

Sensitivity of soil hydrogen uptake to natural and managed moisture dynamics in a semiarid urban ecosystem

Vanessa Buzzard^{Corresp., 1}, Dana Thorne¹, Juliana Gil-Loaiza¹, Alejandro Cueva², Laura K Meredith^{1, 3}

¹ School of Natural Resources and the Environment, University of Arizona, Tucson, Arizona, United States

² Biosphere2, University of Arizona, Oracle, Arizona, United States

³ BIO5 Institute, University of Arizona, Tucson, Arizona, United States

Corresponding Author: Vanessa Buzzard

Email address: vbuzzard@arizona.edu

The North American Monsoon season (June-September) in the Sonoran Desert brings thunderstorms and heavy rainfall. These rains bring cooler temperatures and account for roughly half of the annual precipitation making them important for biogeochemical processes. The intensity of the monsoon rains also increase flooding in urban areas and rely on green infrastructure (GI) stormwater management techniques such as water harvesting and urban rain gardens to capture runoff. The combination of increased water availability during the monsoon and water management provide a broad moisture regime for testing responses in microbial metabolism to natural and managed soil moisture pulses in drylands. Soil microbes rely on atmospheric hydrogen (H_2) as an important energy source in arid and semiarid landscapes with low soil moisture and carbon availability. Unlike mesic ecosystems, transient water availability in arid and semiarid ecosystems has been identified as a key limited driver of microbe-mediated H_2 uptake. We measured soil H_2 uptake in rain gardens exposed to three commonly used water harvesting practices during the monsoon season in Tucson AZ, USA. *In situ* static chamber measurements were used to calculate H_2 uptake in each of the three water harvesting treatments *passive* (stormwater runoff), *active* (stored rooftop runoff), and *greywater* (used laundry water) compared to an unaltered *control* treatment to assess the effects of water management practices on soil microbial activity. In addition, soils were collected from each treatment and brought to the lab for an incubation experiment manipulating the soil moisture to three levels capturing the range observed from field samples. H_2 fluxes from all treatments ranged between $-0.72 \text{ nmol m}^{-2} \text{ s}^{-1}$ and $-3.98 \text{ nmol m}^{-2} \text{ s}^{-1}$ over the monsoon season. Soil H_2 uptake in the greywater treatment was on average 53% greater than the other treatments during pre-monsoon, suggesting that the increased frequency and availability of water in the greywater treatment resulted in higher H_2 uptake during the dry season. H_2 uptake was

significantly correlated with soil moisture ($r = -0.393$, $p = 0.001$, $df = 62$) and temperature ($r = 0.345$, $p = 0.005$, $df = 62$). Our findings suggest that GI managed residential soils can maintain low levels of H_2 uptake during dry periods, unlike unmanaged systems. The more continuous H_2 uptake associated with GI may help reduce the impacts of drought on H_2 cycling in semiarid urban ecosystems.

Sensitivity of soil hydrogen uptake to natural and managed moisture dynamics in a semiarid urban ecosystem

Vanessa R. Buzzard¹, Dana Thorne¹, Juliana Gil-Loaiza¹, Alejandro Cueva², Laura K. Meredith^{1,3}

¹ School of Natural Resources and the Environment, University of Arizona, Tucson, Arizona, United States

² Main affiliation: Biosphere 2, University of Arizona, Tucson, AZ , USA

Current affiliation: Departamento de Ciencias de la Sustentabilidad, El Colegio de la Frontera Sur, Unidad Villahermosa, Tabasco, Mexico

³ BIO5 Institute, University of Arizona, Tucson, Arizona, United States

Corresponding Author:

Vanessa Buzzard¹

1657 E. Helen Street, Tucson, Arizona, 85719, United States

Email address: vbuzzard@email.arizona.edu

26 Abstract

27 The North American Monsoon season (June-September) in the Sonoran Desert brings
 28 thunderstorms and heavy rainfall. These rains bring cooler temperatures and account for roughly
 29 half of the annual precipitation making them important for biogeochemical processes. The
 30 intensity of the monsoon rains also increase flooding in urban areas and rely on green
 31 infrastructure (GI) stormwater management techniques such as water harvesting and urban rain
 32 gardens to capture runoff. The combination of increased water availability during the monsoon
 33 and water management provide a broad moisture regime for testing responses in microbial
 34 metabolism to natural and managed soil moisture pulses in drylands. Soil microbes rely on
 35 atmospheric hydrogen (H_2) as an important energy source in arid and semiarid landscapes with
 36 low soil moisture and carbon availability. Unlike mesic ecosystems, transient water availability
 37 in arid and semiarid ecosystems has been identified as a key driver of microbe-mediated H_2
 38 uptake. We measured soil H_2 uptake in rain gardens exposed to three commonly used water
 39 harvesting practices during the monsoon season in Tucson AZ, USA. *In situ* static chamber
 40 measurements were used to calculate H_2 uptake in each of the three water harvesting treatments
 41 *passive* (stormwater runoff), *active* (stored rooftop runoff), and *greywater* (used laundry water)
 42 compared to an unaltered *control* treatment to assess the effects of water management practices
 43 on soil microbial activity. In addition, soils were collected from each treatment and brought to
 44 the lab for an incubation experiment manipulating the soil moisture to three levels capturing the
 45 range observed from field samples. H_2 fluxes from all treatments ranged between $-0.72 \text{ nmol m}^{-2}$
 46 s^{-1} and $-3.98 \text{ nmol m}^{-2} \text{ s}^{-1}$ over the monsoon season. Soil H_2 uptake in the greywater treatment
 47 was on average 53% greater than the other treatments during pre-monsoon, suggesting that the
 48 increased frequency and availability of water in the greywater treatment resulted in higher H_2
 49 uptake during the dry season. H_2 uptake was significantly correlated with soil moisture ($r = -$
 50 0.393 , $p = 0.001$, $df = 62$) and temperature ($r = 0.345$, $p = 0.005$, $df = 62$). Our findings suggest
 51 that GI managed residential soils can maintain low levels of H_2 uptake during dry periods, unlike
 52 unmanaged systems. The more continuous H_2 uptake associated with GI may help reduce the
 53 impacts of drought on H_2 cycling in semiarid urban ecosystems.

54 Introduction

55 Atmospheric H_2 is an abundant trace gas with a global average of 530 ppb (Schmidt,
 56 1974; Novelli et al., 1999). The primary sources of atmospheric H_2 are identified as
 57 photochemical oxidation, combustion of fossil fuels, and biomass burning (Novelli et al., 1999).
 58 H_2 is an indirect greenhouse gas that contributes to climate change by competing for reactions
 59 with hydroxyl (OH) radicals in the troposphere (Lee, Rahn & Throop, 2012). If OH radicals react
 60 with H_2 , they are no longer available to react with methane (CH_4), a potent greenhouse gas,
 61 leading to more atmospheric CH_4 (Prather, 2003). H_2 is also a source of water vapor production
 62 in the stratosphere, which directly impacts ozone (Lee, Rahn & Throop, 2012). It is estimated
 63 that half of H_2 emissions are from human activity, disproportionately impacting atmospheric H_2

concentrations and the H₂ cycle of urban ecosystems (Novelli et al., 1999; Ehhalt & Rohrer, 2009). Thus, quantifying soil H₂ uptake in urban built landscapes is critical for understanding potential H₂ sinks, and the role of GI on H₂ cycling in drylands.

