

Filamentous mats proliferating at mesophotic depths off Rapa Nui: a new threat to local marine biodiversity? (#59454)

1

First submission

Guidance from your Editor

Please submit by **26 Apr 2021** for the benefit of the authors (and your \$200 publishing discount) .



Structure and Criteria

Please read the 'Structure and Criteria' page for general guidance.



Custom checks

Make sure you include the custom checks shown below, in your review.



Raw data check

Review the raw data.



Image check

Check that figures and images have not been inappropriately manipulated.

Privacy reminder: If uploading an annotated PDF, remove identifiable information to remain anonymous.

Files

Download and review all files from the [materials page](#).

4 Figure file(s)

1 Table file(s)

1 Video file(s)

! Custom checks

Field study



Have you checked the authors [field study permits](#)?



Are the field study permits appropriate?



Structure and Criteria

Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

1. BASIC REPORTING
2. EXPERIMENTAL DESIGN
3. VALIDITY OF THE FINDINGS
4. General comments
5. Confidential notes to the editor

 You can also annotate this PDF and upload it as part of your review

When ready [submit online](#).

Editorial Criteria

Use these criteria points to structure your review. The full detailed editorial criteria is on your [guidance page](#).

BASIC REPORTING

-  Clear, unambiguous, professional English language used throughout.
-  Intro & background to show context. Literature well referenced & relevant.
-  Structure conforms to [PeerJ standards](#), discipline norm, or improved for clarity.
-  Figures are relevant, high quality, well labelled & described.
-  Raw data supplied (see [PeerJ policy](#)).

EXPERIMENTAL DESIGN

-  Original primary research within [Scope of the journal](#).
-  Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
-  Rigorous investigation performed to a high technical & ethical standard.
-  Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

-  Impact and novelty not assessed. Negative/inconclusive results accepted. *Meaningful* replication encouraged where rationale & benefit to literature is clearly stated.
-  All underlying data have been provided; they are robust, statistically sound, & controlled.
-  Speculation is welcome, but should be identified as such.
-  Conclusions are well stated, linked to original research question & limited to supporting results.



The best reviewers use these techniques

Tip

Example

Support criticisms with evidence from the text or from other sources

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Give specific suggestions on how to improve the manuscript

Your introduction needs more detail. I suggest that you improve the description at lines 57- 86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

Comment on language and grammar issues

The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult. I suggest you have a colleague who is proficient in English and familiar with the subject matter review your manuscript, or contact a professional editing service.

Organize by importance of the issues, and number your points

1. Your most important issue
2. The next most important item
3. ...
4. The least important points

Please provide constructive criticism, and avoid personal opinions

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

Comment on strengths (as well as weaknesses) of the manuscript

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

Filamentous mats proliferating at mesophotic depths off Rapa Nui: a new threat to local marine biodiversity?

Javier Sellanes^{Corresp., 1, 2}, Matthias Gorny³, Germán Zapata-Hernández^{1, 2, 4}, Gonzalo Alvarez^{5, 6}, Praxedes Muñoz¹, Fadia Tala^{1, 6, 7}

¹ Departamento de Biología Marina, Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Coquimbo, Chile

² Millennium Nucleus for Ecology and Sustainable Management of Oceanic Islands (ESMOI), Universidad Católica del Norte, Coquimbo, Coquimbo, Chile

³ Oceana, Santiago, Chile

⁴ Programa de Doctorado en Biología y Ecología Aplicada (BEA), Universidad Católica del Norte, Coquimbo, Coquimbo, Chile

⁵ Departamento de Acuicultura, Universidad Católica del Norte, Coquimbo, Coquimbo, Chile

⁶ Centro de Investigación y Desarrollo Tecnológico en Algas y otros Recursos Biológicos (CIDTA), Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Coquimbo, Chile

⁷ Coastal Socio-Ecological Millennium Institute (SECOS), Coquimbo, Coquimbo, Chile

Corresponding Author: Javier Sellanes
Email address: sellanes@ucn.cl

Through the use of a remotely operated vehicle (ROV) previously unexplored habitats (~40 to 350 m depth) around Rapa Nui (Easter Island) have been investigated. We have sighted vast fields of filamentous cyanobacteria-like mats, covering sandy bottoms, as well as over dead corals of mesophotic reefs (mainly composed by *Leptoseris* sp.). The highest coverage of these mats (up to ~100% of total seafloor surface), as well as major mortality of corals occur between 70 and 95 m depth off Hanga Roa, the main village of the island, located on its western side. Healthy *Leptoseris* sp. reefs were documented on the northern and southeastern sites of the island, which are also the least populated. A preliminary morphologic analysis of samples of the mats suggests that the assemblage is composed of at least four filamentous taxa, including two cyanobacteria (*Lyngbya* sp. and *Pseudoanabaena* sp.), a brown (*Ectocarpus* sp.), and a green algae (*Cladophora* sp.). An ongoing eutrophication process off the main town of the island, Hanga Roa village, is suggested as a potential driver of the proliferation of these filamentous mats.

Filamentous mats proliferating at mesophotic depths off Rapa Nui: a new threat to local marine biodiversity?

Javier Sellanes^{1,2*}, Matthias Gorny³, Germán Zapata-Hernández^{1,2,4}, Gonzalo Álvarez^{5,6}, Práxedes Muñoz¹ & Fadia Tala^{1,6,7}

¹Departamento de Biología Marina, Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Chile.

²Millennium Nucleus for Ecology and Sustainable Management of Oceanic Islands (ESMOI), Universidad Católica del Norte, Coquimbo, Chile.

³Oceana Chile.

⁴Programa de Doctorado en Biología y Ecología Aplicada (BEA), Universidad Católica del Norte, Coquimbo, Chile.

⁵Departamento de Acuicultura, Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Chile.

⁶Centro de Investigación y Desarrollo Tecnológico en Algas y otros Recursos Biológicos (CIDTA), Facultad de Ciencias del Mar, Universidad Católica del Norte, Coquimbo, Chile.

