

**Seasonal and diel variations in the vertical distribution,
composition, abundance and biomass of zooplankton
in a deep Chilean Patagonian fjord**

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Abstract

Comau Fjord is a stratified fjord in the northern part of Chilean Patagonia. In spite of aragonite-undersaturated waters causing an energetic burden to coral calcification, this deep body of water harbors dense populations of cold-water corals (CWC). While the paradox has been attributed to a rich supply of zooplankton, quantitative data to support this hypothesis are so far lacking. In this study, we investigated the seasonal and diel changes of the zooplankton community over the entire water column. We used a Nansen net (100 μm mesh) to take stratified vertical hauls between the surface and the bottom (0-50-100-200-300-400-450 m). Samples were scanned with a ZooScan, and abundance, biovolume and biomass were determined for 41 taxa identified on the web-based platform EcoTaxa 2.0. Zooplankton biomass was the highest in summer (209 g dry mass m^{-2}) and the lowest in winter (61 g dry mass m^{-2}). Abundance, however, peaked in spring, suggesting a close correspondence between reproduction and the phytoplankton spring bloom (Chl *a* max. 50.86 mg m^{-3} , 3 m depth). Overall, copepods were the most important group of the total zooplankton community, both in abundance (64-81%) and biovolume (20-70%) followed by mysids and chaetognaths (in terms of biovolume and biomass), and nauplii and Appendicularia (in terms of abundance). Throughout the year, diel changes in the vertical distribution of biomass were found with a daytime maximum in the 100-200 m depth layer and a nighttime maximum in surface waters (0-50 m), associated with the diel vertical migration of the calanoid copepod genus *Metridia*. In addition, our study provides a good background knowledge for the understanding of the ecological connection between zooplankton and CWC. Based on our zooplankton data, we calculated that a zooplankton supply rate of 790 prey $\text{polyp}^{-1} \text{day}^{-1}$ would be necessary to sustain a viable population of CWC in Comau Fjord. Therefore, the high abundance of CWC in the aragonite-undersaturated waters of Comau Fjord may be due to the high food availability provided by the zooplankton biomass.

Keywords. zooplankton, seasonality, diel vertical migration, cold-water corals, ZooScan, EcoTaxa, Comau Fjord, Chilean Patagonia.

Introduction

Coastal marine ecosystems are among the most productive on earth (Mann & Lazier, 1991). They provide substantial economic and ecological services, such as high biological production, nutrient cycling or shoreline stability and erosion control (Escribano et al., 2003; Liu et al., 2010; Pan et al., 2013; Barbier, 2017). The Chilean Fjord Region extends over large and complex hydrographic and geomorphologic conditions, particularly rich in productivity and biodiversity (Försterra et al.,

2017; Häussermann et al., in press), but also endangered by human exploitation, which has been increasing significantly – e.g. salmon aquaculture – over the last two decades (Iriarte et al., 2010; Buschmann et al., 2021; Navedo and Vargas-Chacoff, 2021).

The hydrography of the fjords and channels of Patagonia can be considered a transitional estuarine-marine system where a surface layer of silicate-rich terrestrial freshwater meets nitrate- and phosphate-rich marine waters. These fjords receive freshwater from rivers, surface runoff and groundwater flow due to the high rainfall and glacier melting (Pantoja, Iriarte & Daneri, 2011). The upper brackish layer within the top 10 m water depth is usually poor in nitrate and phosphate but rich in silicate and organic matter from terrestrial inputs (Sánchez, González & Iriarte, 2011). Below the halocline, a water mass with higher salinity (>31), named Modified Subantarctic Water (MSAAW), flowing landward from the adjacent oceanic area provides the fjords with macronutrients (nitrate and phosphate). Both water masses, surface-freshwater and MSAAW, generate a two-layer structure in the water column with sharp vertical and more gradual horizontal salinity gradients (Sievers and Silva, 2008; Pérez-Santos et al., 2014; Meerhoff et al., 2019). The vernal stratification creates a barrier which may hinder the exchange of nutrients within the water column and alter the functioning of the pelagic food web and productivity patterns (Silva et al., 1997; González et al., 2011). During austral spring, the Comau Fjord receives an intense riverine input of fresh water, rich in silicic acid used by bloom-generating diatoms and, thus, leading to high primary production (González et al., 2010). In austral summer, the high concentration of phytoplankton promotes an increase in the abundance and biomass of zooplankton as secondary producers (Antezana, 1999; González et al., 2010). In fjord systems, seasonal patterns are modulated by other oceanographic processes, such as estuarine circulation (Palma & Silva, 2004), tidal regimes and lateral advection (Castro et al., 2011) or water column stratification (Sánchez, González & Iriarte, 2011) influencing the zooplankton biomass and community structure on shorter time scales.

Zooplankton plays an essential role in the functioning of marine ecosystems and in the oceanic carbon cycle. It includes a wide variety of organisms and displays extreme variability in terms of community composition and vertical, seasonal and geographical distribution (Palma & Kaiser, 1993). Many taxa are known to perform diel vertical migrations (DVM), most likely to evade predators (Stich and Lampert, 1981; Iwasa, 1982). According to the predator-evasion hypothesis, migrating zooplankton resides in deep waters during daytime hours where the probability of being perceived by visually orientated predators is lower than if they remained in more illuminated shallow waters, and at night, in the refuge of darkness, they migrate upwards to feed. However, DVM is not performed by all organisms in a zooplankton community or even not by all individuals of one species. For example, while late copepodites and adults of the copepod genus

89 *Metridia* migrate, a large fraction of the young developmental stages remains in surface waters,
90 saving the energy of performing the DVM, suggesting a lower probability of being perceived and
91 consumed by visual predators (Hays, 1995). Both, migrating and resident species are important
92 elements of the biological carbon pump via the production of sinking fecal pellets that transport
93 carbon from the surface to the seafloor (Urrère and Knauer, 1981; Fowler and Knauer, 1986;
94 Emerson and Roff, 1987). Zooplankton also provides a trophic link between primary production
95 and higher consumers such as fish, birds and mammals, but also invertebrate predators, such as
96 corals (Nemoto, 1970; Gili et al., 2006; Höfer et al., 2018).

97 Cold-water corals (CWC) depend on zooplankton as their principal food source to maintain their
98 physiological processes, such as respiratory metabolism and growth (Carrier et al., 2009; Mayr et
99 al. 2011, Naumann et al. 2011). Therefore, their diet is susceptible on the zooplankton seasonal,
100 diel and vertical distribution. In Comau Fjord, azooxanthellate scleractinian CWC are wide-
101 spread despite aragonite-undersaturated waters (Häussermann and Försterra, 2007; Fillinger and
102 Richter, 2013; Jantzen et al., 2013). In aragonite-undersaturated waters, the dissolution of exposed
103 parts of the skeleton, enhanced bioerosion, and reduced CWC growth and survival have been
104 observed (McCulloch et al., 2012; Maier et al., 2016). The calcification of the CWC skeleton is
105 energetically costly, and thus food requirements in these adverse environments are higher
106 compared to aragonite-saturated waters. According to Fillinger & Richter (2013a), in Comau
107 fjord, the CWC *Desmophyllum dianthus* thrive but coral densities decrease below 280 m despite
108 available substrate, suggesting that a shortage of zooplankton could be limiting coral growth.
109 However, up to now little is known about the zooplankton of Comau Fjord in the northern
110 Patagonian region and its role in sustaining the rich CWC communities living in the fjord. Most
111 studies were carried out in the central-southern part of Patagonia (from Penas Gulf to Cape Horn,
112 S 46.50° – S 55.55°), while studies performed in the northern area (from Puerto Montt to San
113 Rafael Lagoon, S 41.20° – S 46.40°) mainly focused on selected microzooplankton taxa in the
114 upper water column, on bulk measurements of zooplankton biomass, or on the carbon flow
115 through the pelagic food web (e.g., Palma, 2008 and references therein; Villenas et al., 2009;
116 González et al., 2010; González et al., 2011; Palma et al., 2011; Sánchez et al., 2011). Other
117 studies addressed the physical oceanographic processes and their effect on zooplankton
118 distribution (e.g., Marín and Delgado, 2009; Castro et al., 2011) and, more recently, investigated
119 zooplankton migration patterns by acoustic backscatter and vertical velocity profiles (Valle-
120 Levinson et al., 2014; Díaz-Astudillo et al., 2017; Pérez-Santos et al., 2017). The information on
121 zooplankton diversity and migration patterns is, however, still very fragmentary and the linkage
122 between the abundance of CWC and zooplankton supply in Comau Fjord remains unknown.

Commenté [A1]: Any reference? Not sure whether this assertion is based on literature data or is an hypothesis?

Commenté [A2]: To me it is not exactly what these authors reported. Fillinger & Richter 2013a "only speculate that the lower oxygen concentration, combined with a lower pH, may entail higher metabolic expenditures for the corals" and further speculate that "The likely reduced water exchange may in addition limit the supply of food to *D. dianthus*", consisting of zooplankton... So your assertion is a bit of a shortcut.

123 In this study, we aim to (i) describe the diel, vertical, and seasonal distribution of
124 mesozooplankton groups of Comau Fjord, with a focus on the dominating taxa, particularly those
125 that migrate and are more likely to aggregate; and (ii) investigate if the zooplankton densities
126 retained in the fjord are sufficient to sustain the population of CWC. Samples were collected day
127 and night with net hauls through the whole water column in spring, summer, autumn and winter.
128 They were processed with a high-resolution image analysis system (ZooScan, Gorsky et al. 2010).
129 The zooplankton taxa were identified using EcoTaxa 2.0 (Picheral, Colin & Irisson, 2017),
130 allowing to assess the influence of the seasonal environmental changes on the zooplankton
131 dynamics, and the natural food available to CWC.

132

133 **Materials & Methods**

134 Field work was carried out in Comau Fjord, Northern Patagonia, Chile (Fig. 1). Zooplankton was
135 sampled four times at a fixed station (24°14.95S, 72°28.83W) in central Comau Fjord: in austral
136 spring (28th September 2016), summer (17th January 2017), autumn (7th May 2017), and winter
137 (29th July 2017), during both: day and night. Samples were collected with a 70 cm-diameter
138 Nansen closing net (mesh size: 100 µm) equipped with a non-filtering cod end. Vertical hauls
139 were carried out at 0.45 m s⁻¹ to sample the depth strata 0-50-100-200-300-400-450 m.
140 Immediately after the collection, the samples were sieved through a 50 µm mesh and preserved in
141 4% borax-buffered formaldehyde for laboratory analyses. After every zooplankton haul, a CTD
142 multi-probe (SBE 19plusV2 Profiler - with RS 232 Interface, Sea-Bird Electronics Inc.) was
143 deployed from the surface to the bottom, measuring conductivity, temperature, oxygen, pH and
144 Chl *a*-fluorescence (Fig. 2).

