Peer

Influence of montane altitudinal ranges on species distribution models; evidence in Andean blow flies

Mariano Altamiranda-Saavedra^{1,2}, Eduardo Amat¹ and Luz Miryam Gómez-P¹

¹ Grupo de Investigación Bioforense, Facultad de Derecho y Ciencias Forenses, Tecnológico de Antioquia Institución Universitaria, Medellín, Colombia

² Grupo de investigación GESTAS, Politécnico Colombiano Jaime Isaza Cadavid, Medellín, Colombia

ABSTRACT

Background. Blow flies are a family of dipterans of medical, veterinary and sanitary importance. We aim to predict the current geographical distribution of six neotropical blowfly species with different altitudinal ranges of distribution (high, medium, and lowlands) and degree of synanthropy (eusynanthropic, hemisynanthropic and asynanthropic) based on their existing fundamental niche (E_A) in Northwestern South America.

Methods. Geographical records were compiled based on data from museum specimens and literature. The accessible area hypothesis (M) was calculated based on three criteria: (1) Altitudinal range, (2) Synanthropy values deducted based on the Human Influence Index (HII) raster dataset, and (3). The mean dispersal capability of flies. The modeling was performed using the Maxent entropy modeling software. The selection of parameters was made with the R Program ENMeval package.

Results. The models were assessed using the area under the operator-partial receiver curve (ROCp). The high statistical performance was evidenced in every modeling prediction. The modeling allowed identifying possible taxonomic inaccuracies and the lack of exhaustive collection in the field, especially for lowlands species. Geographical distribution predicted by the modeling and empirical data was remarkably coherent in montane species.

Discussion. The data obtained evidence that montane elevational ranges affect the performance of the distribution models. These models will allow a more precise predicting of medium and high elevation blow flies than lowlands species. Montane species modeling will accurately predict the fly occurrence to use such biological information for medical, legal, veterinary, and conservation purposes.

Subjects Biogeography, Ecology, Entomology, Zoology, Spatial and Geographic Information Science

Keywords Regionalization, Human influence index, Species distribution models, Synanthropy

INTRODUCTION

Blow flies (Calliphoridae, Oestroidea) are a family of dipterans consisting of approximately 1,600 species worldwide (*Pape, Blagoderov & Mostovski, 2011*), and approximately 100 occurring in the Neotropical region (*Kosmann et al., 2013*). Most carrion-feeding species are of medical, veterinary forensic and sanitary importance. They are of sarcosaprophagous

Submitted 10 July 2020 Accepted 26 October 2020 Published 7 December 2020

Corresponding author Mariano Altamiranda-Saavedra, marianoaltamiranda@elpoli.edu.co, maltamiranda2@gmail.com

Academic editor Brock Harpur

Additional Information and Declarations can be found on page 11

DOI 10.7717/peerj.10370

© Copyright 2020 Altamiranda-Saavedra et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

habits and are among the most active scavengers of the food chain, reducing dead organic matter or waste from primary producers to consumers (*Galante & Marcos-Garcia, 1997*). Most species are strongly attracted by volatile compounds emanated by feces, secretions, decaying materials, and food. This flying tendency between rotten materials and human comestibles makes them efficient mechanical vectors of pathogens (*Greenberg, 1973*). Adults have a well-developed olfactory system that is very sensitive to carrion and allows them to locate it at great distances quickly (*Yan et al., 2018*). Female flies lay eggs (some species deposit larvae) on the decaying material and began the development; larvae are the most important organisms in the carcass reduction process (*Anderson & Cervanka, 2002*), determining its age is used in forensic entomology as a biological clock to calculate the postmortem interval (PMI) (*Greenberg & Kunich, 2002*). Some species can also develop in animals and humans' wounds, where they feed on living tissue, producing myiasis (infestation of alive tissue by maggots) (*Norris, 1965; Francesconi & Lupi, 2012*).

The spatial distribution of flies fundamentally depends on tolerance to environmental conditions, being affected by the degree of human impact (anthropization process), as natural landscapes become urbanized (Kavazos & Wallman, 2012). In entomological studies, the anthropic level of preference is commonly referred to as "synanthropy" (Gregor & Povolný, 1958). Thus, synanthropic, hemisynanthropic, and asynanthropic are categories for classifying flies based upon their degree of attraction or repulsion to human settlements (*Povolný*, 1971). This ecological classification for Necrophagous Diptera has been widely used to understand spatial patterns for habitat association or an urban-rural gradient (Nuorteva, 1963; Hwang & Turner, 2005; Kavazos & Wallman, 2012; Amat & Medina, in press). Potential spatial distribution by modeling an ecological niche has become a frequent subject of study (Austin, 2002). In order to accurately assess the geographical distribution of organisms, several methodologies have been proposed (Anderson, Lew & Peterson, 2003; Qiao et al., 2016a; Qiao et al., 2016b). Models help locate areas of suitable environmental for species thriving and settling based on parameters previously obtained from the field (Peterson et al., 2002). Modeling was useful in assessing distributional and geographical patterns in biogeographical, ecological and conservational contexts (Anderson, Lew & Peterson, 2003).

The use of these models has become popular in recent years, mainly for the study of plants and vertebrates such as mammals, birds, amphibians, and reptiles, and on a smaller scale, of invertebrates and insects (*Elith & Leathwick, 2009; Peterson et al., 2011; Peterson et al., 2020; Qiao et al., 2016a; Qiao et al., 2016b*). Few studies evaluate the relationship between the spatial distribution of blowfly species and their environment using species distribution models (SDMs) (*Lecheta, Corrêa & Moura, 2017; Richards, Williams & Villet, 2009; Mulieri* & *Patitucci, 2019*). In South Africa, the potential distribution of seven important blow flies was analyzed, and it was found that the prediction of the distribution of restricted species was more accurate than those more widespread in the region (*Richards, Williams & Villet, 2009*). Later, *Williams, Richards & Villet (2014)* discovered no correlation between species distributions and sheep farms, or human populations. In South America, the potential geographic distribution of the blow fly *Sarconesia chlorogaster* (Diptera: Calliphoridae) was estimated, and it predicted suitable areas in Ecuador and Colombia, which had no

previous records (*Lecheta, Corrêa & Moura, 2017*). Recently, the potential distribution of *Cochliomyia hominivorax* (Diptera: Calliphoridae) was estimated, and it was determined that large areas in the central-eastern territory of Argentina are environmentally suitable for potential occurrence, and the seasonal temperature fluctuation is the most significant contributor to explain such distribution (*Mulieri & Patitucci, 2019*).

