

Vitamin C supplementation reduces expression of circulating miR-451a in poorly controlled type 2 diabetes mellitus

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Background: Vitamin C is an essential element required for the normal metabolic function. In this study, we investigated the effect of vitamin C supplementation on circulating miRNA (miR) expression in subjects with poorly controlled type 2 diabetes mellitus (T2DM) and correlated with clinical measures.

Methods: Pre- and post-vitamin C supplementation samples from five participants who had increased vitamin C levels, improved oxidative status and polymorphonuclear (PMN) function after receiving 1,000 mg vitamin C daily for six weeks were screened for miRNA expression using the NanoString miRNA assay. Differences in miRNA expression identified from the miRNA screen were validated by qRT-PCR.

Results: Four miRNAs showed significantly different expression post-vitamin C supplementation including the down-regulation of miR-451a (-1.72 fold change (FC), $p=0.036$), and up-regulation of miR-1253 (0.62 FC, $p=0.027$), miR-1290 (0.53 FC, $p=0.036$) and miR-644a (0.5 FC, $p=0.042$). Subsequent validation study showed only miR-451a expression was significantly different with supplementation. The miR-451a expression was negatively correlated with vitamin C levels ($r=-0.497$, $p=0.049$) but positively correlated with levels of malondialdehyde (MDA) ($r=0.584$, $p=0.017$), cholesterol ($r=0.564$, $p=0.022$) and low-density lipoproteins (LDL) ($r=0.522$, $p=0.037$). Bioinformatics analysis of the putative miR-451a target genes indicated gene functions related to signalling pathways involved in cellular processes such as the mammalian target of rapamycin (mTOR) signalling pathway.

Conclusions: Supplementation of vitamin C altered circulating miR-451a expression. This miRNA could be used as a biomarker to indicate the oxidative status in T2DM subjects with poor glycemic control and lead to a novel molecular strategy to reduce oxidative stress in T2DM.

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26

27 Abstract

28 **Background:** Vitamin C is an essential element required for the normal metabolic function. In
29 this study, we investigated the effect of vitamin C supplementation on circulating miRNA (miR)
30 expression in subjects with poorly controlled type 2 diabetes mellitus (T2DM) and correlated
31 with clinical measures.

32 **Methods:** Pre- and post-vitamin C supplementation samples from five participants who had
33 increased vitamin C levels, improved oxidative status and polymorphonuclear (PMN) function
34 after receiving 1,000 mg vitamin C daily for six weeks were screened for miRNA expression
35 using the NanoString miRNA assay. Differences in miRNA expression identified from the
36 miRNA screen were validated by qRT-PCR.

37 **Results:** Four miRNAs showed significantly different expression post-vitamin C
38 supplementation including the down-regulation of miR-451a (-1.72 fold change (FC), p=0.036),
39 and up-regulation of miR-1253 (0.62 FC, p=0.027), miR-1290 (0.53 FC, p=0.036) and miR-644a

40 (0.5 FC, $p=0.042$). Subsequent validation study showed only miR-451a expression was
41 significantly different with supplementation. The miR-451a expression was negatively correlated
42 with vitamin C levels ($r=-0.497$, $p=0.049$) but positively correlated with levels of
43 malondialdehyde (MDA) ($r=0.584$, $p=0.017$), cholesterol ($r=0.564$, $p=0.022$) and low-density
44 lipoproteins (LDL) ($r=0.522$, $p=0.037$). Bioinformatics analysis of the putative miR-451a target
45 genes indicated gene functions related to signalling pathways involved in cellular processes such
46 as the mammalian target of rapamycin (mTOR) signalling pathway.

47 **Conclusions:** Supplementation of vitamin C altered circulating miR-451a expression. This
48 miRNA could be used as a biomarker to indicate the oxidative status in T2DM subjects with
49 poor glycemic control and lead to a novel molecular strategy to reduce oxidative stress in T2DM.

50

51 Introduction

52 Vitamin C, or ascorbic acid, is well known as an antioxidant required for the normal metabolic
53 function of the body and associated with a wide spectrum of biological processes (*Mandl et al.*
54 *2009*). It can diminish the effect of oxidative stress and protect cells or tissue injury from
55 oxidative damages. Several studies reported a beneficial effect of vitamin C supplementation
56 especially in type 2 diabetes mellitus (T2DM), chronic metabolic disease in conjunction with
57 chronic hyperglycemia and excess free fatty acids (FFAs) (*Hawkins et al. 2003*). Overproduction
58 of reactive oxygen species from the disease can cause DNA and protein damage, lipid
59 peroxidation, cellular and vascular dysfunction leading to diabetic complications, which have
60 been associated with high levels of oxidative stress as indicated by MDA and F₂-Isoprostanes
61 (F₂IsoPs) and low levels of vitamin C (*Bhatia et al. 2003; Johansen et al. 2005; Kaviarasan et*
62 *al. 2009*). There is evidence that supplementation with 1000 mg/day of vitamin C for six weeks
63 can improve blood glucose, lipid profiles, Glycated Haemoglobin (HbA1C) and insulin in T2DM
64 (*Afkhami-Ardekani & Shojaoddiny-Ardekani 2007*). Furthermore, previous studies have shown
65 the effect of vitamin C on gene expression through epigenetic regulation including microRNA
66 (miRNA or miR) (*Kolhe et al. 2018*). A study from Kim et al revealed dietary consumption of a
67 high dose of vitamin C enhances anti-oxidation and anti-glycation as well as reduction of
68 inflammatory miRNA in lipoproteins (*Kim et al. 2015*). Treatment of human bone marrow stem
69 cells with vitamin C can modulate miRNA expression and might have an impact on the biology
70 of the cells (*Kolhe et al. 2018*). These studies indicate that vitamin C could affect miRNA
71 expression and may associate with the pathophysiology of T2DM.

72 MiRNAs are a functional non-coding small RNAs typically 18-22 nucleotides (nt) in length that
73 have been identified as an important regulator in fine-tuning of gene expression. These RNA
74 molecules mainly act by repressing mRNA translation at the 3' untranslated region (3'UTR) of
75 the target mRNA and can function in cell-cell communication (*Bartel 2004; Turchinovich et al.*
76 *2016*). One of the properties of these small RNAs is that they are highly stable in various sample
77 types such as serum or plasma and accordingly can be used as a biomarker or target therapeutics
78 in several diseases including T2DM, in which miRNA deregulation has been associated with
79 progression of the disease (*He et al. 2017; Jimenez-Lucena et al. 2018; Maqbool & Ul Hussain*

80 2014; Regazzi 2018). However, less is known about the effect of vitamin C supplementation on
81 miRNAs in T2DM subjects, especially for those subjects with poor glycemic control. In such
82 cases, the high oxidative stress and low levels of plasma vitamin C found in these subjects may
83 be influenced by miRNAs and vice versa.

