_a) and facilitated autogamy (F_a) treatments were grouped. In total, we analyzed viability and germination rate of 1,800 seeds (10 plates per treatment and between 30 and 50 seeds per plate). Germination was induced with gibberellic acid 100 mg/L at 24 h of darkness and constant temperature of 23 °C (Bell et al., 1995). The state of germination and wetting of the plates were controlled on

alternate days; the filter paper was removed every 2-3 days to reduce fungal infection. Seeds with a radicle of at least 2 mm were considered germinated. Seed germinability is the percentage of germinated seeds. We controlled germination rate for a period of 45 days. After 45 days we tested seed viability by crushing the seeds with the tip of the tweezers. Soft and dark seeds were not considered viable and harsh and light-colored seeds were considered viable (as Hesse, Rees & Müller-Schärer, 2007). These seeds along with the germinated seeds were considered the total number of viable seeds.

We calculated the Self-Compatibility Index (SCI) to describe the *G. lutea* breeding system (as Lloyd & Schoen, 1992). SCI is assessed as the average seed set for facilitated autogamy (Fa) divided by the average seed set for facilitated xenogamy (F_{xw} or F_{xa}) and gives information about the self-compatibility of the species. SCI values range from 0 to 1.5 and a species is considered self-incompatible when its values are between 0 and 0.75 (Lloyd & Schoen, 1992).

To assess the occurrence of self-fertilization, we calculated the Auto-Fertility Index (AFI) dividing the seed set for spontaneous autogamy (S_a) by the seed set for facilitated xenogamy within varieties (F_{xw} or F_{xa}) (Lloyd & Schoen, 1992). AFI gives information about the autonomous autogamy degree of the species.

In order to analyze the differences in seed number and seed weight among treatments we used a generalized linear mixed model (GzLMM); the fixed factors were the treatment (five categories: C “Control group”, S_a “Spontaneous autogamy”, F_a “Facilitated autogamy”, F_{xw} “Facilitated xenogamy within varieties”, F_{xa} “Facilitated xenogamy among varieties”), the maternal flower color (color variety) and the interaction between color variety and treatment. Plant individual was a random factor nested within maternal flower color (color variety).

To test for differences in seed viability and germinability depending on treatment, we used a generalized linear model (GzLM) for each response variable. Treatment was a fixed factor

(with four categories, since the seeds of spontaneous and facilitated autogamy treatments were joint to have a sufficient sample). Note that first, we performed a generalized linear mixed model (GzLMM), where treatment was a fixed factor and plate number was a random factor nested within treatment, but it did not converge. Additionally, we could not test if the germination rate varied between colors or individuals because of insufficient sample size. Error distribution and link function were selected to minimize the AIC_C: number of seeds was fitted to a Poisson distribution and a logarithmic link function; whereas weight of seeds was fitted to a Linear distribution and identity link function. Both seed viability and seed germinability were adjusted to a Binomial distribution and probit link function. Analyses were performed with SPSS software (IBM SPSS Statistics for Windows, Version 20.0, IBM Corp., Somers, NY).

Results

The most successful reproductive mechanism for *Gentiana lutea* was found to be cross-pollination between individuals within the same color variety. Conversely, cross-pollination among flower color varieties reduced plant reproductive success. In addition, we found that *G. lutea* depends on pollinators to reproduce.

Number of seeds, seed weight and germinability were similar for within-color variety crossings than for among-color variety crossings (Table 1; Fig. 1; Fig. 2). However, seeds from crossings within varieties present higher viability than seeds from among variety crossings (Fig. 2). Thus, seed viability decreases when pollen donor and receptor belong to different color varieties (Fig. 2). This fact may imply that seed viability decrease is an important component of the partial hybridization barrier between *G. lutea* color morphs. Fruits under autogamy treatments produced fewer and lighter seeds than fruits from inter-

individual crossings. In addition, the effect of treatments on seed weight was the same for both color varieties (no significant color variety * treatment interaction: $p > 0.05$; Table 1; Fig. 1); although we found that under natural pollination, orange-flowering individuals produced more seeds per fruit than yellow-flowering individuals (significant simple effect of flower color: $p < 0.001$; Table 1; Fig. 1). On the contrary, within the spontaneous autogamy treatment, yellow-flowering individuals produced more seeds per fruit ($p < 0.001$; Table 1; Fig. 1). Additionally, we found significant differences among treatments for both, seed viability and seed germinability ($p < 0.001$; Table 1; Fig. 2). Note that both xenogamy treatments resulted in greater seed germinability than the autogamy treatments and control group (Fig. 2).

