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I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

Testosterone and estradiol affect adolescent reinforcement learning

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During adolescence, gonadal hormones influence brain maturation and behavior. The impact of 17 β -estradiol and testosterone on reinforcement learning was previously investigated in adults, but studies with adolescents are rare. We tested 89 German male and female adolescents (mean age \pm sd = 14.7 \pm 1.9 years) to determine to what extent 17 β -estradiol and testosterone influence reinforcement learning capacity in a response time adjustment task. Our data showed that 17 β -estradiol correlated with an enhanced ability to speed up responses for reward in both sexes, while the ability to wait for higher reward correlated with testosterone primarily in males. This suggests that individual differences in reinforcement learning may be associated with variations in these hormones during adolescence, which may shift the balance between a more reward- and an avoidance-oriented learning style.

1 **Testosterone and estradiol affect adolescent reinforcement learning**

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28 Abstract

29 During adolescence, gonadal hormones influence brain maturation and behavior. The impact of
30 17β -estradiol and testosterone on reinforcement learning was previously investigated in adults,
31 but studies with adolescents are rare. We tested 89 German male and female adolescents (mean
32 age \pm sd = 14.7 ± 1.9 years) to determine to what extent 17β -estradiol and testosterone influence
33 reinforcement learning capacity in a response time adjustment task. Our data showed, that 17β -
34 estradiol correlated with an enhanced ability to speed up responses for reward in both sexes,
35 while the ability to wait for higher reward correlated with testosterone primary in males. This
36 suggests that individual differences in reinforcement learning may be associated with variations
37 in these hormones during adolescence, which may shift the balance between a more reward- and
38 an avoidance-oriented learning style.

39 Introduction

40 Sex hormones have a great impact on adolescent (neuro-)physiological maturation. With the
41 onset of puberty at 9 to 10 years in girls and 10 to 12 years in boys, respectively, sex hormone
42 level increases rapidly (Peper & Dahl, 2013). The secretion of hypothalamic gonadotropin
43 releasing hormone (GnRH) thereby initiates the hypothalamic-pituitary-gonadal (HPA) axis.
44 GnRH stimulates the synthesis and secretion of luteinizing hormones (LH) and follicle
45 stimulating hormones (FSH) in the pituitary, which in turn contribute to the maturation of the
46 gonads and sex hormone secretion (Sisk & Foster, 2004).

47 The rising sex hormone level during adolescence significantly contributes to pubertal
48 development. With attainment of sexual maturity, sex hormones maintain the reproductive function
49 (Sisk & Foster, 2004). Neurophysiological investigations demonstrated a different impact of
50 testosterone and 17β -estradiol (E_2) on brain maturation. Testosterone has been related to an
51 increase of global white and grey matter volume in male adolescents (Peper et al., 2009, 2011),
52 whereas in female adolescents E_2 may be negatively associated with gray matter volume (Peper
53 et al., 2009). Furthermore, E_2 seems to predict white matter growth across the entire brain in both
54 sexes (Herting et al., 2014). Moreover, neurophysiological developmental changes during
55 adolescence could be better explained by hormonal and pubertal development (measured by the
56 Pubertal Development Scale or Tanner Stages) than by chronological age (Herting et al., 2014;
57 Wierenga et al., 2018).

58 Sex hormones are very important when it comes to behavior and cognitive function in
59 animals and humans. Besides the impact of E_2 and testosterone on adolescent reward-related
60 risk-taking (i.a. Op de Macks et al., 2016), an influence on reward-related learning and cognition
61 has been assumed as well (Diekhof, 2018; Hamson et al., 2016). In adult women, E_2 may
62 promote verbal memory and fluency (Hamson et al., 2016). In gonadectomized male and female
63 rats, E_2 was found to improve learning and memory even after physiological or psychological
64 stressors (Hamson et al., 2016; Khaleghi et al., 2021). Moreover, studies with castrated male rats
65 suggested that learning may be improved by testosterone treatment (Spritzer et al., 2011). In
66 healthy older men, a short-term testosterone administration improved cognitive performance
67 significantly (Cherrier et al., 2001). Findings from children (6 to 9 years old) further showed a
68 relation between moderate testosterone levels and an average intelligence (IQ between 70 and
69 130), whereas enhanced testosterone concentrations were related to high (IQ > 130), but also low
70 intelligence (IQ < 70) (Ostatníková et al., 2007). Other studies also reported enhanced

71 testosterone concentrations in children and young adolescents (6 to 13 years) with learning
72 disabilities compared to peers without impairments (Kirkpatrick et al., 1993). Given this
73 evidence, one may assume that during early adolescence balanced testosterone concentrations
74 may be important for efficient cognitive processing.

75 One way for sex hormones to modulate aspects of reward processing and reinforcement
76 learning is through the neurotransmitter dopamine. Both estradiol and testosterone can act as
77 natural dopamine-agonists, which promote dopamine release and dopaminergic transmission
78 through various physiological mechanisms (Becker, 1990; Castner et al., 1993; Pasqualini et al.,
79 1995; Sinclair et al., 2014). This is in so far important, since dopamine plays a crucial role in
80 reinforcement learning and determines how proficient individuals learn from positive or negative
81 action outcomes. It has been assumed that changes in dopamine following reward
82 prediction errors possibly act *via* two anatomically distinct pathways in the mesocorticolimbic
83 dopamine system (Maia & Frank, 2011). The activation of the *Go* pathway after the dopamine
84 burst that follows unexpected reward entails in a repetition of the same action. In turn, activation
85 of the *NoGo* pathway results from a dip in the tonic dopamine level, which facilitates learning
86 from unexpected reward reduction, omission, or even punishment. This optimally promotes an
87 adaption of action choice to maximize overall reward (Frank et al., 2004).

