A walking-trace fossil made by a crayfish or crayfish-like arthropod from the Lower Jurassic Moenave Formation of southwestern Utah

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Abstract

Trace fossils (ichnofossils) from the Lower Jurassic Moenave Formation at the St. George Dinosaur Discovery Site at Johnson Farm (SGDS) are relatively well understood and described, but new specimens, particularly of invertebrates, continue to expand the ichnofauna at the site. A previously unstudied arthropod locomotory (walking) trace, SGDS 1290, comprises two widely spaced, thick, gently undulating paramedial impressions flanked externally by small, tapered to elongate tracks with a staggered arrangement. The specimen is not a variant of any existing ichnotaxon, but bears a striking resemblance to modern, experimentally generated crayfish walking traces, suggesting a crayfish or crayfish-like maker for the fossil. Because of its uniqueness, we place it in a new ichnospecies, *Siskemia eurypyge*. It is the first fossil crayfish or crayfish-like locomotion trace ever identified.

Introduction

Ichnology, the study of fossil traces (ichnites), contributes a substantial body of paleobiological information to the understanding of extinct organisms. This is because trace fossils are direct results of ancient animal behavior (Osgood, 1975) that could only otherwise be inferred from body fossils. Furthermore, the global commonness of ichnites compared to body fossils means that the ichnological record often can preserve evidence of the presence of organisms not otherwise represented in the body fossil record (Osgood, 1975), especially of invertebrates that lack hard parts and therefore fossilize only under exceptional conditions. Except for conchostracans (*sensu* Kozur & Weems, 2010) and ostracods, which have biomineralized carapaces, arthropods are among the less commonly preserved body-fossil components of terrestrial (including freshwater) paleoecosystems except in various Konservat-Lagerstätten (fossiliferous sites of exceptional preservational quality) (e.g., Charbonnier et al., 2010; Luque et al., 2019; Selden & Nudds, 2012; Smith, 2012). Yet from the mid-Paleozoic through the Cenozoic, arthropods—especially insects and chelicerates—were certainly the most populous and diverse metazoan components of most terrestrial ecosystems (Labandeira & Beall, 1990), and their paleoecological importance cannot be underestimated.

Arthropod ichnites can be more common and abundant than arthropod body fossils, and indicate the presences of various arthropod taxa in paleoecosystems for which body fossils may be entirely absent. Locomotory tracks (repichnia) of arthropods have an extensive geological history, spanning from the Cambrian (and possibly latest Precambrian [Chen et al., 2018]) through the Holocene. They constitute some of the earliest evidence of metazoan life venturing onto land (reviewed in Minter et al., 2016a; Minter et al., 2016b) and are known from virtually every paleoenvironment, from near shore and shallow marine environments (e.g., Collette, Hagadorn & Lacelle, 2010; MacNaughton et al., 2002; Pirrie, Feldmann & Buatois, 2004; Shillito & Davies, 2018; Trewin & McNamara, 1994) and, terrestrially, from proglacial systems (e.g., Anderson, 1981; Lima, Minter & Netto, 2017; Lima et al., 2015; Uchman, Kazakauskas &

Gaigalas, 2009; Walter, 1985) to desert ergs (e.g., Gilmore, 1927; Good & Ekdale, 2014; Sadler, 1993).

The St. George Dinosaur Discovery Site at Johnson Farm (SGDS) in St. George, Washington County, Utah (Fig. 1) has been called a Konzentrat-Ichnolagerstätte (Hunt & Lucas, 2006a) because of its exceptional concentration of well-preserved ichnites from a broad spectrum of terrestrial organisms. The site preserves a detailed "snapshot" of an earliest Jurassic ecosystem from on- and offshore portions of a freshwater, lacustrine paleoenvironment. The "Dinosaur Discovery" part of the name of the site indicates the concentration of dinosaur tracks at this locality (Milner, Lockley & Johnson, 2006; Milner, Lockley & Kirkland, 2006; Milner et al., 2009), but tracks of other vertebrates (Lockley, Kirkland & Milner, 2004; Milner, Lockley & Johnson, 2006) and a moderately diverse invertebrate ichnofauna (Lucas et al., 2006a) are also preserved. Among the latter are many locomotory trackways made by arthropods, some of which have been referred to cf. *Bifurculapes*, *Diplichnites*, and cf. *Kouphichnium* (Lucas et al., 2006a). Ichnospecies of *Bifurculapes* have been variably attributed to insects, possibly beetles, and to crustaceans (Getty, 2016, 2018; Hitchcock, 1858, 1865); ichnospecies of *Diplichnites* have been attributed to myriapods (Briggs, Rolfe & Brannan, 1979; Davis, Minter & Braddy, 2007; Pollard, Selden & Watts, 2008; Shillito & Davies, 2018), notostracans (Lucas et al., 2006a; Minter et al., 2007), and other arthropods (Melchor & Cardonatto, 2014; Minter et al., 2007); and ichnospecies of *Kouphichnium* have been attributed to limulids (Caster, 1944; King, Stimson & Lucas, 2019; Lomax & Racay, 2012).

A previously unstudied SGDS specimen, SGDS 1290, is an arthropod locomotory trace that differs markedly from any other SGDS specimen, indicating the presence of a heretofore unrecognized component of the SGDS ichnofauna. SGDS 1290 is an arthropod locomotory trace because it includes distinct footprints in a discernible cycle, but lacks any features of vertebrate tracks, such as distinct toes (*sensu* Seilacher, 2007). The trace thus resembles numerous other fossil traces attributed to arthropods, as well as those generated experimentally. The fossil was discovered and collected March 11, 2010 by SGDS volunteer Jon Cross.

Geological Setting

Most of the fossils preserved at the SGDS, including the trace described below, occur in the Whitmore Point Member of the Moenave Formation (Kirkland & Milner, 2006; Kirkland et al., 2014), which conformably overlies the Dinosaur Canyon Member of the Moenave Formation and disconformably underlies the Springdale Sandstone Member, which itself has been assigned as both the lowest member of the Kayenta Formation (Lucas & Tanner, 2006) and the uppermost member of the Moenave Formation (Steiner, 2014a). The richest source of the trace fossils at the SGDS, again including the trace described below, occur within a fine-grained sandstone near the base of the Whitmore Point Member initially called the "Main Track Layer" (Kirkland & Milner, 2006; Milner, Lockley & Johnson, 2006; Milner, Lockley & Kirkland, 2006) and, later and more formally, the Johnson Farm Sandstone Bed (unit 40 of Kirkland et al., 2014) (Fig. 2). SGDS 1290 comes from the uppermost strata of this unit, specifically one of several thinly bedded,

apparently conformable, fine-grained-sandstone "Top Surface Tracksite" horizons (*sensu* Kirkland et al., 2014; Milner, Lockley & Johnson, 2006; Milner, Lockley & Kirkland, 2006). The Whitmore Point Member preserves sediments deposited in and around the large, freshwater Lake Whitmore (formerly called Lake Dixie) (Kirkland & Milner, 2006; Kirkland et al., 2014); at the SGDS itself, the Johnson Farm Sandstone Bed preserves ichnites and sedimentary structures made in both subaerial and subaqueous conditions, indicating a shoreline paleoenvironment (Milner, Lockley & Kirkland, 2006). Ichnologically, invertebrate trace fossils in this paleoenvironment pertain to the *Scoyenia* ichnofacies (Buatois & Mángano, 2004; Lucas et al., 2006a), while the associated vertebrate ichnofauna pertains to the *Eubrontes* ichnocoenosis of the *Grallator* ichnofacies (Hunt & Lucas, 2006b; Hunt & Lucas, 2006c).

Age determinations for the Whitmore Point Member have varied. The unit typically has been placed entirely within the Hettangian (earliest Jurassic) largely on biostratigraphic grounds (see discussions in Kirkland et al., 2014; Milner et al., 2012; Parrish et al., 2019; and Tanner & Lucas, 2009), but was also determined to straddle the Triassic–Jurassic boundary (201.3 ± 0.2 Ma) on magnetostratigraphic grounds (Donohoo-Hurley, Geissman & Lucas, 2010), in which system the Johnson Farm Sandstone Bed would be Rhaetian (latest Triassic) in age. However, Steiner (2014b) recovered Hettangian paleomagnetic sequences from the Whitmore Point Member, and Suarez et al. (2017) calibrated the magnetostratigraphic data of Donohoo-Hurley, Geissman & Lucas (2010) with high-precision U–Pb dates to re-situate the Triassic–Jurassic boundary stratigraphically farther down in the Dinosaur Canyon Member of the Moenave Formation, also making the Whitmore Point Member entirely earliest Jurassic in age. The Johnson Farm Sandstone Bed and its fossils therefore are Hettangian in age, approximately 200 million years old.

Materials & Methods

Measurements of SGDS 1290 were taken using digital calipers. Ichnological terminology for arthropod locomotory traces used herein follows Minter, Braddy & Davis (2007) and Genise (2017). Minter, Braddy & Davis (2007) defined "tracks" as discrete marks made by locomotory appendages, "impressions" as continuous traces made by another portion of the anatomy of a trace maker, and "imprints" as discontinuous such traces; they also provided terms for trackway arrangement and measurements. Genise (2017) outlined various descriptive terms for individual track morphologies.

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BBCCF0E37143. The LSID for the herein described *Siskemia eurypyge* isp. nov. is: urn:lsid:zoobank.org:act:769B0815-8991-4F0E-B32C-99C87A9D293B. The online version of this work is archived and available from the following digital repositories: PeerJ, PubMed Central and CLOCKSS.

Description of SGDS 1290

SGDS 1290, a natural cast (convex hyporelief), consists of two parallel, undulating, paramedial ridges flanked externally by eight closely appressed sets of small tracks that range in shape from ovoid to tapered (teardrop-shaped) to elongate (Fig. 3). The track sets are oriented perpendicular to the trackway axis, though tapered and elongate individual tracks within each set have long axes that parallel or are oblique to the trackway axis. Track sets average 4.5 mm long craniocaudally (anteroposteriorly) and 5.4 mm wide mediolaterally (Table 1). Left (L) and right (R) sets of tracks are arranged in a staggered pattern such that there is half a cycle displacement between each step. Based on Fairchild & Hasiotis (2011), the tapering ends of the tapered tracks are presumed to be cranial (anterior) reflectures, indicating the direction of movement. Most tracks have long axes oriented parallel to the trackway axis; a few (such as in sets L1, L4, and R2) are oblique to the axis. Track R1 is markedly elongate rather than tapered, but also parallel to the trackway axis. Track sets L1, L3, and R4 consist of three distinct but appressed tracks; sets L2, L4, and R2 consist of pairs of appressed tracks, and R1 and R3 appear to consist of single tracks, although the possibility that each comprises multiple, conjoined tracks cannot be ruled out.

The paramedial impressions typically are thick mediolaterally, though they vary and taper briefly to nothingness in a few places (being more continuous than repeating, we consider them "impressions" and not "imprints"). The impressions follow gently undulating (non-linear and low amplitude) pathways. They span approximately 7.5 cm along the slab of rock. Overprints of short segments of the paramedial impressions that are not accompanied by tracks are visible behind the main trace segment on a slightly higher stratum. The impressions vary in width along their lengths, ranging from 0–3.1 mm (mean = 2.0 mm) for the left impressions and 0–3.0 mm (mean = 1.6 mm) for the right (Table 1). The width of the trace from impression to impression averages 9.4 mm when measured from the lateral (external) edges and 5.6 mm when measured between the medial (internal) edges (Table 1). The distances between the impressions thus are greater than the distances between the impressions and their flanking tracks (mean = 1.6 mm). The impressions taper slightly: they are wider at their bases and narrower at their rounded apices.

Comparisons to Arthropod Repichnial Ichnotaxa

Arthropod repichnia lacking medial or paramedial impressions

Several arthropod locomotory (walking) ichnotaxa are readily distinguished from SGDS 1290 by (usually) lacking medial or paramedial impressions, but are worth comparing to SGDS 1290 to ascertain whether or not it might be a morphological variant of such ichnotaxa. These

ichnotaxa are *Acanthichnus* (Hitchcock, 1858), *Asaphoidichnus* (Miller, 1880), *Bifurculapes* (Hitchcock, 1858), *Coenobichnus* (Walker, Holland & Gardiner, 2003), *Copeza* (Hitchcock, 1858), *Danstairia congesta* (Smith, 1909), *Diplichnites* (Dawson, 1873), *Eisenachichnus* (Kozur, 1981), *Foersterichnus* (Pirrie, Feldmann & Buatois, 2004), *Hamipes* (Hitchcock, 1858), *Lithographus* (Hitchcock, 1858), *Hornburgichnium* (Kozur, 1989), *Maculichna* (Anderson, 1975a), *Merostomichnites* (Packard, 1900), *Mirandaichnium* (Aceñolaza, 1978), *Octopodichnus* (Gilmore, 1927), *Petalichnus* (Miller, 1880), *Pterichnus* (Hitchcock, 1865), *Tasmanadia* (Chapman, 1929), and *Umfolozia* (Savage, 1971). Most of these ichnotaxa further differ from SGDS 1290 in the shapes and configurations of their tracks.

Acanthichnus tracks (Fig. 4A), attributed to a chelicerate such as a solifugid, are oppositely arranged, short, elongate impressions in two (or four, per Dalman & Lucas, 2015) parallel rows; tracks either are parallel to or angle slightly away from the trackway axis (Dalman & Lucas, 2015; Hitchcock, 1858). This morphology and organization are both unlike those of SGDS 1290.

Asaphoidichnus tracks (Fig. 4B), attributed to trilobites, are elongate to crescentic, possess 3–4 crescentic branches at one end, and are oriented oblique to the trackway axis (Miller, 1880). They are far more complex in structure than the tracks of SGDS 1290.

Bifurculapes (Fig. 4C), attributed to an insect, possibly a beetle (Getty, 2016), comprises adjacent pairs (rarely triplets) of slightly staggered, elongate, straight to crescentic tracks that lie parallel or slightly oblique to the trackway axis, unlike the tracks of SGDS 1290. Tracks in each pair sometimes converge toward one end in *Bifurculapes*. This ichnotaxon typically does not possess paramedial impressions, but a specimen described by Getty (2016: fig. 1) possess two such traces, albeit faintly, that lie closer to the tracks than to the trackway axis, as in SGDS 1290. These impressions are far less pronounced than their associated tracks, unlike those of SGDS 1290. Getty (2020) ascertained that *Bifurculapes* traces were made subaqueously and may have been made by a terrestrial insect that would have left different tracks subaerially.

Coenobichnus tracks (Fig. 4D), attributed to a hermit crab, are thick, crescentic to ellipsoidal, roughly parallel and closely appressed to the trackway axis, and asymmetrical, with the left tracks larger than the right tracks (Walker, Holland & Gardiner, 2003), all of which differentiate this ichnotaxon from SGDS 1290.

Copeza (possibly a variant and synonym of *Lithographus* [Lull, 1953; Rainforth, 2005]; Fig. 4E) consists of triplets of roughly oppositely arranged, linear, elongate tracks in which the cranialmost lies roughly perpendicular to the trackway axis while the caudalmost pairs lie parallel or oblique to the trackway axis (Lull, 1953). This rare ichnotaxon is thus unlike SGDS 1290.

Danstairia congesta (Fig. 4F) comprises circular to crescentic tracks in closely appressed sets of up to six that are oriented oblique to the trackway axis; tracks often overlap to form V-shaped structures (Walker, 1985), unlike in SGDS 1290.