The Self-compatibility index scored 0.0687. SCI ranges from 0 to 1.5, with values of SCI under 0.75 meaning that the species is self-incompatible. Thus, *Gentiana lutea* is a self-incompatible species which relies in cross-pollination to successfully reproduce. Auto-Fertilization Index (AFI) scored 0.0695 meaning that *G. lutea* is a non-autogamous species.

Discussion

The most advantageous reproductive mechanism for *Gentiana lutea* L. is cross-pollination between the same color morphs. Our results suggest that a partial hybridization barrier exists between *G. lutea* flower color morphs, likely due to a past or ongoing divergence between *G. lutea* varieties. Regardless of the origin of both varieties, it seems that the differentiation is driven or reinforced by the strong dependency of *G. lutea* on pollen vectors which exert selective pressures on flower color (Veiga et al., 2015; Sobral et al., 2015).

It is well known that flower color and other floral traits (aroma, shape, flower sap content) play an essential role on plant reproduction, because they lead animal preferences (Tepedino,

1979; Quattrocchio et al., 1999). Specific sets of floral traits (pollination syndromes) are often associated with particular groups of pollinators. This close animal-plant interaction among species may drive floral evolution (Fenster et al., 2004; Hoballah et al., 2007). Angiosperm diversification may derive from reproductive isolation due to changes in pollinator habitat composition (Bradshaw & Schemske, 2003; Streisfeld & Kohn, 2007; Hoballah et al., 2007; Dauber et al., 2010). A speciation process among flower color varieties of the same species can occur (Straw, 1955; Waser, 1998; Gegear & Burns, 2007) and may be driven by differences in pollinator community, which show higher preference for one morph as many studies suggest (Dronamraju, 1960; Quattrocchio et al., 1999; Hopkins & Rausher, 2012). We know that *Gentiana lutea*'s pollinators show different color preferences among populations at the Cantabrian Mountains (Spain), partially due to differences on the pollinator spectrum within each population and specific flower color preferences from each pollinator species (Sobral et al., 2015). Sympatric speciation could originate due to selective pressures exerted by the pollinators (Quattrocchio et al., 1999; Hopkins & Rausher, 2011) that facilitate isolation between flower color varieties when cross-pollinations are avoided or reduced (Hopkins & Rausher, 2012).

Floral divergence is not always sufficient to produce isolation between morphs and consequent speciation (Fernández-Mazuecos & Vargas, 2011). The geographic isolation produced by the Quaternary climatic changes, which can be accompanied by differences in the pollinator spectra from isolated populations, has been identified as the main cause of divergence in several mountane plant species (Hewitt, 2000; Thompson, 2005; Martín-Bravo et al., 2010; Alarcón et al., 2012; Blanco-Pastor & Vargas, 2013; Fernández-Mazuecos et al., 2013). A subsequent secondary contact between both color morphs in *Gentiana lutea* L. could generate a similar situation to that actually shows our study population and surrounding area, whether is a recent or an old contact, it seems that the maintenance of different color morphs

is reinforced by the fitness reduction on hybrids and the pollinator behavior, as our results confirmed (Table 1; Fig. 1; Fig. 2).

The Self-Compatibility Index obtained here suggests that *Gentiana lutea* L. is a self-incompatible species (Lloyd & Schoen, 1992). Therefore, this species relies on pollen vectors for a successful reproduction, as in the case of the most flowering-plant species (Axelrod, 1960; Tepedino, 1979). The Self-Compatibility Index ($SCI=0.068$) results were similar to the Auto-Fertilization Index ($AFI= 0.069$). Note that SCI is the ratio between the average seeds produced in facilitated autogamy treatment and the average seeds produced in facilitated xenogamy, whereas AFI is assessed from the ratio between seeds set from spontaneous autogamy and facilitated xenogamy treatment. Similar values of the two indexes imply that the number of seeds produced when a flower's own pollen arrives both, naturally and manually is similar. This fact may suggest that self-incompatibility in this species is not caused by a physical barrier separating pollen from the same flower, but is caused by some pre-zygotic barrier mechanism in which pollen from same flower is not fertilizing the ovules, or by some post-zygotic barrier that may produce a lower quality and number of autogamic seeds (Charlesworth & Charlesworth, 1987; Hopkins, 2013).