88 A study using a response time (RT) adaption task, called as “clock task”,
89 demonstrated this relation between dopamine and reinforcement learning by showing that
90 patients with Parkinson’s disease, but pharmacologically normalized dopamine concentration,
91 were better in the *Go learning* aspect of the task. These medicated patients thereby showed an
92 enhanced ability to speed up for a reward (i.e., better ability to acquire a higher reward through
93 quick respond after trial onset). In comparison, in an unmedicated state with
94 pathologically lowered dopamine, the same patients, demonstrated a better *NoGo learning*
95 ability. This was indicated by an increased capacity to slow down responding for reward
96 maximization (i.e., enhanced capacity to wait for higher reward) (Moustafa et al., 2008).

97 With the same task, Diekhof and colleagues characterized the impact of periodically
98 fluctuating sex hormones in women on *Go* as opposed to *NoGo learning* ability. They compared
99 the RT adaptation during the late follicular phase of the menstrual cycle, when the level of E₂
100 was high and progesterone still remained low, with the luteal phase, when progesterone neared its
101 maximum (Reimers et al., 2014), and also with the early follicular phase when both hormones
102 were at their nadir (Diekhof, 2015). Reimers and colleagues (2014) concluded that heightened E₂
103 during the late follicular phase impaired the ability to slow down for reward maximization
104 (*NoGo learning* ability), as opposed to the ability to speed up for higher reward (*Go learning*
105 capacity). Diekhof (2015) extended these findings by showing a positive correlation between E₂
106 and the ability to speed up for reward during the early follicular phase. This latter study indicated
107 a better *Go* vs. *NoGo learning* ability during the early follicular phase and assumed that the
108 boosting influence of the still increasing, yet intermediate E₂ on dopamine probably optimally
109 promotes *Go learning* ability.

110 Regarding the impact of testosterone on reward processing and reinforcement learning,
111 clinical data are currently sparse. Also, rodent studies show inconsistent findings about
112 the influence of testosterone on reward processing. It has been observed that testosterone
113 administration enhanced tyrosine hydroxylase (the rate-limiting enzymes catalyzing dopamine
114 synthesis) in the substantia nigra of gonadectomized adolescent male rats (Purves-Tyson et al.,
115 2012). Yet, testosterone may reduce tyrosine hydroxylase in gonadally intact adolescent male
116 rats in the caudate putamen (Wood et al., 2013). Furthermore, testosterone administration in

117 gonadectomized adolescent male rats enhances mRNA of the dopamine degrading enzymes
118 catechol-O-methyltransferase and monoamine oxidase in the substantia nigra (Purves-Tyson et
119 al., 2012). In contrast, testosterone led to a significant increase of dopamine in the nucleus
120 accumbens and dorsal striatum of gonadally intact male rats. Finally, in humans, testosterone has
121 been found to enhance striatal activity in the context of reward processing, while it decreased
122 activation of the striatum during punishment processing (Morris et al., 2015).

123 Previous studies with early adolescents and young adults could not show a concrete relation
124 between testosterone and performance in cognitive or reward-related tasks (Halari et al., 2005;
125 Ladouceur et al., 2019; White et al., 2020). Therefore, no clear assumptions can be made
126 regarding the influence of testosterone on *Go* and *NoGo learning*. However, in light of its
127 physiological significance for dopaminergic processing, a positive influence on reward
128 processing and the *Go learning* may be assumed.

129 *Current study*

130 In the present study, we assessed response time adjustments and learning behavior in the context
131 of reward maximization in an adolescent sample. The salivary E₂ and testosterone concentration
132 was measured on the test day, which enabled us to examine the effect of the two sex hormones
133 on *Go* and *NoGo* learning capacity. The adolescents performed an RT adjustment task, the so-
134 called *clock task* (modified by Diekhof, 2015; created by Moustafa et al., 2008). In line with
135 findings from adult research, we predicted that *Go learning*, associated with a better capability to
136 speed up responding to maximize reward, would be related to higher E₂ concentrations (e.g.
137 Diekhof, 2015; Reimers et al., 2014). Studies reporting behavioral influences of testosterone on
138 reward-related processing and especially reward learning are scarce. Whether higher testosterone
139 levels would positively influence *Go learning* as well, could not be unconditionally
140 hypothesized. Therefore, we rather explored its relationship with reinforcement learning capacity.
141 Finally, we presumed that the effects of sex hormones on reinforcement learning would be
142 different in female and male adolescents, mostly due to higher E₂ concentrations in females and
143 enhanced testosterone in males.

144 **Materials & Methods**

145 *Participants*

146 In total, 106 healthy German adolescents, between 11 and 18 years old, participated in this study.
147 All participants had no history of psychiatric or neurological disorders and assured no regular
148 medication intake. Fifteen adolescents were excluded from the analysis, because they showed a
149 random response pattern throughout the task, which suggested that the task instructions had not
150 been properly understood or that the respective participant lacked the motivation to perform the
151 task properly. Another two participants were excluded because of technical problems that left the
152 task unfinished. In sum, the data of 89 adolescents (mean age \pm SD = 14.74 \pm 1.9 years; 52
153 females) were analyzed.

154 Every participant had to sign a written declaration of informed consent prior to participation. In
155 the case of minority, a legal guardian (parent) also had to sign a written declaration of informed
156 consent before the testing. The adolescents were recruited in sports and other leisure clubs. The

157 study protocol was approved by the local ethics committee of the Ärztekammer Hamburg (Ref:
158 PV3948) and the study was conducted in accordance with “The Code of Ethics of the World
159 Medical Association” (Declaration of Helsinki).