We aim to evaluate if the altitudinal range influence the current potential geographical distribution of six neotropical blowfly species with different altitudinal ranges of distribution (high, medium and lowlands) and different degree of synanthropy (eusynanthropic, hemisynanthropic and asynanthropic) based on their existing fundamental niche (E_A) in Northwestern South America by considering the accessible area hypothesis (M) calculated based on three criteria: (1) Altitudinal range, (2) Synanthropy values are deducted based on the Human Influence Index (HII) raster dataset, and (3) The mean dispersal capability of flies. This contribution will facilitate the use of ecological and chorological data of flies for medical-legal, veterinary, and conservational purposes.

MATERIALS & METHODS

Study area

The area is located in the northwestern extreme of South America; it includes the political borders of five countries: Northwestern Brazil, Northern Peru, Colombia, Ecuador, and Venezuela. In a biogeographical sense, it is possible to differentiate six natural regions. The blowfly spatial distribution follows the biogeographical regionalization proposed in *Amat* (2017) (Fig. 1).

Presence data

After disposing of spatial autocorrelation with the R package ntbox (*Osorio-Olvera et al., 2020*), the complete occurrence database included 228 records of presence, including asynanthropic and medium lowland (1,100–2,600 m.a.s.l) distribution *Blepharicnema splendens* (Diptera: Calliphoridae) (22) and high land (2,000–3,127 m.a.s.l) distribution *Sarconesia roraima* (Diptera: Calliphoridae) (15); hemisynanthropic and lowland (0–1,000 m.a.s.l) *Chloroprocta idioidea* (Diptera: Calliphoridae) (59) and medium lowland (1,100–2,700 m.a.s.l) distribution *Lucilia purpurascens* (Diptera: Calliphoridae) (30); eusynanthropic and high land (2,200–2,800) distribution *Calliphora vicina* (Diptera: Calliphoridae) (19) and lowland (0–1,400 m.a.s.l) *Cochliomyia macellaria* (Diptera: Calliphoridae) (83) (Table S1). Presence records by species were obtained from the geographical information on specimens revised in eleven entomological museums, and recent literature on blow flies published since 2000. Data for each specie were organized as presence records in a database, with their coordinates in decimal degrees projected in the WGS84 system.

Acronyms of museums, collections and depositaries of specimens are as follows: MIZA Museo del Instituto de Zoología Agrícola, Francisco Fernández Yepes. Universidad Central de Venezuela, Maracay, Venezuela. INPA Coleção de Invertebrados, Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil. MECN Museo Ecuatoriano de Ciencias Naturales. Quito, Ecuador. QCAZ-I Museo de Zoología - Sección Invertebrados, Facultad de Ciencias





Exactas y Naturales, Pontificia Universidad Católica del Ecuador, Quito, Ecuador. IAVH-E Colección entomológica - Instituto de Investigaciones Biológicas Alexander von Humboldt, Villa de Leyva, Boyacá, Colombia. ICN-MHN Instituto de Ciencias Naturales –Museo de Historia Natural, Facultad de Ciencias, Universidad Nacional de Colombia. Bogotá, Colombia. CETdeA Colección entomológica del Tecnológico de Antioquia, Institución Universitaria, Medellín, Colombia. MEFLG Museo Entomológico Francisco Luis Gallego, Universidad Nacional de Colombia, sede Medellín, Medellín, Colombia. UNAB Museo Entomológico, Facultad de Agronomía, Universidad Nacional de Colombia, sede Bogotá, Colombia. UPTC Museo Luis Gonzalo Andrade. Universidad Pedagógica y Tecnológica de Colombia, Tunja, Boyacá, Colombia.

Environmental layers

The variables used to characterize the existing fundamental niche of the species of interest and evaluate their potential distribution were obtained from Worldclim, version 2.0, (spatial resolution = 30 arcseconds, ~ 1 km) These are derived from monthly values of precipitation and temperature with biological significance and represent annual trends of temperature, precipitation, seasonality, and extreme or limiting environmental factors (*Fick & Hijmans, 2017*). In order to reduce collinearity in environmental layers, a Pearson correlation analysis was conducted with the tool known as "SDMtoolbox" on ArcGis 10.3. Variables with correlation value >|0.8| were eliminated (*Raghavan et al., 2019*). Additionally, we used the Jackknife option from the Maxent software to identify variables that did not provide a significant contribution to the robustness of the models.

Model design

A key element in the development of ecological niche models is the hypotheses of areas (M) that have been accessible to the species in relevant periods (*Barve et al., 2011*). With

the available information on the natural history and biogeography of the species of interest, we created hypotheses for the accessible areas (M) by establishing different criteria: (1) reclassifying a digital elevation model (DEM) from Shuttle Radar Topography Mission (SRTM) (*Rabus et al., 2003*) with the information on altitudinal range reported for species (B. splendens 1,000-2,700 masl, S. roraima 1,500-3,500 masl, L. purpurascens 1,000-3,000 masl, C. vicina 1,500–2,500 masl, C. macellaria 0–2,000 masl and C. idioidea 30-1200 masl; 2) the value of synanthropy was measured using a raster from the Human Influence Index (HII) dataset of the Last of the Wild Project, version 2 (Wildlife Conservation Society WCS & University C for IESIN-C-C, 2005). The dataset integrates information from nine layers related to human population densities, land use, infrastructure, and human transportation access. As for asynanthropic species, we took into account the maximum value of this index for occurrence data, the minimum value for eusynanthropy, and finally, the range for hemisynanthropy; (3) the result of the mask of altitude and index of synanthropy was added to a buffer with dispersion capacity data of the species measured in Km2. For species lacking this information, we used the minimum known flight range value for the Calliphoridae family (4.8 km) (Bishopp & Laake, 1921; Lindquist et al., 1951; Quarterman, Mathis & Kilpatrick, 1954; Shura-Bura et al., 1958).

