Fusarium head blight in the Russian Far East: 140 years after description of the 'drunken bread' problem

Tatiana Gagkaeva [Corresp., 1], **Aleksandra Orina** [1], **Olga Gavrilova** [1]

[1] Laboratory of Mycology and Phytopathology, All-Russian Institute of Plant Protection, St. Petersburg, Pushkin, Russian Federation

Corresponding Author: Tatiana Gagkaeva
Email address: t.gagkaeva@yahoo.com

The first appearance of Fusarium head blight (FHB)—and the beginning of scientific research of this disease—occurred the Far East region of Russia at the end of the 19th century. In the summer of 2019, in the Amur region, which comprises 60–70% of grain production in the Russian Far East, flooding caused a state of emergency. The quality of wheat and barley grains grown under natural conditions of FHB outbreaks, including grain infection, fungal species composition, *F. graminearum* DNA content and their chemotypes, and the presence of various mycotoxins, was studied. *Fusarium* infection rates reached extremely high percentages, 51–98%, the majority of which were *F. graminearum* infections. The amount of *F. graminearum* DNA in wheat grain samples was higher than in the barley grain samples and averaged 6.1 and 2.1 pg/ng, respectively. The content of deoxynivalenol (DON) in the wheat samples reached 13,343 ppb and in barley reached 7,755 ppb. A multilocus genotyping assay was conducted on the partially sequenced fragments of the translation elongation factor EF-1a, ammonium ligase gene, reductase gene, and 3-O-acetyltransferase gene in 29 *Fusarium graminearum sensu lato* (s. lat.) strains from the grain harvested in the Amur region. All strains from the Far East region were characterized as *F. graminearum sensu stricto*; 70% were the 15-AcDON chemotype, while the other strains were the 3-AcDON chemotype. According to the results, after 140 years of study of FHB, we are still not very successful in controlling this disease if conditions are favorable for pathogen development. Even at present, some of the grain harvested must be destroyed, as high contamination of mycotoxins renders it unusable.

Fusarium head blight in the Russian Far East: 140 years after description of the 'drunken bread' problem

Tatiana Yu. Gagkaeva, Aleksandra S. Orina, Olga P. Gavrilova

Laboratory of Mycology and Phytopathology, All-Russian Institute of Plant Protection, St. Petersburg, Pushkin 196608, Russia

Corresponding Author:
Tatiana Yu. Gagkaeva
Podbelskogo shosse, 3, St. Petersburg, Pushkin 196608, Russia
Email address: t.gagkaeva@yahoo.com

Abstract

The first appearance of Fusarium head blight (FHB)—and the beginning of scientific research of this disease—occurred the Far East region of Russia at the end of the 19th century.

In the summer of 2019, in the Amur region, which comprises 60–70% of grain production in the Russian Far East, flooding caused a state of emergency. The quality of wheat and barley grains grown under natural conditions of FHB outbreaks, including grain infection, fungal species composition, *F. graminearum* DNA content and their chemotypes, and the presence of various mycotoxins, was studied.

Fusarium infection rates reached extremely high percentages, 51–98%, the majority of which were *F. graminearum* infections. The amount of *F. graminearum* DNA in wheat grain samples was higher than in the barley grain samples and averaged 6.1 and 2.1 pg/ng, respectively. The content of deoxynivalenol (DON) in the wheat samples reached 13,343 ppb and in barley reached 7,755 ppb.

A multilocus genotyping assay was conducted on the partially sequenced fragments of the translation elongation factor EF-1a, ammonium ligase gene, reductase gene, and 3-O-acetyltransferase gene in 29 *Fusarium graminearum sensu lato* (s. lat.) strains from the grain harvested in the Amur region. All strains from the Far East region were characterized as *F. graminearum sensu stricto*; 70% were the 15-AcDON chemotype, while the other strains were the 3-AcDON chemotype.

According to the results, after 140 years of study of FHB, we are still not very successful in controlling this disease if conditions are favorable for pathogen development. Even at present, some of the grain harvested must be destroyed, as high contamination of mycotoxins renders it unusable.

Keywords: chemotype; deoxynivalenol; disease; DNA; epidemic; *Fusarium graminearum*; grain; multilocus genotyping; mycotoxins; Russian Far East.

Introduction

The first description of Fusarium head blight (FHB) within the territory of Russia was in the Far East in 1882 (Palchevsky, 1891; Voronin, 1890). This region is typically a monsoon climate with very damp and warm summers due to the influences of the Sea of Japan and the Pacific Ocean. The scientific investigation of this disease began at the end of the 19th century, but long before this, Chinese peasants and later Russian settlers related the poisoning of people and animals with pinkish grains and heads in the fields.

Between 1882 and 1914, epidemics of this disease in the Far East occurred almost every year (Naumov, 1916). Consumption of affected grain and straw caused numerous cases of food

poisoning of people and farm animals. The initial signs and symptoms of the disease resemble those that can develop after drinking too much alcohol (including dizziness and headache, trembling hands, confusion, and vomiting) and thus were named 'drunken bread'. The extensive research undertaken by Russian mycologists revealed that *Fusarium roseum* Link (*F. graminearum* Schwabe) with teleomorph stage *Gibberella saubinetii* Sacc. (*G. zeae* [Schwein.] Petch) was the principal cause of the disease (Jaczewski, 1904; Naumov, 1916; Voronin, 1890).

N.A. Palchevsky (1891), who lived in this territory and was one of the first to report the disease of grain crops, studied its etiology and deposited diseased grain head specimens in herbaria (kept in the Herbarium LEP of our laboratory, the first specimens are dated 1912). Thanks to this inquisitive individual, drawings of typical symptoms of the disease and pathogens were published (Fig. 1).

FHB was a persistent problem in the Far East during the 20th century (Abramov, 1938; Naumov, 1916) and continues to be today. High severities of FHB are reported nearly every year in the region. Mycological analyses of seed samples from 1998–2002 have shown a high level of FHB-infected wheat and barley seed (23–32%). The most frequently isolated pathogen was *F. graminearum* (Gagkaeva et al., 2002; Ivaschenko et al., 2000).

Potential toxic effects of mycotoxins associated with FHB, particularly trichothecenes, which are secondary metabolites produced by *F. graminearum*, can result in numerous health problems after consumption of infected grain, flour, and processed products. *Fusarium* outbreaks are a concern because of loss of grain yield and quality and mycotoxin contamination.

The development of multilocus sequence typing (MLST) has facilitated the identification of species and chemotypes of the *F. graminearum* species (*Fg*) group (Ward et al., 2008). Among them, the ubiquitous *F. graminearum sensu lato* (s. lat.) includes at least 16 phylogenetic species

(Aoki et al., 2012; O'Donnell et al., 2000, 2004, 2008) united into the *Fg* group. Based on MLST assays, several species of the *Fg* group, including *F. graminearum sensu stricto* (s. str.), *F. ussurianum* T. Aoki, Gagkaeva, Yli-Mattila, Kistler & O'Donnell, and *F. vorosii* B. Tóth, Varga, Starkey, O'Donnell, H. Suga & T. Aoki, were identified in the grain grown in the Russian Far East (Yli-Mattilla et al., 2009). A biogeographic hypothesis suggests that *F. vorosii*, *F. ussurianum*, and *F. asiaticum* O'Donnell, T. Aoki, Kistler & Geiser may be endemic Asian species within the *Fg* group (O'Donnell et al., 2004).

All species within the *Fg* group are capable of producing type B trichothecenes, but the activity of their formation is largely different. Three types of chemotypes have been identified among the strains: deoxynivalenol (DON) and 3-acetyldeoxynivalenol (3-AcDON), deoxynivalenol and 15-acetyldeoxynivalenol (15-AcDON), and nivalenol and 4-acetyl-nivalenol (NIV) (Moss, Thrane, 2004; Ward et al., 2002).