160 On the test day, participants were screened for depressive symptoms with the validated
161 German Depression Inventory for Children and Adolescents (Stiensmeier-Pelster et al., 2014).
162 Individual cognitive capacity was tested *via* the Digit-Span Test by measuring both forward and
163 backward span from the German version of the Wechsler Intelligence Scale for Children (Wechsler,
164 2014) by counting the numbers that were correctly recalled. Self-reported trait impulsivity was
165 examined with a German Version of the Barratt Impulsiveness Scale (BIS-11) for adolescents
166 (Hartmann et al., 2011). Finally, every participant and the corresponding legal guardian filled out
167 a translated version of the Pubertal Development Scale (PDS) (Petersen et al., 1988). A mean of both
168 scores were calculated and used as an indicator of the degree of physical pubertal
development of the given participant.

169 *Experimental task*

170 A modified version of the clock task (Diekhof, 2015), that had been introduced by Moustafa
171 et al. (2008) was used. In the task, three differently colored clock faces were presented. A full
172 rotation of the clock arm lasted 5 seconds. Each clock face was assigned to one of three
173 conditions, namely the fast, the random, and the slow condition. Each of the three clock
174 conditions was shown 50 times in three sessions of 50 trials each, resulting in a total of 150
175 trials. The sequence of clock faces was pseudo-randomized and balanced for trial-type transitions
176 (Diekhof, 2015). The fast clock condition required a
177 fast reaction once the clock arm started to move, in order to maximize reward outcome. The slow
178 clock condition, in contrast, required the participant to postpone responding and slower RTs
179 yielded higher reward. The random condition served as a control variant with no contingency
180 between RT and reward outcome. It was used as an indicator of baseline response preference
181 (Figure 1).

182 The participants had to adapt to the optimal response speed in each condition to
183 maximize their overall reward. The exact reward value of each trial in the fast and slow
184 condition was calculated with a cosine function, ranging between a minimum of 15 and a
185 maximum of 60 points. The random reward value was calculated with the difference between
186 minimum and maximum points of reward multiplied by a random number and added with the
187 minimum reward value (Figure 1). In every condition, a random noise parameter (range
188 between -5 to +4 points) was applied to the reward. This was done to disguise the relation of a
189 specific reward outcome with a specific RT. Immediately after the response, the reward outcome
190 was shown to the participant. For the remaining time of a full clock arm turn, a blank screen was
191 shown. Thus, each trial had the same length. If the participant did not respond within 5 seconds,
192 no reward was presented, and the participant had to wait another 5 seconds before the next trial
193 started.

194 *Saliva collection and analyses*

195 In the morning, three saliva samples were self-collected by the participant in 2 mL Eppendorf tubes
196 at home. Sample collection took place over the course of one hour (half-hourly samples) and
197 started directly after awakening. The participants were allowed to drink water after the first 

198 sample up until 5 min before the second and third sample. They had to refrain from intake of
199 food and beverages other than water during the sampling hour. Saliva samples were stored at -
200 20°C until further use. Before analysis, samples were thawed and centrifuged at RCF 604 x g
201 (i.e., 3000 rpm in a common Eppendorf MiniSpin centrifuge) for 5 min to separate the saliva
202 from mucins. For the E₂ analysis, a 17-β-Estradiol Saliva ELISA was used (Limit of Detection:
203 2.1 pg/mL), coated with anti-17-β-Estradiol antibody (monoclonal) with antibodies derived from
204 donkey and sheep. For the testosterone analysis, a Testosterone Luminescence Immunoassay
205 (Tecan/IBL International) was utilized (Limit of Detection: 1.8 pg/mL), coated
206 with anti-mouse antibody. Intra-assay precision showed a mean CV of 8.8% (17-β-Estradiol
207 Saliva ELISA) and 7.3% (Testosterone Luminescence Immunoassay). Inter-assay precision
208 showed a mean CV of 11.8 (17-β-Estradiol Saliva ELISA) and 7.3% (Testosterone
209 Luminescence Immunoassay).

210 The three morning samples were combined in an aliquot sample that consisted of an
211 equal amount of saliva from every tube (100 μL). The analysis was done as described in the
212 respective manual in our in-house laboratory. Each sample was assayed twice. In addition, a high
213 and a low control were also analyzed. Subsequent behavioral analyses were done with standardized z-
214 transformed values ($z_i = \frac{x_i - \bar{x}}{s_x}$) for each ELISA plate to standardize the measurement inaccuracy of
215 the plates.

216 **Data preprocessing**

217 For each subject, we calculated the mean RTs of each clock type. RTs under 200 ms were
218 discarded, since they were very unlikely to reflect voluntary movements. In all, 125 trials (*mean*
219 *± sd*: 70 ± 72 ms) under 200 ms were excluded. We also calculated the mean RT of the initial 12
220 trials (called first block) and the optimized last 12 trials (called last block) for each condition
221 and participant (Diekhof, 2015; Kohne et al., 2021; Moustafa et al., 2008; Reimers et al.,
222 2014). At the beginning of the experiment (in the first block), the
223 participant did not know which clock face was associated with faster or slower responses for
224 higher reward. Hence, the participant had to try to achieve the optimal outcome via various
225 reactions exploring the task structure. Conversely, at the end of the clock task (in the last block),
226 the participant should have been well adapted and was expected to show optimal RTs that led to
227 the highest reward outcome in relation to individual clock faces.

228 Apart from the mean RT for the three clock types, the actual learning preferences that
229 reflected individual *Go* and *NoGo learning* ability, respectively, were calculated from the last
230 block. They reflected the adaption to the optimal response speed to the slow and fast clock,
231 respectively, and allowed us to test the functional opponency of *Go* versus *NoGo learning*. For
232 this, the RT of the slow and the fast clock were calculated in relation to the random clock, which
233 provided information on the individual baseline response speed of a given participant. In order to
234 calculate the optimized responses to the slow clock condition, we first subtracted the mean RT of
235 the last 12 trials of the random clock condition from the mean slow clock RT of the last block.
236 For standardization, this difference was then divided by the mean RT of the last 12 trials from
237 the random clock. The resulting standardized relative RT reflects “optimized relative slowing”.
238 Correspondingly, the subtraction of the mean fast clock RT from the mean random RT and its
239 division by the mean random RT was used as the “optimized relative speeding” value.