It is an interesting fact that seed number and weight, and to a lesser extent seed germinability, from natural pollination is intermediate between autogamy treatments and xenogamy crossings (see Fig. 1; Fig. 2). This result suggests that pollen vectors carry to a plant, in natural conditions, both pollen from that plant and pollen from different plants. Therefore, those animal pollinators that have higher mobility between plants and lower mobility among flowers within plants would pollinate more successfully (Dauber et al., 2010; Rossi et al., 2014).

On the other hand, yellow-flowering plants set a greater number of seeds when spontaneous self-pollination occurs, which might suggest that the self-incompatibility degree in this

species varies among varieties and is actually higher for orange-flowering individuals (Fig. 1). This implies that the Auto-Fertilization Index might vary among color morphs. Additionally, we found differences in seed number within the natural pollination treatment, in which orange flowers set a greater number of seeds. We know that pollinator assemblage shows preferences for yellowness within our study population, in which yellow-flowering individuals have a greater total seed set (Veiga et al., 2015). However, our results suggest that although orange-flowering plants have lower fitness than yellow ones, they produce more seeds per fruit (see Table1; Fig. 1).

We know that pollinators exert selective pressures on *Gentiana lutea* L. flower color (Veiga et al., 2015) and that these selective pressures drive flower color differentiation in this species (Sobral et al., 2015). Additionally, we know that abiotic factors such as temperature, radiation, elevation and rainfall are not related to flower color variation among *G. lutea* populations (Veiga et al., 'in press'). With the available information, it is not clear whether we are dealing with an ecological speciation process in which the original color differentiation is due to an allopatric or sympatric process; but our results bring to light the existence of phenotypic differentiation and a lower fitness in hybrids. Regardless of the origin of the divergence, it seems that today a divergence process is ongoing among *G. lutea* color morphs driven by selection on flower color exerted by pollinators.

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References

- 296 **Ackerman JD. 2000.** Abiotic pollen and pollination: ecological, functional, and evolutionary
297 perspectives. *Plant Systematics and Evolution* **222** (1–4): 167–185. DOI:
298 10.1007/BF00984101.
- 299 **Alarcón M, Vargas P, Sáez L, Molero J, Juan José Aldasoro JJ. 2012.** Genetic diversity of
300 mountain plants: Two migration episodes of Mediterranean *Erodium* (Geraniaceae). *Molecu-*
301 *lar Phylogenetics and Evolution* **63** (3): 866–876. DOI:10.1016/j.ympev.2012.02.031.
- 302 **Anchisi E, Bernini A, Piaggi E, Polani F. 2010.** *Genziane d'Europa* [Gentians of Europe].
303 Pavia: Verba & Scripta s.a.s. (In Italian)
- 304 **Axelrod DI. 1960.** The evolution of flowering plants. In: Tax S, ed. *Evolution after Darwin I.*
305 *The evolution of life*. Chicago: University of Chicago Press, 227–305.
- 306 **Barrett SCH, Harder LD. 1996.** Ecology and evolution of plant mating. *Tree* **11** (2): 73–79.
- 307 **Bell DT, Rokich DP, McChesney CJ, Plummer JA. 1995.** Effects of temperature, light and
308 gibberellic acid on the germination of seeds of 43 species native to Western Australia. *Journal*
309 *of Vegetation Science* **6** (6): 797–806.
- 310 **Blanco-Pastor JL, Vargas P, Pfeil BE. 2012.** Coalescent simulations reveal hybridization
311 and incomplete lineage sorting in Mediterranean *Linaria*. *PLoS ONE* **7** (6): e39089.
312 DOI:10.1371/journal.pone.0039089.
- 313 **Blanco-Pastor JL, Vargas P. 2013.** Autecological traits determined two evolutionary strate-
314 gies in Mediterranean plants during the Quaternary: low differentiation and range expansion
315 versus geographical speciation in *Linaria*. *Molecular Ecology* **22** (22): 5651–5668. DOI:
316 10.1111/mec.12518.
- 317 **Bradshaw HD, Schemske DW. 2003.** Allele substitution at a flower colour locus produces a
318 pollinator shift in monkeyflowers. *Nature* **426**: 176–178. DOI: 10.1038/nature02106.

