

# Driver versus navigator causation in biology: The case of insulin and fasting glucose

Manawa Diwekar-Joshi<sup>Corresp., 1</sup>, Milind Watve<sup>Corresp. 2</sup>

<sup>1</sup> Biology, Indian Institute of Science Education and Research, Pune, Maharashtra, India

<sup>2</sup> Deenanath Mangeshkar Hospital and Research Centre, Pune, Maharashtra, India

Corresponding Authors: Manawa Diwekar-Joshi, Milind Watve

Email address: manawa.diwekar@students.iiserpune.ac.in, milindwatve@gmail.com

**Background:** In bio-medicine, inferring causal relation from experimental intervention or perturbation is believed to be a more reliable approach than inferring causation from cross-sectional correlation. However, we point out here that even in interventional inference there are logical traps. In homeostatic systems, causality in a steady state can be qualitatively different from that in a perturbed state. On a broader scale there is a need to differentiate driver causality from navigator causality. A driver is essential for reaching a destination but may not have any role in deciding the destination. A navigator on the other hand has a role in deciding the destination and the path but may not be able to drive the system to the destination. The failure to differentiate between types of causalities is likely to have resulted into many misinterpretations in physiology and bio-medicine. **Methods:** We illustrate this by critically re-examining a specific case of the causal role of insulin in glucose homeostasis using five different approaches (1) Systematic review of tissue specific insulin receptor knock-outs, (2) Systematic review of insulin suppression and insulin enhancement experiments, (3) Differentiating steady state and post-meal state glucose levels in streptozotocin treated rats in primary experiments, (4) Mathematical and theoretical considerations and (5) Glucose insulin relationship in human epidemiological data. **Results:** All the approaches converge on the inference that although insulin action hastens the return to a steady state after a glucose load, there is no evidence that insulin action determines the steady state level of glucose. Insulin, unlike the popular belief in medicine, appears to be a driver but not a navigator for steady state glucose level. It is quite likely therefore that the current line of clinical action in the field of type 2 diabetes has limited success largely because it is based on a misinterpretation of glucose-insulin relationship. The insulin-glucose example suggests that we may have to carefully re-examine causal inferences from perturbation experiments and set up revised norms for experimental design for causal inference.

# Driver versus navigator causation in biology: the case of insulin and fasting glucose

Manawa Diwekar-Joshi<sup>1\*</sup>, Milind Gajanan Watve<sup>2\*</sup>

<sup>1</sup>Indian Institute of Science Education and Research, Pune, India

<sup>2</sup>Deenanath Mangeshkar Hospital and Research Centre, Pune, India

\*Corresponding Authors:

Manawa Diwekar-Joshi

Indian Institute of Science Education and Research, Pune

Dr. Homi Bhabha Road, Pashan, Pune 411008, Maharashtra, India

Email address: manawa.diwekar@students.iiserpune.ac.in

Milind Gajanan Watve

Deenanath Mangeshkar Hospital and Research Centre,

Earndwane, Pune 411004, Maharashtra, India

Email address: milindwatve@gmail.com

## Abstract

**Background:** In bio-medicine, inferring causal relation from experimental intervention or perturbation is believed to be a more reliable approach than inferring causation from cross-sectional correlation. However, we point out here that even in interventional inference there are logical traps. In homeostatic systems, causality in a steady state can be qualitatively different from that in a perturbed state. On a broader scale there is a need to differentiate driver causality from navigator causality. A driver is essential for reaching a destination but may not have any role in deciding the destination. A navigator on the other hand has a role in deciding the destination and the path but may not be able to drive the system to the destination. The failure to differentiate between types of causalities is likely to have resulted into many misinterpretations in physiology and bio-medicine.

**Methods:** We illustrate this by critically re-examining a specific case of the causal role of insulin in glucose homeostasis using five different approaches

(1) Systematic review of tissue specific insulin receptor knock-outs,

- (2) Systematic review of insulin suppression and insulin enhancement experiments,
- (3) Differentiating steady state and post-meal state glucose levels in streptozotocin treated rats in primary experiments,
- (4) Mathematical and theoretical considerations and
- (5) Glucose insulin relationship in human epidemiological data.

**Results:** All the approaches converge on the inference that although insulin action hastens the return to a steady state after a glucose load, there is no evidence that insulin action determines the steady state level of glucose. Insulin, unlike the popular belief in medicine, appears to be a driver but not a navigator for steady state glucose level. It is quite likely therefore that the current line of clinical action in the field of type 2 diabetes has limited success largely because it is based on a misinterpretation of glucose-insulin relationship. The insulin-glucose example suggests that we may have to carefully re-examine causal inferences from perturbation experiments and set up revised norms for experimental design for causal inference.

## Introduction

### 1. Causal relationships in Biology:

Inferring causal relationships in biology is a more complex, philosophical and methodological issue than what is generally perceived. Much of the debate has centered around making causal inferences from observed associations or correlations (Boudon, 1965; Chawla et al., 2018; Grace, 2002; A. B. Hill, 1965). Methods of analysis specifically addressing the question of causal inferences have been developed for various contexts and at various levels of reasoning including Hill criteria (A. B. Hill, 1965), path analysis (C. C. Li, 1956; Niles, 1923; Wright, 1960), Granger causality (Granger, 1969), steady state causality (Chawla et al., 2018), genomic and network causality (Kulkarni et al., 2017; Meinshausen et al., 2016; Triantafillou et al., 2017). While on the one hand, complex statistical and computational tools are being developed for determining causality under different contexts, certain simple problems about causality are not sufficiently appreciated and addressed. We focus on one such apparently simple problem here, the lack of appreciation of which has led to serious misinterpretations of experimental results.

In classical experimental physiology, interventions or perturbations are believed to be reliable indicators of causation and there is little debate about it. If the experimenter perturbs A and finds

a significant effect on B after following all fundamental principles of experimental design, the change in A is inferred to be causal to the change in B. However, there are many subtleties in drawing a causal inference from experimental interventions that have not yet attracted sufficient philosophical as well as methodological attention among experimental biologists. One such thinking trap is that in homeostatic systems the nature of causality in a perturbed state can be qualitatively different from that in equilibrium or steady state and the failure to distinguish between the two may have substantially misled biomedical research.

## 2. The role of growth rates in Lotka-Volterra competition models

A well worked out theoretical model that can be used to distinguish clearly between perturbed and steady state causation is the Lotka-Volterra (LV) competition model. This model describes the dynamics for interspecific competition (Gotelli, 2008). The growth of two interacting populations is modelled using logistic equations for both the populations. The changes in populations depend on the individual growth rates and the carrying capacities of the two populations (equations 1 and 2). Additional parameters are the competition coefficients  $\alpha$  and  $\beta$  which represent the effects of the two species on each other. Thus the carrying capacities  $K_1$ ,  $K_2$  and the competition coefficients  $\alpha$  and  $\beta$  determine the equilibrium population (equations 3 and 4) (Gotelli, 2008).

$$\frac{dN_1}{dt} = r_1 \cdot N_1 \left( \frac{K_1 - N_1 - \alpha N_2}{K_1} \right) \quad \text{Equation 1}$$

$$\frac{dN_2}{dt} = r_2 \cdot N_2 \left( \frac{K_2 - N_2 - \beta N_1}{K_2} \right) \quad \text{Equation 2}$$

$$\check{N}_1 = \frac{K_1 - \alpha K_2}{1 - \alpha \beta} \quad \text{Equation 3}$$

$$\check{N}_2 = \frac{K_2 - \beta K_1}{1 - \alpha \beta} \quad \text{Equation 4}$$

where,

$N_1$  and  $N_2$  are the population sizes of the two competing species respectively,  $\check{N}_1$  and  $\check{N}_2$  representing steady state populations.

$r_1$  and  $r_2$  are the growth rates

$K_1$  and  $K_2$  are the carrying capacities

$\alpha$  is the competition coefficient which shows the effect of population 2 on population 1

$\beta$  is the competition coefficient which shows the effect of population 1 on population 2

**Figure 1:** Simulated population dynamics of two competing species A (black lines) and B (grey lines) at different growth rates. Solid lines represent high growth rates reaching the equilibrium faster whereas dotted lines represent slow growth rates reaching the equilibrium more slowly. In both cases species A has slower growth rate but greater carrying capacity than species B. At time T1 the populations are in proportion to their growth rates but at T2, approaching equilibrium, growth rates become increasingly irrelevant in determining population sizes. Thus, growth rates are important determinants of population size in a perturbed state but not at a steady state.

If, in an experiment, we start at a non-equilibrium state, and observe at time T1 (figure 1), the standing populations would be inferred as a function of intrinsic growth rates  $r_1$  and  $r_2$ . But if observed at time T2, the inference would be different. The intrinsic growth rates of both the competing populations do not determine the equilibrium populations of the two species. The magnitude of  $r$  determines the time taken by the population to reach the equilibrium or steady state (figure 1). The role of growth rates in population dynamics is well recognized and is demonstrable in a perturbed state but it needs to be realized that it has no role in determining the steady state populations. Nevertheless, existence of non-zero positive growth rates is essential for attaining the equilibrium or returning to it if perturbed. If either or both the growth rates are made zero, the system will never attain back a stable equilibrium coexistence. Thus, the two growth rates are causal for attaining equilibrium, but they have no causal role in deciding the position of the equilibrium point. Thus, we need to distinguish between the driver cause and the navigator cause. Driver causality is a process that takes a homeostatic system to an equilibrium point but may not have any role in deciding the attributes of the equilibrium. Navigator causality refers to the processes that determine the location of the equilibrium point and lead the driver there, but in the absence of the driver, may not be able to take the system to the steady state. For a homeostatic system, the distinction between perturbed state and steady state causality is practically equivalent to driver and navigator causality. However, the driver-navigator distinction can be applied, in principle, to non-homeostatic systems as well and therefore is a broader concept.

This has relevance to experimental physiology. If knocking out a certain gene, protein or function disables homeostatic control, it does not provide us any clue as to whether it has a driver or navigator function. The experiment does not necessarily demonstrate that the gene, protein or function determines the steady state levels of the controlled variable. Since distinction between

driver and navigator causality has not been explicitly made in experimental physiology, currently there are no norms or methods to resolve between the two types of causations. We use this distinction below to re-examine the role of insulin in glucose homeostasis and show that the failure to distinguish between driver and navigator causality has led to a fundamentally flawed understanding of glucose homeostasis and type 2 diabetes in particular.

### **3. Why is insulin believed to regulate fasting blood sugar: A burden of history?**

After the classical demonstration by Claud Bernard that damage to medulla oblongata causes hyperglycaemia(Bernard, 1879), the second major breakthrough was the demonstration by von Mering and Minkowski that pancreatectomy resulted in hyperglycaemia(Mering & Minkowski, 1890) and further that pancreatic extracts resulted in lowering of plasma glucose. The active principle eventually purified became known as insulin (Karamitsos, 2011). The discovery and success of insulin in treating diabetes was so overwhelming that insulin became the key molecule in glucose homeostasis and the role of brain and other mechanisms were practically forgotten. It should be noted that the prevalent type of diabetes then was what we would call type 1 diabetes (T1D) today in which there is almost complete destruction of pancreatic  $\beta$ -cells. The distinction between type 1 and 2 developed gradually over the next five decades along with the realization that insulin levels may be normal or raised in type 2 diabetes (T2D) and that a substantial population of  $\beta$ -cells survives lifelong (Butler et al., 2003; Clark et al., 1990; Porte & Kahn, 2001). However, by now the thinking about glucose homeostasis was so insulin-centered, that the inability of normal or raised levels of insulin to keep plasma glucose normal was labelled as “insulin resistance”(Reaven, 1988) without adequately examining and eliminating alternative possibilities and the concept got wide uncritical acceptance. Although insulin receptor and downstream functions are known to be highly variable at the cellular level, the question whether altered insulin signalling is solely or mainly responsible for fasting hyperglycaemia of T2D, or other insulin independent mechanisms play a significant role is not clearly answered.

There are multiple reasons to doubt and re-examine the role of insulin in glucose regulation in relation to T2D (Corkey, 2012; Pories & Dohm, 2012; M. Watve, 2013). Exogenous insulin and other insulin-centered lines of treatment have largely failed to reduce diabetic complications and mortality in T2D although short term glucose lowering may be achieved (ACCORD, 2008; King

et al., 2001; Meinert et al., 1970; UK Prospective Diabetes Study Group, 1998a, 1998c, 1998b). In the long run even the glucose normalization goal is not achieved in majority of cases (UK Prospective Diabetes Study Group, 1998b, 1998c). A number of mechanisms are known to influence glucose dynamics, partially or completely independent of insulin signalling, including autonomic signals (Nonogaki, 2000; Schwartz, 2005), glucocorticoids (Di Dalmazi et al., 2012; Gathercole & Stewart, 2010; Goldstein et al., 1993; Kuo et al., 2015), insulin independent glucose transporters (Carruthers et al., 2009) and certain other hormones and growth factors (Clemmons, 2004; Jansen et al., 2006; Messmer-Blust et al., 2012; Suh et al., 2014). Analysis of multi-organ signalling network models have also raised doubts about the central role of insulin and insulin resistance in T2D (Kulkarni et al., 2017).

The definitions as well as clinical measures of insulin resistance are such that the effects of all other mechanisms are accounted for under the name of “insulin resistance”. For example, the HOMA-IR index is calculated as a product of fasting glucose and fasting insulin (Matthews et al., 1985; Turner et al., 1979). The belief that this product reflects insulin resistance is necessarily based on the assumption that insulin signalling alone quantitatively determines glucose level in a fasting steady state. The assumption has seldom been critically examined. If any other mechanisms are contributing to impaired fasting glucose, they will be included in the HOMA-IR index going by the way it is calculated and would be labelled as insulin resistance. This amounts to a circular logic. Insulin resistance is hypothesized to be responsible for the failure of insulin to control fasting glucose, and insulin resistance is measured as the inability of insulin to control fasting glucose. This makes the insulin resistance concept unfalsifiable from clinical data.

We have previously showed using mathematical and statistical tools of causal analysis (Chawla et al., 2018) that the classical pathway of obesity induced insulin resistance leading to a hyperinsulinemic normoglycemic prediabetic state and the faithfulness of HOMA indices in measuring insulin resistance cannot be simultaneously true. Either the HOMA indices do not represent insulin resistance faithfully or the classically believed pathway of compensatory insulin response leading to hyperinsulinemic normoglycemic state is wrong according to this analysis (Chawla et al., 2018). All the mounting inconsistencies and paradoxes in the insulin resistance

concept along with the limited success of type 2 diabetes treatment warrant a re-examination of the foundational concepts.

#### 4. Approach used in this paper

We examine here the long-held belief that altered insulin signalling is responsible for fasting as well as post prandial hyperglycemia in T2D using five different approaches

(1) Systematic review of experiments involving tissue specific insulin receptor knock-outs (IRKOs)

(2) Systematic review of experiments to chronically raise or lower insulin levels

(3) Primary experiments on streptozotocin (STZ) induced hyperglycaemia in rats, that differentiate between steady (fasting) and perturbed (post-feeding) state

(4) Examining the insulin resistance hypothesis for being mathematically possible and theoretically sound

(5) Analysis of insulin-glucose relationship in steady state versus post-meal perturbed state in human epidemiological data for testing the predictions of mathematical models.

The first three approaches have the advantage of using specific molecular interventions where the target is precisely known. For the analyses we chose mechanisms of insulin level/action modification which have been used extensively and have been reproduced by multiple labs world over. The possible disadvantage is that they are mostly animal experiments and doubts are expressed about whether the results are directly relevant to humans (Akhtar, 2015; Ali et al., 2018; Bracken, 2009). However, some of the experiments reported are human and they converge with the inferences of the animal experiments. In the last two approaches, human epidemiological data are used in which the experimental molecular precision is not expected, but we test certain specific predictions of the insulin resistance hypotheses using novel analytical approaches and examine whether they converge on similar inferences. The convergence of human and animal data is important to reach robust conclusions. We will now describe each approach in detail here.

#### 4.1 Systematic review of experiments involving tissue specific insulin receptor knock-outs



The first step in insulin signaling is the binding of insulin to insulin receptor (Bevan, 2001). The downstream actions of this event finally lead to insulin-dependent glucose uptake in insulin dependent tissues of the body. Experimentally, disruption of insulin signaling is achieved by knocking out or inhibiting various players in the signaling cascade. We chose to look at the effects of knocking out the tissue specific insulin receptor on fasting and post-meal/feeding or post glucose load levels in rodent models. Studying tissue specific insulin receptor knockouts enables us to differentiate between the roles of insulin signaling in different tissues. A classical belief is that the post-meal glucose curve is mainly influenced by the rate of glucose uptake by tissues, mainly muscle, whereas the fasting glucose levels are mainly determined by the rate of liver glucose production (Bock et al., 2007). If this belief is true one expects that muscle specific knockout would mainly affect the GTT curve but may not affect fasting glucose level, whereas liver specific knockout would mainly affect the fasting glucose level. We searched the published literature for experiments in which the fasting and post-feeding levels of glucose are measured in tissue specific insulin receptor knock-outs.

## **4.2 Systematic review of experiments to chronically raise or lower insulin levels**

The insulin receptor knockout experiments assume that the main action of insulin is through the specific receptors. It can be argued that insulin acts through other receptors or may have other mechanisms of action yet unknown and therefore receptor knockouts do not fully eliminate insulin action. Alternatively, we can alter insulin level itself to see how it affects glucose level in fasting state or post glucose load. Insulin is known to alter plasma glucose immediately on administration (Galloway et al., 1981), but this is not a steady state response. If insulin levels can be raised or lowered and sustained long enough to reach a steady state, the effect of insulin on glucose in a steady state can be studied. If insulin affects steady state glucose, a sustained rise in insulin will result into a sustained lower steady state glucose level. Conversely a sustained suppression of insulin would lead to higher steady state glucose. We studied published literature for experiments where a stable and sustained increase or decrease in insulin was achieved and then the effect on fasting glucose and GTT studied.

## **4.3 Fasting versus post feeding glucose in STZ rats**

Streptozotocin (STZ) induced diabetes is a popular model of rodent diabetes(Akbarzadeh et al., 2007; Gajdosík et al., 1999). STZ acts by specifically destroying the insulin producing  $\beta$  cells of the pancreatic islets(Szkudelski, 2001). A low dose of STZ that destroys a substantial population of  $\beta$  cells but does not lead to total destruction of their population is often perceived as a good model for T2D, whereas a high dose of STZ that destroys the  $\beta$  cell population almost entirely is perceived as a model of T1D(Akbarzadeh et al., 2007; Asrafuzzaman et al., 2017; Freitas et al., 2015; Gajdosík et al., 1999; Kagami et al., 2008; Kovacs et al., 2011; Szkudelski, 2001; Zhang et al., 2008). We searched literature to look for studies that carefully differentiated between steady state glucose from post load glucose in STZ models but did not find any studies that make this distinction clear. Therefore, we designed and conducted experiments to differentially study the steady state and perturbed state glucose levels in rats treated with STZ.

#### 4.4 Theoretical and mathematical considerations

In this approach we elaborate on the theoretical underpinnings of insulin-glucose relationship. We also explore possible explanations for the unexpectedly consistent failure of experimental insulin signal impairment to alter steady state glucose level. Simultaneously we make differential predictions from alternative homeostasis models that can be tested in human epidemiological data. The two models which we use here answer the critical question in glucose homeostasis: whether the fasting/steady state glucose level is a consequential balance between glucose production and glucose utilization rates (consequential steady state CSS) or whether there is a target glucose level that is maintained by sensing and correcting any changes in it (targeted steady state TSS).

