Peer

Identification of a circadian gene signature that predicts overall survival in lung adenocarcinoma

Xinliang Gao¹, Mingbo Tang¹, Suyan Tian², Jialin Li¹ and Wei Liu¹

¹ Department of Thoracic Surgery, The First Hospital of Jilin University, Changchun, Jilin Province, China ² Division of Clinical Research, The First Hospital of Jilin University, Changchun, Jilin Province, China

ABSTRACT

Background. Lung adenocarcinoma (LUAD) is one of the most common subtypes of lung cancer which is the leading cause of death in cancer patients. Circadian clock disruption has been listed as a likely carcinogen. However, whether the expression of circadian genes affects overall survival (OS) in LUAD patients remains unknown. In this article, we identified a circadian gene signature to predict overall survival in LUAD. **Methods**. RNA sequencing (HTSeq-FPKM) data and clinical characteristics were obtained for a cohort of LUAD patients from The Cancer Genome Atlas (TCGA). A multigene signature based on differentially expressed circadian clock-related genes was generated for the prediction of OS using Least Absolute Shrinkage and Selection Operator (LASSO)-penalized Cox regression analysis, and externally validated using the GSE72094 dataset from the GEO database.

Results. Five differentially expressed genes (DEGs) were identified to be significantly associated with OS using univariate Cox proportional regression analysis (P < 0.05). Patients classified as high risk based on these five DEGs had significantly lower OS than those classified as low risk in both the TGCA cohort and GSE72094 dataset (P < 0.001). Multivariate Cox regression analysis revealed that the five-gene-signature based risk score was an independent predictor of OS (hazard ratio > 1, P < 0.001). Receiver operating characteristic (ROC) curves confirmed its prognostic value. Gene set enrichment analysis (GSEA) showed that Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways related to cell proliferation, gene damage repair, proteasomes, and immune and autoimmune diseases were significantly enriched.

Conclusion. A novel circadian gene signature for OS in LUAD was found to be predictive in both the derivation and validation cohorts. Targeting circadian genes is a potential therapeutic option in LUAD.

Subjects Bioinformatics, Oncology, Respiratory Medicine, Medical Genetics **Keywords** Lung adenocarcinoma, Circadian clock, Overall survival, Gene signature, TCGA

INTRODUCTION

Lung cancer is a leading cause of death in the world (*Bray et al., 2018*). The estimated 5-year survival rate is only 19% (*Siegel, Miller & Jemal, 2019*). In 2019, there were 228,150 new diagnoses of cancers of the lung and bronchus in the United States. Primary lung cancer is divided into two main types: small-cell lung carcinoma and non-small cell lung carcinoma (NSCLC). The latter is further classified into different subtypes according to the

Submitted 3 February 2021 Accepted 16 June 2021 Published 8 July 2021

Corresponding author Wei Liu, l_w01@jlu.edu.cn

Academic editor Katherine Mitsouras

Additional Information and Declarations can be found on page 11

DOI 10.7717/peerj.11733

Copyright 2021 Gao et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

histological origin, such as lung adenocarcinoma (LUAD), squamous cell carcinoma, or large cell carcinoma. Among these, LUAD is the most prevalent subtype, with an increasing incident in recent years (*Cheng et al., 2016*). The prognosis of LUAD is improving due to advances in molecular targeted treatment and immunotherapy (*Hirsch et al., 2016*; *Peters et al., 2019*). However, accurate prognosis prediction models for LUAD are still lacking.

The circadian clock is a molecular time-keeping system that is evolutionarily conserved. It is vital for the maintenance of physiologic homeostasis and normal function in all organisms. It coordinates a variety of biological processes and behaviors (*Fu & Kettner*, 2013; Panda et al., 2002). In the suprachiasmatic nucleus (SCN) of the hypothalamus, a central clock maintains the daily rhythms in the body by neural and humoral communication with peripheral clocks located in peripheral tissues and regulates bodily functions such as sleep/wake cycles and the secretion of many hormones. Disruption of the circadian clock has been listed as a likely carcinogen by the World Health Organization based on both population and laboratory-based findings (*Lunn et al., 2017*; Straif et al., 2007), which raised the interest in research on the relationship between circadian genes and tumor development. Some circadian genes have been demonstrated to control the occurrence and development of NSCLC (*Qiu et al., 2019*). However, the association between circadian genes and prognosis in patients with LUAD remains to be elucidated.

The present study aims to explore the prognostic role of circadian genes in patients with LUAD using The Cancer Genome Atlas (TCGA) data obtained from the NCI Genomic Data Commons, which includes the clinical characteristics and mRNA expression profiles of tumor and tumor-adjacent normal tissues. A prognostic multigene signature will be established using differentially expressed circadian clock genes and then validated with the GSE72094 dataset extracted from the Gene Expression Omnibus (GEO) database. Underlying molecular mechanisms were investigated by performing a Gene set enrichment analysis (GSEA) with Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways.

MATERIALS & METHODS

Data collection

The clinical characteristics and RNA sequencing data (HTSeq-FPKM) of 515 patients with LUAD were retrieved from the NCI Genomic Data Commons (https://portal.gdc. cancer.gov/repository). These 515 patients provided 535 samples from LUAD tumor tissue and 59 samples from adjacent normal tissue. Among the patients, 500 had complete RNA sequencing data and 469 had both complete sequencing data and complete clinical information.

