Variation of soil microbial carbon use efficiency (CUE) and its Influence mechanism in the context of global environmental change: a review

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Soil microbial carbon utilization efficiency (CUE) is the efficiency with which microorganisms convert absorbed carbon (C) into their own biomass C, also referred to as microorganism growth efficiency. Soil microbial CUE is a critical physiological and ecological parameter in the ecosystem's C cycle, influencing the processes of C retention, turnover, soil mineralization, and greenhouse gas emission. Understanding the variation of soil microbial CUE and its influence mechanism in the context of global environmental change is critical for a better understanding of the ecosystem's C cycle process and its response to global changes. In this review, the definition of CUE and its measurement method are reviewed, and the research progress of soil microbial CUE variation and influencing factors is primarily reviewed and analyzed. Soil microbial CUE is usually expressed as the ratio of microbial growth and absorption, which is divided into methods based on the microbial growth rate, microbial biomass, substrate absorption rate, and substrate concentration change, and varies from 0.2 to 0.8. Thermodynamics, ecological environmental factors, substrate nutrient quality and availability, stoichiometric balance, and microbial community composition all influence this variation. In the future, soil microbial CUE research should focus on quantitative analysis of trace metabolic components, analysis of the regulation mechanism of biological-environmental interactions, and optimization of the carbon cycle model of microorganisms' dynamic physiological response process.

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Abstract

Soil microbial carbon utilization efficiency (CUE) is the efficiency with which microorganisms convert absorbed carbon (C) into their own biomass C, also referred to as microorganism growth efficiency. Soil microbial CUE is a critical physiological and ecological parameter in the ecosystem's C cycle, influencing the processes of C retention, turnover, soil mineralization, and greenhouse gas emission. Understanding the variation of soil microbial CUE and its influence mechanism in the context of global environmental change is critical for a better understanding of the ecosystem's C cycle process and its response to global changes. In this review, the definition of CUE and its measurement methods are reviewed, and the research progress of soil microbial CUE variation and influencing factors is primarily reviewed and analyzed. Soil microbial CUE is usually expressed as the ratio of microbial growth and absorption, which is divided into methods based on the microbial growth rate, microbial biomass, substrate absorption rate, and substrate concentration change, and varies from 0.2 to 0.8. Thermodynamics, ecological environmental factors, substrate nutrient quality and availability, stoichiometric balance, and microbial community composition all influence this variation. In the future, soil microbial CUE research should focus on quantitative analysis of trace metabolic components, analysis of the regulation mechanism of biological-environmental interactions, and optimization of the carbon cycle model of microorganisms' dynamic physiological response process.

Introduction

It is the worldwide agreement to deal with climate change by jointly controlling and slowing global warming by effectively increasing carbon (C) retention and reducing C emissions in a reasonable manner [1]. It is critical to accurately simulate and predict the interaction between global warming and the earth's ecosystems, particularly the feedback effects and mechanisms of

40   terrestrial ecosystems on global warming, to formulate effective measures to increase sinks [2],
41   [3]. A large number of studies have found that global warming encourages the release of soil
42   carbon, resulting in positive feedback on global warming [2], [4]. Microorganisms, on the other
43   hand, are increasingly being discovered to play a key role in regulating the feedback of terrestrial
44   ecosystems to global changes, and may even alter the expected feedback effects [5]–[7]. Long-
45   term warming, for example, reduces the decomposition of soil organic carbon by inhibiting
46   microbial biomass and enzyme activity. Microorganisms' physiological metabolic processes, as
47   well as their responses and adaptations to changes in the external environment, have become
48   crucial to the terrestrial ecosystem's feedback effect [5].
49   Soil microbes are involved in almost all material transformation processes in the soil and connect
50   the material circulation of the soil, biosphere, atmosphere, hydrosphere, and lithosphere. The
51   carbon utilization efficiency of soil microorganisms (Microbial carbon use efficiency, CUE), that
52   is, the microorganisms' ability to convert absorbed carbon into biomass carbon, is directly related
53   to their growth. The microbial CUE is set as a constant in many soil C cycle models [8]–[10].
54   Field observations and indoor cultivation experiments, on the other hand, contradict this
55   hypothesis. Changes in the external environment and nutrient conditions can have a significant
56   impact on soil microbial CUE. According to studies, soil microbial CUE increases as soil
57   nutrient availability increases [11], [12] and decreases as temperature rises [5], [13]. However,
58   there is a lack of consensus on the impact of these potential factors. Water stress, for example,
59   inhibits the growth of microorganisms and CUE, according to studies conducted on the prairies
60   of North America [14]. [15] discovered, however, that lowering soil water content did not affect
61   soil microbial CUE. These disparate findings reflect a lack of understanding of soil microbial
62   CUE variation and the mechanisms that influence it, limiting accurate simulation and prediction
63   of terrestrial ecosystem feedback [16].
64   Laboratory data, conceptual and quantitative models, and, to some extent, field-based
65   experiments are all contributing to the development of microbial CUE concepts [1]–[3].
66   However, in order to effectively incorporate agricultural C sequestration, this information must
67   be applied to the complexities and variation of soil microbial carbon use efficiency (CUE) and its
68   impact mechanism in the light of global environmental change. In this light, we illustrate areas
69   where information is missing, such as the complexities of microbial population abiotic, biotic,
70   and interaction interactions, which may be crucial in accurately predicting management
71   outcomes of soil microbial CUE. We think about these uncertainties in terms of influence
72   mechanisms that could improve CUE in soil ecosystems. Many methodological problems have
73   recently been discussed [4] but here we concentrate on the wider influences of other factors on
74   CUE that continue to challenge C sequestration in soil ecosystems
75   The purpose of this current review is to provide a comprehensive understanding of the variation
76   characteristics of soil microbial CUE and its influencing factors, and highlights the focus of
77   future research, by combing and analyzing the existing literature, all to improve the current earth
78   system model and provide a theoretical foundation for scientist, researchers and relevant stake-
79   holders to predict future climate change.