240 The individual learning-related change in RT for each clock condition was calculated by
241 subtracting the RT of the first block from the RT of the last block.

242 **Data analyses**

243 The behavioral data were analyzed with IBM SPSS Statistics 25. First, we performed a repeated
244 measures General Linear Model (GLM) with the factors “clock condition” (fast, random, slow),
245 “block” (first, last), “sex” (female, male) and “age” to test for possible effects of these factors on
246 the RT. In another two GLMs the factor “age” was replaced by either the covariate “pubertal
247 development” (PDS-score) or the z-standardized sex hormone concentration of E₂ (zE₂) and
248 testosterone (zT). This was done to assess the impact of pubertal
249 maturation and sex hormones level on reinforcement learning. Post hoc tests used paired and
250 independent t-tests, which were Bonferroni-corrected for multiple testing. If Levene’s test was
251 significant, Welch’s t-test instead of Student’s t-test was used. The learning preference and
252 effects of covariates were examined with a two-sided Pearson correlation. All effects and
253 differences were considered as significant below a p-value of 0.05, two-tailed.

254 **Results**

255 **Learning preference**

256 Studies with adults revealed a reverse capability for adaptive speeding vs. adaptive slowing of
257 responses in the clock task (Diekhof, 2015; Reimers et al., 2014). Our data demonstrate that this
258 reverse relation in adjustment preferences to either the slow or the fast clock may also exist in
259 adolescents. We found that optimized relative speeding and slowing were negatively correlated
260 in both sexes (*females*: $r = -.48, p < .001$; *males*: $r = -.67, p < .001$) (Figure 2). Adolescents
261 who were better adjusted to the last block of the slow clock had difficulties to speed up for
262 reward. In turn, participants who responded faster to the fast clock in the last block were
263 impaired in the ability to slow down for reward.

264 **General group characteristics**

265 The female and male adolescents did not differ in their age, impulsivity (BIS-11), and zE₂
266 concentration, which was determined by independent t-tests (Table 1). The only significant
267 differences between the two groups were significantly higher zT level in males compared to
268 females ($t_{43.95} = -6.82, p < .001, d = -1.56$) and more advanced pubertal development of
269 females compared to males ($mean_{PDS\ females} \pm se: 3.03 \pm .07$; $mean_{PDS\ males} \pm se: 2.72 \pm .09, t_{87} =$
271 $2.67, p = .009, d = .57$).

272 **Influence of age and sex on response time adjustments**

273 In an initial step, we assessed the influence of chronological age and sex of the participant on
274 learning performance. For this, we used a repeated measures GLM including the covariate “age”,
275 the between-subjects factor “sex” and the within-subject factors “clock condition” (fast, random,
276 slow) and “block” (first, last). We found a significant two-way interaction of clock
277 condition x block ($F_{2, 172} = 4.41, p = .014, \eta^2_p = .05$). This was reflected by a change in the RT
278 from the initial to the optimized last block in the fast ($t_{88} = 11.08, p < .001, d = 1.17$, Bonferroni

279 corrected for three comparisons) and slow condition ($t_{88} = -13.79, p < .001, d = -1.46,$
 280 Bonferroni corrected for three comparisons), but not in the random condition ($t_{88} = .14, p = 1, d$
 281 $= .02, Bonferroni corrected for three comparisons) (Table 2).$

282 ***Influence of pubertal development and sex on response time adjustments***

283 The first GLM was repeated with the factor “pubertal development” (measured with the PDS)
 284 replacing the factor “age”. A significant main effect of clock condition ($F_{2, 172} = 7.28 p = .001,$
 285 $\eta^2_p = .08$), a significant two-way interactions of clock condition x pubertal development ($F_2 =$
 286 $3.4, p = .036, \eta^2_p = .04$) and clock condition x sex ($F_2 = 3.81, p = .024, \eta^2_p = .04$) were emerged.
 287 Furthermore, the interaction between clock condition and block remained significant ($F_{2, 172} =$
 288 $8.04, p < .001, \eta^2 = .09$).

289 Post hoc t-tests showed a significant RT distinction between the three clock conditions
 290 (*fast vs. random*: $p < .001, d = -1.33$; *fast vs. slow*: $p < .001, d = -2.75$; *slow vs. random*: $p <$
 291 $.001, d = 1.61, Bonferroni corrected for two comparisons) (Table 2). Consequently, an$
 292 adjustment to the varying clock conditions in line with the goal of reward maximization could be
 293 assumed. Concerning the interaction between clock condition and sex, a significant difference
 294 only arose in the slow clock condition. Males reacted significantly slower and thereby better in
 295 the slow clock in general than females did ($p = .048, d = -.43$) (Table 2). The interaction of
 296 “pubertal development” and “clock condition” was reflected by a trend-wise positive correlation
 297 between the PDS and the RT of the random condition only ($r = .19, p = .068$) (Table 2).

298 ***Influence of sex hormones and sex on response time adjustments***

299 In a third GLM, we investigated the modulatory influence of zE₂ and zT as a function of the
 300 participants’ sex on RTs in the three clock conditions (fast, random, slow) and the two blocks
 301 (first, last). The main effect of “clock condition” ($F_{2, 160} = 114.83 p < .001, \eta^2_p = .81$) and the
 302 interaction of “clock condition” and “block” ($F_{2, 160} = 7.28 p < 0.001, \eta^2_p = 0.59$) remained
 303 significant. Furthermore, an interaction of block x clock condition x zE₂ concentration ($F_2 = 4.9,$
 304 $p = 0.009, \eta^2_p = 0.06$) and a main effect of block ($F_{1, 80} = 5.29 p = 0.024, \eta^2_p = 0.06$) and of zT ($F_1 =$
 305 $5.28 p = .024, \eta^2_p = .06$) occurred.