The differential expression of the following 14 core genes of the circadian clock according to previous literature was analyzed: *Period 1 (PER1)*, *PER2*, *PER3*, Cryptochrome Circadian Regulator 1 (CRY1), CRY2, Circadian Locomotor Output Cycles Kaput (CLOCK), Aryl Hydrocarbon Receptor Nuclear Translocator Like (ARNTL), Timeless Circadian Regulator (TIMELESS), Neuronal PAS Domain Protein 2 (NPAS2), Nuclear Receptor Subfamily 1 Group D Member 1 (NR1D1), NR1D2, Basic Helix-Loop-Helix Family Member E40 (BHLHE40), BHLHE41, and RAR-Related Orphan Receptor A (RORA) (Chen et al., 2020; *Cox & Takahashi, 2019; Mocellin et al., 2018; Shafi & Knudsen, 2019; Yu et al., 2019).* The validation dataset was obtained from the GSE72094 dataset in the GEO database (https://www.ncbi.nlm.nih.gov/geo/) and included microarray and clinical data for 443 LUAD tumor samples (*Schabath et al., 2016*). The normalized count data were downloaded. The data cut-off date was September 10, 2020. Patients with no follow-up data or information on the expression of circadian genes were excluded.

The TCGA and GEO databases are public data repositories and therefore, ethical approval for this study was not required. This study followed the polices and guidelines for data access and publication specified by the TCGA and GEO databases.

Prognostic validity of the gene signature

Differentially expressed genes (DEGs) involved in the circadian clock were analyzed in the tumor and tumor-adjacent normal tissues of LUAD patients from the TCGA cohort using the "limma" package in R (false discovery rate (FDR) <0.05). Univariate Cox regression analysis was used to identify circadian genes related to overall survival (OS). A gene signature for the prediction of OS was constructed with the DEGs for the circadian clock using Least Absolute Shrinkage and Selection Operator (LASSO)-penalized Cox regression analysis and the "glmnet" package in R. DEGs served as independent variables, and OS as the response variable.

A risk score based on the expression of identified candidate genes was calculated for each patient according to the following formula: score = sum (normalized gene expression level × regression coefficient). Patients were classified as either high- or low-risk using the median score as the cut-off value. The survival analysis of different risk groups was determined with the "survminer" R package. In order to validate the performance of the signature, we used the principal components analysis (PCA) and t-distributed stochastic neighbor embedding (t-SNE) to analyze dimensionality reduction. The "prcomp" function in the R "stats" package was used to carry out the PCA. The data distribution for high-risk and low-risk patients was also mapped using t-SNE and the "Rtsne" package in R. The predictive value of the gene signature was evaluated with time-dependent Receiver operating characteristics (ROC) curve analysis using the "timeROC" package in R. The associations between the risk score, clinical characteristics (gender, age, smoking history, and stage), and OS were assessed with univariate and multivariate Cox regression analyses.

Functional enrichment analysis

The DEGs between the high- and low-risk groups in the TCGA LUAD cohort were identified using the "limma" R package again. GSEA of these DEGs was carried out with KEGG pathways ($|\log 2 \text{ fold change}| \ge 1$, FDR <0.05). Both a nominal *P*-value <0.05 and FDR *q*-value <0.05 were considered statistically significant.

Statistical analysis

All statistical analyses were conducted with R software (Version 3.5.3) and SPSS software (Version 25.0). Gene expression was compared using the two-tailed Student's *t*-test and proportions were compared using the Chi-squared test. The Kaplan–Meier method and the log-rank test were used to assess the differences in OS between high and low-risk patients.

Table 1 Demographical and emiliar characteristics.						
	TCGA LUAD	GSE72094	P value			
No. of patients	469	328				
Age (median, range)	65.1 (33–88)	69.7 (41-89)	P < 0.01			
Gender (%)	257 (54.8%)	172 (52.4%)	P = 0.559			
Female	212 (45.2%)	156 (47.6%)				
Male						
TNM Stage	257 (54.8%)	218 (66.5%)	P = 0.007			
Ι	112 (23.9%)	53 (16.2%)				
II	75 (16.0%)	46 (14.0%)				
III	25 (5.3%)	11 (3.4%)				
IV						
Smoking history	69 (14.7%)	30 (9.1%)	P = 0.025			
Non-smoker	400 (85.3%)	298 (90.9%)				
Smoker						
Median OS (days)	629	842	P = 0.034			

Univariate and multivariate Cox regression analyses were used to identify independent predictors of OS. P < 0.05 (two tailed) was considered statistically significant.

RESULTS

Clinical and demographic characteristics

Table 1 Demographical and clinical characteristics

Two patient cohorts with available data on OS and the RNA expression of circadian clock genes were used to create the prognostic model. The derivation cohort consisted of 500 patients with LUAD and complete RNA sequencing data from the TCGA database while the validation cohort consisted of 398 patients with LUAD from the GSE72094 dataset. Among these patients, 469 patients from TCGA and 328 patients from GSE72094 who not only had complete RNA sequencing data, but also complete clinical data including OS, age, gender, smoking history, and tumor stage, were included in the univariate and multivariate COX analyses. The validation cohort had higher age, lower TNM stage, more smokers, and a higher median OS compared to the derivation cohort. The baseline demographic and clinical characteristics of the included patients are summarized in Table 1.