⁷Coastal Socio-Ecological Millennium Institute (SECOS), Chile

*Corresponding author:

Javier Sellanes

Larrondo 1281, Coquimbo, 178000 Chile

23 Email address: sellanes@ucn.cl

24 Keywords: Cyanophyceae, *Lepstoseris*, mesophotic reefs, marine conservation, Easter island,

25 Polynesia.

Abstract

Through the use of a remotely operated vehicle (ROV) previously unexplored habitats (~40 to 350 m depth) around Rapa Nui (Easter Island) have been investigated. We have sighted vast fields of filamentous cyanobacteria-like mats, covering sandy bottoms, as well as over dead corals of mesophotic reefs (mainly composed by *Leptoseris* sp.). The highest coverage of these mats (up to ~100% of total seafloor surface), as well as **major mortality of corals** occur between 70 and 95 m depth off Hanga Roa, the main village of the island, located on its western side. Healthy *Leptoseris* sp. reefs were documented on the northern and southeastern sites of the island, which are also the least populated. A preliminary morphologic analysis of samples of the mats suggests that the assemblage is composed of at least four filamentous taxa, including two cyanobacteria (*Lyngbya* sp. and *Pseudoanabaena* sp.), a brown (*Ectocarpus* sp.), and a green algae (*Cladophora* sp.). An ongoing eutrophication process off the main town of the island, Hanga Roa village, is suggested as a potential driver of the proliferation of these filamentous mats.

Introduction

Mesophotic coral reef ecosystems typically distribute from 30-40 m to over 150 m depth, they occur in tropical and subtropical waters and are formed mainly by light-dependent corals, but also including other typical reef organisms as sponge and macroalgae species (Baker et al., 2016). Those ecosystems are now recognized as ecologically distinct and independent from shallower counterparts, bearing a significant diversity of unique fauna, which is still unexplored in most parts of the world (Rocha et al., 2018). The insufficient knowledge is a product of the difficult access to those depths, where technical diving (e.g., rebreather system, trimix) or sophisticated submarine equipment (e.g., remotely operated vehicles, autonomous drop-cams and manned submersibles)

are necessary. These ecosystems are vulnerable to a series of anthropogenic stressors, such as fishing, thermal stress, diseases, pollution, invasive species, marine aquarium trade, oil and gas exploration, cables and pipelines (Andradi-Brown et al., 2016).

Rapa Nui (Easter Island; 27° 07' S, 109° 22' W) is a remote island, formed ~0.8 Mya, and located at the westernmost side of the large chain of seamounts comprising the Salas y Gómez ridge, relatively close to the East Pacific Rise (Rodrigo, Díaz & González-Fernández, 2014). Located in the easternmost apex of the Polynesian triangle, it is recognized for the high overall endemism levels of its coastal marine fishes (~22%; Randal & Cea, 2010) and invertebrate taxa (4 to 34%, see Fernández et al., 2014). However, this unique marine biodiversity is severely threatened by several anthropogenic issues, such as coastal erosion, terrestrial runoff (Mieth & Bork, 2005), plastic pollution (Hidalgo-Ruz et al., 2021), exacerbated tourism (Figueroa & Rotarou, 2016), over-fishing (Zylich et al., 2014), and potential pollution by the percolation of domestic sewage and landfill into aquifers (Rosa 2013), among others issues.

Recently (2015-2018), through the use of a remotely operated vehicle (ROV) we have been able to access unexplored habitats (~40 to 350 m depth) around the island, as well as at nearby seamounts, allowing for a first approach to the biodiversity of mesophotic coral ecosystems and deeper sites (Easton et al., 2019), generating new records of fauna including fishes (e.g., Easton et al., 2017), echinoderms (Mecho et al., 2019) as well as reports of vast fields of the solitary mesophotic mushroom coral *Cycloseris vaughani* (Hoeksema, Sellanes & Easton, 2019).

In those surveys, dense and extensive fields of filamentous mats, covering the seafloor and nearby reefs at mesophotic depths, have been also sighted around the island. It is known that cyanobacteria are a usual constituent of coral reef ecosystems (Stal, 2000; Bakker et al., 2017; Ford et al., 2017), having an important role in nitrogen fixation and primary production (Charpy et al., 2007).

However, under certain conditions, they can undergo massive proliferation, affecting the health of the ecosystem and these events have been associated with variations in irradiance, nutrient supply and other natural and anthropogenic disturbances (Ford et al., 2018). These proliferation events seem to be increasing at a global scale due to alterations in local biogeochemical cycles related to climate change (Paul, 2008, Paerl & Paul, 2012). Mats could develop into such dense blooms that could even wash ashore and accumulate in mass, as reported for Guam by Nagle and Paul (1999). At this place, benthic marine cyanobacterial blooms often occur in the presence of diverse assemblages of herbivorous fishes and urchins, but the underlying factors causing these proliferations, as well as the interaction mechanisms between grazers and these mats (since cyanobacteria are known to produce feed-deterrent compounds), are still poorly understood (Nagle & Paul, 1999, Cruz-Rivera & Paul 2000). Cyanobacteria have been also directly linked with ciguatera fish poisoning (CFP) outbreaks (Laurent et al., 2008), and mats could even generate suitable habitats for other toxic microalgae, including toxin-producing dinoflagellates, generating thus co-occurring blooms (Paerl & Otten, 2013). Although several species are not toxic, their growth could produce low oxygen conditions through organic matter accumulation and subsequent degradation processes in the bottom water, affecting also the benthic communities. In some environments mats form associations with sulfate-reducing bacteria, producing sulfide which is toxic for corals and establishing the black band disease (Charpy et al., 2012; Myers & Richardson, 2009).

It has been also reported that in littoral reefs green algae (i.e., Chlorophytes) are one of the most common indicator species of eutrophication (Barile, 2004). Most of the species in this group proliferate due to increased nutrient inputs and tolerate a wide range of environmental conditions, constituting an aggressive competitive species against sensible corals, and having sub-lethal

effects over several biological functions of them (e.g., Koop et al., 2001; Fabricius 2005; Birrel et al., 2008).

In this context, the aims of the present study were: i) to assess the spatial coverage of filamentous mats in the benthic ecosystem around Rapa Nui, ii) to evaluate the extension of mesophotic coral reefs potentially impacted by these mats, and iii) to preliminary describe the taxonomic composition of these mats.