#### 4.5 Analysis of insulin glucose relationship in steady and perturbed state in human data: Epidemiological inquiry

Here we use human epidemiological data to test the correlational predictions made by the CSS versus TSS models of glucose homeostasis.

## Materials & Methods

We will outline the methods used in each of the 5 approaches described in the last section of the introduction.

285

# 286 ***Systematic Review Registration***

- 287 1. The four meta-analyses were registered in PROSPERO.
- 288 2. We did not start the project with the intention of doing these systematic reviews. Our  
289 primary goal was to test the alternative evolutionary hypotheses for the origins of type 2  
290 diabetes (M. Watve & Diwekar-Joshi, 2016). While pursuing this work we noticed  
291 certain basic anomalies in the classical theories of diabetes. At the stage when we thought  
292 of the question addresses in this paper, we had already scanned much of the literature  
293 used here.
- 294 3. To ensure that our review is unbiased and rigorous enough, we repeated the literature  
295 search following PRISMA and PROSPERO guidelines. Therefore, much of the search for  
296 the Meta-analyses was done before the actual PROSPERO registration. The secondary  
297 screening, data extraction and the data analysis was done during the time required after  
298 the submission to the PROSPERO and the actual approval of the registration by them.
- 299 4. But we were glad to find after following proper procedures that our prior work met the  
300 standards of unbiased and rigorous review.
- 301 5. Details about the search strategies, quality assessment and data analysis for the four meta-  
302 analyses are included in Supplementary information 1.
- 303 6. The flowcharts for the search strategies and screening procedures followed are given in  
304 the form of PRISMA flowcharts and checklists and are given as Supplementary  
305 Information files.

306

# 307 ***Systematic Review/Meta-analyses: Search and screening***

- 308 1. Manawa Diwekar-Joshi and Milind Watve decided the search strategy and the actual  
309 searches and screening based on the strategy was done by Manawa Diwekar-Joshi.
- 310 2. Data analysis was done by both the authors.
- 311 3. Disagreements were dissolved by discussion and input from other lab members.
- 312 4. The referees for the manuscript were other post-doc and graduate students of the Watve  
313 Lab.

314

# 315 ***Statistical approach (used for all the meta-analyses)***

Although we short-listed papers that used similar methods, small differences in protocols can make considerable differences in the results. There is substantial variation in results across studies. Therefore, we use non-parametric methods for analyzing the pooled data. We first look at in how many of the experiments the treatment means are greater than the control means and in how many they are smaller. If this difference is significant, we conclude that there is enough qualitative consistency across experiments to reach a reliable inference. If there is a consistent direction of difference, we look at how many are individually significant. As a conservative approach we avoid pooling data quantitatively since across studies there are differences in age or weight of animals, number of days after treatment, number of hours of fasting and other variables. This approach is maintained for the analysis of all the meta-analyses.

### **1. Systematic review of experiments involving tissue specific insulin receptor knock-outs**

The details of the method of the systematic literature review involving the tissue specific insulin receptor knock-outs are given in table 1. The systematic review was registered in PROSPERO (ID: CRD42019132379). The details of the experiments of the shortlisted studies can be seen in table 1 of the supplementary information 1 which shows that similar methods have been utilized to create the knockouts and therefore a comparative analysis is justified.

**Table 1:** Systematic literature review for studies on tissue specific insulin receptor knockouts.

## **2. Systematic review of experiments to chronically raise or lower insulin levels**

### **2.1 Increase in insulin**

A model for sustained increase in insulin levels is a knock-out or inhibition of the insulin degrading enzyme (IDE). An interplay between insulin secretion and insulin degradation maintains the level of insulin in plasma (Authier et al., 1996; Duckworth et al., 1998; Hulse, Ralat, & Wei-Jen, 2009; Shen et al., 2006). Plasma insulin has a half-life of 4 to 9 minutes (Hulse, Ralat, & Wei-Jen, 2009; Tomasi et al., 1967) and it is degraded predominantly by the insulin degrading enzyme (IDE) (Hulse, Ralat, & Wei-Jen, 2009; Shen et al., 2006). Inhibition of IDE has been considered as a therapeutic option for type 2 diabetes with limited and questionable success (Costes & Butler, 2014; Maianti et al., 2014). We performed a systematic literature review to find out experiments in which IDE was inhibited to obtain a sustained high

plasma insulin level and, in such animals, GTT was performed (table 2) (PROSPERO registration ID: CRD42019140619).

**Table 2:** Systematic literature review for studies on insulin degrading enzyme inhibition/knockout.

## 2.2 Decrease in insulin

We performed a systematic literature review for experiments in which there was sustained suppression of insulin production. Two insulin suppressing agents have been repeatedly used to lower insulin production in rodent models as well as in humans.

(i). Diazoxide (DZX): Diazoxide is a potassium channel activator which causes reduction in insulin secretion by the  $\beta$ -cells by keeping the cells in a hyperpolarized state by opening the channel (Panten et al., 1989). It has been used as a drug to modulate insulin secretion for research and therapeutic purposes (Doyle, 2003).

(ii). Octreotide (OCT): Octreotide is a somatostatin analogue which inhibits insulin and growth hormone. It has been used to reduce insulin secretion *in vitro* and *in vivo* (Lamberts et al., 1996).

We searched the literature systematically for studies where the insulin levels have been altered using either DZX or OCT and glucose tolerance has been examined using a GTT after DZX/OCT treatment (table 3) (PROSPERO registration ID for DZX: CRD42020141688; PROSPERO registration ID for OCT: CRD42020141464). It should be noted that this literature includes a significant proportion of human trials. We also searched literature for studies in which insulin was suppressed by other methods.

**Table 3:** Systematic literature review for studies on insulin suppression with diazoxide and octreotide

## 3. Steady state versus perturbed state glucose in STZ rats

### 3.1 Animal model and conditions

#### *Ethics Approval*

The experiments performed on Sprague Dawley (SD) rats had been approved by the Institutional Animal Ethics Committee at IISER, Pune (Protocol Number IISER/IAEC/2016-02/006)

constituted by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Govt. of India.

### ***Housing of the animals***

All the rats were housed in a facility with a temperature of  $23\pm 2^{\circ}\text{C}$  and a 12-hour light/dark cycle with standard rat chow (Altromin rat/mice maintenance diet) and water available ad libitum. The bedding of the cages was changed every three days and every day after injection of STZ. There were no extra measures taken for the enrichment of the animals.

### ***Euthanasia***

The animals were not euthanized before the end of the experiments. At the end of the 12 days of glucose and insulin readings, the animals were euthanized with an intraperitoneal injection of thiopentone (100-120mg/kg body weight).

## **3.2 STZ treatment for insulin suppression**

Male, SD rats weighing 180-200 g were injected with STZ at 50 mg/kg body weight. The STZ was dissolved in Citrate Buffer (Citric Acid: 0.1M and Sodium Citrate: 0.1M). Injection of citrate buffer alone was used as control.

## **3.3 Fasting and post-feeding glucose in 12 day follow up**

Three days after the STZ injection, the rats were fasted for 16 hours and glucose was measured using the hand held Accu-Chek Glucometer. The rats were then given 40 grams of Standard Chow for 8 hours. Food was weighed and post-meal glucose was measured after three hours. The protocol was repeated for 12 days and body weight, food weight and glucose readings were taken daily. 12 animals per group were used for this experiment.

## **3.4 Duration of fasting**

An experiment was also performed to see how much time was required to reach a steady state of glucose after removal of food. The food was removed from the STZ and Control animals after ad libitum availability and glucose readings were taken after 3 hours, 6 hours, 9 hours, 12 hours and 16 hours. After a recovery of three days, glucose levels were measured only at 16 hours after removing the food. 9 STZ treated animals and 10 Control animals (injected with citrate buffer) were used for this experiment.

408

## 409 **4. Theoretical and mathematical considerations**

### 410 **4.1 Choice of models for glucose homeostasis**

411 The fasting state has been generally accepted to be a steady state for glucose concentration for  
 412 several reasons. In a given healthy individual the fasting glucose levels are stable in time (Halter  
 413 et al., 1985; Lerner & Porte, 1972). The post-meal peak of glucose and insulin returns to the  
 414 fasting level within a few hours and remains stable over a long time. The fasting state is  
 415 considered and modelled as a steady state by the widely used HOMA model (Matthews et al.,  
 416 1985; Turner et al., 1979). Classically the negative feedback loops are assumed to work through  
 417 insulin and insulin is taken as a determinant of steady state glucose level. Most popular models  
 418 of glucose homeostasis work on this assumption although non-steady state models of insulin  
 419 resistance exist (Palumbo et al., 2013).

420

### 421 **4.2 Models used for this analysis**

422 A critical question in glucose homeostasis is whether the fasting steady state glucose level is a  
 423 consequential balance between glucose production and glucose utilization rates (consequential  
 424 steady state CSS) or whether there is a target glucose level that is maintained by sensing and  
 425 correcting any changes in it (targeted steady state TSS). The difference in the two can be  
 426 visualized by the tank water level analogy (figure 2). If a tank has an input tap releasing water in  
 427 it at a constant rate and has an outlet at the bottom through which water escapes proportionate to  
 428 the pressure of the water column, a steady state is invariably reached (figure 2A). The steady  
 429 state level is decided by the rate of intake and the size of the outlet. This is a CSS which will  
 430 change with any change in the size/capacity of the input or the outlet tap. In contrast to CSS, in a  
 431 TSS there is a desired water level and sensors are placed above and below the desired level such  
 432 that when the level goes below the lower sensor the input is switched on or its rate increased  
 433 and/or output switched off or its rate decreased (figure 2B). Refer to supplementary information  
 434 2 for the details of the CSS model.

435

436 **Figure 2:** The consequential steady state (CSS) (A) and targeted steady state (TSS) (B) models of homeostasis  
 437 illustrated with a tank water level analogy. In CSS a change in the size of inlet or outlet tap, analogous to insulin  
 438 sensitivity can change the steady state level. In a TSS model, a change in the tap size will alter the time required to

reach a steady state but will not change the steady state level due to the presence of sensors which regulate the inlet and outlet taps

## **5. Analysis of insulin glucose relationship in steady and perturbed state in human data:**

### **Epidemiological inquiry**

#### **5.1 Epidemiological data**

The three data sets used here come from two different studies: (i) Coronary Risk of Insulin Sensitivity in Indian Subjects (CRISIS) study, Pune, India (Yajnik et al., 2007) and (ii) Newcastle Heart Project (NHP), UK (Bhopal et al., 1999). Data from the latter is divided into two groups as the subjects belong to different ethnicities namely European white and south Asian and we will prefer to analyze the two groups separately since certain ethnic differences are likely to be present in the tendency to develop metabolic syndrome (Bhopal, 2013; Gujral et al., 2013). Hence all the comparison of predictions with the data has been done independently for the three data sets. All the studies are population surveys that include non-diabetic (fasting glucose values less than 110mg/dl) and diabetic individuals (fasting glucose values above 110 mg/dl) and the clinical history, morphometric parameters, glucose and insulin during fasting and oral glucose tolerance test (OGTT) of the subjects were recorded. In the analysis, we included only the non-diabetic groups in which the homeostatic mechanism can be assumed to be intact and therefore any hypothesis about it can be tested. Most of the individuals in the diabetic group would be under different drug regime affecting glucose-insulin dynamics in different ways and therefore we exclude that group for the analysis.

#### **5.2 Statistics**

Linear regression and correlation were used to compare the glucose-insulin relationship in steady state (fasting) versus perturbed state (post glucose load) in the three data sets along with the relationships between HOMA-IR and HOMA- $\beta$  derived from the fasting data.

## **Results**

### **1. Systematic review of experiments involving tissue specific insulin receptor knock-outs**



# **Pooled results of IRKO experiments**

We shortlisted 16 papers describing 46 independent experiments in which glucose tolerance curves of insulin receptor knockouts and controls were compared (table 1 of supplementary information 1). The experiments could be segregated in four different tissue specific knockouts for the analysis: fat/adipose insulin receptor knockout or FIRKO (figure 3), Muscle insulin receptor knockout or MIRKO (figure 4), liver insulin receptor knockout or LIRKO (figure 5) and  $\beta$ -cell insulin receptor knockout or  $\beta$ IRKO (figure 6). A generalized trend in the total picture summed up over all four IRKOs was that along the GTT curve, significantly higher glucose levels are seen in the knockouts as compared to the controls, particularly and consistently at 30, 60 and 120 minutes. However, the fasting glucose level was not significantly different. In some studies, fasting glucose was significantly higher in the knockouts than the controls, however in some other studies it was significantly lower as well. In 29 out of 46 experiments there was no significant difference (table 4) in the fasting glucose levels of knockouts and controls. This trend was consistently seen in MIRKO (figure 4), LIRKO (figure 5) and  $\beta$ IRKO (figure 6). Only in FIRKO (figure 3) there were greater number of studies showing fasting glucose significantly higher in the knockouts than in the controls (Guerra et al., 2001; Sakaguchi et al., 2017; Softic et al., 2016), but in the non-parametric analysis the collective trend was not significant table 4 . These inconsistencies in the FIRKO fasting glucose levels compared to the MIRKO LIRKO could be explained with the help of the duration of fasting used for the glucose tolerance tests. Although the glucose levels in the WT are higher than the FIRKO/BATIRKO in the fasting conditions, there could be possible reasons for that. For example, in the study (Sakaguchi et al., 2017), the fasting for the glucose tolerance test was carried out only for 6 hours as compared to 16 hours/ON in other studies. Secondly, one of the knockouts used in this study is a double knockout of insulin receptor as well as the insulin-like growth factor 1 receptor which could be a possible reason for the higher glucose levels in the fasting condition. In case of the (Guerra et al., 2001; Softic et al., 2016), the animals have been fasted ON for the glucose tolerance test and the tests have been performed on FIRKO and WT of different ages. The impairment of glucose tolerance increases with age, through this also is not seen consistently across all the studies. In the case of (Blüher et al., 2002), the fasting duration for the GTT is 16 hours, highest in all the studies and in this case the treated glucose levels in the fasting condition are equal to or lower than that of the controls. It is important to note that in FRIKO, although there are maximum

number of cases where in the treated is higher than the control in the fasting condition, this trend is not significant in the non-parametric analysis (table 4). Also, only in FIRKO, the 30, 60- and 120-minute glucose was not significantly different in the knockouts than the controls (figure 3). It is notable that in none of LIRKO experiments the fasting sugar was significantly higher than the controls. This contradicts the classical belief that liver insulin resistance is mainly responsible for fasting hyperglycemia in T2D (Bock et al., 2007; Johnson et al., 1972).

A possible problem in comparing fasting glucose across different studies was that different fasting intervals have been used ranging from 4 to 16 hours. No study clearly reported how much time is required to reach a steady state in a knockout. In 10 of the experiments in which fasting time was reported as 16 hours, none had fasting sugar significantly different from controls. In the 13 experiments in which it was high, the fasting duration was between 4 to 12 hours or not precisely reported. Therefore, it is likely that in at least some of the experiments, glucose steady state was not yet achieved at the time point defined as fasting. This bias increases the probability that higher fasting glucose is reported for the knockouts. However, since we do not see a significant difference in the collective analysis, the inference that IRKO does not alter fasting glucose is unlikely to be a result of the bias. In fact, any possible correction to the bias might further reduce the apparent residual difference. Therefore, in spite of some inconsistency across studies, a robust generalization is that IRKOs have significantly increased plasma glucose over controls at 30 to 120 minutes post glucose load but they do not appear to affect steady state fasting glucose. The time required to reach the steady state is evidently increased.

**Table 4:** Collective analysis of the fasting and post-feeding glucose levels in the control and IRKOs: The table shows, out of the total number of experiments used for the analysis, in how many the mean of the knockouts (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa. For fasting glucose, the difference is not significant in majority of studies and where there is statistical significance there is lack of consistency across studies. However, at 30, 60 and 120 minutes the knockouts have consistently elevated levels of glucose as compared to the corresponding controls.

**Figure 3:** Glucose levels for control (black squares) and **FIRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the

shortlisted papers in which control and FIRKO were compared using an OGTT. FIRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm 95\%$  CI. Glucose levels of control are expressed as  $0 \pm 95\%$  CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes and (E) 120 minutes.

**Figure 4:** Glucose levels for control (black squares) and **MIRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and MIRKO were compared using an OGTT. MIRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm 95\%$  CI. Glucose levels of control are expressed as  $0 \pm 95\%$  CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that fasting glucose does not differ from the control in any of the experiments.

**Figure 5:** Glucose levels for control (black squares) and **LIRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and LIRKO were compared using an OGTT. LIRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm 95\%$  CI. Glucose levels of control are expressed as  $0 \pm 95\%$  CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that inconsistent with classical belief, liver specific insulin receptor knockout does not show significant effect on fasting glucose in any of the experiments. On the other hand, post load glucose is consistently higher.

**Figure 6:** Glucose levels for control (black squares) and  $\beta$ IRKO (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and  $\beta$ IRKO were compared using an OGTT.  $\beta$ IRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm 95\%$  CI. Glucose levels of control are expressed as  $0 \pm 95\%$  CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that fasting glucose does not differ from the control in any of the experiments.

## 2. Systematic review of experiments to chronically raise or lower insulin levels

### 2.1 Increase in insulin by suppression of IDE

We found 6 publications that described 18 experiments that allowed comparison of GTT between raised insulin groups and control group (table 2 of supplementary information 1).

Analysis revealed no significant difference in the fasting glucose. In only one out of 18 experiments the treatment group had lower fasting glucose than the control. During the GTT curve, at 90 and 120 minutes the difference between treatment and control were significant but in the opposite direction of the expectation (table 5, figure 7). While rise in insulin level should reduce plasma glucose, it increased in 15 out of 18 studies, two of which were individually significant and the difference was significant in non-parametric collective analysis (table 5, figure 7). Across all time points along the GTT, the plasma glucose in the treated group was greater than the control group in majority of the experiments. Thus, in this class of experiments increasing insulin failed to reduce glucose at the steady state as well as post glucose load.

**Table 5:** Analysis of the fasting and post-feeding glucose levels in the control and **IDE-inhibition**: The table shows, out of the total number of experiments used for the analysis, in how many the mean of the IDE inhibition models (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa. There is no significant difference in the control at treated fasting glucose level. At 90 and 120 minutes the difference between the control and treated was significant, but in the opposite direction.

**Figure 7:** Glucose levels for control (black squares) and **IDE-inhibition** (red squares) models at steady and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and IDE-inhibition were compared using an OGTT. IDE-inhibition glucose levels are normalized to that of the control and the difference is expressed with  $\pm 95\%$  CI. Glucose levels of control are expressed as  $0 \pm 95\%$  CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points (B to F) post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that fasting glucose does not differ significantly from the control. At 90 and 120 minutes the trend is higher mean glucose than control which is contrary to the expectation in an experiment with sustainable rise in insulin.

## 2.2 Decrease in insulin: Suppression by diazoxide or octreotide

We found 8 papers describing 14 experiments for diazoxide treatment and 10 papers with 15 experiments for octreotide treatment (tables 3 and 4 from supplementary information 1 respectively). It can be seen from table 6 and figure 8 that for both insulin suppressing agents, suppression of insulin did not result into increased fasting glucose. Further at 120 minutes post glucose load there was a marginally significant rise in glucose in the insulin suppressed group as compared to control group. This demonstrates that pharmacological suppression of insulin was

unable to raise plasma glucose level in a fasting steady state. There was somewhat inconsistent but significant rise post glucose load. Diazoxide is the only agent in which both rodent and human data are available. The results of the two are very similar and separating them does not alter the significance.