Survey methodology

To ensure an inclusive and unbiased analysis of literature and to accomplish the review's objectives, a comprehensive analysis of published articles on soil microbial carbon use efficiency was conducted using the Science Direct (http://sciencedirect.com) database, Web of Science, and Google Scholar. The following keywords were used to retrieve relevant literature: "soil microbial carbon use efficiency", "soil microorganisms", "ecological stoichiometry", "microbial community", and "nutrient limitation in soil ecosystem". While current publications between 2014 and 2019 were considered, publications that did not fall within this time period but contained critical information and were relevant to the review's objectives were also considered. Additionally, the reference lists of the retrieved literature were combed for additional pertinent publications. It is worth noting that this review presents a cross-section of studies on soil microbial carbon use efficiency and does not include all studies on the subject.

Definition of soil microbial carbon utilization rate

Through photosynthesis, vegetation converts CO2 in the atmosphere into organic matter, forming the ecosystem's net primary productivity. To realize the biogeochemical cycle of materials and energy in the ecosystem, the majority of vegetation productivity must be reduced to inorganic nutrients by decomposer-soil microbial decomposition and mineralization, and then absorbed and used by vegetation. Microorganisms' physiological metabolic process is a combination of assimilation and alienation metabolism. Microorganisms convert part of the photosynthesis of plants into microbial biomass, while the rest is released into the atmosphere via respiratory metabolism. Microbial carbon utilization efficiency [12], [17], also known as microbial growth efficiency or substrate utilization efficiency, is the efficiency with which microorganisms convert vegetation productivity into microbial biomass in this process [18]–[20]. Soil microbial CUE is an important ecological parameter in the soil C cycling process. It has a direct impact on the ecosystem's C retention time and turnover rate, as well as the soil's C storage capacity [7], [21], [22].

In ecological research, microbial CUE is usually expressed as the ratio of microbial growth (μ) to absorption (U) [12], [17], that is, CUE=μ/U. Microorganisms absorb C from the outside world mainly for microbial growth (μ), respiratory metabolism (R), secretion of extracellular enzymes and metabolites (EX), and microbial death (BD) (Figure 1). According to the principle of conservation of mass, microorganism U is expressed as;

$$U = \mu + R + EX + BD$$

Among them, soil microbial respiration (R) includes the respiration produced by microorganisms for growth (R_G), maintenance (R_M), extracellular enzyme production (R_E), and overflow process (R_O) [12], and is expressed as:

$$R = R_G + R_M + R_E + R_O$$

120 According to the definition of CUE and the mass conservation equation, the microbial CUE is
121 expressed as:

$$CUE = \frac{\mu}{U} = \frac{\mu}{\mu + R_G + R_M + R_E + R_O + EX + BD}$$

123

124 In natural ecosystems, EX and BD are usually difficult to determine and relative to growth and
125 respiration. The amount of EX and BD is very small and often considered negligible [12].
126 Therefore, CUE is generally considered to be a balanced relationship between the two processes
127 of μ and R, that is,