84 To determine the effect of vitamin C supplementation on miRNA expression in T2DM, we
85 investigated circulating miRNAs after vitamin C supplementation using a miRNA screen. We
86 hypothesized that miRNAs may be affected by vitamin C supplementation and correlate with the
87 clinical characteristics involved in pathophysiological processes. We have chosen subjects who
88 have been affected by vitamin C supplementation on oxidative stress and PMN function. Such
89 associations may provide greater insights into the molecular mechanisms involved in these
90 biological processes and may be used as biomarkers for responses to the dietary treatment of the
91 disease leading to novel strategies in molecular targeting of relevant genes in T2DM.

92

93 **Materials & Methods**

94

95 **Study design and participants**

96 Archived plasma samples were a subset of the participants who have received vitamin C from
97 the project of vitamin C supplementation in T2DM patients (*Chuangchot et al. 2020*). The study
98 of leftover samples was approved by the Human Ethical Committee of Khon Kaen University in
99 accordance with the 1964 Declaration of Helsinki (HE571264). Diagnostic measures of T2DM
100 and clinical data of the subjects were received without personal identification from Srinagarind
101 Hospital, Faculty of Medicine, Khon Kaen, Thailand. The criteria for the screening process
102 included age between 30 and 60 years old, diagnosed as having T2DM for at least 12 months,
103 poor glycemic control (HbA1c $\geq 8.5\%$ or 69.4 mmol/mol), treated with oral diabetic drugs only
104 (no insulin injection), sedentary and lack of any regular exercise program for at least 6 months as
105 well as living in Khon Kaen Province, Thailand. Some subjects were treated with lipid-lowering
106 drugs and/or anti-hypertensive drugs for the duration of the study to maintain blood pressure at \leq
107 140/90 mmHg. The participants received 1,000 mg/day vitamin C for six weeks. Blood samples
108 from the subjects who had increased plasma vitamin C levels, increased phagocytosis and
109 oxidative burst, and reduced oxidative stress at the post-vitamin C supplementation timepoint
110 compared to pre-supplementation were collected to investigate miRNA expression.

111

112 **miRNA extraction**

113 Total RNA was extracted from archived plasma using the miRNeasy Serum/Plasma kit (Qiagen,
114 Germany) according to the manufacturer's instruction. In brief, 200 μ L of the plasma was mixed
115 with five volumes of Qiazol and a fold of chloroform. Spike-in controls, cel-miR-39 and cel-
116 miR-254, were added as a normalizer and indicator of extraction efficiency. After absorption, 14
117 μ L of RNase-free water was added to the spin column to elute total RNA and then miRNA
118 concentration was measured by a Nanophotometer.

119

120 miRNA analysis

121 To explore miRNA profiling in response to vitamin C supplementation, a miRNA microarray
122 screen was performed to discover mature-miRNA expression in plasma samples of five T2DM
123 subjects individually by using the NanoString platform (NanoString Technologies, Seattle, WA,
124 USA). This screen includes 800 human mature-miRNAs and can be profiled without an
125 amplification step. Briefly, 3 uL of total RNA was ligated to a specific miR-Tag and hybridized
126 to the color-coded probe. Counting of miRNA and digital analysis were performed using the
127 nCounter human v3 miRNA expression platform. A conservative background threshold of 100
128 counts was set to eliminate all targets with low expression for further analysis. Six positive
129 controls and cel-miR-254 were used for data normalization.

130

131 qRT-PCR

132 Qualitative real-time polymerase chain reaction (qRT-PCR) was performed to validate miRNA
133 expression for a select set of miRNAs. The small RNAs were converted to cDNA using the
134 miScript II RT kit (Qiagen, Germany) following the manufacturer's protocol. The miScript
135 primer assays and miScript SYBR green PCR kits from the Qiagen were used to measure the
136 specific miRNAs on a Bio-Rad CF96™ real time PCR machine (Bio-Rad, CA, USA) according
137 to the manufacturer protocols. The cel-miR-39 spike-in control was used as a reference gene. Ct
138 values >35 were deemed background. Relative expression of individual miRNAs was calculated
139 by the $2^{-\Delta Ct}$ method.

140

141 Target gene prediction and pathway analysis

142 To identify target genes of relevant miRNAs, four bioinformatics tools were selected to retrieve
143 the predicted target genes namely Tarbase v. 8.0, miRTarBase v. 7.0, TargetScan v. 7.2 and
144 miRDB v. 6.0. The biological functions and enrichment pathways of these target genes were
145 analyzed using the Database for Annotation, Visualization and Integration Discovery (DAVID)
146 v. 6.8. A false discovery rate (FDR) with an adjusted P-value <0.05 was set as the significance
147 threshold.

148

149 Statistical analysis

150 Normal distribution was tested using the Shapiro-Wilk test. Log2 transformed data by nSolver
151 Analysis Software v. 4.0 (NanoString Technologies) was applied to analyze the expression of
152 each miRNA using the visual genomics analysis studio (VGAS) program (an in-house program
153 for genomics analysis and visualization; <http://www.iid.com.au/software/vgas>). Comparison of
154 the pre- and post-vitamin C supplementation data was performed using a paired t-test.
155 Laboratory measures and values from the validation study were compared between pre-and post-
156 supplementation by paired t-test and Wilcoxon signed ranks test using SPSS statistics v. 19
157 (SPSS Inc., Chicago, IL, USA). A power of test ($1-\beta$) was checked by post-hoc analysis using
158 G*Power v. 3.1.9.2 (Heinrich-Heine-Universität Düsseldorf, Germany). Spearman correlation
159 was used to analyze the correlation between circulating miRNA expression and blood parameters

160 using GraphPad Prism v. 5.0 (GraphPad Software Inc., CA, USA). A p-value of <0.05 was set as
161 the significance threshold. All data expressed as mean \pm SD.

162

163 **Results**

164

165 **Characteristics of T2DM subjects at pre- and post-vitamin C supplementation**

166 Subjects had significantly increased levels of plasma vitamin C, raised PMN phagocytosis and
167 oxidative burst, reduced products of lipid peroxidase (MDA and F_2 IsoPs), as well as reduced
168 levels of cholesterol after vitamin C supplementation for six weeks (Table 1).