145 **Sample analysis and image processing**

146 In the laboratory, zooplankton samples were washed with fresh water and prepared for analysis
147 with a ZooScan digital imaging system (Grosjean et al., 2004, Gorsky et al., 2010). ZooScan
148 (CNRS patent, www.hydroptic.com) provides a quick and reliable method for the analysis of
149 preserved plankton samples, storing digitized images for later examination, reprocessing and
150 dissemination. Concentrated samples were subsampled with a Folsom plankton splitter to avoid
151 cluttering the images with more than approximately 1000-1500 individuals. Up to six binary
152 splitting steps were carried out (corresponding to a minimum 1/64th fraction of the original
153 sample). Routinely, the two final splits were scanned with ZooScan yielding images of 2400-dpi
154 resolution (14200×22700 pixels). The hinged base of the ZooScan allowed the recovery of the
155 complete undamaged subsample, which was later stored in 70% ethanol for archiving. Most

Commenté [A3]: Should be 42 not 24? (Fig. 1)

Commenté [A4]: Could you give the hours of sampling?

Commenté [A5]: This is the only subsection in the Mat & Met. So either remove or add a first one such as "sampling", the second being analysis and image processing.

Commenté [A6]: Not sure about the term...

156 overlapping individuals on the scanning surface were manually separated to ensure an even
 157 distribution before scanning. Image analysis was performed with the software ZooProcess
 158 (Gorsky et al., 2010), a plug-in for the image processing and analysis software ImageJ (Schneider,
 159 Rasband & Eliceiri, 2012). The processing involved (1) the automatic subtraction of background
 160 noise, (2) the automatic thresholding and detection of objects, and (3) the automated storage of
 161 detected objects in separate images (“vignettes”). ZooScan detection limit was set at the standard
 162 of 300 µm. Detected zooplankton organisms ranged from 0.3 to 59 mm. Automatic processing
 163 was successful in 75-80% of the cases. Despite the manual separation, some individuals
 164 overlapped, resulting in vignettes with two or more objects. These objects were manually
 165 separated using the “separation with mask” tool of the ZooProcess software. Separated vignettes
 166 were stored, while the original vignette containing multiple objects was eliminated from the
 167 database to avoid duplicate counts. In some cases, the separation of individuals was not possible
 168 as cutting the vignette would mean losing information about the taxonomy of the organisms (i.e.
 169 cutting overlapping urosomes from two different copepods or small copepods embedded in
 170 cnidarians). Overall, the contribution of vignettes with multiple objects was always < 10% of the
 171 total amount of vignettes.

172 Vignettes were subjected to the semi-automated taxonomic classification in EcoTaxa 2.0
 173 (Picheral, Colin & Irsson, 2017). This web-based machine learning application uses training sets
 174 of expert-identified taxa and random forest classification to automatically identify and sort the
 175 objects. Although EcoTaxa contains more than 160 million objects on its server, no ZooScan
 176 training set was available for Patagonian waters. Therefore, manual identification of individuals
 177 on a subset of the images was first necessary to train an initial model, which was later used by the
 178 system to predict the taxonomic classification of the scanned organisms. After the initial
 179 prediction, the automatic classification had an accuracy rate of 44%. We therefore manually
 180 validated the classifications of the initial learning set by sorting more objects into the given
 181 categories. This produced the final learning set for the classification of the entire image data set.
 182 At the end, all classified objects were validated by an expert to assure a correct taxonomic
 183 classification. The organisms were classified to the lowest possible taxonomic level; for most
 184 copepods this was the genus level. However, small calanoid copepods (<1.5 mm) were not
 185 distinguishable on genus level and were comprised as one category: “Calanoids (<1.5 mm)”
 186 including five groups (copepodites (all calanoid taxa, <1mm), *Clausocalanus*, *Microcalanus*,
 187 *Neocalanus* and *Paracalanus*). The category Cnidaria was constituted by organisms from the
 188 class Hydrozoa (mostly medusa and Siphonophorae). Another category contained all images that
 189 were out of focus (“bad focus”) and likely comprised individuals from all copepod taxa. From the
 190 total of 83,516 vignettes, 23,227 could not be assigned to zooplankton taxa, but were labelled as

Commenté [A7]: Probably not easy with 100 µm mesh size. See following sentence: ZooScan detection limit was set at the standard of 300 µm. What does this mean ? Measurement of all objects then removing small ones?

Commenté [A8]: Not clear.

Commenté [A9]: What does this mean?

Commenté [A10]: Does this mean that some individuals smaller than 300 µm could be included here?

Commenté [A11]: Do you mean all or only those that could not be separated?

Commenté [A12]: Which means? The manually annotated vignettes were only identified with 44% success? At what taxonomic level? The success probably depends on this.

Commenté [A13]: What is not clear here is whether your training set was the global one + Patagonian one, or only the Patagonian one.

Commenté [A14]: How many?

191 “detritus”, “fiber”, “leg” “bubble” and “other” and were not considered in our analyses.
 192 ZooProcess provides information about the length and width of each object, allowing the
 193 calculation of its volume as a proxy for its biomass (Gorsky et al., 2010). The program
 194 automatically fits an ellipse around the object, from which the major and minor axis and volume
 195 (V) is computed:

$$196 \quad V (mm^3) = \frac{4}{3} \times \pi \times \frac{major\ axis\ (mm)}{2} \times \left(\frac{minor\ axis\ (mm)}{2} \right)^2$$

197 Biovolume (BV) was then calculated as the sum of the volumes of all objects (ΣV) divided by the
 198 fraction of the sample (e.g., $F=1/64$) and by the volume filtered by the Nansen net (V_N):

$$199 \quad BV (mm^3/m^3) = \frac{\frac{\Sigma V (mm^3)}{F}}{V_N (m^3)}$$

200 V_N was calculated as:

$$201 \quad V_N (m^3) = \left[\pi \times \left[\frac{net\ diameter\ (m)}{2} \right]^2 \right] \times depth\ interval\ (m) \times filtration\ efficiency$$

202 where filtration efficiency was estimated as the theoretical 100% efficiency (value = 1).

203 For the estimation of biomass, a regression between the dry mass of a specimen and its body area
 204 [DM (μg) = ($a A^b$)] was used (Hernández-León and Montero, 2006; Lehet and Hernández-León,
 205 2009), where A is the area (mm^2) of each individual. The conversion of ZooScan-based
 206 biovolume into dry mass required different conversion factors depending on the organism, as for
 207 instance, gelatinous zooplankton with high water content may not be compared to crustaceans or
 208 echinoderms (Table 1). Such coefficients have been successfully published in previous studies for
 209 mid-latitude shelf areas (Marcolin, Gaeta & Lopes, 2015) or Chilean upwelling region (Tutasi &
 210 Escribano, 2020).

211 The biomass (B) of each taxon was then calculated as the sum of the individual dry masses of the
 212 respective taxon (ΣDM) divided by the fraction of the sample (e.g., $F=1/64$) and by the volume
 213 filtered by the Nansen net (V_N):

$$214 \quad B (mg\ dry\ mass/m^3) = \frac{\frac{\Sigma DM(mg)}{F}}{V_N(m^3)}$$

215 Biovolume and biomass were calculated to obtain the sum of the values of all individuals for a
 216 given taxon. In multiple vignettes, the automatic calculation of biovolume and biomass was not

Commenté [A15]: No flowmeter? Not possible for strata?

Commenté [A16]: Not clear what you did... You did not convert volume into dry mass, because you use area, given by the ZooScan. No?

possible because of overlapping specimens from different taxa. Then, the organisms were counted manually and biovolume and biomass were estimated by multiplying the mean volume or DM of the given taxon from all automatic calculations by the extra number of multiple vignettes. Groups with large size variability (e.g., chaetognaths, cnidarians or Euchaetidae) were divided into two categories (big and small) in order to get a better biovolume/biomass assessment.

Commenté [A17]: Please detail: what ranges?

The integrated values of abundance, biovolume and biomass (Fig. 3), were calculated down to 400 m water depth, taking out the last 50 m from summer and winter, to make it comparable to the spring and autumn seasons, where samples were collected down to that specific depth.

Commenté [A18]: Please do not indicate here as it is a result.

225

Results

The physicochemical parameters measured in the water column of Comau Fjord showed stronger seasonal variability in surface waters (0-50 m) than in the deep (50-450 m) (Fig. 2). The temperature profile indicated summer stratification, followed by surface cooling, breakdown of the thermocline in autumn, and reverse temperature gradients in winter and spring. Accordingly, the surface temperature values were the lowest in winter and spring (8.6-11 °C), and the highest in summer (16.7 °C), getting cooler again in autumn (12 °C). In deeper waters, temperatures were more stable with an average value of 11.4 ± 0.2 °C (mean average \pm sd; Fig. 2A). Salinity was between 10-30 in the upper 20 m and 32.9 ± 0.4 below 20 m (Fig. 2B). The pH ranged between 8.5 and 7.7 in the upper 50 m and was 7.9 ± 0.1 in deeper waters in all seasons, except for autumn, where we interpret the sudden drop of pH values as an instrument malfunction (Fig. 2C). Oxygen concentration showed the largest variations in the upper 50 m during the spring season (137.2-410.5 $\mu\text{mol kg}^{-1}$), while below 100 m depth it was on average 180 ± 9.3 $\mu\text{mol kg}^{-1}$ (Fig. 2D). The chlorophyll *a* (Chl *a*) concentration peaked in early spring (50.86 mg Chl *a* m^{-3} at 3 m depth), followed by a decrease towards the end of the summer and low values through autumn and mid-winter (2.5-5 mg Chl *a* m^{-3} at 5-10 m depth) (Fig. 2E). Below 25 m, the concentration of Chl *a* was $< 1.8 \pm 0.5$ mg m^{-3} throughout the year (Fig. 2E).

Commenté [A19]: Please give the depth of the thermocline

Commenté [A20]: What difference do you place in the two terms?

Commenté [A21]: Is it a common unit? Not $\mu\text{mol L}^{-1}$?