High humidity and heavy rainfall stimulate the development of *F. graminearum* s. lat. in grain and, as a result, increase its contamination by DON (Aldred, Magan, 2004; Ramirez et al., 2006). The Russian Far East is often exposed to weather disasters, such as floods, which can lead to negative consequences insurmountable by human efforts and technologies resulting in significant agricultural damage. In the summer of 2019, in the Amur region, which accounts for 60–70% of grain production in the Russian Far East, flooding after substantial rainfall has wreaks havoc with extensive damaged crops. In this region, an emergency regime was established on July 25, 2019, and 'about 250,000 ha was flooded, which amounted to about 20% of the total cultivated area in the region' (TASS, 2019). As a result, the yield of cereals was only partially saved and harvested.

Despite the long history of the problem in the Far East, there is still no objective information on infection and mycotoxin contamination of harvested grain. Epidemics of FHB in the region fuelled our interest in investigating this disease using available modern methods of research.

Broad geographic surveys of *Fusarium* species on cereal crops are important to establish if any present shifts in populations occur in response to environmental change. We expected the diversity of *Fusarium* species belonging to the *Fg* group in this territory to be high, since the conditions were very favorable for fungi and led to the disease epidemic. In addition, we assumed that *F. asiaticum* may appear in the complex of pathogens, since in the neighbouring countries of China and Japan this species is detected on cereals with a high frequency (Gale et al., 2002; Láday et al., 2004; Qu et al., 2007; Suga et al., 2008). In China, where the problem of FHB is also acute, two species of the *Fg* group have been identified: *F. graminearum* s. str. and *F. asiaticum* (Qu et al., 2007). In the north of China, closest to the Amur Region, mostly *F. graminearum* was dominant, and all of the 15-AcDON chemotypes (Shen et al., 2012). But *F. asiaticum* was the predominant species in the Yangtze River Basin, and chemotypes of strains were either 3-AcDON or NIV, with 3-AcDON being predominant.

This study aimed was to investigate (1) the natural *Fusarium* species occurrence and mycotoxin contamination of grain from the Amur region in the Far East in the most favorable conditions for pathogens and (2) provide the multilocus analysis of isolated strains of *F. graminearum* sensu lato to species and trichothecene genotype diversity.

Materials & Methods

Grain samples and climatic conditions of growth

In mid-August 2019, grain harvest samples were collected in various flood-rescued fields located in the Amur Region, the Russian Far East. These samples were spring wheat (nine samples of the most common Aryuna variety) and barley (four samples of the most common Acha variety). Collecting the representative sample from harvesting at these locations was approved by the Russian Science Foundation (project number: 19-76-30005).

The weather in the summer period of 2019 was characterized by disastrous excessive moisture: the total rainfall in July and August was 2.2 and 1.7 times greater, respectively, than the average means of long-term observations (according to https://rp5.ru/). In addition, the number of days with precipitation in these months was 50% and 39% more, respectively, than the average means of the climatic norm (Table 1).

Mycological analysis of grain

Microscopic examination was conducted to reveal the presence of infected grains and perithecia on seed surfaces, and photographs were taken under Olympus BX53 and Olympus SZX16 microscopes.

One hundred seeds per sample were chosen at random and surface disinfected by soaking in a 5% sodium hypochlorite solution for 1–2 min. Then the grains were washed with sterile water and put into Petri dishes on potato sucrose agar medium (PSA) containing 1 mL/L of an antibiotics solution (HyClone ™, Austria). Moreover, a commonly used detergent Triton X-100 (Panreac, Spain) which reduces the linear fungal growth (0.4 μL/L) was added. After 7-14 days of incubation in the dark at 24 °C, identification and demarcation of taxa were carried out (Gerlach, Nirenberg, 1982; Leslie, Summerell, 2006). The grain infection by the specific taxon of fungi was calculated as the ratio of the number of grains from which these fungi were isolated to the total number of analyzed grains and expressed as the incidence percentage.

142 **DNA extraction and quantification**

143 The grain samples (20 g) were homogenized separately using sterilized grinding chambers of
144 a batch mill Tube Mill Control (IKA, Königswinter, Germany). The grain flour was stored at −20
145 °C.

146 The total DNA from 200 mg of grain flour was isolated using the Genomic DNA Purification
147 Kit (Thermo Fisher Scientific, Vilnius, Lithuania) according to the manufacturer's protocol and
148 as previously described in Gagkaeva et al. (2019). Using the same kit, DNA was also isolated
149 from the mycelium of *Fusarium* spp. strains cultivated on PSA. DNA concentrations from the
150 grain samples and fungal strains were determined using a Qubit 2.0 Fluorometer with a Quant-iT
151 dsDNA HS Assay Kit (Thermo Fisher Scientific, Waltham, MA, USA). Before the start of
152 quantitative PCR (qPCR), the concentrations of all DNA samples were normalized to 23–67
153 ng/μL.

154 In every total DNA sample extracted from grain flour, the DNA content of the *F.*
155 *graminearum* and *F. avenaceum* was evaluated by qPCR with TaqMan probes (Yli-Mattila et al.,
156 2008). The reaction was carried out in a 20-μL-volume mixture with 10 μL of a 2 × TaqM master
157 mix (AlkorBio, St. Petersburg, Russia), 300 nM of each primer, 100 nM of a fluorescent sample
158 (Evrogen, Moscow, Russia), and 2 μL of the corresponding DNA solution.

159 Additionally, the DNA content of 3-AcDON and 15-AcDON chemotypes of *F. graminearum*
160 was determined using qPCR with SYBR Green (Nielsen et al., 2012). All qPCR assays were run
161 using the CFX 96 Real-Time System thermocycler (Bio-Rad, Hercules, CA, USA). All samples
162 were analyzed at least twice.

163 **Mycotoxin determination by HPLC-MS/MS**

The HPLC-MS/MS multi-mycotoxin method was used to detect different fungal secondary metabolites. In the grain samples, 3-AcDON, 15-AcDON, alternariol (AOH), alternariol monomethyl ether (AME), beauvericin (BEA), DON, deoxynivalenol-3-glucoside (DON-3gl), diacetoxyscirpenol (DAS), fumonisins B1, B2, and B3, T-2 toxin, HT-2 toxin, T-2 triol, neosolaniol (NEO), fusarenone X, moniliformin (MON), nivalenol (NIV), tentoxin (TEN), tenuazonic acid (TeA), and zearalenone (ZEN) were analyzed.

The analysis of the mycotoxins was carried out following the described procedure (Malachová et al., 2014). Detection and quantification were performed with a QTrap 5500MS/MS system (Applied Biosystems, Foster City, CA, USA) equipped with a TurboV electrospray ionization (ESI) source and a 1290 series UHPLC system (Agilent Technologies, Waldbronn, Germany). Chromatographic separation was performed at 25 °C on a Gemini® C18-column, 150 × 4.6 mm i.d., with a 5-µm particle size, equipped with a C18 SecurityGuard cartridge, 4 × 3 mm i.d. (all from Phenomenex, Torrance, CA, USA). Elution was carried out in binary gradient mode. Both mobile phases contained 5 mM of ammonium acetate and were composed of methanol/water/acetic acid ratios of 10:89:1 (v/v/v; eluent A) and 97:2:1 (v/v/v; eluent B), respectively. The recovery of mycotoxins from grain ranged from 79% to 105%.

Genotyping of *Fusarium* spp.

Among isolated fungi that were morphologically assigned to the *Fg* group (nearly 900), 29 monoconidial strains were randomly selected for further molecular analysis. Additionally, four related *Fusarium* strains with various geographic and substrate origins, the taxonomic status of which requires appraisal, were included in the study (Table 2).

To assess the phylogenetic relationships between all the strains tested, fragments of the translation elongation factor EF-1a (*TEF*), ammonium ligase gene (*URA*), reductase gene (*RED*),

and 3-O-acetyltransferase gene (*Tri101*) were used. Their amplification was carried out using specific primers EF1/EF2, URA11/URA16, RED1d/RED2, and TRI1013E/TRI1015B, respectively, according to the authors' protocols and instructions (O'Donnell et al., 2000, 2004, 2008).