We assessed the current geographical distribution based on the existing fundamental niche concept (E_A) (*Peterson et al., 2011*), using Maxent 3.3.3k (*Phillips, Anderson & Schapire, 2006*) to estimate environmental suitability in these analyses. To assess the optimal parametrization of the suitability estimates in the calibration region, different settings were tested using the ENMeval package of the R program (*Muscarella et al., 2014*), which provides an automated method to execute MaxEnt models across a user-specified range of regularization multiplier (RM) values and feature combinations (FCs) (*Muscarella et al., 2014*). Then, we set the RM range from 0.5 to 4.0 with 0.5 increments and three FCs, i.e., linear (L), linear and quadratic (LQ), linear, quadratic and product (LQP), linear, quadratic, product and threshold (LQPT) linear, quadratic, product threshold and hinge (LQPTH), resulting in 45 possible combinations of features and regularization multipliers (*Muscarella et al., 2014*). Output format (row), number of replicates (10), percent used for testing (25%), type of validation (bootstrapping), maximum number of iterations (5000), convergence threshold (0.00001), and maximum number of background points (10,000), no clamping or extrapolation were fixed for each Maxent run.

The fine-tuned MaxEnt models were made by finding the lowest delta value of Akaike's information criterion corrected for small sample sizes (AICc) among candidate models, which reflects both models, goodness-of-fit and complexity, providing the most conservative results (*Basanta, Rebollar & Parra-Olea, 2019*) (Table 1). Using the R package ntbox (*Osorio-Olvera et al., 2020*), a ROC (Receiver Operating Characteristic) was estimated to assess the performance of the model (*Peterson, Papes & Sober, 2008*). The model was assessed through calibration, randomly selecting 25% of the distribution data and comparing it with the threshold of the area under the curve (AUC). The medians were used through repetitions as a final niche estimate. We established a threshold to convert the raw map of the output of the continuous model into binary assumptions of suitable

Table 1Parameters, set environmental variables used in each model and results of partial ROC analysis (mean value for AUC ratio) to test sta-
tistical significance of ecological niche model predictions. A value of 1.0 is equivalent to the performance of a random classifier. These results were
based on 100 bootstrap replicates, and statistical significance was assessed via bootstrapping and comparison with a random classifier ratio of 1.0.

Species	Records	Environmental variables-relative contribution (%)	Feature classes	Regularization multiplier	ROCp
Blepharicnema splendens	22	bio1 (69.9), bio2 (9), bio8 (1.9), bio12 (1.9), bio14 (1.8), bio15 (3.7), bio19 (10.8)	LQHPT	2	1.840
Sarconesia roraima	15	bio3 (3.7), bio6 (26.8), bio7 (43.2), bio12 (16.9), bio14 (0), bio15 (3.3), bio16 (0.1), bio19 (3.9)	LQHPT	1.5	1.924
Chloroprocta idioidea	59	bio1 (0.3), bio6 (14.5), bio7 (23.2), bio9 (5.6), bio12 (0), bio14 (14.5), bio15 (0), bio16(30.6), bio19 (21.9)	LQ	0.5	1.502
Lucilia purpurascens	30	bio1 (0.9), bio2 (5.1), bio3 (4.4), bio9 (27), bio12 (0.6), bio15 (3.7), bio16 (39), bio19 (36)	LQHPT	1.5	1.675
Calliphora vicina	19	bio2 (2.5), bio3 (3.6), bio6 (8.4), bio7 (0.9), bio14 (3.4), bio15 (12.2), bio19 (54.5)	LQ	0.5	1.979
Cochliomyia macellaria	83	bio1 (8.8), bio3 (7.4), bio6 (0), bio8 (2.8), bio9 (6.4), bio12 (18.8), bio14 (10.9), bio15 (30), bio19 (9.3)	LH	1.5	1.665

Notes.

Bio1, Annual Mean Temperature; Bio2, Mean Diurnal Range; Bio3, Isothermality; Bio4, Temperature Seasonality; Bio5, Max Temperature of Warmest Month; Bio6, Min Temperature of Coldest Month; Bio7, Temperature Annual Range; Bio8, Mean Temperature of Wettest Quarter; Bio9, Mean Temperature of Driest Quarter; Bio10, Mean Temperature of Warmest Quarter; Bio11, Mean Temperature of Coldest Quarter; Bio12, Annual Precipitation; Bio13, Precipitation of Wettest Month; Bio14, Precipitation of Driest Month; Bio15, Precipitation Seasonality; Bio16, Precipitation of Wettest Quarter; Bio17, Precipitation of Driest Quarter; Bio18, Precipitation of Warmest Quarter; Bio19, Precipitation of Coldest Quarter.

versus not suitable, using the threshold of lower training presence (LTPT) (*Pearson et al.*, 2006) under an allowable error rate of E = 5%.