Materials and methods

A remotely operated vehicle (ROV), model Commander MKII (Mariscope Meerestechnik, Kiel, Germany), operated from local fishing boats, was deployed in 56 opportunities around the island (Fig. 1), 18 times each in January 2018 and 2019, and 20 during November/December 2019. The ROV was equipped with two laser pointers, separated by 10 cm, and a front-pointing at an angle of 45° HD video camera (Panasonic SD 909), recording with a resolution of 1920 × 1080 pixels at 30 fps. As mentioned above, some results of these ROV surveys have been presented elsewhere (e.g., Easton et al., 2017; Easton et al., 2019; Hoeksema, Sellanes & Easton, 2019; Mecho et al., 2019, for selected biotic components), but for the present study, the only focus was to analyze the presence of filamentous mats and overall health condition of mesophotic corals. The videos were analyzed at half their normal speed in GOM Player 2.3.19 (GOM & Company; <https://www.gomlab.com/>).

The presence and coverage of filamentous mats were assessed in a semi-quantitative way by observing the seafloor stepwise as the ROV advanced over the ground, in general, 10 to 20 min of video per site. For that, transects were cataloged in four groups: i) no presence of patches of filamentous mats, ii) low coverage, less than 50 % coverage in at least non-overlapping 5 frames

118 of the video of this transect, iii) high coverage, 50 % to 75 % of coverage in at least 5 frames of
119 the video of this transect, and iv) very high coverage, 100 % coverage in at least 5 frames.

120 Using the same methodological approximation, for assessing the health status of corals three
121 criteria were considered: i) healthy reef with > 75% of all the corals alive, ii) 25-75% of the corals
122 alive, and iii) **only dead corals or fragments**. Dead corals were easily identified by being in general
123 of greenish or darker colors, and sometimes also covered by filaments.

124 To characterize the taxonomic composition of the filamentous mat assemblage, in May 2019, a
125 small benthic trawl of 30 cm of horizontal aperture was deployed at a site off Hanga Roa, where
126 patches with 100 % coverage were frequent. Mat samples were fixed with a 4 % aqueous solution
127 of formaldehyde (ACS Reagent, Sigma-Aldrich, Merck Darmstadt, Germany). For morphological
128 characterization, filaments were observed using an Olympus IX71 inverted microscope equipped
129 with phase contrast and epifluorescence (Olympus Co, Tokyo, Japan). Micrographs were taken
130 using a camera ProgRes C3 (JENOPTIK Optical Systems GmbH, Germany) and measurements of
131 cells (length, width) were carried out using ProgRes® CapturePro (JENOPTIK Optical Systems
132 GmbH, Germany) analytical software. For taxonomic identification of the algae composing the
133 **mats at least to genus, monographic publications, floristic studies and systematics articles were**
134 **used (Santelices, 1989; Loiseaux-de Goër & Noailles, 2008; Cormaci, Furnari & Alongi, 2014;**
135 **Ramirez et al., 2018).** Likewise for cyanobacteria identification taxonomic guides and systematics
136 articles were used (Komarek & Anagnostidis, 2007; Yu et al., 2015; Brocke et al., 2018; Zubia et
137 al., 2019).

138 Sample collection was performed under permissions Res. Ext N°41/2016 and N°3314/2017 from
139 SUBPESCA (National Fishing Authority of Chile) to Universidad Católica del Norte. This project

was also presented to the local “Consejo del Mar de Rapa Nui” (Council of the Sea of Rapa Nui), which allowed underwater footage and sampling around the island.

Results

ROV transects around the island covered a range of 43 to 347 m depth. This allowed us to visualize the spatial and bathymetric distribution of filamentous mats (Fig. 1, 2) and mesophotic reefs around the island (Fig. 3). Mesophotic corals were represented by reef-forming *Porites lobata* and *Pocillopora* sp. at shallower depths (<60m), by *Leptoseris* sp. and *Cycloseris* sp. at depths between 70 and 117m depth, and by sea-whips (*Stichopathes* sp.) and other scleractinians between 127 and 327 m (Fig. 2).

Spatial distribution of the filamentous mats

In general, filamentous mats were absent or with a low presence in most of the studied sites around Rapa Nui (Fig. 1A), and medium to very high abundances were usually observed shallower than ~130 m and in the western side of the island (Fig. 1B). Indeed, high coverage was observed in the northwest corner (close to Hanga O’teo) at 123 m depth and high and very high coverages were observed mainly off Hanga Roa (Fig. 1B) from 70 to 95 m depth. In this area, mats were also observed covering sediments (Fig. 2A, B) and fringing recently discovered fields of the zooxanthellate mushroom coral *C. vaughani* (Fig. 2C, Fig. 3A; see Hoeksema, Sellanes & Easton, 2019), and nearby reefs of dead *Leptoseris* sp. (~80 m depth) overgrown, apparently by similar mats (Fig. 2D, Fig. 3B). However, healthy *Leptoseris* sp. reefs were documented in other sites around the island (e.g. Anakena) between 68 and 82 m depth (Figs. 2E, 3B).

Taxonomic characterization of the filamentous mat assemblage

Morphological analyses of samples of mats collected off Hanga Roa suggest that it is an assemblage composed of at least four taxa: one Chlorophyta (*Cladophora* sp.), one Ochrophyta (*Ectocarpus* sp.) and two Cyanobacteria (*Lyngbia* sp. and *Pseudoanabaena* sp.) (Fig. 4) as follows:

Cladophora sp. (Fig. 4A, B): Thallus of green to light green branched uniseriate filaments with 2-3 cm in total length. Basal part of the filaments fixed in the substrate by a primary rhizoid. Presence of unilateral branches inserted laterally or obliquely the filament. Principal axis constituted by cylindrical cells with measures of $998.9 \pm 69.2 \mu\text{m}$ in length and $223.3 \pm 9.5 \mu\text{m}$ in diameter. The apical cells were cylindrical, round ended with a diameter of $250.0 \pm 7.6 \mu\text{m}$ and length of $701.8 \pm 76.0 \mu\text{m}$. Zoosporangia were not observed.

Ectocarpus sp. (Fig. 4C, D): Thallus of light brown to olive sparingly branched filament with 0.1 – 0.5 cm in total length. The cells conform uniseriate filaments that ended in a rounded apical cell. Cells were barrel-shaped, $50.0 \pm 7.3 \mu\text{m}$ in length and $14.1 \pm 3.4 \mu\text{m}$ in diameter. Plurilocular sporangia were present, elongated with cylindroconical form with 80-130 μm in length and 20-30 in diameter.

