

## **Evaluation of resistance to wheat stem rust and identification of resistance genes in wheat lines**

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### **Abstract**

Wheat stem rust, caused by *Puccinia graminis* f. sp. *tritici*, (*Pgt*) is a devastating disease in wheat production. The disease has been effectively controlled since the 1970s due to the widespread use of the *Sr31* resistance gene. However, *Sr31* has lost its effectiveness following the emergence and spread of the Ug99 race variants. Therefore, there is an urgent global effort to identify new germplasm resources effective against those races. In this study, the resistance to *Pgt* of 95 wheat advance lines from Heilongjiang Province was evaluated using three predominant races of *Pgt*, 21C3CTTTM, 34C0MKGSM, and 34C3MTGQM, in China at the seedling and adult plant stage. The presence of 6 *Sr* genes (*Sr2*, *Sr24*, *Sr25*, *Sr26*, *Sr31*, and *Sr38*) was detected using linked molecular markers. The results showed that 86 (90.5%) wheat lines had plant stage resistance to all three races. Molecular marker analysis showed that 24 wheat lines likely carried *Sr38*, 15 wheat lines likely carried *Sr2*, 11 wheat lines likely carried *Sr31*, while none of the wheat lines carried *Sr24*, *Sr25*, or *Sr26*. Furthermore, 6 out of the 95 cultivars tested carried both *Sr2* and *Sr38*, three contained both *Sr31* and *Sr38*, and two wheat lines contained both *Sr2* and *Sr31*. Wheat lines with

known *Sr* genes could be used as donor parents for further breeding programs to provide resistance to stem rust.

**Keywords:** *Puccinia graminis* f. sp. *tritici*, wheat stem rust, resistance genes, molecular marker

## Background

Wheat is the most important cereal grain in the world, contributing 20% of human caloric intake. Although more than 700 million tons of wheat are produced every year, food shortage has become a global problem, due to the rapid growth of the world population (FAO, 2017a). In addition, the yield loss in the production of wheat caused by various wheat pathogens, including the wheat stem rust-causing fungus *Puccinia graminis* f. sp. *tritici* (*Pgt*), accelerated this trend. Over the past 30 years, there has been no large-scale epidemic of wheat stem rust in China, because a large number of cultivars with stem rust resistance genes have been cultivated and popularized, which has played a vital role in controlling this disease (Li *et al.*, 2016). In addition, since the 1980s changes in crop layouts and in cultivation systems in the main overwintering regions that provide initial urediniospores of wheat stem rust (such as Fujian, Guangdong, and other provinces) have played a crucial role in controlling the overwintering of initial pathogens (Cao *et al.*, 2016). Nevertheless, wheat stem rust remains a long-standing threat to global wheat production security because of variations in the virulence of the *Pgt* population, and the ability of urediniospores to spread over long distances by wind (Singh *et al.*, 2015). For example, the emergence of a new virulent race TTKSK (the well-known Ug99), was first identified in Uganda in 1999, and in just a few years its mutant isolates acquired virulence to the important stem rust resistance genes or combinations thereof (such as *Sr24*, *Sr36*, *Sr38*, *Sr31+Sr24*, *Sr31+Sr36*, and *Sr31+SrTmp*), and caused an epidemic of wheat stem rust in 13 countries (FAO, 2017b; Singh *et al.*, 2011). Although the

International Maize and Wheat Improvement Center (CIMMYT) and the International Center for Agricultural Research in the [Dream-Dry](#) Areas (ICARDA) have established a global rust initiative to track and study Ug99 in order to prevent and control the disease on a global scale, in recent years new races of *Pgt* have emerged (e.g., TKTTF, TTTTF) causing a pandemic and stem rust is once again threatening worldwide wheat production (Bhattacharya 2017; Olivera *et al.*, 2015).