RESULTS

The high statistical performance was evidenced in all modeling predictions (ROCp > 1.50) (Table 1). Suitable areas for species of montane distribution (B. splendens, C. vicina, L. purpurascens, and S. roraima) were associated with elevations; the Andes, Central Cordillera, and tabletop Tepuis in Venezuela. Remarkably, the modeling of potential distribution for B. splendens showed suitability areas associated with the Andean and Caribbean regions. These fitting areas correspond to montane ranges along the Andes chain in Peru, Ecuador and Colombia, including the Sierra Nevada de Santa Marta, the Mérida cordillera and the coastal range in Venezuela, but not reaching the tepuis (Fig. 2C). Modeling for S. roraima showed large suitable areas uninhabited within the Tepuyan province in the Guyana shield (Fig. 2B). The smallest potential area predicted was obtained for *C. vicina* modeling in the Andean cordillera from Peru to Venezuela, the Cordillera Central, and Venezuelan Tepuis (Fig. 2C). Modeling prediction for *L. purpurascens* was similar to the previous species, including montane regions of the Andes, Pacific, Caribbean, and Tepuis of Venezuela (Fig. 2F). On the other hand, the modeling of C. macellaria showed a preference for lowlands areas of the Pacific, Orinoquía, and Amazonia regions (Fig. 2D). Finally, the model for C. idioidea with suitable areas was remarkably extended, almost occupying the entire area of study, except for some areas in the Colombian and Venezuelan Orinoquía, and a particular region of the Pacific and Caribbean of Colombia (Fig. 2E).



Figure 2 Potential distribution maps. Potential distribution maps of (A) *Blepharicnema splendens*, (B) *Sarconesia roraima*, (C) *Calliphora vicina*, (D) *Cochliomyia macellaria*, (E) *Chloroprocta idioidea* and (F) *Lucilia purpurascens*. Green circles indicate the locations of the records used for calibrating the model; gray areas are modeled suitable conditions; and white areas are unsuitable conditions. Maps at .kml are available for download in the Supplemental_Information.

Full-size DOI: 10.7717/peerj.10370/fig-2

DISCUSSION

In carrion-flies ensembles had been demonstrated that the seasonality and the habitat and resource availability are critical aspects of the species occurrence, especially in temperate regions (Smith & Wall, 1997; Martínez-Sánchez, Rojo & Marcos-García, 2000; Arias-Robledo, Stevens & Wall, 2019). Here we focus on the latter aspect since the study area assessed has uniform weather, typical of tropical environments (with dry and wet seasons only). The selected blow fly species have different biogeographical histories (*Rognes*, 1997). One group belongs to lowland neotropical species of wide distribution related to warm climates, C. macellaria and C. idioidea (James, 1970); while the second group belongs to temperate species linked to montane medium and high elevations; C. vicina, B. splendens, L. purpurascens, S. roraima (Dear, 1979; Amat, Vélez & Wolff, 2008; Whitworth, 2014). The spatial distribution modeling generated for montane species matches the altitudinal ranges of suitable settlement except for C. idioidea. Being the range altitude an essential element for modeling at these latitudes; this proves that elevation is an indirect variable that may provide a good surrogate for temperature across certain extents and latitudes (Austin, 2002). Montane insects are a highly specialized group with particular ecophysiological adaptations to living in extreme altitude environments (Mani, 1968; Hodkinson, 2005). One adaptation is cold-stenothermy, related to narrow thermal requirements to optimal growth. This condition limits the spatial distribution (*Hodkinson*, 2005), similarly to blow flies montane distributions displayed in this study. This analysis encompasses the environmental and ecological requirements mentioned above, deemed essential for the definition of distribution. However, we found no specific association between the potential distributions of species with a specific bioclimatic variable. The importance of particular environmental variables for a species may vary according to geographic and biotic contexts (Peterson et al., 2011), as evidenced with the modeling for blow flies.

Potential distribution prediction for *B. splendends* matches preliminary observations and occurrences as an endemic Andean taxon distributed from Bolivia to Venezuela made by *Amat & Wolff (2007)*, they reported it as rare and asynanthropic in the Andes inhabiting well preserved forest up to 2,500 m in altitude. The average annual temperature was the most critical variable in explaining the model. This result may suggest that a montane environment with a temperate climate and well-preserved areas under any conservation category (e.g., national park, reserves, sanctuary, and refuges.) may represent suitable conditions for *B. splendens* in the northwestern South America region.

Distribution modeling for *S. roraima* showed a similar montane pattern, being more restricted according to its altitudinal range along the Andean chain reaching Venezuelan Tepuis. *S. roraima* is recorded from Chile at an elevation of 3,250 m (*Dear, 1979*). In Venezuela, it was reported in the Tepuis, Mérida cordillera, and the coastal range at an elevation of 1,900–2,700 m (*Velásquez et al., 2017*). It was recorded in Colombia in the central and the eastern ridge at an elevation of 1,900 to 2,592 m (*Amat, 2009; Wolff & Kosmann, 2016*). Asynanthropic in the surrounding montane areas of Bogotá, Colombia (*Pinilla-Beltran, Segura & Bello, 2012*). Thus, additional records are expected in the Andean region above 1,900 m in a well-preserved forest, avoiding anthropogenic environments

or highly disturbed environments at both Andean slopes in Ecuador and eastern slopes in Peru, in the three Andean ranges of Colombia, including the Sierra Nevada de Santa Marta, and in Venezuela in the Mérida cordillera. Similarly, it is expected to be in the Caribbean region of Venezuela related to the coastal range, especially near the Henri Pittier National Park and elevation ranges at Cueva del Guácharo National Park; and finally, in the Tepuyan region up to Pico da Neblina National Park in Brazil.

L. purpurascens, one of the distinct species of *Lucilia* in the New World region (*Whitworth, 2014*), showed similar montane prediction, including areas above 1,100 m (Fig. 2F) distributed from Argentina to Mexico (*James, 1970*). In the South American Andes, it is relatively common at an elevation of up to 2,200 m (*Whitworth, 2014*). Along the cordillera in Peru, it was a rare fly, hemisynanthropic, and ranged 1,300–1,900 m (*Baumgartner, 1988*). Also, it is known inhabiting the central range in Colombia, up to 2,800 m (*Wolff & Kosmann, 2016*). It was reported reaching the Páramo ecosystem as an uncommon fly at approximately 3,000 m but is commonly related to forested and rural areas (*Amat, 2017*). Additionally, it was recorded in the Mérida cordillera and the coastal range up to 2,000 m in Venezuela (*Velásquez et al., 2017*). The current distributional pattern described above is positively reflected in the modeling obtained.