169

170 **MiRNA screen revealed differences in the expression level for a subset of miRNAs at pre- 171 and post-vitamin C supplementation**

172 A screen of circulating miRNAs in the plasma of five T2DM subjects at pre- and post-
173 supplementation was performed using the NanoString platform. Of the 800 target miRNA in the
174 screen, 26 miRNAs were expressed above background in all samples (Supplementary Fig. S1
175 and Table S1). Compared to the pre-supplementation, four miRNAs showed significantly
176 different expression at post-supplementation. Of these, miR-451a was down-regulated post-
177 supplementation (-1.72 fold change (FC), $p=0.036$), while miR-1253 (0.62 FC, $p=0.027$), miR-
178 1290 (0.53 FC, $p=0.036$) and miR-644a (0.5 FC, $p=0.042$) were up-regulated post-
179 supplementation (Fig. 1A). MiR-451a showed the highest reduced fold change of any of the
180 other miRNAs tested. To validate these differences, samples from three additional subjects were
181 included in the qPCR analysis (total $n=8$). Only miR-451a expression was found to be
182 significantly different post-supplementation compared to the pre-supplementation timepoint
183 ($p=0.030$) (Fig. 1B). The qPCR results for other targets from the screen were not examined
184 further. The Ct values of miR-644a were greater than 35 ($>$ background threshold; data not
185 shown). For miR-1253 and miR-1290, there was no significant difference between pre- and post-
186 supplementation ($p=0.812$ and $p=0.218$, respectively; data not shown).

187

188 **Correlation between circulating miRNA expression and clinical laboratory measures of 189 T2DM**

190 From the initial results, the expression of miR-451a was further correlated with clinical
191 laboratory measures for the T2DM subjects and included both the pre-and post-vitamin C
192 supplementation data to increase power of the statistic. Expression of miR-451a was negatively
193 correlated with the plasma vitamin C levels ($r=-0.497$, $p=0.049$, Fig. 2A) but positively
194 correlated with the MDA ($r=0.584$, $p=0.017$, Fig. 2B), cholesterol ($r=0.564$, $p=0.022$, Fig. 2C)
195 and LDL ($r=0.522$, $p=0.037$, Fig. 2D) levels, suggesting that this miRNA might play a role in
196 oxidative status and lipid metabolism.

197

198 **Target gene prediction and pathway enrichment analysis**

199 To identify target genes and pathway enrichments likely to be affected by miR-451a,
200 bioinformatics analysis was performed using both experimentally validated and prediction tools.
201 A total of 138 target genes of miR-451a were retrieved from four bioinformatics tools to identify
202 relevant KEGG pathways using DAVID. The data indicated the most significant target gene
203 enrichments for miR-451a were involved in the mTOR signalling pathway as well as other
204 signalling pathways that overlap with the target genes such as AKT1 and MAPK1 as shown in
205 Table 2 (Supplementary Table S2).

206

207 Discussion

208 Type 2 diabetes mellitus (T2DM) is a worldwide health problem and continues to increase in
209 prevalence (*Cho et al. 2018*). Long-standing T2DM can result in diabetic complications and
210 increased susceptibility to infections resulting in increased morbidity and mortality, especially in
211 uncontrolled diabetes (*Critchley et al. 2018*). To our knowledge, the present study is the first
212 report to investigate the effect of vitamin C supplementation on miRNA expression in T2DM
213 subjects with poor glycemic control. We hypothesized that supplementation of vitamin C could
214 modulate miRNA expression and associate with the blood parameters of the subjects for which
215 we observed i.e. improved plasma vitamin C levels, reduced oxidative stress and increased PMN
216 function following receiving 1,000 mg of vitamin C daily for six weeks. The result from the
217 initial miRNA screen revealed that miR-451a was downregulated, with the highest negative fold
218 change, after the supplementation and this was validated using qRT-PCR. However, the three
219 significantly upregulated miRNAs from the microarray screen did not show differences in
220 expression in the validation step with additional samples using qPCR. The discrepancy between
221 the two platforms likely reflects the low template of these miRNAs and as such we focused only
222 on miR-451a for further analysis.

223 MiR-451a (or miR-451, miRBase 22) is dicer independent and can be transcribed into a hairpin
224 structure and then alternatively processed with the Argonaute 2 (Ago 2) protein (*Wang et al.*
225 *2019*). This miRNA is stable in peripheral blood and can be found in erythrocytes, PMN,
226 mononuclear cells and platelets (*Ghai et al. 2019; Masaki et al. 2007*). MiR-451a has been
227 reported in the blood circulation of the elderly T2DM subjects and is up-regulated in plasma
228 samples of subjects with diabetic nephropathy as well as in the serum of subjects with acute
229 diabetic Charcot foot, indicating the expressed miRNA may play a role in pathological processes
230 of the disease (*Catanzaro et al. 2018; Pasquier et al. 2018; Sayilar et al. 2016*). Furthermore, the
231 expression of miR-451a was negatively correlated with the vitamin C levels but positively
232 correlated with the MDA levels suggesting its role in oxidative status in the subjects. MiR-451a
233 is also important in the erythroid lineage and plays a role in ROS production in erythrocytes by
234 targeting 14-3-3zeta and via the inhibition of FoxO3 (*Yu et al. 2010*). High oxidative stress may
235 affect miR-451a production as a study from Ranjan et al found that deficient miR-451a
236 expression was associated with defective ROS generation due to reduced Ago2 protein levels in
237 macrophages (*Ranjan et al. 2015*). Inhibition of miR-451a also reduced ROS, lipid peroxidation
238 and DNA damage (*Zhu et al. 2018*). These data suggest that miR-451a expression may positively