Soil uptake is the primary sink for H₂ from the atmosphere and is dominated by high affinity hydrogen-oxidizing bacteria (HA-HOB) (Conrad & Seiler, 1979; Conrad, 1996; Novelli et al., 1999; Ehhalt & Rohrer, 2009; Khdhiri et al., 2015; Piché-Choquette & Constant, 2019) that efficiently consume H₂ at atmospheric abundances. Annually, 75% of tropospheric H₂ is oxidized by soil microbes for use as an energy source (Schmidt, 1974; Rhee, Brenninkmeijer & Röckmann, 2006). The uptake of atmospheric H₂ into soils also contributes to the natural cycling of H₂ and has been studied extensively (Smith-Downey, Randerson & Eiler, 2006, 2008; Constant, Poissant & Villemur, 2009; Meredith et al., 2014, 2017; Greening et al., 2015; Khdhiri et al., 2015; Piché-Choquette & Constant, 2019). Both field and laboratory studies have shown that H₂ uptake increases with soil temperatures and drying (Smith-Downey, Randerson & Eiler, 2006, 2008; Meredith et al., 2017). Although microbial H₂ uptake varies during different life stages and is more common during late growth stages and dormancy (Constant et al., 2010; Meredith et al., 2014; Greening et al., 2015), recent studies have shown microbial uptake during active growth (Islam et al., 2020), which may be stimulated by moisture availability in arid and semiarid environments, highlighting the importance of H₂ for microbial metabolism in desert biomes (Jordaan et al., 2020).

Arid lands account for roughly 41% of the Earth's terrestrial surface, support more than one third of the world population, and are projected to expand with changes in climate (Wang, Chen & Dong, 2006; Feng & Fu, 2013; Maestre et al., 2015; Huang et al., 2016; Prăvălie, 2016). In arid and semiarid ecosystems, natural precipitation regimes are important determinants of ecological activity (Noy-Meir, 1973; Huxman et al., 2004). However, changes in natural hydrological processes due to human activity and management may be as important as natural patterns of precipitation on biogeochemical cycling (Austin et al., 2004). As more of the world's population inhabit and alter drylands, reliance on both natural and managed water sources will also increase.

Tucson, AZ is a city in the Southwest United States which experiences strong seasonal precipitation and increased reliance on green infrastructure (GI) water management practices. The city is home to nearly 1 million residents and is located in the Sonoran Desert, a semiarid ecosystem with two distinct rainy seasons, one during the winter months and one in the summer (Dimmitt, 2000). Unlike the winter rainy season, the summer rains are caused by the North American Monsoon which is characterized by a seasonal shift in the direction of the prevailing winds from the Pacific to winds from the south and southeast, leading to extreme precipitation events in the Southwest. The intensity of summer rain events accounts for half of the annual precipitation and is met by the reduced ability of urban desert soils to rapidly infiltrate such large volumes of water, which can lead to runoff and flash flooding. To combat stormwater runoff and

flooding during the North American Monsoon season, cities like Tucson have implemented GI water management practices to mimic natural hydrological processes by directing and capturing large volumes of rainwater in the soils.

GI techniques include greywater harvesting, rain water harvesting, bioretention basins, and green rooftops. The implementation of these techniques can help mimic the natural water cycle, alter abiotic factors like soil moisture and temperature, and increase microbe-mediated processes that can alter biogeochemical cycling. For example, microbe-mediated fluxes of important trace gases like carbon dioxide (CO_2), nitrous oxide (N_2O), and methane (CH_4) varied with GI design (Grover et al., 2013; McPhillips, Goodale & Walter, 2017; Shrestha, Hurley & Adair, 2018). Moreover, GI designed biofilters commonly used for stormwater management acted as a sink for CH_4 , and were a small source of N_2O during both wet and dry periods with greater CH_4 emissions during extreme wet events (Grover et al., 2013). Although soil moisture has been identified as the primary driver of fluxes, soil temperature was also reported as a driver of small and variable soil N_2O fluxes and CO_2 emissions from roadside bioretention basins (Shrestha, Hurley & Adair, 2018). In addition, flow-through bioretention basins (flat bottom soil bed) using engineered soil media with high phosphorus and C content were sources of CO_2 , N_2O and CH_4 (McPhillips, Goodale & Walter, 2017). Although these studies highlight impacts of GI on greenhouse gas emissions, they are constrained to moist temperate climates which may not translate to understanding natural and managed water inputs from water harvested rain gardens in desert urban ecosystems.

GI water management practices combined with the natural precipitation patterns in dryland environments impact soil moisture and temperature which directly impact biogeochemical cycling (Buzzard et al., 2021). Rain gardens irrigated with harvested water are common GI techniques that alter the nutrient inputs, and the frequency and quantity of wetting events. The soil moisture legacy in GI systems is also dependent on the local monsoonal precipitation regime, which brings cooler temperatures and ensures water is replenished for irrigation during periods of drought. These changes in precipitation imposed by this natural and managed moisture regime provide a unique opportunity to directly measure these critical environmental drivers to better understand the role of soil H_2 uptake in a dryland environment. In this study, we use a residential GI rain garden to assess how seasonal precipitation and irrigation from harvested water sources impact H_2 fluxes. To test the combined effects of seasonal precipitation and GI water management on soil H_2 fluxes, we ask, (i) how does the precipitation regime, specifically monsoons, of the Sonoran desert influence H_2 fluxes?; and (ii) do different GI water harvesting techniques affect soil H_2 fluxes? Greater H_2 uptake has been observed in ecosystems with high temperatures and variable soil moisture highlighting the sensitivity of H_2 to soil moisture availability in water-limited environments (Conrad & Seiler, 1985). Therefore, we hypothesize that increased water availability observed during the monsoon season leads to greater H_2 uptake and that soils from GI water harvesting rain gardens would result in more H_2

uptake during the dry season when managed basins continue to receive water through regular irrigation.

Materials & Methods

Study system

Our study site is located at a residence in central Tucson, Arizona - with a mean annual temperature of 17°C and the average annual rainfall is 32.2 cm (Paul R. Sheppard, Andrew C. Comrie, Gregory D. Packin, Kurt Angersbach, and Malcolm K. Hughes, 1999). In November of 2017, a green infrastructure (GI) water harvesting system was installed by Watershed Management Group Inc. at our study site; site pictures and study design are described in (Buzzard et al., 2021). Specifically, three rain garden basins were dug to direct and capture rainwater in a 3000 gallon plastic rain storage tank as described in (Buzzard et al. 2021). Three GI treatment plots (1 m x 4 m) received different water inputs and sources (*passive* stormwater runoff, *active* irrigated with tank-stored harvested rainwater, and *greywater* from laundry) along with an unaltered control treatment area that represented the initial, flat pea gravel-lain condition. All treatments were composed of four 1 m² batches within a 4 m by 1 m plot that received limited municipal city irrigation to establish plants and were subjected to the same rainfall, but differed in the inputs and frequency of irrigation from harvested water sources, and the type and depth of the mulch layer as described in (Buzzard et al. 2021). Specifically, the *greywater* treatment was irrigated with residential laundry water effluent and rain collected in the storage tank. The *active* water treatment received rainwater irrigation from a rain storage tank that collects and stores roof-top runoff, and may also receive overflow from the tank during large rain events that can flood the basin. The *passive* treatment was a dug basin designed to collect stormwater runoff and overflow from the active basin, yet both the *passive* and *control* (flat-elevated landscape) water treatments received primarily incidental rain.

Soil monitoring system

We installed a meteorological station to measure microclimate variation at the site. We collected local air temperature at 3 meters above ground using a climate sensor (VP-4, METER Group, Inc., Pullman, WA, United States) and rain gauge to measure precipitation (ECRN-100, METER Group, Inc., Pullman, WA, United States). The onsite rain gauge did not work between the 165th and 239th day of the year, and precipitation data from a nearby weather station were used to estimate daily precipitation in the area (“AZMET : The Arizona Meteorological Network : Tucson Station Data Files”). In each treatment, we installed soil moisture and temperature sensors (ECH2O 5TE, METER Group, Inc., Pullman, WA, United States) at 12.5 cm belowground in the center of the treatment, connected to data loggers (EM50, METER Group, Inc., Pullman, WA, United States) collecting hourly data from January 2018 to December 2018 (Buzzard et al., 2021).