Lyngbia sp. (Fig. 4E, F): Thallus caespitose, browning-red, filaments slightly curved, sheet colorless, lamellated with apices not attenuated at the end. Trichome not constricted at the cross-wall, cylindrical cells very short $3.5 \pm 0.3 \mu\text{m}$ in length and $7.1 \pm 0.1 \mu\text{m}$ in diameter, sheath $1.6 \pm 0.3 \mu\text{m}$, end cells rotund, calyptra absent.

Pseudoanabaena sp. (Fig. 4G, H): Trichomes solitary or crowded in clusters, straight or almost straight, pale blue-green, cells. Barrel-shaped, $2.8 \pm 0.8 \mu\text{m}$ in length and $1.2 \pm 0.1 \mu\text{m}$ in diameter, intensely constricted at cross walls, not heterocysts or sheath, end cells round.

186

187 Discussion

188 Although we only have a very preliminary taxonomic characterization of the filamentous mats
 189 covering sandy areas and dead-mesophotic reefs off Rapa Nui, based on morphology, it is
 190 suggested that they are composed of at least **to** cyanobacteria. It is known that cyanobacteria-
 191 dominated microbial mats are typical inhabitants of coral reef systems and often undergo massive
 192 proliferation (Stal, 2000). These events have been associated with natural processes (e.g.,
 193 variations in irradiance), but mostly with anthropogenic disturbances that generate an increase in
 194 the nutrient supply to the marine environment (Ford et al., 2017). Highest coverage of mats was
 195 observed mainly off Hanga Roa village, which concentrates the largest part of the islands'
 196 population (7,750 habitants; www.ine.cl) and where most tourists develop their recreational
 197 activities (~150,000 **visitors** during 2019; www.sernatur.cl). In this sense, the over-tourism, the
 198 absence of wastewater treatment (predominant use of cesspools) and the unlined landfill (Rosa,
 199 2013) could be a great threat, due to the potential input of organic matter, nutrients and
 200 contaminants to the adjacent marine environment. Pollutants could reach the sea by runoff or
 201 through percolation to aquifers that eventually discharge into the sea. At Rapa Nui, submarine
 202 groundwater discharges are ubiquitous in intertidal environments around the island (Brosnan,
 203 Becker & Lipo, 2018), and hypothetically could also seep through deeper sediments (Montgomery
 204 & Associates, 2011), potentially conducting nutrients of anthropogenic origin directly to
 205 mesophotic habitats. Additionally, the permanent coastal erosion around the island and terrestrial
 206 runoff during **rainy seasons** could also increase the nutrient concentrations in the coastal
 207 environments (e.g., ammonia, nitrate and silicate) and potentially reaching mesophotic depths (P.
 208 Muñoz, 2019, unpublished data). However, the volcanic origin of Rapa Nui and their elevated

erosion could also increase the iron concentration in the marine ecosystem, which added to a decrease in the N:P ratio (Ford et al., 2018), could stimulate the proliferation of benthic cyanobacteria in other places around the island.

The damage of mesophotic corals as *Leptoseris* sp. is extensive and evident off Hanga Roa village (also covering the edges of a *C. vaughani* field), in contrast to other sites where healthy corals were documented (e.g. Anakena, La Perouse and Vinapú). Unfortunately, although no previous detailed information about the health condition and community structure of these degraded mesophotic reefs exists, a previous record, obtained in November 1999 during the first ROV survey at the island (Gorny & Retamal, 2000), suggests a healthy status of the *Leptoseris* reef off Hanga Roa (Fig. 2D) ~20 years ago. At that opportunity, the tourist number reached ~20.000 visitants per year (Figueroa & Rotarou, 2016). Despite this circumstantial indication of the status of the mesophotic reefs at Rapa Nui a few decades ago, the ecological impacts on the biodiversity and ecosystem functions associated with anthropogenic causes are still unknown and need to be further addressed in the short-term.

Conclusions

We report for the first time filamentous mats covering sandy areas and dead-mesophotic reefs (*Leptoseris* sp.) off Rapa Nui. A preliminary morphological analysis suggests the presence of at least four filamentous taxa, including two cyanobacteria (*Lyngbya* sp. and *Pseudoanabaena* sp.), a brown (*Ectocarpus* sp.), and a green algae (*Cladophora* sp.). Catastrophic damage was observed at mesophotic *Leptoseris* sp. reefs off the main village of the island. Further research should address a more detailed taxonomic characterization of these mats, e.g., through molecular techniques, the assessment of the seasonal, spatial and structural patterns of the assemblage, their

eventual role in reef deterioration, recognition of eutrophication mechanisms, and long-term monitoring of nutrient dynamics. These studies are encouraged to implement effective and integrated sea-land management actions, including the implementation of waste-water treatment. This information should be key to inform the implementation of management strategies (land-sea integrate) of the recently created Marine Protected Area of multiple uses (MPA-MU) of Rapa Nui, currently the largest in Latin America with ~579,000 km² (Paredes et al., 2019) and aiming to protect this unique world biodiversity heritage place. Finally, this study will serve also as a baseline for future studies of changes over the mesophotic ecosystem off Rapa Nui after the island has been closed for tourism, from March 2019 to date, due to the COVID 19 pandemic.

Acknowledgments

We thank to Poky Tane Haoa, Ricardo Hito and Enrique Hey from the Rapa Nui community. To Erin Easton and Ariadna Mecho for their collaboration during field work, and to Maria Valladares and Valentina Hevia for helping in the collection of the samples. To Bert Hoeksema for his valuable comments on an early version of this manuscript.