306 The interaction of block x clock condition x zE₂ was reflected by a negative correlation
 307 between zE₂ and the initial RT in the fast clock condition ($r = -.24, p = .03$) (Figure 3). In
 308 addition, we also examined the individual learning-related change in the RTs between first and
 309 last block, which demonstrated the adjustment from the initial to the optimized block (RT last
 310 block – RT first block). The learning-related change showed a significant positive correlation
 311 with zE₂ in the fast clock condition ($r = 0.28, p = .01$) (Figure 4). No correlation emerged
 312 with the slow ($r = .08, p = .497$) or random condition ($r = -.18, p = .096$).

313 A post-hoc comparison of the blocks evinced a slower response speed in the initial block
 314 compared to the last block ($t_{88} = -2.67, p = .009, d = -.28$). Further, zT was positively correlated
 315 with a slower RT independent of clock condition or block ($r = .29, p = .007$) (Figure 5).
 316 Since we found a significant difference in the zT of females and males, with higher
 317 concentrations in males (Table 1), we additionally explored the zT effect separately for both
 318 sexes. From this, it became obvious that the correlation probably emerged from the male
 319 adolescents. Accordingly, the mean of both blocks across all clocks was positively correlated

320 with zT in males ($r = .48, p = .002$), but not in females ($r = -.15, p = .298$). In males, a general
321 slowing could also be observed with increasing zT in both blocks of all conditions (*first*: $r = .37,$
322 $p = .025$, *last*: $r = .5, p = .002$) and especially in the slow ($r = .42, p = .01$) and the random ($r =$
323 $.35, p = .032$), but not in the fast condition ($r = .09, p = .579$). Additionally, in the initial ($r =$
324 $.35, p = .036$) and optimized block ($r = .44, p = .007$) of the slow clock positive correlations
325 emerged. Again, these correlations could not be found in females.

326 Discussion

327 This study examined the effects of adolescent E₂ and testosterone concentrations on RT
328 adjustments in the clock task. Results indicate individual differences in the preference for either
329 *Go* or *NoGo learning* (Figure 2) and an adaption to the different clock conditions from the
330 initial to the optimized block. Both findings have already been demonstrated previously in
331 studies with adults (Kohne et al., 2021; Moustafa et al., 2008; Reimers et al., 2014). In addition,
332 we also found that testosterone levels were significantly higher in males than females, while age,
333 impulsivity and E₂ concentrations did not differ between the sexes. We also did not observe an
334 age-dependent influence on the RT, and there was no association between individual pubertal
335 development and *Go* or *NoGo learning*. Solely, a tendency towards a slower baseline response
336 speed with increasing pubertal development emerged. Apart from that, we found a sex difference
337 in the slow clock condition. Male adolescents responded significantly slower (better adapted) to
338 the slow clock condition compared to females. E₂ and testosterone further appeared to modulate
339 learning ability in different ways. Whereas E₂ apparently enhanced initial *Go learning* (Figure 3 and 4),
340 testosterone presumably promoted *NoGo learning* ability (Figure 5), yet
341 primarily in males.

341 Similar to studies with adults, our data confirmed the detection of a preference for *Go* or
342 *NoGo learning* ability with a presumable supporting effect of E₂ on *Go learning* (Diekhof, 2018;
343 Moustafa et al., 2008; Reimers et al., 2014). Furthermore, we observed a relation between
344 habitual testosterone and the ability to slow down for reward, which was especially evident in
345 male adolescents. The observed divergence of females and males in the learning capability
346 related to the slow condition could probably be ascribed to a hormonal sex-difference. Hormonal
347 testosterone concentrations differed significantly between females and males who showed
348 enhanced concentrations. The varying increase of gonadal hormones during puberty could thus
349 be one of the reasons for the different RT adjustments in the slow clock. Accordingly, testosterone was
350 associated with a slower RT and enhanced *NoGo learning* in adolescents. Explorative
351 analysis showed that this result could be traced back to the male adolescents, most likely because
352 testosterone is the main acting gonadal hormone during male pubertal development and by far
353 more variable in pubertal males than in females. In line with adult research, E₂ seemed to
354 stimulate the initially faster responses and therefore *Go learning* in all adolescents. We speculate
355 that the effect of E₂ could have been mediated by its modulatory impact on dopaminergic
356 transmission, which has been assumed for similar findings in adult women (i.a. Diekhof,
357 2015; Reimers et al., 2014). Estrogen receptors can be found in the brain of both sexes with
358 modulating effects on neurotransmission and plasticity (Gillies & McArthur, 2010).

359 The correlation between *Go learning* and E₂ occurred exclusively in the initial block
360 during which participants were still naïve regarding the temporal reward associations of the
361 different clocks. This might indicate that E₂ has only a subtle effect on behavioral responding in

362 the clock task. Once the RT had been optimized in later phases of the task, this correlation was
363 no longer behaviorally measurable (Reimers et al., 2014).

364 Alternatively, E₂ may also support learning through a promotion of signal transduction. E₂
365 administration in young and aged ovariectomized rhesus monkeys led to an increase in spine
366 density in the dorsolateral prefrontal cortex (Hao et al., 2003). An increased spine density on
367 pyramidal neurons are connected to an enhanced number of excitatory synapses per neuron which
368 in turn might improve learning performance in general (Mahmmoud et al., 2015). Moreover, in
369 ovariectomized rats, E₂ administration provoked cell proliferation and an increase of dendritic
370 spine density in the hippocampus (Adams et al., 2002; Tanapat et al., 2005). In a previous study,
371 Davidow and colleagues (2016) demonstrated the positive impact of hippocampal activity and its
372 connectivity to the striatum on reinforcement learning in adolescents (Davidow et al., 2016).
373 Therefore, the potentiating influence of E₂ on the hippocampus may improve reward learning as
374 well. Besides E₂, androgens also positively affect prefrontal and hippocampal processing, but rat
375 studies indicate a greater impact of androgens in males (Hamson et al., 2016).