Soil hydrogen fluxes

Soil H₂ fluxes were measured using vented static chambers during four campaigns 1) pre-monsoon (June 6th), 2) mid-monsoon (August 8th), 3) late-monsoon (September 12th), and 4) post-monsoon (November 14th), in response to variation in soil moisture throughout the monsoon season and to assess variation in microbial activity across GI treatments. For example, the pre-monsoon campaign was characterized by having high temperatures and low soil moisture; the campaigns during mid-monsoon and late-monsoon season were characterized by high temperatures and high soil moisture; and the post-monsoon campaign was characterized by lower temperatures and intermediate (moist) soil moisture. To estimate H₂ fluxes, four cylindrical PVC collars (20 cm diameter, 20 cm height, surface area = 0.0314 m²) were installed 10 cm into the soil within each treatment one week prior to sampling. All gas sampling was performed between 8:30 and 10:30 a.m. to reduce the effects of high temperatures experienced during the summer months in Tucson, AZ. The mulch layer was removed from the soil collar 2 hours prior to sampling to reduce the effect of different mulch types and depths between treatments and allow a direct assessment of the soil surface. Static vented chambers were placed on the soil collar approximately 2 cm to ensure a proper seal and consistent headspace. Immediately following chamber placement, 5 mL gas samples were collected using air-tight syringes from the sampling port at four time points (0, 5, 10, and 15 minutes). All four treatments were sampled at the same time with randomization of samplers (people) and chambers between each batch and treatment to reduce operator-sampling biases. Syringes were placed in a cooler to maintain temperature and reduce exposure to ultraviolet light until they were transferred to the lab. A total of 64 gas samples were collected during a single morning for each time period, totaling 256 gas samples over the monsoon season. H₂ was measured in parts per billion using a reducing compound photometer (RCP model 910-105 series, Peak Laboratories LLC, CA, USA) gas chromatography (GC) within 24 hr of sampling. Each gas sample was measured in triplicates from 1 ml gas injections. Ultra zero grade air was used as the carrier gas. Chamber height and temperature were measured during field sampling and used to calculate H₂ fluxes for each sampling event based on the following equation:

$$F = (\Delta C / \Delta t) * (P_a V / RT) * (1/A)$$

Where F is the H₂ flux (nmol H₂ m⁻² s⁻¹), $\Delta C / \Delta t$ is the rate of change of the H₂ concentration in the headspace of the chamber through time, P_a is the atmospheric pressure (atm), V is the total chamber volume (L) measured for each chamber and sampling event, R is the gas constant (0.08206 atm L mol⁻¹ K⁻¹), T is the temperature of air inside the chamber (K), and A is the surface area covered by the chamber (0.0314 m²). It is important to note that negative F represents H₂ uptake into the soil, whereas positive F values represent emission of H₂ into the atmosphere, so greater rates of H₂ uptake correspond to more negative soil H₂ flux values.

213 Soil moisture incubation experiment

214 To determine if moisture was the dominant influence on H₂ fluxes, we conducted a soil
 215 microcosm experiment that altered soil moisture levels but with constant temperature. Soil
 216 samples from each batch ($n = 4$) within each treatment ($n = 4$) were collected during the post-
 217 monsoon sampling and were air dried for approximately seven days until they had a base soil
 218 moisture level of roughly 2% gravimetric water content (GWC). GWC was calculated for all air
 219 dried samples and water was added to increase the GWC to the three target levels, dry
 220 corresponded to the low range of observed GWC values (2%), moist is the average range
 221 observed (10%), and wet corresponded to the wet end of the range (20%). Each group
 222 represented conditions observed in the field during the 2018 monsoon season. Soils were then
 223 incubated at 25 °C for 7-8 days and then H₂ fluxes were measured for three different moisture
 224 levels within 24 hours. Moisture levels were maintained during incubation by limiting
 225 evaporative loss by covering vials with parafilm wax. Samples were randomized and vials were
 226 closed using a rubber stopper for roughly 20 minutes where 1ml of air was removed using an air
 227 tight syringe at regular intervals during the 20 minute period to calculate fluxes and directly
 228 injected into the GC-RCP to measure H₂ concentrations. Empty serum vials were used as a
 229 control and measured during the experiment.

230 GC Calibration

231
 232 We used a tank of breathing grade air (Airgas, AI B300) as a working standard for this
 233 study. We estimated the amount of H₂ in the breathing air tank to be 664 ppb H₂ by comparison
 234 to atmospheric H₂ measurements. Specifically, we used the median H₂ peak areas in breathing air
 235 and atmospheric gas measurements and assumed a representative value for typical atmospheric
 236 hydrogen concentration of 530 ppb. For each round of field and lab H₂ measurements, we used
 237 the median of at least 21 breathing air tank peak areas measured over the course of the sampling
 238 event to calculate a GC response factor ($[\text{atmospheric H}_2]/[\text{GC H}_2_peak_area]$). The response
 239 factor was used as a scaling factor for single-point calibration of all unknown samples for each
 240 sampling event. This approach allowed us to account for possible drift in the GC response factor
 241 between measurement days, which ranged from a median of 0.00306 ppb area-1 to 0.00361 ppb
 242 area-1. The response factor varied by 5.4% (stdev) across all sampling periods and maintained a
 243 smaller variation than the uncertainty of the GC-RCP analyzer of 10%.

244 Statistical analyses

245 All statistical analyses and figures were completed in R (R Core Team, 2012). Daily
 246 averages for soil moisture and temperature from each treatment were calculated for the four in
 247 situ flux sampling events. The gvlma package in R was used to assess if the model met statistical
 248 assumptions (Peña & Slate, 2006). Visual inspection of residual plots did not reveal any extreme

outliers or obvious deviations from homoscedasticity or normality for soil H₂ fluxes. We used a two-way mixed analysis of variance (ANOVA) to assess the effects of treatment and season on soil H₂ fluxes. Effect size was determined by generalized eta squared (η^2) with small ($\eta^2 = 0.2$), medium ($\eta^2 = 0.5$), and large ($\eta^2 = 0.8$) following Cohen suggested values (Cohen, 1992). Pairwise comparisons were assessed using Tukey's honest significant difference post hoc methods. Soil H₂ fluxes from the microcosm experiment did not meet statistical assumptions for kurtosis. However, results did not differ between transformed and non-transformed H₂ fluxes data from the microcosm, thus nonparametric one way analyses were completed on non-transformed data. A p-value of < 0.05 with Bonferroni adjustment was used to determine statistical significance. Generalized linear regression was used to assess the individual effect of soil temperature and soil moisture on H₂ fluxes.

Results

Meteorological Conditions

Meteorological data were used to measure local climate variability over the monsoon season (Fig. 1; Table S1). In 2018, we recorded 342 mm of precipitation and an average air temperature of 22.88 ± 6.34 °C at our site. The average daily air temperature in our study site was similar for the first three sampling dates (i.e., pre-monsoon, mid-monsoon, late-monsoon), ranging from 32.2 to 33.6 °C, but was much lower during the post-monsoon sampling (14 ± 3.62 °C). No precipitation was recorded in the week prior to each sampling event, with the exception of the mid-monsoon sampling, which received 6.86 mm of rain the day before sampling. However, prior to that rain event, conditions were relatively dry with no observed precipitation for the previous 10 days. Soil and air temperature followed a similar temporal pattern with the highest temperatures recorded between May and October (Fig. 1A; Tables S1 and S2), and the greatest fluctuation in soil temperature was observed in the control treatment (Buzzard et al., 2021).

In situ Soil Hydrogen Fluxes

We found that the interaction between green infrastructure (GI) treatment and increased wetting events observed during the monsoon season significantly affected soil H₂ fluxes (Fig. 2; Table 1; $F(9, 48) = 2.72$, $p = 0.012$, $\eta^2 = 0.338$). In addition, the mean H₂ fluxes from each treatment differed across the four sampling points, with pre-monsoon and late-monsoon H₂ fluxes statistically different between treatments. During the pre-monsoon sampling period we observed significantly higher H₂ uptake into the soil in the greywater treatment compared to the passive and control treatments (Table S3). Whereas the passive treatment had a significantly lower uptake into the soil compared to the other treatments during the late-monsoon (Table S3).