Identification of DEGs related to circadian clock in the TGCA LUAD cohort

In the TCGA LUAD cohort, 9/14 circadian genes were found to be differentially expressed between tumor and tumor-adjacent normal tissues. Five candidate genes were identified to be significantly associated with OS using univariate Cox proportional regression analysis (Figs. 1A–1B). The clustering of the 5 candidate genes are shown with a heatmap in Fig. 1C.

Generation of a prognostic signature in the TGCA LUAD cohort

The 5 identified candidate genes were incorporated into a five-gene-signature based prognostic model using LASSO Cox regression analysis. According to risk scores calculated using the expression levels of these 5 genes, half of the patients were classified as high-risk (n = 250) and the other half as low-risk (n = 250) (Fig. 2A). The chance of survival was



Figure 1 Identification of the candidate genes involved in the circadian cycle in the TCGA cohort. (A) Venn diagram of DEGs and prognostic genes that correlate with OS in tumor and tumor-adjacent normal tissue. (B) Forest plots of the five genes that overlap between DEGs and prognostic genes that relate to OS on univariate Cox regression analysis. (C) The mRNA heatmap of five candidate genes. (D–H) The expression of five candidate genes in tumor and normal tissue.

Full-size 🖾 DOI: 10.7717/peerj.11733/fig-1



Figure 2 Analysis of the prognostic value of the five-gene-signature based risk score in the TCGA cohort. (A) The distribution of the risk scores in the TCGA cohort. (B) The distributions of OS status, OS time and risk score in the TCGA cohort. (C) PCA plot of the TCGA cohort. (D) t-SNE analysis of the TCGA cohort. (E) Kaplan–Meier curves for the OS of patients in the high- respective low-risk group in the TCGA cohort. (F) AUC of time-dependent ROC curves that confirm the prognostic performance of the risk score in the TCGA cohort.

Full-size DOI: 10.7717/peerj.11733/fig-2

lower and the survival time was shorter in the high-risk group than in the low-risk group (Fig. 2B). PCA and t-SNE analysis showed discernible dimensions between high-risk and low-risk patients (Figs. 2C–2D). Kaplan–Meier survival curves confirmed that OS was significantly worse in high-risk than in low-risk patients (Fig. 2E, P < 0.001). The predictive performance of the five-gene-signature based risk score for OS was evaluated using time-dependent ROC curves. The area under the curve (AUC) values were: 1 year, 0.726; 2 years, 0.668; and 3 years, 0.657 (Fig. 2F).

Validation of the five-gene-signature based prognostic model

The stringency of the model developed using the TGCA LUAD cohort was validated in the GSE72094 dataset. Risk scores were calculated for all patients based on the expression levels of the 5 identified candidate genes and patients were classified as high-risk or low-risk accordingly (Fig. 3A). The high-risk group had a significantly higher chance of death and lower OS time (Fig. 3B). PCA and t-SNE analysis showed discernible dimensions between high- and low-risk patients (Figs. 3C–3D). Kaplan–Meier survival curves confirmed that OS was significantly worse in high-risk patients (Fig. 3E, P < 0.001). The AUC values were: 1 year, 0.621; 2 years, 0.657; and 3 years, 0.642 (Fig. 3F).

Prognostic value of the five-gene-signature based risk score

Univariate and multivariate Cox regression analyses were conducted to determine whether the five-gene-signature based risk score was an independent predictor of OS (Table 2). The derivation cohort consisted of 469 patients from the TCGA LUAD cohort; and



Figure 3 Validation of the five-gene-signature based risk score in the GSE72094 dataset. (A) The distribution of the risk scores in the GSE72094 dataset. (B) The distributions of OS status, OS time and risk score in the GSE72094 dataset. (C) PCA plot of the GSE72094 dataset. (D) t-SNE analysis of the GSE72094 dataset. (E) Kaplan–Meier curves for the OS of patients in the high- respective low-risk group in the GSE72094 dataset. (F) AUC of time-dependent ROC curves that confirm the prognostic performance of the risk score in the GSE72094 dataset.

Full-size DOI: 10.7717/peerj.11733/fig-3

Factors	TCGA LUAD					GSE72094						
		Univariate Multivariate		Univariate		Multivariate						
	HR	95% CI	Р	HR	95% CI	Р	HR	95% CI	Р	HR	95% CI	Р
Age	1.01	0.99-1.02	0.302	1.02	1.00-1.03	0.026	1.01	0.98-1.03	0.533	1.00	0.98-1.02	0.973
Gender	1.12	0.83-1.52	0.458	1.09	0.79-1.49	0.606	1.99	1.30-3.05	0.002	2.32	1.49-3.62	0.000
Stage	1.61	1.39–1.85	0.000	1.59	1.37-1.84	0.000	1.70	1.40-2.07	0.000	1.86	1.52-2.29	0.000
Smoking	0.91	0.60-1.38	0.655	0.81	0.53-1.26	0.351	1.31	0.57-3.01	0.523	0.87	0.37-2.03	0.750
Risk score	3.37	2.20-5.17	0.000	3.53	2.26-5.49	0.000	2.16	1.39–3.36	0.001	2.20	1.41-3.41	0.000