376 Similar to E₂, testosterone can modulate dopaminergic transmission and may also impact
377 transmission in other neurotransmitter systems (de Souza Silva et al., 2009; Sinclair et al., 2014).
378 The enhancing effect of testosterone on slowing ability may additionally be explained through an
379 interaction of testosterone and serotonergic processing in males. In male rats, testosterone
380 administration leads to an increase of cerebral serotonin and its metabolites (de Souza Silva et
381 al., 2009; Thiblin et al., 1999). Moreover, a positive correlation between plasma testosterone and
382 serotonin receptor 4 level emerged, leading to the suggestion that higher testosterone is
383 accompanied by a higher cerebral serotonin tonus (Perfalk et al., 2017). Therapeutic approaches
384 use, inter alia, selective serotonin reuptake inhibitors, which enhance synaptic serotonin levels
385 and modulate neuroplasticity (Kraus et al., 2017). For learning and memory formation, synaptic
386 plasticity is exceedingly important. Serotonergic impact on human behavior and
387 neurophysiological processes are commonly investigated through a depletion of the serotonin
388 precursor tryptophan. Studies with healthy humans using tryptophan depletion demonstrate a
389 slowing of responses by pharmacologically increased serotonin (Murphy et al., 2002). We
390 observed a better slowing ability with habitually increased testosterone, which might indicate
391 that this could have been an indirect effect of testosterone on serotonergic transmission. This
392 would also be in line with other studies, which shows that the effect of behavioral slowing in
393 punishment contexts, especially under high incentive motivation, disappeared, if serotonin was
394 pharmacologically depressed (Crockett et al., 2012). Lowered serotonin concentrations after
395 depletion have further been associated with decreased neural sensitivity to punishment
396 (Helmbold et al., 2015). Hence, enhanced testosterone concentration might have driven *NoGo*
397 *learning* and enabled a better slowing down for reward, through its interaction with the
398 serotonergic system.

399 Just as a recent study, we could not observe a relation between reward or punishment sensitivity
400 and the pubertal stage (Chahal et al., 2021). A lowered response speed in further developed
401 adolescents could be a consequence of reduced impulsivity, which may be an indicator of
402 neurophysiological and cognitive maturation. Similar to others, we did not find an association
403 with chronological age (Wierenga et al., 2018). Our results thus support the assumption that
404 pubertal development is a better indicator regarding cognitive performance than chronological
405 age.

406 To date, a non-invasive direct measurement of neurotransmitter processes like dopamine
407 binding or synthesis in the adolescent human brain is not feasible. We used non-invasive

408 measurements to determine steroid hormone concentrations and assessed the individual learning
409 ability for *Go* and *NoGo* learning. By combining both parameters, we tried to apply them as
410 indirect indicators of dopaminergic transmission. Besides E₂ and testosterone, other steroid
411 hormones are presumably attractive for future studies. For instance, the influence of progesterone
412 as a counterpart to E₂ on dopaminergic action may be of increased future interest. Whereas E₂ is
413 assumed to have an agonistic effect on dopaminergic transmission, progesterone supposedly
414 reduces E₂ receptor density (Selcer & Leavitt, 1988) and apparently upregulates monoamine
415 oxidase when it is administered together with E₂, which mimics the luteal phase of a natural
416 menstrual cycle (Luine & Hearn, 1990; Luine & Rhodes, 1983). Additionally, progesterone
417 enhances gamma-aminobutyric acid induced inhibition of dopaminergic neurons (Majewska et
418 al., 1986). Thus, an antagonistic and reducing effect of progesterone on dopaminergic
419 transmission has been suggested (Diekhof, 2018). In future studies, the tracking of the
420 developing menstrual cycle of the female adolescents could probably contribute to a better
421 interpretation of the opposite effects of E₂ and progesterone.

422 Finally, genetic predisposition as such has already been observed to affect reward
423 sensitivity (Richards et al., 2016), and may further interact with steroid hormone level as
424 demonstrated previously (Jakob et al., 2018; Veselic et al., 2021). In addition to previous
425 findings on receptor and transporter polymorphisms of dopamine, serotonin and sex hormones,
426 future studies could examine genetic interactions via genome-wide associations.

427 Conclusion

428 Sex hormones modulate neurophysiological processes and behavior in the context of reward
429 processing in both adult animals and humans. Yet, evidence from adolescent populations is
430 sparse. The present study assessed the impact of E₂ and testosterone on adolescent
431 reinforcement learning. Similar to female adults (e.g. Diekhof, 2015), E₂ promoted initial *Go*
432 learning in both sexes in our adolescent sample. Testosterone, in turn, enhanced *NoGo* learning
433 in males. It could be speculated that individual differences in reinforcement learning are
434 associated with variations in these hormones during adolescence, which shift the balance
435 between a reward and avoidance-related learning style.

436 Future investigations should consider further steroid hormones (e.g. cortisol,
437 progesterone) and neurophysiological processing to specify the impact of hormonal differences
438 on the dopaminergic mechanisms of reinforcement learning.

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Scheme 1

Task design

Fig. 1 A. Reward was calculated using cosine functions for the fast and slow clock. A time-independent function for the random clock was applied as control condition. B. Clock faces were presented pseudo-randomly for 5000 ms. Once a button press was made, the clock arm stopped, and immediate feedback was given. After that, a blank screen was shown for the remaining time that the clock arm would have need to complete the 5000 ms. Therefore, the blank screen ensures a constant time duration of a trial. A trial ended with the achieved points presented 1000 ms.