Soil Microcosm Hydrogen Fluxes

We used a microcosm experiment to test the direct effect of different soil moisture levels on soil H₂ fluxes and did not find a significant difference between treatments (Fig. 3; H(3, 48) = 1.4, $p = 0.706$). However, the mean H₂ fluxes were statistically different between the three soil moisture levels observed for each treatment (Table 1; H(2, 48) = 34.35, $p < 0.001$), with H₂ uptake significantly greater at wet moisture levels (20% gwc) than dry (2% gwc) and moist (10% gwc) soil moisture levels (Table S4).

Abiotic Drivers of Hydrogen Fluxes

Soil temperature and moisture are known drivers of microbial diversity and activity. To test the independent effect of temperature and moisture on H₂ fluxes we assessed the relationship between soil H₂ fluxes from the *in situ* measurements independently with soil temperature and moisture. We found an inverse effect of soil temperature and moisture on soil H₂ fluxes. Soil H₂ uptake decreased with increased temperature ($r = 0.345$, $p = 0.005$, $df = 62$; Fig. 4), and increased with increased soil moisture ($r = -0.393$, $p = 0.001$, $df = 62$; Fig. 4).

Discussion

The primary goal of this study was to examine the interactive effects of green infrastructure (GI) practices and seasonal precipitation on soil H₂ fluxes in a semiarid urban ecosystem. We hypothesized that changes in soil microclimate driven by implementation of GI, specifically changes in soil moisture variability, would lead to increased H₂ uptake in the soils that would be further amplified by seasonal variability in precipitation during the monsoon season. The field observations suggested that soil H₂ uptake was triggered by increased precipitation during the seasonal monsoon across all treatments. In addition, soil H₂ uptake was greater in the greywater treatment compared to the other treatments during pre-monsoon, suggesting that the increased frequency and water availability in the greywater treatment resulted in higher H₂ uptake during the dry season and may support greater microbial consumption of H₂ consistently throughout the year. H₂ uptake was significantly correlated with soil moisture and temperature, which are key environmental factors impacted by both the seasonal precipitation regime and GI water management practices.

The North American Monsoon triggers biological activity by bringing moisture and cooler temperatures to hot-dry soils. However, the large quantities of rain with each storm may lead to flooding and saturation of soils, reducing diffusivity of gases, these factors coupled with lower infiltration rates and soil sealing of urban soils limit or decrease soil H₂ uptake. H₂ uptake into soils is limited by abiotic and biotic processes under dry and wet soil conditions. For example, under dry conditions H₂ diffusion into soils is biotically limited by the presence of dry inactive layers with reduced biological activity in desert soils (Fallon, 1982; Conrad & Seiler, 1985; Smith-Downey, Randerson & Eiler, 2008; Bertagni, Paulot & Porporato, 2021); while wet soil conditions reduce movement into the soils, also leading to H₂ diffusion limitation (Yonemura, Kawashima & Tsuruta, 1999; Yonemura & Kawashima, 2000; Gødde, Meuser &

Conrad, 2000; Ehhalt & Rohrer, 2013). During the pre-monsoon season, H₂ uptake was lower in the control and passive treatments, with greater uptake in the active and greywater GI treatments, consistent with research showing that H₂ uptake even at low moisture levels may promote microbial activity (Fallon, 1982; Conrad & Seiler, 1985; Smith-Downey, Randerson & Eiler, 2006, 2008). Additionally, studies have shown that under very low soil moisture conditions, HOB activity is drastically inhibited (Paulot et al., 2021). As the monsoon season progressed, increased soil moisture resulted in greater H₂ uptake across all treatments, suggesting semiarid urban ecosystems are sensitive to changes in moisture availability and that there is increased uptake throughout the monsoon season as long as there are continual rain events.

As temperatures decreased during the post-monsoon season, we observed median moisture levels and similar soil H₂ uptake across treatments, suggesting an ideal moisture range between 15 and 18% near 15 °C, which is within the ideal temperature range observed for microbial H₂ uptake in the Mojave Desert (Smith-Downey, Randerson & Eiler, 2006). While there are models that robustly assess temperature and moisture effects on H₂ uptake, our results oppose current research that show H₂ uptake increases with increased temperature which may correspond with limitations in our dataset (Ehhalt & Rohrer, 2011, 2013; Yashiro et al., 2011; Bertagni, Paulot & Porporato, 2021). Specifically, our data capture a narrow range of temperatures (12 to 39 °C) compared to the broad range (-20 to 100 °C) modeled in (Ehhalt & Rohrer, 2011) and are missing a critical range between 16 and 27 °C which has been identified as an optimal temperature range of soils with 19% moisture for peak H₂ uptake in the Mojave Desert (Smith-Downey, Randerson & Eiler, 2008). In fact, our results suggest greater variability in H₂ uptake at higher temperatures, with less uptake measured in the passive and control treatments during the driest sampling period (pre-monsoon). Our findings highlight the interaction of temperature and soil moisture as important abiotic drivers of H₂ uptake in semiarid managed systems, with GI altered soil moisture or temperature leading to reduced but sustained H₂ uptake.

The widespread implementation of GI water management practices alter soil moisture legacy by irrigating rain garden basins at different frequencies and providing a more constant water source during the dry season. More frequent and consistent soil moisture events in the greywater compared to the other treatments can lead to shifts in the soil microbiome (Buzzard et al., 2021), nutrient availability, and carbon inputs. Desert soils have low carbon availability, and changes in the water inputs from GI may increase soil carbon by supporting vegetative growth, root development, and increased soil microinvertebrate diversity and abundance (Pavao-Zuckerman & Sookhdeo, 2017). The sustained availability of water and organic resources for energy may further support organoheterotrophic soil microbes, such as *Actinobacteria*, *Proteobacteria*, and *Chloroflexi* (Lynch et al., 2014; Leung et al., 2020; Jordaan et al., 2020), leading to metabolically flexible microbiomes in GI treatments capable of more rapidly switching between resources (Bay et al., 2021a,b). In addition, soil H₂ uptake models project that greater HA-HOB activity is associated with higher temperatures, but can be limited if the

minimum soil moisture threshold is not met (Paulot et al., 2021), and further research should address the uncertainty in HA-HOB activity in arid and semiarid urban ecosystems. As observed in the greywater basin, our findings suggest that a more consistent soil moisture legacy was important for microbial-mediated biogeochemical processes and highlights a sensitivity to increased soil moisture in semiarid urban ecosystems.

Atmospheric trace gases, like H_2 , are important for sustaining soil microbes during periods of low resource availability and may also act to alleviate competition between organoheterotrophs (Fierer, 2017; Bay et al., 2021a,b). Microbial communities with more diverse metabolic strategies may be more resilient to changes in climate and anthropogenic disturbances (Meyer et al., 2004; Allison & Martiny, 2008; Berney & Cook, 2010; Greening et al., 2014). As climate changes, shifts in seasonal precipitation patterns may lead to longer dry periods and more variable precipitation patterns in arid and semiarid regions of the Southwestern United States (Zhang et al., 2021). Although recent work suggests that H_2 uptake may increase with warmer temperatures observed at midlatitudes in the northern hemisphere, altered soil carbon and reduced soil moisture may reduce microbial activity resulting in unknown feedbacks in atmospheric concentrations (Paulot et al., 2021). Here, we show that water management practices, like GI, buffer temperature extremes and reduce periods of drought, increasing H_2 uptake. Our findings suggest that GI managed residential soils can maintain low levels of H_2 uptake year around, above what would be sustained in unmanaged systems (i.e., our control treatment), and may reduce the impacts of drought on H_2 cycling in semiarid urban ecosystems.