Table 2 Risk factors affecting OS in the TCGA LUAD cohort and GSE72094.

the validation cohort consisted of 328 patients from the GSE72094 dataset. Univariate regression analysis revealed that the risk score was significantly associated with OS in both the TGCA LUAD cohort and the GSE72094 dataset (TGCA LUAD cohort: hazard ratio (HR) = 3.373, 95% confidence interval (CI) = 2.199-5.174, P < 0.001; GSE72094 dataset: HR = 2.163 95% CI = 1.393-3.358, P < 0.001). The risk score was found to be an independent predictor of OS even after correcting for confounders in multivariate Cox regression analysis (TGCA LUAD cohort: HR = 3.522, 95% CI = 2.260-5.487, P < 0.001; GSE72094 dataset: HR = 2.195, 95% CI = 1.411-3.415, P < 0.001; Figs. 4A–4B).



Figure 4 Multivariate Cox regression analyses of factors affecting OS in the TCGA LUAD cohort (A) and the GSE72094 dataset (B).

Full-size DOI: 10.7717/peerj.11733/fig-4

Enrichment analysis in the TGCA LUAD cohort

Genes that were differentially expressed in the high- respective low-risk groups were subjected to GSEA for KEGG pathways (Table 3). The results showed that tumorigenesis pathways related to pyrimidine metabolism, cell cycle, proteasome, base excision repair, homologous recombination, and DNA replication were enriched (Fig. 5).

DISCUSSION

Genes of the circadian clock are often abnormally expressed in tumor tissues and may play an important role in tumorigenesis (*Kelleher, Rao & Maguire, 2014*; *Kettner, Katchy & Fu, 2014*). The present study identified 9 DEGs between tumor and tumor-adjacent normal tissues among the 14 circadian genes. The genes *PER3, CRY2, TIMELESS, NPAS2*, and *RORA* were found to be correlated with OS. These results suggest that circadian clock genes may affect the survival outcome in LUAD and that a signature based on the expression of these genes may predict OS and may be an independent prognostic factor.

The PER family is generally considered to have a tumor suppressor effect, and the mechanisms behind the tumor suppressing effects of *PER1* and *PER2* are clear (*Gery et al., 2006*; *Wood et al., 2008*). *PER3* has been confirmed to affect the susceptibility and prognosis of lung cancer through expression changes, methylation, and single nucleotide polymorphisms (SNPs) (*Chu et al., 2018*; *Couto et al., 2014*; *Liu et al., 2014*). However, the exact mechanism for the *PER3* inhibition of tumors is not yet clear. The study by Jun-Sub et al. showed that *PER3* is required for *checkpoint kinase 2* (*CHK2*) activation in human cells, which highlighted its potential role in cell cycle arrest and DNA damage repair (*Im et al., 2010*). Previous studies have linked the circadian clock gene *CRY2* to the occurrence and development of many tumors (*Hasakova et al., 2018*; *Lesicka et al., 2018*; *Relles et al., 2013*; *Tokunaga et al., 2008*). As a transcriptional suppressor, *CRY2* functions as an important regulator of cell cycle, proliferation, DNA damage checkpoint control, and DNA

Table 5 GSEA of DEGs between high-risk group and low-risk group with REGG pathways.									
KEGG pathway	NES	NOM p-val	FDR q-val						
PYRIMIDINE_METABOLISM	2.17	0.000	0.000						
CELL_CYCLE	2.11	0.000	0.003						
SPLICEOSOME	2.08	0.000	0.005						
PROTEASOME	2.05	0.000	0.006						
BASE_EXCISION_REPAIR	2.02	0.000	0.009						
HOMOLOGOUS_RECOMBINATION	1.97	0.000	0.015						
PATHOGENIC_ESCHERICHIA_COLI_INFECTION	1.97	0.000	0.025						
DNA_REPLICATION	1.91	0.000	0.025						
GLYCOSPHINGOLIPID_BIOSYNTHESIS_LACTO_AND_NEOLACTO_SERIES	1.89	0.000	0.030						
PENTOSE_PHOSPHATE_PATHWAY	1.88	0.006	0.030						
THYROID_CANCER	1.87	0.002	0.032						
INTESTINAL_IMMUNE_NETWORK_FOR_IGA_PRODUCTION	-2.11	0.000	0.011						
HEMATOPOIETIC_CELL_LINEAGE	-2.04	0.002	0.018						
ASTHMA	-1.95	0.002	0.039						
AUTOIMMUNE_THYROID_DISEASE	-1.95	0.006	0.030						
CELL_ADHESION_MOLECULES_CAMS	-1.95	0.002	0.025						
PRIMARY_BILE_ACID_BIOSYNTHESIS	-1.91	0.000	0.032						
GLYCOSPHINGOLIPID_BIOSYNTHESIS_GANGLIO_SERIES	-1.90	0.004	0.029						
ALLOGRAFT_REJECTION	-1.89	0.010	0.031						
TYPE_I_DIABETES_MELLITUS	-1.88	0.014	0.031						

...