239 correlate with ROS production. In addition, miR-451a expression showed a positive correlation
240 with cholesterol and LDL. These factors may exacerbate the progression of the disease
241 accompanied with miR-451a expression. However, further work is needed to confirm the
242 correlation and mechanism of miR-451a in oxidative stress found in the current study.
243 We also investigated the target genes of miR-451a and their functional enrichment using both
244 experimentally validated and predicted programs. This was done to include more candidate
245 target genes with a high cut-off set to reduce false positives (*Assmann et al. 2018*). The target
246 genes were significantly involved in signalling pathways that play an important role in cellular
247 functions of biological processes as shown in Fig. 3 and Table 2. To support our results, studies
248 in T2DM cardiomyopathy-induced mice with high ROS production displayed up-regulation of
249 miR-451a that directly targeted calcium-binding protein 39 (CAB39) and resulted in down-
250 regulation of the LKB1/AMPK signaling pathway. The target genes were listed in Table 2.
251 Furthermore, knockout of this miRNA showed the opposite effect (*Kuwabara et al. 2015*). Thus,
252 the miR-451a expression was induced by high oxidative stress leading to suppression of the
253 AMPK pathway, a central energy-sensing of metabolic regulation, which was found to have
254 reduced activity in insulin-resistant individuals (*Li et al. 2019; Xu et al. 2012*). Reduced ROS
255 production found in the T2DM subjects in this study after vitamin C supplementation and down-
256 regulation of miR-451a might be involved through the CAB39/LKB1/AMPK signaling, as
257 PI3K/AKT/mTOR signaling is the central pathway of glucose and lipid metabolism (*Huang et*
258 *al. 2018; Tuo & Xiang 2018*). Hyperglycemia and excess FFAs lead to insulin resistance and
259 high levels of ROS resulting in impaired PI3K/AKT/mTOR signaling found in T2DM subjects
260 (*Huang et al. 2018*). Moreover, reduction of oxidative stress has been reported in the up-
261 regulation of AKT and mTOR after pretreatment with vitamin C (*Lin et al. 2016*). MiR-451a
262 could target multiple genes involved in the PI3K/AKT/mTOR signalling pathway including
263 AKT1, PIK3CA, PIK3R1, and MAPK1 (as indicated in Table 2). Supplementation with vitamin
264 C in the subjects might help to promote these pathways through the regulation of miR-451a.
265 Further studies are required to explore the mechanisms behind the effects of vitamin C
266 supplementation on this circulating miRNA associated with systemic oxidative stress in the
267 subjects.

268 The participants in this study showed enhanced phagocytosis and oxidative burst after
269 supplementation of vitamin C, which is consistent with the previous study that showed the effect
270 of vitamin C in enhancing PMN function such as motility, chemotaxis, phagocytosis and
271 microbial killing (*Bozonet et al. 2015; Carr & Maggini 2017*). However, knowledge of the
272 molecular mechanisms of the effect of vitamin C on these processes, especially phagocytosis and
273 oxidative burst, are limited. MiR-451a has been reported to impact neutrophil chemotaxis by
274 suppressing p38 MAPK through targeting Rab5a and 14-3-3zeta (*Murata et al. 2014*). Rab5a, a
275 small GTPase, regulates intracellular membrane trafficking involved in phagolysosome fusion
276 during bacteria phagocytosis of neutrophils (*Perskvist et al. 2002*). Moreover, treatment of
277 human bladder cancer cells by vitamin C could suppress p38 MAPK activity and ROS
278 production (*Kim et al. 2008*). As suggested from these data, vitamin C may affect miR-451a

279 expression and associate with neutrophil function through targeting Rab5a and the p38 MAPK
280 pathway. However, investigation of intracellular miRNAs in responding to vitamin C
281 supplementation may help us to better understand the molecular mechanism underlying PMN
282 function that we clearly observed in these subjects. A proposed mechanism of actions for miR-
283 451a in response to vitamin C supplementation was shown in Fig. 3.

284 Limitations of the current study include a small sample size. Increasing the number of the
285 participants may better clarify the association of the candidate miRNA and the laboratory
286 measures including improved oxidative status and PMN functions as well as reduced cholesterol
287 after vitamin C supplementation. Nevertheless, the power of test based on difference between the
288 two dependent groups using mean and S.D. of relative miR-451a data revealed that the power
289 was approximated 80%. More research is also needed to confirm and validate the finding that
290 vitamin C intake modulates circulating miR-451a expression and to determine the functional role
291 of this miRNA in subjects with uncontrolled T2DM.

292

293 **Conclusions**

294 In conclusion, this study has identified a novel association between vitamin C supplementation
295 for six weeks and reduced circulating miR-451a expression in poorly controlled T2DM. This
296 candidate miRNA might be used as a biomarker to identify subjects that respond to vitamin C
297 treatment or oxidative status in plasma.

298

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303 **ADDITIONAL INFORMATION AND DECLARATIONS**

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310 University.

311

312 **Competing Interests**

313 The authors declare no competing interests.

314

315 **Author contributions**

316 Laongthip Ruknarong performed experiments, analysed the data, prepared figures and tables,
317 prepared the first draft manuscript and the final draft.

318 Chongchira Boonthongkaew and Nisa Chuangchot enrolled participants, performed experiments
319 and analysed the data
320 Amonrat Jumnainsong, Naruemon Leelayuwat and Apinya Jusakul reviewed and approved the
321 final draft.
322 Silvana Gaudieri and Chanvit Leelayuwat analysed, criticised the data, presentations of the
323 manuscript, reviewed and approved the final draft.
324 Chanvit Leelayuwat design of the study, oversaw and supervised the whole project.
325