Resistance breeding is the most effective, economical, and environmentally friendly strategy to control wheat stem rust. To date, more than 60 *Sr* genes have been identified in wheat and its wild relatives (McIntosh 2016). While most confer race-specific resistance, some confer [race](#) non-specific resistance (*Sr57*, *Sr58*, *Sr55*, and *Sr2*) (Juliana *et al.*, 2017) or high-temperature adult-plant resistance (*Sr13*, *Sr21*) (Chen *et al.*, 2018; Zhang *et al.*, 2017). Therefore, it is of great significance to identify which resistance genes are present in which wheat cultivars and lines in order to guide wheat resistance breeding and rational layout of resistance cultivars, avoiding the large-scale application of a single resistance gene and reducing the selection pressure of wheat cultivars to *Pgt*. The traditional identification approach of genes is gene postulation according to the infection types (ITs) of the different stem rust resistance genes to known *Pgt* races. This strategy is easily affected by environmental conditions and is time-consuming, laborious, and complex (Goutam *et al.*, 2015). In recent years, molecular marker technology has provided a new perspective on wheat disease management and has played an important role in molecular marker-assisted selection (MAS) breeding. One of the most important benefits of this technology is that markers are highly heritable and can be screened at the seedling stage. Due to the emergence and spread of Ug99, the mapping and application of molecular markers that were intricately linked with resistance genes of wheat stem rust have accelerated. So far, many molecular markers have been reported that

are closely linked to wheat stem rust resistance genes (Goutam *et al.*, 2015) many of which have been transformed into Simple Sequence Repeat (SSR), Sequence Characterized Amplified Region (SCAR), and Sequence-Tagged Site (STS) markers that are widely used in wheat disease resistance molecular marker selection and breeding. ~~For example,~~ Haile *et al.* (2013) used SSR and STS markers of 30 single *Sr* genes to detect 58 tetraploid wheat in Ethiopia. The Ug99 resistant genes *Sr2*, *Sr22*, *Sr24*, *Sr36*, and *Sr46* were identified using markers linked with those genes in 99 Kazakh spring wheat (Kokhmetova and Atishova 2012); Mourad *et al.* (2019) have confirmed the presence of stem rust resistance genes *Sr6*, *Sr31*, *Sr1RS*, *Sr24*, *Sr36*, *SrTmp*, *Sr7b*, *Sr9b*, and *Sr38* using gene-specific markers in Nebraska bread wheat germplasm. Wu *et al.*, (2014) screened 139 Chinese wheat cultivars using markers linked with the Ug99 resistance genes *Sr22*, *Sr25*, *Sr26*, and *Sr28*. Xu *et al.* (2017) detected the resistance genes *Sr2*, *Sr24*, *Sr25*, *Sr26*, *Sr31*, and *Sr38* in 75 wheat cultivars in Gansu Province. Therefore, MAS is extremely helpful in identifying the tagged resistance genes which have been pyramided in one genotype.

Northeast China used to be an area of frequent occurrence of wheat stem rust, playing a key role in the large-scale epidemic of the disease. In history, there have been 9 pandemics in this area, and some years this even resulted in an almost total grain failure (Wu and Huang 1987). In recent years, with the adjustment of agricultural structure, wheat production has mainly been distributed in Heilongjiang Province, and the annual planting area is nearly 300 thousand hectares. With the recent outbreak of wheat stem rust around the world, it is of great urgency to evaluate the resistance of wheat cultivars to *Pgt* and to clarify detailed knowledge of resistance genes present in wheat cultivars or lines. Therefore, we previously determined the level of resistance to *Pgt* of the 83 main production cultivars and the prevalence of *Sr2*, *Sr24*, *Sr25*, *Sr26*, *Sr31*, and *Sr38* in this region (Li *et*

Commented [SJ-A1]: 31 markers linked to Sr genes

al., 2016; Xu *et al.*, 2017). Based on these studies, we collected 95 wheat advanced lines to characterize the seedling and adult resistance level to *Pgt*, and to identify the presence of *Sr* genes in those wheat lines using molecular markers. The results of our work will be important for developing potentially durable combinations of effective stem rust resistance genes in wheat cultivars.