$$CUE = \frac{\mu}{U} = \frac{\mu}{\mu + R}$$

129 This definition is widely used in current microbial metabolism and soil carbon cycle models
130 [12], [17].

131

132 ## Determination method of soil microbial carbon utilization

133 Indoor culture, in combination with a mass conservation method and a marker tracing method, is
134 the most common method for determining soil microbial CUE. The method of mass conservation
135 entails directly measuring the change in mass or concentration of a substance and calculating the
136 CUE using the principle of substance conservation. The purpose of the marker tracking method
137 is to effectively track the substrate's utilization path by labeling it and calculating the ratio of
138 substrate used for growth and respiration for determination. It is currently a widely used method.
139 Existing analysis methods can be roughly divided into methods based on microbial growth rate
140 measurement, methods based on microbial biomass measurement, methods based on substrate
141 absorption rate measurement, and methods based on substrate concentration change
142 determination based on different research methods and research objects (microorganisms or
143 substrates). Each method has its own set of benefits, drawbacks, and application range (Table 1).

144

145 **The growth rate method**
146 One of the earliest measurement methods is the growth rate method. It determines the microbial
147 CUE by measuring the rate of microbial biosynthesis and respiration directly. Using 3H-
148 thymidine, ^{3}H-leucine, and other labeling substrates, the synthesized nucleic acid or protein is
149 separated using trichloro-acetic acid digestion, combined with radioisotope analysis, and it is
150 determined that microorganisms synthesize ^{3}H-thymidine into DNA. Alternatively, the rate at
151 which 3H-leucine is converted to protein [23]–[25]. This method is simple and precise, and it is
152 primarily used in aquatic ecosystems. Use heavy water $H_2^{18}O$ labeled with [18]O for cultivation in
153 terrestrial soils, measure the amount of DNA synthesized by microorganisms from the water of
154 ^{18}O, and calculate the soil microbial CUE by combining the microbial DNA and biomass C
155 conversion coefficients. This method can directly measure a microorganism's biosynthesis rate,
156 but it's mostly used for short-term cultivation [26], [27].

157

158   **Biomass method**
159   CUE is typically calculated in terrestrial ecosystems, such as soil, based on changes in microbial
160   biomass. To determine the change in microbial biomass, a labeled activated organic carbon is
161   added as a substrate for indoor culture, along with a chloroform fumigation method. This method
162   is easy to use and maneuver, and it is a widely used standard method. However, because
163   capturing the growth of microbial communities and maintaining respiration in a short period is
164   difficult, the rate of conversion of active markers to biomass measured by this method in a short
165   period is not completely equivalent to the growth rate of microorganisms, but rather the
166   absorption rate of the community, resulting in the phenomenon of overestimation [28]–[30].
167
168   **Substrate absorption rate method**
169   The substrate absorption rate method is used to calculate the CUE by observing how the
170   substrate changes over time. ^{3}H-thymidine, ^{3}H-leucine, and other labeling substrates are used in
171   this method, and a short-term indoor culture is used to determine the rate of absorption and
172   utilization. This method, unlike the microbial biomass measurement method, accounts for the
173   loss of microbial products. However, determining the absorption rate of the substrate is usually
174   difficult and is only appropriate for short-term indoor culture analysis.
175
176   **Substrate concentration change method**
177   The substrate concentration change method, like the substrate absorption rate method, measures
178   CUE in terms of substrate change. After a period of indoor culture, this method adds high
179   concentrations of substrates (glucose, acetic acid, etc.), measures the changes in substrate
180   concentration, and calculates CUE based on the respiratory volume during the same period. This
181   method usually does not necessitate isotope labeling of the substrate, but it does necessitate a
182   substrate medium with a high concentration. The metabolism and secretion processes of
183   microbial products are taken into account because this method necessitates a certain amount of
184   culture time.
185
186   **Stoichiometric method**
187   The stoichiometric method calculates CUE based on the stoichiometric limit theory of elements
188   and does not require direct measurement of CUE. It's widely used in models of soil and litter
189   decomposition [17], [31]. According to the stoichiometric balance theory, when the ratio of
190   essential elements (such as N, P) and carbon in exogenous nutrients is lower than the N/P: C
191   element ratio threshold required for optimal microorganism growth, these Constraints on the
192   effectiveness of elements will affect microorganism growth. $CUE = A_E \times B_{C:E}/TER_{C:E}$ is a first-
193   order functional relationship between CUE and the nutrient element measurement ratio. A_E
194   stands for absorption efficiency of essential elements (E), $B_{C:E}$ stands for the C: E ratio of
195   microbial biomass, and $TER_{C:E}$ stands for the C: E ratio threshold required for optimal
196   microorganism growth [32]–[34]. Although this method has a theoretical foundation, does not