The zooplankton community exhibited large seasonal and diel differences. Abundance, integrated over the upper 400 m of the water column (individuals m^{-2}) showed the highest values in spring and the lowest during autumn (Fig. 3A). Interestingly, the seasonal-integrated biovolume ($\text{cm}^3 \text{m}^{-2}$) and biomass (g dry mass m^{-2}) showed a different pattern with the highest values in summer and the lowest in autumn and winter (Fig. 3B, C). Diel differences in integrated abundances generally showed higher values during the day than at night, except for autumn (Fig. 3A). Diel differences in integrated biovolume and biomass were surprisingly large, with generally higher values during

Commenté [A22]: Maybe you can add "for both day and night".

Commenté [A23]: Unnecessary in the results. Why is it interesting?

250 the night, except for spring (Fig. 3B, C), indicating that during daytime, particularly in spring,
251 zooplankton was more numerous but smaller in size.

252 Zooplankton vertical distribution also showed pronounced seasonal and diel changes.
253 Zooplankton abundance exhibited the highest values in the 0-50 m layer during day and night at
254 all seasons, with the exception of autumn during day time. In autumn, the day-time peak was
255 absent with overall low values throughout the water column (Fig. 4A). During day and night time,
256 the lowest abundances were found in >300 m depth in spring and summer, and in 50-100 m depth
257 in autumn. In winter, the abundance was the lowest in 200-300 m in day time, and below 200 m
258 in night time (Fig. 4A). During the day, the highest values of biovolume (cm^3m^{-3}) and biomass (g
259 dry mass m^{-3}) were observed in 0-50 m and 100-200 m water depth in spring and summer, and in
260 100-300 m in autumn. In winter, day time biovolume and biomass were similarly low over the
261 entire water column (Fig. 4B, C). At night, the highest biovolume and biomass values were found
262 at the surface 0-50 m in all seasons, indicating that the different zooplankton taxa that reside in
263 the 100-200 m water depth performed upward vertical migration. The lowest biovolume and
264 biomass values were found in >300 m depth in spring and summer, in 50-100 and 200-300 m
265 depth in autumn and in >50 m depth in winter (Fig. 4B, C).

266 The taxonomical analysis composition showed strong differences between the biovolume and
267 abundance of zooplankton groups (Fig. 5, 6A; biovolume—not biomass—was used to better
268 represent the gelatinous taxa). Copepoda were the most important group, constituting seasonally
269 20-70% of the total biovolume, and 64-81% of the total abundance. Within the copepod
270 community, individuals smaller than 1.5 mm included (a) copepodites and adults of small
271 calanoid genera, such as *Clausocalanus*, *Microcalanus*, *Neocalanus* and *Paracalanus*; (b)
272 cyclopoid copepods of the genera *Oithona* and *Oncaea*; (c) harpacticoid copepods; and (d)
273 crustacean nauplii (mostly from copepods). Small copepods accounted for 58-86% of the total
274 copepod community. Overall, 14 out of 41 taxa contributed 45-98% of the total biovolume and
275 45-86% of the total zooplankton abundance (Fig. 5). The other 27 taxa were constituted by other
276 copepod taxa (Acartiidae, Aetideidae, Calanoida (non-identifiable), Candaciidae, Centropagidae,
277 Copepoda (non-identifiable), Eucalanidae, Heterorhabdidae, Lucicutiidae, Oncaeidae, Pontellidae,
278 Rhincalanidae, Actinopterygii (eggs and larvae), Appendicularia, Ascidiacea (larvae),
279 Brachiopoda (larvae), Bryozoa (larvae), Cirripedia, Cladocera, Decapoda (zoea), Echinodermata
280 (larvae), eggs, Isopoda, Nemertea (pilidium), Platyhelminthes (larvae), Polychaeta (larvae) and
281 non-identifiable organisms.

282 The most important groups differed regarding biovolume and abundance. For biovolume, Mysida
283 (1-21%) constituted a strong large part of the zooplankton community, followed by Chaetognatha

Commenté [A24]: Overall I think this paragraph is too long and detailed, as everything is in the Fig. 4. Could you try shorten it, keeping only the key results?

Commenté [A25]: Units are useless here.

Commenté [A26]: This is an interpretation and should not be in the results section but in the discussion. At this stage in the paper, it is a bit a very short interpretation that would be discussed, so I suggest to remove and keep for the discussion. Further, at this stage you did not look at the taxonomic composition, so it seems difficult to talk about the "different zooplankton taxa"...

Commenté [A27]: I find this sentence poorly said. If the sense is the same, I suggest: "Looking at the taxonomic composition, the relative contribution of the taxa greatly differed between biovolume and abundance."

Commenté [A28]: Those < 1.5 mm? If yes, it should be clearly stated, for example "these small copepods"

Commenté [A29]: Abundance? If yes clearly state it

Commenté [A30]: It is really not clear if the small copepod above are part of these 14.

Commenté [A31]: At several places you mention the Euchaetidae, but they are not included here. Why? From Fig. 5 they should be in the 14...

Commenté [A32]: This is exactly what you already stated line 266-267. It seems to me that you talk about the 14 taxa most important. True?

284 (3-15%) and Cnidaria (2-23%). Metanauplii and calyptosis stages of Euphausiacea were found
 285 mostly in spring and summer accounting for 0-7% of the total biovolume and when one adult
 286 specimen was found during winter in 0-50 m water depth at night, the total biovolume of this
 287 taxon increased to 65%. Regarding the abundancenumber of specimens, crustacean nauplii (2-9%
 288 with its maximum in spring and minimum in autumn) and Appendicularia (0.4-6%) were the
 289 second and third most abundant groups after Copepoda, respectively. The fourth most abundant
 290 group differed among seasons: Echinodermata larvae (5% in spring), Mollusca larvae (2-5% in
 291 summer, 3-9% in autumn), Ostracoda (4-5% in autumn) and Bryozoa larvae (2-10% in winter). In
 292 Across all samples and seasons, taxa that represented more than 5% of total biovolume were
 293 Cnidaria (13%), *Calanus* (12.9%), Mysida (12.7%), *Metridia* (12.6%), Chaetognatha (9.6%) and
 294 Euchaetidae (8.7%). The most abundant groups with more than 5% of the total abundance were
 295 Harpacticoida (14.0%), Cyclopoida (9.1%) and *Metridia* (8.4%) (Fig. 5, 6A).

Commenté [A33]: Better to use the terms as they appear in the other parts.

296 Seasonally, The integrated abundance of the most dominant taxa followed the phytoplankton
 297 cycle for small and large copepods and chaetognaths, with high values in spring to generally low
 298 values in autumn and winter (Table 2). Highest abundances of crustacean nauplii and cnidarians
 299 were found in early spring with a minimum in autumn and raising up again in late-winter. Mysida
 300 presented a stable abundance over the seasons with a minimum in summer. These groups
 301 presented clearly different vertical distributions (Fig. 6). *Metridia*, a large and important copepod
 302 with regard to both biovolume and abundance, resided generally above 200 m, exhibiting a peak
 303 between 100-200 m during the day and a bimodal distribution (peaks at 0-50 m and 100-200 m)
 304 during the night (Fig. 6B). The largest individuals ($0.62 \pm 0.3 \text{ mm}^3$; average size \pm sd) were found
 305 in the intermediate waters during the day, but in spring, a significant proportion of the *Metridia*
 306 population was also found during the day in shallow waters where smaller individuals ($0.14 \pm$
 307 0.08 mm^3) were identified. The copepod *Calanus* was found mainly in intermediate waters (100-
 308 300 m) during day-time where the largest individuals ($1.37 \pm 0.5 \text{ mm}^3$) resided, whereas at night
 309 part of the population migrated upwards and the other part remained at depth. Small specimen of
 310 *Calanus* ($0.30 \pm 0.003 \text{ mm}^3$) were found in shallow waters also during daytime in spring. At
 311 night, only in spring and winter, individuals were present in shallow waters (<50 m depth).
 312 Despite the low number of *Calanus* specimens found at shallow waters, they contributed
 313 considerably to the biovolume in these layers (Fig. 6C). The Euchaetidae showed higher
 314 abundance and biovolume in the deeper part of the water column during day time, ascending to
 315 shallower waters at night (Fig. 6I). Small copepods of the taxa Harpacticoida and Cyclopoida
 316 were very abundant, but, as expected, they only represented a small fraction of the total
 317 biovolume. Harpacticoida were mainly abundant below 50 m during both day and night time, and
 318 their abundance increased with depth (Fig. 6D). Cyclopoida were mainly present in the upper 200

Commenté [A34]: I suggest you rephrase this sentence: you only have 4 points, you did not test any correlation... You have high abundance in spring which corresponds to high chl-a concentrations. This is the result. The term value is here too vague: I guess it is the abundance but could be also "phytoplankton". So I suggest writing abundance if it is abundance.

Commenté [A35]: What does it mean? Too vague. I guess you mean with high biovol and high abundance. If yes you have to use these words.

Commenté [A36]: Not so clear based on fig. 6C...

319 m during day and night in all seasons (Fig. 6H). Cnidarians were overall not very abundant, but
320 constituted an important fraction of the biovolume, especially in 200-300 m depth during day in
321 all seasons, and 50-100 m depth during day time in summer and autumn. At night, they were
322 mainly present in deeper layers 200-450 m (Fig. 6E). Mysida were mainly present in >100 m
323 depth during the day, while a small fraction was found at night between 0 and 50 m in spring and
324 between 50 and 100 m in autumn and winter (Fig. 6F). Chaetognatha were present mainly in
325 intermediate layers between 100-300 m during day and night in all seasons (Fig. 6G).

326

327 **Discussion**

328 *Physicochemical properties and mesozooplankton seasonal dynamics*

329 The present study provides the first detailed data on the zooplankton community composition,
330 distribution and dynamics in the northern Chilean Patagonia. There are only few zooplankton
331 records with a high taxonomic resolution in Patagonian Chilean waters. Most of such studies were
332 conducted in the central and southern parts. Only recently, zooplankton in the northern fjords is
333 receiving more interest, particularly *Calanus* and *Euphausia* (e.g. Marín and Delgado, 2009;
334 Sánchez et al., 2011; Valle-Levinson et al., 2014; Díaz-Astudillo et al., 2017). Palma (2008)
335 described zooplankton distribution and abundance in the north, central and south of Chilean
336 Patagonia, however, with a gap regarding zooplankton diversity in the northern fjords.