The sequencing was carried out on an ABI Prism 3500 sequencer (Applied Biosystems, Hitachi, Japan) using the BigDye Terminator v3.1 cycle sequencing kit (Applied Biosystems, USA). To address the phylogenetic relationships among taxa maximum likelihood (ML), maximum parsimony (MP) analysis was conducted using the MEGA X 10.2 program (Kumar et al., 2018) as well as Bayesian posterior probability (BP) by MrBayes v. 3.2.1 on the Armadillo 1.1 platform (Lord et al., 2012). Nodal support was assessed by bootstrap analysis on 1,000 replicates. Sequence data were deposited in GenBank.

The *Fusarium* spp. a chemotype (3-AcDON, 15-AcDON, or NIV) was determined using PCR with primers Tri13P1/Tri13P2 according to the authors' protocols and instructions (Wang et al., 2008).

All tested *Fusarium* strains are maintained in the collection of the Laboratory of Mycology and Phytopathology at the All-Russian Institute of Plant Protection.

Statistical analysis

Data were analyzed using Microsoft Office Excel 2010 (Microsoft, Redmond, WA, USA) and Statistica 10.0 (StatSoft, Tulsa, OK, USA). The significance of differences between mean values was estimated by Tukey's test (95% confidence level).

Results

Detection of grain infection with fungi

Visual analysis of grain samples revealed the presence of various deformities, shrunken and with a pink-white coloration of grain heads in the amount of 5–42% (Fig. 2.). Due to prolonged wet weather, the salmon-orange conidia masses of the fungus and blue-black perithecia can be seen on the infected spikelet and glumes in barley. Most of the perithecia were mature, and when placed in a water drop, the ascospores with three septa appeared from asci.

The average germination of wheat grain was 25.1% (12–41%) and of barley grain was 55.3% (48–62%). Almost 100% infection by fungi of all grain samples was noted; often, different fungi were isolated from one grain.

Mycology analyses verified that infection by *Fusarium* spp. was the primary cause of damage in grains, and infection rates reached extremely high percentages (Table 3). Moreover, the proportion of *F. graminearum* s. lat. strains among all isolated *Fusarium* spp. averaged 83.7% in the wheat grain and 89.7% in the barley grain. *Fusarium sporotrichioides* Sherb. strains were detected in 61% of samples, but grain infection was low (1–4%) (the supplemental table). Among the isolated fungi, the occurrence of *F. avenaceum* (Fr.) Sacc., *F. anguioides* Sherb., *F. tricinctum* (Corda) Sacc., *F. poae* (Peck) Wollenw., *F. cerealis* (Cooke) Sacc., *F. equiseti* (Corda) Sacc., *F. incarnatum* (Desm.) Sacc., and *F. heterosporum* Nees et T. Nees as well as four strains belonging to the *Fusarium fujikuroi* species complex were lower (the supplemental table).

Alternaria spp. were the second frequent genera isolated from the grains. Moreover, the infection of wheat grain with *Alternaria* spp. was almost two times lower (12.9%) than that of barley grain (21.5%). *Cladosporium* spp., *Clonostachys rosea* (Link: Fr.) Schroers, Samuels, Seifert & W. Gams, *Cochliobolus* spp., *Epicoccum nigrum* Link, and other fungi were also identified in the grain mycobiota (the supplemental table).

234 **Quantification of *Fusarium* biomass**

235 The amount of *F. graminearum* DNA in grain flour was very high, averaging 4.9 pg/ng
236 (Table 3). In analyzed samples of wheat grain, the amount of *F. graminearum* DNA was higher
237 than in the barley grain samples ($p = 0.032$). The amount of 3-AcDON *F. graminearum* DNA
238 was on average 1.3–1.1 times higher than the content of 15-AcDON genotype DNA. *F.*
239 *avenaceum* DNA was detected in all grain samples in an amount that was on average 160 times
240 less than that of *F. graminearum* DNA.

241 **Detection of mycotoxins**

242 DON was found in all samples. The content of DON reached 13,343 ppb in wheat samples
243 and 7,755 ppb in barley samples. In all analyzed samples, the content of DON exceeded the
244 maximum permissible limits (MPLs) in grain for food (700 ppb for wheat grain, 1,000 ppb for
245 barley grain) and fodder (1,000 ppb for cereal grain), by up to 13 times (TR TS 015/2011; TR TS
246 021/2011). The exception was one barley sample, in which the DON content was lower than the
247 MPL: 911 ppb.

248 In addition, other type B trichothecene mycotoxins, 3-AcDON, 15-AcDON, and DON-3gl,
249 were detected in the grain. Of the total content of trichothecenes, the share of DON in wheat
250 grain was 86.5% and in barley grain was 69.5%.

251 The content of ZEN produced by *F. graminearum* in wheat grain (92–3,670 ppb) was on
252 average 2.1 times higher than in barley grain (111–928 ppb).

253 Low contents of T-2 toxin (5 and 15 ppb) and HT-2 toxin (23 and 58 ppb) produced by *F.*
254 *sporotrichioides* were detected in two barley grain samples.

255 The MON produced by *F. avenaceum* was detected in all samples in amounts up to 218 ppb
256 without differences between crops. The mycotoxin BEA was detected in only two wheat samples

in amounts up to 13 ppb. The fumonisins NEO, DAS, and fusarenone X produced by *Fusarium* fungi were not detected in the analyzed grain samples.

The mycotoxin AOH produced by *Alternaria* fungi was detected in all grain samples in small amounts (8–49 ppb). Moreover, the content of this mycotoxin in barley grain, 11.7 (7.6–17.2), was significantly lower than in wheat grain, 29.0 (14.2–49.1) ($p = 0.032$). AME was found in all analyzed grain samples except for two wheat samples in trace amounts. TeA was detected in all barley grains with a maximal level of 37.4 ppb and in 44% of wheat samples with a maximal level of 75.0 ppb (the supplemental table). Traces of TEN were found in all samples (max 6.4 ppb).

Genotyping of *Fusarium* spp.

Multilocus analyses of the *TEF*, *URA*, *RED*, and *Tri101* sequences were used to determine the genetic relationships among *Fusarium* strains. The dataset included 34 combined sequences of the analyzed strains as well as the 12 reference sequences of *Fusarium* spp. belonging to the *Fg* group and consisted of a total of 2,941 characters (612 bp from the *TEF*, 558 bp from *URA*, 821 bp from *RED*, and 950 bp from *Tri101*). The sequence of the *F. pseudograminearum* type strain NRRL 28334 was used as the outgroup. The resulting phylogenetic tree based on DNA sequence data of *Fusarium* species was constructed (Fig. 3). Maximum likelihood and MP bootstrap support values greater than 70%, followed by Bayesian posterior probability scores greater than 0.95, are shown at the nodes.

The topology of phylogenetic trees constructed by different methods turned out to be similar and demonstrated the phylogenetic relationships between species established earlier (Aoki et al., 2012). Twenty-nine analyzed *Fusarium* strains isolated from Amur grain belonged to the clade with reference strain NRRL 5883 *F. graminearum* s. str. (Fig. 3). Among the analyzed strains of

F. graminearum s. str., nine strains were the 3-AcDON chemotype while 21 strains turned out to be the 15-AcDON chemotype (Table 2).

From four doubtful *Fusarium* strains, one strain MFG 60604, isolated from wheat grain from the Altai Krai (Western Siberia), was clustered with the reference strain *F. vorosii* NRRL 45790 with high bootstrap support (ML/MP/BP: 99/99/1.0). Our phylogenetic analysis indicates that strain MFG 60604 is *F. vorosii* and it is determined as a 15-AcDON chemotype.

Three other doubtful strains, MFG 58836, MFG 59052, and MFG 60755, formed the clade with the reference strains *F. culmorum* NRRL 25475 with high bootstrap support (ML/MP: 98/100). All three *F. culmorum* strains were the 3-AcDON chemotype (Table 2).

Discussion

Despite the long history of the FHB problem in the Russian Far East, objective data on pathogen composition and content of mycotoxins in naturally infected grain is clearly under-published. The mycological analyses of grain from this region in 2019 revealed extremely high infection of grain with *Fusarium* spp.—up to 98%. The predominant cause of FHB was the *Fg* group, which accounted for 86% of all isolated *Fusarium* spp.