Diplichnites (possibly including *Acripes* per Häntzschel [1975] and Hammersburg, Hasiotis & Robison [2018]; also see below) tracks span a range of morphologies. *D. aenigma*

(Fig. 4G), ostensibly the ichnogenoholotype except that no specimen was designated as such (Stimson et al., 2018), typically comprises elongate, closely packed tracks in parallel rows on either side of the trackway axis; the tracks lie perpendicular to the trackway axis (Dawson, 1873). *D. gouldi* Type A (Fig. 4H) comprises parallel rows of closely spaced, oppositely arranged, simple, roughly circular to oblong to comma-shaped or irregular tracks with varying orientations to the trackway axis; *D. gouldi* Type B comprises closely spaced, elongate impressions oriented perpendicular, or nearly so, to the trackway axis, matching the general description of *D. aenigma*; *D. gouldi* Type C is similar to Type B, but the tracks are oriented oblique (~ 45°) to the trackway axis (Trewin & McNamara, 1994). *D. cuithensis*, attributed to large arthropleurid myriapods, is similar to both *D. aenigma* and *D. gouldi* Type B, but is very large and has widely spaced rows of tracks (Briggs, Rolfe & Brannan, 1979). *D. binatus* tracks often occur as closely appressed pairs of imprints (Webby, 1983). *D. triassicus* tracks are small and circular to ovoid rather than elongate, and frequently paired on either side of the trackway axis (e.g., Pollard, Selden & Watts, 2008); this ichnospecies has been alternately suggested to be a synonym of *D. gouldi* (e.g., Lucas et al., 2006b) or to pertain to *Acripes* (e.g., Machalski & Machalska, 1994; Pollard, 1985). *D. metzi* possesses a midline impression that is sometimes interrupted by connections between tracks in the closely appressed rows (Fillmore et al., 2017). The ichnogenus is in need of thorough review (Smith et al., 2003), but in all cases, the tracks are of different morphologies and arrangements than those of SGDS 1290.

Eisenachichnus tracks (Fig. 4I) are elongate, paired, and oblique to the trackway axis (rarely perpendicular), but the patterns of the pairs on either side of the trackway axis normally are asymmetrical: on one side, the paired tracks lie end to end, while on the other, they are more adjacent (Kozur, 1981). This morphology and arrangement are unlike those of SGDS 1290.

Foersterichnus (Fig. 4J), attributed to a crab, consists of widely spaced, paired rows of elongate tracks in sets of 3–4; rows are parallel to oblique to the trackway axis, and tracks in each set frequently overlap (Pirrie, Feldmann & Buatois, 2004). The wide spacing, clustering of tracks, and orientations of the tracks are unlike those of SGDS 1290.

Hamipes tracks (Fig. 4K) consist of closely spaced, paired, elongate to crescentic impressions oriented parallel to the trackway axis; the outer tracks are longer than their accompanying inner tracks, and the tracks are staggered or alternately arranged (Getty, 2018; Hitchcock, 1858). Track morphology readily differentiates *Hamipes* from SGDS 1290.

Lithographus (including *Permichnium sensu* Minter & Braddy, 2009) tracks (Fig. 4L), which match those made by pterygote insects, especially extant cockroaches (Davis, Minter & Braddy, 2007), comprise trios (or pairs, in the case of the *Permichnium* variant) of elongate to comma-shaped, rather than circular or tapering, tracks that are arranged at different angles to one another, some of which are oriented perpendicular to the trackway axis, and others of which are oblique to the trackway axis (Guthörl, 1934; Hitchcock, 1858; Minter & Braddy, 2009). These track morphologies and arrangements are distinct from those of SGDS 1290. *Hornburgichnium* reportedly is similar to *Permichnium*, but has three tracks on either side of the midline instead of two, and at least one of each set is oriented parallel to the trackway axis (Kozur, 1989); it may

also be a variant of *Lithographus* (Lucas et al., 2005a). Trackways of *Lithographus* can transition into trackways that Hitchcock (1858) called *Hexapodichnus* (Davis, Minter & Braddy, 2007; Minter, Braddy & Davis, 2007), so the latter may be considered a behavioral and/or substrate-consistency variant of the former, and also unlike SGDS 1290.

Maculichna (including *Guandacolichnus* and *Paganzichnus* of Pazos [2000] per Buatois & Mángano [2003]) tracks (Fig. 4M) comprise pairs (sometimes more) of small, circular to slightly elongate tracks arranged in closely appressed, slightly staggered rows. Pairings are oriented virtually parallel to the trackway axis (Anderson, 1975a); occasionally, short segments of linear, singular medial or closely spaced, paired paramedial imprints are also preserved that can be offset to one side of the trackway axis (Anderson, 1975a: fig. 8b, d, e). The pairing of *Maculichna* tracks differs from that of SGDS 1290. Aceñolaza & Buatois (1991, 1993) and Archer & Maples (1984) described *Maculichna* traces that exhibit the pairing of classic *Maculichna* from South Africa, but in which tracks are more ellipsoidal to shaped like slightly inflated isosceles triangles; the long axes of the triangles are oriented close to perpendicular to the trackway axis. Pazos (2000) recognized this morphology as the separate ichnotaxon *Paganzichnus*. This morphology is also unlike that of SGDS 1290.

The ichnogenoholotype of *Merostomichnites narragansettensis* (Fig. 4N) consists of parallel rows of roughly oppositely arranged circular to elongate to comma-shaped tracks whose long axes are perpendicular to the trackway axis (Packard, 1900). *Merostomichnites beecheri* tracks are circular and connected across the trackway axis by curvilinear, shallow, M-shaped imprints, creating a sort of segmented midline impression (Packard, 1900: fig. 4). These track and trace morphologies do not match those of SGDS 1290.

Mirandaichnium (Fig. 4O) consists of two rows of elongate, linear tracks that terminate laterally in small, circular impressions. Tracks are oriented perpendicular or oblique to the trackway axis, oppositely situated, and often grouped into series of eight (Aceñolaza & Buatois, 1993; Buatois et al., 1998), unlike those of SGDS 1290.

Octopodichnus (Fig. 4P) ichnospecies, attributed to arachnids, have different morphologies. *O. didactylus* tracks are circular to crescentic to bifurcate or trifurcate oriented parallel to the trackway axis in alternating, arcuate sets of four (Sadler, 1993). *O. minor* tracks have a similar organization, but the tracks are more amorphous; *O. raymondi* tracks consist of clusters of four circular to crescentic marks arranged in checkmark-like patterns (Sadler, 1993). These track morphologies and distinctive arrangements are substantially unlike those of SGDS 1290.

Petalichnus (Fig. 4Q), attributed to trilobites (Braddy & Almond, 1999), comprises sets of 2–3 elongate to crescentic tracks oriented perpendicular to the trackway axis (Miller, 1880). Anderson (1975b) and Braddy & Almond (1999) diagnosed *Petalichnus* tracks as sometimes bifurcate and occurring in series of 9–12; they further noted that the ichnotaxon needs review. Both track morphology and organization are unlike those of SGDS 1290.

Pterichnus tracks (Fig. 4R), attributed to isopods, frequently are segmented and are more linear and elongate than any in SGDS 1290. Tracks (or series of segments) are oriented oblique

(Types 1 and 2 of Gaillard et al., 2005) or parallel (Types 3 and 4 of Gaillard et al., 2005) to the trackway axis, and approximately symmetrically arranged in two parallel rows (Gaillard et al., 2005; Hitchcock, 1865) that are somewhat closer together than are those of the morphologically similar *Diplichnites*. Types 3 and 4 of Gaillard et al. (2005) morphologically grade into *Diplopodichnus* (Uchman et al., 2011). Hammersburg, Hasiotis & Robison (2018) suggested that *Pterichnus* comprises undertracks of *Lithographus* and is thus a junior synonym of that ichnotaxon. In any case, *Pterichnus* tracks are readily distinguished from SGDS 1290.

Tasmanadia traces (Fig. 4S) consist of two rows of closely packed, elongate, linear tracks oriented generally perpendicular to the trackway axis; occasionally, tracks overlap at one end, creating narrow, V-shaped structures (Chapman, 1929; Glaessner, 1957). Morphologically, this ichnotaxon resembles *Diplichnites gouldi* Type B and *Umfolozia* (but lacks the organization of the latter), and differs from SGDS 1290 for the same reasons as those ichnotaxa.

Umfolozia (Fig. 4T), attributed to syncarid and peracarid crustaceans (Lima, Minter & Netto, 2017; Savage, 1971), consists of parallel rows of irregularly shaped to crescentic tracks oriented perpendicular or oblique to the trackway axis that follow a unique repeating pattern (Anderson, 1981; Savage, 1971) unlike anything discernible in SGDS 1290. Aceñolaza & Buatois (1993) noted morphological similarities between *Mirandaichnium*, *Tasmanadia*, and *Umfolozia* and postulated similar track makers.

In summary, SGDS 1290 is not a variant of any of these ichnotaxa.

Arthropod repichnia possessing one medial impression

Several other arthropod locomotory (walking) ichnotaxa are also readily distinguished from SGDS 1290 by possessing singular medial, rather than paired paramedial, impressions; again, comparison is warranted to ascertain whether or not SGDS 1290 is a morphological variant of such ichnotaxa. These ichnotaxa are *Kouphichnium* (Caster, 1938; Nopcsa, 1923), "*Merostomichnites* isp." (Hanken & Stormer, 1975), *Oniscoidichnus* (Brady, 1947, 1949), *Palmichnium* (Richter, 1954), *Paleohelcura* (Gilmore, 1926), *Protichnites* (Owen, 1852), *Robledoichnus* (Kozur & Lemone, 1995), *Shalemichnus* (Kozur & Lemone, 1995), *Stiallia* (Smith, 1909), and *Stiaria* (Smith, 1909). As with traces lacking any medial impressions, these ichnotaxa further differ from SGDS 1290 in track morphology.

Kouphichnium traces (Fig. 4U) are attributed to limulids and occur in a variety of configurations. Tracks in clear *Kouphichnium* walking traces that possess singular medial impressions (e.g., many *K. lithographicum*, but not, for example, *K. minkinensis* [King, Stimson & Lucas, 2019; q.v. Gaillard, 2011a; Shu et al., 2018]) typically occur in sets of up to five in rows oriented oblique to the medial impression and trackway axis; individual tracks range from circular and ellipsoidal to elongate, and can split into anywhere from 2–5 branches at their ends (Caster, 1938, 1944; King, Stimson & Lucas, 2019; Shu et al., 2018). Well-preserved *Kouphichnium* tracks are dissimilar to those of SGDS 1290.

Traces referred to as "*Merostomichnites*" (Fig. 4V) and attributed to the eurypterid *Mixopterus* by Hanken & Stormer (1975) consist of three elongate and crescentic tracks in

oblique rows on either side of an intermittent medial impression; the tracks increase in size laterally, and some split into two or more branches on one end. In any of these cases, however, the tracks are substantially more complex than those of SGDS 1290.

Oniscoidichnus tracks (Fig. 4W) are elongate to crescentic, oriented perpendicular or oblique to the trackway axis, closely packed and closely appressed to the single midline impression (Brady, 1947; Davies, Sansom & Turner, 2006). In all these details, *Oniscoidichnus* traces differ markedly from SGDS 1290.

Ichnospecies of *Palmichnium* (Fig. 4X), also attributed to eurypterids, vary in morphology. Generally, they comprise complex sets of tracks lying lateral to a medial impression that can be either continuous or discontinuous. Tracks range in shape from elongate to crescentic to ovoid to chevron shaped, and they generally parallel the trackway axis. The tracks occur in oblique rows in sets of up to four; in some traces, the lateralmost tracks are elongate and curved, while the more medial tracks are linear and oriented parallel to the trackway axis (Braddy & Milner, 1998; Minter & Braddy, 2009; Poschmann & Braddy, 2010; Richter, 1954). Tracks are both more numerous and differently shaped than those of SGDS 1290.

Paleohelcura (including *Mesichnium* per Braddy [1995] and *Triavestigia* per Kozur, Löffler & Sittig [1994]; possibly a junior synonym of *Stiaria*; Fig. 4Y) traces, attributed to scorpions (Brady, 1947; Davis, Minter & Braddy, 2007), comprise small, circular tracks in sets of three in either rows, triangular arrangements, or checkmark-like patterns that lie external and oblique to the medial impression (Gilmore, 1926; Lagnaoui et al., 2015; Sadler, 1993). This distinctive layout is unlike that of SGDS 1290. Peixoto et al. (2020) attributed traces lacking a medial impression and comprising closely appressed pairs or triplets of mostly elliptical tracks from the Upper Jurassic or Lower Cretaceous of Brazil to a new ichnospecies of *Paleohelcura* and attributed them to a pterygote insect track maker. Tracks in this ichnospecies are arranged in rows oriented oblique to the trackway axis, and track sets in this ichnospecies lie close to the midline. This morphology is also unlike that of SGDS 1290.

Protichnites traces (Fig. 4Z) comprise thick, often segmented medial impressions (sometimes absent except on trackway turns) flanked by oppositely arranged, subcircular to ellipsoidal to irregularly shaped tracks with varying orientations to the trackway axis (Burton-Kelly & Erickson, 2010; Collette, Gass & Hagadorn, 2012; Hagadorn & Seilacher, 2009). They differ substantially from the tracks of SGDS 1290.

Robledoichnus tracks (Fig. 4AA), attributed to flying insects, resemble tracks of *Eisenachichnus* but possess a discontinuous, faint medial trace consisting entirely of periodic, V-shaped marks flanked by asymmetrical pairs of tracks. On one side, the tracks are short, tapered, and oriented oblique to the trackway axis; on the other side, the tracks are longer and crescentic, oriented closer to perpendicular to the trackway axis (Kozur & Lemone, 1995). Lucas et al. (2005b) considered *Robledoichnus* a probable junior synonym of *Paleohelcura* or *Stiaria*, and the ichnotaxon differs from SGDS 1290 for similar reasons as those ichnotaxa, in addition to the asymmetry.

Shalemichnus traces (Fig. 4BB), for which only half a trackway is known, consist of a straight medial impression punctuated at intervals by V-shaped marks. This impression is flanked by sets of three tapered tracks in straight rows oriented perpendicular to the trackway axis; individual tracks have their long axes parallel to the trackway axis (Kozur & Lemone, 1995). Minter & Braddy (2009) considered *Shalemichnus* a junior synonym of *Stiaria*. The tracks of *Shalemichnus* bear some similarity to those of SGDS 1290, but the paramedial impressions of SGDS 1290 lack the V-shaped markings of the medial impression of *Shalemichnus*.

Stiallia traces (Fig. 4CC) consist of paired rows of long, linear impressions parallel or slightly oblique to the trackway axis and that frequently overlap. *Stiallia pilosa* lacks any medial or paramedial impressions, but *Stiallia* (*Carrickia* of Smith [1909]) *berriana* possesses a medial row of crescentic to chevron-shaped marks (Smith, 1909; Walker, 1985). Pollard (1995) suggested that *Stiallia* could be an arthropod swimming, rather than a walking, trace, though it also resembles traces made by bristletail insects walking in highly saturated mud (Getty et al., 2013: fig. 6F, G). *Stiallia* tracks are markedly unlike those of SGDS 1290.

Stiaria tracks (including some ichnospecies of *Danstairia* of Smith [1909]; Fig. 4DD)), attributed to scorpionids (Braddy, 2003; Lucas, Lerner & Voigt, 2013) and monuran insects (Genise, 2017; Kopaska-Merkel & Buta, 2013), are oppositely situated groups of 2–4 generally circular to tapered tracks in a linear to crescentic arrangement lying roughly perpendicular to the trackway axis (Walker, 1985). In some specimens and ichnospecies of *Stiaria*, the singular medial impression actually varies in position, meandering from medial to almost lateral to their tracks (Fillmore, Lucas & Simpson, 2012: fig. 26d; Walker, 1985: fig. 5b, c). In some Mississippian specimens from Pennsylvania, the medial impression is flanked by thin, discontinuous, but closely appressed paramedial imprints (Fillmore, Lucas & Simpson, 2012: fig. 26d-g). Track arrangement alone differentiates *Stiaria* from SGDS 1290. Genise (2017) asserted that *Stiaria* should be considered a junior synonym of *Siskemia* (the latter has page priority over the former).

As with locomotory traces lacking medial impressions, SGDS 1290 is not a variant of any of these ichnotaxa.