337 The Comau Fjord is connected to the Pacific Ocean by the Chacao Channel and Ancud Gulf. It
338 possesses a two-layer system in water masses causing an estuarine circulation, characterized by a
339 strong pycnocline, in which salinity increases from 10 to 32 over a depth of 15 m (Fig. 2B). The
340 physicochemical observations in our study are consistent with earlier descriptions from Comau
341 Fjord, subjected to strong terrestrial inputs, where surface waters (0-50 m) present higher
342 variability than the deeper and quasi-homogeneous layer (Fig. 2). Clear differences were observed
343 in temperature, pH, oxygen and Chl *a* in relation to the season, likely caused by the strong
344 seasonal variability in solar radiation (maximum between spring and summer), precipitation and
345 river discharge (maximum in late autumn and winter) (González et al., 2010).

346 In Comau Fjord, a thermal inversion of the surface layer (0-50 m) in winter is visible (Fig. 2),
347 probably due to heat loss in the surface layer caused by winds and the discharge of cold
348 freshwater from rivers and glaciers (Silva, Calvete & Sievers, 1997). Later in the year, the thermal
349 density stratification stabilizes the water column, triggering a phytoplankton bloom in austral
350 spring (Iriarte et al., 2007) with a strong peak in chlorophyll *a* (Fig. 2E). As shown by previous

351 studies in the area (Palma and Silva, 2004; Vargas et al., 2008; González et al., 2010), this peak in
352 chlorophyll *a* was most likely due to blooming chain-forming diatoms. The high phytoplankton
353 biomass is expected to be grazed predominantly by copepods, increasing their biomass and
354 establishing the classical diatom-to-zooplankton food web (Palma and Silva, 2004; Vargas et al.,
355 2008; González et al., 2010). Accordingly, we found the highest zooplankton abundance during
356 the most productive season, the austral spring. At this time, crustacean nauplii and young stages
357 of calanoid copepods, which are indicative of intense zooplankton reproduction, accounted for a
358 large proportion of the zooplankton community. During summer, biovolume and biomass reached
359 their maxima (Fig. 3), indicating the growth of the zooplankton organisms. Subsequently, in
360 autumn and winter, zooplankton abundance, biovolume and biomass decreased. At this time,
361 primary production should be low, as reflected by low Chl *a* values (Fig. 2E), and likely
362 zooplankton growth was food limited (Escribano et al., 2007). In winter, it is known that the
363 plankton in the fjord shifts towards a microbial loop based community grazed by heterotrophic
364 nanoflagellates, which become the main mesozooplankton prey (Vargas et al., 2008; González et
365 al., 2010) but do not support a high secondary production.

366 Pronounced seasonality of environmental variables often results in high biological production and
367 are associated to seasonal changes in the holoplankton community (Mauchline, 1998; Balbotín
368 and Bustos, 2005; Aracena et al., 2011), and meroplankton abundance (Ladah et al., 2005;
369 Landaeta & Castro, 2006). The northern part of the fjord region, the area between Puerto Montt
370 and Guafo Mouth, represents the most productive area of Chilean Patagonia in terms of primary
371 production and zooplankton biomass (Palma, 2008). In contrast, the phytoplankton production in
372 the southern area is low due to the influence of glaciers, resulting in cold, fresh and turbid waters
373 (Palma and Rosales, 1997; Palma and Silva, 2004; Iriarte et al., 2007; Palma, 2008), and
374 consequently low zooplankton survival and growth (Giesecke et al., 2019). Previous studies
375 described ranges for zooplankton biomass, expressed as the plankton wet volume, of 65 to more
376 than 1386 ml zooplankton 1000 m⁻³ outside Comau Fjord, in the Inner Sea of Chiloé (Palma,
377 2008). This is in line with the present results (250-1500 ml zooplankton 1000 m⁻³), showing an
378 especially high biovolume during the summer season in Comau Fjord. Palma and Rosales (1997)
379 also found the highest values of zooplankton biomass in the northern part (interior of Reloncaví
380 Fjord and Ancud Gulf) with values that ranged between 56-1626 ml zooplankton 1000 m⁻³, but a
381 low zooplankton biomass in the inner of Comau Fjord. The observed variations may potentially
382 be due to (a) temporal differences with a much lower Chl *a* concentration in their year of study for
383 the same season (Ramírez, 1995); and/or (b) methodological and analytical differences, e.g.
384 different sampling gears and proxies for biomass estimation (i.e. measurement of zooplankton wet
385 volume vs. image analysis in this study).

Commenté [A37]: so more a biovolume?

Commenté [A38]: Please give values

386 In fjord systems, oceanographic processes such as estuarine circulation, tidal mixing or water
387 column stratification may influence the composition and abundance of zooplankton communities
388 (Palma & Silva, 2004; Sánchez, González & Iriarte, 2011). Overall, copepods were the main
389 contributors to the total biomass and biovolume of the zooplankton community, especially during
390 summer (69-78%). This is in agreement with previous studies showing that in Chilean fjords,
391 planktonic crustaceans, such as copepods and euphausiids, have the highest abundances and
392 biomasses, followed by chaetognaths and gelatinous plankton (Defren-Janson et al., 1999; Palma
393 and Silva, 2004). Copepods are the most abundant and diverse components of marine zooplankton
394 worldwide (Mauchline, 1998), and the abundance of small copepods (<1.5 mm) generally
395 surpasses the abundance of larger ones (Fransz, 1988; Gallienne and Robins, 1998; Gallienne et
396 al., 2001). Similarly, small copepods accounted for 58-86% of the total copepod community in
397 the present study. Small copepods are an important link between primary producers and
398 epipelagic consumers, and it is thus important to quantify their contribution to the zooplankton
399 community (Turner, 2004). Another important contributor to the zooplankton communities in
400 Chilean fjords is the ~~most abundant and common~~ euphausiid *Euphausia vallentini*. This species
401 mainly occurs in MSAAW, from the ocean to the interior waters of the fjords, where they are
402 often found in dense swarms performing strong vertical migrations (Palma and Silva, 2004;
403 Hamame and Antezana, 2010). The present study revealed young stages of euphausiids during
404 spring and summer but only one adult specimen in winter, indicating that euphausiids are present
405 in Comau fjord, but have not been caught quantitatively in our samples. This is likely related to
406 the small volume filtered by our net and the patchy distribution of *E. vallentini*, as well as the
407 ability of these micronekton organisms to avoid small nets (Brinton, 1962).

408 ***Zooplankton diel vertical migration and population dynamics***

409 Diel Vertical Migration (DVM) is usually associated with differences in light intensity within the
410 photic zone, taking place periodically in 24 h cycles (Brierley, 2014). During day-time,
411 zooplankton organisms migrate to deeper, darker waters to avoid visual predators, such as fishes,
412 while they come to the surface for feeding at night (Hays, Webb & Frears, 1998). In Comau
413 Fjord, we ~~see-saw~~ patterns of DVM related to the size of the zooplankton. Here, a large
414 proportion of zooplankton organisms that inhabit the intermediate waters (100-300 m) during day
415 time ascended to shallow waters at night. But small organisms (individuals <1.5 mm), composed
416 mainly of cyclopoids and harpacticoids, did not perform ~~ed~~ a clear DVM. The two copepod
417 groups, however, differed with regard to their vertical distribution. The highest abundances of
418 cyclopoid copepods were found in the upper 100 m, whereas harpacticoids were found between
419 200-400 m (Fig. 6D). Their population dynamics followed the phytoplankton seasonal cycle, with

the highest abundance in spring, while diminishing in summer to minima abundances in autumn and winter (Table 2).

Large individuals from several calanoid copepod taxa (*Metridia*, *Calanus*, Euchaetidae), mysids, chaetognaths and cnidarians did perform DVM over 300 m. This agrees with the findings by Hays (1995) that DVM is pronounced in large and pigmented species due to their susceptibility of being perceived by visually orientated predators. Similarly, studies in northern Patagonia have shown that conspicuous zooplankton organisms tend to avoid well illuminated waters (Villenas, Soto & Palma, 2009). According to Hays et al. (2001), large individuals of *Metridia* usually reside in deep waters, but a fraction of these ascends to shallow waters at night, whereas smaller individuals stay at the surface continuously. In Comau Fjord, *Metridia* showed the highest values of biovolume and abundance at intermediate depths (100-200 m) during daytime, but in the surface layers at night (Fig. 6B), [which suggested the migration of the largest individuals towards the surface](#). In spring, a significant proportion of the *Metridia* population was found in shallow waters during the day. This was probably due to the smaller size of the individuals and the higher amount of food available here. Following the same pattern as *Metridia*, larger individuals of *Calanus* were found in deeper waters during day-time, whereas small specimens were found in shallow waters also during daytime in spring, where they likely escaped visual predators due to their small size. It is possible that the size differences between day and night might be a consequence of the presence of different species or life stages. Euchaetidae also performed DVM where most of the organisms lived continuously in the deepest waters (200-450 m) during the day, but a small proportion of the population migrated to the surface during night (Fig. 6I). These three copepod [groups](#) presented higher abundances in spring. In upwelling areas, where seasonal changes of environmental conditions are also pronounced, the highest abundances of *Calanus chilensis* were observed in spring and low abundances through autumn and winter (Hidalgo & Escribano, 2007). Therefore, the high abundance of their populations during these seasons is in association to the maximum concentration in phytoplankton, which decreased over time with minima during fall and winter.

Mysids were mainly detected in deep waters (>200 m water depth) accounting for up to 70% of the total zooplankton biovolume. Despite their high variability in their geographic distribution, mysids are typically hyperbenthic and able to feed on a wide range of food sources (Mauchline, 1980), from organic detritus on the seafloor during day (where they reside), and zooplankton, especially cyclopoids and harpacticoids, during the night when they migrate upwards. However, in Comau Fjord they were rarely found in the surface layer. According to Sánchez et al. (2011), the distribution of mesozooplankton in the upper 30 m is strongly influenced by the salinity stratification, which may act as a barrier for the DVM of some oceanic larger zooplankton

455 organisms. Thus, the variability in surface temperature, salinity, pH, and oxygen could have acted
456 as a barrier to the mysids' upward migration in Comau Fjord. Besides their important contribution
457 to the total zooplankton biovolume in deep waters, mysids are often overlooked because of
458 sampling limitations (Mauchline, 1980). Like euphausiids, mysids can form dense swarms,
459 making them a potential food resource for a wide range of organisms, from predatory fishes to
460 benthic CWC, but despite their importance in the marine food web, poor attention has been given
461 to their presence in Chilean Patagonia. To our knowledge, there are only two studies describing
462 mysids in this area: Guglielmo and Ianora (1997) found that the most abundant species for the
463 Strait of Magellan is the deep-dwelling *Boreomysis rostrata*; Díaz-Astudillo et al. (2017) found
464 higher abundances of mysids during night and inside the fjord for the Reloncaví Fjord and Ancud
465 Gulf. Thus, this study constitutes the first record of mysids in Comau Fjord.