326 References

- 327 **Afkhami-Ardekani M, and Shojaoddiny-Ardekani A. 2007.** Effect of vitamin C on blood
328 glucose, serum lipids & serum insulin in type 2 diabetes patients. *Indian J Med Res*
329 **126**:471-474.
- 330 **Assmann TS, Recamonde-Mendoza M, Punales M, Tschiedel B, Canani LH, and Crispim**
331 **D. 2018.** MicroRNA expression profile in plasma from type 1 diabetic patients: Case-
332 control study and bioinformatic analysis. *Diabetes Res Clin Pract* **141**:35-46.
333 10.1016/j.diabres.2018.03.044
- 334 **Bartel DP. 2004.** MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* **116**:281-
335 297. 10.1016/s0092-8674(04)00045-5
- 336 **Bhatia S, Shukla R, Venkata Madhu S, Kaur Gambhir J, and Madhava Prabhu K. 2003.**
337 Antioxidant status, lipid peroxidation and nitric oxide end products in patients of type 2
338 diabetes mellitus with nephropathy. *Clin Biochem* **36**:557-562.
- 339 **Bozonet SM, Carr AC, Pullar JM, and Vissers MC. 2015.** Enhanced human neutrophil
340 vitamin C status, chemotaxis and oxidant generation following dietary supplementation
341 with vitamin C-rich SunGold kiwifruit. *Nutrients* **7**:2574-2588. 10.3390/nu7042574
- 342 **Carr AC, and Maggini S. 2017.** Vitamin C and Immune Function. *Nutrients* **9(11)**:1211.
343 10.3390/nu9111211
- 344 **Catanzaro G, Besharat ZM, Chiacchiarini M, Abballe L, Sabato C, Vacca A, Borgiani P,**
345 **Dotta F, Tesauro M, Po A, and Ferretti E. 2018.** Circulating MicroRNAs in Elderly
346 Type 2 Diabetic Patients. *Int J Endocrinol* **2018**:6872635. 10.1155/2018/6872635
- 347 **Cho NH, Shaw JE, Karuranga S, Huang Y, da Rocha Fernandes JD, Ohlrogge AW, and**
348 **Malanda B. 2018.** IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017
349 and projections for 2045. *Diabetes Res Clin Pract* **138**:271-281.
350 10.1016/j.diabres.2018.02.023
- 351 **Chuangchot N, Boonthongkaew C, Phoksawat W, Jumnainsong A, Leelayuwat C, and**
352 **Leelayuwat N. 2020.** Oral vitamin C treatment increases polymorphonuclear cell
353 functions in type 2 diabetes mellitus patients with poor glycemic control. *Nutr Res* **79**:50-
354 59. 10.1016/j.nutres.2020.05.010
- 355 **Critchley JA, Carey IM, Harris T, DeWilde S, Hosking FJ, and Cook DG. 2018.** Glycemic
356 Control and Risk of Infections Among People With Type 1 or Type 2 Diabetes in a Large
357 Primary Care Cohort Study. *Diabetes Care* **41**:2127-2135. 10.2337/dc18-0287
- 358 **Ghai V, Kim TK, Etheridge A, Nielsen T, Hansen T, Pedersen O, Galas D, and Wang K.**
359 **2019.** Extracellular Vesicle Encapsulated MicroRNAs in Patients with Type 2 Diabetes
360 Are Affected by Metformin Treatment. *J Clin Med* **8(5)**:617. 10.3390/jcm8050617

- 361 **Hawkins M, Tonelli J, Kishore P, Stein D, Ragucci E, Gitig A, and Reddy K. 2003.**
362 Contribution of elevated free fatty acid levels to the lack of glucose effectiveness in type
363 2 diabetes. *Diabetes* **52**:2748-2758. 10.2337/diabetes.52.11.2748
- 364 **He Y, Ding Y, Liang B, Lin J, Kim TK, Yu H, Hang H, and Wang K. 2017.** A Systematic
365 Study of Dysregulated MicroRNA in Type 2 Diabetes Mellitus. *Int J Mol Sci* **18(3)**: 456.
366 10.3390/ijms18030456
- 367 **Huang X, Liu G, Guo J, and Su Z. 2018.** The PI3K/AKT pathway in obesity and type 2
368 diabetes. *Int J Biol Sci* **14**:1483-1496. 10.7150/ijbs.27173
- 369 **Jimenez-Lucena R, Rangel-Zuniga OA, Alcala-Diaz JF, Lopez-Moreno J, Roncero-Ramos**
370 **I, Molina-Abril H, Yubero-Serrano EM, Caballero-Villarraso J, Delgado-Lista J,**
371 **Castano JP, Ordovas JM, Perez-Martinez P, Camargo A, and Lopez-Miranda J.**
372 **2018.** Circulating miRNAs as Predictive Biomarkers of Type 2 Diabetes Mellitus
373 Development in Coronary Heart Disease Patients from the CORDIOPREV Study. *Mol*
374 *Ther Nucleic Acids* **12**:146-157. 10.1016/j.omtn.2018.05.002
- 375 **Johansen JS, Harris AK, Rychly DJ, and Ergul A. 2005.** Oxidative stress and the use of
376 antioxidants in diabetes: linking basic science to clinical practice. *Cardiovasc Diabetol*
377 **4**:5. 10.1186/1475-2840-4-5
- 378 **Kaviarasan S, Muniandy S, Qvist R, and Ismail IS. 2009.** F(2)-isoprostanes as novel
379 biomarkers for type 2 diabetes: a review. *J Clin Biochem Nutr* **45**:1-8. 10.3164/jcbtn.08-
380 266
- 381 **Kim JE, Jin DH, Lee SD, Hong SW, Shin JS, Lee SK, Jung DJ, Kang JS, and Lee WJ.**
382 **2008.** Vitamin C inhibits p53-induced replicative senescence through suppression of ROS
383 production and p38 MAPK activity. *Int J Mol Med* **22**:651-655.
- 384 **Kim SM, Lim SM, Yoo JA, Woo MJ, and Cho KH. 2015.** Consumption of high-dose vitamin
385 C (1250 mg per day) enhances functional and structural properties of serum lipoprotein to
386 improve anti-oxidant, anti-atherosclerotic, and anti-aging effects via regulation of anti-
387 inflammatory microRNA. *Food Funct* **6**:3604-3612. 10.1039/c5fo00738k
- 388 **Kolhe R, Mondal AK, Pundkar C, Periyasamy-Thandavan S, Mendhe B, Hunter M, Isaies**
389 **CM, Hill WD, Hamrick MW, and Fulzele S. 2018.** Modulation of miRNAs by Vitamin
390 C in Human Bone Marrow Stromal Cells. *Nutrients* **10(2)**:186.. 10.3390/nu10020186
- 391 **Kuwabara Y, Horie T, Baba O, Watanabe S, Nishiga M, Usami S, Izuhara M, Nakao T,**
392 **Nishino T, Otsu K, Kita T, Kimura T, and Ono K. 2015.** MicroRNA-451 exacerbates
393 lipotoxicity in cardiac myocytes and high-fat diet-induced cardiac hypertrophy in mice
394 through suppression of the LKB1/AMPK pathway. *Circ Res* **116**:279-288.
395 10.1161/CIRCRESAHA.116.304707
- 396 **Li J, Wan W, Chen T, Tong S, Jiang X, and Liu W. 2019.** miR-451 Silencing Inhibited
397 Doxorubicin Exposure-Induced Cardiotoxicity in Mice. *Biomed Res Int* **2019**:1528278.
398 10.1155/2019/1528278
- 399 **Lin CJ, Chen TL, Tseng YY, Wu GJ, Hsieh MH, Lin YW, and Chen RM. 2016.** Honokiol
400 induces autophagic cell death in malignant glioma through reactive oxygen species-
401 mediated regulation of the p53/PI3K/Akt/mTOR signaling pathway. *Toxicol Appl*
402 *Pharmacol* **304**:59-69. 10.1016/j.taap.2016.05.018
- 403 **Mandl J, Szarka A, and Banhegyi G. 2009.** Vitamin C: update on physiology and
404 pharmacology. *Br J Pharmacol* **157**:1097-1110. 10.1111/j.1476-5381.2009.00282.x
- 405 **Maqbool R, and Ul Hussain M. 2014.** MicroRNAs and human diseases: diagnostic and
406 therapeutic potential. *Cell Tissue Res* **358**:1-15. 10.1007/s00441-013-1787-3