## **Materials and Methods**

### **Plant and fungal materials**

A total of 95 advance wheat lines were collected from Heilongjiang Academy of Agricultural Sciences (Harbin, Jiusan, Hongxinglong, Heihe, Jiamusi) and Heilongjiang Bayi Agricultural University, covering most important wheat-producing regions. Thirty-six monogenic lines with known stem rust resistance (*Sr*) genes, which were used in our study to test the virulence spectrum of *Pgt* and confirm the validity of these molecular markers, were provided by the Institute of Plant Immunity, Shenyang Agricultural University. The cultivar LC was used as a universal susceptible control. All wheat lines have not been deposited in a publicly available herbarium. Three races (21C3CTTM, 34C0MKGSM, and 34C3MTGQM) of *Pgt* with different virulence spectrum (STable 1), which were used to evaluate the resistance level of the advance wheat lines to *Pgt*, were identified using an international system of nomenclature for *Pgt* by the Institute of Plant Immunity, Shenyang Agricultural University.

### **Seedling infection type (IT) assays**

Xianxin Wu undertook the formal identification of the plant material used in your study. Seedling infection type assays were conducted in duplicate in a greenhouse. The wheat lines were planted in a 12 cm diameter clay pot. The seeding infection type assays were carried out when the wheat

seedlings grew to the two-leaf stage (one leaf and one sprout). First, the leaves were sprayed with a 0.05% Tween 20 solution using a handheld atomizer to form a water film on the leaves. Then, fresh urediniospores (1 g) and dried talc, mixed in a ratio of 1:20 (w/w), were inoculated on the seedlings. Following hydration in the dark for 16 hours at 18 to 20°C, the inoculated seedlings were transferred to a glass greenhouse with a temperature of  $20 \pm 1^\circ\text{C}$ . When the universal susceptible control wheat line LC was fully infected (14 days after inoculation), the seedling ITs were investigated and recorded according to the 0-4 scale described by Stakman *et al.* (1962). According to this scale, 0 - 2 was classified as low infection type (resistant) while 3 and 4 were classified as high infection type (susceptible).

#### **Field stem rust evaluation**

Resistance in adult plants was measured in 2016 and 2017 at the experimental site of the College of Plant Protection, Shenyang Agricultural University (latitude  $41^\circ49'\text{N}$ , longitude  $123^\circ33'\text{E}$ , altitude 67 m). Seeds of each cultivar (line) were planted in double 1 m-rows, spaced 25 cm apart. The susceptibility control, LC, was planted perpendicular to all wheat cultivars (monogenic lines) between the double 1 m-rows. The various lines of wheat were inoculated at the green-and-jointing stage. Watering was achieved through sprinkling irrigation to ensure the soil was fully humid prior to inoculation, which was conducted in the evening. After spraying the leaves with a 0.05% Tween-20 aqueous solution, diluted urediniospores (urediniospores to talcum powder = 1:30 (w/w)) were sprayed as a powder onto the leaves for inoculation. A plastic cover maintained the moisture for 12-14 hours. The IRs were assessed as immune ('I'), resistant ('R'), moderately resistant ('MR'), moderately susceptible ('MS'), or fully susceptible ('S'). Stem rust severity was assessed using a modified Cobb scale as described by Roelfs *et al.* (1992). When the LC was fully infected (14 days

after inoculation), the first disease assessment was conducted which was repeated every 3 days. The highest IR and severity for each wheat variety was recorded.

### **Polymerase chain reaction (PCR) amplification**

DNA was extracted from the young leaves of 10-day old seedlings grown to the one-leaf stage, using a DNA extraction kit (<http://www.sangon.com>; China). Polymerase chain reactions (PCR) were carried out using an S1000™ Thermal Cycler in a volume of 25  $\mu\text{L}$ , including 2  $\mu\text{L}$  of 50  $\text{ng}\cdot\mu\text{L}^{-1}$  DNA, 1  $\mu\text{L}$  of each primer ( $10\ \mu\text{mol}\cdot\text{L}^{-1}$ ), 2.5  $\mu\text{L}$  of  $10\times$  buffer (including  $\text{Mg}^{2+}$  at a final concentration of 2.5 mM), 0.2  $\mu\text{L}$  of *Taq* polymerase ( $5\ \text{U}\cdot\mu\text{L}^{-1}$ ), and 0.5  $\mu\text{L}$  of deoxyribonucleoside triphosphates ( $10\ \text{mmol}\cdot\text{L}^{-1}$  each). PCR amplifications were done as previously reported (Xu *et al.*, 2017). Six markers were used in this study, and their effectiveness was confirmed using 36 monogenic lines with known *Sr* genes (Supplemental Table 1). Primers were synthesized by Sangon Biotech (China) (Table 4), and PCR amplification conditions were as described in previous studies (Xu *et al.*, 2018). Fragments of the targeted genes were separated by electrophoresis using 2% (w/v) agarose gels, stained with ethidium bromide, and observed under UV light.