466 Chaetognaths were distributed throughout the water column with the highest biovolumes between
467 100-300 m, during both day and night (Fig. 6G). This is in accordance with the distribution found
468 by Guglielmo and Ianora (1995) for the Strait of Magellan. Cnidarian's highest biovolumes were
469 found in 50-100 m and in deeper layers (<200 m) during day and night, respectively. South of
470 Comau Fjord, between Boca del Guafo and Pulluche Channel, the vertical distribution of
471 cnidarians (i.e. Hydromedusae and Siphonophorae) showed that most of the specimens were
472 distributed in the upper 100 m water depth and in deeper waters only in Moraleda and Darwin
473 channels (Palma, Apablaza & Soto, 2007). Considering that copepods compose the principal
474 source of food for cnidarians and chaetognaths (Palma & Kaiser, 1993), a higher abundance of
475 carnivorous zooplankton was observed in spring, probably as a result of the increasing copepod
476 abundance in Comau Fjord during this time. Therefore, the overall decline in copepod abundance
477 from spring to winter may be explained by the decrease in phytoplankton concentration and the
478 increase of the predation pressure imposed by carnivorous zooplankton.

479 In the 50-100 m water layer a minimum in zooplankton abundance and biomass was found in all
480 seasons (Fig. 4). This "zooplankton gap" could be related to a high concentration of predators in
481 this water depth. In the northern fjords, the high biomasses and abundances of gelatinous
482 organisms are correlated to a decrease in chitinous biomass of other organisms (Palma and Silva,
483 2004; Villenas et al., 2009; this study). It is known that chaetognaths and cnidarians can grow at
484 high rates, form dense aggregations and seasonally dominate the zooplankton biomass (Casanova,
485 1999; Brodeur et al., 2002). Large densities of chaetognaths and gelatinous organisms, such as
486 medusae and siphonophores, could decimate a zooplankton population as they are characterized
487 by being voracious consumers of copepods and larvae from other organisms (Lie et al., 1983;
488 Palma and Rosales, 1997). For instance, large aggregations of the siphonophore *Muggiaea*
489 *atlantica*, common in the area (Palma & Rosales, 1997; Palma & Silva, 2004; Villenas, Soto &

Commenté [A39]: should be >, because greater depth (> 200m depth)

490 Palma, 2009), lead to a decrease in fish stocks, due to competition over the same
491 mesozooplankton prey (Purcell, 1985). In our study, we found a disproportionately large
492 biovolume of carnivorous organisms (68-82% of the total zooplankton biovolume) in this water
493 layer, particularly in summer and autumn, although it was not evident for spring and winter (Fig.
494 7). This may be due to the fact that euphausiids were not quantitatively sampled and they may peak
495 in this layer. According to Försterra and Häussermann (2012), euphausiids may reside around 150
496 m during daytime and migrate upwards close to the pycnocline at night. Because euphausiids are
497 a potential food for many organisms, they could attract many predators, such as anchovies or blue
498 whales, into the 50-100 m water depth which may reduce the mesozooplankton community.
499 Therefore, this suggest that carnivorous zooplankton may have depleted zooplankton in this
500 twilight zone and predation may play a role in structuring the vertical distribution of species in
501 Comau Fjord.

502 In the present study, zooplankton abundance, biovolume and biomass, integrated over the entire
503 water column, differed between day and night. It is difficult to reconcile this with a resident
504 zooplankton community, where diel vertical migrations change the vertical distribution but not
505 the water-column-integrated values. These differences were probably due to the high patchiness
506 the zooplankton exhibited during day and night, especially by larger zooplankton individuals, and
507 those able to form swarms (i.e. mysids and euphausiids). Biological processes, such as diel
508 vertical migration, predator avoidance, location of food patches and mating are mechanisms by
509 which the underlying zooplankton behavior present high spatial heterogeneity (Folt & Burns,
510 1999). Therefore, in Comau Fjord, those biological drivers are considered to be the main factors
511 governing the distribution of zooplankton.

512 ***Ecological connection with cold-water corals***

513 Mesozooplankton plays an important role in the food web as a food source for many pelagic and
514 benthic organisms (González et al., 2013), including cold-water corals (CWC) (Gili et al., 2006;
515 Carlier et al., 2009; Mayr et al., 2011). In Comau Fjord, CWC thrive despite the naturally
516 acidified waters, in which the pH ranges from 8.4 to 7.4, representing aragonite over- to aragonite
517 undersaturated waters (Häussermann and Försterra, 2007; Försterra et al., 2017). The occurrence
518 of dense CWC banks colonizing the steep walls of the fjord is a long-standing enigma
519 (Häussermann and Försterra, 2007; Fillinger and Richter, 2013a), as coral calcification is a
520 challenge in aragonite-undersaturated waters (Orr et al., 2005; Guinotte et al., 2006). According to
521 a laboratory study by Martínez-Dios et al. (2020), a high ingestion rate of the CWC
522 *Desmophyllum dianthus* showed a positive impact on their calcification rates, regardless of the
523 seawater pH.

524 Due to the difficulty of studying CWC *in situ*, little is known about their natural diet and their
525 feeding capacity. Only recently, a study confirmed that *D. dianthus* preyed on medium and large
526 sized calanoid copepods and euphausiids (Höfer et al., 2018). Based on our recent DVM data,
527 shallow-dwelling CWC in Comau Fjord might feed on small copepods (e.g. cyclopoid and
528 calanoid) and/or nauplii (Tsounis et al., 2010) during the day, and on larger organisms during
529 night hours when zooplankton migrates upwards. Deeper-dwelling corals, by contrast, may
530 mainly encounter larger prey, such as mysids or large calanoid copepods (Calanidae,
531 Euchaetidae). Large seasonal variability affected the abundance and biomass of the zooplankton
532 in Comau Fjord. Therefore, CWC must be adapted to differences in food availability, i.e. high
533 zooplankton abundances during spring and summer and low concentrations during autumn and
534 winter. In summer, CWC in Comau Fjord display high growth rates (Jantzen et al., 2013b) which
535 may be associated with the high zooplankton availability. In winter, when zooplankton biomass is
536 low, CWC may slow down their metabolism to cope with the low concentration of food available
537 (Naumann et al., 2011).

538 Höfer et al. (2018) observed in laboratory experiments that *D. dianthus* showed no feeding
539 saturation, when prey was up to five times more abundant than in nature. They suggested that
540 CWC are able to exploit dense swarms of zooplankton which may aggregate near boundaries
541 (Mauchline, 1980; Genin et al., 2005). *D. dianthus* is found in high densities, attached to the hard
542 substratum of the steep walls from Comau Fjord, which can be found from 8 m water depth
543 (directly below the pycnocline) down to the seafloor (~480 m) (Cairns, Häussermann & Försterra,
544 2005; Fillinger & Richter, 2013a). The highest abundances were, however, found above 270 m,
545 where *D. dianthus* forms coral banks (Fillinger & Richter, 2013b). In these depths, as the present
546 study shows, also the zooplankton biomass was the highest. If we assume a freestream flow of 5
547 cm s⁻¹ and a current of 2.5 cm s⁻¹ in the coral canopy near the wall, with a dense tentacle crown of
548 7 cm in diameter for a given coral individual capturing zooplankton in a volume of 250 ml at 0.5
549 efficiency (every second individual retained), we found that below 300 m water depth, CWC
550 would have an annual mean capture of less than 790 preys per polyp a day. In shallow waters,
551 CWC would have a supply rate of 5809-10754 preys per polyp a day between day and night. This
552 could explain, why despite the availability of hard substrate, *D. dianthus* is not found in high
553 numbers below 270 m (Fillinger & Richter, 2013a). We then conclude that 790 prey polyp⁻¹ day⁻¹
554 could be the zooplankton flux necessary to sustain a viable coral population, beyond which a
555 surplus of occasional krill swarms would be needed. Therefore, the high abundance of CWC in
556 the aragonite-undersaturated waters of the Comau Fjord might be due to the high food availability
557 provided by the zooplankton biomass (Cairns et al., 2005; González et al., 2010; this study).

558

559 Conclusions

560 The seasonal changes of zooplankton over the entire water column showed that abundance peaked
561 in spring, likely due to a phytoplankton bloom during this time. In summer, biovolume and
562 biomass ~~was~~ were the highest and decreased thereafter over time, reaching the lowest values in
563 late autumn and mid-winter. Probably, the low concentration of Chl-*a* during the cold seasons
564 could not support secondary production. The vertical distribution of zooplankton biovolume and
565 biomass differed between day and night, with a daytime maximum in the 100-200 m water depth
566 and a nighttime maximum in surface waters (0-50 m) associated with the diel vertical migration
567 of the calanoid copepod *Metridia*. Overall, copepods were the dominant group of the total
568 zooplankton community with an important contribution of the small organisms (individuals < 1.5
569 mm), followed by mysids, chaetognaths and cnidarians (biovolume and biomass), and nauplii and
570 Appendicularia (abundance).

571 In Comau Fjord, cold-water corals (CWC) thrive despite its naturally acidified waters. To
572 maintain their metabolic functions, CWC need to ingest a high amount of zooplankton. This study
573 provides a description of the natural food spectrum for CWC where shallow corals may feed on
574 small copepods or nauplii during day and larger organisms during night, while deeper-dwelling
575 corals may mainly feed on mysids, euphausiids or larger calanoid copepods. CWC need to be
576 adapted to the seasonal zooplankton variability with higher zooplankton abundances and biomass
577 in spring and summer, and lower concentrations in autumn and winter. The high amount of
578 zooplankton in Comau Fjord provides sufficient nourishment to maintain a viable coral
579 population despite the aragonite under-saturated waters. Nevertheless, future studies are necessary
580 to understand how CWC cope with small and large zooplankton preys and how much of the
581 carbon and nitrogen ingested are directed to which metabolic pathways. Therefore, the Comau
582 Fjord constitutes a natural laboratory where we can understand the benthic-pelagic coupling
583 between zooplankton and CWC under low pH, in an area where anthropogenic activities are
584 gaining importance jeopardizing the natural conditions for the marine biota.