Interestingly, the amount of fungal DNA in the wheat grain was on average higher than in the barley grain, while the percentage of infected grains was the same. The revealed differences may be due to the abundance of fungal biomass concentrated on the surface of barley grains (husk, palea, pericarp), while the wheat grain is completely permeated with fungal hyphae. In general, in this situation in 2019, the infection rates for both wheat and barley were off the scale. In our opinion, the highest DON content detected in this study, in the amount of 13,343 ppb, exceeds the maximum amounts of this mycotoxin in grain previously detected in the Russian territory. During the outbreak of FHB in southern Europe in 1985–1991 the maximal content of DON in

303 grain reached 10,000 ppb (Kononenko, 2005). Recently, in 2017, a DON amount of 7,920 ppb
304 was detected in wheat grain grown in southern Europe (Kononenko et al., 2020).
305 The content of 3-AcDON in wheat and barley grain, as well as 15-AcDON, was similar and
306 did not exceed 293 ppb. In the plant, DON can be present as a metabolite, DON-3gl, which is
307 represents up to 46% of the total amount of DON in infected wheat and maize varieties
308 (Berthiller et al. 2009). It has been shown that DON-3gl can be converted back to DON in
309 mammals (Dall'Erta et al., 2013; Tucker et al., 2019). Therefore, DON-3gl is also frequently
310 referred to as a masked mycotoxin. In our study, the maximum content of DON-3gl reached
311 3,803 ppb and was twice as high, on average, in barley grain than in wheat grain. The amounts of
312 DON-3gl come to 13.5% and 39.5% of the total amounts of DON in infected wheat and barley
313 samples, respectively. However, there were no significant differences in the content of the
314 trichothecene mycotoxin average between wheat and barley grains.
315 Using morphology to accurately assess species limits for the *Fg* group is not reliable. Before
316 this study, we hypothesized that in the extremely humid and warmest conditions of 2019, in the
317 area where FHB outbreaks were observed for at least 140 years, several species of the Asian
318 clade of the *Fg* group will be identified. Especially considering that earlier we have already
319 found three species of the *Fg* group in this region: *F. graminearum* s. str., *F. ussurianum* and *F.*
320 *vorosii* (Yli-Mattila et al., 2009). Selecting freshly isolated fungi for analysis, we took cultures
321 for a detailed study, which included all the morphological diversity present within the limits
322 possible for the *Fg* group (pigmentation, rate of formation of macroconidia, size, and shape).
323 Multilocus phylogenetic analysis revealed that all strains from the Amur grains belonged to the
324 *F. graminearum* s. str.

Molecular methods make it possible to reveal the intraspecific diversity of *F. graminearum* and to establish the quantitative presence of two different chemotypes. The *F. graminearum* strains are divided into 3-AcDON and 15-AcDON chemotypes depending on the prevailing acetylated form of DON (Alexander et al., 2011; Foround et al., 2019). Regional differences have been reported regarding the occurrence of chemotypes within the *Fg* group (Foround et al., 2019; Pasquali et al., 2016). In our study, on average, the DNA content of the 3-AcDON and 15-AcDON fungus chemotypes in the grain was similar, but the DNA of the 15-AcDON chemotype in wheat grain was significantly higher (4.6 times) than in barley ($p = 0.014$), whereas the difference in DNA content of the 3-AcDON fungus chemotype in wheat and barley grain was insignificant. It is not known whether the observed differences are related to chemotype-specific plant-host preferences. There may be a difference in pathogenicity between the 3- and 15-ADON chemotypes to wheat and barley (Foroud et al., 2019; Clear et al., 2013).

According to our results, 30% of the analyzed *F. graminearum* strains were the 3-AcDON chemotype, while 70% of the strains were the 15-AcDON chemotype. Previously, the chemotype analysis of the 105 *F. graminearum* strains collected in the Russian Far East in 1998–2006 revealed the approximately equal occurrence of 3-AcDON (48%) and 15-AcDON (52%) chemotypes (Yli-Mattila et al., 2009). An increase in the 15-ADON chemotype has recently been shown in regions of Europe, where the 3-ADON chemotype was previously dominant, although many of the factors affecting their distribution are still unclear (Nielsen et al., 2012; Aamot et al., 2015; Pasquali et al., 2016; Foround et al., 2019). The third chemotype of *F. graminearum* s. str. producing nivalenol (NIV) has not yet been identified in Russia or China (Shen et al., 2012), although it is known to be found in Europe (Pasquali et al., 2016).

In our analysis, *Fusarium* sp. strain MFG 60604 was included that was isolated from wheat grain in the Western Siberia region (the Altai Krai); phenotypically, this strain was a dubious representative of the *Fg* group. In this region, the occurrence of *F. graminearum* was previously not typical, but in recent years, we have been identifying this pathogen in cereal grains (Gagkaeva et al., 2019). The strain MFG 60604, isolated from wheat grain from West Siberia, was clustered with the reference strain *F. vorosii* NRRL 45790 with high bootstrap support (ML/MP/BP: 98/99/1.0), which allows for accurate establishment of its species affiliation. A single strain (MFG 60604) identified as *F. vorosii* in this study, is the only third strain of *F. vorosii* found in Russia and the first one identified in the Siberian region. Previously identified strains of *F. vorosii* from the Russian Far East belonged to the 15-AcDON chemotype (Yli-Mattila et al., 2009) and so did the strain identified in this study. However, among six *F. vorosii* strains originating from Korea, five were the NIV chemotype, while only one was the 15-AcDON (Lee et al., 2016). Among *F. vorosii*, no strains of the 3-AcDON chemotype have been identified, which, probably, were not detected due to the small number of strains of this species analyzed to date. In the limited surveys to date, strains of several species of the *Fg* group were found to represent only a single chemotype (Aoki et al., 2012).

Two strains of *F. culmorum* of closely related taxon to *Fg* group from the Western Siberia and Ural regions and one from the South European region of Russia were included in the study. The high genetic similarity of analyzed *F. culmorum* strains collected from remote regions characterized by different climatic conditions (the distance between isolation points is about 2,500 km) is consistent with the previously shown information that *F. culmorum* is a single phylogenetic species with little or no differences between lineages, despite the geographic separation of genotypes (Obanor et al., 2010).

The studies analyzing the occurrence of *F. culmorum* chemotypes in different regions, as a rule, show a significant excess of the occurrence of the DON chemotype compared to the NIV chemotype (Laraba et al., 2017; Pasquali et al., 2016; Scherm et al., 2012). Strains of the 15-AcDON chemotype typical for *F. graminearum* were not identified among the strains of *F. culmorum*. A previous analysis of a few strains of *F. culmorum* from the Russian territory has characterized them as the 3-AcDON chemotype (Yli-Mattila et al., 2009).

Fusarium spp. continue to pose a threat to farmers, destroying crops or dramatically reducing yields, as well as to animal and human health due to the production of mycotoxins. Even in our time, when we know much more about the nature of *Fusarium* spp. then 140 years ago, we are still not very successful in controlling the diseases they cause on crops if conditions are favorable for the development of pathogens. Indeed, in the process of our study, it was shown in the mass media that although the grain was harvested with great difficulty, due to the significant contamination of the grain, part of the crop, 240 tons, had to be destroyed by fire.

Conclusions

The high prevalence of Fusarium head blight in cereal grains cultivated in the Far East is particularly alarming and strongly indicates the need for increased measures to prevent plant infection and improved food safety interventions. The maximum DON content in wheat grains reached 13,141 ppb in this study. The multilocus sequence revealed that the majority of the strains used in this study belonged to *F. graminearum* s. str.

Acknowledgments

We are grateful to the managers of the Russian Branch of BASF company for their help with the grain sample collection and Nadezhda N. Gogina from the All-Russian Scientific and Technological Institute of Poultry (Moscow region) for carrying out HPLC-MS/MS analysis.