Conclusions

Atmospheric H_2 is impacted by changes in climate and increased anthropogenic emissions. Characterizing the changes in microbial mediated soil H_2 uptake has important implications for semiarid urban systems where local land management practices alter the abiotic and biotic environment, increasing uncertainty in the atmospheric hydrogen cycle. Here, we showed that environmental factors impacted by seasonal precipitation and GI water management practices in the semiarid deserts of Arizona increased soil H_2 uptake during seasonally dry periods. As temperatures and drought increase in the Southwestern United States, uptake of atmospheric H_2 by soils may be reduced, unless cities continue to implement GI water management practices, which decrease dry periods and support greater H_2 uptake. Continued investigation into how managed systems impact soil microbial community composition and function is suggested to better understand the distribution and functional role of HA-HOB microbial communities in sustaining H_2 uptake in semiarid urban climates during drought. Our findings highlight the interaction between natural and managed water regimes on soil H_2 uptake as corroborated by field and lab measurements, reinforcing the unique role of H_2 in microbial metabolism in semiarid urban landscapes.

Acknowledgements

We would like to thank Erik Arcos, Andreas Brændholt, and Leslie Dominguez for help with field sampling; and Peter Moma and L. Tate Montgomery for laboratory assistance.

References

- Allison SD, Martiny JBH. 2008. Colloquium paper: resistance, resilience, and redundancy in microbial communities. *Proceedings of the National Academy of Sciences of the United States of America* 105 Suppl 1:11512–11519.
- Austin AT, Yahdjian L, Stark JM, Belnap J, Porporato A, Norton U, Ravetta DA, Schaeffer SM. 2004. Water pulses and biogeochemical cycles in arid and semiarid ecosystems. *Oecologia* 141:221–235.
- AZMET : The Arizona Meteorological Network : Tucson Station Data Files. *Available at* <https://cals.arizona.edu/AZMET/01.htm> (accessed July 5, 2021).
- Bay SK, Dong X, Bradley JA, Leung PM, Grinter R, Jirapanjawat T, Arndt SK, Cook PLM, LaRowe DE, Nauer PA, Chiri E, Greening C. 2021a. Trace gas oxidizers are widespread and active members of soil microbial communities. *Nature Microbiology* 6:246–256.
- Bay SK, Waite DW, Dong X, Gillor O, Chown SL, Hugenholtz P, Greening C. 2021b. Chemosynthetic and photosynthetic bacteria contribute differentially to primary production across a steep desert aridity gradient. *The ISME journal*. DOI: 10.1038/s41396-021-01001-0.
- Berney M, Cook GM. 2010. Unique Flexibility in Energy Metabolism Allows Mycobacteria to Combat Starvation and Hypoxia. *PloS one* 5:e8614.
- Bertagni MB, Paulot F, Porporato A. 2021. Moisture fluctuations modulate abiotic and biotic limitations of H₂ soil uptake. *Global biogeochemical cycles* 35. DOI: 10.1029/2021gb006987.

421 Buzzard V, Gil-Loaiza J, Graf Grachet N, Talkington H, Youngerman C, Tfaily MM, Meredith
 422 LK. 2021. Green infrastructure influences soil health: Biological divergence one year after
 423 installation. *The Science of the total environment* 801:149644.

424 Cohen J. 1992. A power primer. *Psychological bulletin* 112:155–159.

425 Conrad R. 1996. Soil microorganisms as controllers of atmospheric trace gases (H₂, CO, CH₄,
 426 OCS, N₂O, and NO). *Microbiological reviews* 60:609–640.

427 Conrad R, Seiler W. 1979. The role of hydrogen bacteria during the decomposition of hydrogen
 428 by soil. *FEMS microbiology letters* 6:143–145.

429 Conrad R, Seiler W. 1985. Influence of temperature, moisture, and organic carbon on the flux of
 430 H₂ and CO between soil and atmosphere: Field studies in subtropical regions. *Journal of*
 431 *geophysical research* 90:5699.

432 Constant P, Chowdhury SP, Pratscher J, Conrad R. 2010. Streptomycetes contributing to
 433 atmospheric molecular hydrogen soil uptake are widespread and encode a putative high-
 434 affinity [NiFe]-hydrogenase. *Environmental microbiology* 12:821–829.

435 Constant P, Poissant L, Villemur R. 2009. Tropospheric H₂ budget and the response of its soil
 436 uptake under the changing environment. *The Science of the total environment* 407:1809–
 437 1823.

438 Dimmitt MA. 2000. Biomes and communities of the Sonoran Desert region. *A natural history of*
 439 *the Sonoran Desert*:3–18.

440 Ehhalt DH, Rohrer F. 2009. The tropospheric cycle of H₂: a critical review. *Tellus. Series B,*
 441 *Chemical and physical meteorology* 61:500–535.

442 Ehhalt DH, Rohrer F. 2011. The dependence of soil H₂ uptake on temperature and moisture: a
 443 reanalysis of laboratory data. *Tellus. Series B, Chemical and physical meteorology*

63:1040–1051.

Ehhalt D, Rohrer F. 2013. Deposition velocity of H₂: a new algorithm for its dependence on soil moisture and temperature. *Tellus. Series B, Chemical and physical meteorology* 65:19904.

Fallon RD. 1982. Influences of pH, Temperature, and Moisture on Gaseous Tritium Uptake in Surface Soils. *Applied and environmental microbiology* 44:171–178.

Feng S, Fu Q. 2013. Expansion of global drylands under a warming climate. *Atmospheric Chemistry and Physics* 13:10081–10094.

Fierer N. 2017. Embracing the unknown: disentangling the complexities of the soil microbiome. *Nature reviews. Microbiology* 15:579–590.

Gödde M, Meuser K, Conrad R. 2000. Hydrogen consumption and carbon monoxide production in soils with different properties. *Biology and fertility of soils* 32:129–134.

Greening C, Constant P, Hards K, Morales SE, Oakeshott JG, Russell RJ, Taylor MC, Berney M, Conrad R, Cook GM. 2015. Atmospheric hydrogen scavenging: from enzymes to ecosystems. *Applied and environmental microbiology* 81:1190–1199.

Greening C, Villas-Bôas SG, Robson JR, Berney M, Cook GM. 2014. The Growth and Survival of *Mycobacterium smegmatis* Is Enhanced by Co-Metabolism of Atmospheric H₂. *PloS one* 9:e103034.

Grover SPP, Cohan A, Chan HS, Livesley SJ, Beringer J, Daly E. 2013. Occasional large emissions of nitrous oxide and methane observed in stormwater biofiltration systems. *The Science of the total environment* 465:64–71.

Huang J, Yu H, Guan X, Wang G, Guo R. 2016. Accelerated dryland expansion under climate change. *Nature Climate Change* 6:166–171. DOI: 10.1038/nclimate2837.

Huxman TE, Snyder KA, Tissue D, Leffler AJ, Ogle K, Pockman WT, Sandquist DR, Potts DL,

Schwinning S. 2004. Precipitation pulses and carbon fluxes in semiarid and arid ecosystems. *Oecologia* 141:254–268.

Islam ZF, Welsh C, Bayly K, Grinter R, Southam G, Gagen EJ, Greening C. 2020. A widely distributed hydrogenase oxidises atmospheric H₂ during bacterial growth. *The ISME journal* 14:2649–2658.

Jordaan K, Lappan R, Dong X, Aitkenhead IJ, Bay SK, Chiri E, Wieler N, Meredith LK, Cowan DA, Chown SL, Greening C. 2020. Hydrogen-Oxidizing Bacteria Are Abundant in Desert Soils and Strongly Stimulated by Hydration. *mSystems* 5. DOI: 10.1128/mSystems.01131-20.