**Table 6:** Analysis of the fasting and post-feeding glucose levels in the control and **treatment with Diazoxide or Octreotide:** The table shows, out of the total number of experiments used for the analysis, in how many the mean of the treated (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa.

**Figure 8:** Glucose levels for control (black squares) and treated (red squares) models at steady and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and treated were compared using an OGTT. Glucose levels after treatment with Diazoxide/Octreotide are normalized to that of the control and the difference is expressed with  $\pm 95\%$  CI. Glucose levels of control are expressed as  $0 \pm 95\%$  CI. For Diazoxide treatment, steady state is represented by the fasting glucose (A) and the perturbed state is represented by (B) at 120 minutes post glucose load. For Octreotide treatment, steady state is represented by the fasting glucose (C) and the perturbed state is represented by (D) at 120 minutes post glucose load. Note the inconsistencies across studies.

We found more means of insulin suppression in which GTT after suppression was reported, but there were not many published replications of the experiments coming independently from different research groups. Therefore, any systematic review was not warranted. We briefly review their results here.

### 2.3 Suppression by Protein restriction:

Dietary protein deprivation is another method of insulin suppression. This also led to a decrease in plasma insulin levels; however fasting glucose levels did not increase (Schteingart et al., 1979).

**2.4 Suppression by insulin siRNA:** Transgenic mice for insulin-siRNA along with IDE overexpression, showed decreased levels of insulin. Again the fasting glucose levels remained normal while there was a change in glucose tolerance curve (figure 9) (Hwang et al., 2007). The

curves in figure 9 are typical of insulin receptor knockout or insulin suppression experiments where, in individuals with impaired insulin signalling the glucose peak is higher which returns to steady state much later than the controls, but the fasting steady state level is not different.

**Figure 9:** Intra peritoneal glucose tolerance test of control and siRNA treated mice. ). Fasting glucose levels in both the siRNA treated (represented by solid lines) and untreated group (represented by dotted lines) remain unaltered in male (represented by black lines) and female (represented by grey lines) mice. 15 minutes after the glucose injection, the treated mice show higher glucose levels relative to the untreated mice in both the male and female groups; and this effect is seen throughout till 120 minutes. Figure redrawn from data by (Hwang et al., 2007).

## 2.5 Suppression of insulin by partial gene ablation:

In rodents, there are two insulin genes *Ins1* and *Ins2*(Duvillie et al., 1997). A double knockout of both the genes results in death, but ablation of either of the genes does not alter the glucose tolerance significantly suggesting redundancy (Mehran et al., 2012). There are studies in which one gene is completely knocked out and the other one is a heterozygote (Dionne et al., 2016; Mehran et al., 2012; Page et al., 2019; Templeman et al., 2015, 2017). Reduced insulin gene dosage did not consistently result into fasting hyperglycemia in these studies although it offered protection against some of the effects of hyperinsulinemia.

## 3.Steady state versus perturbed state glucose in STZ rats

Among the STZ treated rats, all the animals showed significantly higher post load glucose than the control group on all the 12 days sampled. However, in 10 out of the 12 days the 16-hour fasting glucose was not significantly different from the control although the variance was substantially greater than that of the control (figure 10).

A close look at the time course of fasting in the two groups revealed that in 4 out of 9 STZ animals the glucose levels reached the normal range but with substantial delay as compared to control animals. In two more animals the levels did not reach the normal range till 16 hours but a monotonic decrease continued throughout the period, indicating that their blood glucose may not have reached a steady state in 16 hours. Only in 3 animals the 16-hour glucose was higher than the control range with some indications of stabilizing at a higher level. In the time course

experiment, the animals are handled frequently leading to some unavoidable stress, which may have influenced the glucose levels. In the 12 day follow up experiment, plasma glucose is estimated only after 16 hours and here there is no significant difference in the control and STZ animals on 10 out of 12 days. Furthermore, the individuals that showed higher 16 hour fasting glucose did not do so consistently. In the 12 days follow up, the distribution of 16-hour fasting glucose was typically skewed with one or two outliers having high glucose levels. Interestingly the outliers were not the same animals every day. There was considerable day to day variation in individuals and averaged over the 12 days, none of the STZ animals showed significantly higher fasting glucose than the controls although they consistently showed higher post feeding glucose. Thus, these experiments show on the one hand that STZ treatment failed to increase steady state glucose levels significantly and consistently. On the other, the STZ animals took substantially longer and rather unpredictable time to reach a steady state and even at 16-hours of fasting, all individuals need not have attained a steady state. These results warrant caution against considering fixed hours fasting glucose as steady state glucose in experimental or epidemiological data. While in healthy individuals it is well established that following glucose load a steady state is regained in about two hours, it is possible that in experimental impairment of insulin signalling or in clinical diabetes, plasma glucose takes substantially longer time to reach a steady state and overnight fasting need not represent a steady state in all cases.

**Figure 10:** Treatment of SD rats with STZ. (A): The 16-hour fasting and post-meal glucose values of treated (STZ 50mg/kg) and control rats (Citrate buffer CB) over 12 days. N= 12 for each group. Note that on 10 out of 12 days the mean fasting glucose of the treated group (STZ-F) was not significantly different from the control mean (CB-F). Post feeding the treated group (STZ-P) has substantially greater mean than the control (CB-P). (B) Time course of glucose during 16 hours fasting. X axis represents the time after removal of food when the glucose readings are taken and Y axis represent the glucose levels. The grey band represents the upper and lower bounds of 95% CI of the control group with the mean glucose values represented by filled circles. Filled squares represent individuals that showed a monotonic decrease in glucose levels. In three animals the glucose levels reduced at or below the control levels and in two others they showed a continued monotonic decrease but did not reach the normal level in 16 hours. Filled triangles with dotted lines represent the individual time courses of the three STZ treated rats which showed some indications of stabilizing at a steady state above the normal.

#### 4. Theoretical and mathematical considerations

In glucose homeostasis, in a fasting state, liver glucose production is analogous to the inlet tap and tissue glucose uptake analogous to the size of the outlet (figure 2), both being a function of insulin signalling. Most models of glucose regulation assume CSS (Bergman, 1989; Bergman, 2005; Makroglou et al., 2006; Matthews et al., 1985; Tomasi et al., 1967; Turner et al., 1979; Palumbo et al., 2013). According to the CSS, the steady state/fasting level of glucose (level of water in the water tank analogy) is decided by the difference between the inlet tap (liver glucose production) and outlet (tissue glucose uptake) (figure 2A). Both liver glucose production and tissue glucose uptake depend on insulin signalling, hence a change in insulin signalling (change in inlet and outlet flow rates) would result in an altered level of glucose (steady-state water level would change).

It has not been critically examined whether CSS or TSS describes glucose homeostasis more appropriately. This is important because if TSS model is appropriate, insulin resistance and relative insulin deficiency will not result into altered steady state glucose levels although the time required for reaching a steady state after perturbation might change. Comparing the TSS with the water tank analogy, if insulin resistance affects the liver glucose production (inlet tap) or the tissue glucose uptake (outlet tap), because of the presence of upper and lower set points, insulin resistance and the relative insulin insufficiency might change the time required to attain the steady state, but not the steady state itself (figure 2B). If CSS model is appropriate, insulin resistance or altered insulin levels are bound to change fasting glucose levels. The failure of insulin receptor knockouts and insulin suppression experiments to alter the fasting steady state, along with the delay in reaching the steady state indicates that TSS model is likely to describe glucose homeostasis more appropriately. The TSS model requires mechanisms of sensing any departure from the targeted steady state. Such mechanisms are not known in peripheral systems but glucose sensing neurons are certainly known to be present in the brain. Therefore, if TSS is a more appropriate model, the CNS mechanisms are likely to be central to glucose homeostasis, particularly in determining the steady state levels; whereas insulin signalling would play a role in determining the rate at which a steady state is reached after perturbation.

### **Testable predictions from the models:**

It is possible to make other testable predictions of TSS and CSS models. In the normal healthy individual, increased glucose utilization is expected to decrease fasting glucose levels by the CSS



model but not by the TSS model. Human experiments have shown that sustained exercise does not reduce plasma glucose, in fact it might increase (Coggan, 1991). In order to match with experimental data, CSS based models of glucose dynamics during exercise need to include additional terms which involve neuronal mechanisms such as direct stimulation of liver glucose production in response to exercise through sympathetic route (Roy & Parker, 2006). This brings the model close to a TSS model. If TSS model describes glucose homeostasis more appropriately, reduced insulin signalling is not expected to change steady state glucose but only alter the time course to reach a steady state.

The mechanism of attaining a hyperinsulinemic normoglycemic prediabetic state is different by the CSS and TSS models. By the classical CSS based pathway, obesity induced insulin resistance is believed to be primary. The insulin resistance reduces glucose uptake and the excess glucose triggers a compensatory insulin response. The resultant hyperinsulinemia compensates for insulin resistance keeping the fasting glucose levels normal. Detailed analysis of the model and matching its prediction with empirical data has refuted this model (Chawla et al., 2018). One of the intuitively appealing reasons for this refutation is that after the heightened insulin levels normalize glucose, there is no reason why insulin levels remain high. Therefore, a steady state with hyperinsulinemia and normoglycemia is impossible by the CSS model but it exists in a prediabetic state. If a “compensatory” insulin response is mediated by glucose, one would expect a positive correlation between fasting glucose (FG) and fasting insulin (FI) and no correlation between insulin resistance and  $\beta$  cell responsiveness.

By the TSS model, on the other hand, compensatory response is possible in either way. Primary insulin resistance may increase the glucose levels transiently, but when glucose sensing mechanisms detect the change a compensatory response can be operational. By this mechanism a hyperinsulinemic normoglycemic state is possible. Alternatively, primary hyperinsulinemia (Corkey, 2012; Garvey et al., 1986; Shanik et al., 2008; Weyer et al., 2000) can also be compensated by increased insulin resistance by hitting the lower level of sensing which would trigger compensatory insulin resistance. Even in this case a hyperinsulinemic normoglycemic state is possible. Both glucose sensing neurons and neuronal regulation of insulin release and liver glucose production are well known. In the compensatory response mediated by TSS

pathways there need not be a correlation between fasting insulin and fasting glucose, but insulin resistance and  $\beta$  cell response would be correlated.

Also using a simple CSS model (see supplementary information 2 for details), simulations show that the correlation coefficient and regression slope in the insulin-glucose relationship would remain the same in the fasting as well as post-meal state although the range of glucose and insulin levels will be different. On the other hand, in a TSS model the post-meal glucose and insulin levels are expected to be correlated but the steady state levels may not. We test these predictions by the two alternative models using human epidemiological data below.

We argued above that since on impairment of insulin signalling, the time required to reach a steady state can be substantially longer, overnight fasting may not ensure a steady state in all individuals. Fasting hyperglycaemia in T2D can have two alternative (but not mutually exclusive) causes. Either it represents the failure to reach a steady state in the specified fasting period, or it is because of mechanisms other than reduced insulin action. The TSS model can make differential predictions from the two alternative causes since it predicts a positive correlation between plasma glucose and plasma insulin in the post-meal state but loss of this correlation on reaching a steady state. In population data, if some individuals have reached a steady state but a few others haven't we would expect a correlation significantly weaker than the post-meal correlation. These predictions can be tested in epidemiological data.

## **5. Analysis of insulin glucose relationship in steady and perturbed state in human data: Epidemiological inquiry**

In all the three data sets there was weak ( $R^2$  range 0.017 to 0.057) but significant correlation between fasting glucose (FG) and fasting insulin (FI) and strong correlation between HOMA-IR and HOMA- $\beta$  ( $R^2$  range 0.20 to 0.83) (figure 11).

**Figure 11:** The Fasting Glucose-Fasting Insulin, Post-meal Glucose-post-meal Insulin and HOMA-IR HOMA- $\beta$  scatter plots in non-diabetic populations in the three data sets. The FG-FI correlation is weak as compared to post-

meal correlation. The HOMA-IR and HOMA- $\beta$  correlations are very strong in all the three data sets, which is not expected by the classical insulin resistance theory.

It was seen in all three data sets that the correlation coefficients for glucose and insulin were an order of magnitude higher in the post-meal cross sectional data than in the fasting state (Table 7 and fig 11). Also, the regression slopes in the post-meal data were substantially different from fasting data unlike what is expected by the CSS model (Supplementary information 2). By both the sets of predictions the CSS model predictions are rejected. The HOMA-IR HOMA- $\beta$  correlation, as well as the difference between the regression correlation parameters between fasting and post-meal data are compatible with predictions of the TSS model. However, although weak, there is significant correlation between FG and FI unlike what may be expected by a steady state TSS model. This incompatibility is not sufficient to falsify the TSS model since the failure of a small proportion of individuals to have reached a steady state at overnight fasting is sufficient to explain the weak correlation. It is also likely that the assumption of fasting may not be true for the entire sample. Even if a small number of individuals do not comply with the overnight fasting instructions, a positive correlation can result and this possibility is extremely difficult to exclude in human data.

The support of TSS model over CSS model is important because it accounts for the failure of impairment of insulin signalling to alter fasting glucose but increase only post load glucose.

**Table 7:** Correlation and regression parameters of glucose-insulin relationship at steady and perturbed states.

## Discussion

The five approaches examined above fail to support the classical belief about glucose insulin relationship. The insulin receptor knock-out experiments and insulin suppression or enhancement experiments converge to show that alteration in insulin levels or insulin sensitivity does not change the steady state glucose levels. Evidence that it changes the shape of the glucose curve after food intake or glucose loading is more convincing in spite of some inconsistency across different experiments. Typically return to the steady state is delayed by impaired insulin

signalling but the steady state glucose level remains unchanged. Convergence of experiments using other means of causing specific alterations in insulin action strengthens the inference. A number of mathematical models attempt to capture the dynamics of glucose homeostasis. A good model should be able to explain all the empirical results summed up here namely the inability of insulin receptor knockouts, insulin suppression and insulin enhancement experiments to alter steady state glucose levels; the difference in the regression correlation parameters between insulin and glucose in the steady versus perturbed state; the extremely weak correlation between fasting glucose and fasting insulin, but very strong correlation between HOMA-IR and HOMA- $\beta$ ; the hyperinsulinemic-normoglycemic prediabetic state and the phenomenon of impaired glucose tolerance but normal fasting glucose. Reviewing models of glucose homeostasis is beyond the scope of this paper, but we outline here what a good model of glucose homeostasis needs to explain. In our observation, all existing models explain only some of the empirical findings. We suggest here that this inability is because of a questionable common baseline assumption of all models that insulin signalling determines the glucose level in the fasting as well as post feeding conditions. It should be possible to construct a model which is compatible with all experimental and epidemiological findings, if we realize that insulin affects glucose only in the post feeding but not in fasting conditions.

It is difficult to defend the classical assumptions about glucose-insulin relationship against the multiple convergent lines of evidence. Although results of these experiments have been there in the published literature for about two decades, these results were mostly explained away giving different excuses for different sets of experiments. The possible lines of defense would include difference between homeostatic mechanisms in rodents and humans or the possibility of non-linear nature of glucose-insulin relationship. The evidence reviewed here comes from rodents as well as humans and the glucose insulin scatters do not show any clear indication of non-linearity. Further it would be prudent to avoid making inferences based on dietary or other complex interventions since they can have multiple mechanisms of action. Specific genetic or molecular interventions are more revealing with respect to the underlying mechanisms since we can be more confident about their specificity of action. Therefore our inference that insulin action does not influence fasting glucose levels is the most straightforward and parsimonious inference. Any

other explanations will have to be supported by giving evidence for the assumptions made in those explanations.

The failure of experimental alteration in insulin signalling to alter steady state glucose raises two distinct possibilities about fasting hyperglycaemia in T2D. One is that fasting hyperglycaemia in T2D is a result of processes independent of insulin signalling such as autonomic signalling or other insulin independent mechanisms. The sympathetic tone is known to be altered in metabolic syndrome (Thorp & Schlaich, 2015) and increased sensitivity of liver to sympathetic signal is likely to be mainly responsible to fasting hyperglycaemia (Bruce et al., 1992). The other possibility is that with impaired insulin signalling overnight fasting is not sufficient to reach a steady state, therefore fasting hyperglycaemia in T2D is a non-steady state phenomenon in type 2 diabetes. The considerably weaker but still significant correlation between glucose and insulin in fasting as compared to post glucose load data suggests that both the factors are likely to be operational differentially in different individuals.

In either case certain fundamental concepts in our understanding of T2D need to be revised. First of all, the definition and measurement of insulin resistance using steady state glucose and insulin levels needs to be questioned. Most commonly used indices of insulin resistance are based on the assumption that insulin signalling decides the fasting steady state glucose levels, although non-equilibrium methods of assessing insulin resistance have been described (Patarrão et al., 2012). In the classical view other mechanisms of glucose regulation are assumed to be absent or non-significant. If increased sympathetic signalling increases liver glucose production, HOMA-IR will still account it as “insulin resistance”. The same is true about insulin resistance measured by hyperinsulinemic euglycemic clamp. The way insulin resistance is measured at the clinical level eliminates the chance of separately accounting for other mechanisms of glucose regulation. Even when experiments show that certain agents affect glucose dynamics independent of insulin action, they are typically labelled as “insulin sensitizing” agents (Hossain et al., 2018). As a result, the belief that insulin is the only mechanism of glucose regulation relevant to T2D is artificially strengthened. There is a subtle circularity in the working definition of insulin resistance. Insulin resistance is blamed for the failure of normal or elevated levels of insulin to regulate glucose. In order to test this hypothesis, we should have an independent definition and

measure of insulin resistance. Only then we can test whether and to what extent insulin resistance can alter glucose dynamics. However, clinically insulin resistance is measured by the inability of insulin to regulate glucose. Such a measure cannot be used to test the hypothesis that insulin resistance leads to the failure of insulin to regulate glucose. The unfalsifiability of the insulin resistance hypothesis arising out of this circularity has halted any attempts towards realistic assessments of the true causes of fasting hyperglycaemia in type 2 diabetes. In the molecular approach to induce insulin resistance, we have an independent definition and causality for insulin resistance and therefore such experiments are free from circularity of definition. The results of such experiments reviewed here are therefore more revealing and reliable. Since all of them converge to show that altering insulin signalling does not alter steady state glucose levels, the insulin resistance and inadequate compensation hypothesis for steady state hyperglycaemia stands clearly rejected.

The question can be turned upside down to examine whether steady state glucose level determines steady state insulin. If glucose is infused with a constant rate over a long time, insulin levels will come back to the baseline levels if glucose is not a determinant of fasting insulin. If it is, then insulin levels will stabilize at a new heightened steady state level. Jetton et al. (2008) infused intra venous glucose (20% glucose w/v) continuously for 4 days in rats. Both glucose and insulin levels increased significantly after the infusion. However, later both glucose and insulin levels came back to normal even as the infusion continued. Increase in the concentration of the infused glucose (up to 35%) also yielded similar results (Steil et al., 2001). Thus, immediately on perturbation, glucose affected insulin levels, however after allowing sufficient time to regain steady state, the continued infusion of glucose had no significant effect on insulin levels. This demonstrates that even glucose does not hold a causal relationship with insulin in a steady state whereas glucose level perturbation is certainly known to stimulate insulin response. The evidence reviewed above indicates that insulin is a driver but not a navigator of glucose homeostasis. A non-zero level of insulin is required for reaching a homeostatic steady state. In type 1 diabetes, the almost complete absence of insulin prevents glucose homeostasis. In type 2 diabetes there are non-zero insulin levels and therefore, a steady state is possible, but insulin itself plays little role in deciding the steady state glucose level. It is more likely that neuronal and other hormonal-metabolic factors affect the steady state glucose in T2D.