197 require experimental measurements, and can be parameterized and simulated, it is an empirical
198 relationship that is primarily used in the simulation of soil carbon cycle process models.
199

Variability of soil microbial carbon utilization rate

201 Soil microbial CUE is not constant in natural ecosystems. Microorganisms would only assimilate
202 organic matter and completely assimilate the substrate in an ideal state, so the carbon utilization
203 rate would be 1. Microorganisms' growth, on the other hand, is always accompanied by the
204 consumption of respiratory metabolic products. Microorganisms have a maximum CUE
205 (CUE_{max}) of less than 1 due to thermodynamic constraints [35], [36]. Microorganisms' actual
206 growth is governed by their stoichiometric balance, which shows that CUE varies with changes
207 in essential element absorption efficiency (E), the microorganisms' C: E ratio, and the optimal
208 growth C: E ratio threshold. When the nitrogen absorption efficiency A_N=1, the soil microbial
209 CUE can approach the maximum CUE when the C: N ratio threshold $TER_{C: N}$ is 15 (CUE_{max} =
210 0.6); when the $TER_{C: N}$ is 30, the soil microbial CUE drops to 1/2 of CUE_{max} (CUE_{max}/20.3).
211 Because A_N is usually less than one in the real world, it's difficult for microorganisms' actual
212 CUE to reach CUE_{max}. The results of three different methods of ATP generation, electron
213 transfer, and energy conversion show that the actual maximum CUE of microorganisms is
214 around 0.6 due to thermodynamic limitations [35]. After accounting for the maintenance of
215 metabolic consumption by microorganisms, [28] found that the maximum thermodynamic CUE
216 is around 0.55.
217 The soil microbial CUE can range from 0.2 to 0.8, according to the integration results based on
218 measured data. [28], [37] used a stoichiometric model of ecological enzyme activity, biomass
219 composition, and nutrient concentration to find that the average soil microbial CUE is around
220 0.29, which is close to CUE_{max}/2. The average soil microbial CUE is about 0.270.11, according
221 to the estimation result based on the C: N element stoichiometric equilibrium equation, which is
222 similar to the estimation result based on the C: P element stoichiometric equilibrium equation
223 (0.250.12) [17]. However, the results of the experiments show that the microbial CUE in
224 different soil layers varies. For example, the CUE of microorganisms in the mineral layer
225 (0.2840.005) is higher than the CUE of microorganisms in the organic layer (0.2050.008) [38].
226 The CUEs of different microbial populations are also different. Based on an integrated analysis
227 of experimental observation data, [39] discovered that the CUE of bacteria is around 0.3360.213,
228 which is higher than that of fungi (0.3260.196). The CUE of soil microbial varies depending on
229 the type of vegetation. The research and study of [40] on Siberian vegetation transects revealed
230 that the CUE of soil microorganisms decreased as they progressed from the meadow steppe to
231 the Taiga forest and tundra.
232 Soil microbial CUE is usually set as a parameter in the current large number of biogeochemical
233 cycle models, with values ranging from 0.25 to 0.6. (Table 2). Because different substrate
234 compositions and other influences are taken into account in different biogeochemical models, the
235 CUE parameter values vary. For example, the decomposition of underground and above-ground
236 organic matter in the CENTURY model uses different soil microbe CUs of 0.45 and 0.55,