References

Andradi-Brown D, Laverick J, Bejarano I, Bridge T, Colin PL, Eyal G, Jones R, Kahng SE, Reed J, Smith TB, Spalding H, Well E, Wood E. 2016. Threats to mesophotic coral ecosystems and management options. In: Baker EK, Puglise, KA and Harris, PT, eds. *Mesophotic coral ecosystems—A lifeboat for coral reefs?* The United Nations Environment Programme and GRID-Arendal, Nairobi and Arendal, pp. 98

Baker E, Puglise KA, Colin PL, Harris PT, Kahng SE, Rooney JJ, Sherman C, Slattery M, Spalding HL. 2016. What are mesophotic coral ecosystems? In: Baker EK, Puglise KA and Harris PT, eds. *Mesophotic coral ecosystems—A lifeboat for coral reefs?* The United Nations Environment Programme and GRID-Arendal, Nairobi and Arendal, pp. 98

Bakker D, van Duyl F, Bak R, Nugues MM, Nieuwland G, Meesters E. 2017. 40 Years of benthic community change on the Caribbean reefs of Curacao and Bonaire: the rise of slimy cyanobacterial mats. *Coral Reefs* DOI: 10.1007/s00338-016-1534-9

Barile PJ. 2004. Evidence of anthropogenic nitrogen enrichment of the littoral waters of East Central Florida. *Journal of Coastal research* 20:137–1245

Birrel C, McCook L, Willis B, Diaz-Pulido G. 2008. Effects of benthic algae on the replenishment of corals and the implications for the resilience of coral reefs. *Oceanography and Marine Biology: An Annual Review* 46:25–63

Brocke HJ, Piltz B, Herz N, Abed RMM, Palinska KA, John U, Haan JD, de Beer D, Nugues MM. 2018. Nitrogen fixation and diversity of benthic cyanobacterial mats on coral reefs in Curaçao. *Coral Reefs* 37(3): 861–874

Brosnan T, Becker MW, Lipo CP. 2018. Coastal groundwater discharge and the ancient inhabitants of Rapa Nui (Easter Island), Chile. *Hydrogeology Journal* <https://doi.org/10.1007/s10040-018-1870-7>

278

279 Charpy L, Casareto BE, Langlade MJ, Suzuki Y.2012. Cyanobacteria in coral reef ecosystems: a
280 review. *Journal of Marine Biology* Doi: 10.1155/2012/259571

281

282 Cormaci M, Furnari G, Alongi G, 2014. Flora marina bentónica del Mediterráneo: Chlorophyta.
283 Boll. Accad. Gioenia Sc. Nat., *Catania* 47:11–436

284

285 Cruz-Rivera E, Paul VJ. 2000. Coral reef benthic cyanobacteria as food and refuge: Diversity,
286 chemistry and complex interactions. Proceedings 9th International Coral Reef Symposium, Bali,
287 Indonesia 23-27 October 2000, Vol. 1

288

289 Easton EE, Sellanes J, Gaymer CF, Morales N, Gorny M, Berkenpass E. 2017. Diversity of deep-
290 sea fishes of the Easter Island Ecoregion. *Deep Sea Res Part II* 137:78–88.
291 <https://doi.org/10.1016/j.dsr2.2016.12.006>

292

293 Easton EE, Gorny, M, Mecho A, Sellanes J, Gaymer CF, Slapding HL, Aburto J. 2019. Chile and
294 the Salas y Gómez Ridge. In: Loya Y, Puglise KA, Bridge TCL, eds. *Mesophotic Coral*
295 *Ecosystems*. Springer International Publishing, pp 477–490. [https://doi.org/10.1007/978-3-319-](https://doi.org/10.1007/978-3-319-92735-0)
296 [92735-0](https://doi.org/10.1007/978-3-319-92735-0)

297

298 Fabricius K.2005. Effects of terrestrial runoff on the ecology of corals and coral reefs: review and
299 synthesis. *Marine Pollution Bulletin* 50:125–146

300

301 Fernandez M, Pappalardo P, Rodriguez Ruiz MC, Castilla JC. 2014. Synthesis of the state of
302 knowledge about species richness of macroalgae, macroinvertebrates and fishes in coastal and
303 oceanic waters of Easter and Salas y Gomez islands. *Latin American Journal of Aquatic Research*
304 42(4):760–802

305

306 Figueroa E, Rotarou ES. 2016. Sustainable Development or Eco-Collapse: Lessons for Tourism
307 and Development from Easter Island. *Sustainability* 8:1093; doi:10.3390/su8111093

308

309 Ford A, Van Hoytema N, Moore B, Pandiha L, Wild C, Ferse S. 2017. High sedimentary oxygen
310 consumption indicates that sewage input from small islands drives benthic community shifts on
311 overfished reefs. *Environmental Conservation* 44(4):405–414. doi:10.1017/S0376892917000054

312

313 Ford AK, Bejarano S, Nugues MM, Visser PM, Albert S, Ferse SCA. 2018. Reefs under siege—
314 the rise, putative drivers, and consequences of benthic cyanobacterial mats. *Frontiers in Marine*
315 *Science* 5:18 Doi:10.3389/fmars.2018.00018

316

317 Gorny M, Retamal M. 2000. Estudio sobre la biodiversidad del megabentos de Isla de Pascua y
318 Salas y Gómez mediante video subacuático. Libro de Resúmenes. Taller sobre los resultados del
319 Crucero Cimar-Fiordo 5. Valparaíso: 119-121

320

321 Hidalgo-Ruz V, Luna-Jorquera G, Eriksen M, Frick H, Miranda-Urbina D, Porflitt-Toro M,
322 Rivadeneira M, Robertson CJR, Scofield RP, Serratos J, Suazo CG, Thiel M. 2021. Factors (type,
323 colour, density, and shape) determining the removal of marine plastic debris by seabirds from the

South Pacific Ocean: Is there a pattern? *Aquatic Conservation: Marine and Freshwater Ecosystem*
31:389–407

Hoeksema BW, Sellanes J, Easton EE. 2019. A high-latitude, mesophotic *Cycloseris* field at 85 m
depth off Rapa Nui (Easter Island). *Bulletin of Marine Science* 95:
<https://doi.org/10.5343/bms.2018.0053>

Komarek J, Anagnostidis K. 2007. Süßwasserflora von Mitteleuropa, Bd. 19/2: Cyanoprokaryota.
Bd. 2 / Part 2: Oscillatoriales. Springer Spektrum

Koop K, Booth D, Broadbent A, Drodie J, Bucher D, Capone D, Coll J, Dennison W, Erdmann
M, Harrison P, Hoegh-Guldberg O, Hutchings P, Jones GB, Larkum AWD, O’Neil J, Steven A,
Tentori E, Ward S, Williamson J, Yellowless D. 2001. ENCORE: The effect of nutrient enrichment
on coral reefs. Synthesis of results and conclusions. *Marine Pollution Bulletin* 42:91–120