## **Results**

### **Evaluation of wheat lines for stem rust resistance at the seedling stage**

The ITs of 95 main wheat advance lines in Heilongjiang to the *Pgt* races 21C3CTTMM, 34C0MKGSM, and 34C3MTGQM at seedling stage are shown in Table 1. Nine wheat cultivars were susceptible (ITs 3-4) to all tested isolates at the seedling stage, accounting for 9.4% of the tested lines (Table 2). The remaining 86 (90.5%) wheat cultivars were resistant to all tested isolates.

### **Evaluation of wheat lines for stem rust resistance at the adult plant stage**

The infection responses (IRs) of 95 wheat lines to all tested isolates at the adult plant stage were determined during the 2017 and 2018 cropping seasons, and are presented in Table 1. Based on the IRs, the tested wheat lines were classified into three groups. The first group (I) contained 21 (22.1%) wheat lines immune to all tested isolates displaying no visible symptoms (IT: 0) in two seasons. In the second group, 65 (68.4%) wheat lines showed MR-R (IT: 1, 1-, 1, 1+) with severity between 5%-50% to all tested isolates. In the third group, the remaining 9 (9.5%) wheat lines showed MS-S (IT: 3-, 3, 4) with severity between 60%-90% to all tested isolates (Table 3). In addition, the resistance levels of all tested lines to different isolates were different in different seasons.

#### **Molecular identification**

The adult plant resistance gene *Sr2*, originating from the tetraploid Yaroslav emmer, is located on chromosome arm 3BS. Mago *et al.* (2011) showed that a DNA marker *Xgwm533* is closely linked to this gene and that a 120 bp specific band could be amplified by PCR from wheat cultivars (lines) carrying this gene. This marker was used to determine the presence of *Sr2* in the 95 advance wheat lines from Heilongjiang Province. Fifteen wheat lines as well as the positive control line 'Hope' produced the 120 bp band (Fig 1A), indicating that these wheat lines carry the resistance gene *Sr2*.

A specific molecular marker, *Sr24#12*, was developed to detect the presence of the *Sr24* gene. A specific band of about 500 bp could be amplified by PCR in the wheat line LcSr24Ag, known to contain *Sr24*. The results showed that this fragment was only amplified in the positive control, LcSr24Ag, but not in the negative control Little Club (LC) or in any of the tested lines, indicating the likely absence of the *Sr24* gene in those wheat lines.

The resistance genes *Sr25* and *Sr26* are derived from *Thinopyrum elongatum*. These two genes provide good resistance to Ug99 and its variants. For this reason, the 95 wheat lines were subjected



to PCR amplification with marker *Gb* (130 bp) linked with *Sr25* and with *Sr26#43* (207 bp) linked with *Sr26*. No specific fragments corresponding to these two primers were amplified in any of the tested wheat lines, except for their positive controls *Agatha/9\*LMPG* and *Eagle*, respectively, indicating the absence of those two genes in all 95 wheat lines.

Wheat stem rust gene *Sr31* originated from rye and has been deployed worldwide in many wheat cultivars. The molecular marker *SCSS30.2576*, producing a 576 bp specific PCR fragment, was used to characterize the absence or presence of *Sr31*. Out of 95 wheat genotypes tested using *SCSS30.2576*, the 576 bp fragment was identified in 11 wheat lines (*Jiusan07-6378*, *Jiusan06-6203*, *Jiusan07-6086*, *Long10-0449*, *Long10-7767*, *Long10-0453*, *Long11H1336*, *Long11-2097*, *Long11-1027*, *Longfu10K329*, and *Longfu08-6564*) as well as in the positive control *Sr31/6\*LMPG* (Fig 1B), indicating that those 11 advance wheat lines carry the *Sr31* gene.