References

Aamot HU, Ward TJ, Brodal G, Vrålstad T, Larsen GB, Klemsdal SS, Elameen A, Uhlig S, Hofgaard IS. 2015. Genetic and phenotypic diversity within the *Fusarium graminearum* species complex in Norway. European Journal of Plant Pathology **142:** 501–519 https://doi.org/10.1007/s10658-015-0629-4

Abramov IN. 1938. Diseases of agricultural plants on the Far East. Far Eastern State Regional Publishing House, Khabarovsk. (in Russian)

Aldred D, Magan N. 2004. Prevention strategies for trichothecenes. Toxicology Letters **153:**165–171 https://doi.org/10.1016/j.toxlet.2004.04.031

Alexander NJ, McCormick SP, Waalwijk C, van der Lee T, Proctor RH. 2011. The genetic basis for 3-ADON and 15-ADON trichothecene chemotypes in *Fusarium graminearum*. Fungal Genetics and Biology **48:**485–495 https://doi.org/10.1016/j.fgb.2011.01.003

Aoki T, Ward T, Kistler H, O'Donnell K. 2012. Systematics, phylogeny and trichothecene mycotoxin potential of Fusarium head blight cereal pathogens. Mycotoxins **62:**91–102 https://doi.org/10.2520/myco.62.91

Clear RM, Tucker JR, Gaba D, Patrick SK, Lee SJ, Demeke T, Tittlemier SA, Legge WG, Gräfenhan T. 2013. Deoxynivalenol levels and chemotype frequency in barley cultivars inoculated with two chemotypes of *Fusarium graminearum*. Canadian Journal of Plant Pathology **35:** 37–45 https://doi.org/10.1080/07060661.2012.751622

418 Dall'Erta A, Cirlini M, Dall'Asta M, Rio DD, Galaverna G. 2013. Masked mycotoxins are
419 efficiently hydrolyzed by human colonic microbiota releasing their aglycones. Chemical
420 Toxicology Research **26:**305–312. http://dx.doi.org/10.1021/tx300438c
421 Foround NA, Baines D, Gagkaeva TYu, Thakor N, Badea A., Steiner B., Bürstmayr M,
422 Bürstmayr H. 2019. Trichothecenes in cereal grains – an update. Toxins **11:**634
423 http://dx.doi.org/10.3390/toxins11110634
424 Gagkaeva T, Gavrilova O, Orina A, Lebedin Y, Shanin I, Petukhov P, Eremin S. 2019.
425 Analysis of toxigenic *Fusarium* species associated with wheat grain from three regions of
426 Russia: Volga, Ural, and West Siberia. Toxins **11:**252 https://doi.org/10.3390/toxins11050252
427 Gagkaeva TYu, Levitin MM, Zuev E, Terentjeva I. 2002. Evaluation of genetic resources
428 of wheat and barley from Far East of Russia for resistance to Fusarium head blight. Journal of
429 Applied Genetics **43A:**229–236
430 Gale LR, Chen L–F, Hernick CA, Takamura K, Kistler HC. 2002 Population analysis of
431 *Fusarium graminearum* from wheat fields in Eastern China. Phytopathology **92:**1315–1322
432 https://doi.org/10.1094/PHYTO200292121315
433 O'Donnell K, Ward TJ, Geiser DM, Kistler HC, Aoki T. 2004. Genealogical
434 concordance between the mating type locus and seven other nuclear genes supports formal
435 recognition of nine phylogenetically distinct species within the *Fusarium graminearum* clade.
436 Fungal Genetics and Biology **41**: 600–623 https://doi.org/10.1016/j.fgb.2004.03.003
437 Gerlach W, Nirenberg HI. 1982 The genus *Fusarium*—A Pictorial Atlas. In: *Mitteilungen*
438 *aus der Biologischen Bundesanstalt fur Land- und Forstwirtschaft, Berlin-Dahlem*;
439 Herausgegeben von der Biologischen Bundesanstalt für Land- und Forstwirtschaft, Berlin, 209

Ivashchenko VG, Shipilova NP, Levitin MM. 2000. Species composition of *Fusarium* fungi on cereals in the Asian part of Russia. Mikologia I Fitopatologia **34:**54–68 (In Russian)

Jaczewski AA. 1904. About drunken bread. Sheet for diseases and damages of cultivated and wild useful plants **11:**89–92 (In Russian)

Karlovsky P. 2011. Biological detoxification of the mycotoxin deoxynivalenol and its use in genetically engineered crops and feed additives. Applied Microbiology and Biotechnology **91:**491–504 https://doi.org/10.1007/s00253-011-3401-5

Kononenko GP. 2005. The system of mycotoxicological control of objects of veterinary, sanitary and environmental supervision. Theses of the doctoral dissertation, Moscow

Kononenko GP, Burkin AA, Zotova YeV. 2020. Mycotoxicological monitoring Part 3 Feedstuffs from raw grain processing. Veterinary Science Today **3:**213–219 https://doi.org/10.29326/2304-196X-2020-3-34-213-219

Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: Molecular evolutionary genetics analysis across computing platforms. Molecular Biology and Evolution **35:**1547–1549 https://doi.org/10.1093/molbev/msy096

Láday M, Juhász. Á, Mulé G, Moretti A, Szécsi A, Logrieco A. 2004. Mitochondrial DNA diversity and lineage determination of European isolates of *Fusarium graminearum* (*Gibberella zeae*). European Journal Plant Pathology **110:**545–550 https://doi.org/10.1023/B:EJPP0000032394391302c

Laraba I, Boureghda H, Abdallah N, Bouaicha O, Obanor F, Moretti A, Geiser DM, Kim H-S, McCormick SP, ProctorRH, KellyAC, Ward TJ, O'Donnell K. 2017. Population genetic structure and mycotoxin potential of the wheat crown rot and head blight pathogen *Fusarium*

culmorum in Algeria. Fungal Genetics and Biology **103:**34–41 https://doi.org/10.1016/jfgb201704001

Lee T, Paek J-S, Lee K, Lee S, Choi J-H, Ham H, Hong S, Ryu J-G. 2016. Occurrence of toxigenic *Fusarium vorosii* among small grain cereals in Korea. Plant Pathology Journal **32:**407–413 https://doi.org/10.5423/PPJOA0520160123

Leslie JF, Summerell BA. 2006. The *Fusarium* laboratory manual. Blackwell Publishing, Ames https://doi.org/10.1002/9780470278376

Lord E, Leclercq M, Boc A, Diallo AB, Makarenkov V. 2012. Armadillo 11: An original workflow platform for designing and conducting phylogenetic analysis and simulations. PLoS ONE **7:**e29903 https://doi.org/10.1371/journalpone0029903

Malachová A, Sulyok M, Beltrán E, Berthiller F, Krska. R. 2014. Optimization and validation of a quantitative liquid chromatography–tandem mass spectrometric method covering 295 bacterial and fungal metabolites including all regulated mycotoxins in four model food matrices. Journal of ChromatographyA **1362:**145–156 https://doi.org/10.1016/jchroma201408037

Moss MO, Thrane U. 2004. *Fusarium* taxonomy with relation to trichothecene formation. Toxicology Letters **153:**23–28 https://doi.org/10.1016/jtoxlet200404021

Naumov NA. 1916. Drunken bread Observations under some species of genus *Fusarium* Bureau of Mycology and Phytopathology, Petrograd (In Russian)

Nielsen LK, Jensen JD, Rodríguez A, Jørgensen LN, Justesen AF. 2012. TRI12 based quantitative real-time PCR assays reveal the distribution of trichothecene genotypes of *F graminearum* and *F culmorum* isolates in Danish small grain cereals. International Journal of Food Microbiology **157:**384–392 https://doi.org/10.1016/jijfoodmicro201206010

O'Donnell K, Kistler HC, Tacke BK, Casper HH. 2000. Gene genealogies reveal global phylogeographic structure and reproductive isolation among lineages of *Fusarium graminearum*, the fungus causing wheat scab. PNAS **97:**7905–7910 https://doi.org/10.1073/pnas130193297

O'Donnell K, Ward TJ, Aberra D, Kistler HC, Aoki T, Orwig N, Kimura M, Bjornstad A, Klemsdal SS. 2008. Multilocus genotyping and molecular phylogenetics resolve a novel head blight pathogen within the *Fusarium graminearum* species complex from Ethiopia. Fungal Genetics and Biology **45:**1514–1522 https://doi.org/10.1016/jfgb200809002