585

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593 **References**

- 594 Antezana T. 1999. Plankton of Southern Chilean fjords: trends and linkages. *Scientia Marina*
595 63:69–80. DOI: 10.3989/scimar.1999.63s169.
- 596 Aracena C, Lange CB, Iriarte JL, Rebolledo L, Pantoja S. 2011. Latitudinal patterns of export
597 production recorded in surface sediments of the Chilean Patagonian fjords (41–55°S) as a
598 response to water column productivity. *Continental Shelf Research* 31:340–355. DOI:
599 10.1016/j.csr.2010.08.008.
- 600 Barbier EB. 2017. Marine ecosystem services. *Current Biology* 27:R507–R510. DOI:
601 10.1016/j.cub.2017.03.020.
- 602 Brierley AS. 2014. Diel vertical migration. *Current Biology* 24:R1074–R1076. DOI:
603 10.1016/j.cub.2014.08.054.
- 604 Brinton E. 1962. Variable factors affecting the apparent range and estimated concentration of
605 euphausiids in the North Pacific. *Pacific Science* 16:374–408.
- 606 Brodeur RD, Sugisaki H, Hunt Jr GL. 2002. Increases in jellyfish biomass in the Bering Sea:
607 Implications for the ecosystem. *Marine Ecology Progress Series* 233:89–103. DOI:
608 10.3354/meps233089.
- 609 Buschmann AH, Niklitschek EJ, Pereda S. 2021. Acuicultura y sus impactos en la conservación
610 de la Patagonia chilena. In: Castilla JC, Armesto JJ, Martínez-Harms MJ eds. *Conservación
611 en la Patagonia chilena: evaluación del conocimiento, oportunidades y desafíos*. Santiago,
612 Chile: Ediciones Universidad Católica de Chile, 367–387.
- 613 Cáceres M, Valle-Levinson A, Sepúlveda HH, Holderied K. 2002. Transverse variability of flow
614 and density in a Chilean fjord. *Continental Shelf Research* 22:1683–1698. DOI:
615 10.1016/S0278-4343(02)00032-8.
- 616 Cairns SD, Häussermann V, Försterra G. 2005. A review of the Scleractinia (Cnidaria: Anthozoa)
617 of Chile, with the description of two new species. *Zootaxa* 1018:15–46. DOI:
618 10.11646/zootaxa.1018.1.2.
- 619 Carlier A, Le Guilloux E, Olu K, Sarrazin J, Mastrototaro F, Taviani M, Clavier J. 2009. Trophic
620 relationships in a deep mediterranean cold-water coral bank (Santa Maria di Leuca, Ionian
621 sea). *Marine Ecology Progress Series* 397:125–137. DOI: 10.3354/meps08361.
- 622 Casanova JP. 1999. Chaetognatha. In: Boltovskoy D ed. *South Atlantic Zooplankton*. Backuys
623 Publ., Leiden, 1353–1374.
- 624 Castro LR, Cáceres MA, Silva N, Muñoz MI, León R, Landaeta MF, Soto-Mendoza S. 2011.
625 Short-term variations in mesozooplankton, ichthyoplankton, and nutrients associated with
626 semi-diurnal tides in a patagonian Gulf. *Continental Shelf Research* 31:282–292. DOI:
627 10.1016/j.csr.2010.09.005.
- 628 Díaz-Astudillo M, Cáceres MA, Landaeta MF. 2017. Zooplankton structure and vertical
629 migration: Using acoustics and biomass to compare stratified and mixed fjord systems.
630 *Continental Shelf Research* 148:208–218. DOI: 10.1016/j.csr.2017.09.004.
- 631 Emerson CW, Roff JC. 1987. Implications of fecal pellet size and zooplankton behaviour to
632 estimates of pelagic-benthic carbon flux. *Marine Ecology Progress Series* 35:251–257. DOI:
633 10.3354/meps035251.
- 634 Escribano R, Hidalgo P, González H, Giesecke R, Riquelme-Bugueño R, Manríquez K. 2007.
635 Seasonal and inter-annual variation of mesozooplankton in the coastal upwelling zone off

central-southern Chile. *Progress in Oceanography* 75:470–485. DOI: 10.1016/j.pocean.2007.08.027.

Fillinger L, Richter C. 2013a. Vertical and horizontal distribution of *Desmophyllum dianthus* in Comau Fjord, Chile: a cold-water coral thriving at low pH. *PeerJ*:1–22. DOI: 10.7717/peerj.194.

Fillinger L, Richter C. 2013b. Abundance of *Desmophyllum dianthus* during expedition Comau2012. In: Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research, Bremerhaven, PANGAEA,. DOI: <https://doi.org/10.1594/PANGAEA.811898>.

Folt C, Burns C. 1999. Biological drivers of zooplankton patchiness. *Trends in Ecology and Evolution* 14:300–305. DOI: 10.1016/S0169-5347(99)01616-X.

Försterra G, Häussermann V. 2012. Report on blue whales sightings (*Balaenoptera musculus* linnaeus, 1758) in a narrow fjord during autumn-winter in southern Chile. *Spixiana* 35:237–245.

Försterra G, Häussermann V, Laudien J. 2017. Animal forests in the Chilean Fjords; discoveries, perspectives, and threats in shallow and deep waters. In: Rossi S, Bramanti L, Gori A, Orejas C eds. *Marine Animal Forests*. Springer, Cham,. DOI: http://doi-org-443.webvpn.fjmu.edu.cn/10.1007/978-3-319-21012-4_3.

Fowler SW, Knauer GA. 1986. Role of large particles in the transport of elements and organic compounds through the oceanic water column. *Progress in Oceanography* 16:147–194. DOI: [https://doi.org/10.1016/0079-6611\(86\)90032-7](https://doi.org/10.1016/0079-6611(86)90032-7).

Franz HG. 1988. Vernal abundance, structure and development of epipelagic copepod populations of the eastern Weddell Sea (Antarctica). *Polar Biology* 9:107–114. DOI: 10.1007/BF00442037.

Gallienne CP, Robins DB. 1998. Trans-oceanic characterization of zooplankton community size structure using an optical plankton counter. *Fisheries Oceanography* 7:147–158.

Gallienne CP, Robins DB, Woodd-Walker RS. 2001. Abundance, distribution and size structure of zooplankton along a 20° west meridional transect of the northeast Atlantic Ocean in July. *Deep-Sea Research II* 48:925–949. DOI: 10.1016/S0967-0645(00)00114-4.

Genin A, Jaffe JS, Reef R, Richter C, Franks PJS. 2005. Swimming against the flow: a mechanism of zooplankton aggregation. *Science* 308:860–862. DOI: 10.1126/science.1107834.

Giesecke R, Höfer J, Vallejos T, González HE. 2019. Death in southern Patagonian fjords: copepod community structure and mortality in land- and marine-terminating glacier-fjord systems. *Progress in Oceanography* 174:162–172. DOI: 10.1016/j.pocean.2018.10.011.

Gili JM, Rossi S, Pagès F, Orejas C, Teixidó N, López-González PJ, Armtz WE. 2006. A new trophic link between the pelagic and benthic systems on the Antarctic shelf. *Marine Ecology Progress Series* 322:43–49. DOI: 10.3354/meps322043.

González HE, Calderón MJ, Castro L, Clement A, Cuevas LA, Daneri G, Iriarte JL, Lizárraga L, Martínez R, Menschel E, Silva N, Carrasco C, Valenzuela C, Vargas CA, Molinet C. 2010. Primary production and plankton dynamics in the Reloncaví Fjord and the Interior Sea of Chiloé, Northern Patagonia, Chile. *Marine Ecology Progress Series* 402:13–30. DOI: 10.3354/meps08360.

González HE, Castro LR, Daneri G, Iriarte JL, Silva N, Tapia F, Teca E, Vargas CA. 2013. Land-ocean gradient in haline stratification and its effects on plankton dynamics and trophic carbon fluxes in Chilean Patagonian fjords (47–50°S). *Progress in Oceanography* 119:32–47. DOI: 10.1016/j.pocean.2013.06.003.

682 González HE, Castro L, Daneri G, Iriarte JL, Silva N, Vargas CA, Giesecke R, Sánchez N. 2011.
683 Seasonal plankton variability in Chilean Patagonia fjords: Carbon flow through the pelagic
684 food web of Aysen Fjord and plankton dynamics in the Moraleda Channel basin. *Continental*
685 *Shelf Research* 31:225–243. DOI: 10.1016/j.csr.2010.08.010.

686 Gorsky G, Ohman MD, Picheral M, Gasparini S, Stemmann L, Romagnan JB, Cawood A, Pesant
687 S, García-Comas C, Prejger F. 2010. Digital zooplankton image analysis using the ZooScan
688 integrated system. *Journal of Plankton Research* 32:285–303. DOI: 10.1093/plankt/fbp124.

689 Grosjean P, Picheral M, Warembourg C, Gorsky G. 2004. Enumeration, measurement, and
690 identification of net zooplankton samples using the ZOOSCAN digital imaging system.
691 *ICES Journal of Marine Science* 61:518–525. DOI: 10.1016/j.icesjms.2004.03.012.

692 Guglielmo L, Ianora A (eds.). 1995. *Atlas of Marine Zooplankton. Straits of Magellan. Copepods*.
693 Springer-Verlag, Berlin Heidelberg. DOI: 10.1007/978-3-642-79139-0.

694 Guglielmo L, Ianora A (eds.). 1997. *Atlas of Marine Zooplankton. Strait of Magellan. Amphipods,*
695 *Euphausiids, Mysids, Ostracods, and Chaetognaths*. Springer-Verlag Berlin Heidelberg.
696 DOI: 10.1007/978-3-642-60340-2.

697 Guinotte JM, Orr J, Cairns S, Freiwald A, Morgan L, George R. 2006. Will human-induced
698 changes in seawater chemistry alter the distribution of deep-sea scleractinian corals?
699 *Frontiers in Ecology and the Environment* 4:141–146. DOI: 10.1890/1540-
700 9295(2006)004[0141:WHCISC]2.0.CO;2.

701 Hamame M, Antezana T. 2010. Vertical diel migration and feeding of *Euphausia vallentini*
702 within southern Chilean fjords. *Deep-Sea Research II* 57:642–651. DOI:
703 10.1016/j.dsr2.2009.10.013.

704 Häussermann V, Försterra G. 2007. Large assemblages of cold-water corals in Chile: a summary
705 of recent findings and potential impacts. *Bulletin of Marine Science* 81:195–207.

706 Häussermann V, Försterra G, Laudien J. Hard bottom marobenthos of Chilean Patagonia:
707 emphasis on conservation of sublittoral invertebrate algal forests. In: Castilla JC, Armesto JJ,
708 Martínez-Harms MJ eds. *Conservación en la Patagonia Chilena: evaluación del*
709 *conocimiento, oportunidades y desafíos*. Santiago, Chile: Ediciones Universidad Católica de
710 Chile, 321–343.