237 respectively [8]. The Daisy, NCSOL, ICBM, and other models take carbon pool activity into
238 account. CUE is 0.6 for activated carbon pools and less than 0.6 for inert carbon pools [9], [10],
239 [41] (Table 2). However, [31] pointed out that measured soil microbial CUE results are
240 frequently lower than the model's preset value, implying that the current model understates the
241 true hetero-oxygen respiration flux to some extent.
242
Influencing factors of soil microbial carbon utilization.
244 Thermodynamics, physiology, and ecology, as well as environmental factors like temperature
245 and moisture, soil texture, substrate, and nutrient availability, and microbial community
246 composition, influence soil microbial CUE.
247
Thermodynamic limit
249 Microorganisms' carbon absorption process is a combination of assimilation and alienation.
250 Microorganisms convert photosynthesis from vegetation into their biomass, but this is always
251 accompanied by the metabolic consumption of their respiration. Microorganisms have a
252 theoretical maximum CUE of about 0.8 at the thermodynamic limit [36]. This indicates that
253 microorganisms can use no more than 80% of absorbed organic C for their growth and that at
254 least 20% of the C must be used to maintain the energy consumption of breathing. However, the
255 actual soil microbial CUE is difficult to reach the theoretical maximum because the microbes
256 still need to invest more energy in processes like respiration and nutrient absorption due to
257 physiological and ecological environmental constraints.
258
Limits of ecological environmental factors
260 Temperature, humidity, precipitation, and soil moisture all influence microbial metabolism,
261 altering the balance between and R and thus affecting CUE [5], [12]. [5], [12], [13], [42], [43]
262 have found that soil microbial CUE has a negative feedback on temperature increase, with CUE
263 decreasing as temperature rises. This is because, when the temperature is controlled,
264 microorganisms' growth and metabolic rate increase as the temperature rises [44]–[46].
265 However, the temperature sensitivity of microbial respiration metabolism is higher than that of
266 growth response [44], and microbial respiration increases faster than microbial growth, reducing
267 CUE [5], [43], [47]. According to [48], soil microbial CUE decreased by about 0.009 for every
268 1°C increase in temperature. Under high-temperature stress, the negative feedback effect of
269 microbial CUE is more pronounced [42]. According to the simulation results, 30 years of
270 continuous temperature rise has reduced the proportion of absorbed C used for microbial growth,
271 lowering the CUE from 0.31 to 0.23. [5]. However, some studies have found that the soil
272 microbial CUE does not change significantly as the temperature rises [49], [50]. The
273 composition of the substrate and the metabolic stage influence the response of soil microbial
274 CUE to temperature. The CUE of soil microorganisms under the supply of a single-molecule
275 structure substrate decreases with increasing temperature, while the CUE of soil microorganisms
276 under the supply of a polymer structure substrate does not change with increasing temperature,

277 according to [51]. Long-term warming, according to some studies, will make microorganisms
278 adaptable. Long-term warming will cause microorganisms to reduce their basal respiration rate
279 [52]. The continuous warming experiment in Harvard Forest revealed that a 5°C increase in
280 temperature over 18 years reduced the degree of soil microbial CUE, with an increasing
281 temperature lower than the warming effect of two consecutive years [6]. Because
282 microorganisms' thermal adaptability is linked to changes in microbial community composition,
283 reduced nutrient availability, and changes in microbial metabolic pathways, as well as substrates
284 and observation methods, there are still many unknowns about how microorganisms respond to
285 temperature and how they do so.
286 Another important environmental factor that influences microorganism growth and respiration,
287 and thus CUE, is soil moisture and water availability [14]. The effect of soil water availability on
288 CUE is complex and variable, similar to the effect of temperature, and is influenced by intensity,
289 duration, soil type, and soil water stress. Short-term water stress stimulates a microbial response
290 to water stress, promotes microbes to reduce the impact of drought by increasing osmotic
291 pressure regulation or short-term dormancy, and increases soil microbial CUE, according to
292 studies [15], [53]. Long-term water stress, on the other hand, reduces the solubility and
293 absorption of soil substrates, inhibiting microorganism growth [12], [54]. Long-term water
294 stress, on the other hand, will increase the metabolic consumption of microorganisms, lowering
295 CUE [14]. The CUE of soil microorganisms in an anaerobic environment is lower than in an
296 aerobic environment, according to studies [55]. The metabolites of soil microorganisms are
297 released from CO_2 to CH_4, which cannot be completely oxidized, and CUE decreases in an
298 anaerobic environment [56].
299
Substrate quality and effectiveness
301 Microorganisms use soil organic carbon and vegetation litter as a source of food. Soil microbial
302 CUE will be affected significantly by substrate quality [16]. The different material compositions
303 of the substrate, the decomposition process, the degree of reducibility, and the effectiveness all
304 contribute to this effect. Complex substrates (such as lignin and phenols, for example)
305 necessitate more enzymatic decomposition reactions, which increases microorganism respiratory
306 metabolism and lowers CUE [51], [57]. Degradation of refractory substrates increases
307 extracellular enzyme metabolism and lowers CUE [58]. The geographic distribution of soil
308 microorganism CUE is primarily influenced by differences in the quality of the soil substrate at
309 various points, according to [40].
310 The degree of C reduction of the substrate (γS) is another important factor that affects the CUE
311 of soil microorganisms. The degree of C reducibility refers to the chemical energy per mole of C
312 and is usually expressed by the electron equivalent per mole of C. The γS of the main substrate
313 used by microorganisms is usually in the range of 3-5 (such as organic acids, glucose,
314 carbohydrates, and lipids), which is equivalent to the degree of C reduction of soil
315 microorganisms ($\gamma B \approx 4.2$) [35]. When the γs of the substrate is lower than the microbial γB, the
316 CUE of soil microorganisms is lower because the energy per unit of the substrate cannot meet