- 319 **Campbell DR, Alarcon R, Wu CA. 2003.** Reproductive isolation and hybrid pollen disad-
320 vantage in *Ipomopsis*. *Journal of Evolutionary Biology* **16** (3): 536–540. DOI:
321 10.1046/j.1420-9101.2003.00538.x.
- 322 **Charlesworth D, Charlesworth B. 1987.** Inbreeding depression and its evolutionary conse-
323 quences. *Annual Review of Ecology and Systematics* **18**:237–268. DOI:
324 10.1146/annurev.es.18.110187.001321.
- 325 **Charlesworth D. 2006.** Evolution of plant breeding systems. *Current Biology* **16**: 726–735.
326 DOI: 10.1016/j.cub.2006.07.068.
- 327 **Conner JK, Rush S. 1996.** Effects of flower size and number on pollinator visitation to wild
328 radish, *Raphanus raphanistrum*. *Oecologia* **105** (4): 509–516. DOI: 10.1007/BF00330014.
- 329 **Darwin CR. 1859.** Chapter IV: Natural selection. In: Murray J, ed. *On the origin of species*
330 *by means of natural selection, or the preservation of favoured races in the struggle for life*.
331 London: W. Clowes and Sons, 92–95.
- 332 **Darwin CR. 1862.** Chapter I: Structure of orchis. In: Murray J, ed. *On the various*
333 *contrivances by which British and foreign orchids are fertilised by insects and on the good*
334 *effects of intercrossing*. London: W. Clowes and Sons, 34–44.
- 335 **Darwin CR. 1877.** Chapter XII: General results. In: Appleton D and Company, ed. *The*
336 *effects of cross and self-fertilization in the vegetable kingdom*. New York: Appleton, 436–469.
- 337 **Dauber J, Biesmeijer, JC, Gabriel D, Kunin WE, Lamborn E, Meyer B, Nielsen A, Potts**
338 **SG, Roberts PM, Sober V, Settele J, Steffan-Dewenter I, Stout JC, Teder T, TscheulinT,**
339 **Vivarelli D, Petanidou T. 2010.** Effects of patch size and density on flower visitation and
340 seed set of wild plants: a pan-European approach. *Journal of Ecology* **98**: 188–196. DOI:
341 10.1111/j.1365-2745.2009.01590.x.
- 342 **Dronamraju KR. 1960.** Selective visits of butterflies to flowers, a possible factor in
343 sympatric speciation. *Nature* **186**:178–179. DOI: 10.1038/186178a0.

- 344 **Fenster CB, Armbruster WS, Wilson P, Dudash MR, Thomson JD. 2004.** Pollination
- 345 syndromes and floral specialization. *Annual Review of Ecology, Evolution and Systematics*
- 346 **35:** 375–403. DOI: 10.1146/annurev.ecolsys.34.011802.132347.
- 347 **Fernández-Mazuecos M, Vargas P. 2011.** Historical isolation *versus* recent long-distance
- 348 connections between Europe and Africa in bifid toadflaxes (*Linaria* sect. *Versicolores*). *PLoS*
- 349 *ONE* **6** (7): e22234. DOI:10.1371/journal.pone.0022234.
- 350 **Fernández-Mazuecos M, Blanco-Pastor JL, Gómez JM, Vargas P. 2013.** Corolla
- 351 morphology influences diversification rates in bifid toadflaxes (*Linaria* sect. *Versicolores*).
- 352 *Annals of Botany* **112:** 1705–1722. DOI: 10.1093/aob/mct214.
- 353 **Gegeer RJ, Burns JG. 2007.** The birds, the bees, and the virtual flowers: Can pollinator
- 354 behavior drive ecological speciation in flowering plants? *The American Naturalist* **170** (4):
- 355 551–566. DOI: 10.1086/521230.
- 356 **Ghazoul J. 2006.** Floral diversity and the facilitation of pollination. *Journal of Ecology* **94:**
- 357 295–304. DOI: 10.1111/j.1365-2745.2006.01098.x.
- 358 **Gómez A, Lunt DH. 2007.** Refugia within refugia: patterns of phylogeographic concordance
- 359 in the Iberian Peninsula. In: Weiss S, Ferrand N, eds. *Phylogeography of Southern European*
- 360 *refugia*. Dordrecht: Springer, 155–188. DOI: 10.1007/1-4020-4904-8_5.
- 361 **González-López O, Polanco C, György Z, Pedryc A, Casquero PA. 2014.** Genetic
- 362 variation of the endangered *Gentiana lutea* L. var. *aurantiaca* (Gentianaceae) in populations
- 363 from the Northwest Iberian Peninsula. *International Journal of Molecular Sciences* **15** (6):
- 364 10052–10066. DOI: 10.3390/ijms150610052.
- 365 **Harder LD, Barrett SCH. 1996.** Pollen dispersal and mating patterns in animal-pollinated
- 366 plants. In: Lloyd DG, Barrett SCH, eds. *Floral biology: Studies on floral evolution in animal-*
- 367 *pollinated plants*. New York: Chapman & Hall, 140–190.
- 368 **Hauser TP, Jørgensen RB, Østergård H. 1998.** Fitness of backcross and F2 hybrids

between weedy *Brassica rapa* and oilseed rape (*B. napus*). *Heredity* **81**: 436–443.
DOI:10.1046/j.1365-2540.1998.00425.x.