Khdhiri M, Hesse L, Popa ME, Quiza L, Lalonde I, Meredith LK, Röckmann T, Constant P. 2015. Soil carbon content and relative abundance of high affinity H₂-oxidizing bacteria predict atmospheric H₂ soil uptake activity better than soil microbial community composition. *Soil biology & biochemistry* 85:1–9.

Lee H, Rahn T, Throop HL. 2012. A novel source of atmospheric H₂: abiotic degradation of organic material. *Biogeosciences* 9:4411–4419.

Leung PM, Bay SK, Meier DV, Chiri E, Cowan DA, Gillor O, Woebken D, Greening C. 2020. Energetic Basis of Microbial Growth and Persistence in Desert Ecosystems. *mSystems* 5. DOI: 10.1128/mSystems.00495-19.

Lynch RC, Darcy JL, Kane NC, Nemergut DR, Schmidt SK. 2014. Metagenomic evidence for metabolism of trace atmospheric gases by high-elevation desert Actinobacteria. *Frontiers in microbiology* 5:698.

Maestre FT, Delgado-Baquerizo M, Jeffries TC, Eldridge DJ, Ochoa V, Gozalo B, Quero JL, García-Gómez M, Gallardo A, Ulrich W, Bowker MA, Arredondo T, Barraza-Zepeda C,

- 490 Bran D, Florentino A, Gaitán J, Gutiérrez JR, Huber-Sannwald E, Jankju M, Mau RL,
491 Miriti M, Naseri K, Ospina A, Stavi I, Wang D, Woods NN, Yuan X, Zaady E, Singh BK.
492 2015. Increasing aridity reduces soil microbial diversity and abundance in global drylands.
493 *Proceedings of the National Academy of Sciences of the United States of America*
494 112:15684–15689.
- 495 McPhillips L, Goodale C, Walter MT. 2017. Nutrient leaching and greenhouse gas emissions in
496 grassed detention and bioretention stormwater basins. *Journal of Sustainable Water in the*
497 *Built Environment* 4:04017014.
- 498 Meredith LK, Commane R, Keenan TF, Klosterman ST, Munger JW, Templer PH, Tang J,
499 Wofsy SC, Prinn RG. 2017. Ecosystem fluxes of hydrogen in a mid-latitude forest driven
500 by soil microorganisms and plants. *Global change biology* 23:906–919.
- 501 Meredith LK, Rao D, Bosak T, Klepac-Ceraj V, Tada KR, Hansel CM, Ono S, Prinn RG. 2014.
502 Consumption of atmospheric hydrogen during the life cycle of soil-dwelling actinobacteria.
503 *Environmental microbiology reports* 6:226–238.
- 504 Meyer AF, Lipson DA, Martin AP, Schadt CW, Schmidt SK. 2004. Molecular and metabolic
505 characterization of cold-tolerant alpine soil *Pseudomonas sensu stricto*. *Applied and*
506 *environmental microbiology* 70:483–489.
- 507 Novelli PC, Lang PM, Masarie KA, Hurst DF, Myers R, Elkins JW. 1999. Molecular hydrogen
508 in the troposphere: Global distribution and budget. *Journal of geophysical research*
509 104:30427–30444.
- 510 Noy-Meir I. 1973. Desert ecosystems: Environment and producers. *Annual review of ecology*
511 *and systematics* 4:25–51.
- 512 Paulot F, Paynter D, Naik V, Malyshev S, Menzel R, Horowitz LW. 2021. Global modeling of

hydrogen using GFDL-AM4.1: Sensitivity of soil removal and radiative forcing.
International journal of hydrogen energy 46:13446–13460.

Paul R. Sheppard, Andrew C. Comrie, Gregory D. Packin, Kurt Angersbach, and Malcolm K. Hughes. 1999. *The Climate of the Southwest*. University of Arizona.

Pavao-Zuckerman MA, Sookhdeo C. 2017. Nematode Community Response to Green Infrastructure Design in a Semiarid City. *Journal of environmental quality* 46:687–694.

Peña EA, Slate EH. 2006. Global Validation of Linear Model Assumptions. *Journal of the American Statistical Association* 101:341.

Piché-Choquette S, Constant P. 2019. Molecular Hydrogen, a Neglected Key Driver of Soil Biogeochemical Processes. *Applied and environmental microbiology* 85. DOI: 10.1128/AEM.02418-18.

Prather MJ. 2003. Atmospheric science. An environmental experiment with H₂? *Science* 302:581–582.

Právělie R. 2016. Drylands extent and environmental issues. A global approach. *Earth-Science Reviews* 161:259–278.

Rhee TS, Brenninkmeijer CAM, Röckmann T. 2006. The overwhelming role of soils in the global atmospheric hydrogen cycle. *Atmospheric Chemistry and Physics* 6:1611–1625.

Schmidt U. 1974. Molecular hydrogen in the atmosphere. *Tell'Us* 26:78–90.

Shrestha P, Hurley S, Adair E. 2018. Soil Media CO₂ and N₂O Fluxes Dynamics from Sand-Based Roadside Bioretention Systems. *WATER* 10:185.

Smith-Downey NV, Randerson JT, Eiler JM. 2006. Temperature and moisture dependence of soil H₂ uptake measured in the laboratory. *Geophysical research letters* 33. DOI: 10.1029/2006gl026749.

- Smith-Downey NV, Randerson JT, Eiler JM. 2008. Molecular hydrogen uptake by soils in forest, desert, and marsh ecosystems in California. *Journal of geophysical research* 113. DOI: 10.1029/2008jg000701.
- Wang X, Chen F, Dong Z. 2006. The relative role of climatic and human factors in desertification in semiarid China. *Global environmental change: human and policy dimensions* 16:48–57.
- Yashiro H, Sudo K, Yonemura S, Takigawa M. 2011. The impact of soil uptake on the global distribution of molecular hydrogen: chemical transport model simulation. *Atmospheric chemistry and physics discussions: ACPD* 11:4059–4103.
- Yonemura S, Kawashima S. 2000. Carbon monoxide, hydrogen, and methane uptake by soils in a temperate arable field and a forest. *Journal of geophysical research*.
- Yonemura S, Kawashima S, Tsuruta H. 1999. Continuous measurements of CO and H₂ deposition velocities onto an andisol: uptake control by soil moisture. *Tellus B* 51:688–700. DOI: 10.1034/j.1600-0889.1999.t01-2-00009.x.
- Zhang F, Biederman JA, Dannenberg MP, Yan D, Reed SC, Smith WK. 2021. Five decades of observed daily precipitation reveal longer and more variable drought events across much of the western United States. *Geophysical research letters* 48. DOI: 10.1029/2020gl092293.

Figure 1

Meteorological data measured hourly on site.

(A) Site level air temperature ($^{\circ}\text{C}$) measured at 3 m aboveground. Light grey represents the hourly temperature. Dark grey line represents the smooth fit function from a generalized additive model estimation of air temperature. (B) Daily cumulative sum of precipitation (mm) for at the site. Dashed red lines represent the static chamber hydrogen sampling dates.