The *Sr38* gene, linked with leaf rust gene *Lr37* and stripe rust gene *Yr17*, originated from *Triticum ventricosum* and is located on a 2NS/2AS translocation. The 2NS-specific STS marker *VENTRIUP-LN2* was used to detect the presence of the gene cluster. *VENTRIUP-LN2* amplified a 259 bp band in the positive control and in 24 (25.3%) wheat lines (Fig 1C, Table 1), confirming the presence of the *Sr38* gene in those 24 lines.

## Discussion

The stem rust resistance gene *Sr2* originated from diploid wheat which displays adult plant stage resistance (Singh *et al.*, 2011). The gene is located on chromosome 3BS and causes resistance to many *Pgt* races. It was introduced into North America and the CIMMYT wheat breeding program in 1925. Since then, it has been widely deployed in many countries, including China. This gene was combined with *Sr33* in production for nearly 70 years and remained resistant

(Periyannan *et al.*, 2013). Here, using a molecular marker, we identified fifteen wheat lines that carry *Sr2*, and that display all-stage resistance to the tested *Pgt* races 21C3CTTMM, 34C0MRGSM, and 34C3MTGQM. However, the Hope line, which carries a single *Sr2* gene, was susceptible to *Pgt* races 21C3CTTMM, 34C0MRGSM, and 34C3MTGQM at the seedling stage, therefore those 15 wheat lines ~~many may~~ contain another unknown resistance gene that confers resistance to the above three races at the seedling stage. Therefore, these resistant materials can be purposefully used to improve the resistance level of Heilongjiang wheat varieties to Chinese *Pgt* and Ug99 in future disease resistance breeding.

The *Sr24* gene, originating from *Thinopyrum ponticum* and located on 3DL of the wheat chromosome, is widely used in wheat production in the world. Because the gene does not confer resistance to Ug99, it was added to a North American system of nomenclature for *Pgt* in 2008 to identify Ug99 and its variants. Although *Sr24* did not provide resistance to some variants of Ug99, it provided excellent resistance to most Chinese races and to the new races TKTTF and TTTTF that caused disease epidemics in Ethiopia and Italy in 2014 and 2016, respectively. In the previous study, the molecular marker *Sr24#12* was used to screen wheat cultivars from Heilongjiang province. Unexpectedly, no wheat varieties that might contain this gene were found in 83 tested wheat materials (Li *et al.*, 2019). As we expected, no wheat lines that contain the gene were found in 95 tested wheat lines, in agreement with our previous study ~~that also found that~~ indicating no main commercial wheat cultivars carry this gene.

The *Sr25* and *Sr26* genes were derived from *Thinopyrum ponticum*. These two genes provided excellent resistance to Ug99 strains, TKTTF and TTTTF and to all races of *Pgt* that are found in China (Li *et al.*, 2019). Recently, with the diversification of breeding methods, considering their

excellent ability to provide resistance to Ug99 and its variants, wheat breeders in various countries began to use these two genes in order to improve the resistance to stem rust of wheat. Since *Sr25* is a temperature-sensitive gene, its resistance is affected by growth period and temperature. The resistance at the seedling stage is higher than at the adult stage, and ~~is~~ the plants are more susceptible at high temperatures. Research has shown that the gene is almost absent from wheat varieties in China, and our results confirm this (Li *et al.*, 2016). *Sr26* is mainly applied to wheat breeding in Australia, and it is seldom used in China. No wheat lines containing *Sr26* were found among the tested varieties in this study. Combining our results with those from previous reports, *Sr26* was not found in nearly 400 wheat materials collected from different regions of China (Li *et al.*, 2016; Li *et al.*, 2019; Xu *et al.*, 2017, 2018). Therefore, it is suggested that the introduction of this gene into wheat breeding would enrich the diversity of resistance sources of wheat varieties in China.