Hauser TP, Shaw RG, Østergård H. 1998. Fitness of F1 hybrids between weedy *Brassica rapa* and oilseed rape (*B. napus*). *Heredity* **81** (4): 429–435. DOI:10.1046/j.1365-2540.1998.00424.x.

Hegi G. 1927. *Illustrierte Flora von Mitteleuropa*. München: JF Lehmanns-Verlag.

Hesse E, Rees M, Müller-Schärer H. 2007. Seed bank persistence of clonal weeds in contrasting habitats: implications for control. *Plant Ecology* **190**: 233–243. DOI: 10.1007/s11258-006-9203-7.

Hewitt GH. 2000. The genetic legacy of quaternary ice ages. *Nature* **405**: 907–913. DOI: 10.1038/35016000.

Hoballah ME, Gübitz T, Stuurman J, Broger L, Barone M, Mandel T, Dell’Olivo A, Arnold M, Kuhlemeier C. 2007. Single gene– mediated shift in pollinator attraction in *Petunia*. *Plant Cell* **19** (3): 779–790. DOI: 10.1105/tpc.106.048694.

Hopkins R, Rausher MD. 2011. Identification of two genes causing reinforcement in the Texas wildflower *Phlox drummondii*. *Nature* **469**: 411–414. DOI: 10.1038/nature09641.

Hopkins R, Rausher MD. 2012. Pollinator-mediated selection on flower color allele drives reinforcement. *Science* **335** (6070): 1090–1092. DOI: 10.1126/science.1215198.

Hopkins R. 2013. Reinforcement in plants. *New Phytologist* **197** (4): 1095–1103. DOI: 10.1111/nph.12119.

IBM Corp. Released. 2011. *IBM SPSS Statistics for Windows, Version 20.0*. Armonk, NY: IBM Corp.

Kéry M, Matthies D, Spillmann HH. 2000. Reduced fecundity and offspring performance in small populations of the declining grassland plants *Primula veris* and *Gentiana lutea*. *Journal of Ecology* **88**: 17–30. DOI: 10.1046/j.1365-2745.2000.00422.x.

- Kingsolver JG, Hoekstra HE, Hoekstra JM, Berrigan D, Vignieri SN, Hill CE, Hoang A, Gibert P, Beerli P. 2001.** The strength of phenotypic selection in natural populations. *The American Naturalist* 157 (3): 245–261. DOI: 10.1086/319193.
- Kozuharova EK. 1994.** Modes of pollination of some *Gentiana* L. species distributed in Bulgaria. *Annals of the University of Sofia, 2, Botanika* 85: 215–224.
- Kozuharova EK, Anchev ME. 2006.** Nastic corolla movements of nine *Gentiana* species (Gentianaceae), presented in the Bulgarian flora. *Phytologia Balcanica* 12 (2): 255–265.
- Laínz M. 1982.** *Mis contribuciones al conocimiento de la flora de Asturias* [My contributions to the knowledge of the Asturian flora]. Oviedo: Diputación Provincial de Asturias, Instituto de Estudios Asturianos (CSIC).
- Lloyd DG, Schoen DJ. 1992.** Self- and cross-fertilization in plants. I. Functional dimensions. *International Journal of Plant Sciences* 153 (3): 358–369.
- Martín-Bravo S, Valcárcel V, Vargas P, Luceño M. 2010.** Geographical speciation related to Pleistocene range shifts in the Western Mediterranean Mountains (*Reseda* sect. *Glaucoreседа*, *Resedaceae*). *Taxon* 59 (2): 466–482.
- Ollerton J, Winfree R, Tarrant S. 2011.** How many flowering plants are pollinated by animals? *Oikos* 120: 321–326. DOI: 10.1111/j.1600-0706.2010.18644.x.
- Quattrocchio F, Wing J, van der Woude K, Souer E, de Vetten N, Mol J, Koes R. 1999.** Molecular analysis of the anthocyanin2 gene of *Petunia* and its role in the evolution of flower color. *The Plant Cell* 11: 1433–1444.
- Renobales G. 2003.** Notas acerca del tratamiento de las Gentianaceae para flora Ibérica [Notes about the treatment of the Gentianaceae for Iberian flora]. *Anales del Jardín Botánico de Madrid* 60 (2): 461–469. (In Spanish)
- Renobales G. 2012.** *Gentiana lutea* L. In: Castroviejo S, Talavera S, Andrés C, Arista M, Fernández Piedra MP, Gallego MJ, Ortiz PL, Romero Zarco C, Salgueiro FJ, Silvestre S,