Arthropod repichnia possessing three or more medial and paramedial impressions

Mitchellichnus (Fig. 4EE), attributed to archaeognathan insects (Getty et al., 2013), is distinguished from SGDS 1290 by possessing three medial impressions (Walker, 1985). *Mitchellichnus* tracks are complex, comprising two distinct types and arrangements. An inner set, lying close to the medial impressions, consists of apparently elongate tracks in sets of up to six that lie parallel to slightly oblique to the trackway axis; an outer set consists of larger, amorphous impressions (Walker, 1985). Tracks are thus more numerous in *Mitchellichnus* than in SGDS 1290, and the tracks differ in arrangement and morphology. Like *Stiaria*, Genise (2017) asserted that *Mitchellichnus* should be considered a junior synonym of *Siskemia*.

Keircalia (Fig. 4FF) is distinguished from SGDS 1290 by possessing four medial impressions (Smith, 1909; Walker, 1985). *Keircalia* tracks are crescentic to irregularly shaped, generally are oriented perpendicular to the trackway axis, and have no discernible arrangement (Walker, 1985). Both track morphology and organization are unlike those of SGDS 1290.

Arthropod repichnia possessing paired paramedial impressions

A few ichnotaxa, as well as some experimentally produced tracks of extant arthropods, resemble SGDS 1290 by possessing paired paramedial impressions in at least some specimens. Such ichnotaxa are *Danstairia vagusa* (Smith, 1909), *Glaciichnium* (Walter, 1985), *Warvichnium* (Walter, 1985), and *Siskemia* (Smith, 1909); similar extant traces include those made by notostracans (Trusheim, 1931) and crayfish (Fairchild & Hasiotis, 2011).

Danstairia vagusa (Fig. 4GG) possesses intermittent, thin, linear imprints that do not always parallel their accompanying trackways. Tracks are circular to triangular, generally have their long axes perpendicular to the trackway axis, and lack any coherent layout (Walker, 1985), unlike those of SGDS 1290. *D. vagusa* somewhat resembles *Keircalia* traces, but its tracks are spaced more widely apart.

Glaciichnium traces (Fig. 4HH), which resemble traces made by isopods (Gibbard & Stuart, 1974; Lima, Minter & Netto, 2017; Uchman, Kazakauskas & Gaigalas, 2009; Uchman et al., 2011), comprise 1–3 elongate, linear tracks ("bars" that are divided into segments [Uchman, Kazakauskas & Gaigalas, 2009]) that lie oblique to the trackway axis and are staggered on either side of that axis, unlike the tracks of SGDS 1290; their linear, serial but discontinuous paramedial imprints are widely spaced, abutting the medial ends of the tracks (Walter, 1985), farther apart than those of SGDS 1290. Walter (1985) and Lima et al. (2015) described the paramedial imprints in Brazilian specimens as comprising successive pairs of C-shaped imprints rather than strictly linear structures, further unlike SGDS 1290. Some *Glaciichnium* traces also possess a medial imprint as well (Uchman, Kazakauskas & Gaigalas, 2009; Walter, 1985).

Warvichnium traces (Fig. 4II) are complex, comprising pairs to multiple sets of linear, discontinuous medial and paramedial imprints flanked by varying numbers of linear to crescentic tracks in two or more sets: an inner set, close to the medial imprints, that are oriented slightly oblique to the trackway axis, and an outer set oriented closer to perpendicular to the trackway axis (Walter, 1985), quite unlike SGDS 1290. Getty (2020) noted similarities between *Warvichnium* and subaqueous *Bifurculapes*.

Among described arthropod repichnia, SGDS 1290 most closely resembles *Siskemia bipediculus* (Fig. 4JJ) and *S. elegans* (Fig. 4KK), which have been attributed to archaeognathan insects (Getty et al., 2013). These two ichnospecies differ primarily in the orientations of their track rows to the trackway axis and the continuities and thicknesses of their paramedial impressions (Walker, 1985), though these easily could be behavioral and/or substrate-driven variants. *Siskemia* tracks comprise pairs (in *S. bipediculus*) or trios (in some *S. elegans*) of roughly circular tracks in rows oriented oblique or perpendicular to a pair of paramedial impressions; the tracks of SGDS 1290 vary more in morphology, but share this general layout.

Siskemia tracks have a staggered distribution, similar to, but less pronounced than, in SGDS 1290. However, in both of these *Siskemia* ichnospecies, the paramedial impressions are thin, linear (especially in *S. bipediculus*), and much closer together and to the trackway axis than those of SGDS 1290. In fact, all of the ichnospecies of *Siskemia* erected by Smith (1909), as well as both specimens later attributed to this ichnotaxon (e.g., Getty et al., 2013; McNamara, 2014; Pollard, Steel & Undersrud, 1982) and *Siskemia*-like traces made by extant, archaeognathan insects (Getty et al., 2013), have such thin, linear, closely appressed paramedial impressions (sometimes offset toward one side of the trackway). The only time when archaeognathan traces approach the paramedial impression spacing of SGDS 1290 is when both abdominal styli (laterally) and gonostyli (medially) of the trace makers register impressions and imprints in wet mud, producing two sets of paramedial traces (Getty et al., 2013: fig. 6K, L), but even then the linear, lateralmost paramedial impressions still do not resemble the thick, undulating impressions of SGDS 1290. Simultaneously, in such wet mud, archaeognathan tracks are elongate and oriented oblique to the trackway axis, unlike those in SGDS 1290. In total, SGDS 1290 does not fit neatly into any known *Siskemia* ichnospecies and does not seem to be an archaeognathan insect trace.

Among traces made by extant arthropods, SGDS 1290 bears similarities to traces made by both notostracans and crayfish. Interpretive drawings of experimental traces made by notostracans figured by Trusheim (1931) depict elongate, crescentic, or tapered tracks oriented perpendicular to paramedial impressions; the tracks are arranged oppositely, unlike those of SGDS 1290. Additionally, the thin, linear paramedial impressions figured by Trusheim (1931) lie so far from the trackway axis that they often contact their accompanying tracks, a phenomenon that only occurs in SGDS 1290 near L4 and R4, where the lateral margins of its undulating paramedial impressions meander particularly far laterally. Tasch (1969) noted, though, that the drawings presented by Trusheim (1931) were misleading compared to his own experimentally generated notostracan traces. However, he described the morphologies of his notostracan tracks only as “minute en echelon stripes” (Tasch, 1969: 327), which does not adequately specify how they differed from those of Trusheim (1931); track details are impossible to discern in his lone photographic figure (Tasch, 1969: pl. 1.2). Gand et al. (2008) also conducted neoichnological experiments with notostracans, recovering locomotory traces that were less orderly than those illustrated by Trusheim (1931) (Gand et al., 2008: figs. 16.1, 16.2, 17.1). Their extant notostracan tracks comprised multiple tracks with rather chaotic distributions lateral to their paramedial impressions, unlike the regular distribution seen in SGDS 1290. Gand et al. (2008) found their notostracan traces to fall within the “etho-morphotype” of *Acripes*, as exemplified by their novel ichnospecies *A. multiformis* from the Permian of France. (Linck [1943] and Pollard [1985] also referred *Acripes* [*Merostomichnites* of Linck, 1943] tracks to notostracans, but not based on neoichnological experiments.) *A. multiformis* traces, unlike classic *Acripes* (Matthew, 1910), possess paramedial imprints, albeit inconsistently. Hammersburg, Hasiotis & Robison (2018), Häntzschel (1975), Miller (1996), and Pemberton, MacEachern & Gingras (2007) all supported classic *Acripes* as a junior synonym of *Diplichnites*; the issue of

synonymy is beyond the scope of this paper, but we note at least that the tracks in fossils that Gand et al. (2008) called *A. multiformis* differ from SGDS 1290 in the same ways that *Diplichnites* tracks do (see above). Lastly, Knecht et al. (2009: figs. 5, 6) also illustrated extant notostracan traces (Fig. 4LL), which are "tidier" than those of Gand et al. (2008) and resemble those of classic *Acripes* and *Diplichnites*, albeit with discontinuous paramedial and curvilinear lateral (external) imprints. The tracks in these traces comprise irregular, ellipsoidal sets oriented oblique to the trackway axis, unlike those of SGDS 1290. In total, SGDS 1290 is unlikely to be a notostracan trace.

Fairchild & Hasiotis (2011) conducted neoichnological experiments with crayfish to examine their locomotory traces. Although extant crayfish traces vary in morphology depending on substrate conditions and slope, in general, when clear, they consist of sets of 1–4 circular, tapering, ellipsoidal, or elongate tracks, occasionally of different sizes, that are oriented parallel to the trackway axis and that lie lateral to a pair of undulating, variably thick paramedial impressions that lie closer to their flanking tracks than to each other (Fig. 4MM). Morphologically, the tracks and impressions match those of SGDS 1290, although the tracks made by extant crayfish often are larger than those of SGDS 1290, depending on the substrate. Track sets in extant crayfish traces have complex arrangements: when comprised of multiple traces, they frequently are in rows oriented perpendicular to the trackway axis, but sometimes rows are oblique to the trackway axis. When fewer tracks are preserved, they can appear to lie in single, parallel rows on either side of the paramedial impressions. Track positions can be opposite to staggered, also as in SGDS 1290. In both track and paramedial impression morphology, as well as in overall trace architecture, SGDS 1290 strongly resembles crayfish traces made in damp silt and clay (Fairchild & Hasiotis, 2011: fig. 2e, f), dry and saturated, very fine-grained sand (Fairchild & Hasiotis, 2011: fig. 4a, b, e, f), dry and damp, fine-grained sand (Fairchild & Hasiotis, 2011: fig. 5a–d), and saturated medium sand (Fairchild & Hasiotis, 2011: fig. 6e, f). SGDS 1290 is preserved in, and was presumably registered in, a fine-grained sandstone, lithologically matching one set of experimental conditions in Fairchild & Hasiotis (2011). However, SGDS 1290 is not as detailed as many of the experimentally generated crayfish traces in comparable sediments. This could indicate one or more things: the fossil could be a slight overtrack (*sensu* Bertling et al., 2006) rather than a direct natural cast; the trace maker may have been partly buoyant; and/or trace-maker behavior and sediment consistency combined such that the lighter limbs did not impress as deeply as the heavier tail.

Discussion

Trace maker

The stronger resemblance of SGDS 1290 to experimental crayfish locomotion traces than to any known ichnotaxon, or other documented extant arthropod trace, implies a crayfish or crayfish-like maker for the fossil. Whether SGDS 1290 had a crayfish-like or an actual crayfish maker depends on whether the term "crayfish" is used to refer to members of a monophyletic clade (Parastacidae + (Astacidae + (Cambaridae + Cricoidoscelosidae)); Karasawa, Schweitzer

& Feldmann, 2013) of freshwater (and brackish water if *Protastacus* is included, *sensu* Karasawa, Schweitzer & Feldmann [2013]) lobsters, or, more broadly, to any freshwater, lobster-like crustacean regardless of phylogenetic position, which presumes that more than one crayfish-like lineage colonized terrestrial environments in the past. Here we use the term in the monophyletic sense: true crayfish comprise Astacida (*sensu* Karasawa, Schweitzer & Feldmann, 2013; Schram & Dixon, 2004; Shen, Braband & Scholtz, 2015). Whether or not the maker of SGDS 1290 was a true crayfish is unclear: the oldest undisputed crayfish body fossils are from the Early Cretaceous (Garassino, 1997; Martin et al., 2008; Shen, 2003; Taylor, Schram & Shen, 1999), although unnamed, Late Jurassic specimens from western Colorado also have been called crayfish (Hasiotis, Kirkland & Callison, 1998). Despite this, a Triassic or earlier origin for true crayfish has been hypothesized frequently (Breinholt, Pérez-Losada & Crandall, 2009; Crandall & Buhay, 2008; Porter, Pérez-Losada & Crandall, 2005; Schram, 2001; Schram & Dixon, 2004; Wolfe et al., 2019) and possibly substantiated by fossil burrows (domichnia) referred to crayfish from the Early Permian (Hembree & Swaninger, 2018) and Late Permian–Early Triassic (Baucon et al., 2014; Hasiotis & Mitchell, 1993).

Several Late Triassic body-fossil specimens also have been reported as crayfish (Hasiotis, 1995; Hasiotis & Mitchell, 1993; Miller & Ash, 1988; Olson & Huber, 1997; Santucci & Kirkland, 2010), but the identities of these specimens as true astacidans has not been established. Miller & Ash (1988) placed a Late Triassic specimen from Petrified Forest National Park, Arizona in *Enoploclytia*, which is an erymid lobster, not an astacidan, genus. That generic placement subsequently has been contested (Amati, Feldmann & Zonneveld, 2004; Schweitzer et al., 2010; Urreta, 1989), so the specimen needs detailed restudy, but if it pertains to Erymidae rather than Astacida, then it indicates that a lineage of erymid lobsters colonized terrestrial environments, possibly before true (monophyletic) crayfish. Some older analyses (reviewed in Rode & Babcock, 2003) postulated crayfish origins within Erymidae, but more recent phylogenetic analyses (Devillez, Charbonnier & Barreil, 2019; Karasawa, Schweitzer & Feldmann, 2013; Rode & Babcock, 2003; Schram & Dixon, 2004; Stern & Crandall, 2015) have recovered (a frequently paraphyletic) Erymidae with members at varying distances from Astacida. If those hypothesized phylogenetic relationships are correct, then no erymid can, in a monophyletic sense, be considered a crayfish, even if it was a freshwater taxon. But regardless of semantics or phylogenetic relationships, crayfish and erymid morphological similarities suggest that their locomotory traces might be indistinguishable, making a definitive attribution of SGDS 1290 impossible.

A crayfish or crayfish-like trace maker for SGDS 1290 is tenable both chronologically and ecologically. As mentioned above, multiple crayfish or crayfish-like morphotypes have been found in the Upper Triassic Chinle Formation of Arizona and Utah (Hasiotis, 1995; Miller & Ash, 1988; Santucci & Kirkland, 2010). The Moenave Formation overlies the Chinle Formation in southwestern Utah, so crayfish or crayfish-like decapods plausibly could have been present in and around freshwater Lake Whitmore both geographically and stratigraphically. As-yet-undescribed, crayfish or crayfish-like body fossils also have been recovered from lacustrine

sediments of the uppermost Triassic Chatham Group (Newark Supergroup) in North Carolina (Olsen & Huber, 1997), attesting to how widespread such arthropods were in terrestrial environments in North America even prior to the Jurassic.

Ichnotaxonomy

To date, no fossil arthropod locomotory ichnotaxon has been attributed definitively to a crayfish or crayfish-like trace maker. Several such ichnotaxa have been attributed, for various reasons, to crustaceans (e.g., Braddy, 2003; Gand et al., 2008; Lima, Minter & Netto, 2017; Pirrie, Feldmann & Buatois, 2004; Savage, 1971; Walker, Holland & Gardiner, 2003); additionally, some purported crustacean tracks have not been assigned to particular ichnotaxa (e.g., Imaizumi, 1967; Karasawa, Okamura & Naruse, 1990; Matsuoka et al., 1993), including mortichnial trackways leading to *Eryma*, *Eryon*, and *Mecochirus* lobster body fossils from the marine, Upper Jurassic lithographic limestones of Germany (Glaessner, 1969: fig. 243A; Seilacher, 2008: fig. 2; Viohl, 1998: fig. 6). None of these German taxa are crayfish, though morphological similarities of *Eryma* and *Mecochirus* to crayfish might mean that they would have produced indistinguishable locomotory trace fossils during normal, forward locomotion. None of their traces have been granted detailed description or ichnotaxonomic assignment.