711 Hays GC. 1995. Ontogenetic and seasonal variation in the diel vertical migration of the copepods
712 *Metridia lucens* and *Metridia longa*. *Limnology and Oceanography* 40:1461–1465.

713 Hays GC, Kennedy H, Frost BW. 2001. Individual variability in diel vertical migration of a
714 marine copepod: Why some individuals remain at depth when others migrate. *Limnology and*
715 *Oceanography* 46:2050–2054. DOI: 10.4319/lo.2001.46.8.2050.

716 Hays GC, Webb PI, Frears SL. 1998. Diel changes in the carbon and nitrogen content of the
717 copepod *Metridia lucens*. *Journal of Plankton Research* 20:727–737. DOI:
718 10.1093/plankt/20.4.727.

719 Hernández-León S, Montero I. 2006. Zooplankton biomass estimated from digitalized images in
720 Antarctic waters: A calibration exercise. *Journal of Geophysical Research* 111:1–6. DOI:
721 10.1029/2005JC002887.

722 Hidalgo P, Escribano R. 2007. Coupling of life cycles of the copepods *Calanus chilensis* and
723 *Centropages brachiatus* to upwelling induced variability in the central-southern region of
724 Chile. *Progress in Oceanography* 75:501–517. DOI: 10.1016/j.pocean.2007.08.028.

725 Höfer J, González HE, Laudien J, Schmidt GM, Häussermann V, Richter C. 2018. All you can
726 eat: the functional response of the cold-water coral *Desmophyllum dianthus* feeding on krill
727 and copepods. *PeerJ*. DOI: 10.7717/peerj.5872.

728 Iriarte JL, González HE, Liu KK, Rivas C, Valenzuela C. 2007. Spatial and temporal variability
729 of chlorophyll and primary productivity in surface waters of southern Chile (41.5-43° S).
730 *Estuarine, Coastal and Shelf Science* 74:471–480. DOI: 10.1016/j.ecss.2007.05.015.

731 Iriarte JL, González HE, Nahuelhual L. 2010. Patagonian fjord ecosystems in Southern Chile as a
732 highly vulnerable region: problems and needs. *Ambio* 39:463–466. DOI: 10.1007/s13280-
733 010-0049-9.

734 Iwasa Y. 1982. Vertical migration of zooplankton : a game between predator and prey. *The*
735 *American Naturalist* 120:171–180.

736 Jantzen C, Häussermann V, Försterra G, Laudien J, Ardelan M, Maier S, Richter C. 2013a.
737 Occurrence of a cold-water coral along natural pH gradients (Patagonia, Chile). *Marine*
738 *Biology* 160:2597–2607. DOI: 10.1007/s00227-013-2254-0.

739 Jantzen C, Laudien J, Sokol S, Försterra G, Häussermann V, Kupprat F, Richter C. 2013b. In situ
740 short-term growth rates of a cold-water coral. *Marine and Freshwater Research* 64:631–641.
741 DOI: 10.1071/MF12200.

742 Ladah LB, Tapia FJ, Pineda J, López M. 2005. Spatially heterogeneous, synchronous settlement
743 of *Chthamalus* spp. larvae in northern Baja California. *Marine Ecology Progress Series*
744 302:177–185.

745 Landaeta MF, Castro LR. 2006. Larval distribution and growth of the rockfish, *Sebastes capensis*
746 (Sebastidae, Pisces), in the fjords of southern Chile. *ICES Journal of Marine Science*:714–
747 724.

748 Lehette P, Hernández-León S. 2009. Zooplankton biomass estimation from digitized images: a
749 comparison between subtropical and Antarctic organisms. *Limnology and Oceanography*:
750 *Methods* 7:304–308. DOI: 10.4319/lom.2009.7.304.

751 Lie U, Magnesen T, Tunberg B, Aksnes D. 1983. Preliminary studies on the vertical distribution
752 of size-fractions in the zooplankton community in Lindås- Pollene, Western Norway. *Sarsia*
753 68:65–80. DOI: 10.1080/00364827.1983.10420558.

754 Maier C, Popp P, Sollfrank N, Weinbauer MG, Wild C, Gattuso JP. 2016. Effects of elevated
755 pCO₂ and feeding on net calcification and energy budget of the Mediterranean cold-water
756 coral *Madrepora oculata*. *Journal of Experimental Biology* 219:3208–3217. DOI:
757 10.1242/jeb.127159.

758 Mann KH, Lazier JRN. 1991. *Dynamics of marine ecosystems: biological-physical interactions in*
759 *the oceans*. Oxford, Blackwell Scientific Publications.

760 Marcolin C da R, Gaeta S, Lopes RM. 2015. Seasonal and interannual variability of zooplankton
761 vertical distribution and biomass size spectra off Ubatuba , Brazil. *Journal of Plankton*
762 *Research* 37:808–819. DOI: 10.1093/plankt/fbv035.

763 Marín VH, Delgado LE. 2009. Diversidad y distribución espacial de copépodos superficiales (0-
764 10 metros) en la zona costera del mar interior de Chiloé (Cimar 10 Fiordos). *Ciencia y*
765 *Tecnología del Mar* 32:95–100.

766 Martínez-Dios A, Pelejero C, López-Sanz À, Sherrell RM, Ko S, Häussermann V, Försterra G,
767 Calvo E. 2020. Effects of low pH and feeding on calcification rates of the cold-water coral
768 *Desmophyllum dianthus*. *PeerJ*. DOI: 10.7717/peerj.8236.

769 Mauchline J. 1980. The biology of mysids and euphausiids. Chapter 4 - Vertical distribution and
770 migration. *Advances in Marine Biology*:66–80.

771 Mauchline J. 1998. The biology of calanoid copepods. *Advances in Marine Biology* 33:1–710.

772 Mayr CC, Försterra G, Häussermann V, Wunderlich A, Grau J, Zieringer M, Altenbach A V.
773 2011. Stable isotope variability in a Chilean fjord food web: implications for N- and C-

cycles. *Marine Ecology Progress Series* 428:89–104. DOI: 10.3354/meps09015.

McCulloch M, Trotter J, Montagna P, Falter J, Dunbar R, Freiwald A, Försterra G, López Correa M, Maier C, Rüggeberg A, Taviani M. 2012. Resilience of cold-water scleractinian corals to ocean acidification: boron isotopic systematics of pH and saturation state up-regulation. *Geochimica et Cosmochimica Acta* 87:21–34. DOI: 10.1016/j.gca.2012.03.027.

Meerhoff E, Castro LR, Tapia FJ, Pérez-Santos I. 2019. Hydrographic and biological impacts of a Glacial Lake Outburst Flood (GLOF) in a Patagonian Fjord. *Estuaries and Coasts* 42:132–143. DOI: 10.1007/s12237-018-0449-9.

Naumann MS, Orejas C, Wild C, Ferrier-Pagès C. 2011. First evidence for zooplankton feeding sustaining key physiological processes in a scleractinian cold-water coral. *Journal of Experimental Biology* 214:3570–3576. DOI: 10.1242/jeb.061390.

Navedo JG, Vargas-Chacoff L. 2021. Salmon aquaculture threatens Patagonia. *Science* 372:695–696. DOI: 10.1126/science.abj0016.

Nemoto T. 1970. Feeding pattern of baleen whales in the ocean. In: Steele JH ed. *Marine food chains*. University of California Press, 551.

Orr JC, Fabry VJ, Aumont O, Bopp L, Doney SC, Feely RA, Gnanadesikan A, Gruber N, Ishida A, Joos F, Key RM, Lindsay K, Maier-Reimer E, Matear R, Monfray P, Mouchet A, Najjar RG, Plattner G-K, Rodgers KB, Sabine CL, Sarmiento JL, Reiner S, Slater RD, Totterdell IJ, Weirig M, Yamanaka Y, Yool A. 2005. Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature* 437:681–686. DOI: 10.1038/nature04095.

Palma S. 2008. Zooplankton distribution and abundance in the austral Chilean channels and fjords. In: Silva N, Palma S eds. *Progress in the oceanographic knowledge of Chilean interior waters, from Puerto Montt to Cape Horn*. Comité Oceanográfico Nacional - Pontificia Universidad Católica de Valparaíso, Valparaíso, 107–113.

Palma S, Apablaza P, Soto D. 2007. Diversity and aggregation areas of planktonic cnidarians of the southern channels of Chile (Boca del Guafo to Pulluche Channel). *Investigaciones Marinas* 35:71–82. DOI: 10.4067/S0717-71782007000200008.

Palma S, Kaiser K. 1993. *Plancton marino de aguas Chilenas*. Ediciones Universitarias de Valparaíso.

Palma S, Rosales SA. 1997. Sifonóforos epipelágicos de los Canales Australes Chilenos (41° 30' - 46° 40' S). *Ciencia y Tecnología del Mar* 20:125–145.

Palma S, Silva N. 2004. Distribution of siphonophores, chaetognaths, euphausiids and oceanographic conditions in the fjords and channels of southern Chile. *Deep-Sea Research II* 51:513–535. DOI: 10.1016/j.dsr2.2004.05.001.

Palma S, Silva N, Retamal MC, Castro L. 2011. Seasonal and vertical distributional patterns of siphonophores and medusae in the Chiloé Interior Sea, Chile. *Continental Shelf Research* 31:260–271. DOI: 10.1016/j.csr.2010.04.007.

Pan J, Marcoval MA, Bazzini SM, Vallina M V., Marco SG de. 2013. Coastal marine biodiversity challenges and threats. In: Arias AH, Menendez MC eds. *Marine Ecology in a Changing World*. CRC Press, 43–67. DOI: 10.1201/b16334-3.

Pantoja S, Iriarte JL, Daneri G. 2011. Oceanography of the Chilean Patagonia. *Continental Shelf Research* 31:149–153. DOI: 10.1016/j.csr.2010.10.013.

Pérez-Santos I, Castro L, Ross L, Niklitschek E, Mayorga N, Cubillos L, Gutierrez M, Escalona E, Castillo M, Alegría N, Daneri G. 2018. Turbulence and hypoxia contribute to dense zooplankton scattering layers in Patagonian Fjord System. *Ocean Science* 14:1185–2018.

DOI: 10.5194/os-2017-89.

Pérez-Santos I, Garcés-Vargas J, Schneider W, Ross L, Parra S, Valle-Levinson A. 2014. Double-diffusive layering and mixing in Patagonian fjords. *Progress in Oceanography* 129:35–49. DOI: 10.1016/j.pocean.2014.03.012.

Picheral M, Colin S, Irisson J-O. 2017. EcoTaxa, a tool for the taxonomic classification of images. <http://ecotaxa.obs-vlfr.fr>.