- Quintanar A, eds. *Flora ibérica XI: Gentianaceae-Boraginaceae* [Iberian flora XI: Gentianaceae-Boraginaceae]. Madrid: Real Jardín Botánico (CSIC), 10–13. (In Spanish)
- Rossi M. 2012.** Taxonomy, phylogeny and reproductive ecology of *Gentiana lutea* L. D. Phil. Thesis, University of Bologna, Italy.
- Rossi M, Fisogni A, Nepi M, Quaranta M, Galloni M. 2014.** Bouncy versus idles: On the different role of pollinators in the generalist *Gentiana lutea* L. *Flora - Morphology, Distribution, Functional Ecology of Plants* **209** (3-4): 164–171. DOI: 10.1016/j.flora.2014.02.002.
- Sobral M, Veiga T, Domínguez P, Guitián JA, Guitián P, Guitián J. 2015.** Selective pressures explain differences in flower color among *Gentiana lutea* populations. *PLoS ONE* **10** (7): e0132522. DOI:10.1371/journal.pone.0132522.
- Stebbins GL. 1970.** Adaptive radiation of reproductive characteristics in angiosperms, I: Pollination mechanisms. *Annual Review of Ecology and Systematics* **1**: 307–326. DOI: 10.1146/annurev.es.01.110170.001515.
- Straw RM. 1955.** Hybridization, homogamy and sympatric speciation. *Evolution* **9** (4): 441–444.
- Streisfeld MA, Kohn JR. 2007.** Environment and pollinator-mediated selection on parapatric floral races of *Mimulus aurantiacus*. *Journal of Evolutionary Biology* **20**: 122–132. DOI: 10.1111/j.1420-9101.2006.01216.x.
- Struwe L, Albert VA. 2002.** *Gentianaceae: systematics and natural history*. Cambridge: Cambridge University Press.
- Thompson JN. 1994.** *The co-evolutionary process*. Chicago: University of Chicago Press.
- Thompson JD. 2005.** *Plant evolution in the Mediterranean*. Oxford: Oxford University Press.
- Tepedino VJ. 1979.** The importance of bees and other insect pollinators in maintaining floral