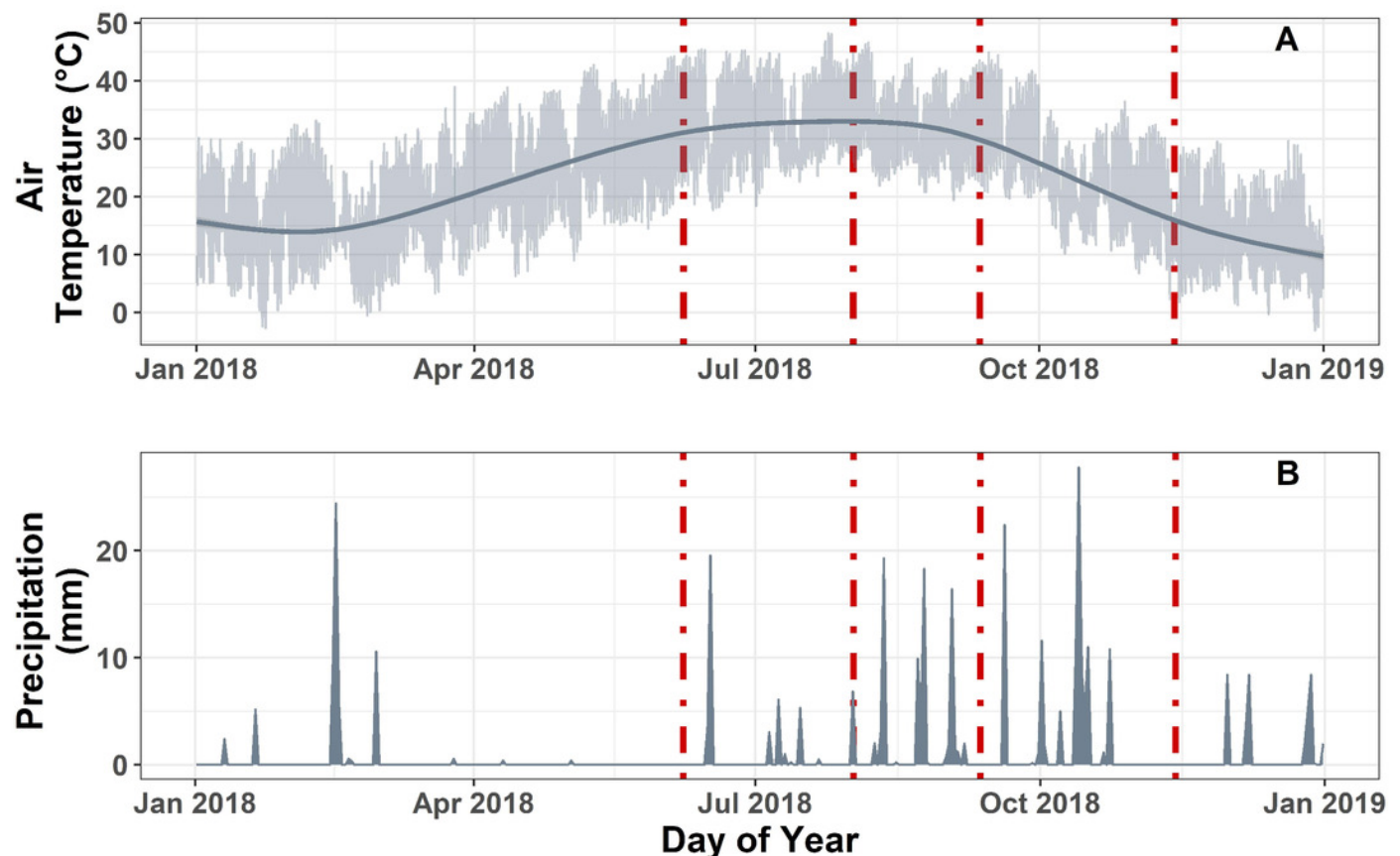


Figure 2

The effects of green infrastructure management and time during the monsoon season on soil hydrogen fluxes ($\text{nmol H}_2 \text{ m}^{-2} \text{ s}^{-1}$).

Boxplots displayed with point distributions for each batch within a treatment. The center line represents the median, and the lower and upper lines correspond to the first and third quartiles (25% and 75% quartiles). Whiskers correspond to the 95% confidence intervals. The two-way ANOVA presented F statistic, p-value and generalized eta squared (η^2). The grey dashed line at 0 on the y-axis represents the transition between soil uptake (negative values) and emission into the atmosphere (positive values).

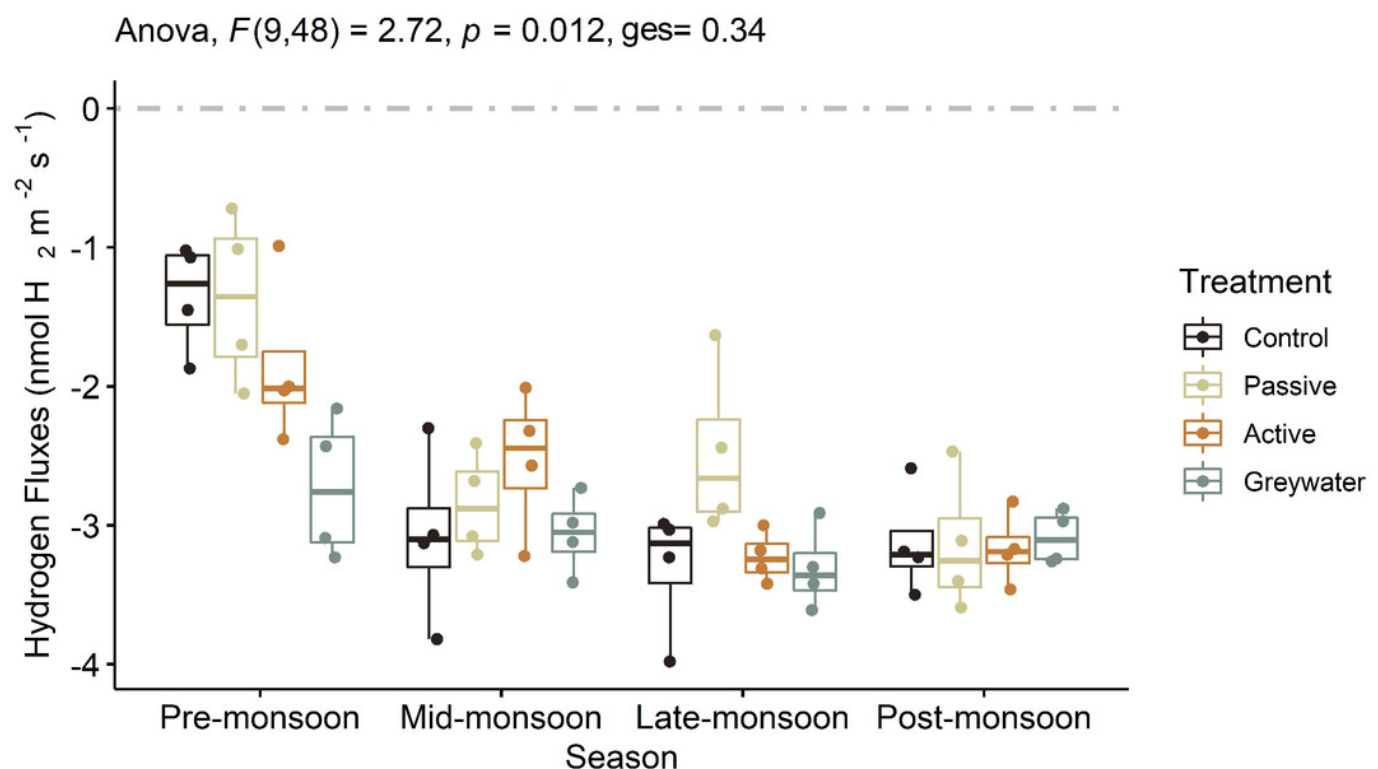


Figure 3

Microcosms assessing the effects of green infrastructure management and soil moisture on hydrogen fluxes ($\text{nmol H}_2 \text{ m}^{-2} \text{ s}^{-1}$).

Boxplots displayed with point distributions for each batch within a treatment. The center line represents the median, and the lower and upper lines correspond to the first and third quartiles (25% and 75% quartiles). Whiskers correspond to the 95% confidence intervals. The one-way nonparametric Kruskal-Wallis ANOVA was performed independently for both treatment and moisture level on H_2 . Corresponding p-values are recorded with treatment first and moisture level second. The grey dashed line at 0 on the y-axis represents the transition between soil uptake (negative values) and emission into the atmosphere (positive values).