Laurent D, Kerbrat AS, Darius HT, Girard E, Golubic S, Benoit E, Sauviat MP, Chinain M, Molgo
J, Pauillac S. 2008. Are cyanobacteria involved in Ciguatera Fish Poisoning-like outbreaks in New
Caledonia? *Harmful Algae* 7:827–838

Loiseaux-de Goër S, Noailles MC. 2008. Algues de Roscoff. Editions de la Station Biologique de
Roscoff

Mecho A, Easton EE, Sellanes J, Gorny M. Mah C. 2019. Unexplored diversity of the mesophotic echinoderm fauna of the Easter Island Ecoregion. *Marine Biology* Doi.org/10.1007/s00227-019-3537-x

Mieth A, Bork H-R. 2005. History, origin and extent of soil erosion on Easter Island (Rapa Nui). *Catena* 63:244–260

Montgomery EL & Associates INC. 2011. Condiciones hidrogeológicas Isla de Pascua, Chile. Dirección general de aguas. Pp. 48

Myers J, Richardson L. 2009. Adaptation of cyanobacteria to the sulfide-rich microenvironment of black band disease of coral. *FEMS Microbiology Ecology* 67:242–251

Nagle DG, Paul VJ. 1999. Production of secondary metabolites by filamentous tropical marine cyanobacteria: ecological functions of the compounds. *Journal of Phycology* 35:1412–1421

Paerl HW, Otten TG. 2013. Harmful Cyanobacterial Blooms: Causes, Consequences and Controls. *Microbial Ecology* 65:995–1010

Paerl H, Paul V. 2012. Climate change: Links to global expansion of harmful cyanobacteria. *Water Research* 46(5):1349–1363

Paredes F, Flores D, Figueroa A, Gaymer C, Aburto J. 2019. Science, capacity building and conservation knowledge: The empowerment of the local community for marine conservation in Rapa Nui. *Aquatic Conservation: Marine and Freshwater Ecosystem* 130–137
Doi.org/10.1002/aqc.3114

Paul VJ. 2008. Global warming and cyanobacterial harmful algal blooms. In: Hudnell HK, Ed. *Cyanobacterial Harmful Algal Blooms: State of the Science and Research Needs*. Advances in Experimental Medicine and Biology, Springer 619:239–257

Ramirez ME, Bulboa C, Contreras L, Mora AM. 2018. Algas marinas de Quintay. RiL Editores

Rocha LA, Pinheiro HT, Shepherd B, Papastamatiou YP, Luiz OJ, Pyle RL, Bongaerts P. 2018. Mesophotic coral ecosystems are threatened and ecologically distinct from shallow water reefs. *Science* 361:281–284. <https://doi.org/10.1126/science.aaq1614>

Rodrigo C, Díaz J, González-Fernández. 2014. Origin of the Easter Submarine Alignment: morphology and structural lineaments. *Latin American Journal of Aquatic Research* 42(4):857–870 <http://dx.doi.org/10.3856/vol42-issue4-fulltext-12>

Rosa K. 2013. A hydrologic overview and discussion of sources of groundwater pollution on Rapa Nui. *Rapa Nui Journal* 27(2): 51–59

Santelices B. 1989. Algas Marinas de Chile. Ediciones Universidad Católica de Chile

391

392 Stal LJ. 2000. Cyanobacterial mats and stromatolites. In: Whitton PA, Potts M, eds. *The ecology*
393 *of cyanobacteria: their diversity in time and space*. Kluwe Academic, Dordrecht, pp 61–120

394

395 Yu G, Zhu M, Chen Y, Pan Q, Chai W, Li R. 2015. Polyphasic characterization of four species of
396 *Pseudanabaena* (Oscillatoriales, Cyanobacteria) from China and insights into polyphyletic
397 divergence within the *Pseudanabaena* genus. *Phytotaxa* 192(1):1–12

398

399 Zubia M, Vieira C, Palinska KA, Roué M, Gaertner J-C, Zloch I, Grellier M, Golubic S. 2019.
400 Benthic cyanobacteria on coral reefs of Moorea Island (French Polynesia): diversity response to
401 habitat quality. *Hydrobiologia* 843(1):61–78

402

403 Zyllich K, Harper S, Licandeo R, Vega R, Zeller D, Pauly D. 2014. Fishing in Easter Island, a
404 recent history (1950-2010). *Latin American Journal of Aquatic Research* 42(4):845–856

Figure 1

Study area showing different ROV dive sites around Rapa Nui and coverage of filamentous mats at them

A) Dive sites and levels of coverage of filamentous mats on the benthic ecosystems. B) Depth range of ROV transect for the different categories of filamentous mats coverage used in the study. Green: no mats observed, Yellow: low coverage, Orange: high coverage, and Red: very high coverage. Box plots indicate median and 1 Standard Deviation, whiskers indicate depth range for this category.