# Conclusions

From the insulin-glucose lesson we can learn certain general principles and norms for making causal inferences from experimental results. There are certainly more examples and implications of the driver navigator distinction than the ones captured by the insulin example. We see these phenomena in simple physico-chemical systems as well as in complex biological systems. An azeotropic mixture or a constant boiling solution has a steady state proportion of two or more liquids. If one starts from a non-equilibrium mixture, the rate of heating affects the time required to reach the azeotropic equilibrium but does not affect the equilibrium composition. This is because the molecular dynamics that decides the equilibrium has more complex elements than simple evaporation(D. Li et al., 2020) It is possible that in a given system some causal factors affect both steady and perturbed states where as others affect only one of them. For example, stirring increases the rate of dissolution of a solute, but does not affect the saturation concentration. Heating, on the other hand, can affect both.

Some of the well studies examples in complex systems include the regulation of plasmid numbers in bacteria. The rate of replication of plasmids does not decide the stable plasmid copy number since there is a complex set of mechanisms that decide the steady state independent of the rate of DNA replication (M. M. Watve et al., 2010).

The differences between driver and navigator causes may also be specific to the context. For example, the rate of lexicon building at an early age may or may not be determining the adult vocabulary and language skills depending upon what is being tested (F. Hill et al., 2017; Hurtado et al., 2008; Peter et al., 2019)

In warm blooded animals, in a healthy state, the body temperature is maintained constant by homeostatic mechanisms. If the environmental temperature changes suddenly, heat transfer between the body and the environment is inevitable and as a result the body temperature will change transiently. However, the homeostatic loops will be operative and bring the body temperature back to normal soon. If an experimenter measures the body temperature immediately after changing the environmental temperature, a causal relationship might be apparent. If the body temperature is measured after reaching the steady state, one may infer that environmental temperature has no causal relationship with body temperature. If the experimenter is measuring heat transfer as the mechanism of causation, then it will be demonstrable at any

time. So, a causal mechanism exists all the time but demonstration of a causal mechanism or pathway is not sufficient to establish a causal role.

These set of examples demonstrate that causation may work differently in steady versus perturbed states and the need to differentiate between driver and navigator causality is widespread in different fields of science. There is a need for a detailed philosophical account on this issue as well as a set of working norms for practicing science. We suggest a few norms specifically for the field of experimental physiology.

Certain kinds of experimental interventions are unable to distinguish between driver versus navigator causality. Knocking out a driver or a navigator will disable the journey to the destination. Therefore, complete knockout of a cause may not distinguish between driver and navigator causality. On the other hand, experiments quantitatively altering the level of the causal factor while keeping it non-zero has a different set of implications. If a perturbation is momentary or transient, the results obtained would certainly reflect perturbed state causality, but may not reflect steady state causality. On the other hand, sustained perturbations held constant for sufficiently long to allow the system to regain a steady state are necessary to establish steady state causality. If upon sustainably altering a causal factor the effect variable returns to the same steady state, it reflects only perturbed state and not steady state causality. If, on the other hand, sustained alteration in the causal factor results into an altered steady state, it indicates steady state causality. A sub-normal driver will delay the time to destination but will not change the destination. On the other hand, changing the navigator may or may not alter the time, but will alter the position of the destination. In the history of insulin research, early experiments such as total pancreatectomy demonstrated the necessary role of insulin in glucose homeostasis but the distinction between driver or navigator causality was not even conceptually perceived. So, it was assumed that insulin does both the roles. Although the absence of correlation between fasting glucose and insulin but good correlation after perturbation was noted as early as 1969 (Goodner et al., 1969) in the absence of conceptual differentiation between steady state and perturbed state causality, a clear interpretation did not emerge. Now in the presence of multiple experiments showing the precise role of insulin, we need to revive our concepts of causality. Although the experiments reviewed above have been there in the literature, the inability of insulin to alter



fasting glucose was not appreciated before, only because the distinction between driver and navigator causality was not there in the philosophical foundation of biological causality.

For any homeostatic system in which there can be one or more feedback loops working as homeostatic mechanisms and one or many possible perturbations, all the perturbations can have a demonstrable causal role in a non-equilibrium state but only some of the components of the feedback loops may have steady state causal relationship with the controlled parameter. If there are more than one homeostatic mechanisms operating on a controlled variable, it is possible that components of any one loop do not affect the steady state level of the variable. This distinction is important in building the philosophy of experimental physiology and medicine. Most of the experiments in physiology are perturbation experiments and often we try to understand the steady state in a homeostatic system using such experiments. This situation warrants care in making inferences in the absence of which our understanding of physiology can remain flawed.

## Acknowledgements

We would like to thank Prof. Raj Bhopal, University of Edinburgh for providing clinical data from the Newcastle Heart Project. We would also like to thank Prof. ChittaranjanYajnik, KEM Hospital, Pune for providing the clinical data from the CRISIS study. We are immensely grateful for the critical and insightful comments they provided on the earlier version of the manuscript. We would like to acknowledge Indian Institute of Science Education and Research for providing the funding for this project. We would also like to thank Geetanjali Nerurkar for her help during the animal experiments.

## References

- ACCORD. (2008). Effects of Intensive Glucose Lowering in Type 2 Diabetes. *New England Journal of Medicine*, 358(24), 2545–2559. <https://doi.org/10.1056/NEJMoa0802743>
- Akbarzadeh, A., Norouzzian, D., Mehrabi, M. R., Jamshidi, S., Farhangi, A., Verdi, A. A., Mofidian, S. M. A., & Rad, B. L. (2007). Induction of diabetes by Streptozotocin in rats. *Indian Journal of Clinical Biochemistry*, 22(2), 60–64. <https://doi.org/10.1007/BF02913315>
- AKHTAR, A. (2015). The Flaws and Human Harms of Animal Experimentation. *Cambridge Quarterly of Healthcare Ethics*, 24(04), 407–419. <https://doi.org/10.1017/S0963180115000079>
- Ali, Z., Chandrasekera, P. C., & Pippin, J. J. (2018). Animal research for type 2 diabetes mellitus, its limited translation for clinical benefit, and the way forward. *Alternatives to*

- 1015 *Laboratory Animals : ATLA*, 46(1), 13–22. <http://www.ncbi.nlm.nih.gov/pubmed/29553794>
- 1016 Asrafuzzaman, M., Cao, Y., Afroz, R., Kamato, D., Gray, S., & Little, P. J. (2017). Animal
- 1017 models for assessing the impact of natural products on the aetiology and metabolic
- 1018 pathophysiology of Type 2 diabetes. *Biomedicine & Pharmacotherapy*, 89, 1242–1251.
- 1019 <https://doi.org/10.1016/j.biopha.2017.03.010>
- 1020 Authier, F., Posner, B. I., & Bergeron, J. J. (1996). Insulin-degrading enzyme. *Clinical and*
- 1021 *Investigative Medicine. Medecine Clinique et Experimentale*, 19(3), 149–160.
- 1022 <http://www.ncbi.nlm.nih.gov/pubmed/8724818>
- 1023 Bergman, R. N. (1989). Toward Physiological Understanding of Glucose Tolerance: Minimal-
- 1024 Model Approach. *Diabetes*, 38(12), 1512–1527. <https://doi.org/10.2337/diab.38.12.1512>
- 1025 Bergman, Richard N. (2005). Minimal Model: Perspective from 2005. *Hormone Research in*
- 1026 *Paediatrics*, 64(3), 8–15. <https://doi.org/10.1159/000089312>
- 1027 Bernard, C. (1879). Leçons de physiologie opératoire. *Librairie J.-B. Baillie et Fils*, 650.
- 1028 Bevan, P. (2001). Insulin signalling. *Journal of Cell Science*, 114(Pt 8), 1429–1430.
- 1029 <http://www.ncbi.nlm.nih.gov/pubmed/11282018>
- 1030 Bhopal, R. S. (2013). A four-stage model explaining the higher risk of Type 2 diabetes mellitus
- 1031 in South Asians compared with European populations. *Diabetic Medicine*, 30(1), 35–42.
- 1032 <https://doi.org/10.1111/dme.12016>
- 1033 Bhopal, R., Unwin, N., White, M., Yallop, J., Walker, L., Alberti, K. G. M. M., Harland, J.,
- 1034 Patel, S., Ahmad, N., Turner, C., Watson, B., Kaur, D., Kulkarni, A., Laker, M., &
- 1035 Tavridou, A. (1999). Heterogeneity of coronary heart disease risk factors in Indian,
- 1036 Pakistani, Bangladeshi, and European origin populations: cross sectional study. *BMJ*,
- 1037 319(7204), 215–220. <https://doi.org/10.1136/bmj.319.7204.215>
- 1038 Blüher, M., Michael, M. D., Peroni, O. D., Ueki, K., Carter, N., Kahn, B. B., & Kahn, C. R.
- 1039 (2002). Adipose tissue selective insulin receptor knockout protects against obesity and
- 1040 obesity-related glucose intolerance. *Developmental Cell*. [https://doi.org/10.1016/S1534-](https://doi.org/10.1016/S1534-5807(02)00199-5)
- 1041 [5807\(02\)00199-5](https://doi.org/10.1016/S1534-5807(02)00199-5)
- 1042 Bock, G., Chittilapilly, E., Basu, R., Toffolo, G., Cobelli, C., Chandramouli, V., Landau, B. R.,
- 1043 & Rizza, R. A. (2007). Contribution of Hepatic and Extrahepatic Insulin Resistance to the
- 1044 Pathogenesis of Impaired Fasting Glucose: Role of Increased Rates of Gluconeogenesis.
- 1045 *Diabetes*, 56(6), 1703–1711. <https://doi.org/10.2337/db06-1776>
- 1046 Boudon, R. (1965). A Method of Linear Causal Analysis: Dependence Analysis. *American*
- 1047 *Sociological Review*, 30(3), 365. <https://doi.org/10.2307/2090717>
- 1048 Bracken, M. B. (2009). Why animal studies are often poor predictors of human reactions to
- 1049 exposure. *Journal of the Royal Society of Medicine*, 102(3), 120–122.
- 1050 <https://doi.org/10.1258/jrsm.2008.08k033>
- 1051 Bruce, D. G., Chisholm, D. J., Storlien, L. H., Kraegen, E. W., & Smythe, G. A. (1992). The
- 1052 effects of sympathetic nervous system activation and psychological stress on glucose
- 1053 metabolism and blood pressure in subjects with Type 2 (non-insulin-dependent) diabetes
- 1054 mellitus. *Diabetologia*, 35(9), 835–843. <https://doi.org/10.1007/BF00399929>
- 1055 Butler, A. E., Janson, J., Bonner-Weir, S., Ritzel, R., Rizza, R. A., & Butler, P. C. (2003). -Cell
- 1056 Deficit and Increased -Cell Apoptosis in Humans With Type 2 Diabetes. *Diabetes*, 52(1),
- 1057 102–110. <https://doi.org/10.2337/diabetes.52.1.102>
- 1058 Carruthers, A., DeZutter, J., Ganguly, A., & Devaskar, S. U. (2009). Will the original glucose
- 1059 transporter isoform please stand up! *American Journal of Physiology-Endocrinology and*
- 1060 *Metabolism*, 297(4), E836–E848. <https://doi.org/10.1152/ajpendo.00496.2009>

- Chawla, S., Pund, A. B., V., Kulkarni, S., Diwekar-Joshi, M., & Watve, M. (2018). Inferring causal pathways among three or more variables from steady-state correlations in a homeostatic system. *PLOS ONE*, 13(10), e0204755. <https://doi.org/10.1371/journal.pone.0204755>
- Clark, A., Saad, M. F., Nezzar, T., Uren, C., Knowler, W. C., Bennett, P. H., & Turner, R. C. (1990). Islet amyloid polypeptide in diabetic and non-diabetic Pima Indians. *Diabetologia*, 33(5), 285–289. <https://doi.org/10.1007/BF00403322>
- Clemmons, D. R. (2004). Role of Insulin-Like Growth Factor I in Maintaining Normal Glucose Homeostasis. *Hormone Research in Paediatrics*, 62(1), 77–82. <https://doi.org/10.1159/000080763>
- Coggan, A. R. (1991). Plasma Glucose Metabolism During Exercise in Humans. *Sports Medicine*, 11(2), 102–124. <https://doi.org/10.2165/00007256-199111020-00003>
- Corkey, B. E. (2012). Banting Lecture 2011: Hyperinsulinemia: Cause or Consequence? *Diabetes*, 61(1), 4–13. <https://doi.org/10.2337/db11-1483>
- Costes, S., & Butler, P. C. (2014). Insulin-Degrading Enzyme Inhibition, a Novel Therapy for Type 2 Diabetes? *Cell Metabolism*, 20(2), 201–203. <https://doi.org/10.1016/j.cmet.2014.07.016>
- Di Dalmazi, G., Pagotto, U., Pasquali, R., & Vicennati, V. (2012). Glucocorticoids and Type 2 Diabetes: From Physiology to Pathology. *Journal of Nutrition and Metabolism*, 2012, 1–9. <https://doi.org/10.1155/2012/525093>
- Dionne, D. A., Skovsø, S., Templeman, N. M., Clee, S. M., & Johnson, J. D. (2016). *in Mice With Genetically Reduced Insulin*. 157(July), 2724–2734. <https://doi.org/10.1210/en.2016-1102>
- Doyle, M. E. (2003). Pharmacological Agents That Directly Modulate Insulin Secretion. *Pharmacological Reviews*, 55(1), 105–131. <https://doi.org/10.1124/pr.55.1.7>
- Duckworth, W. C., Bennett, R. G., & Hamel, F. G. (1998). Insulin Degradation: Progress and Potential 1. *Endocrine Reviews*, 19(5), 608–624. <https://doi.org/10.1210/edrv.19.5.0349>
- Duvillie, B., Cordonnier, N., Deltour, L., Dandoy-Dron, F., Itier, J.-M., Monthieux, E., Jami, J., Joshi, R. L., & Bucchini, D. (1997). Phenotypic alterations in insulin-deficient mutant mice. *Proceedings of the National Academy of Sciences*, 94(10), 5137–5140. <https://doi.org/10.1073/pnas.94.10.5137>
- Freitas, S. C. F., Harthmann, Â. d'Avila, Rodrigues, B., Irigoyen, M.-C., & De Angelis, K. (2015). Effect of aerobic exercise training on regional blood flow and vascular resistance in diabetic rats. *Diabetology & Metabolic Syndrome*, 7, 115. <https://doi.org/10.1186/s13098-015-0109-1>
- Gajdosík, A., Gajdosíková, A., Stefek, M., Navarová, J., & Hozová, R. (1999). Streptozotocin-induced experimental diabetes in male Wistar rats. *General Physiology and Biophysics*, 18 Spec No, 54–62. <http://www.ncbi.nlm.nih.gov/pubmed/10703720>
- Galloway, J. A., Spradlin, C. T., Nelson, R. L., Wentworth, S. M., Davidson, J. A., & Swarner, J. L. (1981). Factors Influencing the Absorption, Serum Insulin Concentration, and Blood Glucose Responses After Injections of Regular Insulin and Various Insulin Mixtures. *Diabetes Care*, 4(3), 366–376. <https://doi.org/10.2337/diacare.4.3.366>
- Garvey, W. T., Olefsky, J. M., & Marshall, S. (1986). Insulin Induces Progressive Insulin Resistance in Cultured Rat Adipocytes: Sequential Effects at Receptor and Multiple Postreceptor Sites. *Diabetes*, 35(3), 258–267. <https://doi.org/10.2337/diab.35.3.258>
- Gathercole, L. L., & Stewart, P. M. (2010). Targeting the pre-receptor metabolism of cortisol as

- 1107 a novel therapy in obesity and diabetes. *The Journal of Steroid Biochemistry and Molecular*  
1108 *Biology*, 122(1–3), 21–27. <https://doi.org/10.1016/j.jsbmb.2010.03.060>
- 1109 Goldstein, R. E., Wasserman, D. H., McGuinness, O. P., Lacy, D. B., Cherrington, A. D., &  
1110 Abumrad, N. N. (1993). Effects of chronic elevation in plasma cortisol on hepatic  
1111 carbohydrate metabolism. *American Journal of Physiology-Endocrinology and Metabolism*,  
1112 264(1), E119–E127. <https://doi.org/10.1152/ajpendo.1993.264.1.E119>
- 1113 Goodner, C. J., Conway, M. J., & Werrbach, J. H. (1969). Control of insulin secretion during  
1114 fasting hyperglycemia in adult diabetics and in nondiabetic subjects during infusion of  
1115 glucose. *Journal of Clinical Investigation*, 48(10), 1878–1887.  
1116 <https://doi.org/10.1172/JCI106154>
- 1117 Gotelli, N. J. (2008). *A Primer of Ecology*.
- 1118 Grace, J. B. (2002). Shipley, B. Cause and correlation in Biology. *Annals of Botany*, 90(6), 777–  
1119 778. <https://doi.org/10.1093/aob/mcf249>
- 1120 Granger, C. W. J. (1969). Investigating Causal Relations by Econometric Models and Cross-  
1121 spectral Methods. *Econometrica*, 37(3), 424. <https://doi.org/10.2307/1912791>
- 1122 Guerra, C., Navarro, P., Valverde, A. M., Arribas, M., Brüning, J., Kozak, L. P., Kahn, C. R., &  
1123 Benito, M. (2001). Brown adipose tissue – specific insulin receptor knockout shows  
1124 diabetic phenotype without insulin resistance. *The Journal of Clinical Investigation*, 108(8),  
1125 1205–1213. <https://doi.org/10.1172/JCI200113103>.Introduction
- 1126 Gujral, U. P., Pradeepa, R., Weber, M. B., Narayan, K. M. V., & Mohan, V. (2013). Type 2  
1127 diabetes in South Asians: similarities and differences with white Caucasian and other  
1128 populations. *Annals of the New York Academy of Sciences*, 1281(1), 51–63.  
1129 <https://doi.org/10.1111/j.1749-6632.2012.06838.x>
- 1130 Halter, J. B., Ward, W. K., Porte, D., Best, J. D., & Pfeifer, M. A. (1985). Glucose regulation in  
1131 non-insulin-dependent diabetes mellitus. Interaction between pancreatic islets and the liver.  
1132 *The American Journal of Medicine*, 79(2B), 6–12.  
1133 <http://www.ncbi.nlm.nih.gov/pubmed/2863979>
- 1134 Hill, A. B. (1965). The Environment and Disease: Association or Causation? *Proceedings of the*  
1135 *Royal Society of Medicine*, 58(5), 295–300. <https://doi.org/10.1177/003591576505800503>
- 1136 Hill, F., Clark, S., Hermann, K. M., & Blunsom, P. (2017). *Understanding Early Word Learning*  
1137 *in Situated Artificial Agents*. <http://arxiv.org/abs/1710.09867>
- 1138 Hossain, Z., Valicherla, G. R., Gupta, A. P., Syed, A. A., Riyazuddin, M., Chandra, S., Siddiqi,  
1139 M. I., & Gayen, J. R. (2018). Discovery of pancreastatin inhibitor PSTi8 for the treatment of  
1140 insulin resistance and diabetes: Studies in rodent models of diabetes mellitus. *Scientific*  
1141 *Reports*, 8(1), 1–13. <https://doi.org/10.1038/s41598-018-27018-8>
- 1142 Hulse, R. E., Ralat, L. A., & Wei-Jen, T. (2009). *Chapter 22 Structure, Function, and Regulation*  
1143 *of Insulin-Degrading Enzyme* (pp. 635–648). [https://doi.org/10.1016/S0083-](https://doi.org/10.1016/S0083-6729(08)00622-5)  
1144 [6729\(08\)00622-5](https://doi.org/10.1016/S0083-6729(08)00622-5)
- 1145 Hulse, R. E., Ralat, L. a, & Wei-Jen, T. (2009). Structure, function, and regulation of insulin-  
1146 degrading enzyme. *Vitamins and Hormones*, 80(08), 635–648.  
1147 [https://doi.org/10.1016/S0083-6729\(08\)00622-5](https://doi.org/10.1016/S0083-6729(08)00622-5)
- 1148 Hurtado, N., Marchman, V. A., & Fernald, A. (2008). Does input influence uptake? Links  
1149 between maternal talk, processing speed and vocabulary size in Spanish-learning children.  
1150 *Developmental Science*, 11(6), F31–F39. <https://doi.org/10.1111/j.1467-7687.2008.00768.x>
- 1151 Hwang, D., Seo, S., Kim, Y., Kim, C., Shim, S., Jee, S., Lee, S., Sin, J., Cho, J., Kang, B., Jang,  
1152 I., & Cho, J. (2007). Significant change in insulin production, glucose tolerance and ER