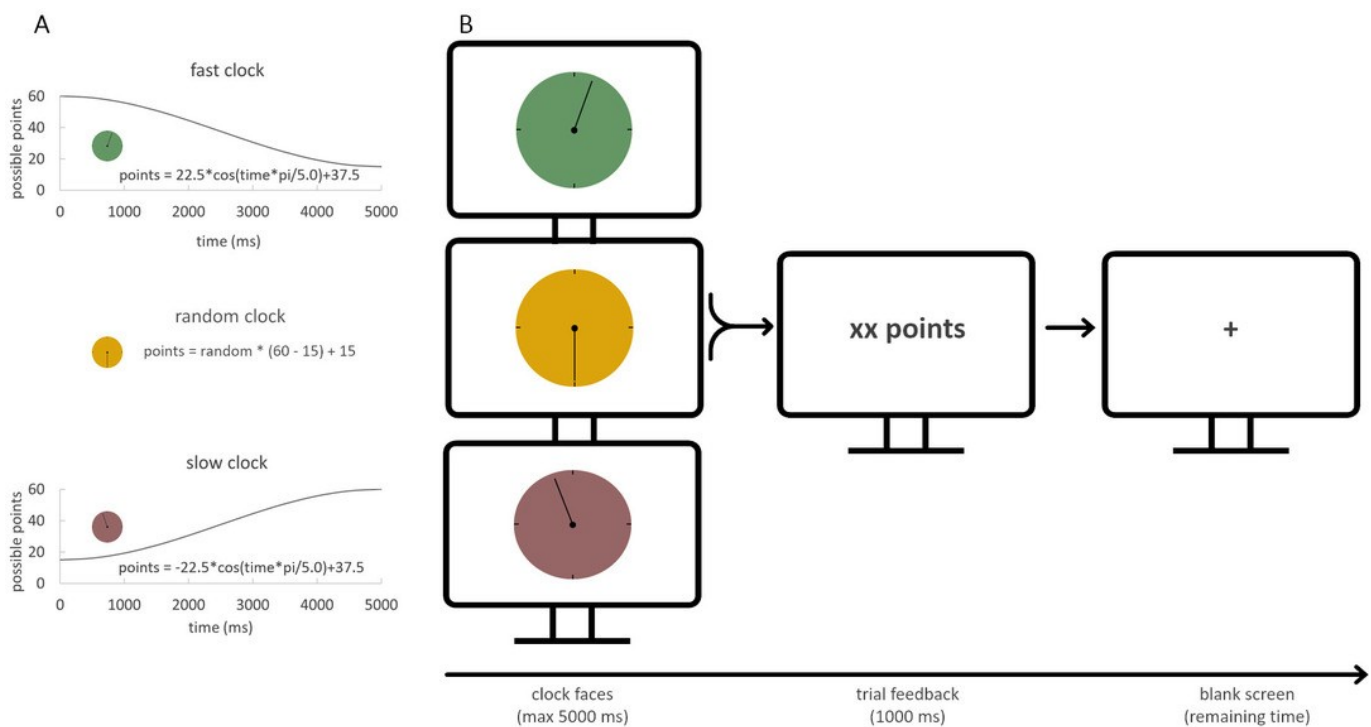


Figure 1

Reverse relation of slowing and speeding.

Optimized relative speeding and slowing were negatively correlated in females, and males ($p < .001$).

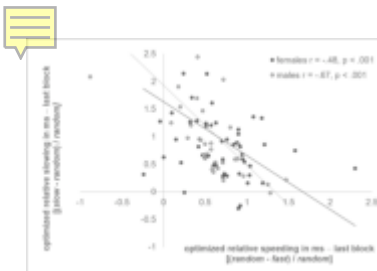


Figure 2

Negative correlation between zE_2 and the initial fast clock.

Subjects who had higher zE_2 concentrations responded faster during the initial fast clock condition ($r = -0.24$, $p = 0.03$).

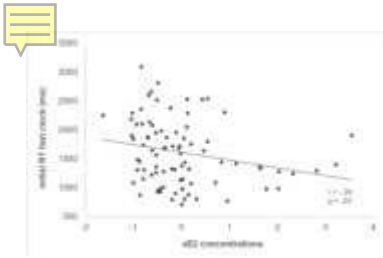


Figure 3

Positive correlation between zE_2 and the learning-related change of the fast clock.

Subjects who had lower zE_2 concentrations showed a higher adjustment from the initial to the optimized block in the fast clock condition ($r = 0.28$, $p = 0.01$).

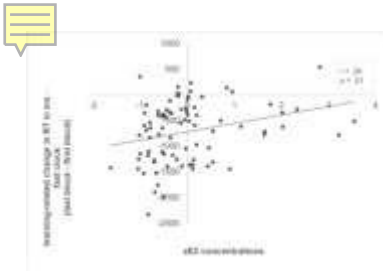


Figure 4

Positive correlation between zT and the response time of all clocks and both blocks.

Subjects who had higher zT concentrations responded generally slower ($r = 0.29$, $p = 0.007$).

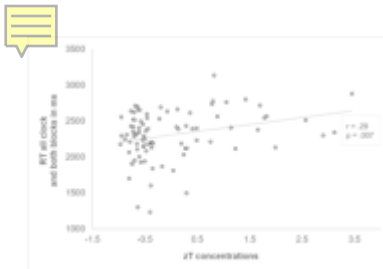


Table 1 (on next page)