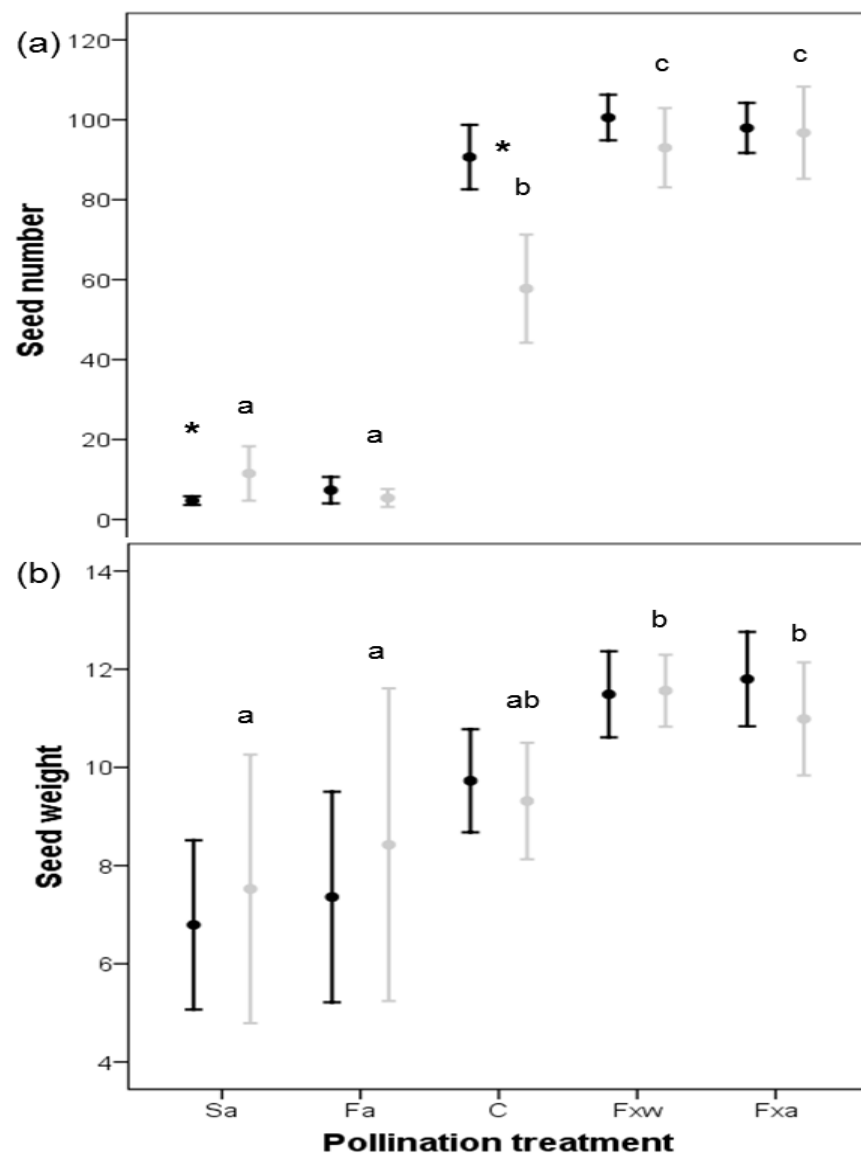
- species composition. *Great Basin Naturalist Memoires* **3**: 139–150.
- Vargas P, Carrió E, Guzmán B, Amat E, Güemes J. 2009.** A geographical pattern of *Antirrhinum* (Scrophulariaceae) speciation since the Pliocene based on plastid and nuclear DNA polymorphisms. *Journal of Biogeography* **36** (7): 1297–1312. DOI: 10.1111/j.1365-2699.2008.02059.x.
- Veiga T, Guitián JA, Guitián P, Guitián J, Sobral M. 2015.** Are pollinators and seed predators selective agents on flower color in *Gentiana lutea*? *Evolutionary Ecology* **29** (3): 451–464. DOI: 10.1007/s10682-014-9751-6.
- Veiga T, Guitián J, Guitián P, Guitián JM, Munilla I, Sobral M. 'in press'.** Flower color variation in the montane plant *Gentiana lutea* L. (Gentianaceae) is unrelated to abiotic factors. *Plant Ecology & Diversity*.
- Waser NM. 1983.** The adaptive nature of floral traits: ideas and evidence. In: Real L ed. *Pollination biology*. Orlando: Academic press, 241–285.
- Waser NM. 1998.** Pollination, angiosperm speciation, and the nature of species boundaries. *Oikos* **82** (1): 198–201. DOI: 10.2307/3546930.
- Zhu C, Yamamura S, Koiwa H, Nishihara M, Sandmann G. 2002.** cDNA cloning and expression of carotenogenic genes during flower development in *Gentiana lutea*. *Plant Molecular Biology* **48** (3): 277–285. DOI: 10.1023/A: 1013383120392.
- Zhu C, Yamamura S, Nishihara M, Koiwa H, Sandmann G. 2003.** cDNAs for the synthesis of cyclic carotenoids in petals of *Gentiana lutea* and their regulation during flower development. *Biochimica et Biophysica Acta (BBA)* **1625** (3): 305–308. DOI: 10.1016/S0167-4781(03)00017-4.

Table 1: Effect of different treatments of pollination on female reproductive success (number of seeds, seeds weight, seed viability and seed germinability).

We marked in bold the statistically significant factors ($P < 0.05$).

Response Variable	N	Factors	Variance	S.E.	Wald Chi-Square	d.f.	P
Seed number	130	<i>Random effect</i>					
		Plant (Color variety)	0.050	0.016			
		<i>Fixed effects</i>					
		Treatment			504.389	4	0.000
		Color morph			0.001	1	0.971
		Color morph*Treatment			23.222	4	0.000
Seed weight	108	<i>Random effect</i>					
		Plant (Color variety)	6.748	3.112			
		<i>Fixed effects</i>					
		Treatment			44.347	4	0.003
		Color morph			0.004	1	0.949
		Color morph *Treatment			0.177	4	0.950
Seed viability	1,800	<i>Fixed effects</i>					
		Treatment			30.906	3	0.000
Seed germinability	1,800	<i>Fixed effects</i>					
		Treatment			32.966	3	0.000