- 407 **Masaki S, Ohtsuka R, Abe Y, Muta K, and Umemura T. 2007.** Expression patterns of
408 microRNAs 155 and 451 during normal human erythropoiesis. *Biochem Biophys Res*
409 *Commun* **364**:509-514. 10.1016/j.bbrc.2007.10.077
- 410 **Murata K, Yoshitomi H, Furu M, Ishikawa M, Shibuya H, Ito H, and Matsuda S. 2014.**
411 MicroRNA-451 down-regulates neutrophil chemotaxis via p38 MAPK. *Arthritis*
412 *Rheumatol* **66**:549-559. 10.1002/art.38269
- 413 **Pasquier J, Ramachandran V, Abu-Qaoud MR, Thomas B, Benurwar MJ, Chidiac O,**
414 **Hoarau-Vechot J, Robay A, Fakhro K, Menzies RA, Jayyousi A, Zirie M, Al**
415 **Suwaidi J, Malik RA, Talal TK, Najafi-Shoushtari SH, Rafii A, and Abi Khalil C.**
416 **2018.** Differentially expressed circulating microRNAs in the development of acute
417 diabetic Charcot foot. *Epigenomics* **10**:1267-1278. 10.2217/epi-2018-0052
- 418 **Perskvist N, Roberg K, Kulyte A, and Stendahl O. 2002.** Rab5a GTPase regulates fusion
419 between pathogen-containing phagosomes and cytoplasmic organelles in human
420 neutrophils. *J Cell Sci* **115**:1321-1330.
- 421 **Ranjan R, Lee YG, Karpurapu M, Syed MA, Chung S, Deng J, Jeong JJ, Zhao G, Xiao L,**
422 **Sadikot RT, Weiss MJ, Christman JW, and Park GY. 2015.** p47phox and reactive
423 oxygen species production modulate expression of microRNA-451 in macrophages. *Free*
424 *Radic Res* **49**:25-34. 10.3109/10715762.2014.974037
- 425 **Regazzi R. 2018.** MicroRNAs as therapeutic targets for the treatment of diabetes mellitus and its
426 complications. *Expert Opin Ther Targets* **22**:153-160. 10.1080/14728222.2018.1420168
- 427 **Sayilar EI, Gullulu M, Tuncel E, Peynirci H, Alemdar A, Tunca B, Egeli U, Cecener G,**
428 **Bayindir M, and Cosgun G. 2016.** Biomarker Potential of Urine miR-451 at Different
429 Stages of Diabetic Nephropathy. *Journal of Diabetes & Metabolism* **7**:2. Unsp 1000650
430 10.4172/2155-6156.1000650
- 431 **Tuo Y, and Xiang M. 2018.** mTOR: A double-edged sword for diabetes. *J Leukoc Biol*
432 **106**(2):385-395. 10.1002/JLB.3MR0317-095RR
- 433 **Turchinovich A, Tonevitsky AG, and Burwinkel B. 2016.** Extracellular miRNA: A Collision
434 of Two Paradigms. *Trends Biochem Sci* **41**:883-892. 10.1016/j.tibs.2016.08.004
- 435 **Wang T, Wu F, and Yu D. 2019.** miR-144/451 in hematopoiesis and beyond. *ExRNA* **1**:16.
436 10.1186/s41544-019-0035-8
- 437 **Xu XJ, Gauthier MS, Hess DT, Apovian CM, Cacicedo JM, Gokce N, Farb M, Valentine**
438 **RJ, and Ruderman NB. 2012.** Insulin sensitive and resistant obesity in humans: AMPK
439 activity, oxidative stress, and depot-specific changes in gene expression in adipose tissue.
440 *J Lipid Res* **53**:792-801. 10.1194/jlr.P022905
- 441 **Yu D, dos Santos CO, Zhao G, Jiang J, Amigo JD, Khandros E, Dore LC, Yao Y, D'Souza**
442 **J, Zhang Z, Ghaffari S, Choi J, Friend S, Tong W, Orange JS, Paw BH, and Weiss**
443 **MJ. 2010.** miR-451 protects against erythroid oxidant stress by repressing 14-3-3zeta.
444 *Genes Dev* **24**:1620-1633. 10.1101/gad.1942110
- 445 **Zhu H, Zhang L, Xu J, Zhu C, Zhao H, Zhu Y, and Lv G. 2018.** AntogomiR-451 protects
446 human gastric epithelial cells from ethanol via activating AMPK signaling. *Biochem*
447 *Biophys Res Commun* **497**:339-346. 10.1016/j.bbrc.2018.02.082
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449 **Figure legends**

450 **Figure 1 Patterns of miRNA expression.** (A) Analysis of circulating miRNA expression
451 combined with clinical measures and qPCR data in response to vitamin C supplementation in
452 T2DM patients with poor glycemic control (n=5). For the miRNA, blue and red bubbles

453 represent pre-and post-supplementation, respectively. The size of the bubbles reflects the count
454 of the specific miRNA relative to other miRNAs. The laboratory measures and miR-451a qRT-
455 PCR data are indicated as a heat map from minimum value to maximum value of each parameter
456 independently of other parameters; green to red represents the low to high range. Pre; pre-
457 vitamin C, Post; post-vitamin C supplementation. Numbers represent subjects 1-5. (B) Plot
458 represents relative expression of miR-451a as validated by qRT-PCR (n=8).

459

460 **Figure 2 Correlation between miR-451a expression and laboratory variables.** (A) Vitamin C
461 levels, (B) MDA, (C) Cholesterol and (D) LDL. Blue and red represent pre-and post-
462 supplementation samples, respectively.