....

repair (Hoffman et al., 2010). CRY2 acts as a tumor suppressor gene. It can limit tumor formation by increasing c-MYC turnover (Huber et al., 2016), or increase the elimination of premalignant and malignant cells through the activation of p53-independent apoptosis pathways (Lee & Sancar, 2011). The circadian genes NPAS2 and TIMELESS, on the other hand, are both correlated with poor OS. A recent study has shown that upregulated NPAS2 promoted the survival of hepatocellular carcinoma cells through the upregulation of cell division cycle 25 A (CDC25A) and inhibition of mitochondria-dependent intrinsic apoptosis (Yuan et al., 2017). Knockout or inhibition of TIMELESS can lead to cell cycle stagnation and subsequent apoptosis, which limits the growth of liver cancer cells (*Elgohary* et al., 2015). The circadian gene RORA was found to be downregulated in LUAD tissue and negatively correlated with LUAD prognosis in this study. RORA is a versatile gene. Besides the circadian clock, it is also a well-known regulator of inflammation and lipid metabolism. Moreover, recent studies have suggested that RORA may also play a role in the progression and prognosis of colon cancer and breast cancer (*Lee et al., 2010*). The recruitment of RORA can induce the expression of the tumor suppressor genes F-box/WD repeat-containing protein 7 (FBXW7), Semaphorin 3F (SEMA3F), and P21, leading to apoptosis and suppression of tumor cell proliferation (Wang et al., 2017).

Results from the enrichment analysis revealed that metabolic pathways related to the substrates of DNA synthesis (pyrimidine metabolism and pentose phosphate pathway) were enriched in the high-risk group, as well as pathways regulating cell cycle and DNA replication. Increasing evidence suggests a regulatory effect of circadian genes on cellular



Figure 5 The KEGG enrichment plots of tumorigenesis pathways. (A) Pyrimidine metabolism. (B) Cell cycle. (C) Proteasome. (D) Base excision repair. (E) Homologous recombination. (F) DNA replication. Full-size DOI: 10.7717/peerj.11733/fig-5

proliferation (Chakrabarti & Michor, 2020), and their involvement in the proliferation of a variety of tumor cells (Abreu et al., 2018; Wang et al., 2016; Yu et al., 2018). A recent study on lung cancer demonstrated that the loss of the central clock components led to increased c-MYC expression, which enhanced proliferation (*Papagiannakopoulos et al.*, 2016). Base excision repair and homologous recombination pathways were also found to be enriched in the high-risk group, which may indicate that the circadian clock disorder affects the repair of gene damage to influence the survival of malignant tumors. Both CRY and TIMELESS are known to be involved in DNA damage repair. Tae et al. found that CRY s are related to the nucleotide excision repair gene XPA (Kang, Reardon & Sancar, 2011). TIMELESS can modulate CHK1 and serine/threonine-protein kinase (ATR) downstream of single-strand DNA breaks and activate CHK2 via ATM modulation downstream of double strand breaks (Yang, Wood & Hrushesky, 2010). The proteasome pathway was enriched in the high-risk group. Recent studies have also confirmed that some ubiquitin ligases participate in the degradation of core circadian clock genes through the ubiquitinproteasome pathway, thereby controlling the biological functions of cells, including cell senescence (Chen et al., 2018; Ullah et al., 2020). This cross-talk between circadian clock genes and the ubiquitin-proteasome pathway may be related to the prognosis of LUAD. Some immune and autoimmune disease pathways were enriched in the low-risk group. This shows that the disturbance of the circadian clock is accompanied by alterations in the function of the immune system (Aiello et al., 2020), which may be related to the

occurrence, development, and prognosis of LUAD. Wu and his colleagues have shown that abnormal circadian genes contribute to T cell exhaustion and global upregulation of immune inhibitory molecules, such as programmed death-ligand 1 (PD-L1) and cytotoxic T-lymphocyte antigen (CTLA)-4, which promote tumor development (*Wu et al., 2019*).

There are several limitations to this study. Firstly, the present study is a retrospective study with data from publicly available databases. This makes the study more prone to selection bias and it is also impossible to draw conclusions regarding cause–effect. Experimental studies should be conducted to understand the mechanisms behind the role of the circadian genes. Secondly, using tumor-adjacent normal tissue as a control has the advantages of minimizing biological variation, but one cannot be sure if the seemingly "normal" tissue adjacent to a tumor is truly "normal". Thirdly, while there might be many other genes that are important in LUAD, we only focused on 14 core genes of the circadian clock. It is possible that other more important genes were excluded from the design.

CONCLUSIONS

In summary, we constructed a novel five-gene signature with genes involved in the circadian clock to predict the prognosis of LUAD. The signature could successfully separate LUAD patients with a low risk of non-survival from those with a high risk in both the derivation and validation cohorts. The underlying molecular mechanisms between circadian genes and tumor proliferation, DNA repair, ubiquitin-proteasome pathway, and immunity in LUAD remain poorly understood. and warrant further investigation. Circadian genes might be potential targets for future cancer therapy.