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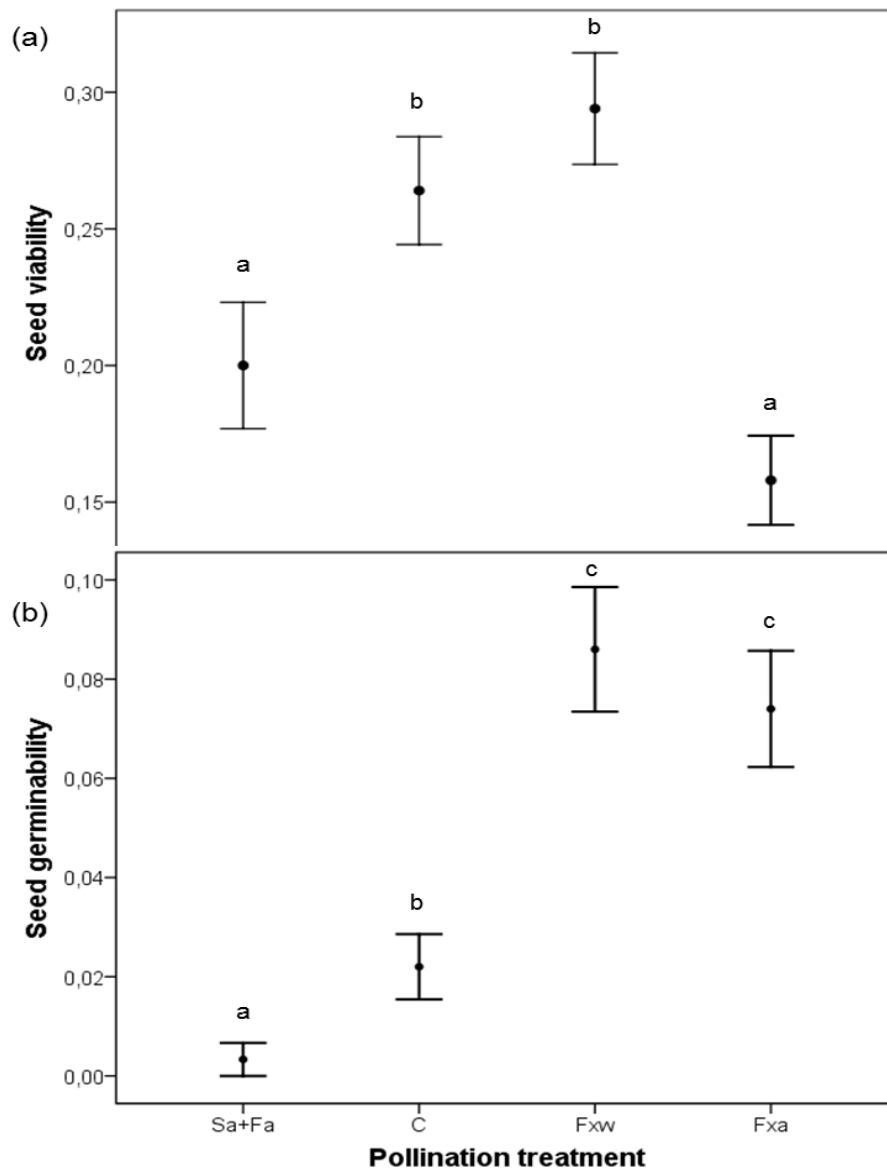


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476 **Figure 1: Reproductive success assessed by seed number (A) and seed weight in mg (B),**
477 **in function of different pollination treatments.**

478 Pollination treatments were: Spontaneous autogamy (Sa); Facilitated autogamy (Fa); Control
479 group (C), Facilitated xenogamy within varieties (Fxw) and Facilitated xenogamy among va-
480 rieties (Fxa). Significant statistical differences between treatments are marked with different
481 letters, and significant statistical differences between color varieties (orange: black-colored
482 bars; yellow: grey-colored bars) are marked with an asterisk. Bars show the Standard Error
483 (S.E.).

484



485

486 **Figure 2: Reproductive success assessed by seed viability (A) and seed germinability (B),**
 487 **in function of different pollination treatments.**

488 Pollination treatments were the following: Spontaneous autogamy (Sa) + Facilitated autoga-
 489 my (Fa); Control group (C), Facilitated xenogamy within varieties (Fxw) and Facilitated xe-
 490 nogamy among varieties (Fxa). The statistical significant differences between treatments are
 491 marked with different letters. Bars show the Standard Error (S.E.).