463

464 **Figure 3 Proposed mechanism of the effect of miR-451a in response to vitamin C**
465 **supplementation.** Red lines represent the results of this study and blue lines indicate potential
466 mode of interaction for miR-451a. Poor glycemic controlled T2DM subjects receiving vitamin C
467 1,000 mg dairy for six weeks showed significant down-regulation of circulating miR-451a
468 accompanied with increased levels of vitamin C, reduced oxidative stress (MDA and F₂IsoPs)
469 and increased PMN function (phagocytosis and oxidative burst). Changes in oxidative status may
470 modulate miR-451a expression by altering Ago2 protein and/or increased involvement of
471 signalling pathways such as AMPK signalling by repressing its target genes. MiR-451a may play
472 a role in neutrophil chemotaxis by targeting RAB5A and 14-3-3zeta resulting in the activation of
473 p38 MAPK signalling. However, the role of this miRNA in phagocytosis and oxidative burst of
474 neutrophils is still to be explored (dotted line) and may be reflected in intracellular miRNA
475 changes rather than circulating changes as identified in this study.

Table 1 (on next page)

Table 1 Characteristics of the participants at the pre-and post-vitamin C supplementation timepoints.

Age, gender, BMI and blood chemistry data of participants pre- and post- vitamin C supplementation.

1 **Table 1:**
 2 **Characteristics of the participants at the pre-and post-vitamin C supplementation**
 3 **timepoints.**
 4

Parameters	Pre-supplementation (n=8)	Post-supplementation (n=8)	P-value
Gender (female/male)	7/1	7/1	-
Age (yr)	58.8±5.9	58.8±5.9	-
BMI (Kg/m ²)	26.2±4.1	26.2±3.9	0.861
HbA1c (mmol/mol)	95.0±15.0	95.0±14.8	0.909
FBS (mmol/L)	13.0±4.3	10.0±2.0	0.059
Plasma vitamin C levels (μmol/L)	57.8±11.0	90.5±55.5	0.017
Plasma MDA levels (μmol/mL)	17.0±8.6	10.6±3.8	0.031
Plasma F ₂ IsoPs (pg/mL)	16.9±4.8	12.0±4.3	0.012
Phagocytosis (%)	20.4±8.6	30.7±11.4	0.012
Oxidative burst (%)	5.9±3.5	11.1±4.5	0.006
Cholesterol (mg/dL)	239.3±60.1	187.9±45.9	0.050
Triglyceride (mg/dL)	168.8±63.8	195.9±100.1	0.217
HDL (mg/dL)	44.9±10.6	45.9±15.0	0.735
LDL (mg/dL)	164.9±53.7	126.5±32.3	0.154
Insulin (IU/mL)	14.1±4.6	11.5±3.6	0.057

5 Data represented as mean ± S.D. P-value was calculated using paired t-test.

6

Table 2 (on next page)

Table 2 KEGG signaling pathways of miR-451a.

KEGG signaling pathways, Target gene, p-value and FDR prediction of miR-451a.

1 **Table 2:**
 2 **KEGG signaling pathways of miR-451a.**
 3

Term	Gene targets	P-value	FDR
mTOR signaling pathway	AKT1, MAPK1, TSC1, STK11, PIK3CA, PRKAA1, CAB39, IKBKB, PIK3R1	1.29E-08	0.00002
Hepatitis B	AKT1, MAPK1, YWHAZ, IL6, MAP3K1, BCL2, MMP9, PIK3CA, IKBKB, MYC, PIK3R1, ATF2	1.75E-08	0.00002
PI3K-Akt signaling pathway	AKT1, MAPK1, IL6, YWHAZ, TSC1, STK11, BCL2, PIK3CA, PRKAA1, LPAR1, IL6R, IKBKB, MYC, PIK3R1, ATF2	2.5E-07	0.00031
FoxO signaling pathway	AKT1, MAPK1, IL6, CDKN2B, STK11, CDKN2D, PIK3CA, PRKAA1, IKBKB, PIK3R1	6.56E-07	0.00080
Estrogen signaling pathway	AKT1, MAPK1, GNAQ, MMP9, PIK3CA, MMP2, PIK3R1, ATF2	8.51E-05	0.01033
TNF signaling pathway	AKT1, MAPK1, IL6, MMP9, PIK3CA, IKBKB, PIK3R1, ATF2	1.87E-05	0.02266
Non-alcoholic fatty liver disease (NAFLD)	AKT1, IL6, PIK3CA, PRKAA1, IL6R, IKBKB, UQCRQ, EIF2AK3, PIK3R1	2.95E-05	0.03578
AMPK signaling pathway	AKT1, TSC1, STK11, RAB14, PIK3CA, PRKAA1, CAB39, PIK3R1	3.94E-05	0.04776

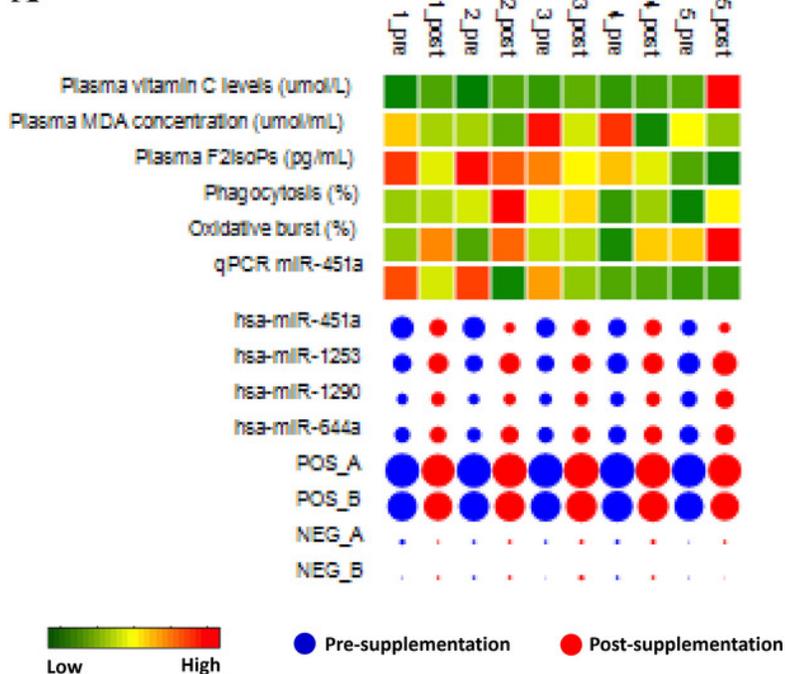
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Figure 1

Figure 1 Patterns of miRNA expression.