Only three locomotory ichnotaxa have been attributed specifically to crayfish. First, Heidtke (1990) erected *Pollichianum repichnum* for Early Permian trace fossils from Germany that he attributed to the "crawfish" (in the English abstract; "Krebses" in the German abstract) *Uronectes fimbriatus*, also from the Early Permian of Germany. However, *Uronectes* has long been recognized as a syncarid (Brooks, 1962; Calman, 1934; Perrier et al., 2006), not an astacidan, or even a decapod, so the term appearing in the English abstract likely is a simple translation error. Furthermore, however, *P. repichnum* is not differentiable from the resting trace (cubichnion) *P. cubichnum* (O'Brien, Braddy & Radley, 2009) and therefore is a junior synonym and is not a locomotory trace. In any case, *Pollichianum* is morphologically quite unlike both SGDS 1290 and experimentally generated crayfish traces (Fairchild & Hasiotis, 2011). Second, Bolliger & Gubler (1997) hypothesized that their novel ichnospecies *Hamipes molassicus* was made by a buoyed (presumably swimming) crayfish. Getty (2018) referred these specimens to *Conopsoides*; later, Getty & Burnett (2019) suggested that at least some of the specimens may pertain to *Acanthichnus*. Third, de Gibert et al. (2000) attributed Early Cretaceous, Spanish specimens that they assigned to *Hamipes didactylus* to crayfish. Getty (2018) attributed these tracks to *Bifurculapes* and maintained a crustacean track maker for *H. didactylus*, but was not more specific. However, neither *Bifurculapes* nor *Hamipes* resemble experimentally generated crayfish traces (Fairchild & Hasiotis, 2011), or any of the mortichnial decapod traces, and thus are unlikely to have been made by a crayfish-like decapod, at least while walking.

As detailed above, SGDS 1290 does not fit neatly into any existing ichnotaxon. Whether or not to place it in a novel ichnotaxon is, therefore, an open question. Bertling et al. (2006), Gaillard (2011b), and Minter, Braddy & Davis (2007) provided solid criteria for the erection of new ichnotaxa, the latter particularly for arthropods. One criterion is that a new ichnotaxon

ideally should be represented by a substantial number of specimens that demonstrate behavioral and substrate-based morphological variation; this prevents erecting several ichnotaxa for minor, readily explained variations in trace morphology. SGDS 1290, as a singular specimen, certainly does not meet that criterion, but Minter, Braddy & Davis (2007) also allowed that truly unique morphologies exhibited by singular specimens can support an ichnotaxon. In terms of uniqueness, another criterion is whether or not a new morphotype falls onto a continuum, established or hypothetical, of morphologies within an established ichnotaxon. SGDS 1290 is closest morphologically to ichnospecies of *Siskemia*, but has several distinctions from any established ichnospecies therein, particularly the thick and undulating paramedial impressions and the wider spacing between the paramedial impressions and consequent closer appression of the paramedial impressions to the tracks. No published specimen of *Siskemia* demonstrates these features; nor do *Siskemia*-like traces made by archaeognathan insects in experimental conditions (Getty et al., 2013). Thus, SGDS 1290 does not appear to fall within the established *Siskemia* continuum. The greater prominence (depth) of the paramedial impressions of SGDS 1290 than their associated tracks suggests either a trace maker with heavier tail elements than the gonostyli of an archaeognathan insect or an archaeognathan trace maker with unusually large styli adopting an unusual posture (possibly partly buoyant), flexing its caudal region downward to create deep styli impressions but not deep track impressions. We consider the latter unlikely; thus, SGDS 1290 does not appear to fall within a hypothetical *Siskemia* continuum, either. However, SGDS 1290 falls within the continuum of trace morphologies made by extant crayfish in experimental conditions (Fairchild & Hasiotis, 2011). No philosophical basis has been established for the recognition of novel ichnotaxa on the basis of comparison with traces made by extant organisms; only by comparison with fossil ichnotaxa because extant traces cannot be the basis for an ichnotaxon (Bertling et al., 2006; International Commission on Zoological Nomenclature, 1999).

SGDS 1290 clearly is morphologically distinctive. Lacking a sufficient number of specimens with which to determine ranges of morphological variation, however, erecting a new ichnogenus for it clearly is unwarranted. Yet we feel that its unique morphology warrants ichnotaxonomic distinction. Given its distant similarity to *Siskemia* ichnospecies, we therefore herein place it in a new ichnospecies of that ichnogenus.

Systematic Ichnology

Ichnofamily Protichnidae Uchman, Gaździcki & Błażejowski 2018

Ichnogenus *Siskemia* Smith 1909

Type Ichnospecies *Siskemia elegans* Smith 1909

Diagnosis. Trace consisting of parallel rows of grouped tracks on either side of two parallel, paramedial impressions. Each group of tracks consists of up to four imprints arranged in series,

transversely or obliquely to the midline of the trackway (following Walker, 1985). Walker (1985) further specified that *Siskemia* was diagnosed by paramedial impressions with maximum widths of 0.5 mm, but following Bertling et al. (2006), size is not a suitable ichnotaxobase.

Ichnospecies *Siskemia eurypyge* isp. nov.
Figure 3

Diagnosis. Two parallel, undulating, paramedial impressions flanked externally by closely appressed sets of 1–3 small, ovoid to tapered to elongate tracks; tapered and elongate tracks have long axes parallel or oblique to the trackway axis. Track sets are oriented perpendicular to the trackway axis. Left and right tracks are arranged in a staggered pattern. Paramedial impressions are mediolaterally thick, but discontinuous, tapering out of existence briefly in some places. Impressions are gently undulating (low amplitude). The paramedial impressions lie far from the trackway axis, generally closer to (and sometimes in contact with) the tracks than to the midline axis or each other.
Holotype. Natural cast specimen SGDS 1290, St. George Dinosaur Discovery Site, St. George, Utah, USA.
Type locality. "Bug Crossing Quarry," SGDS Loc. 87, St. George Dinosaur Discovery Site, St. George, Washington County, Utah, USA (Fig. 1).
Stratigraphy. "Top Surface" of Johnson Farm Sandstone Bed (unit 40 of Kirkland et al., 2014), Whitmore Point Member, Moenave Formation (Fig. 2). Hettangian, Lower Jurassic.
Derivation of name. From the Greek ευρυς (eurys), meaning "broad" or "wide," and πυγή (pyge), meaning "rump." The ichnospecies name refers to the wider spacing between the paramedial impressions, made by the rear end of the trace maker, than those of other *Siskemia* ichnospecies.

Conclusions

SGDS 1290, from the Lower Jurassic (Hettangian) Whitmore Point Member of the Moenave Formation, consists of two paramedial impressions that are flanked by alternating sets of tapered or elongate tracks. The traces closely resemble those made by extant crayfish (Fairchild & Hasiotis, 2011) and are similar in components to traces placed in the ichnogenus *Siskemia* (Smith 1909; Walker, 1985). In previously recognized *Siskemia* ichnospecies, the paramedial impressions are thin, relatively linear, and closely appressed to the trackway axis. But in SGDS 1290, paramedial impressions have the opposite morphology and arrangement: they are thick and undulating, and lie closer to their tracks than the medial axis of the trackway. Thus, we erect a new ichnospecies, *Siskemia eurypyge*, to house SGDS 1290 and as-yet undiscovered traces with this morphology and arrangement.

Placing SGDS 1290 in *Siskemia* extends the known range of the ichnogenus into the Early Mesozoic. All other reported occurrences of the ichnogenus are Paleozoic in age: Early Silurian (McNamara, 2014; Trewin & McNamara, 1994), Late Silurian (Davies, Sansom &

690 Turner, 2006), Early Devonian (Pollard, Steel & Undersrud, 1982; Pollard & Walker, 1984;
691 Smith, 1909; Walker, 1985), and Pennsylvanian (Getty et al., 2013). Age should not be a factor
692 in ichnotaxonomy, however (Bertling et al., 2006). Paleozoic *Siskemia* traces likely were made
693 by archaeognathan insects (Getty et al., 2013), for which body fossils are known as early as the
694 Devonian and which are extant. Based on their similarity to traces made by extant crayfish
695 (Fairchild and Hasiotis, 2011), *S. eurypyge* likely was made by a crayfish or crayfish-like
696 crustacean, for which body fossils are known as early as the Late Triassic and which also are
697 extant. Thus, *Siskemia* ispp. traces would be expected to occur from Early Silurian to Recent, but
698 thus far have not been documented except for the occurrences noted above.
699 SGDS 1290 expands the ichnological record of crayfish and crayfish-like animals to
700 include repichnia in addition to domichnia. Fossil burrows (*Camborygma* ispp.), usually
701 attributed to crayfish, are well known at some sites and in some formations (e.g., Hasiotis, 1995;
702 Hasiotis & Bown, 1996; Hasiotis & Honey, 1995; Hasiotis & Mitchell, 1993; Hasiotis, Kirkland
703 & Callison, 1998; see Schram & Dixon [2004] concerning pre-Cretaceous examples), attesting to
704 the presences—and, in some places, abundances—of crayfish and/or crayfish-like taxa in
705 Mesozoic–Cenozoic freshwater paleoecosystems. Yet locomotion traces made by these
706 burrowers oddly have never before been documented as ichnofossils, possibly because they
707 infrequently venture far from their burrows in substrates suitable for registering locomotory
708 traces, as with modern crayfish (Martin, 2013). SGDS 1290 is the first documented locomotory
709 trace fossil made by a freshwater crayfish or crayfish-like organism, as well as the first fossil
710 evidence of such a taxon in the Lower Jurassic Moenave Formation and indeed the Early Jurassic
711 of the southwestern US. The absence of *Camborygma* burrows in the Moenave Formation that
712 would have been made by the SGDS 1290 trace maker is puzzling, and may be a consequence of
713 a lack of recognition; alternatively, the producer of SGDS 1290 was not a burrower.
714

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725 References Cited

726 Aceñolaza FG. 1978. Trazas fosiles de la Formación Patquía en el Bordo Atravesado, Sierra de
727 Famatina, La Rioja. *Acta Geológica Lilloana* 15:19–29.

728 Aceñolaza FG, Buatois LA. 1991. Trazas fosiles del Paleozoico superior continental argentino.
729 *Ameghiniana* 28:89–108.
730 Aceñolaza FG, Buatois LA. 1993. Nonmarine perigondwanic trace fossils from the Late
731 Paleozoic of Argentina. *Ichnos* 2:183–201. DOI: 10.1080/10420949309380092.
732 Amati L, Feldmann RM, Zonneveld J-P. 2004. A new family of Triassic lobsters (Decapoda:
733 Astacidea) from British Columbia and its phylogenetic context. *Journal of Paleontology*
734 78:150–168. DOI: 10.1666/0022-3360(2004)078<0150:ANFOTL>2.0.CO;2.
735 Anderson AM. 1975a. Turbidites and arthropod trackways in the Dwyka glacial deposits (Early
736 Permian) of southern Africa. *Transactions of the Geological Society of South Africa*
737 78:265–273.
738 Anderson AM. 1975b. The "trilobite" trackways in the Table Mountain Group (Ordovician) of
739 South Africa. *Palaeontologia Africana* 18:35–45.
740 Anderson AM. 1981. The *Umfolozia* arthropod trackways in the Permian Dwyka and Ecca series
741 of South Africa. *Journal of Paleontology* 55:84–108.
742 Archer AW, Maples CG. 1984. Trace-fossil distribution across a marine-to-nonmarine gradient
743 in the Pennsylvanian of southwestern Indiana. *Journal of Paleontology* 58:448–466.
744 Baucon A, Ronchi A, Felletti F, de Carvalho CN. 2014. Evolution of crustaceans at the edge of
745 the end-Permian crisis: ichnonetwork analysis of the fluvial succession of Nurra
746 (Permian–Triassic, Sardinia, Italy). *Palaeogeography, Palaeoclimatology, Palaeoecology*
747 410:74–103. DOI: 10.1016/j.palaeo.2014.05.034.
748 Bertling M, Braddy SJ, Bromley RG, Demathieu GR, Genise J, Mikuláš R, Nielsen JK, Nielsen
749 KSS, Rindsberg AK, Schlirf M, Uchman, A. 2006. Names for trace fossils: a uniform
750 approach. *Lethaia* 39:265–286. DOI: 10.1080/00241160600787890.
751 Bolliger, T, Gubler, T. 1997. Arthropodenlaufspuren von Hochdorf (LU) und weitere
752 Arthropodennachweise aus der Oberen Süsswassermolasse der Schweiz. *Eclogae*
753 *Geologicae Helvetiae* 90:617–627.
754 Braddy SJ. 1995. The ichnotaxonomy of the invertebrate trackways of the Coconino Sandstone
755 (Lower Permian), northern Arizona. In: Lucas SG, Heckert AB, eds. *Early Permian*
756 *Footprints and Facies*. *New Mexico Museum of Natural History and Science Bulletin* 6:
757 219–224.
758 Braddy SJ. 2003. Trackways—arthropod locomotion. In: Briggs DEG, Crowther PR, eds.
759 *Palaeobiology II*. Malden: Blackwell Publishing, 389–393.
760 Braddy SJ, Almond JE. 1999. Eurypterid trackways from the Table Mountain Group
761 (Ordovician) of South Africa. *Journal of African Earth Sciences* 29:165–177. DOI:
762 10.1016/S0899-5362(99)00087-1.
763 Braddy SJ, Milner ARC. 1998. A large arthropod trackway from the Gaspé Sandstone Group
764 (Middle Devonian) of eastern Canada. *Canadian Journal of Earth Science* 35:1116–
765 1122. DOI: 10.1139/cjes-35-10-1116.
766 Brady LF. 1947. Invertebrate tracks from the Coconino Sandstone of northern Arizona. *Journal*
767 *of Paleontology* 21:466–472.

768 Brady LF. 1949. *Oniscoidichnus*, new name for *Isopodichnus* Brady 1947 not Bornemann 1889.
769 *Journal of Paleontology* 23:573.
770 Breinholt J, Pérez-Losada M, Crandall KA. 2009. The timing and diversification of freshwater
771 crayfishes. In: Martin JW, Crandall KA, Felder DJ, eds. *Decapod Crustacean*
772 *Phylogenetics*. Boca Raton: CRC Press, 343–356.
773 Briggs DEG, Rolfe WDI, Brannan J. 1979. A giant myriapod trail from the Namurian of Arran,
774 Scotland. *Palaeontology* 22:273–291.
775 Brooks HK. 1962. On the fossil Anaspidacea, with a revision of the classification of the
776 Syncarida. *Crustaceana* 4:229–242.
777 Buatois LA, Mángano MG. 2003. Caracterización ichnológica y paleoambiental de la localidad
778 tipo de *Orchesteropus atavus* Frenguelli, Huerta de Huachi, Provincia de San Juan,
779 Argentina. *Ameghiniana* 40:53–70.
780 Buatois LA, Mángano MG. 2004. Animal–substrate interactions in freshwater environments:
781 applications of ichnology in facies and sequence stratigraphic analysis of fluvio-
782 lacustrine successions. In: McIlroy D, ed. *The Application of Ichnology to*
783 *Palaeoenvironmental and Stratigraphic Analysis*. *Geological Society of London Special*
784 *Publication* 228:311–333.
785 Buatois LA, Mángano MG, Maples CG, Lanier WP. 1998. Ichnology of an Upper Carboniferous
786 fluvio-estuarine paleovalley: the Tonganoxie Sandstone, Buildex Quarry, eastern Kansas,
787 USA. *Journal of Paleontology* 72:152–180. DOI: 10.1017/S0022336000024094.
788 Burton-Kelly ME, Erickson JM. 2010. A new occurrence of *Protichnites* Owen, 1852, in the
789 Late Cambrian Potsdam Sandstone of the St. Lawrence Lowlands. *Open Paleontology*
790 *Journal* 3:1–13. DOI: 10.2174/1874425701003010001.
791 Calman WT. 1934. Notes on *Uronectes fimbriatus* (Jordan), a fossil crustacean of the division
792 Syncarida. *Annals and Magazine of Natural History, 10th Series* 75:321–330.
793 Caster KE. 1938. A restudy of the tracks of *Paramphibius*. *Journal of Paleontology* 12:3–60.
794 Caster KE. 1944. Limuloid trails from the Upper Triassic (Chinle) of the Petrified National
795 Forest National Monument, Arizona. *American Journal of Science* 242:74–84. DOI:
796 10.2475/ajs.242.2.
797 Chapman F. 1929. On some remarkable annelid remains from Arthur River, N.W. Tasmania.
798 *Papers and Proceedings of the Royal Society of Tasmania* 1928:1–5.
799 Charbonnier S, Vannier J, Hantzpergue P, Gaillard C. 2010. Ecological significance of the
800 arthropod fauna from the Jurassic (Callovian) La Voulte Lagerstätte. *Acta*
801 *Palaeontologica Polonica* 55:111–132. DOI: 10.4202/app.2009.0036.
802 Chen Z, Chen X, Zhou C, Yuan X, Xiao, S. 2018. Late Ediacaran trackways produced by
803 bilaterian animals with paired appendages. *Science Advances* 4:eaao669. DOI:
804 10.1126/sciadv.aao6691.
805 Collette JH, Gass KC, Hagadorn JW. 2012. *Protichnites eremita* unshelled? Experimental
806 model-based neoichnology and new evidence for a euthycarcinoid affinity for this
807 ichnospecies. *Journal of Paleontology* 86:442–454. DOI: 10.1666/11-056.1.