- stress signaling in transgenic mice coexpressing insulin-siRNA and human IDE. *International Journal of Molecular Medicine*. <https://doi.org/10.3892/ijmm.19.1.65>
- Jansen, C., Lundquist, I., Salehi, A., Axelson, J., & Ohlsson, B. (2006). Does Epidermal Growth Factor Participate in the Regulation of Glucose, Insulin and Glucagon Levels? *European Surgical Research*, 38(4), 377–384. <https://doi.org/10.1159/000094533>
- Jetton, T. L., Everill, B., Lausier, J., Roskens, V., Habibovic, A., LaRock, K., Gokin, A., Peshavaria, M., & Leahy, J. L. (2008). Enhanced  $\beta$ -cell mass without increased proliferation following chronic mild glucose infusion. *American Journal of Physiology-Endocrinology and Metabolism*, 294(4), E679–E687. <https://doi.org/10.1152/ajpendo.00569.2007>
- Johnson, M. E., Das, N. M., Butcher, F. R., & Fain, J. N. (1972). The regulation of gluconeogenesis in isolated rat liver cells by glucagon, insulin, dibutyryl cyclic adenosine monophosphate, and fatty acids. *The Journal of Biological Chemistry*, 247(10), 3229–3235. <http://www.ncbi.nlm.nih.gov/pubmed/4337509>
- Kagami, K., Morita, H., Onda, K., Hirano, T., & Oka, K. (2008). Protective effect of caffeine on streptozotocin-induced beta-cell damage in rats. *Journal of Pharmacy and Pharmacology*, 60(9), 1161–1165. <https://doi.org/10.1211/jpp.60.9.0007>
- Karamitsos, D. T. (2011). The story of insulin discovery. *Diabetes Research and Clinical Practice*, 93, S2–S8. [https://doi.org/10.1016/S0168-8227\(11\)70007-9](https://doi.org/10.1016/S0168-8227(11)70007-9)
- King, P., Peacock, I., & Donnelly, R. (2001). The UK Prospective Diabetes Study (UKPDS): clinical and therapeutic implications for type 2 diabetes. *British Journal of Clinical Pharmacology*, 48(5), 643–648. <https://doi.org/10.1046/j.1365-2125.1999.00092.x>
- Kovacs, B., Lumayag, S., Cowan, C., & Xu, S. (2011). microRNAs in Early Diabetic Retinopathy in Streptozotocin-Induced Diabetic Rats. *Investigative Ophthalmology & Visual Science*, 52(7), 4402. <https://doi.org/10.1167/iovs.10-6879>
- Kulkarni, S., Sharda, S., & Watve, M. (2017). Bi-stability in type 2 diabetes mellitus multi-organ signalling network. *PLOS ONE*, 12(8), e0181536. <https://doi.org/10.1371/journal.pone.0181536>
- Kuo, T., McQueen, A., Chen, T.-C., & Wang, J.-C. (2015). *Regulation of Glucose Homeostasis by Glucocorticoids* (pp. 99–126). [https://doi.org/10.1007/978-1-4939-2895-8\\_5](https://doi.org/10.1007/978-1-4939-2895-8_5)
- Lamberts, S. W. J., van der Lely, A.-J., de Herder, W. W., & Hofland, L. J. (1996). Octreotide. *New England Journal of Medicine*, 334(4), 246–254. <https://doi.org/10.1056/NEJM199601253340408>
- Lerner, R. L., & Porte, D. (1972). Acute and steady-state insulin responses to glucose in nonobese diabetic subjects. *Journal of Clinical Investigation*, 51(7), 1624–1631. <https://doi.org/10.1172/JCI106963>
- Li, C. C. (1956). The Concept of Path Coefficient and Its Impact on Population Genetics. *Biometrics*, 12(2), 190. <https://doi.org/10.2307/3001760>
- Li, D., Gao, Z., Vasudevan, N. K., Li, H., Gao, X., Li, X., & Xi, L. (2020). Molecular Mechanism for Azeotrope Formation in Ethanol/Benzene Binary Mixtures through Gibbs Ensemble Monte Carlo Simulation. *The Journal of Physical Chemistry B*, 124(16), 3371–3386. <https://doi.org/10.1021/acs.jpcc.9b12013>
- Maianti, J. P., McFedries, A., Foda, Z. H., Kleiner, R. E., Du, X. Q., Leissring, M. A., Tang, W. J., Charron, M. J., Seeliger, M. A., Saghatelian, A., & Liu, D. R. (2014). Anti-diabetic activity of insulin-degrading enzyme inhibitors mediated by multiple hormones. *Nature*, 511(7507), 94–98. <https://doi.org/10.1038/nature13297>
- Makroglou, A., Li, J., & Kuang, Y. (2006). Mathematical models and software tools for the

- 1199 glucose-insulin regulatory system and diabetes: an overview. *Applied Numerical*
- 1200 *Mathematics*, 56(3–4), 559–573. <https://doi.org/10.1016/j.apnum.2005.04.023>
- 1201 Matthews, D. R., Hosker, J. P., Rudenski, A. S., Naylor, B. A., Treacher, D. F., & Turner, R. C.
- 1202 (1985). Homeostasis model assessment: insulin resistance and  $\beta$ -cell function from fasting
- 1203 plasma glucose and insulin concentrations in man. *Diabetologia*, 28(7), 412–419.
- 1204 <https://doi.org/10.1007/BF00280883>
- 1205 Mehran, A. E., Templeman, N. M., Brigidi, G. S., Lim, G. E., Chu, K., Hu, X., Botezelli, J. D.,
- 1206 Asadi, A., Hoffman, B. G., Kieffer, T. J., Bamji, S. X., & Clee, S. M. (2012). Article
- 1207 Hyperinsulinemia Drives Diet-Induced Obesity Independently of Brain Insulin Production.
- 1208 *Cell Metabolism*, 16(6), 723–737. <https://doi.org/10.1016/j.cmet.2012.10.019>
- 1209 Meinert, C. L., Knatterud, G. L., Prout, T. E., & Klimt, C. R. (1970). A study of the effects of
- 1210 hypoglycemic agents on vascular complications in patients with adult-onset diabetes. II.
- 1211 Mortality results. *Diabetes*, 19, Suppl:789-830.
- 1212 <http://www.ncbi.nlm.nih.gov/pubmed/4926376>
- 1213 Meinshausen, N., Hauser, A., Mooij, J. M., Peters, J., Versteeg, P., & Bühlmann, P. (2016).
- 1214 Methods for causal inference from gene perturbation experiments and validation.
- 1215 *Proceedings of the National Academy of Sciences*, 113(27), 7361–7368.
- 1216 <https://doi.org/10.1073/pnas.1510493113>
- 1217 Mering, J. von J., & Minkowski, O. (1890). Diabetes mellitus nach Pankreasexstirpation. *Vogel,*
- 1218 *Leipzig*, 28.
- 1219 Messmer-Blust, A. F., Philbrick, M. J., Guo, S., Wu, J., He, P., Guo, S., & Li, J. (2012). RTEF-1
- 1220 Attenuates Blood Glucose Levels by Regulating Insulin-Like Growth Factor Binding
- 1221 Protein-1 in the Endothelium. *Circulation Research*, 111(8), 991–1001.
- 1222 <https://doi.org/10.1161/CIRCRESAHA.112.268110>
- 1223 Niles, H. E. (1923). The Method of Path Coefficients an Answer to Wright. *Genetics*, 8(3), 256–
- 1224 260. <http://www.ncbi.nlm.nih.gov/pubmed/17246012>
- 1225 Nonogaki, K. (2000). New insights into sympathetic regulation of glucose and fat metabolism.
- 1226 *Diabetologia*, 43(5), 533–549. <https://doi.org/10.1007/s001250051341>
- 1227 Page, M. M., Skovsø, S., Cen, H., Chiu, A. P., Dionne, D. A., Hutchinson, D. F., Lim, G. E.,
- 1228 Szabat, M., Flibotte, S., Sinha, S., Nislow, C., Rodrigues, B., & Johnson, J. D. (2019).
- 1229 *Reducing insulin via conditional partial gene ablation in adults reverses diet-induced*
- 1230 *weight gain*. <https://doi.org/10.1096/fj.201700518R>
- 1231 Palumbo, P., Ditlevsen, S., Bertuzzi, A., & De Gaetano, A. (2013). Mathematical modeling of
- 1232 the glucose–insulin system: A review. *Mathematical Biosciences*, 244(2), 69–81.
- 1233 <https://doi.org/10.1016/j.mbs.2013.05.006>
- 1234 Panten, U., Burgfeld, J., Goerke, F., Rennie, M., Schwanstecher, M., Wallasch, A., Zünkler, B.
- 1235 J., & Lenzen, S. (1989). Control of insulin secretion by sulfonylureas, meglitinide and
- 1236 diazoxide in relation to their binding to the sulfonylurea receptor in pancreatic islets.
- 1237 *Biochemical Pharmacology*, 38(8), 1217–1229. [https://doi.org/10.1016/0006-](https://doi.org/10.1016/0006-2952(89)90327-4)
- 1238 [2952\(89\)90327-4](https://doi.org/10.1016/0006-2952(89)90327-4)
- 1239 Patarrão, R. S., Lutt, W. W., Afonso, R. A., Ribeiro, R. T., Fernandes, A. B., Boavida, J. M., &
- 1240 Macedo, M. P. (2012). Postprandial but not fasting insulin resistance is an early identifier of
- 1241 dysmetabolism in overweight subjects. *Canadian Journal of Physiology and Pharmacology*,
- 1242 90(7), 923–931. <https://doi.org/10.1139/y2012-086>
- 1243 Peter, M. S., Durrant, S., Jessop, A., Bidgood, A., Pine, J. M., & Rowland, C. F. (2019). Does
- 1244 speed of processing or vocabulary size predict later language growth in toddlers? *Cognitive*

- 1245 *Psychology*, 115, 101238. <https://doi.org/10.1016/j.cogpsych.2019.101238>
- 1246 Pories, W. J., & Dohm, G. L. (2012). Diabetes: Have We Got It All Wrong?: Hyperinsulinism as
- 1247 the culprit: surgery provides the evidence. *Diabetes Care*, 35(12), 2438–2442.
- 1248 <https://doi.org/10.2337/dc12-0684>
- 1249 Porte, D., & Kahn, S. E. (2001). Beta-cell dysfunction and failure in type 2 diabetes: potential
- 1250 mechanisms. *Diabetes*, 50(Supplement 1), S160–S163.
- 1251 <https://doi.org/10.2337/diabetes.50.2007.S160>
- 1252 Reaven, G. M. (1988). Role of Insulin Resistance in Human Disease. *Diabetes*, 37(12), 1595–
- 1253 1607. <https://doi.org/10.2337/diab.37.12.1595>
- 1254 Roy, A., & Parker, R. S. (2006). DYNAMIC MODELING OF EXERCISE EFFECTS ON
- 1255 PLASMA GLUCOSE AND INSULIN LEVELS. *IFAC Proceedings Volumes*, 39(2), 509–
- 1256 514. <https://doi.org/10.3182/20060402-4-BR-2902.00509>
- 1257 Sakaguchi, M., Fujisaka, S., Cai, W., Winnay, J. N., Konishi, M., O'Neill, B. T., Li, M., García-
- 1258 Martín, R., Takahashi, H., Hu, J., Kulkarni, R. N., & Kahn, C. R. (2017). Adipocyte
- 1259 Dynamics and Reversible Metabolic Syndrome in Mice with an Inducible Adipocyte-
- 1260 Specific Deletion of the Insulin Receptor. *Cell Metabolism*, 25(2), 448–462.
- 1261 <https://doi.org/10.1016/j.cmet.2016.12.008>
- 1262 Schteingart, D. E., McKenzie, A. K., Victoria, R. S., & Tsao, H. S. (1979). Suppression of
- 1263 insulin secretion by protein deprivation in obesity. *Metabolism*, 28(9), 943–949.
- 1264 [https://doi.org/10.1016/0026-0495\(79\)90095-7](https://doi.org/10.1016/0026-0495(79)90095-7)
- 1265 Schwartz, M. W. (2005). Diabetes, Obesity, and the Brain. *Science*, 307(5708), 375–379.
- 1266 <https://doi.org/10.1126/science.1104344>
- 1267 Shanik, M. H., Xu, Y., Skrha, J., Dankner, R., Zick, Y., & Roth, J. (2008). Insulin Resistance and
- 1268 Hyperinsulinemia: Is hyperinsulinemia the cart or the horse? *Diabetes Care*, 31(Supplement
- 1269 2), S262–S268. <https://doi.org/10.2337/dc08-s264>
- 1270 Shen, Y., Joachimiak, A., Rich Rosner, M., & Tang, W.-J. (2006). Structures of human insulin-
- 1271 degrading enzyme reveal a new substrate recognition mechanism. *Nature*, 443(7113), 870–
- 1272 874. <https://doi.org/10.1038/nature05143>
- 1273 Softic, S., Boucher, J., Solheim, M. H., Fujisaka, S., Haering, M. F., Homan, E. P., Winnay, J.,
- 1274 Perez-Atayde, A. R., & Kahn, C. R. (2016). Lipodystrophy due to adipose tissue-specific
- 1275 insulin receptor knockout results in progressive NAFLD. *Diabetes*, 65(8), 2187–2200.
- 1276 <https://doi.org/10.2337/db16-0213>
- 1277 Steil, G. M., Trivedi, N., Jonas, J.-C., Hasenkamp, W. M., Sharma, A., Bonner-Weir, S., & Weir,
- 1278 G. C. (2001). Adaptation of  $\beta$ -cell mass to substrate oversupply: enhanced function with
- 1279 normal gene expression. *American Journal of Physiology-Endocrinology and Metabolism*,
- 1280 280(5), E788–E796. <https://doi.org/10.1152/ajpendo.2001.280.5.E788>
- 1281 Suh, J. M., Jonker, J. W., Ahmadian, M., Goetz, R., Lackey, D., Osborn, O., Huang, Z., Liu, W.,
- 1282 Yoshihara, E., van Dijk, T. H., Havinga, R., Fan, W., Yin, Y.-Q., Yu, R. T., Liddle, C.,
- 1283 Atkins, A. R., Olefsky, J. M., Mohammadi, M., Downes, M., & Evans, R. M. (2014).
- 1284 Endocrinization of FGF1 produces a neomorphic and potent insulin sensitizer. *Nature*,
- 1285 513(7518), 436–439. <https://doi.org/10.1038/nature13540>
- 1286 Szkudelski, T. (2001). The mechanism of alloxan and streptozotocin action in B cells of the rat
- 1287 pancreas. *Physiological Research*, 50(6), 537–546.
- 1288 <http://www.ncbi.nlm.nih.gov/pubmed/11829314>
- 1289 Templeman, N. M., Clee, S. M., & Johnson, J. D. (2015). *Suppression of hyperinsulinaemia in*
- 1290 *growing female mice provides long-term protection against obesity*. 2392–2402.

- 1291 <https://doi.org/10.1007/s00125-015-3676-7>
- 1292 Templeman, N. M., Flibotte, S., Chik, J. H. L., Foster, L. J., Nislow, C., Johnson, J. D.,
- 1293 Templeman, N. M., Flibotte, S., Chik, J. H. L., Sinha, S., Lim, G. E., Foster, L. J., Nislow,
- 1294 C., & Johnson, J. D. (2017). Sensitivity in Old Mice and Extends Lifespan Reduced
- 1295 Circulating Insulin Enhances Insulin Sensitivity in Old Mice and Extends Lifespan.
- 1296 *CellReports*, 20(2), 451–463. <https://doi.org/10.1016/j.celrep.2017.06.048>
- 1297 Thorp, A. A., & Schlaich, M. P. (2015). Relevance of Sympathetic Nervous System Activation
- 1298 in Obesity and Metabolic Syndrome. *Journal of Diabetes Research*, 2015, 1–11.
- 1299 <https://doi.org/10.1155/2015/341583>
- 1300 Tomasi, T., Sledz, D., Wales, J. K., & Recant, L. (1967). Insulin Half-Life in Normal and
- 1301 Diabetic Subjects. *Experimental Biology and Medicine*, 126(1), 315–317.
- 1302 <https://doi.org/10.3181/00379727-126-43434>
- 1303 Triantafyllou, S., Lagani, V., Heinze-Deml, C., Schmidt, A., Tegner, J., & Tsamardinos, I.
- 1304 (2017). Predicting Causal Relationships from Biological Data: Applying Automated Causal
- 1305 Discovery on Mass Cytometry Data of Human Immune Cells. *Scientific Reports*, 7(1),
- 1306 12724. <https://doi.org/10.1038/s41598-017-08582-x>
- 1307 Turner, R. C., Holman, R. R., Matthews, D., Hockaday, T. D. R., & Peto, J. (1979). Insulin
- 1308 deficiency and insulin resistance interaction in diabetes: Estimation of their relative
- 1309 contribution by feedback analysis from basal plasma insulin and glucose concentrations.
- 1310 *Metabolism*, 28(11), 1086–1096. [https://doi.org/10.1016/0026-0495\(79\)90146-X](https://doi.org/10.1016/0026-0495(79)90146-X)
- 1311 UK Prospective Diabetes Study Group. (1998a). United Kingdom Prospective Diabetes Study
- 1312 24: A 6-Year, Randomized, Controlled Trial Comparing Sulfonylurea, Insulin, and
- 1313 Metformin Therapy in Patients with Newly Diagnosed Type 2 Diabetes That Could Not Be
- 1314 Controlled with Diet Therapy. *Annals of Internal Medicine*, 128(3), 165.
- 1315 <https://doi.org/10.7326/0003-4819-128-3-199802010-00001>
- 1316 UK Prospective Diabetes Study Group. (1998b). Intensive blood-glucose control with
- 1317 sulphonylureas or insulin compared with conventional treatment and risk of complications
- 1318 in patients with type 2 diabetes (UKPDS 33). *The Lancet*, 352(9131), 837–853.
- 1319 [https://doi.org/10.1016/S0140-6736\(98\)07019-6](https://doi.org/10.1016/S0140-6736(98)07019-6)
- 1320 UK Prospective Diabetes Study Group. (1998c). Tight blood pressure control and risk of
- 1321 macrovascular and microvascular complications in type 2 diabetes: UKPDS 38. *BMJ*,
- 1322 317(7160), 703–713. <https://doi.org/10.1136/bmj.317.7160.703>
- 1323 Watve, M. (2013). *Doves, Diplomats, and Diabetes*. Springer New York.
- 1324 <https://doi.org/10.1007/978-1-4614-4409-1>
- 1325 Watve, M., & Diwekar-Joshi, M. (2016). What to expect from an evolutionary hypothesis for a
- 1326 human disease: The case of type 2 diabetes. *HOMO*, 67(5), 349–368.
- 1327 <https://doi.org/10.1016/j.jchb.2016.07.001>
- 1328 Watve, M. M., Dahanukar, N., & Watve, M. G. (2010). Sociobiological Control of Plasmid Copy
- 1329 Number in Bacteria. *PLoS ONE*, 5(2), e9328. <https://doi.org/10.1371/journal.pone.0009328>
- 1330 Weyer, C., Hanson, R. L., Tataranni, P. A., Bogardus, C., & Pratley, R. E. (2000). A high fasting
- 1331 plasma insulin concentration predicts type 2 diabetes independent of insulin resistance:
- 1332 evidence for a pathogenic role of relative hyperinsulinemia. *Diabetes*, 49(12), 2094–2101.
- 1333 <https://doi.org/10.2337/diabetes.49.12.2094>
- 1334 Wright, S. (1960). The Treatment of Reciprocal Interaction, with or without Lag, in Path
- 1335 Analysis. *Biometrics*, 16(3), 423. <https://doi.org/10.2307/2527693>
- 1336 Yajnik, C. S., Joglekar, C. V., Lubree, H. G., Rege, S. S., Naik, S. S., Bhat, D. S., Uradey, B.,