ACKNOWLEDGEMENTS

This manuscript has been edited and proofread by Medjaden Bioscience Limited.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by the Jilin Province Scientific and Technological Department, International Scientific and Technological Cooperation Project (20190701043GH), Wu Jieping Medical Foundation (No. 320.6750.19092-1), and the Development Center for Medical Science & Technology, National Health Commission of the People's Republic of China (No. WA2020RW18). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors: Jilin Province Scientific and Technological Department, International Scientific and Technological Cooperation Project: 20190701043GH. Wu Jieping Medical Foundation: 320.6750.19092-1. Development Center for Medical Science & Technology,National Health Commission of the People's Republic of China: WA2020RW18.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Xinliang Gao conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Mingbo Tang and Jialin Li performed the experiments, prepared figures and/or tables, and approved the final draft.
- Suyan Tian conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Wei Liu conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

Data is available at TCGA-LUAD (https://portal.gdc.cancer.gov/repository), and at NCBI GEO: GSE72094.

REFERENCES

- Abreu M, Basti A, Genov N, Mazzoccoli G, Relógio A. 2018. The reciprocal interplay between TNF α and the circadian clock impacts on cell proliferation and migration in Hodgkin lymphoma cells. *Scientific Reports* 8:11474 DOI 10.1038/s41598-018-29847-z.
- Aiello I, Fedele MLM, Román F, Marpegan L, Caldart C, Chiesa JJ, Golombek DA, Finkielstein CV, Paladino N. 2020. Circadian disruption promotes tumor-immune microenvironment remodeling favoring tumor cell proliferation. *Science Advances* 6:eaaz4530 DOI 10.1126/sciadv.aaz4530.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. 2018. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians* 68:394–424 DOI 10.3322/caac.21492.
- Chakrabarti S, Michor F. 2020. Circadian clock effects on cellular proliferation: Insights from theory and experiments. *Current Opinion in Cell Biology* 67:17–26 DOI 10.1016/j.ceb.2020.07.003.
- Chen J, Liu A, Lin Z, Wang B, Chai X, Chen S, Lu W, Zheng M, Cao T, Zhong M, Li R, Wu M, Lu Z, Pang W, Huang W, Xiao L, Lin D, Wang Z, Lei F, Chen X, Long W, Zheng Y, Chen Q, Zeng J, Ren D, Li J, Zhang X, Huang Y. 2020. Downregulation of the circadian rhythm regulator HLF promotes multiple-organ distant metastases in non-small cell lung cancer through PPAR/NF-κb signaling. *Cancer Letters* 482:56–71 DOI 10.1016/j.canlet.2020.04.007.
- Chen S, Yang J, Yang L, Zhang Y, Zhou L, Liu Q, Duan C, Mieres CA, Zhou G, Xu G. 2018. Ubiquitin ligase TRAF2 attenuates the transcriptional activity of the core clock

protein BMAL1 and affects the maximal Per1 mRNA level of the circadian clock in cells. *Febs Journal* **285**:2987–3001 DOI 10.1111/febs.14595.

- Cheng TY, Cramb SM, Baade PD, Youlden DR, Nwogu C, Reid ME. 2016. The international epidemiology of lung cancer: latest trends, disparities, and tumor characteristics. *Journal of Thoracic Oncology* 11:1653–1671 DOI 10.1016/j.jtho.2016.05.021.
- Chu LW, Till C, Yang B, Tangen CM, Goodman PJ, Yu K, Zhu Y, Han S, Hoque AM, Ambrosone C, Thompson I, Leach R, Hsing AW. 2018. Circadian genes and risk of prostate cancer in the prostate cancer prevention trial. *Molecular Carcinogenesis* 57:462–466 DOI 10.1002/mc.22770.
- Couto P, Miranda D, Vieira R, Vilhena A, De Marco L, Bastos-Rodrigues L. 2014. Association between CLOCK, PER3 and CCRN4L with non-small cell lung cancer in Brazilian patients. *Molecular Medicine Reports* 10:435–440 DOI 10.3892/mmr.2014.2224.
- **Cox KH, Takahashi JS. 2019.** Circadian clock genes and the transcriptional architecture of the clock mechanism. *Journal of Molecular Endocrinology* **63**:R93–r102 DOI 10.1530/jme-19-0153.
- Elgohary N, Pellegrino R, Neumann O, Elzawahry HM, Saber MM, Zeeneldin AA, Geffers R, Ehemann V, Schemmer P, Schirmacher P, Longerich T. 2015. Protumorigenic role of Timeless in hepatocellular carcinoma. *International Journal of Oncology* 46:597–606 DOI 10.3892/ijo.2014.2751.
- **Fu L, Kettner NM. 2013.** The circadian clock in cancer development and therapy. *Progress in Molecular Biology and Translational Science* **119**:221–282 DOI 10.1016/b978-0-12-396971-2.00009-9.
- Gery S, Komatsu N, Baldjyan L, Yu A, Koo D, Koeffler HP. 2006. The circadian gene per1 plays an important role in cell growth and DNA damage control in human cancer cells. *Molecular Cell* 22:375–382 DOI 10.1016/j.molcel.2006.03.038.
- Hasakova K, Vician M, Reis R, Zeman M, Herichova I. 2018. Sex-dependent correlation between survival and expression of genes related to the circadian oscillator in patients with colorectal cancer. *Chronobiology International* **35**:1423–1434 DOI 10.1080/07420528.2018.1488722.
- Hirsch FR, Suda K, Wiens J, Bunn Jr PA. 2016. New and emerging targeted treatments in advanced non-small-cell lung cancer. *The Lancet* **388**:1012–1024 DOI 10.1016/s0140-6736(16)31473-8.
- Hoffman AE, Zheng T, Ba Y, Stevens RG, Yi CH, Leaderer D, Zhu Y. 2010. Phenotypic effects of the circadian gene Cryptochrome 2 on cancer-related pathways. *BMC Cancer* 10:110 DOI 10.1186/1471-2407-10-110.
- Huber AL, Papp SJ, Chan AB, Henriksson E, Jordan SD, Kriebs A, Nguyen M, Wallace M, Li Z, Metallo CM, Lamia KA. 2016. CRY2 and FBXL3 Cooperatively Degrade c-MYC. *Molecular Cell* 64:774–789 DOI 10.1016/j.molcel.2016.10.012.
- Im JS, Jung BH, Kim SE, Lee KH, Lee JK. 2010. Per3, a circadian gene, is required for Chk2 activation in human cells. *FEBS Letters* **584**:4731–4734 DOI 10.1016/j.febslet.2010.11.003.