(A) Analysis of circulating miRNA expression combined with clinical measures and qPCR data in response to vitamin C supplementation in T2DM patients with poor glycemic control (n=5). For the miRNA, blue and red bubbles represent pre- and post-supplementation, respectively. The size of the bubbles reflects the count of the specific miRNA relative to other miRNAs. The laboratory measures and miR-451a qRT-PCR data are indicated as a heat map from minimum value to maximum value of each parameter independently of other parameters; green to red represents the low to high range. Pre; pre-vitamin C, Post; post-vitamin C supplementation. Numbers represent subjects 1-5. (B) Plot represents relative expression of miR-451a as validated by qRT-PCR (n=8).

A



B

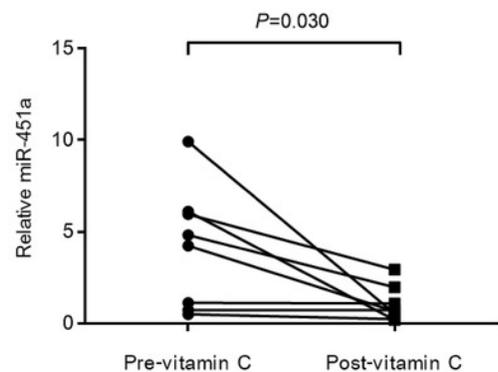


Figure 2

Figure 2 Correlation between miR-451a expression and laboratory variables.

(A) Vitamin C levels, (B) MDA, (C) Cholesterol and (D) LDL. Blue and red represent pre-and post-supplementation samples, respectively.

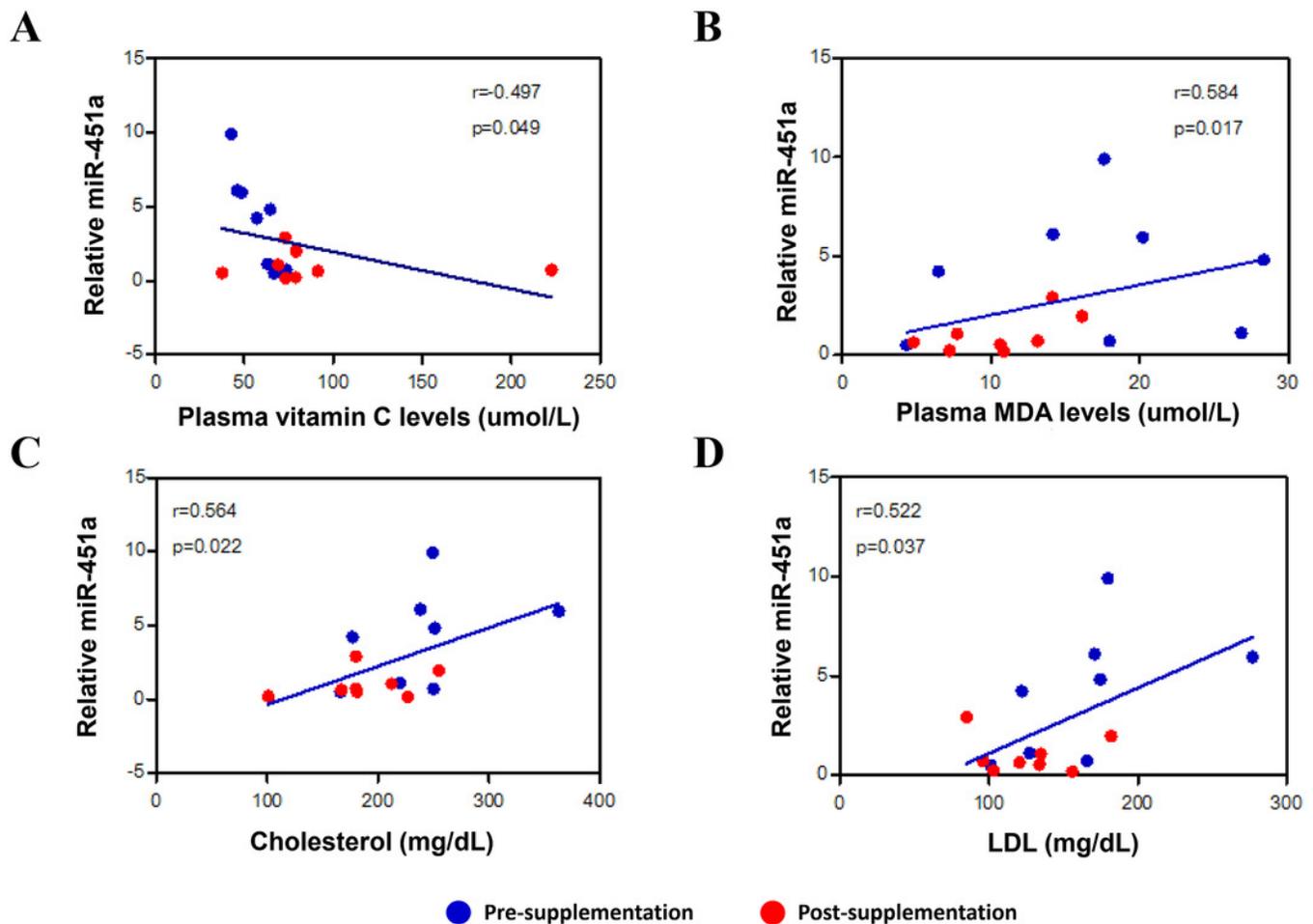


Figure 3

Figure 3 Proposed mechanism of the effect of miR-451a in response to vitamin C supplementation.

Red lines represent the results of this study and blue lines indicate potential mode of interaction for miR-451a. Poor glycemic controlled T2DM subjects receiving vitamin C 1,000 mg daily for six weeks showed significant down-regulation of circulating miR-451a accompanied with increased levels of vitamin C, reduced oxidative stress (MDA and F₂IsoPs) and increased PMN function (phagocytosis and oxidative burst). Changes in oxidative status may modulate miR-451a expression by altering Ago2 protein and/or increased involvement of signalling pathways such as AMPK signalling by repressing its target genes. MiR-451a may play a role in neutrophil chemotaxis by targeting RAB5A and 14-3-3zeta resulting in the activation of p38 MAPK signalling. However, the role of this miRNA in phagocytosis and oxidative burst of neutrophils is still to be explored (dotted line) and may be reflected in intracellular miRNA changes rather than circulating changes as identified in this study.