317   the energy requirement of a unit of biomass production [35], [36]. The analysis results of 16
318   substrates CUE with different degrees of reduction showed that sugar CUE (0.667)> amino sugar
319   CUE (0.601)> amino acid CUE (0.551)> organic acid CUE (0.498).
320   The availability of substrate nutrients will also have an impact on soil microbial CUE.
321   Microorganisms will respond to nutrient changes by changing the carbon assimilation pathways
322   regulated by their chemical enzymes when the availability and composition of nutrients changes
323   [59], [60]. A large number of studies have found that as nutrient availability increases, soil
324   microbial CUE increases [11], [12]. The microbial carbon absorption rate and nutrient
325   concentration have a saturation function relationship. Microorganisms will maintain the optimal
326   carbon absorption rate to meet the absorption system's resource consumption when resources are
327   limited [37], [60]. When the availability of nutrients increases and the nutrient concentration
328   exceeds the microorganisms' equilibrium concentration, carbon absorption increases, increasing
329   CUE. Nutrient restriction, on the other hand, will lower CUE. The microorganism's
330   decomposition and synthesis, as well as the coupled metabolic process, will be altered by
331   nutrient restriction, which will increase metabolites such as extracellular enzymes and
332   polysaccharides, and a decrease in CUE [61], [62]. This has also been proven by a large number
333   of N addition experiments. The addition of nutrients such as nitrogen can either stimulate or
334   inhibit microorganism activity and respiration metabolism. Microbes have a high CUE,
335   especially in environments limited by N elements [19], [63]–[65]. The addition of nitrogen
336   boosts microorganisms' ability to use active carbon [65]. Studies have also shown that after long-
337   term N addition, the active C pool decomposes gradually, and the microbial community becomes
338   restricted by energy such as C, lowering the CUE of soil microorganisms. More in-depth long-
339   term observational studies are needed to reveal how microorganisms respond and adapt to
340   changes in nutrient availability, as most nutrient addition experiments are still limited in time.
341   Soil microorganisms will adjust and redistribute resources within cells to meet the demand for a
342   variety of nutrient elements, which will further affect microorganism growth and CUE [66]–[68].
343   When microorganisms are restricted by a nutrient, they will expend more energy to obtain the
344   missing nutrient elements, inhibiting microorganism growth and CUE [11], [12]. When
345   microorganisms are restricted by the P element, for example, they will increase the input for P
346   element resource acquisition, lowering CUE [28]. Experiments have shown that nutrient
347   deficiency inhibits microorganism growth and lowers CUE [51]. This nutrient scarcity is usually
348   the result of the availability of multiple nutrients being constrained at the same time [28].
349
350   **Limits of ecological stoichiometry**
351   Changes in the stoichiometric ratio of the substrate's elements have a significant impact on
352   microorganism growth and CUE [12], [69]. Decomposer microbes have a stable stoichiometric
353   ratio of elements, which differs from the external environment's and the decomposing substrate's
354   stoichiometric composition. According to studies, soil microorganisms have a C/N ratio of about
355   7-8.6 [69]–[71], which is higher than plankton (6.6) and aquatic organisms (8.3) but lower than
356   soil organic carbon (12.3). (18.2-75). Soil microorganisms have a C/P ratio of about 23-60 [69],

357   [70], which is much lower than plankton (106), aquatic ecosystems (166), soil organic carbon
358   (72), and vegetation litter (100-837) [69]–[73]. To help maintain their stoichiometric balance,
359   soil microorganisms will preferentially absorb and utilize the elements that are most suitable for
360   their growth, according to the stoichiometric restriction theory. It is conducive to the
361   mineralization of elements when the ratio of the C content of the substrate to the essential
362   element E content (C: E) is lower than the threshold value of the element ratio required for
363   optimal microorganism growth (TERC: E); in TERC: E, it is conducive to the retention of
364   elements. Soil microorganisms are C-limited when decomposing substrates with high N and P
365   element content, and CUE is higher. When the content of N and P elements in the decomposition
366   substrate is low, soil microorganisms may respond in one of two ways: they may inhibit substrate
367   C and nutrient absorption, or they may release excess C through overflow respiration or
368   metabolic secretions [34]. The first reaction does not affect microbial CUE, but the second
369   reaction reduces microbial CUE in the soil due to increased respiratory metabolism and
370   secretion.
371   There is a negative correlation between soil microbial CUE and soil organic matter, C: N, and
372   C:P ratios, according to numerous studies [32], [38], [39]. The soil microbial CUE in farmland
373   and deciduous forest decreased as soil soluble C: N increased, according to [64]. [40]
374   investigated the variation of soil microbial CUE at various altitudes and found that as the latitude
375   increased, the soil C: N increased, and the soil microbial CUE decreased.
376