- Kang TH, Reardon JT, Sancar A. 2011. Regulation of nucleotide excision repair activity by transcriptional and post-transcriptional control of the XPA protein. *Nucleic Acids Research* 39:3176–3187 DOI 10.1093/nar/gkq1318.
- Kelleher FC, Rao A, Maguire A. 2014. Circadian molecular clocks and cancer. *Cancer Letters* 342:9–18 DOI 10.1016/j.canlet.2013.09.040.
- Kettner NM, Katchy CA, Fu L. 2014. Circadian gene variants in cancer. Annals of Medicine 46:208–220 DOI 10.3109/07853890.2014.914808.
- Lee JM, Kim IS, Kim H, Lee JS, Kim K, Yim HY, Jeong J, Kim JH, Kim JY, Lee H, Seo SB, Kim H, Rosenfeld MG, Kim KI, Baek SH. 2010. RORalpha attenuates Wnt/beta-catenin signaling by PKCalpha-dependent phosphorylation in colon cancer. *Molecular Cell* 37:183–195 DOI 10.1016/j.molcel.2009.12.022.
- Lee JH, Sancar A. 2011. Regulation of apoptosis by the circadian clock through NF-kappaB signaling. *Proceedings of the National Academy of Sciences U S A* 108:12036–12041 DOI 10.1073/pnas.1108125108.
- Lesicka M, Jabłońska E, Wieczorek E, Seroczyńska B, Siekierzycka A, Skokowski J, Kalinowski L, Wąsowicz W, Reszka E. 2018. Altered circadian genes expression in breast cancer tissue according to the clinical characteristics. *PLOS ONE* 13:e0199622 DOI 10.1371/journal.pone.0199622.
- Liu B, Xu K, Jiang Y, Li X. 2014. Aberrant expression of Per1, Per2 and Per3 and their prognostic relevance in non-small cell lung cancer. *International Journal of Clinical and Experimental Pathology* 7:7863–7871.
- Lunn RM, Blask DE, Coogan AN, Figueiro MG, Gorman MR, Hall JE, Hansen J, Nelson RJ, Panda S, Smolensky MH, Stevens RG, Turek FW, Vermeulen R, Carreón T, Caruso CC, Lawson CC, Thayer KA, Twery MJ, Ewens AD, Garner SC, Schwingl PJ, Boyd WA. 2017. Health consequences of electric lighting practices in the modern world: A report on the National Toxicology Program's workshop on shift work at night, artificial light at night, and circadian disruption. *Science of the Total Environment* 607-608:1073–1084 DOI 10.1016/j.scitotenv.2017.07.056.
- Mocellin S, Tropea S, Benna C, Rossi CR. 2018. Circadian pathway genetic variation and cancer risk: evidence from genome-wide association studies. *BMC Medicine* 16:20 DOI 10.1186/s12916-018-1010-1.
- Panda S, Antoch MP, Miller BH, Su AI, Schook AB, Straume M, Schultz PG, Kay SA, Takahashi JS, Hogenesch JB. 2002. Coordinated transcription of key pathways in the mouse by the circadian clock. *Cell* 109(02):307–320 DOI 10.1016/s0092-8674(02)00722-5.
- Papagiannakopoulos T, Bauer MR, Davidson SM, Heimann M, Subbaraj L,
 Bhutkar A, Bartlebaugh J, Vander Heiden MG, Jacks T. 2016. Circadian
 Rhythm Disruption Promotes Lung Tumorigenesis. *Cell Metabolism* 24:324–331
 DOI 10.1016/j.cmet.2016.07.001.
- Peters S, Reck M, Smit EF, Mok T, Hellmann MD. 2019. How to make the best use of immunotherapy as first-line treatment of advanced/metastatic non-small-cell lung cancer. *Annals of Oncology* **30**:884–896 DOI 10.1093/annonc/mdz109.