O'Donnell K, Ward TJ, Geiser DM, Kistler HC, Aoki T. 2004. Genealogical concordance between the mating type locus and seven other nuclear genes supports formal recognition of nine phylogenetically distinct species within the *Fusarium graminearum* clade. Fungal Genetics and Biology **41:**600–623 https://doi.org/10.1016/jfgb200403003

Obanor F, Erginbas-Orakci G, Tunali B, Nicol JM, Chakraborty S. 2010. *Fusarium culmorum* is a single phylogenetic species based on multilocus sequence analysis. Fungal Biology **114:**753–765 https://doi.org/10.1016/jfunbio201007001

Palchevsky NA. 1891 Diseases of cultivated cereals of the South Ussuri region Public Benefit Partnership, Saint Petersburg (In Russian)

Pasquali M, Beyer M, Logrieco A, Audenaert K, Balmas V, Ryan Basler R, BoutignyA-L, Chrpová J, Czembor E, Gagkaeva T, González-Jaén MT, Hofgaard IS, Köycü ND, Hoffmann L, Lević J, Marin P, Miedaner T, Migheli Q, Moretti A, Müller MEH, Munaut F, Parikka P, Pallez-Barthel M, Piec J, Scauflaire J, Scherm B, Stanković S, Thrane U, Uhlig S, Vanheule A, Yli-Mattila T, Vogelgsang S. 2016. A European database of *Fusarium graminearum* and *F culmorum* trichothecene genotypes. Frontiers in Microbiology **7:**406 https://doi.org/10.3389/fmicb201600406

Qu B, Li HP, Zhang JB, Xu YB, Huang T, Wu AB, Zhao CS, Carter J, Nicholson P, Liao YC. 2007. Geographic distribution and genetic diversity of *Fusarium graminearum* and *Fusarium asiaticum* on wheat spikes throughout China. Plant Pathology **57:**15–24 https://doi.org/10.1111/j1365-3059200701711x

Ramirez ML, Chulze S, Magan N. 2006. Temperature and water activity effects on growth and temporal deoxynivalenol production by two Argentinean strains of *Fusarium graminearum* on irradiated wheat grain. International Journal of Food Microbiology **106:**291–296 https://doi.org/10.1016/jijfoodmicro200509004

Scherm B, Balmas V, Spanu F, Pani G, Delogu G, Pasquali M, Migheli Q. 2012. *Fusarium culmorum*: causal agent of foot and root rot and head blight on wheat. Molecular Plant Pathology **14:**1–19 https://doi.org/10.1111/mpp12011

Shen CM, Hu YC, Sun HY, Li W, Guo JH, Chen HG. 2012. Geographic distribution of trichothecene chemotypes of the *Fusarium graminearum* species complex in major winter wheat production areas of China. Plant Disease **96:**1172–1178 https://doi.org/10.1094/PDIS-11-11-0974-RE

Suga H, Karugia GW, Ward T, Gale LR, Tomimura K, Nakajima T, Miyasaka A, Koizumi S, Kageyama K, Hyakumachi M. 2008. Molecular characterization of the *Fusarium graminearum* species complex in Japan. Phytopathology **98:**159–166 https://doi.org/10.1094/PHYTO-98-2-0159

TASS, 2019. The flood damage in 2019 in the Amur region is estimated at more than 6 billion rubles. 16 Sept. 2019. https://tass.ru/ekonomika/6888626 (In Russian)

TR TS 015/2011 Technical regulation of the Customs Union 015/2011 "About grain safety" as amended on September 15, 2017 Supplement No 2 Available online:

http://www.eurasiancommission.org/ru/act/texnreg/deptexreg/tr/Documents/TP_зерно.pdf (In Russian) (accessed 7 May 2021)

TR TS 021/2011 Technical regulation of the Customs Union 021/2011 " About food safety" as amended on August 8, 2019 Supplement No 3 Available online: http://www.eurasiancommission.org/ru/act/texnreg/deptexreg/tr/Documents/TR%20TS%20PishevayaProd.pdf (In Russian) (accessed 7 May 2021)

Tucker JR, Badea A, Blagden R, Pleskach K, Tittlemier SA, Fernando WGD. 2019. Deoxynivalenol-3-glucoside content is highly associated with deoxynivalenol levels in two-row barley genotypes of importance to Canadian barley breeding programs. Toxins **11:**319 https://doi.org/10.3390/toxins11060319

Voronin MC. 1890. About drunken bread in the South Ussuri region. Botanical notes **3:**13–21 (In Russian)

Wang J, Li H, Qu B, Zhang J, Huang T, Chen F, Liao Y. 2008. Development of a generic PCR detection of 3-acetyldeoxynivalenol, 15-acetyldeoxynivalenol- and nivalenol-chemotypes of *Fusarium graminearum* clade. International Journal of Molecular Sciences **9:**2495–2504 https://doi.org/10.3390/ijms9122495

Ward TJ, Bielawski JP, Kistler HC, Sullivan E, O'Donnell K. 2002. Ancestral polymorphism and adaptive evolution in the trichothecene mycotoxin gene cluster of phytopathogenic *Fusarium.* PNAS **99:**9278–9283 https://doi.org/10.1073/pnas142307199

Ward TJ, Clear R, Rooney A, O'Donnell K, Gaba D, Patrick S, Starkey DE, Gilbert J, Geiser DM, Nowicki TW. 2008. An adaptive evolutionary shift in Fusarium head blight pathogen populations is driving the rapid spread of more toxigenic *Fusarium gramienarum* in

North America. Fungal Genetics and Biology **45:**473–484 https://doi.org/10.1016/j.fgb.2007.10.003

Yli-Mattila T, GagkaevaT, Ward TJ, Aoki T, Kistler HC, O'Donnell K. 2009. A novel Asian clade within the *Fusarium graminearum* species complex includes a newly discovered cereal head blight pathogen from the Far East of Russia. Mycologia **101:**841–852 https://doi.org/10.3852/08-217

Yli-Mattila T, Paavanen-Huhtala S, Parikka P, Hietaniemi V, Jestoi M, Gagkaeva T, Sarlin T, Haikara A, Laaksonen S, Rizzo A., 2008. Real-time PCR detection and quantification of *Fusarium poae*, *F graminearum*, *F sporotrichioides* and *F langsethiae* as compared to mycotoxin production in grains in Finland and Russia. Archives of Phytopathology and Plant Protection **41:**243–260 https://doi.org/10.1080/03235400600680659

Figure 1

The fungal perithecia and spores, and the symptoms of *Fusarium* disease of cereals from the Far East of Russia presented in the book by N. A. Palchevsky (1891).

Figure 2

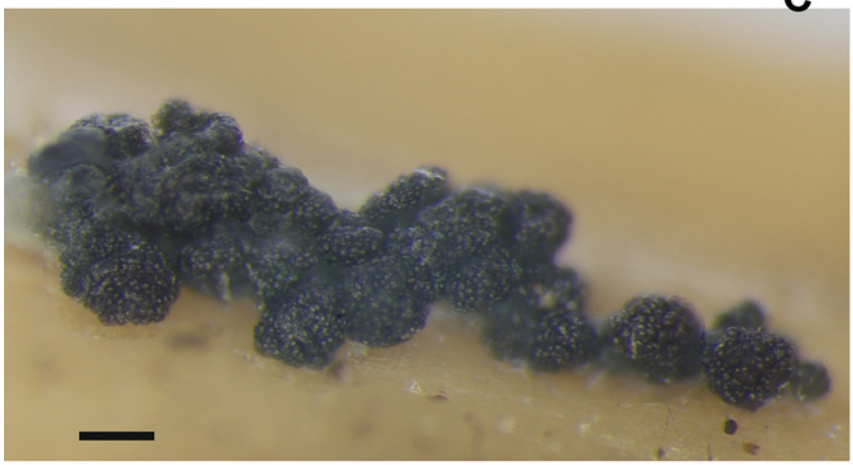
The diseased wheat (A) and barley grains (C); colony of *F. graminearum* on PSA, 10 days (B); macroconidia *F. graminearum* (D); the perithecia (E); expulsion of asci and ascospores from the perithecia (F). Scale bars: D = 20 µm; E = 200 µm; F = 5