C. vicina, a cosmopolitan species, is strongly associated with urban locations (Rognes & Whitworth, 2012), and cold temperatures (Greco, Brandmayr & Bonacci, 2014). Due to the high degree of synanthropy in the Andean region, it is expected to occur in medium and large cities along the Andean mountain chain above 2,200 m, such as La Mesa (Trujillo), Mérida (Mérida), Tovar (Mérida), San Cristóbal (Táchira), Ocaña (Santander), Pamplona (Norte de Santander), Sogamoso (Boyacá), Tunja (Boyacá), Bogotá, Pasto (Nariño), Ipiales (Nariño), Ibarra (Imbabura), Quito (D.C.), Ambato (Tungurahua), Riobamba (Chimborazo), Cuenca (Azuay), and Loja (Loja). In the Caribbean region, it is expected in Caracas (D.C.) and within a considerable area in the province of Sucre. Finally, in the Tepuvan region, despite the modeling showing relatively suitable environmental conditions, doubts regarding its occurrence raised since no large urban places exist for their settlement. This species has been recorded in the low and intermediate elevations of Chile and Argentina. However, in Northwestern South America, it was collected in high elevations in the Andean region related to cold and large cities in Ecuador and Colombia (Amat, 2017). Pinilla-Beltran, Segura & Bello (2012) reported it as eusynanthropic in Bogotá at an elevation of 2,600 m. In temperate latitudes C. vicina is a year-round species, more commonly collected in the winter (Zabala, Díaz & Saloña Bordas, 2014; Arias-Robledo, Stevens & Wall, 2019). Based on South American records from Argentina and Chile, C. vicina seems to extend the elevation range upward as it approaches the Equator line (Rognes & Whitworth, 2012), following an occupancy tendency related to cold temperature, as evidenced by the model.

The potential distribution map for *C. macellaria* shows extensive suitable areas in the Amazonian region of Colombia, Venezuela and Brazil, the Orinoquía and inter-Andean valleys of Colombia, the Pacific region of Colombia and Ecuador, the Caribbean region of Colombia and Venezuela, and finally, in low elevations areas surrounding the Venezuelan Tepuis. This wide distribution matches the occurrence pattern mentioned by *Dear* (1985). *Baumgartner & Greenberg* (1985) stated that *C. macellaria* was presumably the most common blowfly in the neotropical region before the arrival of exotic *Chrysomya* spp. They referred to this species as premontane under 2,450 m in the central Andes of Peru, similar to Colombia, where it is spread across its five natural regions in forested, rural and urban areas (*Amat, 2009*). *Velásquez et al.* (2017) recently cited this species occurring in northern Venezuela up to 1,100 m. Despite *C. macellaria* being classified as eusynanthropic in Peru (S.I = +79) (*Baumgartner & Greenberg, 1985*) and Colombia (S.I = +69) (*Montoya, Sanchez & Wolff, 2009*), it is also undoubtedly related to rural and well-preserved environments (hemisynanthropic), as evidenced here by some collection records used for the modeling. Besides, to the suitable areas mentioned above, we expect its occurrence beyond these borders, including a larger portion of the Brazilian, Colombian, Ecuadorian and Peruvian Amazon, a larger area of the Orinoquía in Venezuela and the montane regions ranging up to 2,200 m. These results evidence the lack of exhaustive collection at these sites.

A puzzling result for C. idioidea modeling was evidenced since it is usually related to well-preserved rainforests in lowlands (Baumgartner, 1988; Amat, 2009). This distributional pattern contrasts with suitable areas here reflected (Fig. 2E), which includes large regions of montane and highlands environments (e.g., high Andean forest and Páramo) exceeding their expected altitudinal range. Several montane records above 1,000 m were included to build the model; these records belong to polymorphic specimens with the pattern of body and wings coloration being darker. C. idioidea was initially considered as a species complex (Shannon, 1926; Hall, 1948). However, Dear (1985) mentioned a notable variation of coloration pattern depending on the geographical distribution but still considered it a single species. Moreover, in Peru, Baumgartner & Greenberg (1985) noticed two types of phenotypes differing in anterior facets' size. Yusseff-Vanegas & Agnarsson (2017) in recent molecular studies based on mitochondrial (COI) and nuclear (ITS2) DNA sequences, in specimens from Central America and the Caribbean indicate at least the existence of two species in C. idioidea, they also evidenced some morphological differences. Morphological differences here detected and the extended distribution pattern predicted, led us to believe that the current model was probably performed based on at least two different life-histories. We suspect that montane specimens here reviewed may be part of a cryptic species of montane distribution, contrasting the taxonomic arrangement of *Dear* (1985). Caution is suggested to use this modeling since the current taxonomic status of C. idioidea and its populations must be revised.

CONCLUSIONS

In general, our results showed that montane distributional patterns affect the performance of SDMs in neotropical blow flies. Montane species models fit better with the known current distribution and the biogeographical region (Andean province). Contrarily, lowland species showed a puzzling distribution mismatching the expected areas, neither empirical data.

The use of flies synanthropy values derived from the Human Influence Index (HII) raster data set was a pivotal aspect of modeling; this variable improved the SDMs regardless

of their synanthropic classification (Asynanthropic, Hemisynantropic, or Eusynanthropic). While the warm areas of low lands occupy a vast portion of the area surveyed (68.7%), the temperate and cold areas are restricted to the montane environments, being these a smaller fraction (31.3%) of the complete area surveyed. These differences in terms of size proportion may intrinsically influence the performance of the fly models obtained. In other words, the larger the area to evaluate, the lower the performance of the model and vice-versa. Furthermore, spatial distribution is not only related to climate variables but also other ecological interactions and bionomical attributes (besides synanthropy) not included in the input data to modeling. Analyzing potential distribution based on this methodology allowed identifying possible taxonomic inaccuracies and the lack of exhaustive collection, especially for lowlands species, while better performance was evidenced for species with montane and temperate distribution. The information here provided, contributes to biogeographical knowledge and will certainly serve as a contribution to the use of blow flies' biological data for conservational and medical-legal purposes.