Group differences by sex

| | females | | males | | females vs. males | | 95% CI | |
|----------------------------|-------------------------|----------|-------------------------|----------|--------------------|------------------|--------|--------|
| | <i>mean</i> ± <i>SD</i> | <i>n</i> | <i>mean</i> ± <i>SD</i> | <i>n</i> | <i>t</i> | <i>p</i> | lower | upper |
| | | | | | | | | |
| Age (years) | 14.67 ± 1.96 | 52 | 14.84 ± 1.83 | 37 | -.4 ^a | 0.689 | -0.98 | -65 |
| zE₂ | .14 ± 1.11 | 49 | -.2 ± 0.56 | 35 | 1.59 ^b | 0.177 | -0.09 | 0.77 |
| E₂ | 5.89 ± 2.63 pg/mL | 49 | 5.27 ± 2.08 pg/mL | 35 | 0.80 | 0.425 | -0.64 | 1.49 |
| zT | -.53 ± 0.42 | 52 | .74 ± 1.07 | 37 | -6.82 ^c | <0.001 | -1.64 | -0.89 |
| T | 21.58 ± 14.1 pg/mL | 52 | 89.61 ± 63.28 pg/mL | 37 | -6.43 ^e | <0.001 | -89.45 | -46.61 |
| BIS-11 | 63 ± 6.45 | 52 | 63.83 ± 9.57 | 36 | -0.46 ^d | 0.65 | -4.5 | 2.83 |
| PDS | 3.03 ± 0.53 | 52 | 2.72 ± 0.56 | 37 | 2.67 ^a | 0.009 | 0.08 | 0.55 |
| DICA | 11.58 ± 6.37 | 52 | 9.39 ± 3.94 | 36 | 1.99 ^e | 0.05 | -0.01 | 4.39 |
| Digit span forward | 6.31 ± 0.9 | 52 | 6.31 ± 0.79 | 36 | 0.01 ^f | 0.991 | -0.37 | 0.37 |
| Digit span backward | 4.85 ± 1.29 | 52 | 4.89 ± 1.13 | 37 | -0.17 ^a | 0.862 | -0.57 | 0.47 |

^a *t*₈₇, ^b *t*₈₂, ^c *t*_{43.95}, ^d *t*_{56.62}, ^e *t*_{85.09}, ^f *t*_{81.25}, ^g *t*_{38.55}

1 Table 1 Group differences by sex

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Table 2 (on next page)

Comprehensive summary of RTs and post-hoc results

1 **Table 2 Comprehensive summary of RTs and post-hoc results**

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| block | clock | mean RT ± SE | | | females vs. males | | | | correlations of all participants | | | | | | |
|-------------------------|-------------------|------------------------------|-----------------------------|-----------------------------|-------------------|----------------|--------|--------|----------------------------------|----------------|--------------|-----------------|------------|----------------|---|
| | | females & males | | females | males | t (df = 87) | p | 95% CI | | zT | | zE ₂ | | PDS | |
| | | | | | | | | lower | upper | r | p | r | p | r | p |
| first & last | <i>FAST</i> | 1264 ± 37ms ^{ab***} | 1302 ± 54ms | 1212 ± 293ms | 1.2 | 0.234 | -60ms | 241ms | -0.12 | 0.327 | -0.08 | 0.497 | -0.12 | 0.274 | |
| | <i>RANDOM</i> | 2196 ± 65ms ^{ac***} | 2157 ± 562ms | 2253 ± 695ms | -0.71 | 0.477 | -360ms | 170ms | 0.23 | 0.032** | -0.02 | 0.843 | 0.19 | 0.068* | |
| | <i>SLOW</i> | 3458 ± 67ms ^{bc***} | 3346 ± 653ms ^{d**} | 3617 ± 593ms ^{d**} | -2 | 0.048** | -539ms | -2ms | 0.28 | 0.009** | -0.04 | 0.731 | 0.1 | 0.359 | |
| | <i>ALL CLOCKS</i> | 2307 ± 333ms | 2269 ± 311ms | 2360 ± 360ms | -1.28 | 0.203 | -234ms | 50ms | 0.29 | 0.007** | -0.09 | 0.412 | 0.14 | 0.185 | |
| first | <i>FAST</i> | 1610 ± 58ms ^{d***} | 1655 ± 571ms | 1547 ± 524ms | 0.92 | 0.363 | -127ms | 345ms | -0.08 | 0.441 | -0.24 | 0.03** | -0.12 | 0.249 | |
| | <i>RANDOM</i> | 2203 ± 78ms | 2104 ± 685ms | 2343 ± 794ms | -1.52 | 0.132 | -552ms | 73ms | 0.18 | 0.084* | 0.08 | 0.469 | 0.1 | 0.354 | |
| | <i>SLOW</i> | 2945 ± 87ms ^{e***} | 2791 ± 807ms ^{f**} | 3163 ± 803ms ^{f**} | -2.15 | 0.034** | -717ms | -29ms | 0.3 | 0.004** | -0.06 | 0.572 | 0.1 | 0.366 | |
| last | <i>FAST</i> | 919 ± 36ms ^{d***} | 949 ± 382ms | 877 ± 275ms | 1.04 | 0.3 | -74ms | 219ms | -0.04 | 0.718 | 0.1 | 0.383 | -0.04 | 0.692 | |
| | <i>RANDOM</i> | 2190 ± 80ms | 2211 ± 703ms | 2162 ± 834ms | 0.3 | 0.765 | -276ms | 374ms | -0.14 | 0.195 | -0.12 | 0.275 | .22 | 0.038** | |
| | <i>SLOW</i> | 3972 ± 66ms ^{e***} | 3902 ± 694ms | 4071 ± 503ms | -1.33 | 0.188 | -435ms | 97ms | 0.23 | 0.03** | <.01 | 0.991 | .07 | 0.491 | |

Note: Equal letters mean significant paired t-Test results (**p < .001, *p < .05, †p < .1).

^a t₈₈ = -12.51; 95CI -1080ms, -784ms ; ^b t₈₈ = -25.93; 95CI -2362ms, 2026ms ; ^c t₈₈ = 15.2; 95CI 1097ms, 1427ms ; ^d t₈₈ = -11.08; 95CI -815ms, -567ms ; ^e t₈₈ = 13.79; 95CI 879ms, 1175ms