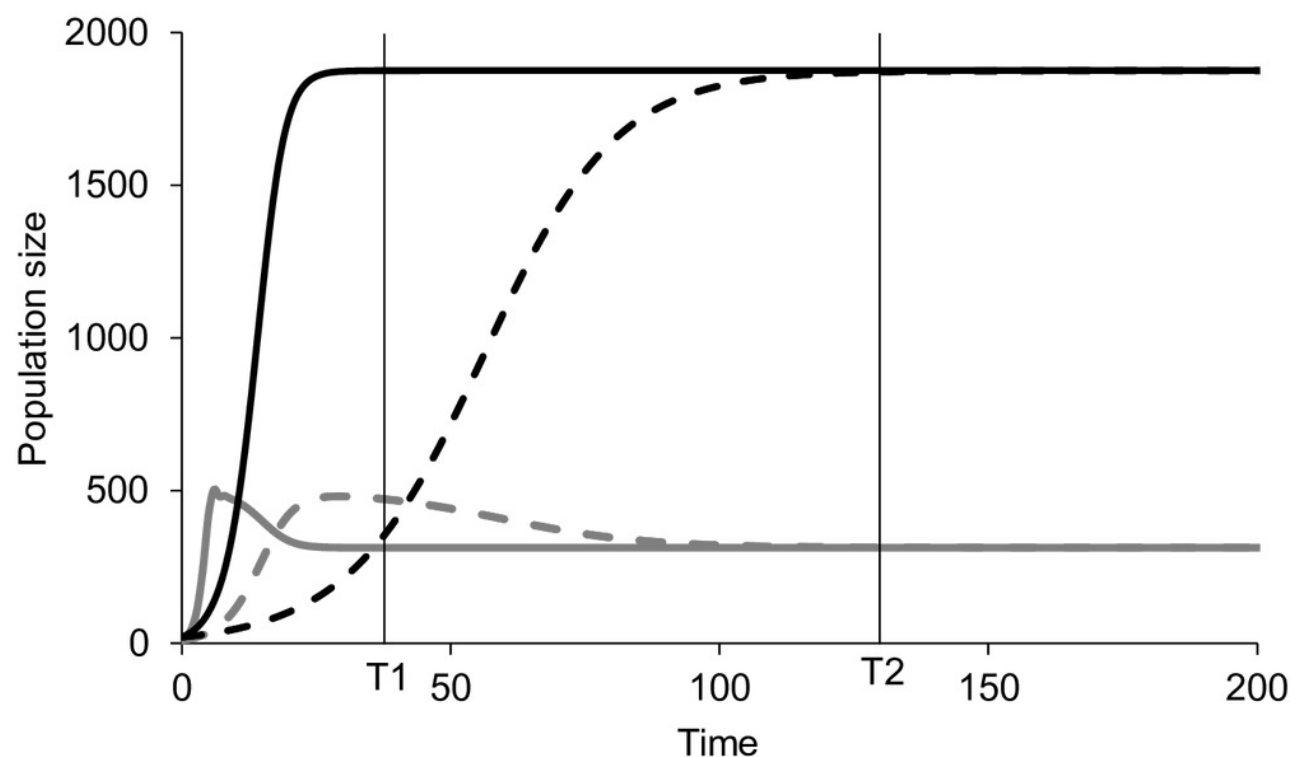


1337 Raut, K. N., Shetty, P., & Yudkin, J. S. (2007). Adiposity, inflammation and  
 1338 hyperglycaemia in rural and urban Indian men: Coronary Risk of Insulin Sensitivity in  
 1339 Indian Subjects (CRISIS) Study. *Diabetologia*, 51(1), 39–46.  
 1340 <https://doi.org/10.1007/s00125-007-0847-1>  
 1341 Zhang, M., Lv, X.-Y., Li, J., Xu, Z.-G., & Chen, L. (2008). The Characterization of High-Fat  
 1342 Diet and Multiple Low-Dose Streptozotocin Induced Type 2 Diabetes Rat Model.  
 1343 *Experimental Diabetes Research*, 2008, 1–9. <https://doi.org/10.1155/2008/704045>  
 1344

# Figure 1

Lotka-Volterra model of two competing species at different growth rates

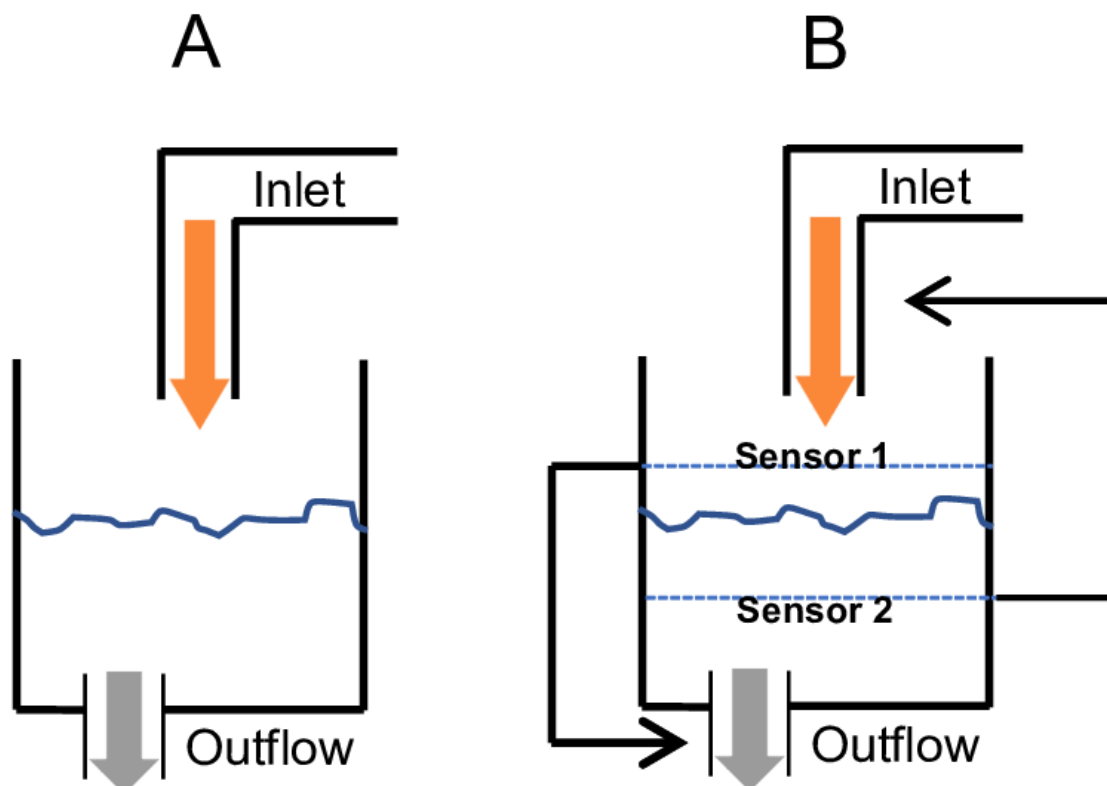
Simulated population dynamics of two competing species A (black lines) and B (grey lines) at different growth rates. Solid lines represent high growth rates reaching the equilibrium faster whereas dotted lines represent slow growth rates reaching the equilibrium more slowly. In both cases species A has slower growth rate but greater carrying capacity than species B. At time T1 the populations are in proportion to their growth rates but at T2, approaching equilibrium, growth rates become increasingly irrelevant in determining population sizes. Thus, growth rates are important determinants of population size in a perturbed state but not at a steady state.



# Figure 2

Consequential steady state (CSS) and targeted steady state (TSS) models of glucose homeostasis.

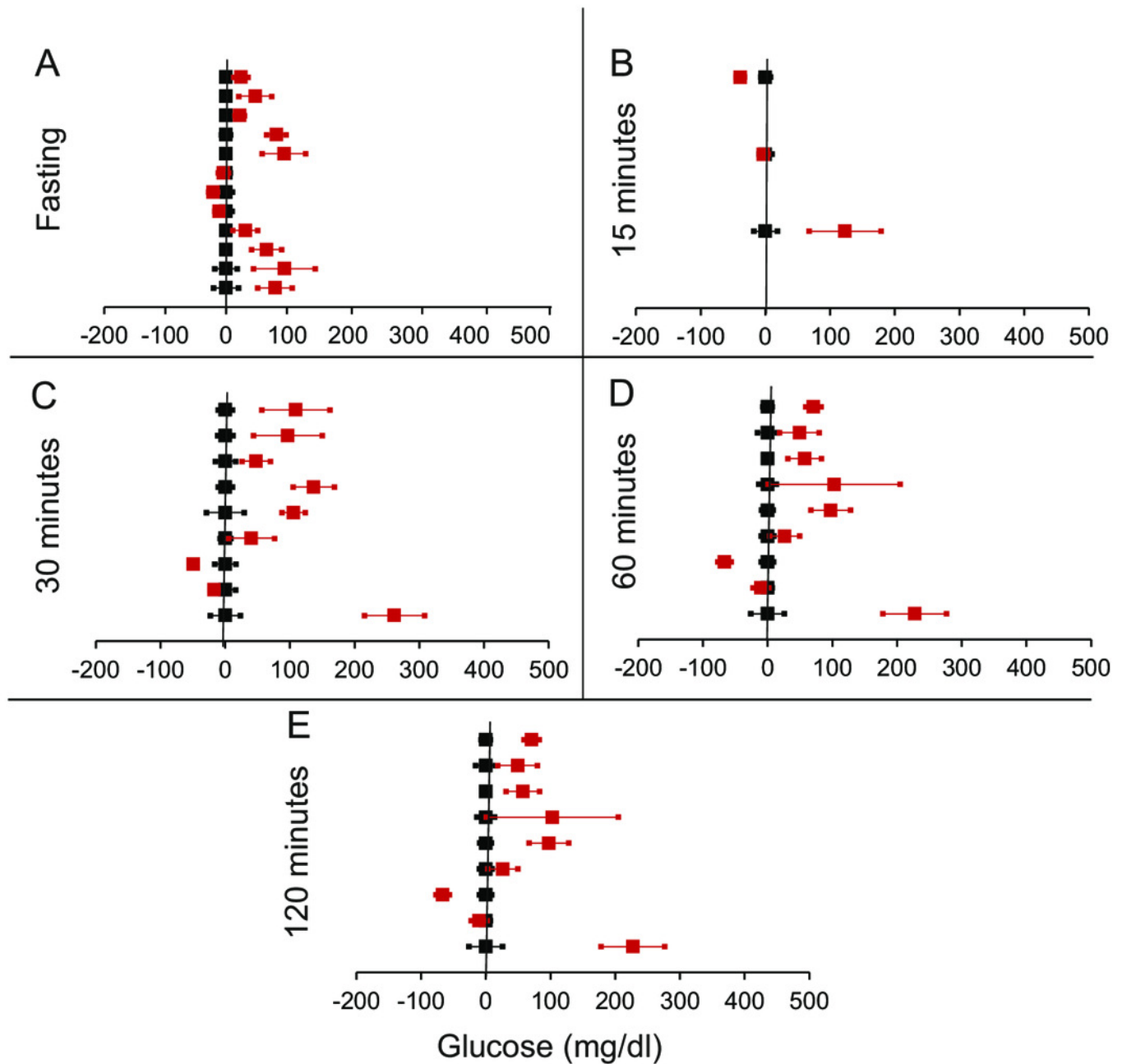
The consequential steady state (CSS) (A) and targeted steady state (TSS) (B) models of homeostasis illustrated with a tank water level analogy. In CSS a change in the size of inlet or outlet tap, analogous to insulin sensitivity can change the steady state level. In a TSS model, a change in the tap size will alter the time required to reach a steady state but will not change the steady state level.



# Figure 3

Steady state and perturbed state glucose for control and FIRKO

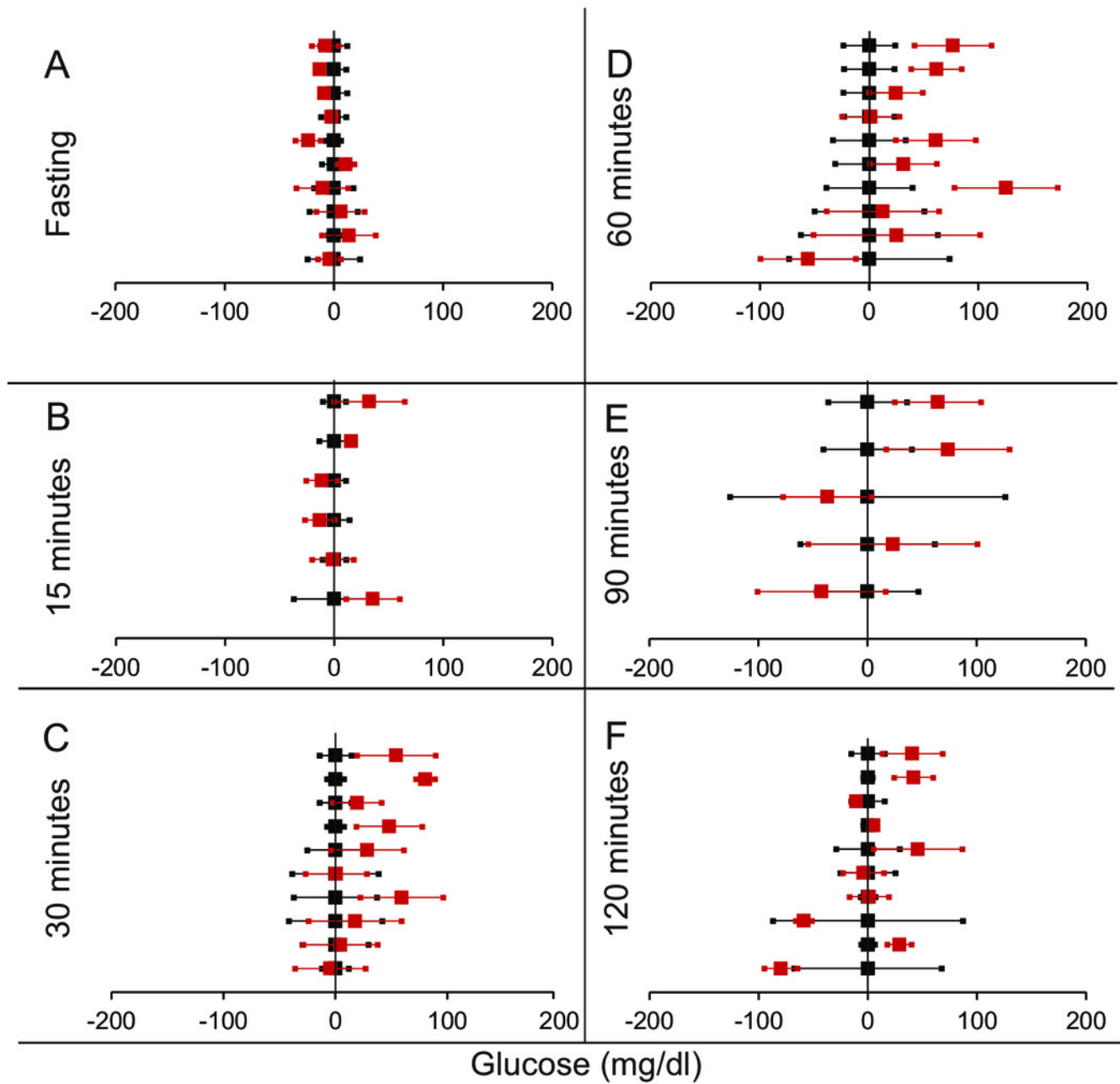
: Glucose levels for control (black squares) and **FIRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and FIRKO were compared using an OGTT. FIRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm$  95% CI. Glucose levels of control are expressed as  $0 \pm$  95% CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes and (E) 120 minutes.



# Figure 4

Steady state and perturbed state glucose for control and MIRKO

Glucose levels for control (black squares) and **MIRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and MIRKO were compared using an OGTT. MIRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm$  95% CI. Glucose levels of control are expressed as  $0 \pm$  95% CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that fasting glucose does not differ from the control in any of the experiments.