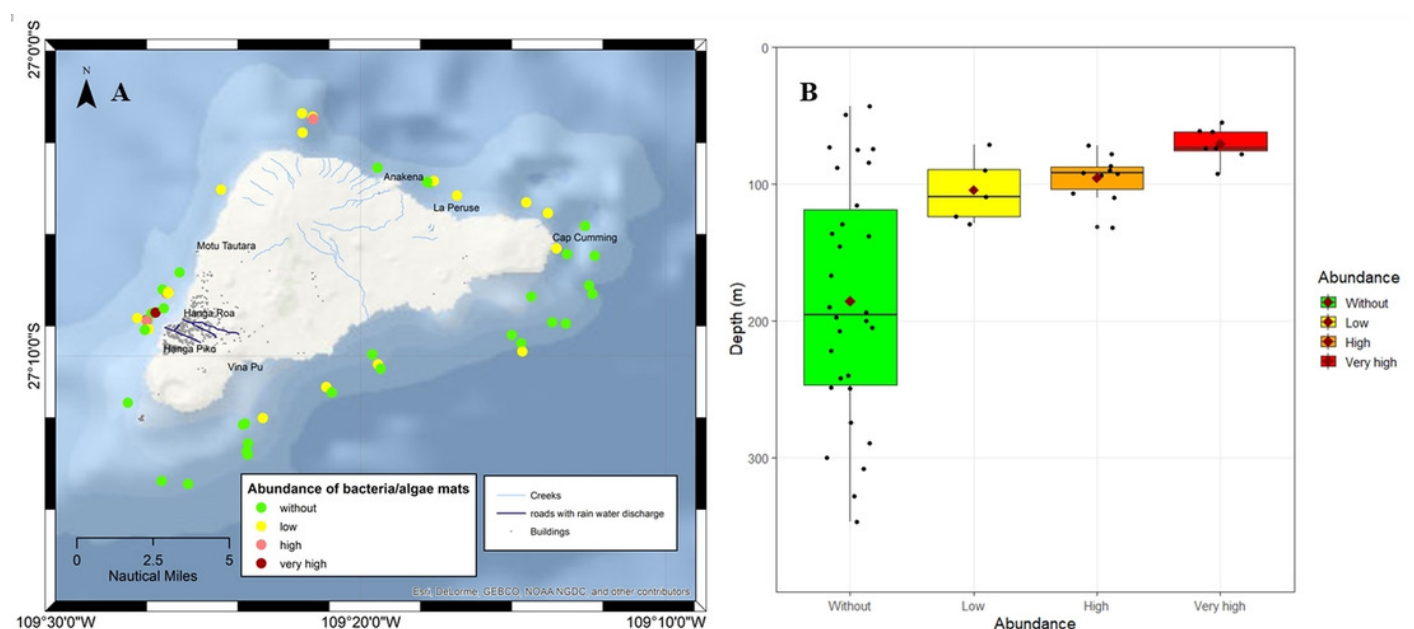


Figure 2

ROV images of the filamentous mats and mesophotic reefs off Rapa Nui. A) filamentous mats field ~80 m depth off Hanga Roa, Rapa Nui.

B) Close up of the filaments. C) Filaments among *C. vauhani* individuals. D) Dead *Leptoseris* sp. reef ca. 80 m depth overgrown by filaments. E) Healthy *Leptoseris* reef off Anakena ~ 80 m depth. F) Healthy *Leptoseris* reef off Hanga Roa filmed during prospective ROV surveys in “CIMAR-5 Islas” cruise made in 1999. Scale bars: 10 cm (A, B, C), 25 cm (D, E, F). Images: Matthias Gorny, OCEANA.

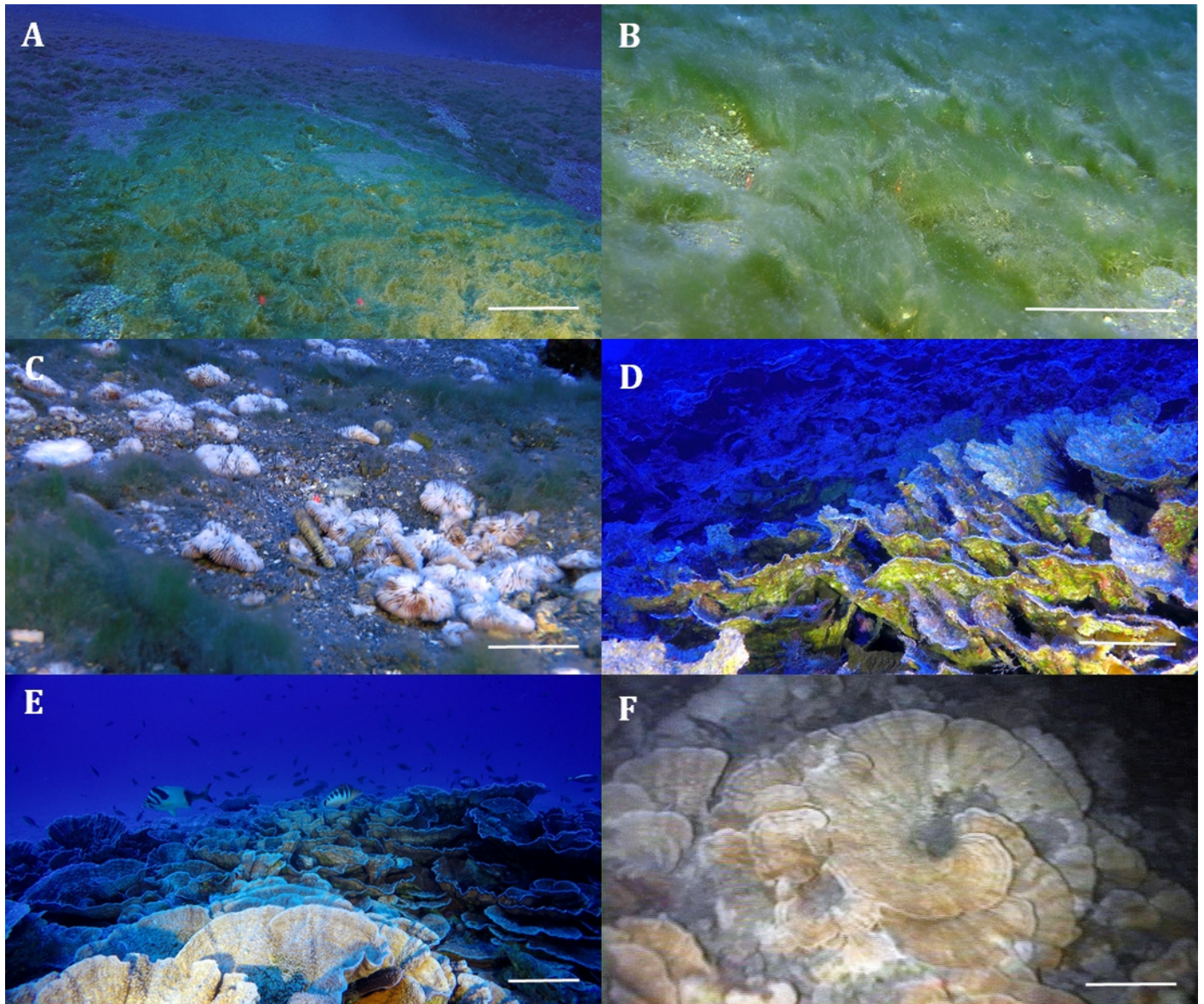


Figure 3

Location of mesophotic reefs off Rapa Nui in this study.