- Qiu M, Chen YB, Jin S, Fang XF, He XX, Xiong ZF, Yang SL. 2019. Research on circadian clock genes in non-small-cell lung carcinoma. *Chronobiology International* 36:739–750 DOI 10.1080/07420528.2018.1509080.
- Relles D, Sendecki J, Chipitsyna G, Hyslop T, Yeo CJ, Arafat HA. 2013. Circadian gene expression and clinicopathologic correlates in pancreatic cancer. *Journal of Gastrointestinal Surgery* 17:443–450 DOI 10.1007/s11605-012-2112-2.
- Schabath MB, Welsh EA, Fulp WJ, Chen L, Teer JK, Thompson ZJ, Engel BE, Xie M, Berglund AE, Creelan BC, Antonia SJ, Gray JE, Eschrich SA, Chen DT, Cress WD, Haura EB, Beg AA. 2016. Differential association of STK11 and TP53 with KRAS mutation-associated gene expression, proliferation and immune surveillance in lung adenocarcinoma. *Oncogene* 35:3209–3216 DOI 10.1038/onc.2015.375.
- Shafi AA, Knudsen KE. 2019. Cancer and the Circadian Clock. *Cancer Research* 79:3806–3814 DOI 10.1158/0008-5472.Can-19-0566.
- Siegel RL, Miller KD, Jemal A. 2019. Cancer statistics. *CA: A Cancer Journal for Clinicians* 69:7–34 DOI 10.3322/caac.21551.
- Straif K, Baan R, Grosse Y, Secretan B, El Ghissassi F, Bouvard V, Altieri A, Benbrahim-Tallaa L, Cogliano V. 2007. Carcinogenicity of shift-work, painting, and fire-fighting. *The Lancet Oncology* 8(07):1065–1066 DOI 10.1016/s1470-2045(07)70373-x.
- Tokunaga H, Takebayashi Y, Utsunomiya H, Akahira J, Higashimoto M, Mashiko M, Ito K, Niikura H, Takenoshita S, Yaegashi N. 2008. Clinicopathological significance of circadian rhythm-related gene expression levels in patients with epithelial ovarian cancer. Acta. *Obstetrics and Gynecology Scand* **87**:1060–1070 DOI 10.1080/00016340802348286.
- **Ullah K, Chen S, Lu J, Wang X, Liu Q, Zhang Y, Long Y, Hu Z, Xu G. 2020.** The E3 ubiquitin ligase STUB1 attenuates cell senescence by promoting the ubiquitination and degradation of the core circadian regulator BMAL1. *Journal of Biological Chemistry* **295**:4696–4708 DOI 10.1074/jbc.RA119.011280.
- Wang Q, Ao Y, Yang K, Tang H, Chen D. 2016. Circadian clock gene Per2 plays an important role in cell proliferation, apoptosis and cell cycle progression in human oral squamous cell carcinoma. *Oncology Reports* 35:3387–3394 DOI 10.3892/or.2016.4724.
- Wang Z, Xiong F, Wang X, Qi Y, Yu H, Zhu Y, Zhu H. 2017. Nuclear receptor retinoidrelated orphan receptor alpha promotes apoptosis but is reduced in human gastric cancer. *Oncotarget* 8:11105–11113 DOI 10.18632/oncotarget.14364.
- Wood PA, Yang X, Taber A, Oh EY, Ansell C, Ayers SE, Al-Assaad Z, Carnevale K, Berger FG, Peña MM, Hrushesky WJ. 2008. Period 2 mutation accelerates ApcMin/+ tumorigenesis. *Molecular Cancer Research* 6:1786–1793 DOI 10.1158/1541-7786.Mcr-08-0196.
- Wu Y, Tao B, Zhang T, Fan Y, Mao R. 2019. Pan-cancer analysis reveals disrupted circadian clock associates with T cell exhaustion. *Front Immunol* 10:2451 DOI 10.3389/fimmu.2019.02451.

- Yang X, Wood PA, Hrushesky WJ. 2010. Mammalian TIMELESS is required for ATMdependent CHK2 activation and G2/M checkpoint control. *Journal of Biological Chemistry* 285:3030–3034 DOI 10.1074/jbc.M109.050237.
- Yu CC, Chen LC, Chiou CY, Chang YJ, Lin VC, Huang CY, Lin IL, Chang TY, Lu TL, Lee CH, Huang SP, Bao BY. 2019. Genetic variants in the circadian rhythm pathway as indicators of prostate cancer progression. *Cancer Cell Int* 19:87 DOI 10.1186/s12935-019-0811-4.
- Yu M, Li W, Wang Q, Wang Y, Lu F. 2018. Circadian regulator NR1D2 regulates glioblastoma cell proliferation and motility. *Oncogene* **37**:4838–4853 DOI 10.1038/s41388-018-0319-8.
- Yuan P, Li J, Zhou F, Huang Q, Zhang J, Guo X, Lyu Z, Zhang H, Xing J. 2017. NPAS2 promotes cell survival of hepatocellular carcinoma by transactivating CDC25A. *Cell Death & Disease* 8:e2704 DOI 10.1038/cddis.2017.131.