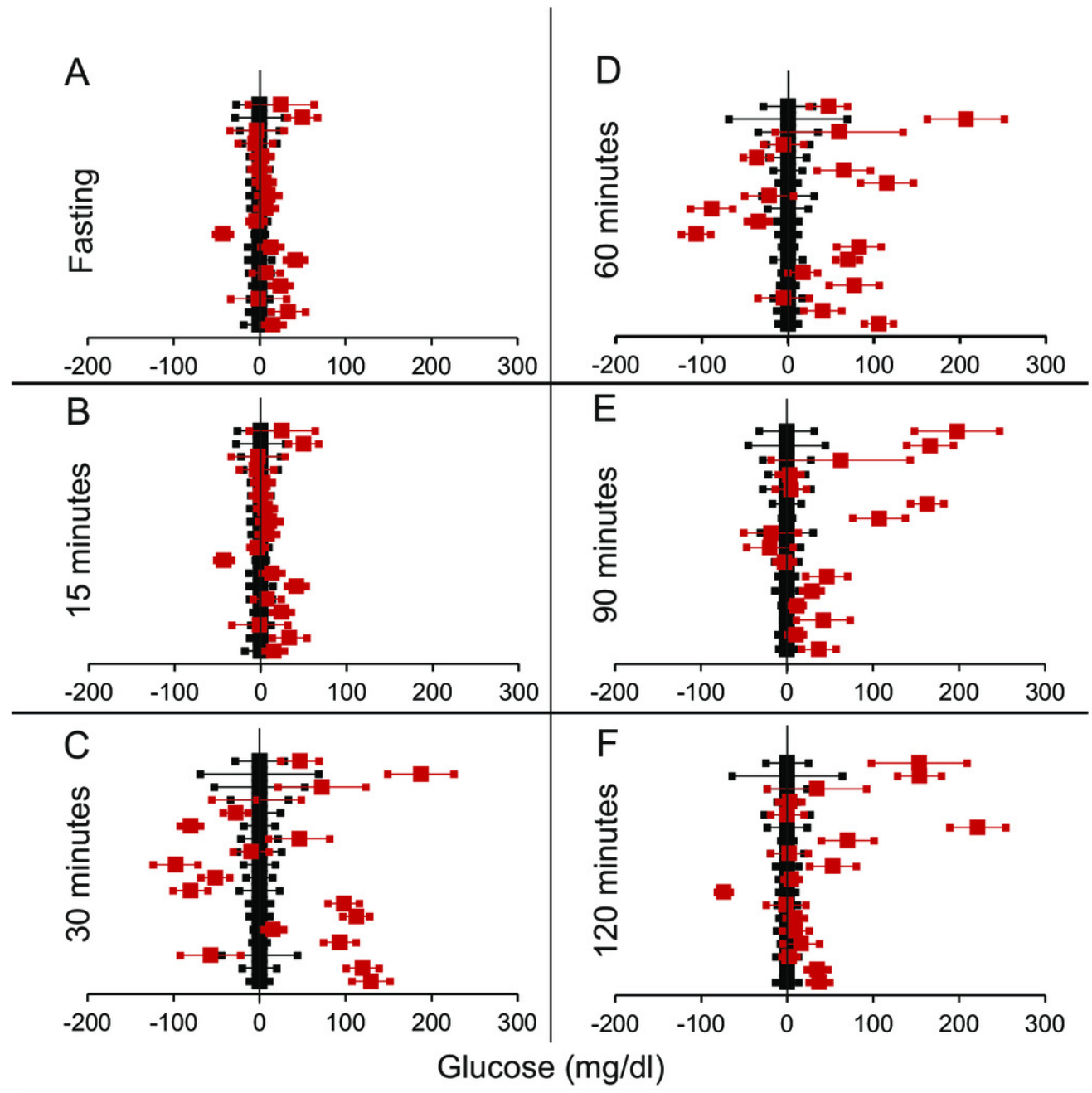


# Figure 5

Steady state and perturbed state glucose for control and LIRKO

Glucose levels for control (black squares) and **LIRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and LIRKO were compared using an OGTT. LIRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm$  95% CI. Glucose levels of control are expressed as  $0 \pm$  95% CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that inconsistent with classical belief, liver specific insulin receptor knockout does not show significant effect on fasting glucose in any of the experiments. On the other hand, post load glucose is consistently higher.

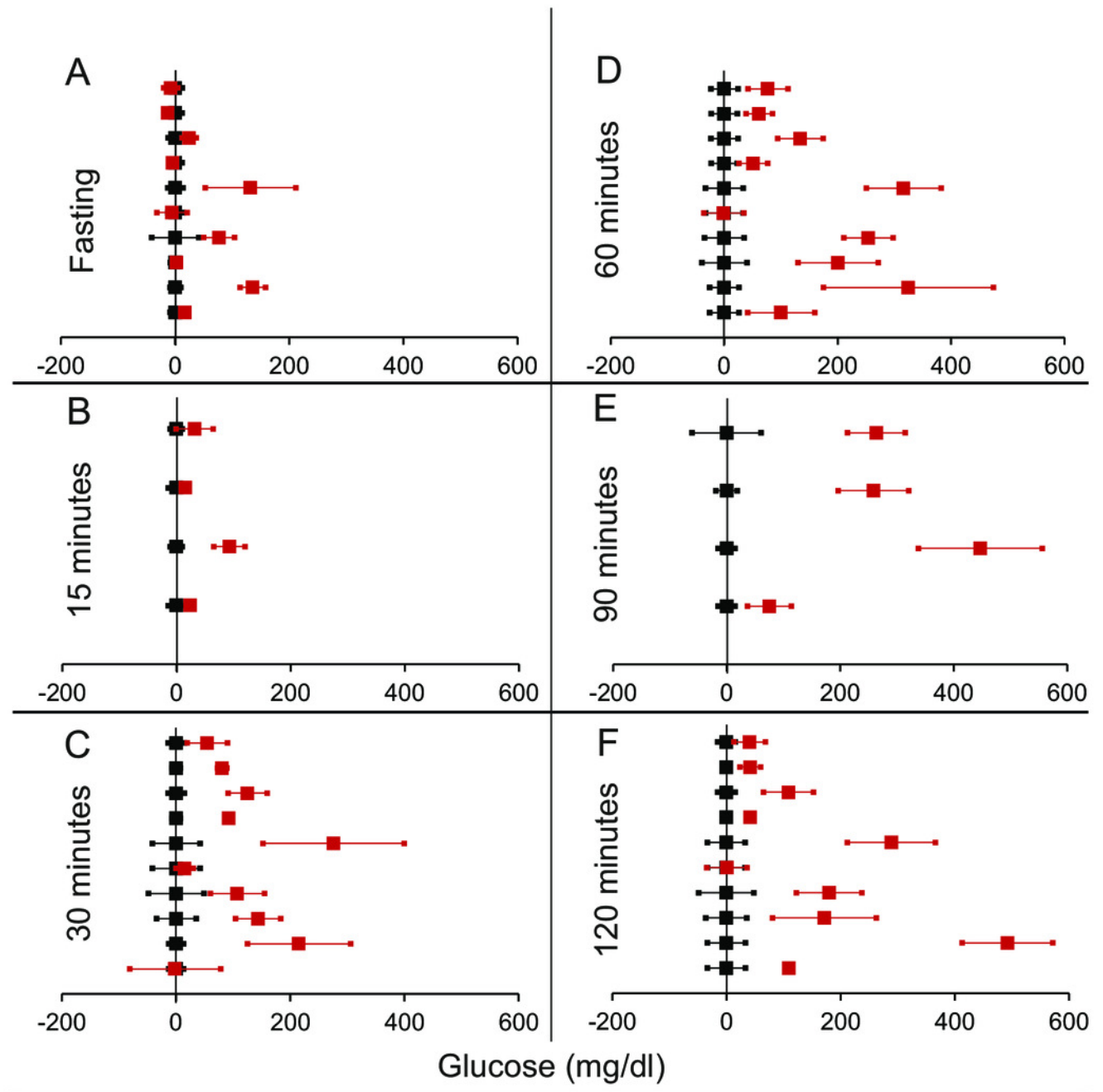




# Figure 6

Steady state and perturbed state glucose for control and  $\beta$ IRKO

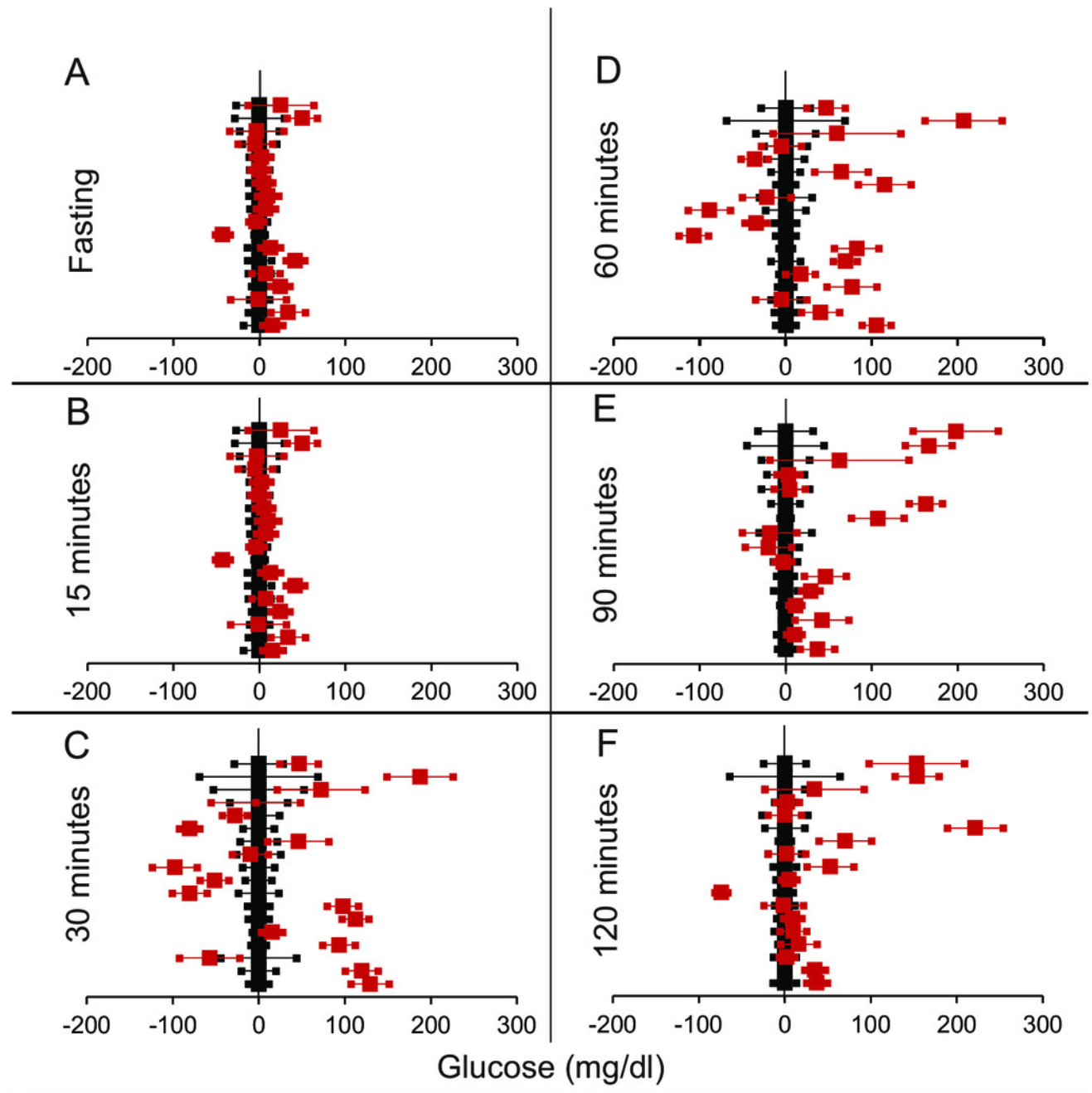
Glucose levels for control (black squares) and  **$\beta$ IRKO** (red squares) at steady state and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and  $\beta$ IRKO were compared using an OGTT.  $\beta$ IRKO glucose levels are normalized to that of the control and the difference is expressed with  $\pm$  95% CI. Glucose levels of control are expressed as  $0 \pm$  95% CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that fasting glucose does not differ from the control in any of the experiments.



# Figure 7

Steady state and perturbed state glucose for control and IDE inhibition.

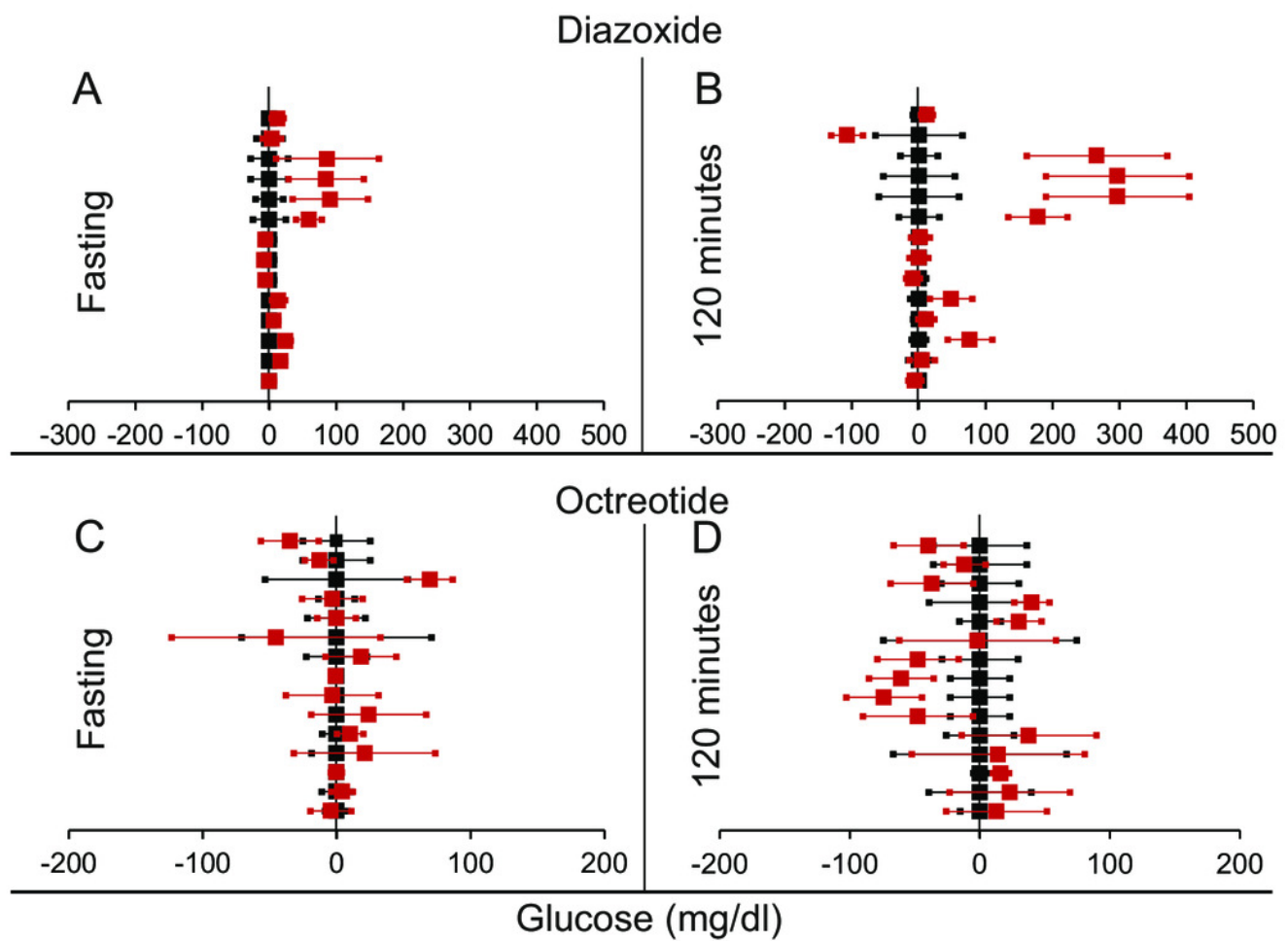
Glucose levels for control (black squares) and **IDE-inhibition** (red squares) models at steady and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and IDE-inhibition were compared using an OGTT. IDE-inhibition glucose levels are normalized to that of the control and the difference is expressed with  $\pm$  95% CI. Glucose levels of control are expressed as  $0 \pm$  95% CI. Steady state is represented by the fasting glucose (A) and the perturbed state is represented by different time points (B to F) post glucose load when the readings are taken: (B) 15 minutes (C) 30 minutes (D) 60 minutes (E) 90 minutes and (F) 120 minutes. Note that fasting glucose does not differ significantly from the control. At 90 and 120 minutes the trend is higher mean glucose than control which is contrary to the expectation in an experiment with sustainable rise in insulin.



# Figure 8

Steady state and perturbed state glucose for control and insulin suppression.

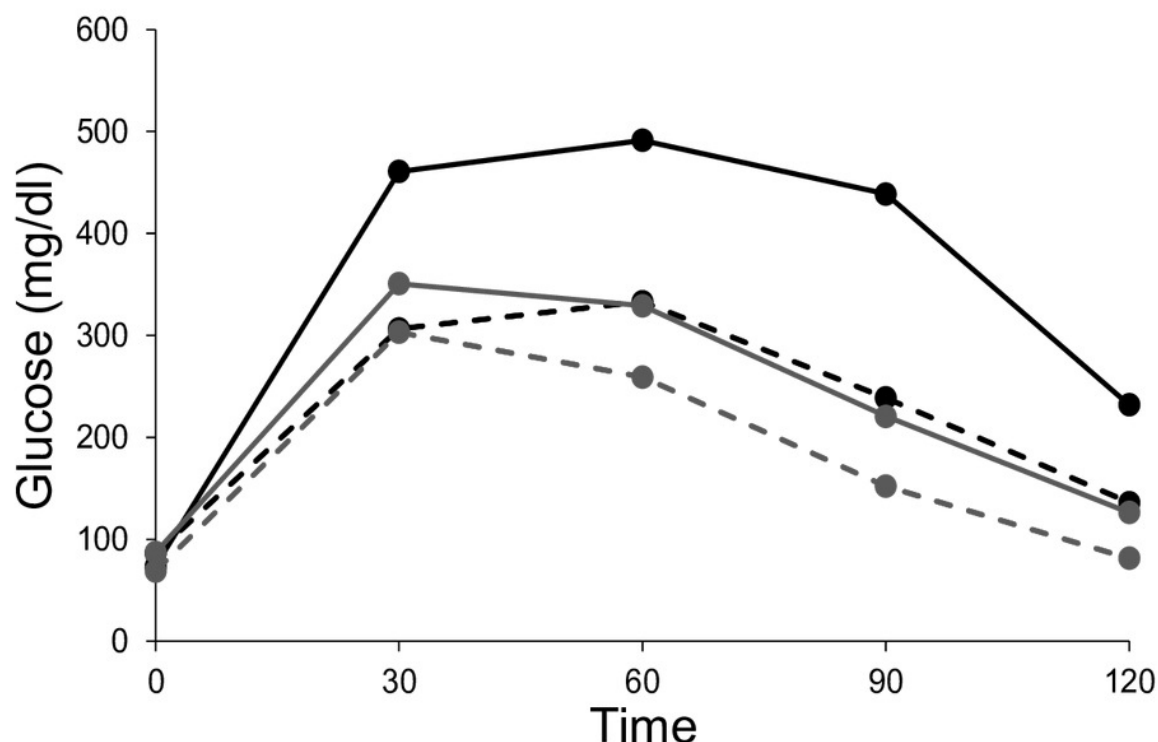
Glucose levels for control (black squares) and treated (red squares) models at steady and perturbed state. The X-axis represents the glucose levels and the Y-axis represents experiments from different studies from the shortlisted papers in which control and treated were compared using an OGTT. Glucose levels after treatment with Diazoxide/Octreotide are normalized to that of the control and the difference is expressed with  $\pm$  95% CI. Glucose levels of control are expressed as  $0 \pm$  95% CI. For Diazoxide treatment, steady state is represented by the fasting glucose (A) and the perturbed state is represented by (B) at 120 minutes post glucose load. For Octreotide treatment, steady state is represented by the fasting glucose (C) and the perturbed state is represented by (D) at 120 minutes post glucose load. Note the inconsistencies across studies.



# Figure 9

## Steady state and perturbed state glucose after insulin suppression by siRNA

Intra peritoneal glucose tolerance test of control and siRNA treated mice. Fasting glucose levels in both the siRNA treated (Tg, represented by solid squares) and untreated group (non-Tg, represented by solid circles) remain unaltered in male (A) and female (B) mice. 15 minutes after the glucose injection, the treated mice show higher glucose levels relative to the untreated mice and this effect is seen throughout till 120 minutes. Figure redrawn from data by (Hwang et al., 2007) .