377   **Composition of microbial community**
378   Because different microbial populations decompose and absorb the organic matter at different
379   rates, the structure and composition of the microbial community have an impact on soil
380   microbial CUE [74], [75]. Fast-growing microorganisms that use the 'opportunity' growth
381   strategy are more adapted to high-nutrient environments and have lower CUE than slow-growing
382   microorganisms [76], [77]. The CUE of a microbial community dominated by fungi is often
383   higher than that of a microbial community dominated by bacteria, according to [78]. Fungi have
384   a wider C: N:P variation range than bacteria, and their C: N ratio is higher than bacteria's, which
385   has a higher requirement for C and has a high CUE [76]. However, some studies have found that
386   the CUE of soil communities does not differ significantly [18], [19]. [19] found no significant
387   difference in CUE between communities with a high fungi/bacteria ratio and communities with a
388   low fungi/bacteria ratio, which was 0.59±0.02 and 0.61±0.01, respectively, in a study of
389   farmland ecosystems. Increased substrate C: E can also increase the CUE of the fungal
390   community while decreasing the CUE of the bacterial community, according to studies [76]. The
391   microbial community's interspecific competition will reduce the microorganisms' CUE [79].
392   Because the microbial community's composition is highly susceptible to changes in the external
393   environment and human activities, as well as changes in the substrate's quality and composition,
394   quantifying the differences in the composition of different communities remains a work in
395   progress [18], [76].
396

Conclusions
Problems and Prospects
Analysis of Research Scales

Microbial CUE generally refers to the efficiency of microorganisms in absorbing and using C and converting it into their products, but it has specific time and space properties [80]. Microbial CUE is divided into three levels by [81]: population CUE (CUE_P), community CUE (CUE_C), and ecosystem CUE (CUE_E). These three levels of CUE describe the biosynthesis efficiency of microorganisms at various scales, from population to ecosystem, and include a variety of physiological and chemical processes in microbes. [81]. A large number of early studies concentrated on microbial CUE at the population and community levels. More and more research has recently focused on changes in the ecosystem-scale CUE and how it responds to the environment [82], [83]. The comparison of CUE results obtained under various research scales, observation stages, and measurement conditions will make the analytical results more difficult and uncertain [80]. For accurate comparison and application in future CUE research, it is critical to more clearly identify the specific research scale and process of microbial CUE.

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Analysis of metabolic components

The current soil CUE calculation method is not consistent, and there is an overestimation problem. Microorganisms' respiration is made up of several components, including growth respiration for growth, maintenance respiration for organism operation, secreting extracellular enzymes respiration, and overflow respiration. Extracellular enzymes and overflow respiration components are rarely considered in most current microbial CUE studies, which mostly focus on growth or growth and maintenance respiration of microorganisms [12]. As a result, CUE is overestimated, and the CUE calculation results are extremely disparate. The loss of extracellular enzymes and overflow respiration must be fully considered in future research. Select an appropriate culture time to observe the entire process, including the metabolism and secretion of microbial products, in the measurement process of indoor culture, for example. Enzyme

436 production and conversion are also measured using a combination of isotope labeling and
437 metabolomics.
438

Research on multi-element interactions

440 The impact of environmental changes on soil microbial CUE, as well as the response and
441 feedback of soil microbial CUE to global changes, remain highly uncertain due to the complex
442 soil-plant-microbe interaction. Is the soil microbial CUE, for example, decreasing as the
443 temperature rises? Is it possible to adjust the temperature? How will long-term exogenous
444 nutrient input affect soil microbial CUE? Because the response and adaptation of soil microbial
445 CUE to environmental changes is complicated and variable due to differences in substrate
446 quality, effectiveness, element stoichiometric composition, microbial activity, and community
447 composition [47], it is necessary to integrate and discuss the physiological response process of
448 microbial CUE in future process mechanism research.
449