A

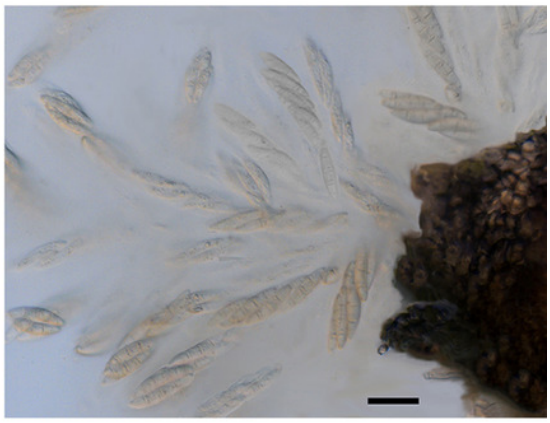
B

C

D



E



F

Figure 3

Maximum likelihood (ML) phylogenetic tree based on DNA sequence data from fragments of the translation elongation factor EF-1a (*TEF*), ammonium ligase gene (*URA*), reductase gene (*RED*), and 3-O-acetyltransferase gene (*Tri101*) of [i]F

Numbers on the nodes are ML and maximum parsimony bootstrap support values greater than 70%, followed by Bayesian posterior probability scores greater than 0.95. Reference *Fusarium* isolates with NRRL number are indicated in bold. *F. pseudograminearum* was used as an outgroup.

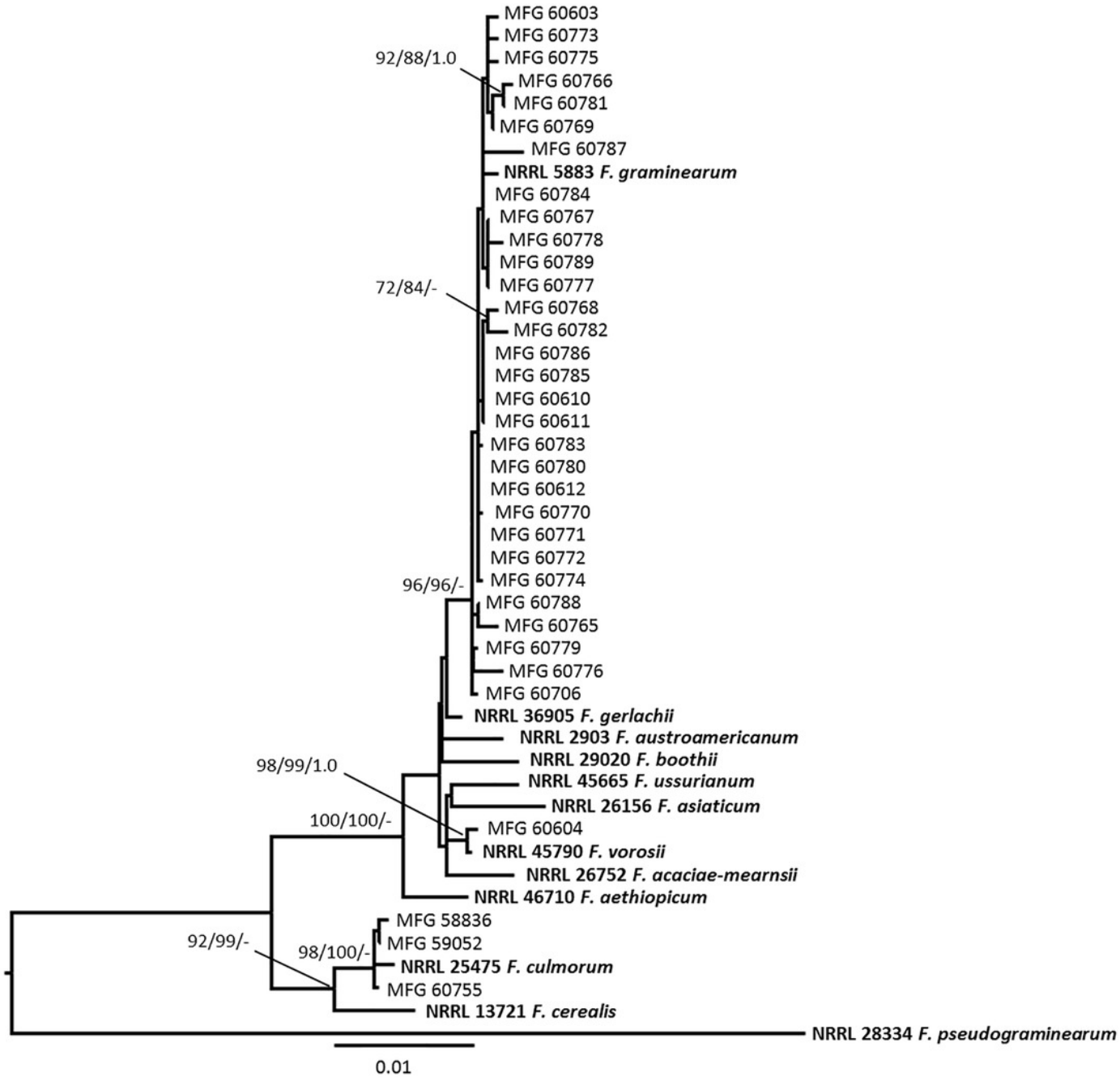

MFG 60603
MFG 60773
92/88/1.0
MFG 60775
MFG 60766
MFG 60781
MFG 60769
MFG 60787
NRRL 5883 F. graminearum
MFG 60784
MFG 60767
MFG 60778
MFG 60789
72/84/-
MFG 60777
MFG 60768
MFG 60782
MFG 60786
MFG 60785
MFG 60610
MFG 60611
MFG 60783
MFG 60780
MFG 60612
MFG 60770
MFG 60771
MFG 60772
96/96/-
MFG 60774
MFG 60788
MFG 60765
MFG 60779
MFG 60776
MFG 60706
NRRL 36905 F. gerlachii
NRRL 2903 F. austroamericanum
98/99/1.0
NRRL 29020 F. boothii
NRRL 45665 F. ussurianum
NRRL 26156 F. asiaticum
100/100/-
MFG 60604
NRRL 45790 F. vorosii
NRRL 26752 F. acaciae-mearnsii
NRRL 46710 F. aethiopicum
MFG 58836
92/99/-
MFG 59052
98/100/-
NRRL 25475 F. culmorum
MFG 60755
NRRL 13721 F. cerealis
NRRL 28334 F. pseudograminearum
0.01

Table 1(on next page)

Climatic data during the growing season of 2019 in the Amur region (https://rp5.ru/)

Month	Average temperature, °C			Average humidity, %	Total rainfall, mm	Days with precipitation
	mean	min	max			
May	+12.0	−1.6	+27.5	56	17	19
June	+18.2	+7.2	+30.1	66	46	21
July	+21.1	+13.5	+30.2	83	300	27
August	+18.6	+10.6	+29.2	82	206	25

1

Table 2(on next page)

Fusarium strains included in the study

[a] NRRL– the ARS Culture Collection (USA); nucleotide sequences of these reference strains were used in phylogenetic analysis. MFG – the fungal collection of Laboratory of Mycology and Phytopathology, All-Russian Institute of Plant Protection (Russia); the studied strains. [b] The translation elongation factor EF-1a (*TEF*), ammonium ligase gene (*URA*), reductase gene (*RED*), and 3-O-acetyltransferase gene (*Tri101*). [c] Bold indicates the number of sequence obtained in this study.