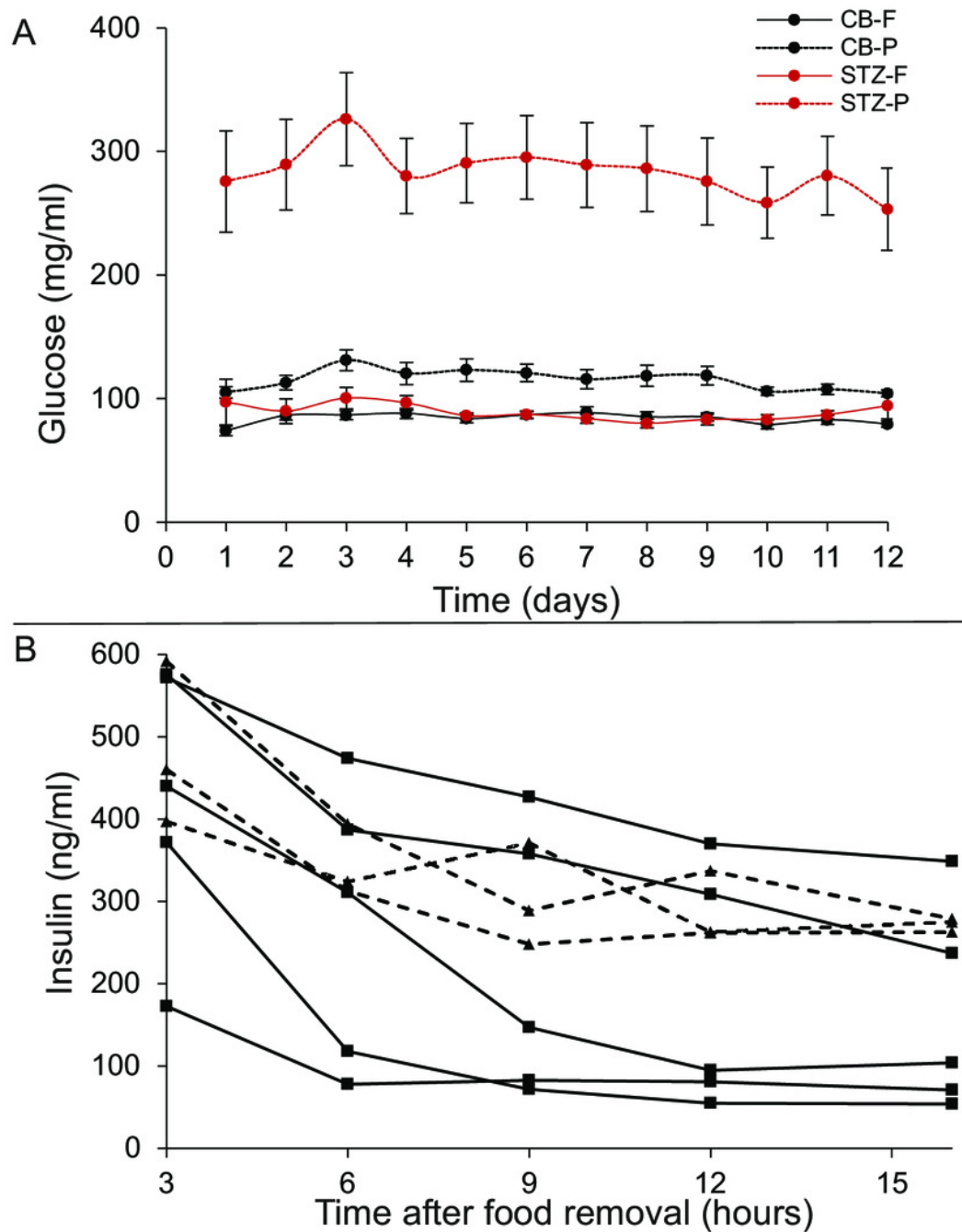




# Figure 10

Steady state and perturbed state glucose after insulin suppression by STZ

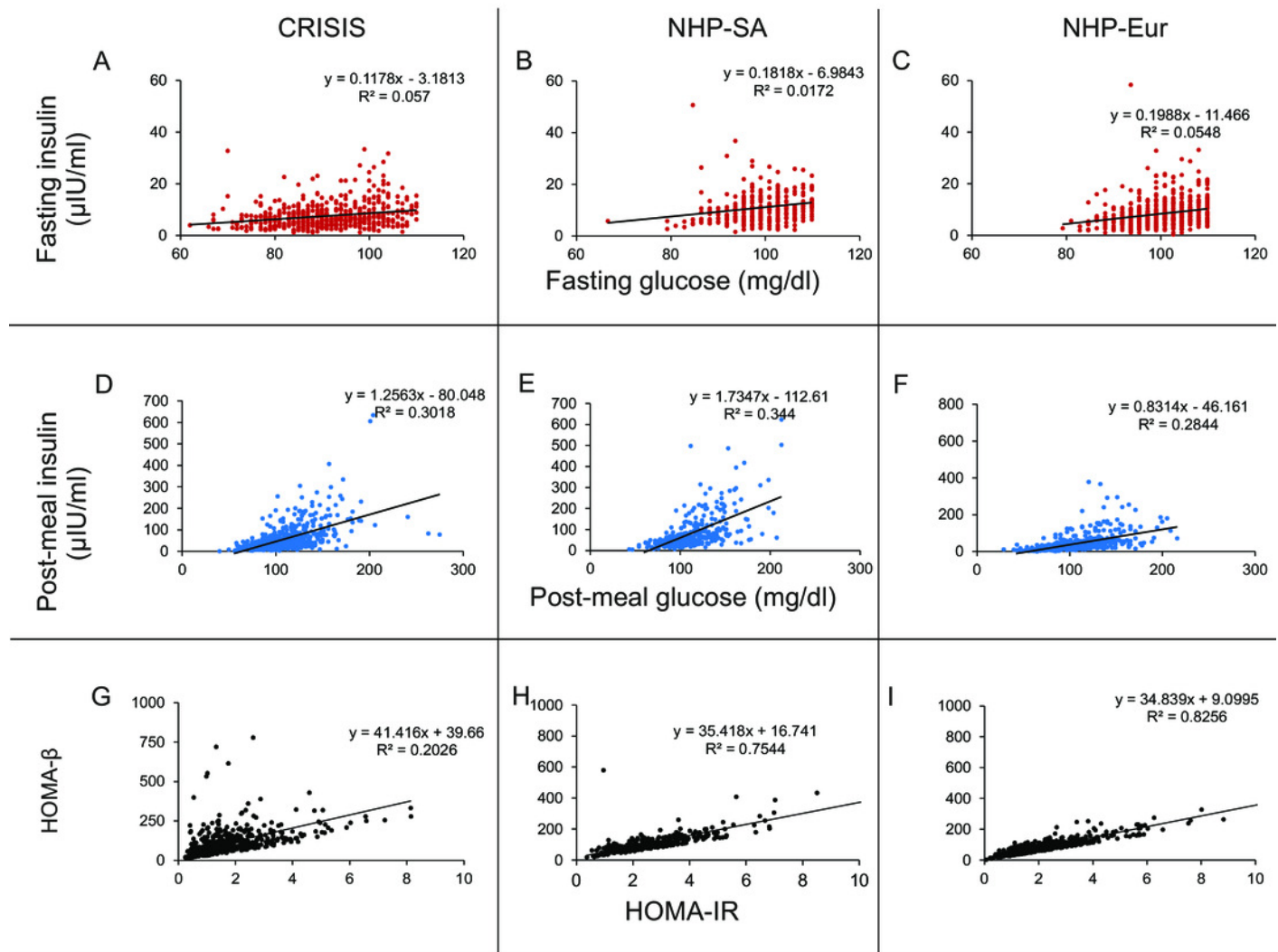
Treatment of SD rats with STZ. (A): The 16-hour fasting and post-meal glucose values of treated (STZ 50mg/kg) and control rats (Citrate buffer CB) over 12 days. N= 12 for each group. Note that on 10 out of 12 days the mean fasting glucose of the treated group (STZ-F) was not significantly different from the control mean (CB-F). Post feeding the treated group (STZ-P) has substantially greater mean than the control (CB-P). (B) Time course of glucose during 16 hours fasting. X axis represents the time after removal of food when the glucose readings are taken and Y axis represent the glucose levels. The grey band represents the upper and lower bounds of 95% CI of the control group with the mean glucose values represented by filled circles. Filled squares represent individuals that showed a monotonic decrease in glucose levels. In three animals the glucose levels reduced at or below the control levels and in two others they showed a continued monotonic decrease but did not reach the normal level in 16 hours. Filled triangles with dotted lines represent the individual time courses of the three STZ treated rats which showed some indications of stabilizing at a steady state above the normal.



# Figure 11

Correlations in the human data:

The Fasting Glucose-Fasting Insulin, Post-meal Glucose-post-meal Insulin and HOMA-IR HOMA- $\beta$  scatter plots in non-diabetic populations in the three data sets. The FG-FI correlation is weak as compared to post-meal correlation. The HOMA-IR and HOMA-  $\beta$  correlations are very strong in all the three data sets, which is not expected by the classical insulin resistance theory



**Table 1**(on next page)

IRKO meta analysis

Systematic literature review for studies on tissue specific insulin receptor knockouts.

1 **Table 1: IRKO meta analysis**

2 Systematic literature review for studies on tissue specific insulin receptor knockouts.

<b>Meta-analysis→ Task performed ↓</b>	<b>Insulin receptor knockout</b>
Key word(s) used for the first search on the PubMed/MEDLINE data-base	“insulin receptor knockout”
Number of hits in the first search	78
Inclusion criteria for primary screening	Study showing experiments with IRKOs in which GTT curve has been reported
Number of papers shortlisted based on primary screening	36
Inclusion criteria for secondary screening	Study showing similar methods of making the insulin receptor knockout; had fasting and post glucose bolus readings of the control and knockout
Number of papers shortlisted based on screening the full-text and back referencing (data extracted from these papers)	16
List of publications used in the final analysis: references numbers	Blüher et al., 2002; Brüning et al., 1998; Cohen et al., 2004; Dodson Michael et al., 2000; Ealey et al., 2008; Escribano et al., 2009; Guerra et al., 2001; Haas et al., 2012; Kawamori et al., 2009; Lauro et al., 1998; Mauvais-Jarvis et al., 2000; Okada et al., 2007; Otani, 2003; Sakaguchi et al., 2017; Softic et al., 2016; Wojtaszewski et al., 1999)

3  
4

## **Table 2**(on next page)

IDE inhibition meta analysis

Systematic literature review for studies on insulin degrading enzyme inhibition/knockout

1 **Table 2: IDE inhibition meta analysis**

2 Systematic literature review for studies on insulin degrading enzyme inhibition/knockout.

<b>Meta-analysis→ Task performed ↓</b>	<b>Insulin degrading enzyme</b>
Key word used for the first search on the PubMed/MEDLINE database	“insulin degrading enzyme”
Number of hits in the first search	1179
Inclusion criteria for primary screening	Studies showing experiments with inhibition of IDE and GTT
Number of papers shortlisted based on primary screening	33
Inclusion criteria for secondary screening	Study showed experiments with IDE inhibition, had fasting and post-glucose load readings of control and IDE inhibition
Number of papers shortlisted based on screening the full-text and back referencing (data extracted from these papers)	6
List of the studies used in the final analysis (all were rodent studies; reference numbers given here)	(Abdul-Hay et al., 2011; Deprez-Poulain et al., 2015; Durham et al., 2015; Farris et al., 2003; Maianti et al., 2014; Villa-Pérez et al., 2018)

3



# **Table 3**(on next page)

Diazoxide and Octreotide meta analyses

Systematic literature review for studies on insulin suppression with diazoxide and octreotide

1 **Table 3: Diazoxide and Octreotide meta analyses**

2 Systematic literature review for studies on insulin suppression with diazoxide and octreotide

<b>Meta-analyses→ Task performed ↓</b>	<b>Diazoxide</b>	<b>Octreotide</b>
Key word used for the first search on the PubMed/MEDLINE data-base	“diazoxide and diabetes”; “insulin suppression”	“octreotide and diabetes”; “insulin suppression
Number of hits in the first search	1043	1202
Inclusion criteria for primary screening	Study shows stable insulin suppression using diazoxide and a GTT has been performed after insulin suppression.	Study shows stable insulin suppression using octreotide and a GTT has been performed after insulin suppression.
Papers shortlisted based on primary screening	239	289
Inclusion criteria for secondary screening	Study showed similarities in the concentration of diazoxide used; and had fasting and post glucose bolus readings of the control and diazoxide subjects	Study showed similarities in the concentration of octreotide used; and had fasting and post glucose bolus readings of the control and octreotide subjects
Papers shortlisted based on screening the full-text and back referencing (data extracted from these papers)	Rodent studies (2) Human studies (6)	Rodent studies (0) Human studies (10)
List of human studies used in the final analysis	(Brauner et al., 2016; Due et al., 2007; Ramanathan et al., 2011; Schreuder et al., 2005; van Boekel et al., 2008; Wigand & Blackard, 1979)	(Breckenridge et al., 2007; Candrina et al., 1988; Davies et al., 1986; Giustina et al., 1991; Johnston et al., 1986; Madsen et al., 2011; Parkinson et al., 2002; Ronchi et al., 2002; Williams et al., 1986, 1988)
List of rodent studies used in the final analysis	(Leahy et al., 1994; Matsuda et al., 2002)	None

3

# **Table 4**(on next page)

: Analysis of IRKO meta analysis

Collective analysis of the fasting and post-feeding glucose levels in the control and IRKOs: The table shows, out of the total number of experiments used for the analysis, in how many the mean of the knockouts (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa. It can be seen that for fasting glucose the difference is not significant in majority of studies and where there is statistical significance there is lack of consistency across studies. However, at 30, 60 and 120 minutes the knockouts have consistently elevated levels of glucose as compared to the corresponding controls.

**Table 4: Analysis of IRKO meta analysis**

Collective analysis of the fasting and post-feeding glucose levels in the control and IRKOs: The table shows, out of the total number of experiments used for the analysis, in how many the mean of the knockouts (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa. It can be seen that for fasting glucose the difference is not significant in majority of studies and where there is statistical significance there is lack of consistency across studies. However, at 30, 60 and 120 minutes the knockouts have consistently elevated levels of glucose as compared to the corresponding controls.

	Total studies	T>C	T<C	<i>p</i> using chi square	T > C individually significant	T < C individually significant
<b>All IRKOs</b>						
Fasting	46	25	20	0.454	13	4
15 minutes	14	7	7	0.999	4	2
30 minutes	40	36	4	<0.0001*	22	1
60 minutes	40	36	4	<0.0001*	24	1
120 minutes	46	37	9	<0.0001*	24	2
<b>FIRKO</b>						
Fasting	12	9	3	0.083	9	1
15 minutes	3	1	2	0.566	1	1
30 minutes	9	7	2	0.095	6	1
60 minutes	9	7	2	0.095	5	1
120 minutes	12	9	3	0.83	7	2
<b>MIRKO</b>						
Fasting	10	3	7	0.205	0	2
15 minutes	6	3	3	0.999	1	0
30 minutes	10	9	1	0.011*	3	0
60 minutes	10	9	1	0.011*	3	0
120 minutes	10	6	4	0.527	3	0
<b>LIRKO</b>						
Fasting	9	4	5	0.739	0	0
15 minutes	1	1	0	N.A.	0	0
30 minutes	9	9	0	0.003*	6	0
60 minutes	9	9	0	0.003*	7	0
90 minutes	9	9	0	0.003*	5	0
120 minutes	9	7	2	0.094	4	0
<b>βIRKO</b>						
Fasting	8	6	2	0.157	4	0
15 minutes	2	2	0	0.157	2	0
30 minutes	8	7	1	0.033*	6	0
60 minutes	8	7	1	0.033*	7	0
90 minutes	4	4	0	0.046*	4	0
120 minutes	8	8	0	0.0046*	7	0

11  
12

# Table 5 (on next page)

## Analysis of IDE inhibition meta analysis

**Analysis of the fasting and post-feeding glucose levels in the control and IDE-inhibition:** The table shows, out of the total number of experiments used for the analysis, in how many the mean of the IDE inhibition models (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa. There is no significant difference in the control at treated fasting glucose level. At 90 and 120 minutes the difference between the control and treated was significant, but in the opposite direction.

**Table 5: Analysis of IDE inhibition meta analysis**

Comparison between steady state (fasting) and perturbed state (post glucose load) of control and IDE suppression.

	<b>Total experiments</b>	<b>T&gt;C</b>	<b>T&lt;C</b>	<b><i>p</i> using chi square</b>	<b>T &gt; C individually significant</b>	<b>T &lt; C individually significant</b>
Fasting	18	12	6	0.157	0	1
15 minutes	14	7	7	0.999	3	5
30 minutes	18	10	8	0.637	5	2
60 minutes	18	11	7	0.346	5	1
90 minutes	16	13	3	0.012*	4	0
120 minutes	18	15	2	0.002*	2	1

# Table 6 (on next page)

Analysis of DZX and OCT meta analyses

**Analysis of the fasting and post-feeding glucose levels in the control and treatment with Diazoxide or Octreotide:** The table shows, out of the total number of experiments used for the analysis, in how many the mean of the treated (T) was greater than the control means (C) and in how many the trend was reverse. This relative position of the means across studies is compared non-parametrically to see whether the trend across studies was non-random, significant ones being indicated by asterisk. The table also gives in how many studies T was significantly greater than C and vice versa.



1 **Table 6: Analysis of DZX and OCT meta analyses**

2 Comparison between steady state(fasting) and perturbed state (post glucose load) of control and insulin suppression.

	<b>Total studies</b>	<b>T&gt;C</b>	<b>T&lt;C</b>	<b><i>p</i> using chi square</b>	<b>T &gt; C individually significant</b>	<b>T &lt; C individually Significant</b>
<b>Diazoxide treatment</b>						
Fasting	14	10	3	0.052	5	0
15 minutes	7	6	1	0.059	1	1
30 minutes	12	10	2	0.021*	2	1
60 minutes	13	9	4	0.166	4	2
90 minutes	3	3	0	0.083	2	0
120 minutes	14	10	3	0.052	6	1
<b>Octreotide treatment</b>						
Fasting	15	6	7	0.781	0	0
30 minutes	14	4	10	0.108	0	2
60 minutes	14	4	10	0.108	2	1
90 minutes	13	5	8	0.405	1	0
120 minutes	15	7	8	0.797	1	1

3

**Table 7** (on next page)

Correlations from human data

Correlation and regression parameters of glucose-insulin relationship at steady and perturbed states.

1 **Table 7: Correlations from human data**

2 Correlation and regression parameters of glucose-insulin relationship at steady and perturbed states.

	Steady state(fasting)			Perturbed state (2 hours post glucose bolus)		
Parameter → Data set ↓	R-squared (variance explained)	p value	Slope (95% CI)	R-squared (variance explained)	p value	Slope (95% CI)
CRISIS (N=522)	0.0570 (5.7%)	<0.0001	0.1178 (0.0765 to 0.1591)	0.3018 (30.18%)	<0.0001	1.2563 (1.0917 to 1.4209)
NHP-South Asian (N=310)	0.0172 (1.72%)	0.021	0.1818 (0.0279 to 0.3356)	0.344 (34.4%)	<0.0001	1.7347 (1.4661 to 2.0033)
NHP-European (N=574)	0.0548 (5.48%)	<0.0001	0.1988 (0.131 to 0.2666)	0.2844 (28.44%)	<0.0001	0.8314 (0.7231 to 0.9397)

3