The *Sr31* gene is one of the most widely used stem rust resistance genes in wheat breeding in the world. It is located on 1BL/1RS chromosome and was first transferred from ‘Petkus’ rye to bread wheat (Mago *et al.*, 2002). In the 1960s, China began to introduce ‘Soviet Union’ and ‘Romania’ wheat strains containing *Sr31* (Jiang *et al.*, 2007). Since then, this gene has been widely used in wheat breeding in China, and the cultivated area of wheat varieties carrying this gene accounts for more than 60%. Although *Sr31* has “lost” its effectiveness to Ug99 races, it has always provided excellent resistance to all domestic stem rust isolates in China’s wheat production. Knowing the distribution of this gene in domestic cultivars is of practical significance for monitoring for Ug99 and preventing the occurrence of stem rust in China. In this study, the *Sr31*-linked marker SCSS30.2576 was used to detect the distribution of this gene in 95 wheat lines from Heilongjiang

province, and pedigree analysis revealed that 11 of those wheat lines carried *Sr31*. The characterization of the resistance of these wheat lines to three races of *Pgt* also supported this result, since all of these wheat lines produced low ITs (0 to 2) at the seedling stage, and were immune (I), resistant (R) or moderately resistant (MR) at the adult-plant stage with relatively low severity (< 30%). Thus, our results suggest that there are relatively few wheat varieties containing *Sr31* in Heilongjiang province, less than in other provinces in China (Cao *et al.*, 2019; Xu *et al.*, 2017, 2018).

The *Sr38* gene originated from *Aegilops ventricosa* L. It was first transferred into the winter wheat variety ‘VPM1’ and is closely related to the stripe rust resistance gene *Yr17* and the leaf rust resistance gene *Lr37* in wheat (Bariana and McIntosh 1993). The *Yr17-Lr37-Sr38* gene cluster is widely used in the world since it provides excellent combined resistance to stripe rust, leaf rust, and stem rust of wheat. In this study, specific fragments were amplified in 19 wheat lines, indicating that these lines may contain *Sr38*. Similar to *Sr31*, *Sr38* has also “lost” its ability to provide resistance to the Ug99 races, but no *Pgt* isolate can overcome this resistance in China. Therefore, *Sr38* will still play a role in the prevention and control of domestic stem rust, but Ug99-resistant genes should be aggregated in breeding to improve the resistance level of Chinese wheat cultivars to this disease.

Our results also showed that the wheat lines from Heilongjiang province displayed good resistance to tested *Pgt* races. Of the 95 wheat lines tested, 86 (90.5%) not only had good resistance to the races 21C3CTTMM, 34C0MKGSM, and 34C3MTGQM at the seedling stage, but also showed good resistance to three races in the resistance evaluation of two consecutive years at the adult stage, and had low severity (< 30%). Therefore, those 86 wheat lines have all-stage

resistance to tested races. This may be related to the fact that resistance to *Pgt* is a breeding goal of wheat lines, and wheat cultivars approved in Heilongjiang province must be resistant to wheat stem rust. All wheat lines are screened with the predominant race group 21C3 and the sub-dominant race group 34 by the Plant Immunity Laboratory of Shenyang Agricultural University at the field nursery before registration, and only wheat lines with medium resistance or above can be registered as new varieties through variety examination and approval. From the results of molecular detection, the wheat lines have abundant resistant material containing broad-spectrum stem rust resistance genes *Sr2* as well as *Sr31* and *Sr38* that provide resistance to all wheat stem rust in China. It may also contain other unknown resistance genes. This excellent material can be used as precious germplasm for the breeding of resistant wheat lines in the future.

### **Conclusion**

Breeding resistant cultivars is the most cost-effective and eco-soundly-friendly strategy to protect wheat from wheat stem rust. In this study, ~~the~~ resistance to *Pgt* of 95 wheat advance lines from Heilongjiang Province was evaluated using three predominant races of *Pgt*, 21C3CTTM, 34C0MKGSM, and 34C3MTGQM, in China at the seedling and adult plant stage. Overall, the ~~resistant-resistance~~ level of wheat lines to wheat stem rust were strong in Heilongjiang Province. Base on it, the presence of genes *Sr2*, *Sr24*, *Sr25*, *Sr26*, *Sr31*, and *Sr38* in these lines were detected using gene specific DNA markers. The results showed that 42 tested wheat lines might carry one of these genes. This information can be used in wheat-breeding plans for stem rust resistance in the future.

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