ACKNOWLEDGEMENTS

We acknowledge Adriana Pérez-Hoyos for compiling geographical data, Natalia Torres Moreno for data analysis support, all curators of the entomological collections visited, Álvaro Barragán, Emilia Moreno and Santiago Villamarín in Ecuador, José Clavijo and Quintin Arias in Venezuela, Marcio Oliveira in Brazil, Claudia Medina, Francisco Serna, Fredy Molano[†], Jhon Alveiro Quiroz "Black" and German Amat in Colombia.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This research was funded by TECNOLÓGICO DE ANTIOQUIA, CODEI-grant number 206001114. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors: TECNOLÓGICO DE ANTIOQUIA, CODEI: 206001114.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Mariano Altamiranda-Saavedra conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Eduardo Amat conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Luz Miryam Gómez-P conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability

The following information was supplied regarding data availability: Raw data are available as a Supplemental File.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.10370#supplemental-information.

REFERENCES

- **Amat E. 2009.** Contribución al conocimiento de las Chrysomyinae y Toxotarsinae (Diptera: Calliphoridae) de Colombia. *Revista Mexicana de Biodiversidad* **80:**693–708.
- Amat E. 2017. Calliphoridae (Diptera) do noroeste da América do Sul: Diversidade, distribuição e código de barras genético. D. Phil. Thesis, Instituto Nacional de Pesquisas da Amazônia. https://bdtd.inpa.gov.br/handle/tede/2608 (accessed on 21 November 2020).
- **Amat E, Medina CA. 2020.** A rapid ecological assessment for necrophagous flies (Diptera, 2020.07.24.22049 Calyptratae) in a mosaic landscape of the Colombian. In Press.
- Amat E, Vélez MC, Wolff M. 2008. Illustrated key for identification to genera and species of blowflies (Diptera: Calliphoridae) of Colombia. *Caldasia* **30**:231–244 DOI 10.1111/j.1365-3113.2010.00536.x.
- **Amat E, Wolff M. 2007.** New records of *Blepharicnema splendens* (Calliphoridae: Calliphorinae, Luciliini) from Colombia. *Revista de la Sociedad Entomológica Argentina* **66**:187–190.
- Anderson GS, Cervanka VJ. 2002. Insects associated with the body: their use and analysis. In: Haglund WD, Sorg MH, eds. Adv. Forensic Taphon. Method, Theory, Archeol. Perspect. Boca Raton: CRC Press, 173–200.
- Anderson RP, Lew D, Peterson AT. 2003. Evaluating predictive models of species' distributions: criteria for selecting optimal models. *Ecological Modelling* 162:211–232 DOI 10.1016/S0304-3800(02)00349-6.
- Arias-Robledo G, Stevens JR, Wall R. 2019. Spatial and temporal habitat partitioning by calliphorid blowflies. *Medical and Veterinary Entomology*.
- Austin MP. 2002. Spatial prediction of species distribution: an interface between ecological theory and statistical modelling. *Ecological Modelling* **33(2)**:228–237 DOI 10.1111/mve.12354.
- Barve N, Barve V, Jiménez-Valverde A, Lira-Noriega A, Maher SP, Peterson AT, Soberón J, Villalobos F. 2011. The crucial role of the accessible area in ecological niche modeling and species distribution modeling. *Ecological Modelling* 222:1810–1819 DOI 10.1016/j.ecolmodel.2011.02.011.
- Basanta MD, Rebollar EA, Parra-Olea G. 2019. Potential risk of *Batrachochytrium sala*mandrivorans in Mexico. PLOS ONE 14:1–13 DOI 10.1371/journal.pone.0211960.

- **Baumgartner D. 1988.** Spread of introduced *Chrysomya* blow flies (Diptera: Calliphoridae) in Neotropics with records new to Venezuela. *Biotropica* **20**:167–168 DOI 10.2307/2388191.
- Baumgartner DL, Greenberg B. 1985. Distribution and medical ecology of the blow flies (Diptera: Calliphoridae) of Peru1. Annals of the Entomological Society of America 78:565–587 DOI 10.1093/aesa/78.5.565.
- **Bishopp FC, Laake EW. 1921.** Dispersal of flies by flight. *Journal of Agricultural Research* **21**:726–766.
- **Dear JP. 1979.** Revision of the Toxotarsinae (Diptera: Calliphoridae). *Papeis Avulsos de Zoologia* **32(13)**:145–182.
- **Dear JP. 1985.** A revision of the New World Chrysomyini (Diptera: Calliphoridae). *Revista Brasileira de Zoologia* **3**:109–169 DOI 10.1590/S0101-81751985000300001.
- Elith J, Leathwick JR. 2009. Species distribution models: ecological explanation and prediction across space and time. *Annual Review of Ecology, Evolution, and Systematics* 40:677–697 DOI 10.1146/annurev.ecolsys.110308.120159.
- Fick SE, Hijmans RJ. 2017. Worldclim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* **9**:4302–4315.
- Francesconi F, Lupi O. 2012. Myiasis. *Clinical Microbiology Reviews* 25(1):79 LP–105 DOI 10.1128/CMR.00010-11.
- Galante E, Marcos-Garcia MA. 1997. Detritívoros, Coprófagos y Necrófagos. *Boletín de la Sociedad Entomológica Aragonesa* 20:57–64.
- Greco S, Brandmayr P, Bonacci T. 2014. Synanthropy and temporal variability of calliphoridae living in Cosenza (Calabria, Southern Italy). *Journal of Insect Science* 14(1):216 DOI 10.1093/jisesa/ieu078.
- **Greenberg B. 1973.** *Flies and disease II: biology and disease transmission*. Princeton: Princeton University Press.
- **Greenberg B, Kunich JC. 2002.** *Entomology and the law: flies as forensic indicators.* Cambridge: Cambridge University Press.