Collette JH, Hagadorn JW, Lacelle MA. 2010. Dead in their tracks—Cambrian arthropods and their traces from intertidal sandstones of Quebec and Wisconsin. *Palaios* 25:475–486. DOI: 10.2110/palo.2009.p09-134r.

Crandall KA, Buhay JE. 2008. Global diversity of crayfish (Astacidae, Cambaridae, and Parastacidae—Decapoda) in freshwater. *Hydrobiologia* 595:295–301. DOI: 10.1007/s10750-007-9120-3.

Dalman SG, Lucas SG. 2015. Lower Jurassic arthropod resting trace from the Hartford Basin of Massachusetts, USA. *Ichnos* 22:177–182. DOI: 10.1080/10420940.2015.1059337.

Davies NS, Sansom IJ, Turner P. 2006. Trace fossils and paleoenvironments of a Late Silurian marginal-marine/alluvial system: the Ringerike Group (Lower Old Red Sandstone), Oslo region, Norway. *Palaios* 21:46–62. DOI: 10.2110/palo.2003.p03-08.

Davis RB, Minter NJ, Braddy SJ. 2007. The neoichnology of terrestrial arthropods. *Palaeogeography, Palaeoclimatology, Palaeoecology* 255:284–307. DOI: 10.1016/j.palaeo.2007.07.013.

Dawson JW. 1873. Impressions and footprints of aquatic animals and imitative markings, on Carboniferous rocks. *American Journal of Science* 105:16–24. DOI: 10.2475/ajs.s3-5.25.16.

de Gibert JM, Fregenal-Martínez MA, Buatois LA, Mángano MG. 2000. Trace fossils and their palaeoecological significance in Lower Cretaceous lacustrine conservation deposits, El Montsec, Spain. *Palaeogeography, Palaeoclimatology, Palaeoecology* 156:89–101. DOI: 10.1016/S0031-0182(99)00133-9.

Devillez J, Charbonnier S, Barreil, V. 2019. An attempt to clarify phylogenetic affinities of erymid lobsters (Decapoda) using morphological characters. *Arthropod Systematics and Phylogeny* 77:365–395. DOI: 10.26049/ASP77-3-2019-1.

Donohoo-Hurley LL, Geissman JW, Lucas SG. 2010. Magnetostratigraphy of the uppermost Triassic and lowermost Jurassic Moenave Formation, western United States: correlation with strata in the United Kingdom, Morocco, Turkey, Italy, and eastern United States. *Geological Society of America Bulletin* 122:2005–2019. DOI: 10.1130/B30136.1.

Fairchild JM, Hasiotis ST. 2011. Terrestrial and aquatic neoichnological laboratory experiments with the freshwater crayfish *Oronectes*: trackways on media of varying grain size, moisture, and inclination. *Palaios* 26:790–804. DOI: 10.2110/palo.2011.p11-066r.

Fillmore DL, Lucas SG, Simpson EL. 2012. Ichnology of the Mississippian Mauch Chunk Formation, eastern Pennsylvania. *New Mexico Museum of Natural History and Science Bulletin* 54:1–136.

Fillmore DL, Szajna MA, Lucas SG, Hartline BW, Simpson EL. 2017. Ichnology of a Late Triassic lake margin: the Lockatong Formation, Newark Basin, Pennsylvania. *New Mexico Museum of Natural History and Science Bulletin* 76:1–107.

Gaillard C. 2011a. A giant limulid trackway (*Kouphichnium lithographicum*) from the lithographic limestones of Cerin (Late Kimmeridgian, France): ethological and

environmental implications. *Swiss Journal of Geosciences* 104(suppl. 1):57–72. DOI: 10.1007/s00015-010-0032-2.

Gaillard C. 2011b. Ichnotaxinomie et notion d'ichnoespèce. *Comptes Rendus Palevol* 10:209–218. DOI: 10.1016/j.crpv.2010.10.007.

Gaillard C, Hantzpergue P, Vannier J, Margerard AL, Mazin JM. 2005. Isopod trackways from the Crayssac Lagerstätte, Upper Jurassic, France. *Palaeontology* 48:947–962.

Gand G, Garric J, Schneider J, Walter H, Lapeyrie J, Martin C, Thiery, A. 2008. Notostraca trackways in Permian playa environments of the Lodève Basin (France). *Journal of Iberian Geology* 34:73–108.

Garassino A. 1997. The macruran decapod crustaceans of the Lower Cretaceous (lower Barremian) of Las Hoyas (Cuenca, Spain). *Atti della Società Italiana di Scienze Naturali e del Museo Civico di Storia Naturale di Milano* 137:101–126.

Genise JF. 2017. *Ichnoentomology: Insect Traces in Soils and Paleosols*. Switzerland: Springer International Publishing AG. DOI: 10.1007/978-3-319-28210-7.

Getty PR. 2016. *Bifurculapes* Hitchcock 1858: a revision of the ichnogenus. *Atlantic Geology* 52:247–255. DOI: 10.4138/atlgeol.2016.013.

Getty PR. 2018. Revision of the Early Jurassic arthropod trackways *Camurichnus* and *Hamipes*. *Boletín de la Sociedad Geológica Mexicana* 70:281–292. DOI: 10.18268/BSGM2018v70n2a1.

Getty PR. 2020. Evidence that the fossil insect trackway *Bifurculapes laqueatus* Hitchcock, 1858 was made underwater. *Acta Geologica Polonica* 70:125–133. DOI: 10.24425/agp.2019.126456.

Getty PR, Burnett J. 2019. *Conopsoides* Hitchcock 1858: an ichnological chimera of *Acanthichnus* and *Bifurculapes*. *Atlantic Geology* 55:389–398. DOI: 10.4138/atlgeol.2019.014.

Getty PR, Sproule R, Wagner DL, Bush AM. 2013. Variation in wingless insect trace fossils: insights from neoichnology and the Pennsylvanian of Massachusetts. *Palaios* 28:243–258. DOI: 10.2110/palo.2012.p12-108r.

Gibbard PL, Stuart AJ. 1974. Trace fossils from proglacial lake sediments. *Boreas* 3:69–74.

Gilmore CW. 1926. Fossil footprints from the Grand Canyon. *Smithsonian Miscellaneous Collections* 77(9):1–41.

Gilmore CW. 1927. Fossil footprints from the Grand Canyon: second contribution. *Smithsonian Miscellaneous Collections* 80(3):1–78.

Glaessner MF. 1957. Paleozoic arthropod trails from Australia. *Paläontologische Zeitschrift* 31:103–109. DOI: 10.1007/BF02988968.

Glaessner MF. 1969. Decapoda. In: Moore, RC, ed. *Treatise on Invertebrate Paleontology Part R: Arthropoda 4, Vol. 2*. Lawrence: University of Kansas Printing Service, 399–566.

Good TR, Ekdale AA. 2014. Paleoecology and taphonomy of trace fossils in the eolian Upper Triassic/Lower Jurassic Nugget Sandstone, northeastern Utah. *Palaios* 29:401–413. DOI: 10.2110/palo.2014.013.

887 Guthörl P. 1934. Die Arthropoden aus dem Carbon und Perm des Saar-Nahe-Pfalz-Gebietes.
888 *Abhandlungen der Preußischen Geologischen Landesanstalt, Neue Folge* 164:1–219.
889 Hagadorn JW, Seilacher A. 2009. Hermit arthropods 500 million years ago? *Geology* 37:295–
890 298. DOI: 10.1130/G25181A.1.
891 Hammersburg SR, Hasiotis ST, Robison RA. 2018. Ichnotaxonomy of the Cambrian Spence
892 Shale Member of the Langston Formation, Wellsville Mountains, northern Utah, USA.
893 *Paleontological Contributions* 2018(20):1–66. DOI: 10.17161/1808.26428.
894 Hanken NS, Stormer L. 1975. The trail of a large Silurian eurypterid. *Fossils and Strata* 4:255–
895 270.
896 Häntzschel W. 1975. *Treatise on Invertebrate Paleontology Part W: Miscellanea, Supplement 1:*
897 *Trace Fossils and Problematica, 2nd Ed.* Lawrence: University of Kansas Printing
898 Service.
899 Hasiotis ST. 1995. Crayfish fossils and burrows from the Upper Triassic Chinle Formation,
900 Canyonlands National Park, Utah. In: Santucci VL, McClelland L, eds. *National Park*
901 *Service Research Volume 2.* National Park Service Technical Report NPS/NRPO/NRTR-
902 95/16:49–53.
903 Hasiotis ST, Bown TM. 1996. A short note about crayfish burrows from the Paleocene–Eocene
904 Claron Formation, southwestern Utah, USA. *Freshwater Crayfish* 11:121–129.
905 Hasiotis ST, Honey JG. 1995. Crayfish burrows and their paleohydrologic significance in
906 Paleocene continental deposits, Washakie, Great Divide, and Hanna Basins, Wyoming,
907 USA. *Freshwater Crayfish* 10:105–117.
908 Hasiotis ST, Kirkland JI, Callison G. 1998. Crayfish fossils and burrows from the Upper Jurassic
909 Morrison Formation of western Colorado. In: Carpenter K, Chure DJ, Kirkland JI, eds.
910 *The Upper Jurassic Morrison Formation: An Interdisciplinary Study, Part 2. Modern*
911 *Geology* 22:481–491.
912 Hasiotis ST, Mitchell CE. 1993. A comparison of crayfish burrow morphologies: Triassic and
913 Holocene fossil, paleo- and neo-ichnological evidence, and the identification of their
914 burrowing signatures. *Ichnos* 2:291–314. DOI: 10.1080/10420949309380104.
915 Heidtke U. 1990. *Pollichianum cubichnum* nov. ichnogen. et ichnospec. und *Pollichianum*
916 *repichnum* nov. ichnospec., zwei neue Invertebratenichnia aus dem saar-pfälzischen
917 Rotliegend (Unter-Perm, SW-Deutschland). *Mitteilungen der Pollichia* 77:133–139.
918 Hembree DI, Swaninger ES. 2018. Large *Camborygma* isp. in fluvial deposits of the Lower
919 Permian (Asselian) Dunkard Group, southeastern Ohio, U.S.A. *Palaeogeography,*
920 *Palaeoclimatology, Palaeoecology* 491:137–151. DOI: 10.1016/j.palaeo.2017.12.003.
921 Hitchcock E. 1858. *A Report on the Sandstone of the Connecticut Valley, Especially Its Fossil*
922 *Footmarks, Made to the Government of the Commonwealth of Massachusetts*. Boston:
923 William White.
924 Hitchcock E. 1865. *Supplement to the Ichnology of New England. A Report to the Government of*
925 *Massachusetts, in 1863*. Boston: Wright and Potter State Printers.

Hunt AP, Lucas SG. 2006a. The significance of the St. George Dinosaur Discovery Site at Johnson Farm: a Jurassic Konzentrat-Ichnolagerstätte, Utah, U.S.A. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:282–288.

Hunt AP, Lucas SG. 2006b. Triassic–Jurassic tetrapod ichnofacies. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:12–22.

Hunt AP, Lucas SG 2006c. Tetrapod ichnofacies: a new paradigm. *Ichnos* 14: 59–68. DOI:10.1080/10420940601006826.

Imaizumi R. 1967. On some Lebensspuren from the Tatsunokuchi Formation at Tatsunokuchi Gorge, Sendai, Miyagi Prefecture. In: Commemorative Association of Prof. Ichirō Hayasaka's 76th Birthday, ed. *Contributions to Celebrate Prof. Ichirō Hayasaka's 76th Birthday*. Kanazawa: Commemorative Association of Prof. Ichirō Hayasaka's 76th Birthday, 205–210.

International Commission on Zoological Nomenclature. 1999. *International Code of Zoological Nomenclature, Fourth Edition.* London: The International Trust for Zoological Nomenclature.

Karasawa H, Okumura Y, Naruse A. 1990. Discovery of crab footprints from the Miocene Mizunami Group, central Japan. *Bulletin of the Mizunami Fossil Museum* 17:93–96.

Karasawa H, Schweitzer CE, Feldmann RM. 2013. Phylogeny and systematics of extant and extinct lobsters. *Journal of Crustacean Biology* 33:78–123. DOI: 10.1163/1937240X-00002111.

King OA, Stimson MR, Lucas SG. 2019. The ichnogenus *Kouphichnium* and related xiphosuran traces from the Steven C. Minkin Paleozoic footprint site (Union Chapel Mines), Alabama, USA: ichnotaxonomic and paleoenvironmental implications. *Ichnos* 26:266–302. DOI: 10.1080/10420940.2018.1561447.

Kirkland JI, Milner ARC. 2006. The Moenave Formation at the St. George Dinosaur Discovery Site at Johnson Farm, St. George, southwestern Utah. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:289–309.

Kirkland JI, Milner ARC, Olsen PE, Hargrave JE. 2014. The Whitmore Point Member of the Moenave Formation in its type area in northern Arizona and its age and correlation with the section in St. George, Utah: evidence for two major lacustrine sequences. In: MacLean JS, Biek RF, Huntoon JE, eds. *Geology of Utah's Far South*. *Utah Geological Association Publication* 43:321–356.

Knecht RJ, Benner JS, Rogers DC, Ridge JC. 2009. *Surculichnus bifurcauda* n. igen., n. isp., a trace fossil from late Pleistocene glaciolacustrine varves of the Connecticut River Valley, USA, attributed to notostracan crustaceans based on neoichnological experimentation. *Palaeogeography, Palaeoclimatology, Palaeoecology* 272:232–239. DOI: 10.1016/j.palaeo.2008.10.013.

Kopaska-Merkel DC, Buta RJ. 2013. Field trip guidebook to the Steven C. Minkin Paleozoic footprint site, Walker County, Alabama. In: Lucas SG, DiMichele WA, Barrick JE, Schneider JW, Spielmann JA, eds. *The Carboniferous–Permian Transition*. *New Mexico Museum of Natural History and Science Bulletin* 60:178–198.

Kozur H. 1981. Weitere Beiträge zur Paläontologie und Stratigraphie des Perm. *Geologische–Paläontologische Mitteilungen, Innsbruck* 11:243–257.

Kozur H. 1989. Biostratigraphic zonations in the Rotliegendes and their correlations. *Acta Musei Reginaehradecensis S.A.: Scientiae Naturales* 22:15–30.

Kozur HW, Lemone DV. 1995. New terrestrial arthropod trackways from the Abo Member (Sterlitamakian, Late Sakmarian, late Wolfcampian) of the Shalem Colony Section, Robledo Mountains, New Mexico. In: Lucas SG, Heckert AB, eds. *Early Permian Footprints and Facies*. *New Mexico Museum of Natural History and Science Bulletin* 6:107–113.