A) Map showing the position of reefs and main taxa constituting them recorded with ROV between 40-350 m. B) Map showing the different health statuses of *Leptoseris* sp. reefs. Green: Healthy (no noticeable impact), Yellow: Some damage (25-75% of corals damaged), and Red: Destroyed (only dead corals or fragments).

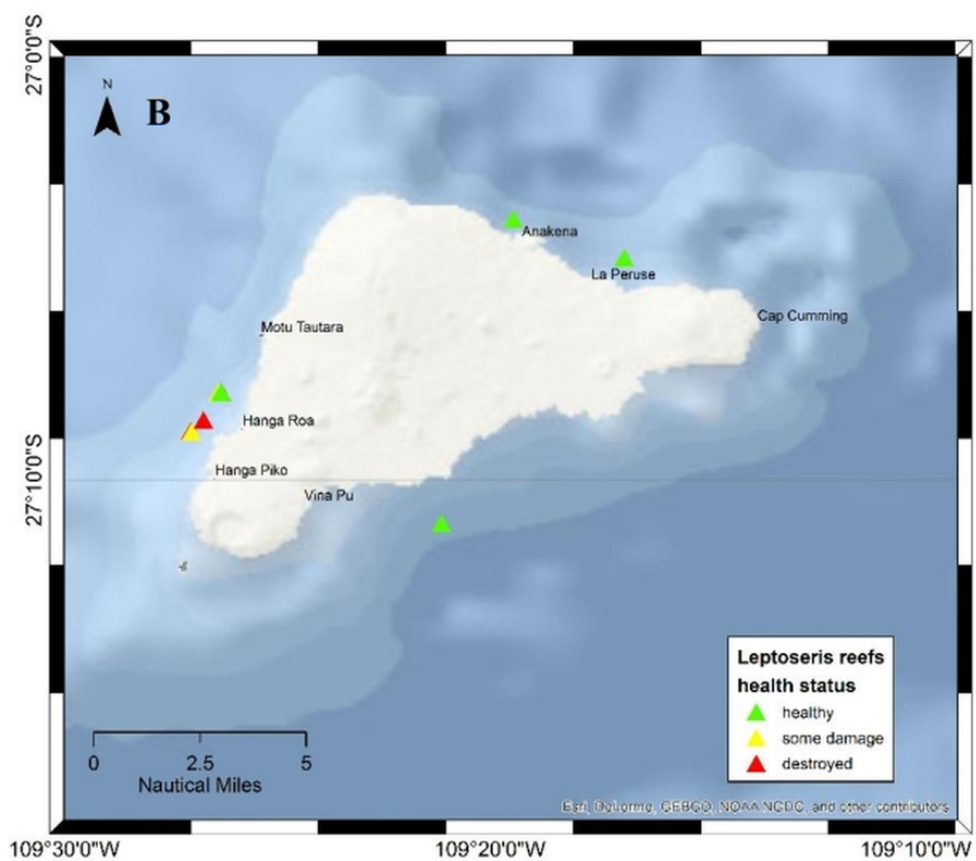
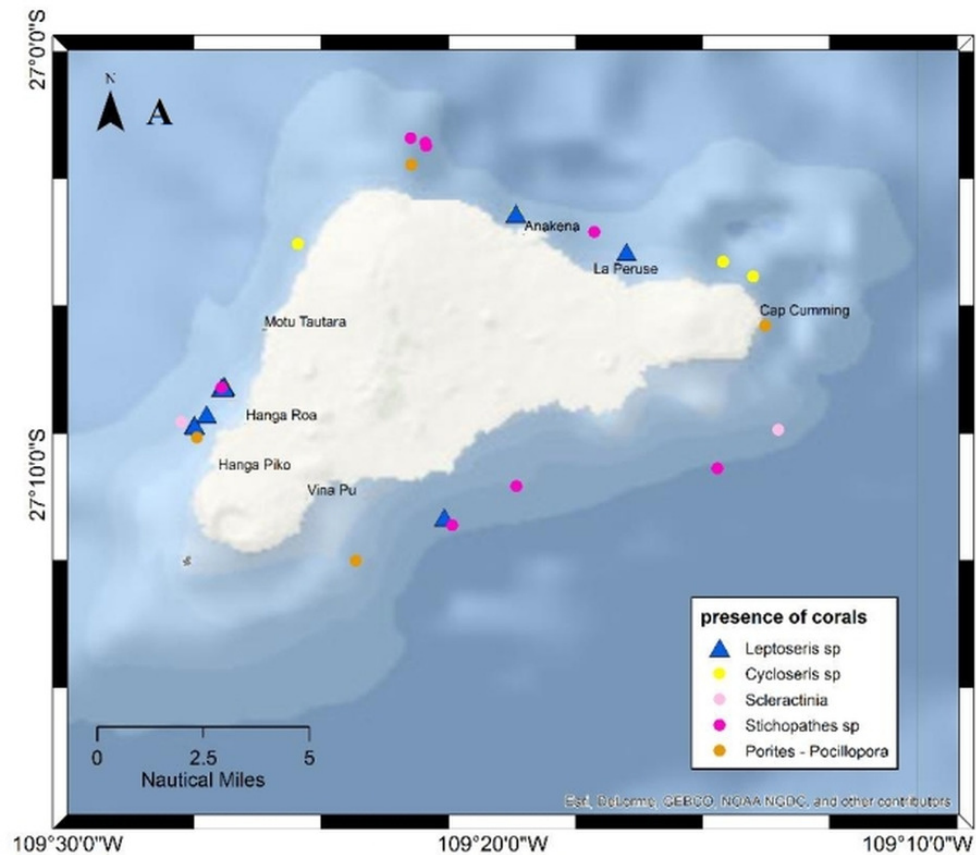


Figure 4

Micrographs of the four filamentous taxa collected off Hanga Roa, Rapa Nui at mesophotic depths using phase-contrast (A-E, and G) and epifluorescence (F and H) techniques.

(A, B) *Cladophora* sp. (C, D) *Ectocarpus* sp. (E, F) *Lyngbya* sp. (G, H) *Pseudoanabaena* sp.

Scale bars = A: 500 μm , B: 200 μm , C: 100 μm , D: 30 μm , E and F: 20 μm , G and H: 30 μm .