Species	Strain[a]	Host	Geographic original	Year	GenBank accession number[b]				Chemo-type
					TEF	*URA*	*RED*	*Tri101*	
F. acaciae-mearnsii	NRRL 26752	acacia	South Africa		AF212447	AF212705	AF212558	AF212594	
F. aethiopicum	NRRL 46710	wheat	Ethiopia		FJ240296	FJ240274	FJ240252	FJ240339	
F. asiaticum	NRRL 26156	wheat	China		AF212452	AF212710	AF212563	AF212599	
F. austroamericanum	NRRL 2903		Brazil		AF212438	AF212696	AF212549	AF212585	
F. boothii	NRRL 29020	corn	USA		AF212443	AF212701	AF212554	AF212590	
F. cerealis	NRRL 13721	potato	Poland		AF212464	AF212722	AF212575	AF212611	
F. culmorum	NRRL 25475	barley	Denmark		AF212463	AF212721	AF212574	AF212610	

F. culmorum	MFG 58836	wheat, grain	Russia, Omsk region	2015	**MW273182**[c]	**MW273250**	**MW273216**	**MW892041**	3-AcDON
F. culmorum	MFG 59052	wheat, grain	Russia, Krasnodar region	2017	**MW273183**	**MW273251**	**MW273217**	**MW892042**	3-AcDON
F. culmorum	MFG 60755	barley, grain	Russia, Tyumen region	2015	**MW273187**	**MW273255**	**MW273221**	**MW892043**	3-AcDON
F. gerlachii	NRRL 36905	wheat	USA		DQ459742	DQ459776	DQ459793	DQ452409	
F. graminearum	NRRL 5883	corn	USA		AF212455	AF212713	AF212566	AF212602	
F. graminearum	MFG 60765	wheat, grain	Russia, Amur region	2019	**MW273157**	**MW273225**	**MW273191**	**MW273259**	15-AcDON

F. graminearum	MFG 60766	wheat, grain	Russia, Amur region	2019	**MW273168**	**MW273236**	**MW273202**	**MW273270**	15-AcDON
F. graminearum	MFG 60767	wheat, grain	Russia, Amur region	2019	**MW273176**	**MW273244**	**MW273210**	**MW273278**	15-AcDON
F. graminearum	MFG 60768	wheat, grain	Russia, Amur region	2019	**MW273177**	**MW273245**	**MW273211**	**MW273279**	15-AcDON
F. graminearum	MFG 60769	wheat, grain	Russia, Amur region	2019	**MW273181**	**MW273249**	**MW273215**	**MW273283**	15-AcDON
F. graminearum	MFG 60770	wheat, grain	Russia, Amur region	2019	**MW273186**	**MW273254**	**MW273220**	**MW273286**	3-AcDON
F. graminearum	MFG	wheat,	Russia,	2019	**MW273188**	**MW273256**	**MW273222**	**MW273287**	15-

	60771	grain	Amur region						AcDON
F. graminearum	MFG 60772	wheat, grain	Russia, Amur region	2019	**MW273189**	**MW273257**	**MW273223**	**MW273288**	3-AcDON
F. graminearum	MFG 60773	wheat, grain	Russia, Amur region	2019	**MW273190**	**MW273258**	**MW273224**	**MW273289**	15-AcDON
F. graminearum	MFG 60774	wheat, grain	Russia, Amur region	2019	**MW273158**	**MW273226**	**MW273192**	**MW273260**	15-AcDON
F. graminearum	MFG 60775	wheat, grain	Russia, Amur region	2019	**MW273159**	**MW273227**	**MW273193**	**MW273261**	15-AcDON
F. graminearum	MFG 60776	wheat, grain	Russia, Amur	2019	**MW273160**	**MW273228**	**MW273194**	**MW273262**	15-AcDON

			region						
F. graminearum	MFG 60777	wheat, grain	Russia, Amur region	2019	**MW273161**	**MW273229**	**MW273195**	**MW273263**	15-AcDON
F. graminearum	MFG 60778	wheat, grain	Russia, Amur region	2019	**MW273162**	**MW273230**	**MW273196**	**MW273264**	3-AcDON
F. graminearum	MFG 60779	wheat, grain	Russia, Amur region	2019	**MW273163**	**MW273231**	**MW273197**	**MW273265**	15-AcDON
F. graminearum	MFG 60780	wheat, grain	Russia, Amur region	2019	**MW273164**	**MW273232**	**MW273198**	**MW273266**	3-AcDON
F. graminearum	MFG 60781	barley, grain	Russia, Amur region	2019	**MW273165**	**MW273233**	**MW273199**	**MW273267**	15-AcDON

F. graminearum	MFG 60782	barley, grain	Russia, Amur region	2019	**MW273166**	**MW273234**	**MW273200**	**MW273268**	3-AcDON
F. graminearum	MFG 60783	barley, grain	Russia, Amur region	2019	**MW273167**	**MW273235**	**MW273201**	**MW273269**	3-AcDON
F. graminearum	MFG 60784	barley, grain	Russia, Amur region	2019	**MW273169**	**MW273237**	**MW273203**	**MW273271**	3-AcDON
F. graminearum	MFG 60785	barley, grain	Russia, Amur region	2019	**MW273170**	**MW273238**	**MW273204**	**MW273272**	3-AcDON
F. graminearum	MFG 60786	barley, grain	Russia, Amur region	2019	**MW273171**	**MW273239**	**MW273205**	**MW273273**	15-AcDON
F. graminearum	MFG	barley,	Russia,	2019	**MW273172**	**MW273240**	**MW273206**	**MW273274**	15-

	60787	grain	Amur region						AcDON
F. graminearum	MFG 60788	barley, grain	Russia, Amur region	2019	**MW273173**	**MW273241**	**MW273207**	**MW273275**	15-AcDON
F. graminearum	MFG 60789	wheat, grain	Russia, Amur region	2019	**MW273174**	**MW273242**	**MW273208**	**MW273276**	15-AcDON
F. graminearum	MFG 60603	barley, grain	Russia, Amur region	2019	**MW273175**	**MW273243**	**MW273209**	**MW273277**	15-AcDON
F. graminearum	MFG 60612	wheat, grain	Russia, Kemerovo region	2019	**MW273178**	**MW273246**	**MW273212**	**MW273280**	3-AcDON
F. graminearum	MFG 60610	wheat, grain	Russia, Amur	2019	**MW273179**	**MW273247**	**MW273213**	**MW273281**	15-AcDON

			region						
F. graminearum	MFG 60611	wheat, grain	Russia, Amur region	2019	**MW273180**	**MW273248**	**MW273214**	**MW273282**	15-AcDON
F. graminearum	MFG 60706	soybean, leaves	Russia, Amur region	2019	**MW273185**	**MW273253**	**MW273219**	**MW273285**	15-AcDON
F. pseudograminearum	NRRL 28334	*Medicago* sp.	South Africa		AF212470	AF212729	AF212580	AF212617	
F. ussurianum	NRRL 45665	wheat, grain	Russia, Jewish autonomous region	2002	FJ240300	FJ240279	FJ240257	FJ240344	
F. vorosii	NRRL 45790	wheat, grain	Russia, Primorsky Krai	2006	FJ240302	FJ240281	FJ240259	FJ240346	

F. vorosii	MFG 60604	wheat, grain	Russia, Altay Krai	2018	**MW273184**	**MW273252**	**MW273218**	**MW273284**	15-AcDON

1

Table 3(on next page)

Infection of wheat and barley grain grown in the Amur region in the Russian Far East, 2019

Parameters		Samples of grain	
		Wheat	Barley
Grain infected with fungi	*Fusarium* spp.	**81.1** (58–98)	**80.5** (64–94)
on average (range), %	incl. *F. graminearum*	**68.0** (47–88)	**72.0** (61–92)
Content of mycotoxins	DON	**7,498** (3,207–13,343)	**5,390** (912–7,756)
on average (range), ppb	3-AcDON	**122** (27–293)	**131** (0–192)
	15-AcDON	**85.5** (23–179)	**93.5** (19–154)
	3-DON-glucoside	**1,011** (299–2,001)	**2,128** (98–3,803)
	ZEN	**1,153** (92–3,670)	**537** (111–928)
	MON	**70.2** (10–218)	**72.7** (5–207)
Amount of *Fusarium*	*F. graminearum*	**6,089** (2,658–11,342)	**2,102** (163–3,557)
DNA × 10^{-3} on average	3-AcDON genotype	**1,084** (395–2,007)	**508** (107–783)
(range), pg/ng	15-AcDON genotype	**1,708** (755–2,776)	**371** (101–713)
	F. avenaceum	**40** (6–97)	**13** (3–38)

1