Kozur H, Löffler M, Sittig E. 1994. First evidence of *Paleohelcura* (arthropod trackway) in the Rotliegend of Europe: *Neues Jahrbuch für Geologie und Paläontologie Monatshefte* 1994:618–632. DOI: 10.1127/njgpm/1994/1994/618.

Kozur HW, Weems RE. 2010. The biostratigraphic importance of conchostracans in the continental Triassic of the northern hemisphere. In: Lucas SG, ed., *The Triassic Timescale*. *Geological Society of London Special Publications* 334:315–417. DOI: 10.1144/ SP334.13.

Labandeira CC, Beall BS. 1990. Arthropod terrestriality. In: Mikulic DG, ed. *Arthropod Paleobiology: Short Courses in Paleobiology* 3. Knoxville: Paleontological Society, 214–256.

Lagnaoui A, Voigt S, Belahmira A, Saber H, Schneider JW, Hminna A. 2015. New ichnospecie of the ichnogenus *Paleohelcura* Gilmore, 1926 from Late Permian red-beds of Argana Basin (Western High Atlas Mountains, Morocco). In: Saber H, Lagnaoui A, Belahmira A, eds. *Proceedings of Abstracts, 1st International Congress on Continental Ichnology*. *Arabian Journal of Earth Sciences Special Issue* ICCI-2015:52–54.

Lima JHD, Minter NJ, Netto RG. 2017. Insights from functional morphology and neoichnology for determining tracemakers: a case study of the reconstruction of an ancient glacial arthropod-dominated fauna. *Lethaia* 50:576–590. DOI: 10.1111/let.12214.

Lima JHD, Netto RG, Corrêa CG, Lavina ELC. 2015. Ichnology of deglaciation deposits from the Upper Carboniferous Rio do Sul Formation (Itararé Group, Paraná Basin) at central-east Santa Catarina State (southern Brazil). *Journal of South American Earth Sciences* 63:137–148. DOI: 10.1016/j.jsames.2015.07.008.

Linck O. 1943. Die Buntsandstein-Kleinfährten von Nagold. (*Limulidichnulus nagoldensis* n. g. n. sp., *Merostomichnites triassicus* n. sp.). *Neues Jahrbuch für Mineralogie, Geologie und Paläontologie Monatshefte Abt. B* 1943:9–27.

Lockley MG, Kirkland JI, Milner ARC. 2004. Probable relationships between the Lower Jurassic crocodilomorph trackways of *Batrachopus* and *Selenichnus*: evidence and implications

based on new finds from the St. George area southwestern Utah. *Ichnos* 11:143–149. DOI: 10.1080/10420940490442340.

Lomax DE, Racay CA. 2012. A long mortichnial trackway of *Mesolimulus walchi* from the Upper Jurassic Solnhofen lithographic limestone near Wintershof, Germany. *Ichnos* 19:175–183. DOI: 10.1080/10420940.2012.702704.

Lucas SG, Lerner AJ, Milner ARC, Lockley MG. 2006a. Lower Jurassic invertebrate ichnofossils from a clastic lake margin, Johnson Farm, southwestern Utah. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:128–136.

Lucas SG, Lerner AJ, Spielmann JA, Hunt AP. 2006b. Invertebrate ichnofossils from the Upper Triassic Bull Canyon Formation, east-central New Mexico. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:118–121.

Lucas SG, Lerner AJ, Voigt S. 2013. Scorpionid resting trace from the Lower Permian of southern New Mexico, USA. *Ichnos* 20:195–201. DOI: 10.1080/10420940.2013.845096.

Lucas SG, Minter NJ, Spielmann JA, Hunt AP, Braddy SJ. 2005b. Early Permian ichnofossil assemblage from the Fra Cristobal Mountains, southern New Mexico. In: Lucas SG, Zeigler KE, Spielmann JA, eds. *The Permian of Central New Mexico*. *New Mexico Museum of Nature and Science Bulletin* 31:140–150.

Lucas SG, Minter NJ, Spielmann JA, Smith JA, Braddy SJ. 2005a. Early Permian ichnofossils from the northern Caballo Mountains, Sierra County, New Mexico. In: Lucas SG, Zeigler KE, Spielmann JA, eds. *The Permian of Central New Mexico*. *New Mexico Museum of Nature and Science Bulletin* 31:151–162.

Lucas SG, Tanner LH. 2006. The Springdale Member of the Kayenta Formation, Lower Jurassic of Utah–Arizona. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:71–76.

Lull RS. 1953. Triassic life of the Connecticut Valley. *State of Connecticut State Geological and Natural History Survey Bulletin* 81:1–336.

Luque J, Feldmann RM, Vernygora O, Schweitzer CE, Cameron DB, Kerr KA, Vega FJ, Duque A, Strange M, Palmer AR, Jaramillo, C. 2019. Exceptional preservation of mid-Cretaceous marine arthropods and the evolution of novel forms via heterochrony. *Science Advances* 5:eaav3875. DOI: 10.1126/sciadv.aav3875.

Machalski M, Machalska K 1994. Arthropod trackways, "*Diplichnites*" *triassicus* (Linck, 1943), from the Lower Triassic (Buntsandstein) fluvial deposits of the Holy Cross Mts, central Poland. *Acta Geologica Polonica* 44:267–275.

MacNaughton RB, Cole JM, Dalrymple RW, Braddy SJ, Briggs DEG, Lukie, TD. 2002. First steps on land: arthropod trackways in Cambrian–Ordovician eolian sandstone,

1045 southeastern Ontario, Canada. *Geology* 30:391–394. DOI: 10.1130/0091-
1046 7613(2002)030<0391:FSOLAT>2.0.CO;2.
1047 Martin AJ. 2013. *Life Traces of the Georgia Coast: Revealing the Unseen Lives of Plants and*
1048 *Animals*. Bloomington: Indiana University Press.
1049 Martin AJ, Rich TH, Poore GCB, Schultz MB, Austin CM, Kool L, Vickers-Rich P. 2008. Fossil
1050 evidence in Australia for oldest known freshwater crayfish of Gondwana. *Gondwana*
1051 *Research* 14:287–296. DOI: 10.1016/j.gr.2008.01.002.
1052 Matsuoka K, Karasawa H, Yoshimura A, Ieda K. 1993. Footprint fossils discovered from the
1053 Miocene Shidara Group, Toei-cho, Kitashitara-gun, Aichi Prefecture, Japan. *Science*
1054 *Reports of the Toyohashi Museum of Natural History* 3:27–36.
1055 Matthew GF. 1910. Remarkable forms of the Little River Group. *Transactions of the Royal*
1056 *Society of Canada* 4:115–125.
1057 McNamara KJ. 2014. Early Paleozoic colonisation of the land: evidence from the Tumblagooda
1058 Sandstone, southern Carnarvon Basin, western Australia. *W A Science—Journal of the*
1059 *Royal Society of Western Australia* 97:111–132.
1060 Melchor RN, Cardonatto MC. 2014. Insights on behavior of Late Paleozoic aquatic crustaceans
1061 (Pygocephalomorpha?): compound trace fossils from western Argentina. *Ichnos* 21:76–
1062 99. DOI: 10.1080/10420940.2013.879868.
1063 Miller GL, Ash SR. 1988. The oldest freshwater decapod crustacean, from the Triassic of
1064 Arizona. *Palaeontology* 31:273–279.
1065 Miller RF. 1996. Location of trace fossils and problematica of George Frederic Matthew from
1066 Part W, Treatise on Invertebrate Paleontology. *Journal of Paleontology* 70:161–171.
1067 DOI: 10.1017/S0022336000023210.
1068 Miller SA. 1880. Silurian ichnolites, with definitions of new genera and species. *Journal of the*
1069 *Cincinnati Society of Natural History* 2(1):217–222.
1070 Milner ARC, Birthisel TA, Kirkland JI, Breithaupt BH, Matthews NA, Lockley MG, Santucci
1071 VL, Gibson SZ, DeBlieux DD, Hurlbut M, Harris JD, Olsen PE. 2012. Tracking Early
1072 Jurassic dinosaurs across southwestern Utah and the Triassic–Jurassic transition. In:
1073 Bonde JW, Milner ARC, eds. *Field Trip Guide Book for the 71st Annual Meeting of the*
1074 *Society of Vertebrate Paleontology*. *Nevada State Museum Paleontological Papers* 1:1–
1075 107.
1076 Milner ARC, Harris JD, Lockley MG, Kirkland JI, Matthews NA. 2009. Bird-like anatomy,
1077 posture, and behavior revealed by an Early Jurassic theropod dinosaur resting trace. *PLoS*
1078 *ONE* 4:e4591. DOI: 10.1371/journal.pone.000459.
1079 Milner ARC, Lockley MG, Johnson SB. 2006. The story of the St. George Dinosaur Discovery
1080 Site at Johnson Farm: an important new Lower Jurassic dinosaur tracksite from the
1081 Moenave Formation of southwestern Utah. In: Harris JD, Lucas SG, Spielmann JA,
1082 Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial*
1083 *Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:329–345.

Milner ARC, Lockley MG, Kirkland JI. 2006. A large collection of well-preserved theropod dinosaur swim tracks from the Lower Jurassic Moenave Formation, St. George, Utah. In: Harris JD, Lucas SG, Spielmann JA, Lockley MG, Milner ARC, Kirkland JI, eds. *The Triassic–Jurassic Terrestrial Transition*. *New Mexico Museum of Nature and Science Bulletin* 37:315–328.

Minter NJ, Braddy SJ. 2009. Ichnology of an Early Permian intertidal flat: the Robledo Mountains Formation of southern New Mexico, USA. *Special Papers in Palaeontology* 82:1–107.

Minter NJ, Braddy SJ, Davis RB. 2007. Between a rock and a hard place: arthropod trackways and ichnotaxonomy. *Lethaia* 40:365–375. DOI: 10.1111/j.1502-3931.2007.00035.x.

Minter NJ, Buatois LA, Mángano MG, Davies NS, Gibling MR, Labandeira C. 2016b. The establishment of continental ecosystems. In: Mángano MG, Buatois LA, eds. *The Trace-Fossil Record of Major Evolutionary Events Vol. 1: Precambrian and Paleozoic*. Dordrecht: Springer Science+Business Media, 205–324. DOI: 10.1007/978-94-017-9600-2_6.

Minter NJ, Buatois LA, Mángano MG, MacNaughton RB, Davies NS, Gibling MR. 2016a. The prelude to continental invasion. In: Mángano MG, Buatois LA, eds. *The Trace-Fossil Record of Major Evolutionary Events Vol. 1: Precambrian and Paleozoic*. Dordrecht: Springer Science+Business Media, 157–204. DOI: 10.1007/978-94-017-9600-2_5.

Minter NJ, Krainer K, Lucas SG, Braddy SJ, Hunt AP. 2007. Palaeoecology of an Early Permian playa lake trace fossil assemblage from Castle Peak, Texas, USA. *Palaeogeography, Palaeoclimatology, Palaeoecology* 246:390–423. DOI: 10.1016/j.palaeo.2006.10.009.

Nopcsa FB. 1923. Die Familien der Reptilien. *Forschritte der Geologie und Paläontologie* 2:1–210.

O'Brien LJ, Braddy SJ, Radley JD. 2009. A new arthropod resting trace and associated suite of trace fossils from the Lower Jurassic of Warwickshire, England. *Palaeontology* 52:1099–1112. DOI: 10.1111/j.1475-4983.2009.00901.x.

Olsen PE, Huber P. 1997. Stop 3: Triangle Brick Quarry. In: Clark TW, ed. *TRIBI: Triassic Basin Initiative, Abstracts with Programs and Field Trip Guidebook*. Durham: Duke University, 22–29.

Osgood RG Jr. 1975. The paleontological significance of trace fossils. In: Frey RW, ed. *The Study of Trace Fossils*. Berlin: Springer, 87–108. DOI: 10.1007/978-3-642-65923-2_6.

Owen R. 1852. Description of the impressions and foot-prints of the *Protichnites* from the Potsdam Sandstone of Canada. *Quarterly Journal of the Geological Society* 8:214–225. DOI: 10.1144/GSL.JGS.1852.008.01-02.26.

Packard AS. 1900. On supposed merostomatous and other Paleozoic arthropod trails, with notes on those of *Limulus*. *Proceedings of the American Academy of Arts and Sciences* 36(4):63–71.

Parrish JT, Rasbury ET, Chan MA, Hasiotis ST. 2019. Earliest Jurassic U–Pb ages from carbonate deposits in the Navajo Sandstone, southeastern Utah, USA. *Geology* 47:1015–1019. DOI: 10.1130/G46338.1.

Pazos PJ. 2000. Trace fossils and facies in glacial to postglacial deposits from the Paganzo basin (Late Carboniferous), central Precordillera, Argentina. *Ameghiniana* 37:23–38.

Peixoto B de CPeM, Mángano MG, Minter NJ, dos Reis Fernandes LB, Fernandes MA. 2020. A new insect trackway from the Upper Jurassic–Lower Cretaceous eolian sandstones of São Paulo State, Brazil: implications for reconstructing desert paleoecology. *PeerJ* 8:ee8880. DOI: 10.7717/peerj.8880.

Pemberton SG, MacEachern JA, Gingras MK. 2007. The antecedents of invertebrate ichnology in North America: the Canadian and Cincinnati schools. In: Miller W III, ed. *Trace Fossils: Concepts, Problems, Prospects.* Amsterdam: Elsevier, 14–31.

Perrier V, Vannier J, Racheboeuf PR, Charbonnier S, Chabard D, Sotty D. 2006. Syncarid crustaceans from the Montceau Lagerstätte (Upper Carboniferous; France). *Palaeontology* 49:647–672. DOI: 10.1111/j.1475-4983.2006.00553.x.

Pirrie D, Feldmann RM, Buatois LA. 2004. A new decapod trackway from the Upper Cretaceous, James Ross Island, Antarctica. *Palaeontology* 47:1–12. DOI: 10.1111/j.0031-0239.2004.00343.x.

Pollard JE. 1985. *Isopodichnus*, related arthropod trace fossils and notostracans from Triassic fluvial sediments. *Transactions of the Royal Society of Edinburgh* 76:273–285. DOI: 10.1017/S026359330001049X.

Pollard JE. 1995. John Smith's discoveries of trace fossils from Old Red Sandstone and Carboniferous rocks of southwest Scotland. In: Ayrshire Archaeological and Natural History Society, ed. *John Smith of Dalry: Geologist, Antiquarian and Natural Historian Part 1—Geology.* Ayrshire: Walker & Connell, Ltd., 30–39.

Pollard J, Selden P, Watts S. 2008. Trace fossils of the arthropod *Camptophyllia* from the Westphalian (Carboniferous) of Lancashire, UK and their palaeoenvironmental context. *Palaeogeography, Palaeoclimatology, Palaeoecology* 270:399–406. DOI: 10.1016/j.palaeo.2008.01.032.

Pollard JE, Steel RJ, Undersrud E. 1982. Facies sequences and trace fossils in lacustrine/fan delta deposits, Hornelen Basin (M. Devonian), western Norway. *Sedimentary Geology* 32:63–87. DOI: 10.1016/0037-0738(82)90014-8.

Pollard JE, Walker EF. 1984. Reassessment of sediments and trace fossils from Old Red Sandstone (Lower Devonian) of Dunure, Scotland, described by John Smith (1909). *Geobios* 17:567–576. DOI:10.1016/S0016-6995(84)80029-7.

Porter ML, Pérez-Losada M, Crandall KA. 2005. Model-based multi-locus estimation of decapod phylogeny and divergence times. *Molecular Phylogenetics and Evolution* 37:355–369. DOI: 10.1016/j.ympev.2005.06.021.

Poschmann M, Braddy SJ. 2010. Eurypterid trackways from Early Devonian tidal facies of Alken an der Mosel (Rheinisches Schiefergebirge, Germany). *Palaeobiodiversity and Palaeoenvironments* 90:111–124. DOI: 10.1007/s12549-010-0024-2.