Optimization of soil carbon cycle model

451 In the soil C cycle model, microbial CUE is an important parameter. Many existing models
452 assume CUE is constant or only take it into account infrequently, making it impossible to
453 accurately simulate the dynamic response process of soil microbial CUE to changes in external
454 environmental conditions and its impact on the soil C cycle [7], [84]. Some models represent
455 microbial CUE as a first-order functional relationship between temperature, nutrient availability,
456 or nutrient element ratio, ignoring physiological processes such as changes in microbial
457 metabolic pathways, community composition, and inter-species relationships. Furthermore, the
458 microbial respiration process is generally considered to be mainly related to the microbial C
459 balance, but not to the N balance, in a large number of biogeochemical models, and it lacks
460 consideration of the dynamic changes of the P element [28], [34]. The balance of elements, the
461 dynamic response process of microorganisms to changes in external environmental conditions,
462 and changes in physiological processes such as microbial metabolic pathways, community
463 composition, inter-species relationships, and microbial turnover must all be considered in the
464 future optimization of the soil C turnover model.
465

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468

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tables

2   Table 1. Different microbial carbon use efficiency measurement methods

Measurement	Microbe		Substrate				Model
Measurement principle expression	Based on growth rate $$CUE = \frac{\mu}{\mu + R}$$		Based on changes in biomass $$CUE = \frac{\Delta C_B}{\Delta C_B + R_{CUM}}$$	Based on absorption rate $$CUE = \frac{U}{U - R}$$	Based on absorption rate $$CUE = \frac{\Delta C_S - R_{CUM}}{\Delta C_S}$$		Based on stoichiometric ratio $$CUE = \frac{\Delta A_E}{TER_{CE}}$$
Measurement parameters	Microbial growth rate and respiration		Changes and accumulation of microbial biomass respiration	Substrate absorption rate and respiration	Changes in substrate concentration and cumulative respiration		Element E absorption rate, microbial C:E and the threshold element for optimal growth of microbes
Substrate	³H-thymidine, ³H-leucine	¹⁸O-H₂O	¹⁸C-glucose, ¹⁴C-acetate	¹³C-glucose, ¹⁴C-acetate, ³H-thymidine	sugars, amino sugars, amino acids and organic acids		
Label needed time scale	needed short time	needed short time	needed short time	needed short time	not needed Long time		
Advantages	Direct measurement of microbial biosynthesis rate	Direct measurement of microbial biosynthesis rate	Simple and easy to operate	Consider microbial productivity flow	Consider the loss of microbial productivity		No measurement required
Disadvantages	Unsuitable for soil	Only suitable for short-term determination	Need to be converted into biomass, overestimating CUE	Only suitable for short-term determination	Need to measure the adsorption of the substrate and provide a high concentration of the substrate		There are model assumptions, empirical coefficients
Application field	Waters	land	land	land	land		
Reference	[23]–[25]	[26], [27]	[15], [28]	[29]	[13], [14]		[30]

3   CUE, carbon utilization; μ, microbial growth rate; R, total microbial respiration rate; U, substrate absorption rate; ΔC_B, change in microbial
4   biomass; ΔC_S, change in substrate concentration; R_{CUM}, cumulative respiration rate; A_E; The absorption efficiency of element (E); BC: E, the ratio of
5   C: E of microbial biomass to the C; $TER_{C:E}$, C: required for optimal growth of microorganisms

14   Table 2. Variation of microbial carbon use efficiency

Type of model	Carbon use efficiency	Attributes	References
Measured values	0.39		[44]
	0.58		[45]
	0.44-0.73		[46]
	0.58-0.70		[47]
	0.26-0.68		[48]
	0.45-0.75		[28]
	0.46-0.62		[15]
	0.35-0.83		[14]
	0.24±0.08		[49]
	0.49-0.79		[16]
	0.42-0.84		[42]
Stoichiometric model	0.29		[39]
Q-model	0.25		[50]
CENTURY-model	0.45	Decomposition of underground organic matter	[8]
	0.55	Decomposition of surface organic matter	[8]
Daisy, NCSOIL, ICBM model	0.6	Activated carbon pool	[9], [10], [43]
Daisy, NCSOIL, ICBM model	<0.6	Most inert carbon pools	[9], [10], [43]

Figure 1

Figure 1. microbial metabolic components and equilibrium equation

Sketched according to the definition of soil microorganism CUE and the mass balance equation of soil microorganism metabolism proposed by [12] ; U, microbial carbon absorption; μ, microbial growth; R_G, microbial growth respiration; R_m, microbial maintenance respiration; R_E, extracellular enzyme Respiration; R_O, overflow respiration; EX, secretion of extracellular enzymes and metabolites; BD, microbial death.

$$U = \mu + R_G + R_M + R_E + R_O + EX + BD$$

Carbon absorption (U)

Extracellular enzyme respiration (R_E)

Growth respiration (R_G)

Overflow breathing (R_O)

Microbial growth μ

Respiratory maintenance (R_m)

Microbial death (BD)

Extracellular enzyme and metabolite secretion (EX)