Gregor F, Povolný D. 1958. Versuch einer Klassifikation der synanthropen fliegen (Diptera). Journal Hygiene Epidemiology Microbiology & Inmunology 2:205–216.

- Hall DJ. 1948. The blowflies of North America. Lafayette: Thomas Say Foundation.
- Hodkinson ID. 2005. Terrestrial insects along elevation gradients: species and community responses to altitude. *Biological Reviews of the Cambridge Philosophical Society* 80:489–513 DOI 10.1017/S1464793105006767.
- Hwang C, Turner BD. 2005. Spatial and temporal variability of necrophagous Diptera from urban to rural areas. *Medical and Veterinary Entomology* 19:379–391 DOI 10.1111/j.1365-2915.2005.00583.x.
- James MT. 1970. Family Calliphoridae. In: Papavero N, ed. *A catalogue of the diptera of Americas south of the United States*. São Paulo: Museu de Zoologia, Universidade de São Paulo, 28.
- Kavazos CRJ, Wallman JF. 2012. Community composition of carrion-breeding blowflies (Diptera: Calliphoridae) along an urban gradient in south-eastern Australia. *Landscape and Urban Planning* 106:183–190 DOI 10.1016/j.landurbplan.2012.03.002.

- Kosmann C, Mello RP de, Harterreiten-Souza ÉS, Pujol-Luz JR. 2013. A list of current valid blow fly names (Diptera: Calliphoridae) in the Americas South of Mexico with key to the Brazilian species. *EntomoBrasilis* 6:74–85 DOI 10.12741/ebrasilis.v6i1.266.
- Lecheta MC, Corrêa RC, Moura MO. 2017. Climate shapes the geographic distribution of the blowfly *Sarconesia chlorogaster* (Diptera: Calliphoridae): an environmental niche modeling approach. *Environmental Entomology* **46**:1051–1059 DOI 10.1093/ee/nvx124.
- Lindquist AW, Yates WW, Hoffman RA, Butts JS. 1951. Studies of the flight habits of three species of flies tagged with radioactive phosphorus1. *Journal of Economic Entomology* 44:397–400 DOI 10.1093/jee/44.3.397.
- Mani MS. 1968. Ecological specializations of high altitude insects. In: *Ecology and biogeography of high altitude insects*. Dordrecht: Springer Netherlands, 51–74 DOI 10.1007/978-94-017-1339-9_3.
- Martínez-Sánchez A, Rojo S, Marcos-García MA. 2000. Annual and spatial activity of dung flies and carrion in a Mediterranean holm-oak pasture ecosystem. *Medical and Veterinary Entomology* 14:56–63 DOI 10.1046/j.1365-2915.2000.00205.x.
- Montoya GA, Sanchez JD, Wolff M. 2009. Sinantropía de Calliphoridae (Diptera) del Municipio La Pintada, Antioquia—Colombia. *Revista Colombiana de Entomologia* 35:73–82.
- Mulieri PR, Patitucci LD. 2019. Using ecological niche models to describe the geographical distribution of the myiasis-causing *Cochliomyia hominivorax* (Diptera: Calliphoridae) in southern South America. *Parasitology Research* 118:1077–1086 DOI 10.1007/s00436-019-06267-0.
- Muscarella R, Galante PJ, Soley-Guardia M, Boria RA, Kass JM, Uriarte M, Anderson RP. 2014. ENMeval: an R package for conducting spatially independent evaluations and estimating optimal model complexity for Maxent ecological niche models. *Methods in Ecology and Evolution* **5**:1198–1205 DOI 10.1111/2041-210X.12261.
- Norris KR. 1965. The bionomics of blow flies. *Annual Review of Entomology* 10:47–68 DOI 10.1146/annurev.en.10.010165.000403.
- Nuorteva P. 1963. Synanthropy of blowflies (Dipt., Calliphoridae) in Finland. *Annales Entomologicae Fennicae* 29:1–49.
- Osorio-Olvera L, Lira-Noriega A, Soberón J, Peterson AT, Falconi M, Contreras-Díaz RG, Barve V, Barve N. 2020. ntbox: an R package with graphical user interface for modeling and evaluating multidimensional ecological niches. *Methods in Ecology and Evolution* 2041–210X.13452 DOI 10.1111/2041-210X.13452.
- Pape T, Blagoderov V, Mostovski MB. 2011. Order Diptera Linnaeus, 1758. In: Zhang ZQ, ed. *Animal biodiversity: an outline of higher-level classification and survey of taxonomic richness. Zootaxa*. vol. 3148. 222–229.
- **Pearson RG, Raxworthy CJ, Nakamura M, Townsend Peterson A. 2006.** Predicting species distributions from small numbers of occurrence records: a test case using cryptic geckos in Madagascar. *Journal of Biogeography* **34**:102–117 DOI 10.1111/j.1365-2699.2006.01594.x.