Purcell JE. 1985. Predation of fish eggs and larvae by pelagic cnidarians and ctenophores. *Bulletin of Marine Science* 37:739–755.

Ramírez B. 1995. Distribución de la biomasa pigmentaria en los Canales Australes. In: *Resúmenes Resultados Crucero Cimar Fiordo 1*. 74–75.

Sánchez N, González HE, Iriarte JL. 2011. Trophic interactions of pelagic crustaceans in Comau Fjord (Chile): Their role in the food web structure. *Journal of Plankton Research* 33:1212–1229. DOI: 10.1093/plankt/fbr022.

Schneider CA, Rasband WS, Eliceiri KW. 2012. NIH Image to ImageJ: 25 years of image analysis. *Nature Methods* 9:671–675. DOI: 10.1038/nmeth.2089.

Sievers HA, Silva N. 2008. Water masses and circulation in austral Chilean channels and fjords. In: Silva N, Palma S eds. *Progress in the oceanographic knowledge of Chilean interior waters, from Puerto Montt to Cape Horn*. Comité Oceanográfico Nacional - Pontificia Universidad Católica de Valparaíso, Valparaíso, 53–58.

Silva N, Calvete C, Sievers HA. 1997. Características Oceanográficas Físicas y Químicas de Canales Australes Chilenos entre Puerto Montt y Laguna San Rafael (Crucero Cimar-Fiordo 1). *Ciencia y Tecnología del Mar* 20:23–106.

Stich HB, Lampert W. 1981. Predator evasion as an explanation of diurnal vertical migration by zooplankton. *Nature* 293:396–398. DOI: 10.1038/293396a0.

Sullivan BK. 1993. How does water column structure influence copepod populations in coastal marine systems? *Bulletin of Marine Science* 53:240–246.

Tsounis G, Orejas C, Reynaud S, Gili JM, Allemand D, Ferrier-Pagès C. 2010. Prey-capture rates in four Mediterranean cold water corals. *Marine Ecology Progress Series* 398:149–155. DOI: 10.3354/meps08312.

Turner JT. 2004. The importance of small planktonic copepods and their roles in pelagic marine food webs. *Zoological Studies* 43:255–266.

Tutasi P, Escribano R. 2020. Zooplankton diel vertical migration and downward C flux into the oxygen minimum zone in the highly productive upwelling region off Northern Chile. *Biogeosciences* 17:455–473. DOI: 10.5194/bg-2019-127.

Urrère MA, Knauer GA. 1981. Zooplankton fecal pellet fluxes and vertical transport of particulate organic material in the pelagic environment. *Journal of Plankton Research* 3:369–387. DOI: 10.1093/plankt/3.3.369.

Valle-Levinson A, Castro L, Cáceres M, Pizarro O. 2014. Twilight vertical migrations of zooplankton in a Chilean fjord. *Progress in Oceanography* 129:114–124. DOI: 10.1016/j.pocean.2014.03.008.

Vandromme P, Stemmann L, Garcia-Comas C, Berline L, Sun X, Gorsky G. 2012. Assessing biases in computing size spectra of automatically classified zooplankton from imaging systems: a case study with the ZooScan integrated system. *Methods in Oceanography* 1–2:3–21. DOI: 10.5194/bgd-7-9175-2010.

Vargas CA, Martínez RA, González HE, Silva N. 2008. Contrasting trophic interactions of microbial and copepod communities in a fjord ecosystem, Chilean Patagonia. *Aquatic*

866 *Microbial Ecology* 53:227–242. DOI: 10.3354/ame01242.
867 Villenas F, Soto D, Palma S. 2009. Cambios interanuales en la biomasa y biodiversidad de
868 zooplankton gelatinoso en aguas interiores de Chiloé, sur de Chile (primaveras 2004 y 2005).
869 *Revista de Biología Marina y Oceanografía* 44:309–324. DOI: 10.4067/s0718-
870 19572009000200005.
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873 **Tables:**

874

875 **Table 1:** Regression equations between individual dry mass and body area to estimate biomass
876 for different taxonomic groups given by Lehet and Hernández-León (2009)

877

878 **Table 2:** Integrated abundances (ind m⁻²) of the most important zooplankton taxa over the entire
879 water column sampled on four dates throughout a year.

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881

882 **Figures:**

883

884 **Fig. 1: Study site.** (A) Overview of Chilean Patagonia. Red square denotes area in panel B. (B)
885 Inner Sea of Chiloé where Comau Fjord (red dot) is located. (C) Comau Fjord with the
886 bathymetry and location of the station where zooplankton samples were taken (red dot). Adapted
887 from Fillinger & Richter (2013a).

888

889 **Fig. 2: Vertical profiles of physico-chemical parameters in the Comau Fjord.** ~~Physico-chemical~~
890 ~~parameters for~~ (A) temperature, (B) salinity, (C) pH, (D) oxygen and (E) Chlorophyll-~~a~~ of Comau
891 ~~Fjord~~. Note the break at 100 m with the different scales above and below.

892

893 **Fig. 3: Zooplankton seasonal and diel distribution.** Seasonal distribution of integrated (A)
894 abundance, (B) biovolume and (C) biomass of the zooplankton community during day and night.

895

896 **Fig 4: Zooplankton vertical distribution.** Seasonal, diel and vertical distribution of (A)
897 abundance, (B) biovolume and (C) biomass of the zooplankton community during day and night.

898

899 **Fig 5: Taxonomic composition.** ~~Percentage of taxonomic composition of~~ Relative (A) biovolume
900 and (B) abundance of major zooplankton groups. Taxa comprising less than 3% of the total
901 zooplankton community (27 taxa) were pooled together as “other”.

902

903 **Fig 6: Seasonal, diel and vertical distribution of the most important taxa.** Vertical, diel and
904 seasonal distribution of the most important taxa representing their biovolume and abundance (%)
905 in relation to the rest of the zooplankton community. In (A) is presented the total sum of
906 biovolume and abundance in all samples. Vertical distributions of (B) Metridinidae, (C)
907 Calanidae, (D) Harpacticoida, (E) Cnidaria, (F) Mysida, (G) Chaetognatha, (H) Cyclopoida and
908 (I) Euchaetidae is represented. The group “small calanoids (<1.5 mm)”, although higher than 5%
909 both in biovolume and abundance, is not represented here as it is composed of a mix of taxa with
910 different functions.

911

912 **Fig 7: Copepods versus carnivorous zooplankton.** Percentages of copepods versus carnivorous
913 zooplankton for (A) total biovolume and (B) total abundance.

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919 Table 1 Regression equations between individual dry mass and body area to estimate biomass for
 920 different taxonomic groups given by Lehet and Hernández-León (2009)
 921

Organism	<i>a</i>	<i>b</i>	Area (mm ²)
Actinopterygii (eggs and larvae)	43.38	1.54	0.079 – 1.198
Appendicularia	43.38	1.54	0.056 – 6.071
Ascidacea (larvae)	43.38	1.54	0.072 – 1.652
Amphipoda	43.38	1.54	0.103 – 59.854
Brachiopoda (larvae)	43.38	1.54	0.193 – 0.366
Bivalvia	43.38	1.54	0.071 – 2.040
Bryozoa (larvae)	43.38	1.54	0.067 – 0.240
Chaetognatha	23.45	1.19	0.068 – 15.935
Cirripedia	43.38	1.54	0.071 – 0.286
Cladocera	43.38	1.54	0.072 – 0.455
Cnidaria	4.03	1.24	0.051 – 95.743
Copepoda	43.97	1.52	0.068 – 9.177
Crustacea (nauplii)	43.38	1.54	0.070 – 0.878
Decapoda (zoea)	43.38	1.54	0.072 – 6.733
Echinodermata	43.38	1.54	0.070 – 0.757
Euphausiacea	43.38	1.54	0.145 – 461.813
Eggs	43.38	1.54	0.070 – 1.952
Gastropoda	43.38	1.54	0.071 – 2.266
Isopoda	43.38	1.54	0.073 – 0.930
Mysidacea	43.38	1.54	0.126 – 43.504
Nemertea (pilidium)	43.38	1.54	0.082 – 0.777
Ostracoda	43.38	1.54	0.066 – 1.270
Platyhelminthes (larvae)	43.38	1.54	0.075 – 0.162
Polychaeta (larvae)	43.38	1.54	0.068 – 7.535

Commenté [A40]: Inadequate caption: the table does not give the regression equation, but the coefficients of this equation. You can give the general equation here also. Please explain what the “Area” column means: is it the range over which the coefficients are applicable (i.e. range from Lehet) or the range of areas you observed for each category in your own data?

Commenté [A41]: Larvae, I guess

Commenté [A42]: Is it a mix between nauplii of copepods and those of cirripeds?

Commenté [A43]: I suggest you choose between the name of a larva, here pilidium, or the more general term “larvae”. For example, for Bryozoa you can have cyphonautes

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930 Table 2 Integrated abundances (ind m⁻²) of the most important zooplankton taxa over the entire
 931 water column sampled on four dates throughout a year.

932

		SPRING		SUMMER		AUTUMN		WINTER	
		Day	Night	Day	Night	Day	Night	Day	Night
Cyclopoida		47,894	28,198	5,974	6,501	2,227	4,342	3,209	3,477
	Total	76,093		12,475		6,569		6,686	
Harpacticoida		48,632	47,978	9,664	12,421	9,929	7,486	6,514	4,345
	Total	96,610		22,084		17,415		10,859	
Nauplii		20,600	18,129	1,481	2,583	431	2,094	2,463	1,616
	Total	38,730		4,064		2,526		4,080	
Metridia		27,772	9,287	25,691	8,105	12,696	18,262	4,438	1,684
	Total	37,059		33,795		30,958		6,122	
Calanus		11,288	7,447	5,898	2,827	3,622	3,087	1,977	889
	Total	18,735		8,726		6,709		2,866	
Euchaetidae		1,840	4,259	1,731	1,110	790	873	1,060	733
	Total	6,091		2,840		1,663		1,793	
Cnidaria		4,895	2,546	1,520	3,713	702	2,432	4,636	2,848
	Total	7,442		5,233		3,134		7,484	
Chaetognatha		3,108	1,216	2,528	1,187	884	1,390	1,000	598
	Total	4,324		3,716		2,235		1,598	
Mysida		239	171	62	164	151	411	239	166
	Total	411		226		561		405	

Commenté [A44]: I think you cannot sum day and night abundances, because it artificially increases the abundance: the organisms present at nighttime are not added to those of the daytime, they replace them. So I suggest using a mean, not a sum.

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