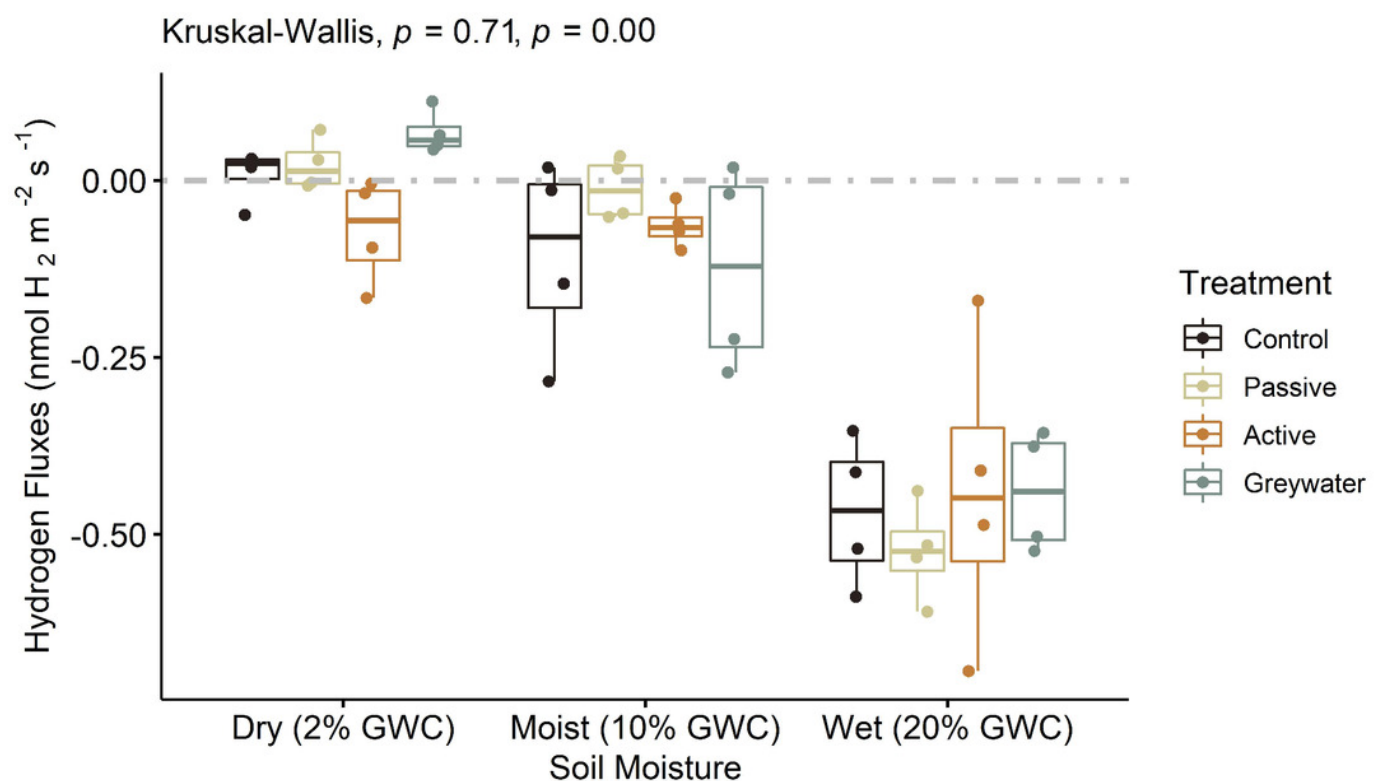


Figure 4

Effect of selected environmental drivers during in situ measurements of soil hydrogen fluxes ($\text{nmol H}_2 \text{ m}^{-2} \text{ s}^{-1}$).

(A) Positive correlation of hydrogen fluxes and soil temperature $^{\circ}\text{C}$ shows H_2 uptake into the soil is greater at lower temperatures, and (B) Negative correlation between hydrogen fluxes and soil moisture indicates that H_2 uptake increases with increased moisture. Boxplots display the median, 25th and 75th interquartile range, and whiskers show 1.5 times the interquartile for soil temperature (top left), soil moisture (top right), and soil hydrogen fluxes (right side) averaged during the in situ sampling period. Negative H_2 fluxes indicate uptake from the atmosphere into the soil.

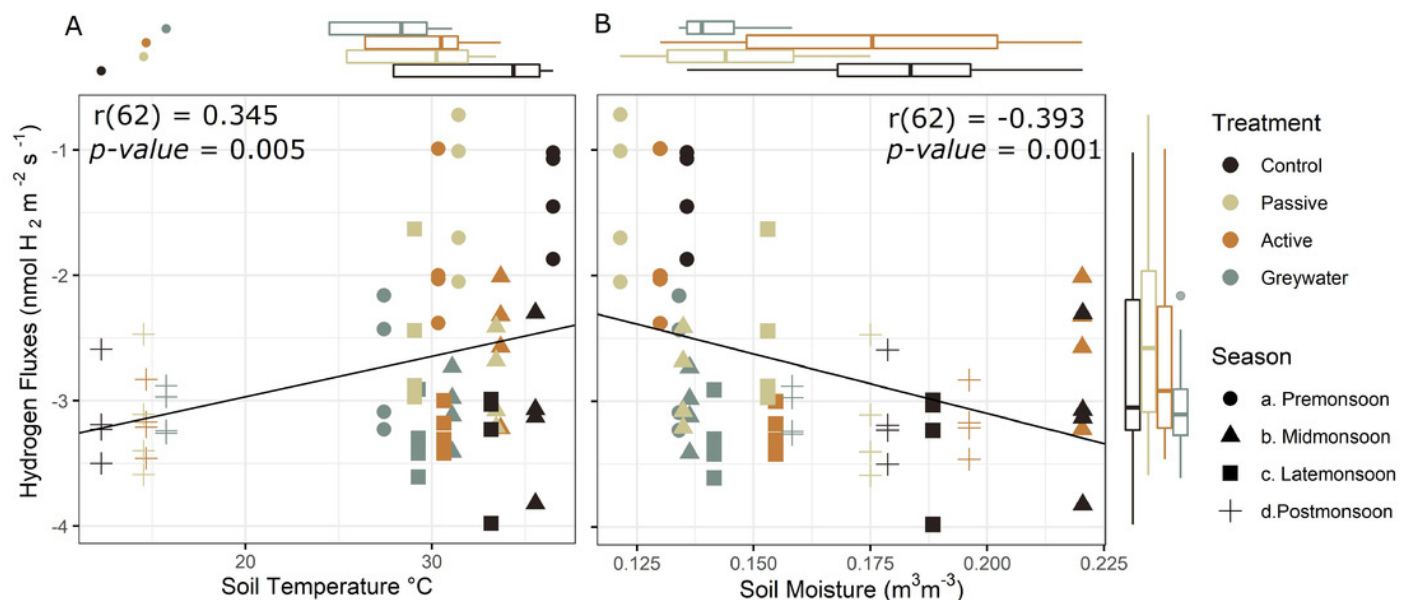


Table 1(on next page)

Analysis of variance (ANOVA) assessing the effects of green infrastructure management and soil moisture on hydrogen fluxes ($\text{nmol H}_2 \text{ m}^{-2} \text{ s}^{-1}$).

Two-way ANOVA for assessing hydrogen fluxes as a function of treatment and season for in situ field sampling. One-way nonparametric Kruskal-Wallis ANOVA independently assessing hydrogen fluxes as a function of treatment and moisture level for microcosm lab experiment.

DFn: Degrees of freedom in the numerator. DFd: Degrees of freedom in the denominator.

Test Statistic: F for two-way parametric ANOVA; H for one-way nonparametric Kruskal-Wallis test.

Experiment Location	Effect	DFn	DFd	Test Statistic	p-value	p<.05	Effect Size
Field	Treatment	3	48	4.64	0.006	*	0.225
Field	Season	3	48	29.85	0.000	*	0.651
Field	Treatment:Season	9	48	2.72	0.012	*	0.338
Lab	Treatment	3	48	1.40	0.706		-0.036
Lab	Moisture Level	2	48	34.35	0.000	*	0.719