Rainforth E. 2005. Ichnotaxonomy of the Fossil Footprints of the Connecticut Valley (Early Jurassic, Newark Supergroup, Connecticut and Massachusetts). D. Phil. Dissertation, Columbia University.

Richter R. 1954. Fährte eines "Risenkrebses" im Rheinischen Schiefergebirge. *Natur und Volk* 84:261–269.

Rode AL, Babcock LE. 2003. Phylogeny of fossil and extant crayfish and some closely related nephropid lobsters. *Journal of Crustacean Biology* 23:418–435. DOI: 10.1163/20021975-99990351.

Sadler CJ. 1993. Arthropod trace fossils from the Permian De Chelly Sandstone, northeastern Arizona. *Journal of Paleontology* 67:240–249. DOI: 10.1017/S0022336000032169.

Santucci VL, Kirkland JI. 2010. An overview of National Park Service paleontological resources from the parks and monuments in Utah. In: Sprinkel DA, Chidsey TC Jr, Anderson PB, eds. *Geology of Utah's Parks and Monuments, 3rd Ed. Utah Geological Association Publication* 28:589–623.

Savage NM. 1971. A varvite ichnocoenosis from the Dwyka series of Natal. *Lethaia* 4:217–233. DOI: 10.1111/j.1502-3931.1971.tb01290.x.

Schindewolf OH. 1928. Studien aus dem Marburger Buntsandstein 4. *Isopodichnus problematicus* (Schindewolf) in Unteren und Mittleren Buntsandstein. *Senckenbergiana* 10:27–37.

Schram FR. 2001. Phylogeny of decapods: moving towards a consensus. *Hydrobiologia* 449:1–20. DOI: 10.1023/a:1017543712119.

Schram FR, Dixon CJ. 2004. Decapod phylogeny: addition of fossil evidence to a robust morphological cladistic data set. *Bulletin of the Mizunami Fossil Museum* 31:1–19.

Schweitzer CE, Feldmann RM, Garassino A, Karasawa H, Schweiger G. 2010. *Systematic List of Fossil Decapod Crustacean Species*. Leiden: Brill NV. DOI: 10.1163/ej.9789004178915.i-222.

Seilacher A. 2007. *Trace Fossil Analysis.* Berlin: Springer. DOI: 10.1007/978-3-540-47226-1.

Seilacher A. 2008. Biomats, biofilms, and bioglue as preservational agents for arthropod trackways. *Palaeogeography, Palaeoclimatology, Palaeoecology* 270:252–257. DOI: 10.1016/j.palaeo.2008.07.011.

Selden PA, Nudds JR. 2012. *Evolution of Fossil Ecosystems, 2nd Ed.* Amsterdam: Elsevier.

Shen H, Braband A, Scholtz, G. 2015. The complete mitogenomes of lobsters and crayfish (Crustacea: Decapoda: Astacidea) reveal surprising differences in closely related taxa and convergences to Priapulida. *Journal of Zoological Systematics and Evolutionary Research* 53:273–281. DOI: 10.1111/jzs.12106.

1198 Shen Y-B. 2003. Shrimps. In: Chang M-M, Chen P-J, Wang Y-Q, Wang Y, Miao D-S, eds. *The*
1199 *Jehol Biota: The Emergence of Feathered Dinosaurs, Beaked Birds and Flowering*
1200 *Plants.* Shanghai: Shanghai Scientific & Technical Press, 53–57.
1201 Shillito AP, Davies NS. 2018. Death near the shoreline, not life on land: Ordovician arthropod
1202 trackways in the Borrowdale Volcanic Group, UK. *Geology* 47:55–58. DOI:
1203 10.1130/G45663.1.
1204 Shu W, Tong J, Tian L, Benton MJ, Chu D, Yu J, Guo W. 2018. Limuloid trackways from
1205 Permian–Triassic continental successions of North China. *Palaeogeography,*
1206 *Palaeoclimatology, Palaeoecology* 508:71–90. DOI: 10.1016/j.palaeo.2018.07.022.
1207 Smith A, Braddy SJ, Marriott SB, Briggs DEG. 2003. Arthropod trackways from the Early
1208 Devonian of South Wales: a functional analysis of producers and their behaviour.
1209 *Geological Magazine* 140:63–72. DOI: 10.1017/S0016756802006982.
1210 Smith DM. 2012. Exceptional preservation of insects in lacustrine environments. *Palaios*
1211 27:346–353. DOI: 10.2110/palo.2011.p11-107r.
1212 Smith J. 1909. *Upland Fauna of the Old Red Sandstone Formation of Carrick, Ayrshire.*
1213 Kilwinning: A.W. Cross.
1214 Steiner M. 2014a. Age of Lower Jurassic Springdale Sandstone of southwestern Utah from its
1215 magnetic polarity sequence. *Volumina Jurassica* 12(2):23–30. DOI:
1216 10.5604/17313708.1130124.
1217 Steiner M. 2014b. New magnetostratigraphy and paleopole from the Whitmore Point Member of
1218 the Moenave Formation at Kanab, Utah. *Volumina Jurassica* 12(2):13–22. DOI:
1219 10.5604/17313708.1130122.
1220 Stern D, Crandall KA. 2015. Phylogenetic estimate of the freshwater crayfish (Decapoda:
1221 Astacidea) using morphology and molecules. In: Kawai T, Faulkes Z, Scholtz G, eds.
1222 *Freshwater Crayfish: A Global Overview.* Boca Raton: CRC Press, 298–309.
1223 Stimson M, MacRae A, Calder J, Hebert B, Reid D, Reid L. 2018. The *Diplichnites aenigma*
1224 enigma: ichnotaxonomic implications of a restudy of Dawson's type locality at Coal
1225 Mine Point, Joggins, Nova Scotia. *1st Joggins Research Symposium, Joggins Fossil*
1226 *Institute, Abstracts. Atlantic Geology* 54:487. DOI: 10.4138/atlgeol.2018.016.
1227 Suarez CA, Knobbe TK, Crowley JL, Kirkland JI, Milner ARC. 2017. A chronostratigraphic
1228 assessment of the Moenave Formation, USA using C-isotope chemostratigraphy and
1229 detrital zircon geochronology: implications for the terrestrial end Triassic extinction.
1230 *Earth and Planetary Science Letters* 475:83–93. DOI: 10.1016/j.epsl.2017.07.028.
1231 Tanner LH, Lucas SG. 2009. The Whitmore Point Member of the Moenave Formation: Early
1232 Jurassic dryland lakes on the Colorado Plateau, southwestern USA. *Volumina Jurassica*
1233 6(1):11–21.
1234 Tasch P. 1969. New observations on the notostracan *Triops longicaudatus* in natural and cultural
1235 situations. *Transactions of the Kansas Academy of Science* 72:323–329.

1236 Taylor RS, Schram FR, Shen Y-B. 1999. A new crayfish family (Decapoda: Astacida) from the
1237 Upper Jurassic of China, with a reinterpretation of other Chinese crayfish taxa.
1238 *Paleontological Research* 3:121–136. DOI: 10.2517/prpsj.3.121.
1239 Trewin NH, McNamara KJ. 1994. Arthropods invade the land: trace fossils and
1240 palaeoenvironments of the Tumblagooda Sandstone (?Late Silurian) of Kalbarri, western
1241 Australia. *Transactions of the Royal Society of Edinburgh: Earth Sciences* 85:177–210.
1242 DOI: 10.1017/S026359330000359X.
1243 Trusheim F. 1931. Aktuo-paläontologische Beobachtungen an *Triops cancriformis* Schaeffer
1244 (Crust. Phyll.). *Senckenbergiana* 13:234–243.
1245 Uchman A, Gaździcki A, Błażejowski B. 2018. Arthropod trace fossils from Eocene cold climate
1246 continental strata of King George Island, West Antarctica. *Acta Palaeontologica*
1247 *Polonica* 63:383–396. DOI: 10.4202/app.00467.2018.
1248 Uchman A, Hu B, Wang Y, Song H. 2011. The trace fossil *Diplopodichnus* from the Lower
1249 Jurassic lacustrine sediments of central China and the isopod *Armadillidium vulgare*
1250 (pillbug) Lebensspuren as its Recent analogue. *Ichnos* 18:147–155. DOI:
1251 10.1080/10420940.2011.601372.
1252 Uchman A, Kazakauskas V, Gaigalas A. 2009. Trace fossils from Late Pleistocene varved
1253 lacustrine sediments in eastern Lithuania. *Palaeogeography, Palaeoclimatology,*
1254 *Palaeoecology* 272:199–211. DOI: 10.1016/j.palaeo.2008.08.003.
1255 Urreta MBA. 1989. The Cretaceous decapod Crustacea of Argentina and the Antarctic Peninsula.
1256 *Palaeontology* 32:499–552.
1257 Viohl G. 1998. Die Solnhofener Plattenkalke—Entstehung und Lebensräume. *Archaeopteryx*
1258 16:37–68.
1259 Walker EF. 1985. Arthropod ichnofauna of the Old Red Sandstone at Dunure and Montrose,
1260 Scotland. *Transactions of the Royal Society of Edinburgh: Earth Sciences* 76:287–297.
1261 DOI: 10.1017/S0263593300010506.
1262 Walker SE, Holland SM, Gardiner L. 2003. *Coenobichnus currani* (new ichnogenus and
1263 ichnospecies): fossil trackway of a land hermit crab, early Holocene, San Salvador,
1264 Bahamas. *Journal of Paleontology* 77:576–582. DOI: 10.1017/S0022336000044255.
1265 Walter H. 1985. Zur Ichnologie des Pleistozäns von Liebegast. *Freiberger Forschungshefte C*
1266 400:101–116.
1267 Webby BD. 1983. Lower Ordovician arthropod trace fossils from western New South Wales.
1268 *Proceedings of the Linnean Society of New South Wales* 107:61–76.
1269 Wolfe JM, Breinholt JW, Crandall KA, Lemmon AR, Lemmon EM, Timm LE, Siddall ME,
1270 Bracken-Grissom HD. 2019. A phylogenomic framework, evolutionary timeline and
1271 genomic resources for comparative studies of decapod crustaceans. *Proceedings of the*
1272 *Royal Society of London B* 286:20190079. DOI: 10.1098/rspb.2019.0079.

FIGURE CAPTIONS

Figure 1: Location of the St. George Dinosaur Discovery Site at Johnson Farm. (A) Map showing the location of the St. George Dinosaur Discovery Site in St. George, Utah. (B) The museum at the St. George Dinosaur Discovery Site. Photograph by Jerald D. Harris.

Figure 2: Stratigraphic section at and immediately around the St. George Dinosaur Discovery Site (SGDS) in St. George, Utah. Arthropod locomotory trackway SGDS 1290 comes from the "Top Surface Tracksite" layers of the Johnson Farm Sandstone Bed (red arrow).

Figure 3: Arthropod locomotory trace fossil SGDS 1290. (A) Photograph of specimen; scale in mm. (B) Schematic diagram of specimen. Arrow indicates direction of movement. L = left track; R = right track; Ov = overtracks. Numbers indicate position of tracks in sequence from caudal (posterior) to cranial (anterior). Photograph by Andrew R.C. Milner.

Figure 4: Schematic diagrams of locomotory (presumably walking) trace fossils attributed to arthropods (A–KK) and extant walking traces made by arthropods (LL–MM) in comparison to SGDS 1290 (NN). Diagrams not to scale. (A) *Acanthichnus cursorius* (traced from Hitchcock, 1858). (B) *Asaphoidichnus trifidus* (traced from Miller, 1880). (C) *Bifurculapes laqueatus* (traced from Getty, 2016). (D) *Coenobichnus currani* (traced from Walker, Holland & Gardiner, 2003). (E) *Copeza triremis* (traced from Hitchcock, 1858). (F) *Danstairia congesta* (traced from Walker, 1985). (G) *Diplichnites aenigma* (traced from Dawson, 1873). (H) *Diplichnites gouldi* Type A (traced from Trewin and McNamara, 1994). (I) *Eisenachichnus inaequalis* (traced from Kozur, 1981). (J) *Foersterichnus rossensis* (traced from Pirrie, Feldmann & Buatois, 2004). (K) *Hamipes didactylus* (traced from Getty, 2018). (L) *Lithographus hieroglyphicus* (traced from Hitchcock, 1858). (M) *Maculichna varia* (traced from Anderson, 1975a). (N) *Merostomichnites narrangansettensis* (traced from Packard, 1900). (O) *Mirandaichnium famatinense* (traced from Aceñolaza, 1978). (P) *Octopodichnus didactylus* (traced from Gilmore, 1927). (Q) *Petalichnus multipartatus* (Miller, 1880). (R) *Pterichnus centipes* (traced from Hitchcock, 1865). (S) *Tasmanadia twelvetreesi* (traced from Glaessner, 1957). (T) *Umfolozia sinuosa* (traced from Anderson, 1981). (U) *Kouphichnium lithographicum* (traced from Gaillard, 2011). (V) "*Merostomichnites* isp." (traced from Hanken & Stormer, 1975). (W) *Oniscoidichnus filiciformis* (traced from Brady, 1947). (X) *Palmichnium antarcticum* (traced from Braddy & Milner, 1998). (Y) *Paleohelcura tridactyla* (traced from Gilmore, 1926). (Z) *Protichnites septemnotatus* (traced from Owen, 1852). (AA) *Robledoichnus lucasi* (traced from Kozur & Lemone, 1995). (BB) *Shalemichnus sittigi*, half of trackway (traced from Kozur & Lemone, 1995). (CC) *Stiallia berriana* (traced from Smith, 1909). (DD) *Stiaria quadripedia* (traced from Walker, 1985). (EE) *Mitchellichnus ferrydenensis* (traced from Walker, 1985). (FF) *Keircalia multipedia* (traced from Walker, 1985). (GG) *Danstairia vagusa* (traced from

Walker, 1985). (HH) *Glaciichnium liebegastensis* (traced from Walter, 1985). (II) *Warvichnium ulbrichi* (traced from Walter, 1985). (JJ) *Siskemia bipediculus* (traced from Walker, 1985). (KK) *Siskemia elegans* (traced from Walker, 1985). (LL) Extant notostracan traces (traced from Knecht et al., 2009). (MM) Extant crayfish traces in very fine sand (traced from Fairchild & Hasiotis, 2011). (NN) SGDS 1290.

TABLE CAPTIONS

Table S1: Measurements (in mm) of arthropod locomotory trace fossil SGDS 1290.
Measurements in parentheses are approximated based on faint portions of paramedial impressions.

Figure 1

Location of the St. George Dinosaur Discovery Site at Johnson Farm.

(A) Map showing the location of the St. George Dinosaur Discovery Site in St. George, Utah.

(B) The museum at the St. George Dinosaur Discovery Site. Photograph by Jerald D. Harris.

Figure 2

Stratigraphic section at and immediately around the St. George Dinosaur Discovery Site (SGDS) in St. George, Utah.

Arthropod locomotory trackway SGDS 1290 comes from the “Top Surface Tracksite” layers of the Johnson Farm Sandstone Bed (red arrow).