- **Peterson AT, Papes M, Sober J. 2008.** Rethinking receiver operating characteristic analysis applications in ecological niche modeling. *Ecological Modelling* **3**:63–72 DOI 10.1016/j.ecolmodel.2007.11.008.
- Peterson AT, Sánchez-cordero V, Ben BC, Ramsey JM. 2002. Ecologic niche modeling and potential reservoirs for chagas disease, Mexico. *Emerging Infectious Diseases* 8(7):662–667 DOI 10.3201/eid0807.010454.
- Peterson AT, Soberon J, Pearson RG, Anderson RP, Martinez-Meyer E, Nakamura M, Araujo MB. 2011. *Ecological niches and geographic distributions*. Princeton University Press DOI 10.23943/princeton/9780691136868.001.0001.
- Peterson AT, Soberón J, Ramsey J, Osorio-Olvera L. 2020. Co-occurrence networks do not support identification of biotic interactions. *Biodiversity Informatics* 15:1–10 DOI 10.17161/bi.v15i1.9798.
- Phillips SJ, Anderson RP, Schapire RE. 2006. Maximum entropy modeling of species geographic distributions. *Ecological Modelling* 190:231–259 DOI 10.1016/j.ecolmodel.2005.03.026.
- Pinilla-Beltran YT, Segura NA, Bello FJ. 2012. Synanthropy of Calliphoridae and Sarcophagidae (Diptera) in Bogotá, Colombia. *Neotropical Entomology* 41(3):237–242 DOI 10.1007/s13744-012-0036-x.
- **Povolný D. 1971.** Synanthropy. In: Greenberg B, ed. *Flies and disease, ecology, classification, and biotic associations, 1.* Princeton: Princeton University Press 17–54.
- **Qiao H, Peterson AT, Campbell LP, Soberón J, Ji L, Escobar LE. 2016b.** NicheA: creating virtual species and ecological niches in multivariate environmental scenarios. *Ecography* **39**:805–813 DOI 10.1111/ecog.01961.
- Qiao H, Saupe EE, Soberón J, Peterson AT, Myers CE. 2016a. Impacts of Niche Breadth and Dispersal Ability on Macroevolutionary Patterns. *The American Naturalist* 188(2):149–162 DOI 10.1086/687201.
- Quarterman KD, Mathis W, Kilpatrick JW. 1954. Urban fly dispersal in the area of Savannah, Georgia. *Journal of Economic Entomology* 47:405–412 DOI 10.1093/jee/47.3.405.
- **Rabus B, Eineder M, Roth A, Bamler R. 2003.** The shuttle radar topography mission—a new class of digital elevation models acquired by spaceborne radar. *Journal of Pho-togrammetry and Remote Sensing* **57**:241–262 DOI 10.1016/S0924-2716(02)00124-7.
- Raghavan RK, Barker SC, Cobos ME, Barker D, Teo EJM, Foley DH, Nakao R, Lawrence K, Heath ACG, Peterson AT. 2019. Potential spatial distribution of the newly introduced long-horned tick, *Haemaphysalis longicornis* in North America. *Scientific Reports* 9:498 DOI 10.1038/s41598-018-37205-2.
- Richards CS, Williams KA, Villet MH. 2009. Predicting geographic distribution of seven forensically significant blowfly species (Diptera: Calliphoridae) in South Africa. *African Entomology* **17**:170–182 DOI 10.4001/003.017.0207.
- **Rognes K. 1997.** The Calliphoridae (Blowflies) (Diptera: Oestroidea) are not a monophyletic group. *Cladistics* **13**:27–66 DOI 10.1006/clad.1997.0031.

- **Rognes T, Whitworth T. 2012.** Identification of Neotropical blow flies of the genus *Calliphora* Robineau- Desvoidy (Diptera: Calliphoridae) with the description of a new species. *Zootaxa* **3209**:1–27 DOI 10.11646/zootaxa.3209.1.1.
- **Shannon RC. 1926.** Synopsis of the American Calliphoridae (Diptera). *Proceedings of the Entomological Society of Washington* **28**:115–139.
- Shura-Bura EV, Shaikow AD, Ivanova EV, Gazunova AL, Mitruikova MS, Fedorova KG. 1958. The character of dispersion from the point of release in certain species of flies of medical importance. *Entomologie Obozrenie* 37:336–346.
- Smith KE, Wall R. 1997. The use of carrion as breeding sites by the blowfly Lucilia sericata and other Calliphoridae. *Medical and Veterinary Entomology* 11:38–44 DOI 10.1111/j.1365-2915.1997.tb00287.x.
- Velásquez Y, Martínez-Sánchez AI, Thomas A, Rojo S. 2017. Checklist and distribution maps of the blow flies of Venezuela (Diptera, Calliphoridae, Mesembrinellidae). *ZooKeys* 645:103–132 DOI 10.3897/zookeys.645.6972.
- Whitworth T. 2014. A revision of the Neotropical species of *Lucilia* Robineau-Desvoidy (Diptera: Calliphoridae). *Zootaxa* 3810:76 DOI 10.11646/zootaxa.3810.1.1.
- Wildlife Conservation Society WCS, University C for IESIN-C-C. 2005. Last of the Wild Project, Version 2, 2005 (LWP-2): Global Human Influence Index (HII) Dataset (Geographic). 38108(1):001–076.
- Williams KA, Richards CS, Villet MH. 2014. Predicting the geographic distribution of *Lucilia sericata* and *Lucilia cuprina* (Diptera: Calliphoridae) in South Africa. *African Invertebrates* 55:157–170 DOI 10.5733/afin.055.0109.
- Wolff M, Kosmann C. 2016. Families Calliphoridae and Mesembrinellidae. *Zootaxa* 4122:856–875 DOI 10.11646/zootaxa.4122.1.7.
- Yan G, Liu S, Schlink AC, Flematti GR, Brodie BS, Bohman B, Greeff JC, Vercoe PE, Hu J, Martin GB. 2018. Behavior and electrophysiological response of gravid and nongravid *Lucilia cuprina* (Diptera: Calliphoridae) to carrion-associated compounds. *Journal of Economic Entomology* 111(4):1958–1965 DOI 10.1093/jee/toy115.
- Yusseff-Vanegas SZ, Agnarsson I. 2017. DNA-barcoding of forensically important blow flies (Diptera: Calliphoridae) in the Caribbean region. *PeerJ* 5:e3516 DOI 10.7717/peerj.3516.
- Zabala J, Díaz B, Saloña Bordas MI. 2014. Seasonal blowfly distribution and abundance in fragmented landscapes. Is it useful in forensic inference about where a corpse has been decaying? *PLOS ONE* 9(6):e99668 DOI 10.1371/journal.pone.0099668.