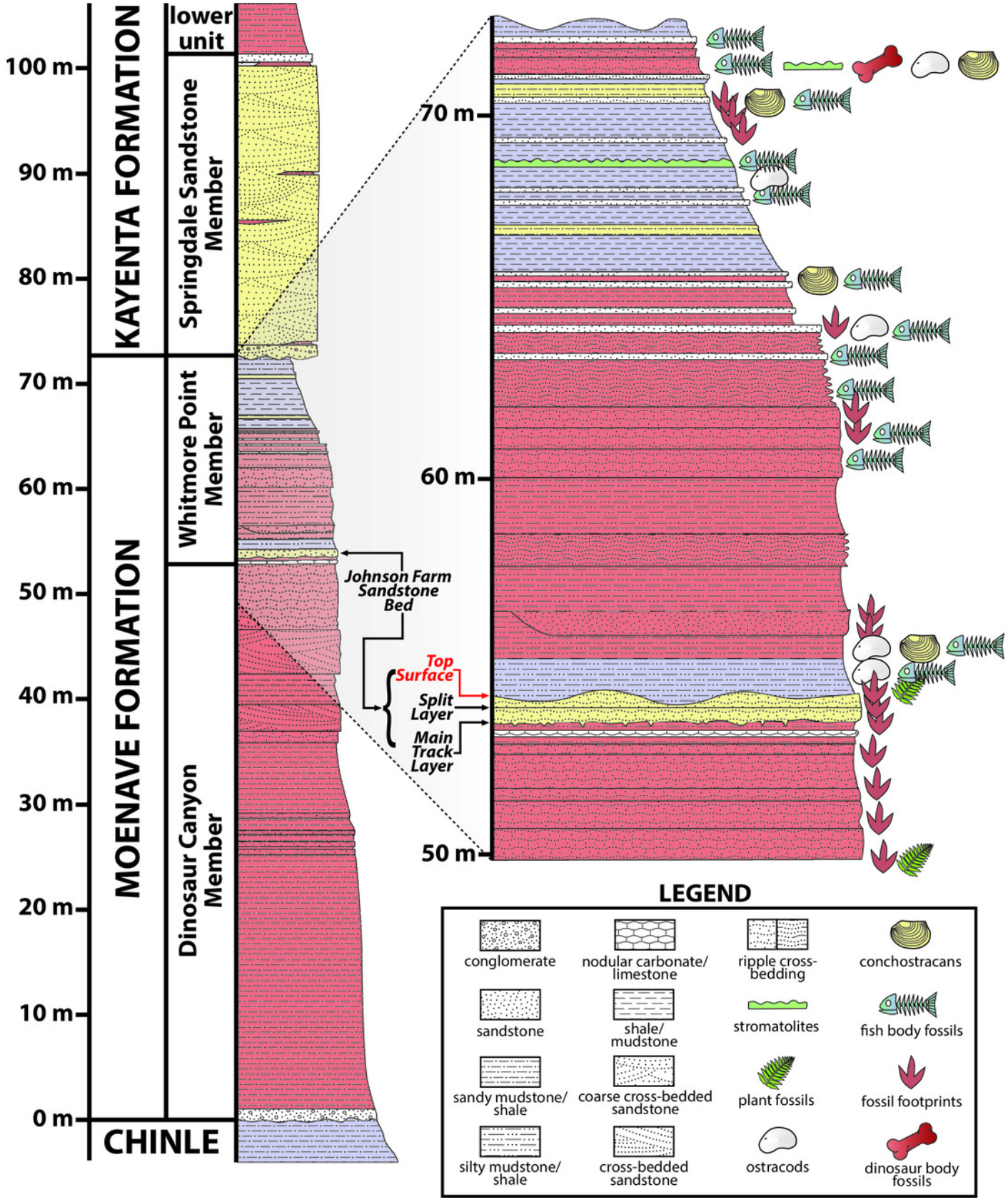
KAYENTA FORMATION
lower unit
Springdale Sandstone Member
MOENAVE FORMATION
Whitmore Point Member
Dinosaur Canyon Member
CHINLE
100 m
90 m
80 m
70 m
60 m
50 m
40 m
30 m
20 m
10 m
0 m
Johnson Farm Sandstone Bed
Top Surface
Split Layer
Main Track Layer
LEGEND
conglomerate
nodular carbonate/ limestone
ripple cross-bedding
conchostracans
sandstone
shale/ mudstone
stromatolites
fish body fossils
sandy mudstone/ shale
coarse cross-bedded sandstone
plant fossils
fossil footprints
silty mudstone/ shale
cross-bedded sandstone
ostracods
dinosaur body fossils

Figure 3

Arthropod locomotory trace fossil SGDS 1290.

(A) Photograph of specimen; scale in mm. (B) Schematic diagram of specimen. Arrow indicates direction of movement. L = left track; R = right track; Ov = overtracks. Numbers indicate position of tracks in sequence from caudal (posterior) to cranial (anterior). Photograph by Andrew R.C. Milner.

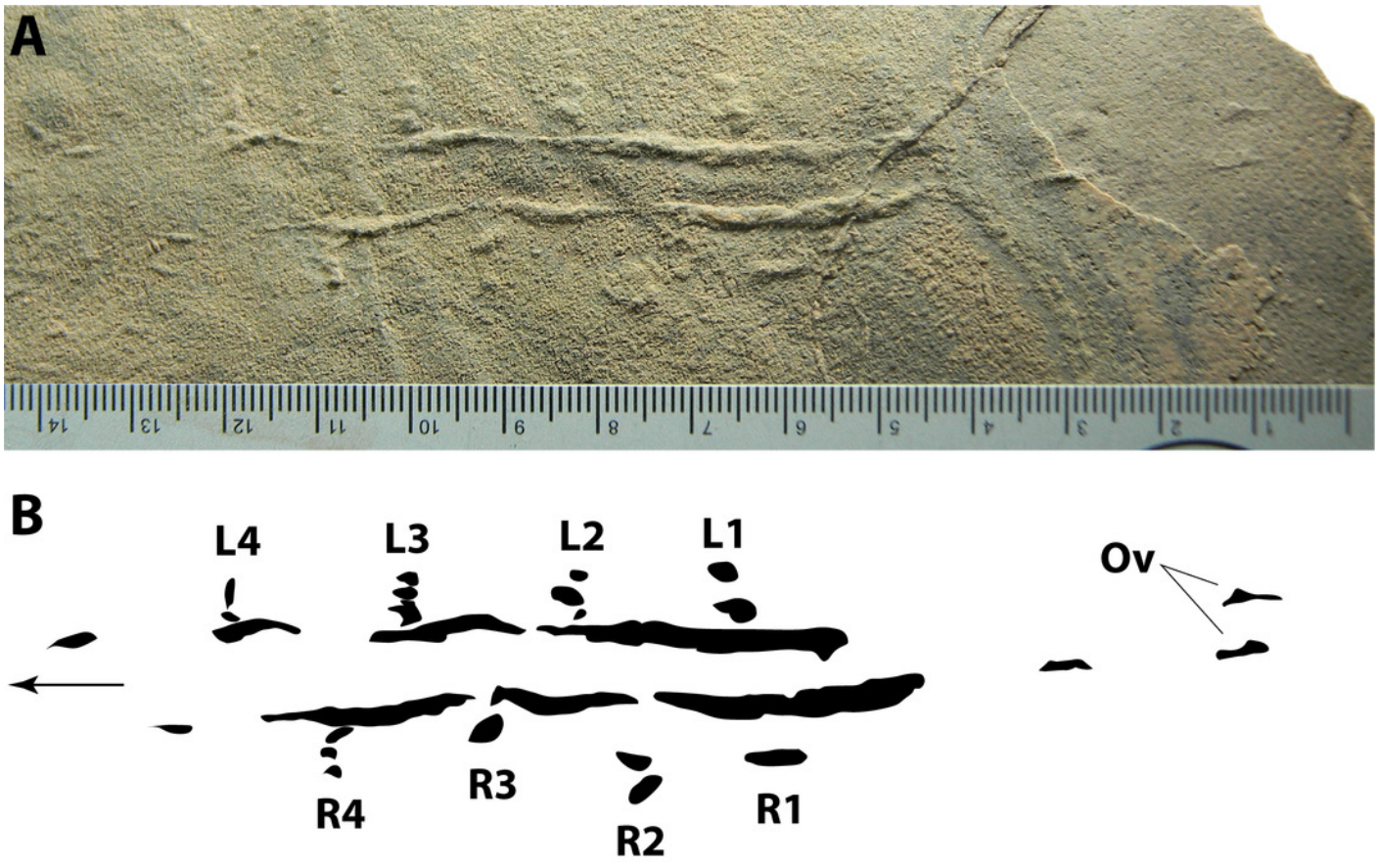

Figure 4

Schematic diagrams of locomotory (presumably walking) trace fossils attributed to arthropods (A–KK) and extant walking traces made by arthropods (LL–MM) in comparison to SGDS 1290 (NN).

Diagrams not to scale. (A) *Acanthichnus cursorius* (traced from Hitchcock, 1858). (B) *Asaphoidichnus trifidus* (traced from Miller, 1880). (C) *Bifurculapes laqueatus* (traced from Getty, 2016). (D) *Coenobichnus currani* (traced from Walker, Holland & Gardiner, 2003). (E) *Copeza triremis* (traced from Hitchcock, 1858). (F) *Danstairia congesta* (traced from Walker, 1985). (G) *Diplichnites aenigma* (traced from Dawson, 1873). (H) *Diplichnites gouldi* Type A (traced from Trewin and McNamara, 1994). (I) *Eisenachichnus inaequalis* (traced from Kozur, 1981). (J) *Foersterichnus rossensis* (traced from Pirrie, Feldmann & Buatois, 2004). (K) *Hamipes didactylus* (traced from Getty, 2018). (L) *Lithographus hieroglyphicus* (traced from Hitchcock, 1858). (M) *Maculichna varia* (traced from Anderson, 1975a). (N) *Merostomichnites narrangansettensis* (traced from Packard, 1900). (O) *Mirandaichnium famatinense* (traced from Aceñolaza, 1978). (P) *Octopodichnus didactylus* (traced from Gilmore, 1927). (Q) *Petalichnus multipartatus* (Miller, 1880). (R) *Pterichnus centipes* (traced from Hitchcock, 1865). (S) *Tasmanadia twelvetreesi* (traced from Glaessner, 1957). (T) *Umfolozia sinuosa* (traced from Anderson, 1981). (U) *Kouphichnium lithographicum* (traced from Gaillard, 2011). (V) "*Merostomichnites* isp." (traced from Hanken & Stormer, 1975). (W) *Oniscoidichnus filiciformis* (traced from Brady, 1947). (X) *Palmichnium antarcticum* (traced from Braddy & Milner, 1998). (Y) *Paleohelcura tridactyla* (traced from Gilmore, 1926). (Z) *Protichnites septemnotatus* (traced from Owen, 1852). (AA) *Robledoichnus lucasi* (traced from Kozur & Lemone, 1995). (BB) *Shalemichnus sittigi*, half of trackway (traced from Kozur & Lemone, 1995). (CC) *Stiallia berriana* (traced from Smith, 1909). (DD) *Stiaria quadripedia* (traced from Walker, 1985). (EE) *Mitchellichnus ferrydenensis* (traced from Walker, 1985). (FF) *Keircalia*

multipedia (traced from Walker, 1985). (GG) *Danstairia vagusa* (traced from Walker, 1985). (HH) *Glaciichnium liebegastensis* (traced from Walter, 1985). (II) *Warvichnium ulbrichi* (traced from Walter, 1985). (JJ) *Siskemia bipediculus* (traced from Walker, 1985). (KK) *Siskemia elegans* (traced from Walker, 1985). (LL) Extant notostracan traces (traced from Knecht et al., 2009). (MM) Extant crayfish traces in very fine sand (traced from Fairchild & Hasiotis, 2011). (NN) SGDS 1290.

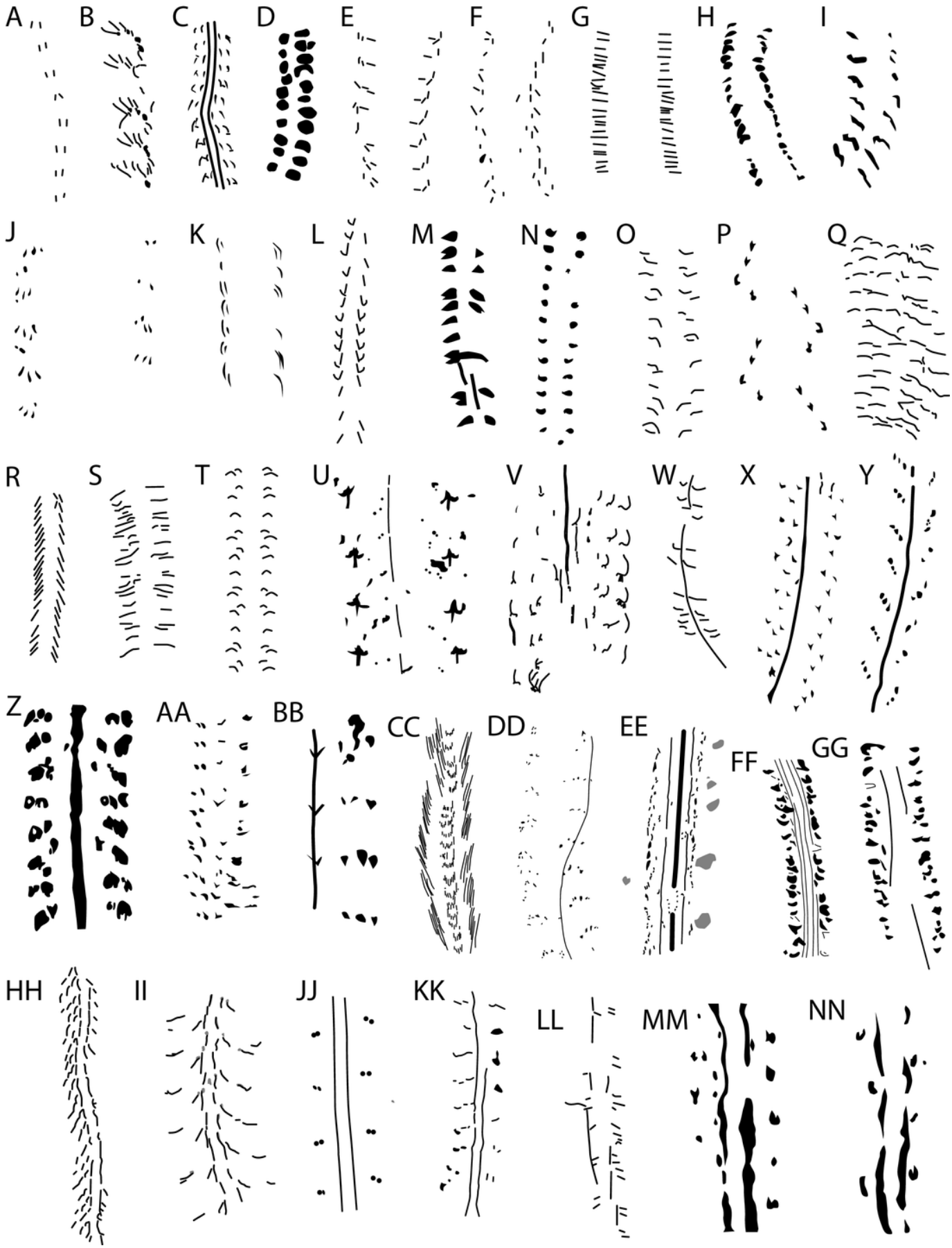
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Table 1(on next page)

Measurements (in mm) of arthropod locomotory trace fossil SGDS 1290.

Measurements in parentheses are approximated based on faint portions of paramedial impressions.

Table 1: Measurements (in mm) of arthropod locomotory trace fossil SGDS 1290.
Measurements in parentheses are approximated based on faint portions of paramedial impressions.

Track Position	Length	Width	Impression Internal Width	Impression External Width	Track to Impression Distance	Left Impression Width	Right Impression Width
L1	4.3	6.4	4.7	9.1	1.8	3.1	2.5
L2	4.2	5.7	5.9	9.5	1.5	2.6	2.2
L3	5.2	5.8	5.8	9.7	0.0	1.6	1.6
L4	3.2	5.6	(8.1)	(11.2)	0.0	1.9	0.0
R1	6.8	2.5	4.2	9.2	3.0	3.0	2.4
R2	4.6	5.3	5.4	8.8	4.2	2.7	0.0
R3	4.5	6.4	6.5	9.2	2.4	1.3	1.3
R4	3.0	5.8	6.9	10.6	0.0	0.0	3.0